

ORIGINAL RESEARCH

# Whole-Genome Analysis of Two Copies of bla<sub>NDM-1</sub> Gene Carrying Acinetobacter johnsonii Strain Acsw 19 Isolated from Sichuan, China

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Guoyuan Zou Institute of Plant Nutrition and Resources, Beijing Academy of Agriculture and Forestry Science, Beijing, People's Republic of China Email gyzou@163.com **Purpose:** To characterize the genetic feature of the carbapenems resistant *Acinetobacter johnsonii* strain Acsw19 isolated from municipal sludge. This strain was found to carry two copies of  $bla_{\text{NDM-1}}$ , cmlB1-like gene, and  $bla_{\text{OXA-211}}$ -like gene along with other 8 antimicrobial resistance genes, 3 plasmids, 15 genomic islands and 8 prophages.

**Methods:** A carbapenem-resistant *Acinetobacter johnsonii* strain Acsw19 isolated from municipal sludge was subjected to whole-genome sequencing (WGS) via the PacBio and Illumina MiSeq platforms. Thereafter, the characteristic was analyzed by a series of bioinformatics software.

**Results:** The results showed that the genome of Acsw19 was consisted of a 3,433,749 bp circular chromosome and 3 circular plasmids, pAcsw19-1 (11,161 bp), pAcsw19-2 (351,885 bp) and pAcsw19-3 (38,391bp), respectively. Resistome analysis showed that Acsw19 carried 12 antimicrobial resistance genes, including 6 [cmB1-like,  $bla_{\text{NDM-1}}$ ,  $bla_{\text{OXA-58}}$ , aph (3')-Vla, msr(E) and mph(E)] in the plasmid pAcsw19-2 and 6 ( $bla_{\text{OXA-211}}$ -like,  $bla_{\text{NDM-1}}$ , aph(3'')-lb, aph(6)-ld, sul2, and floR) in the chromosome genome. Specifically, the cmlB1-like gene shared 86.33%, 71.7% and 71.9% similarities with the cmlB1, cmlA4 and cmlA8 gene, and the  $bla_{\text{OXA-643}}$  and  $bla_{\text{OXA-652}}$ , at the nucleotide level, respectively. Phylogenetic analysis showed that the  $bla_{\text{OXA-611}}$ -like gene and cmlB1-like gene had the closest evolutionary relationship with  $bla_{\text{OXA-643}}$  and cmlB1, respectively. These results indicated that the  $bla_{\text{OXA-211}}$ -like and cmlB1-like genes identified in the current study should be the novel variant resistance genes.

**Conclusion:** Carrying of two copies of  $bla_{\text{NDM-1}}$ , cmlB1-like,  $bla_{\text{OXA-211}}$ -like and along with other 8 antimicrobial resistance genes, 3 plasmids, 15 genomic islands and 8 prophages *Acinetobacter johnsonii* strain might increase the possibility of spreading of resistance genes.

Keywords: Acinetobacter johnsonii, bla<sub>NDM-1</sub>, bla<sub>OXA</sub>, genomic island

#### Introduction

Producing of carbapenemases, including the  $\beta$ -lactamases of Ambler classes A, B (metallo- $\beta$ -lactamases) and D, are the most common mechanism of bacterial carbapenems resistance. <sup>1-3</sup> Especially, the New Delhi Metallo- $\beta$ -lactamase (NDM), Klebsiella pneumoniae carbapenemase (KPC) and some Class D  $\beta$ -lactamases (CHDLs) have been identified worldwide in gram-negative bacterial isolates from clinical, environmental samples, and food animals, especially in *Enterobacteriaceae*, <sup>1,4-9</sup> and also in *Pseudomonas aeruginosa* and *Acinetobacter species*. <sup>10-12</sup> Carbapenem-resistant

Acinetobacter species are mainly associated with the carbapenem-hydrolyzing NDM and CHDLs, such as  $bla_{\rm OXA-23}$ -like,  $bla_{\rm OXA-24/40}$ -like,  $bla_{\rm OXA-21}$ -like,  $bla_{\rm OXA-211}$ -like gene. Bepecially, co-carrying of the  $bla_{\rm NDM}$  and  $bla_{\rm OXA}$  in clinical, food, environmental derived isolates of Acinetobacter species were prevalent in the world. In addition, the genes encoding  $bla_{\rm NDM}$  and  $bla_{\rm OXA}$  are known to be carried on some mobile genetic elements that inserted into the chromosome or plasmids, and it is suspected the mechanism of horizontal gene transfer (HGT) promotes the exchange of resistance genes among pathogenic microorganisms isolated from the clinical, environmental samples, and food animals. 11,22–25

