



Molecular Mechanisms in Clear Cell Renal Cell Carcinoma: Role of miRNAs and Hypermethylated miRNA Genes in Crucial Oncogenic Pathways and Processes

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Clear cell renal cell carcinoma (ccRCC) is the third most common urological cancer, and it has the highest mortality rate. The increasing drug resistance of metastatic ccRCC has resulted in the search for new biomarkers. Epigenetic regulatory mechanisms, such as genome-wide DNA methylation and inhibition of protein translation by interaction of microRNA (miRNA) with its target messenger RNA (mRNA), are deeply involved in the pathogenesis of human cancers, including ccRCC, and may be used in its diagnosis and prognosis. Here, we review oncogenic and oncosuppressive miRNAs, their putative target genes, and the crucial pathways they are involved in. The contradictory behavior of a number of miRNAs, such as suppressive and anti-metastatic miRNAs with oncogenic potential (for example, miR-99a, miR-106a, miR-125b, miR-144, miR-203, miR-378), is examined. miRNAs that contribute mostly to important pathways and processes in ccRCC, for instance, PI3K/AKT/mTOR, Wnt-β, histone modification, and chromatin remodeling, are discussed in detail. We also separately consider their participation in crucial oncogenic processes, such as hypoxia and angiogenesis, metastasis, and epithelial-mesenchymal transition (EMT). The review also considers the interactions of long non-coding RNAs (IncRNAs) and miRNAs of significance in ccRCC. Recent advances in the understanding of the role of hypermethylated miRNA genes in ccRCC and their usefulness as biomarkers are reviewed based on our own data and those available in the literature. Finally, new data and perspectives concerning the clinical applications of miRNAs in the diagnosis, prognosis, and treatment of ccRCC are discussed.

Keywords: clear cell renal cell carcinoma, microRNA, target genes, angiogenesis, epithelial-mesenchymal transition, metastasis, hypermethylated miRNA genes, long non-coding RNA

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1

INTRODUCTION

Renal cell carcinoma (RCC) is diagnosed in approximately 90% of patients with kidney cancer. RCC has the highest mortality rate among urogenital cancers (Vasudev et al., 2012). Approximately 270,000 new cases of RCC and 116,000 RCC-related deaths are reported globally every year (Randall et al., 2014). RCC is a heterogeneous group of epithelial tumors, among which clear cell RCC (ccRCC) is the most common and accounts for 70–80% of the reported cases of RCC (Cairns, 2010). The severity of ccRCC is higher than that of papillary kidney cancer and chromophobe tumors (Cairns, 2010). Lack of effective diagnostics in the early stages of the disease, increasing mortality rate, and resistance to therapies in patients with metastatic ccRCC emphasize the need to discover new biomarkers that are applicable for the early diagnosis of ccRCC and detection of metastasis.

Epigenetic regulatory mechanisms involving DNA methylation at the genomic transcriptional level and the interaction of non-coding RNAs, in particular microRNAs (miRNAs), with target messenger RNA (mRNA) at the post-transcriptional level are important in the regulation of genes and proteins (Baylin and Jones, 2016). miRNAs are involved in the regulation of cell proliferation, differentiation, and stress responses, and in the regulation of other fundamental biological processes and signaling pathways (Cora et al., 2017). Alterations in expression and regulatory functions of miRNA can be one of the key factors of various pathogeneses. miRNAs are involved in the development of more than 300 diseases, including oncological diseases. The number of publications aimed at identifying target genes and signaling pathways that involve miRNAs has increased (Reddy, 2015; Dragomir et al., 2018; Gattolliat et al., 2018). miRNAs are important positive or negative regulators of all processes characteristic of the pathogenesis of tumors, which include control of the cell cycle, apoptosis, neo-angiogenesis, tissue invasion, and metastasis (Exposito-Villen et al., 2018; Goradel et al., 2018; Kashyap et al., 2018; Mens and Ghanbari, 2018).