In this study, we mainly characterized the genetic feature of a carbapenems resistant strain *Acinetobacter johnsonii* Acsw19 isolated from municipal sludge. This strain was found to carry two copies of  $bla_{\text{NDM-1}}$ , cmlB1-like gene, and  $bla_{\text{OXA-211}}$ -like gene along with other 8 antimicrobial resistance genes, three plasmids, 15 genomic islands and 8 prophages.

#### **Materials and Methods**

# Bacterial Isolate, Identification and Antimicrobial Susceptibility Testing

The sample of municipal sewage was obtained from the influx of wastewater-related plant in Luzhou City (Sichuan Province, China) in March 2019. The sewage was 1:10 diluted and an aliquot (10 µL) was streaked onto a CHROM Agar Orientation (CHROMAgar, Paris, France) agar plate containing 2 mg/L meropenem (Solarbio, China) and then incubated at 37 °C overnight. Bacterial species identification was carried out by the Vitek2 system (BioMérieux, France), 16srRNA sequencing and matrix-assisted laser desorption ionization time of flight (MALDI-TOF) mass spectrometry. The minimal inhibitory concentrations (MICs) of 15 antimicrobial agents (Solarbio, China) including meropenem, imipenem, cefepime, cefotaxime, ceftazidime, piperacillin-tazobactam, amoxicillinclavulanic acid, gentamicin, amikacin, aztreonam, erythromycin, chloramphenicol, sulfadiazine, colistin and ciprofloxacin were determined by broth microdilution method according to the recommendations of the Clinical and Laboratory Standards Institute (CLSI 2013, M100-S23). Escherichia coli strain ATCC 25,922 was used as quality control. Polymerase chain reaction (PCR) amplification and DNA sequencing were performed to identify the key carbapenemase-encoding genes  $(bla_{NDM})$  and  $bla_{KPC}$ ) as previously reference.<sup>7</sup>

### Whole-Genome Sequencing and Analysis

Genomic DNA of the strain A. johnsonii Acsw19 was extracted using the DNA Kit (QIAGEN, Germany). The 10kb sequencing library and a 300 bp paired-end library were constructed using the standard PacBio RS sample and Illumina DNA sample preparation instructions, and then sequenced on Pacific Biosciences RS II and MiSeq systems sequencing platforms (Novogene, China). The reads were de novo assembled using the software Celera Assembler (version 8.0). Gene prediction was performed for the whole genome with Glimmer 3.02 (http://www.cbcb.umd.edu/soft ware/glimmer/).<sup>26</sup> And the annotation of the Acsw19 genome was achieved using the NCBI Prokaryotic Genome Annotation Pipeline. Pairwise alignment was performed by BLASTn search (http://blast.ncbi.nlm.nih.gov/Blast. cgi). The average nucleotide identity (ANI) analysis was performed by the computer.<sup>27</sup> The resistome was identified using ResFinder 2.1 (https://cge.cbs.dtu.dk/services/ ResFinder/)<sup>28</sup> (minimum threshold for identity, 85%; minimum coverage, 60%) and Comprehensive Antibiotic Resistance Database (CARD) (https://card.mcmaster.ca/). The genomic island sequences were predicted based on three different genomic islands (GIs) prediction software (IslandPATH-DIMOB, IslandPick, and SIGI-HMM)<sup>29-31</sup> and the Prophage was predicted by using phiSpy.<sup>32</sup>

#### **Results and Discussion**

# Bacterial Isolate, Identification and Resistance Gene Detection

A gram-stain-negative,  $bla_{\text{NDM-1}}$  and  $bla_{\text{OXA}}$  producing Acinetobacter johnsonii Acsw19 was isolated and identified by the Vitek2 system (BioMérieux, France), 16srRNA sequencing and matrix-assisted laser desorption ionization time of flight (MALDI-TOF) mass spectrometry in Luzhou City, Southwestern China. Acinetobacter johnsonii strain Acsw19 was resistant to meropenem, imipenem, cefepime, cefotaxime, ceftazidime, piperacillintazobactam, amoxicillin-clavulanic acid, gentamicin, amikacin, aztreonam, erythromycin, chloramphenicol and sulfadiazine and it was susceptible to colistin and ciprofloxacin according to CLSI breakpoints (M100-S23) (Supplementary Table 1). To the best of our knowledge, the occurrence of multiple antibiotic genes in the multidrug resistant bacterial isolates from sewage has been evidenced in numerous studies, with the involvement of numerous species. 33,34