miRNAs are classified as being oncogenic or oncosuppressive (tumor suppressive) based on their stimulating or inhibiting effects, respectively, on tumor development. The targets of oncogenic miRNAs usually include the mRNAs of the tumor suppressor genes. In contrast, the targets of miRNAs that act to suppress tumor development are oncogenes and genes involved in tumor progression. The regulation of tumor suppressor miRNAs involves methylation of the promoter regions of their genes. The methylation suppresses their expression and subsequently inhibits their suppressor function. These processes occur more often in genes encoding tumor suppressor miRNAs than in genes encoding tumor suppressor proteins (Kunej et al., 2011; Piletic and Kunej, 2016). Methylation of tumor suppressor miRNA genes and the interaction of miRNAs with target mRNAs have a systemic effect on key processes and signaling pathways involved in carcinogenesis (Lopez-Serra and Esteller, 2012; Loginov et al., 2015; Baylin and Jones, 2016). Studies of methylation and profiling of miRNA expression have driven the design of minimally invasive diagnostics, and recent advances in the field of cancer epigenomics have highlighted the profound

possibilities of these approaches in clinical practice (Li et al., 2015; Xing and He, 2016; Morris and Latif, 2017).

Epigenetic mechanisms involved in ccRCC genesis have received less attention than other cancers, such as colorectal, lung, breast, and prostate carcinomas. Nevertheless, many genes susceptible to hypermethylation have been identified, including *VHL*, *RASSF1A*, *CDH1*, and *APAF1*, and reported as promising biomarkers of ccRCC (for example, Dmitriev et al., 2014; Braga et al., 2015; Xing and He, 2016; Morris and Latif, 2017). Data on the expression profiles of miRNAs, their target genes in ccRCC, and their potential use in the clinic are accumulating and have been covered in earlier reviews (for example, Li et al., 2015; He Y.H. et al., 2018). However, the contribution of methylation of miRNA genes to epigenetic regulatory mechanisms in ccRCC remains unclear, although interesting results have been reported.

This review examines the expression profiles, targets, and functions of oncogenic and tumor suppressor miRNAs, their role in molecular mechanisms and signaling pathways of ccRCC, as well as their clinical potential. In addition, based on data obtained in our studies and the published literature, we consider in detail the advances made in studies of hypermethylated miRNA genes in ccRCC and their usefulness as biomarkers.

Several long non-coding RNAs (lncRNAs) have been recently described in RCC, as has their involvement in regulatory interactions as competing endogenous RNA (ceRNA), whereby lncRNAs can act as miRNA sponges to modulate gene expression (Li et al., 2017; Chan and Tay, 2018). Moreover, integrative analyses have enabled construction of a ccRCC regulatory ceRNA network comprising 89 lncRNAs, 10 miRNAs, and 22 mRNAs (Yin et al., 2018). In general, however, very few publications have addressed lncRNAs in relation to ccRCC (60 papers on the topic were found indexed in PubMed as of January 2019). In this review, we also consider some data on lncRNAs and their interactions with miRNAs that are relevant to ccRCC.

ONCOGENIC AND SUPPRESSIVE MIRNAS IN MAIN SIGNALING PATHWAYS OF CCRCC

Expression microchips and quantitative PCR have been used to identify miRNAs with decreased expression levels in kidney tumors, presumably oncosuppressor miRNAs, and with increased expression levels in ccRCC, presumably oncogenic miRNAs (for example, Li et al., 2015). The expression of the miRNA genes is generally evaluated by determining the levels of mature miRNA, which is very stable and thus an effective biomarker (Andorfer et al., 2011).

Tumor suppressor miRNAs include miR-34a (Yamamura et al., 2012; Zhang C. et al., 2014; Toraih et al., 2017), miR-30c (Huang J. et al., 2013), miR-30d (Wu et al., 2013), miR-99a (Cui et al., 2012), miR-133a (Kawakami et al., 2012), miR-133b (Wu et al., 2014), miR-138 (Ding et al., 2018), miR-141 (Li W. et al., 2014), miR-143 (Yoshino et al., 2013), miR-182 (Kulkarni et al., 2018), miR-187 (Zhao et al., 2013), miR-199a (Qin et al., 2018), miR-200c (Ding et al., 2018), miR-205 (Xu et al., 2018), and others. These miRNAs target mRNAs of oncogenes

or genes encoding proteins mediating the progression of kidney tumors. For example, the targets of the crucially important miR-34a are *c-MET* (Toraih et al., 2017), *c-MYC* (Yamamura et al., 2012), and *NOTCH1* (Zhang C. et al., 2014) oncogenes. All are involved in the proliferation and activation of the cell cycle. Many targets of suppressor miRNAs are associated with invasion and migration. For example, matrix metalloproteinase 9 (MMP-9) is a validated target of miR-133b (Wu et al., 2014). *VIM*, *EZH2*, *ZEB2* (Ding et al., 2018), and *HIF1A* (Song et al., 2011) are regulated by miR-138.