# Characterization of the Whole Genome of Acinetobacter johnsonii Strain Acsw 19

We got 1,876,876,663 bp data of the whole genome by the WGS. The genome of *Acinetobacter johnsonii* Acsw19 consisted of a 3,433,749bp circular chromosome and three circular plasmids, pAcsw19-1, pAcsw19-2 and pAcsw19-3 in the size of 11,161bp, 351, 885bp and 38, 391 with the 35.66%, 38.79% and 33.76% G+C content, respectively. The chromosome has a 41.78% G+C content, 21 rRNA operons, 88 tRNAs, 44ncRNAs and 3413 predicted protein coding sequences (CDSs) (Table 1).

### Resistome Analysis

A total of 12 drug-resistance genes was detected in the whole genome of Acsw19. Specifically, 6 resistance genes [(bla<sub>NDM-1</sub>, bla<sub>OXA-211</sub>-like, aph(3")-Ib (strB), aph(6)-Id (strA), sul2, and floR)] were located in the chromosome genome and 6 [cmlB1-like, mph(E), msr(E), bla<sub>OXA-58</sub>, bla<sub>NDM-1</sub> and aph(3")-VIa] in plasmid pAcsw19-2 (Table 2). None of the drug-resistance gene was detected in plasmid pAcsw19-1 and

**Table I** Characteristic of the Whole Genome of Acinetobacter johnsonii Acsw19

Context	Size (bp)	G+C (%)	No. of Predicted ORFs
Chromosome	3,433,749	41.78%	3413
Plasmid pAcsw19-1	11,161	35.66%,	18
Plasmid pAcsw19-2	351,885	38.79%	383
Plasmid pAcsw19-3	38,391	33.76%	58

pAcsw19-3. The full lengthen of *cmlB1*-like gene consisted of 1266 nucleotides encoding a protein with 422 amino acids. Sequence analysis showed that the cmlB1-like gene shared 86.33%, 71.7% and 71.9% sequence similarities with the known cmlB1, cmlA4, and cmlA8 genes at nucleotide level and 67.8%, 44.8% and 44.5% at amino acid level, respectively. The full lengthen of bla<sub>OXA-211</sub>-like gene consisted of 825 nucleotides encoding a protein with 275 amino acids. Sequence analysis showed that the bla<sub>OXA-211</sub>-like gene shared 94.4%, 95.39%, 95.52%, 96% and 96.36% identity with the known bla<sub>OXA-211</sub>, bla<sub>OXA-643</sub>, bla<sub>OXA-281</sub>, bla<sub>OXA-645</sub> and bla<sub>OXA-652</sub> gene at nucleotide level and 94.4%, 95.4%, 95.5%, 96% and 96.4% at amino acid level, respectively. Phylogenetic analysis showed that the bla<sub>OXA-211</sub>-like gene and cmlB1-like gene had the closest evolutionary relationship with bla<sub>OXA-643</sub> (Figure 1) and cmlB1 (Figure 2). These results indicated that the  $bla_{OXA-211}$ -like and cmlB1-like genes maybe two new allelic variant of the gene  $bla_{OXA}$  and cmlB1. To our the knowledge, there are more new allelic variant of the resistance genes have been found from the sewage derived isolates. 33,35,36

#### Characterization of Three Plasmids

Plasmid pAcsw19-1 contained 18 putative coding open reading frames (ORFs). Sequence analysis showed that pAcsw19-1 had 100%, 92%, 68% and 68% query cover and 96.55%, 91.5%, 87.8% and 87.8% sequence similarities with the plasmids p2\_010062 (CP033122), p4\_010055 (CP032283), p3\_010030 (CP029391) and pALWEK1.4 (CP032107) at nucleotide level, and these reported plasmids

Table 2 Distribution of the Resistance Genes in Acinetobacter johnsonii Strain Acsw19

	Resistance Gene	Identity %	Query/ Template Length	Position in Context	Predicted Phenotype	Accession Number
Chromosome	bla <sub>NDM-1</sub> bla <sub>OXA-211-like</sub> aph(3")-lb(strB) aph(6)-ld (strA) sul2 floR	100 95.64 99.88 100 100 98.35	813/813 826/825 804/804 837/837 816/816 1214/1215	3,091,502.3092314 16,507.17331 2,803,602.2804405 2,802,766.2803602 2,797,798.2798613 2,801,117.2802330	Beta-lactam resistance Beta-lactam resistance Aminoglycoside resistance Aminoglycoside resistance Sulphonamide resistance Phenicol resistance	FN396876 HG931732 AF321551 M28829 AY034138 AF118107
pAcsw19-2	mph(E) msr(E)  aph(3')-Vla bla <sub>NDM-1</sub> bla <sub>OXA-58</sub> cmlB l -like	100 100 100 100 100 86.51	885/885 1476/1476 780/780 813/813 843/843 1268/1266	47,669.48553 48,609.50084 256,925.257704 250,228.251040 57,613.58455 28,217.29482	Macrolide resistance Macrolide, Lincosamide and Streptogramin B resistance Aminoglycoside resistance Beta-lactam resistance Beta-lactam resistance Phenicol resistance	DQ839391 FR751518 X07753 FN396876 AY665723 AM296481

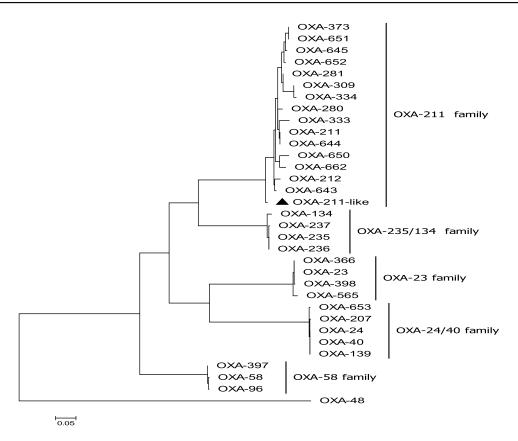


Figure I Molecular phylogenetic analysis by maximum likelihood method with 1000 bootstraps of OXA-211 like (a novel variant of OXA-211 family) with the six Class D  $\beta$ -lactamases (CHDLs) OXA family (OXA-23, OXA-24/40, OXA-235/134, OXA-58, OXA-48, OXA-211) representative sequences retrieved from GenBank database. The black triangle was indicated the gene which was found in this study.

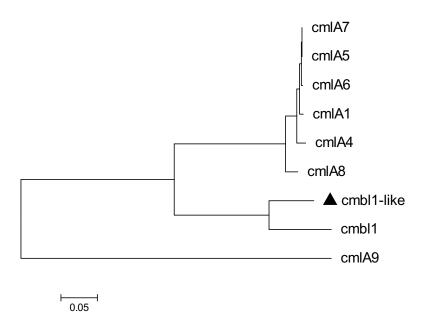


Figure 2 Molecular phylogenetic analysis by maximum likelihood method with 1000 bootstraps of CmIA/FloR family chloramphenicol efflux MFS (cmIB1-like) with the eight CmIA/FloR family chloramphenicol efflux MFS representative sequences retrieved from GenBank database. The black triangle was indicated the gene which was found in this study.

were all harbored by the *Acinetobacter* species. None of antimicrobial resistance gene was determined in plasmid pAcsw19-1. Plasmid pAcsw19-1 carried two copies of

plasmid replicons, two mobilization proteins (*mobL*-like), and several hypothetical ORFs. Complete sequence analysis showed that the 351,885 bp plasmid pAcsw19-2 contained

383 putative coding ORFs. Plasmid pAcsw19-2 had 87.34%, 14.76% and 7.06% query cover and 99.28%, 99.78% and 100% sequence similarities with the plasmids pXBB1-9 (CP010350), pACI-df08 (CP026426)<sup>37</sup> and pM131-2 (JX101647) at nucleotide level, respectively.<sup>11</sup>