Typical oncogenic miRNAs include miR-7 (He H. et al., 2018), miR-21 (Li X. et al., 2014; Chen J. et al., 2017; Cui et al., 2017), miR-155 (Ji et al., 2017), miR-590-5p (Xiao et al., 2013), and others. Their targets, on the contrary, are tumor suppressor genes, which include *PDCD4* (Li X. et al., 2014), *PTEN* (Cui et al., 2017), *TIMP3* (Chen J. et al., 2017), *FOXO3A* (Ji et al., 2017), *PBRM1* (Xiao et al., 2013), non-coding *MEG3* (He H. et al., 2018), and others.

Modern methodologies have provided a great deal of information concerning the miRNA expression profiles in ccRCC (He H. et al., 2015; Wotschofsky et al., 2016). Sixty-three differentially expressed miRNAs have been identified by analyzing the massive sequencing data published in The Cancer Genome Atlas (Liang et al., 2017).

To date, a substantial amount of data has been obtained on the role of miRNA in the regulation of target genes in ccRCC and its pathogenesis (Jung et al., 2009; Redova et al., 2011; Heinzelmann et al., 2014). A study of miRNA and mRNA gene networks constructed on the basis of their expression profiles in ccRCC reported a key role for miR-106a-5p. The loss of this miRNA led to the increased expression of 49 putative targets (Müller and Nowak, 2014). Other miRNAs implicated in this study were miR-135a-5p (32 targets), miR-206 (28 targets), miR-363-3p (22 targets), and miR-216b (13 targets) (Müller and Nowak, 2014). The targets included genes that affect apoptosis, metastasis, cellular mobility, and oncogenes (c-MET, VEGFA, NRP2, and FLT1). A similar study (Butz et al., 2015) pinpointed miR-124-3p, miR-30a-5p, and miR-200c-3p as the most significant miRNAs affecting protein expression in ccRCC; the expressions of these miRNAs were often reduced.

Notably, since the von Hippel Lindau (*VHL*) gene has an important role in both familial and sporadic kidney cancer, the genes associated with VHL-dependent regulation are important candidates in studying the spectrum of miRNAs that alter *VHL* expression (Neal et al., 2010). Loss of VHL function results in constitutive activation of the hypoxia-inducible factor (HIF) pathway, which leads to hypoxia and subsequent expression of angiogenic factors. Oncogenic miRNAs that enhance the development of hypoxia and angiogenesis and their targets have been identified. The next section of this review is devoted to a detailed look at the VHL/HIF pathways and the involved miRNAs.

The significance of the phosphoinositide 3-kinase (PI3K)/protein kinase B (AKT)/mechanistic target of rapamycin (mTOR) pathway (Joosten et al., 2018) associated with the escape from apoptosis, growth, and proliferation of cells in kidney cancer must be noted. The Cancer Genome Atlas Research

Network indicates frequent mutations in the genes of this pathway in ccRCC. The PI3K/AKT/mTOR pathway can also stimulate HIF through mTOR modulation (Cancer Genome Atlas Research, 2013).

miR-148a targets *AKT2*, and this interaction leads to the suppression of cell growth, colony formation, migration, and invasion and tumor growth in xenografts. The miR-148a level is decreased in RCC and is inversely correlated with tumor size, stage, and metastasis to lymph nodes (Cao et al., 2017). In cultured RCC cells, the inhibition of *FLOT1* by miR-182-5p reduces the phosphorylation and activation of AKT2 and subsequent phosphorylation of FOXO3a (Xu et al., 2014). Oncosuppressor and transcription factor FOXO3A is translocated out of the nucleus upon phosphorylation by proteins such as Akt (protein kinase B). FOXO3a decreases proliferation and arrests cells in the G1 phase. The level of miR-182-5p is often reduced in RCC (Xu et al., 2014).

On the contrary, miR-122 mimic increases the level of phosphorylation of AKT2 and mTOR. In addition to its enhanced expression in RCC, miR-122 reduces the expression of the Sprouty RTK signaling antagonist 2 (*SPRY2*) gene. The product of this gene is the inhibitor of Ras/MAPK signaling pathway (Wang Z. et al., 2017). Thus, while miR-148a and miR-182-5p act as tumor suppressors (Fan Y. et al., 2016; Cao et al., 2017), miR-122 acts as an oncogene (Lian et al., 2013).

The expression of miR-99a was first shown to be downregulated in RCC tumor tissues and correlated with poor survival (Cui et al., 2012). The authors also reported that the use of its mimic suppressed cell growth, migration, invasion *in vitro* and *in vivo*, and induced cell arrest at the G1 phase (Cui et al., 2012). However, in contrast to these results, which indicated tumor suppression activity of miR-99a, a recent study (Oliveira et al., 2017) described the upregulation of miR-99a and downregulation of its target gene mTOR in most of the ccRCC samples examined. These findings indicated the oncogenic activity of miR-99a. Both studies implicated mTOR as the direct target of miR-99a in renal cancer cells. Further studies are needed to definitively determine the suppressive or oncogenic activity of miR-99a in ccRCC.

miR-144 also targets *mTOR* in RCC. The reduced expression of miR-144 has been inversely correlated with the stage and size of the tumor, while increased miR-144 expression can suppress cell growth and arrest cells in the G1 phase (Xiang et al., 2016). However, another study (Xiao et al., 2017) identified miR-144-3p as an oncomiRNA. The authors described that the overexpression of miR-144-3p promoted proliferation, migration, invasion, and chemoresistance in ccRCC cells. They also identified AT-rich interactive domain 1A (*ARID1A*) as a direct target gene of miR-144-3p in ccRCC. ARID1A is a transcription regulator that is a part of the SNF/SWI remodeling complex. The data to date highlight the lack of clarity concerning the role of miR-144 in ccRCC. Further in-depth studies are required.

miR-137 expression is reduced in RCC and miR-137 suppresses the activation of the PI3K/AKT signaling pathway. This leads to reduced proliferation, migration, and invasiveness of cells, enhanced apoptosis, and suppressed tumor growth in

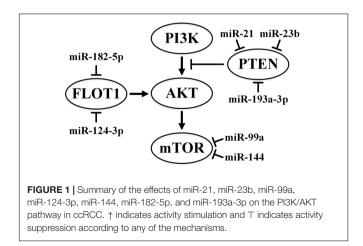
xenografts (Zhang and Li, 2016). The lncRNA *SNHG1* also negatively regulates this miRNA in RCC (Zhao S. et al., 2018).

The target of most oncogenic miRNAs in ccRCC is the tumor suppressor phosphate and tensin homolog (*PTEN*) gene, which is the negative regulator of the PI3K/AKT/mTOR pathway. In particular, *PTEN* directly interacts with miR-23b (Zaman et al., 2012), miR-193-3p (Liu L. et al., 2017), miR-21 (Cui et al., 2017), and miR-22 (Fan W. et al., 2016), although miR-22 acts as a tumor suppressor. Appropriate interactions that affect the expression of PTEN may be significant in the development of the disease. Aggressive ccRCC specimens are characterized by a reduced level of PTEN (Cancer Genome Atlas Research, 2013). There are some indications that miR-193a-3p and miR-224 can affect the PI3K/AKT pathway by targeting the ST3 beta-galactoside alpha-2,3-sialyltransferase 4 (*ST3GalIV*) gene in RCC (Pan et al., 2018a).

The influence of some miRNAs on the PI3K/AKT pathway is depicted in **Figure 1**.

It is believed that the effect of the reduced expression of miR-124-3p in ccRCC is due to an increase in the expression of its putative targets, *CAV1* and *FLOT1*, which are cytoplasmic membrane proteins associated with caveolae-mediated endocytosis (Butz et al., 2015). CAV1 expression causes the AKT-dependent formation of lamellipodia, which increases cell migration and invasion. In addition, it activates the Ras-extracellular signal-regulated kinase (Ras-ERK) pathway and suppresses anoikis, a form of programmed cell death specific to non-malignant cells induced by the detachment of anchorage-dependent cells. FLOT1 is also associated with endocytosis and activation of MAPK (Butz et al., 2015).