Plasmid pAcsw19-2 carried 6 resistance genes [cmlB1like, mph(E), msr(E), bla<sub>OXA-58</sub>, bla<sub>NDM-1</sub>, and aph(3')-VIa] which were distributed in three regions. The first region carried the cmlB1-like gene. The genetic context of this region was ISL3-like transposase-cmlB1-like-LysR (Figure 2). This region was similar to the corresponding area of plasmid pOXA58 010030 (CP029396), pOXA58 010055 (CP032285), and pXBB1-9 (CP010351). The second region carried three resistance genes  $[mph(E), msr(E), and bla_{OXA-58}]$ . The first two resistance gene mph(E) and msr(E) were carried by the genetic context which was constituted by IS30 family transposase, helix-turn-helix-ORF, mph(E), msr(E), helix-turn -helix-ORF, plasmid stability associated protein-coding ORFs (addiction module antidote protein, RelE/ParE, brnA/T, and LysE family), ISAba1, and AraC transcriptional regulator gene. The AraC was linked to the third resistance gene bla<sub>OXA-58</sub> carrying area which was constituted by ISAba3bla<sub>OXA58</sub>-ISAba3 (Figure 3). The context of bla<sub>OXA-58</sub> carrying area (ISAba3-bla<sub>OXA58</sub>-ISAba3) was same to the previous reports. 11,38-40

The third region carried 2 resistance genes [plasmid borne *bla*<sub>NDM-1</sub> and *aph* (3')-*VIa*]. Nucleotide sequence analysis revealed that the *bla*<sub>NDM-1</sub> gene was flanked in the upstream region of *IS*5 transposase-*dmc*-unknown ORF-*umuD*-Y-family DNA polymerase-unknown ORF-

trpF-ble<sub>MBL</sub> and downstream by the IS30 family transposase and 6 unknown ORFs carried region, which linked to the resistance gene aph(3')-Via carrying genetic context [ISAba125-aph(3')-Via -ISAba125]. This resistance area (ISAba125-aph(3')-Via-ISAba125) was similar to the corresponding region of the plasmid pAP43-OXA58-NDM1 (CP043053), which was harbored by Acinetobacter pittii.

The plasmid pAcsw19-3 contained 58 putative coding ORFs. Sequence analysis showed that pAcsw19-3 had 72%, 76%, and 83% query cover and 99.90%, 99.96% and 99.84% sequence similarities with the plasmids p3\_010055 (CP032282.1), p2\_010030 (CP029390), and p4\_010060 (CP031712) at the nucleotide level, and these plasmids were all harbored by the *Acinetobacter species* strains. None of the antimicrobial resistance gene was determined in plasmid pAcsw19-3, either.

# Characterization of the Genomic Islands (GI)

Fifteen genomic islands, named GI Acsw19-1 to GI Acsw19-15, were identified by the software IslandPATH-DIMOB, IslandPick and SIGI-HMM. Sequence analysis showed that the length of the 15 genomic islands were ranged from ~5.1 kb to ~94.86 kb with the average G+C context of 32.59% to 54.18%, respectively. Moreover, 13 genomic islands (GI Acsw19-1 to GI Acsw19-13) were located in the chromosome and 2 genomic islands (GI Acsw19-14 and GI Acsw19-15) in the plasmid pAcsw19-2. Among the 15 GIs, two (GI Acsw19-11 and GI Acsw19-12) were the resistant

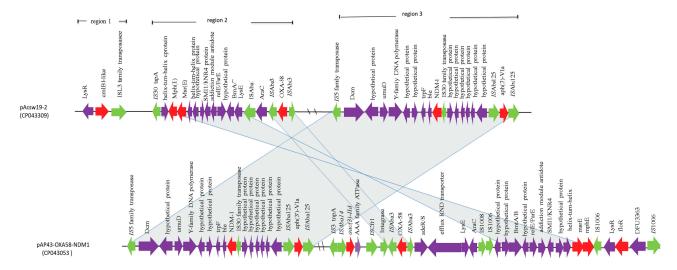


Figure 3 Schematic map of the genetic context of resistance gene regions in pAcSW19-2. The resistance genes are indicated by red arrows, the mobile genes are indicated by the green arrows and other function genes are indicated by the purple arrows. Gray areas between open reading frames (ORFs) denote nucleotide identities with the similarity context.