Studies have also shown the importance of the Wnt/ β -catenin signaling pathway in RCC and indicated that many WNT antagonist genes are inhibited by methylation of their promoters in ccRCC (Joosten et al., 2018). In ccRCC, miR-106b-5p can activate this pathway, having as its targets the following negative regulators of the Wnt/ β -catenin pathway: *LZTFL1*, *SFRP1*, and *DKK2* (Lu et al., 2017). Thus, being an oncogene, miR-106b-5p supports the aggressiveness and stemness phenotypes of ccRCC (Lu et al., 2017). miR-1260b also targets *DKK2* and stimulates the



proliferation and invasiveness of RCC cells, and its expression is enhanced in RCC (Hirata et al., 2013).

miR-203a targets the glycogen synthase kinase-3 β (*GSK3* β) gene (Hu et al., 2014), whose product is involved in β -catenin degradation (Joosten et al., 2018). Thus, miR-203a also acts as an oncogene in RCC, being a predictor of poor prognosis (Hu et al., 2014). IGF-II mRNA binding protein 1 (IGF2BP1), which participates in the Wnt/ β -catenin pathway, is a target of miR-372 in the RCC cell lines, thus miR-372 acts as a tumor suppressor (Huang et al., 2015).

The roles of miRNAs in important processes in ccRCC, which include cell migration, invasion, and proliferation, were highlighted in a recent review (He Y.H. et al., 2018).

Notably, mutations in known oncogenes and tumor suppressors, such as RAS, TP53, RB, and PTEN, are not typical of RCC (Joosten et al., 2018). However, sporadic mutations occur with high frequency in the genes encoding proteins associated with histone modification and chromatin remodeling. These genes include SETD2 (3-12%), KDM5C (3-8%), KDM6A (1%), BAP1 (8-15%), and PBRM1 (21-41%) (Cancer Genome Atlas Research, 2013). miRNA-mediated repression also contributes to the altered expression of these genes in RCC. For example, miR-106a-5p, which targets SETD2, functions as an oncogene in this case. SETD2 is instrumental in suppressing and halting the cell cycle at the transition from G0 to G1, and also inhibits proliferation and stimulates apoptosis in RCC (Xiang et al., 2015). SETD2 increases the binding of H3K36me3 to the p53 promoter that enhances the expression of p53. In ccRCC, miR-590-5p acts as an oncogene and its PBRM1 target acts as a tumor suppressor (Xiao et al., 2013).

Immune checkpoint molecules play an important role in the control of carcinogenesis. In ccRCC, the B7 homolog 3 (*B7-H3*) immune checkpoint molecules are validated targets of miR-187. This miRNA exhibits the characteristics of a tumor suppressor, and its concentration is reduced in both tumor tissue and plasma that provides opportunities for use in diagnosis of patients with ccRCC. A low level of miR-187 is associated with advanced stages of the disease and a worse 5-year survival rate. *In vitro*, it reduces cell proliferation and mobility and suppresses tumor growth (Zhao et al., 2013). Non-classical human leukocyte antigen G (HLA-G) inhibits the cytotoxic activity of T-lymphocytes and natural killer cells. *HLA-G* is often expressed in RCC and is a target of miR-148a and miR-133a, and overexpression of these miRNAs suppresses its effect (Jasinski-Bergner et al., 2015).

It is interesting to note the dual functions of some miRNAs in ccRCC. For instance, one of the targets of miR-106a-5p is the P21 (RAC1) Activated Kinase 5 (*PAK5*) gene. PAK5 is a serine/threonine kinase (activated by RAC) that regulates cytoskeleton dynamics, cell proliferation, and survival. miR-106a-5p is reportedly a tumor suppressor that reduces metastasis in xenografts and reduces migration and invasiveness of RCC cell lines (Pan et al., 2017). These data contradict the studies that reported oncogenic effects of miR-106a-5p, such as its ability to target *SETD2* and indirectly downregulate TP53 (Xiang et al., 2015) and VHL protein levels (Oliveira et al., 2017). The contradictory nature of the influence of miR-106a-5p on ccRCC is summarized in **Figure 2**.

Thus, some miRNAs (for example, miR-106a and miR-144) can show both tumor suppressor and oncogenic properties depending on the context, i.e., their effect is apparently dependent on the target gene and pathways involved. However, some contradictory data, for instance, for miR-99a (Cui et al., 2012; Oliveira et al., 2017), may be the result of inconvenient methodology or inadequate sampling of examined tumors.