 Table 3 Overall Features of the Acinetobacter johnsonii Strain Acsw19 Genomic Islands

Gls_id	Location (Start-End)	Length (bp)	G+C %	Closest Match in Genbank (Query Cover and Identity)	Resistance Genes and Mobile Genes Carried
GI_Acsw19-1	Chromosome (1,005,689–1,014,205)	8,517	38.56	Acinetobacter johnsonii strain M19 chromosome genome (99%, 98.97%)	None
GI_Acsw19-2	Chromosome (1,533,242–1,543,640)	10,399	39.81	Acinetobacter johnsonii strain IC001 chromosome genome (96% 98.97%)	IS3 family transposase
GI_Acsw19-3	Chromosome (1,804,441–1,809,505)	5,065	36.92	Acinetobacter haemolyticus strain TJS01 chromosome genome (58%, 98.68%)	Integrase
GI_Acsw19-4	Chromosome (1,816,879–1,835,718)	18,840	39.44	Acinetobacter sp. WCHA45 plasmid pNDMI_010045 (34%, 90.83%)	IS1, IS3 and three IS5 family transposase
GI_Acsw19-5	Chromosome (1,886,561–1,897,546)	10,986	34.84	Acinetobacter johnsonii strain XBBI chromosome genome (88%, 96.43%)	ISAha1 family transposase
GI_Acsw19-6	Chromosome (1,919,728–1,925,691)	5964	35.78	Acinetobacter haemolyticus strain AN54 chromosome gemome (35%, 98.66%)	Two IS5 family transposases
GI_Acsw19-7	Chromosome (2,033,708–2,038,927)	5220	37.09	Acinetobacter johnsonii strain XBBI chromosome genome (26%, 85%)	None
GI_Acsw19-8	Chromosome (2,531,192–2,545,937)	1,4746	38.74	Acinetobacter johnsonii strain M19 chromosome genome (99%, 96.27%)	IS5 family transposase
GI_Acsw19-9	Chromosome (2,555,415–2,582,484)	27,070	36.29	Acinetobacter johnsonii strain LXL_C1 chromosome genome (79%, 99.09%)	Two IS3 family transposases, ISAha1 family transposase
GI_Acsw19-10	Chromosome (2,586,932–2,600,597)	13,666	39.02	Acinetobacter sp. LoGeW2-3 chromosome, genome (70%, 92.78%)	Integrase
GI_Acsw19-11	Chromosome (2,780,884–2,807,016)	26,133	42.83	Acinetobacter baumannii MRSN 15313 chromosome genome (47%, 100%)	ISI family transposase, Integrase, ISAba1, sul2, IS91-like transposase, aph(3")-lb, aph(6)-ld, ISAba, floR
Gl_Acsw19-12	Chromosome (3,082,734–3,095,042)	12,309	54.18	Acinetobacter baumannii strain IOMTU 433 complete genome (96%, 100%)	IS91 family transposase, NDM-1 and Two ISAba125
Gl_Acsw19-13	Chromosome (3,233,129–3,327,995)	94,867	43.12	Acinetobacter nosocomialis strain KAN01 chromosome genome (64%, 87.11%)	ISS family transposase, Integrase
GI_Acsw19-14	pAcsw19-2 (59,672–67,958)	8287	32.59	Acinetobacter sp. WCHA55 pOXA58_010055 (84%, 99.98%)	IS6-like transposase
GI_Acsw19-15	pAcsw19-2 (293,855–301,112)	7258	42.37	Acinetobacter sp. WCHA55 pOXA58_010055 (99%, 100%),	IS4 family transposase

genomic islands. Genomic island GI Acsw19-11 (26,133) bp) carried the aminoglycosides resistance genes strA and strB, sulphonamides resistance gene sul2 and phenical resistance gene floR. Genomic island GI Acsw19-12 (12,309 bp) carried the chromosome borne  $bla_{NDM-1}$ (Table 3). Sequence analysis showed that GI Acsw19-11 had 47%, 37%, and 34% query cover and 100% sequence similarities with the DNA sequence of A. baumannii MRSN15313 chromosome genome (CP033869), A. pittii WCHAP100004 plasmid pOXA58 100004 (CP027249), and uncultured bacterium HHV216 plasmid pHHV216 (FJ012880). Based on the specific genetic content, GI Acsw19-11 could be divided into two regions (regions A and regions B) (Figure 4). The region A was 15,713bp in length which served as the backbone of GI Acsw19-11. It mainly carried the tyrosine-type recombinase/integrase gene (1137 bp) and an IS1 family tnpA, which might be responsible for encoding the site-specific resolvase and transposition. Additionally, the other genes of region A, including ferrous iron transporter gene *feoAB*, Cd(II)/Pb(II)-responsive transcriptional regulator encoding gene *cadR*, hydrolase encoding gene, some of hypothetical protein-encoding genes, were found to be located in the backbone. Sequence of region A was high similar to the corresponding region of other various *Acinetobacter* species. The 4 resistance genes (*strA* and *strB*, *sul2* and *floR*) carried region B was 8980 bp in length (Figure 3). These four resistance genes genetic context is *ISAba1-sul2-glmM-ISVsa3-LysR-floR-DUF3363-strB-strA-ISAba1*. The *sul2 and floR* carrying area (6814 bp) was similar to the corresponding region of plasmid pOXA58\_100004,

while the *strB* and *strA* carrying area (2832 bp) was similar to corresponding region of plasmid pNDM-AP \_882 harbored by the *A. pittii* AP\_882 (CP014478.1).

Sequence analysis showed that GI Acsw19-12 had 96%,

Sequence analysis showed that GI\_Acsw19-12 had 96%, 96%, and 96% query cover and 100%, 99.9%, and 99.9% sequence similarities with the corresponding region of sequences of *A. baumannii* AR\_0083 genome (CP027528),

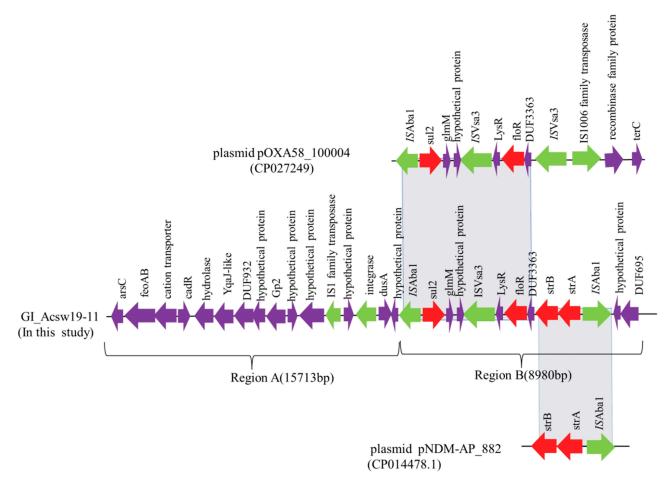


Figure 4 Schematic map of the genetic context of the GI\_Acsw19-11. The resistance genes are indicated by red arrows, the mobile genes are indicated by the green arrows and other function genes are indicated by the purple arrows. Gray areas between open reading frames (ORFs) denote nucleotide identities with the similarity context.

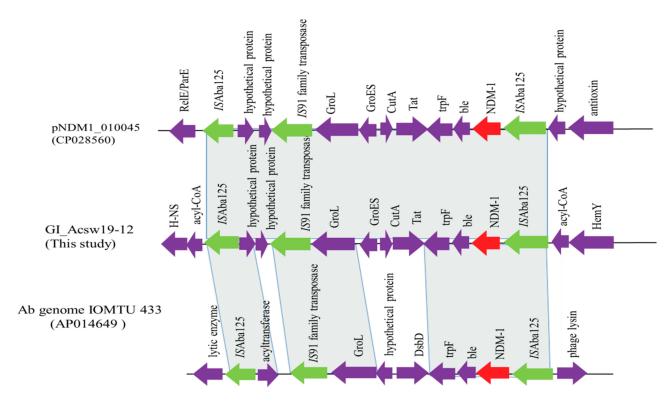


Figure 5 Schematic map of the genetic context of the GI\_Acsw19-2. The resistance genes are indicated by red arrows, the mobile genes are indicated by the green arrows and other function genes are indicated by the purple arrows. Gray areas between open reading frames (ORFs) denote nucleotide identities with the similarity context.

A. pittii ST220 genome (CP029610), and A. baumannii AR\_0083 genome (CP027528). GI\_Acsw19-12 had 82%, 82%, and 84% query cover and 100%, 100%, and 99.9%

sequence similarities with the plasmids including the pNDM1\_060092 (CP035935), pNDM-GJ01 (KT965092)<sup>42</sup> and pNDM-JVAP01(KM923969)<sup>43</sup> at nucleotide level.

Table 4 Overall Features of the Acinetobacter johnsonii Strain Acsw19 Prophage

Prophage_ID	Location (Start- End)	Length (bp)	G+C %	Closest Match in Genbank (Covery % and Identity %)	Resistance Genes and Mobile Genes Carried
Pp_Acsw19-1	Plasmid pAcsw19-2 (4463-36,078)	31,616	42.82	Acinetobacter defluvii WCHA30 plasmid pOXA58_010030 (99%, 99.98%)	cm/BI-like, Integrase, IS3, ISL3, IS1006 and two IS6 family transposases
Pp_Acsw19-2	Chromosome (1,557,755–171,680)	159,049	40.78	Acinetobacter johnsonii strain M19 chromosome genome (97%, 97.04%)	None
Pp_Acsw19-3	Chromosome (1,757,515–182,333)	65,817	40.03	Acinetobacter johnsonii strain M19 chromosome genome (64%, 95.15%)	Integrase, ISI family transposase, ISS family transposase
Pp_Acsw19-4	Chromosome (1,889,367–1,972,910)	83,544	40.3	Acinetobacter johnsonii strain M19 chromosome genome (92%, 97.11%)	4 IS5 family transposases
Pp_Acsw19-5	Chromosome (2,031,361–2,061,501)	30,141	40.65	Acinetobacter johnsonii strain M19 chromosome genome (67%, 95.92%)	Site-specific integrase
Pp_Acsw19-6	Chromosome (2,957,108–297,432)	17,222	43.01	Acinetobacter johnsonii XBB1 genome (99%, 97.34%)	None
Pp_Acsw19-7	Chromosome (3,085,102–3,130,818)	45,717	47.12	Acinetobacter sp. WCHA55 chromosome genome (99%, 93.93%)	IS91 family transposase, bla <sub>NDM-1</sub> and two ISAba125
Pp_Acsw19-8	Chromosome (3,156,419–3,188,118)	31,700	42.5	Acinetobacter johnsonii strain M19 chromosome genome (99%, 96.24%)	None

Moreover, the chromosome borne *bla*<sub>NDM</sub> was located in the genetic element (10,023 bp) *ISA*ba125-*IS*91-family tnpA-*groL-groES-cutA-tat-trpF-ble-bla*<sub>NDM-1</sub>-*ISA*ba125 (Figure 5). This *bla*<sub>NDM</sub> carrying element was similar equal to the corresponding region of the pNDM1 010045.

### Characterization of Prophages

A total of eight prophages, named Pp Acsw19-1 to Pp Acsw19-8, was identified by phiSpy (Table 4). Sequence analysis showed that one prophage (Pp Acsw19-1) was located in the plasmid pAcsw19-2 and 7 prophages (Pp Acsw19-2 to Pp Acsw19-8) were located in the chromosome genome. The lengthen of eight prophages was ranged from ~17.22 kb to ~159 kb with average G+C context of 40.03% to 47.12%, respectively. Interestingly, the  $bla_{NDM}$ carrying genomic islands GI Acsw19-12 was located in the prophage Pp\_Acsw19-7 and the cmlB1-like gene carrying prophage Pp Acsw19-1 was located in the plasmid pAcsw19-2. These findings suggested that some mobile genetic elements can be excised from chromosome or mobile genetic elements and then integrated into the chromosome or other mobile genetic elements. 44–47 These situations may promote the exchange of resistance genes among pathogenic microorganisms.

### **Conclusions**

The current study characterize the complete genome sequence of *Acinetobacter* Acsw19, which carried two copies of *bla*<sub>NDM-1</sub>, *cmlB1*-like gene, *bla*<sub>OXA-211</sub>-like gene along with other 8 antimicrobial resistance genes in 3 plasmids, 15 genomic islands and 8 prophages, which could provide great genetic plasticity for the host dissemination of antimicrobial resistance. The occurrence of multiple antibiotic genes in bacterial isolates from sewage has been evidenced in numerous studies, with the involvement of numerous species, it may be serving as an important reservoir of resistance genes and a hot spot for the transfer of resistance genes and mobile genetic elements. We should be vigilant these isolates or resistance genes transfer to the clinical bacterial.

# Nucleotide Sequence Accession Numbers

The genome sequence of *Acinetobacter johnsonii* Acsw19 is deposited in the NCBI database under accession numbers CP043307 (chromosome) and CP043308 to CP043310 (plasmid pAcsw19-1 to pAcsw19-3).

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### **Disclosure**

The authors declare no conflict of interest.

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