The next few sections of this review will be devoted to discussing the regulation of angiogenesis and epithelial-mesenchymal transition (EMT), given the great importance of these processes in RCC.

MOLECULAR MECHANISMS IN HYPOXIA AND ANGIOGENESIS IN CCRCC: THE ROLE OF MIRNA

In most clear cell carcinomas of the kidney, the expression of the VHL gene is dysregulated in a similar manner (98% of cases feature gene deletion, with hypermethylation and point mutations in the remaining 2%). VHL binds HIF1A and degrades it only in the presence of oxygen. In the absence of oxygen, HIF1A dimerizes with HIF1B and leads to the activation of transcription of more than 40 genes, including the vascular endothelial growth factor (VEGF) family and erythropoietin (EPO) genes, genes encoding growth factors (e.g., EGF, PDGF) and glycolytic enzymes, and genes involved in glucose metabolism (e.g., GLUT1/4) (Roskoski, 2017). In the presence of VHL mutations downregulating its expression, the aforementioned regulatory influences of VHL on HIF1 are disrupted. Inactivation by oxygen is also a characteristic of HIF2A (Serocki et al., 2018). The VEGF-A protein of the VEGF family stimulates the dimerization and activation of the VEGF receptor 2 (VEGFR2). This interaction regulates, in particular, the angiogenesis process, ensuring the supply of oxygen and nutrients to the tumor and encouraging metastasis. The activation of VEGFR2, in turn, leads to the activations of the ERK1/2, AKT, Src, and p38

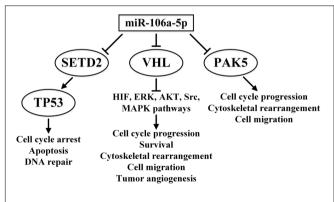


FIGURE 2 | Multiple features of the effects of miR-106a-5p in ccRCC signaling pathways. The overwhelming effect of miR-106a-5p on tumor suppressors TP53 and VHL and oncogene PAK5 is presented. ↑ indicates activity stimulation and T indicates activity suppression according to any of the mechanisms.

MAPK pathways, which are important for tumor development and metastasis. These activations occur through a cascade of phosphorylation events involving the tyrosine moiety of various proteins, which further recruit phosphotyrosine-binding proteins. These pathways are associated with key angiogenesis events that include cell proliferation, migration, survival, and vascular permeability (Roskoski, 2017). In patients with ccRCC being treated with VEGF and mTOR blockers, angiogenesis can be stimulated by angiopoietin 2, c-MET, or interleukin (Malouf et al., 2016).

While HIF1 responds to acute hypoxia, the adaptation to chronic hypoxia is mediated by the expression of HIF2 and HIF3 in endothelial tissues, while the expression of HIF1 is downregulated. miRNAs play a significant role in this adaptation. While there is a significant overlap of HIF1 and HIF2 targets, the role of HIF3 is much less clear. It has at least six isoforms. One suppresses the activity of HIF1 and HIF2, while others may activate the transcription of a number of genes (Serocki et al., 2018).

In response to prolonged hypoxia, 786-O, a VHL-defective cell line of ccRCC, displays upregulation of the expression of *L1CAM* and *FBN1* genes. In addition to the downregulated expression of miR-100 and miR-378, 786-O hypoxic cells have downregulated expression levels of *AUTS2*, *MAPT*, *AGT*, and *USH1C* genes. Expression of miR-100 and miR-378 was also found to be significantly reduced in bone metastases of ccRCC (Chen S.C. et al., 2017).

In ccRCC, the response to hypoxia and metastasis is also modulated by the androgen receptor (AR). AR increases the expression of miR-185-5p, which targets *VEGF-C* (a gene encoding lymphangiogenetic factor). At the same time, binding of miR-185-5p to the promoter region of *HIF2A* mRNA increases HIF2A expression and the subsequent expression of *VEGFA* (an angiogenetic factor). This explains why AR-positive ccRCC metastasizes to the lung rather more often than to lymph nodes (Huang et al., 2017).

In RCC patients, especially those diagnosed with ccRCC and VHL disorders caused by familial mutations, disruption of signaling pathways inhibited by tyrosine kinase inhibitors and other related inhibitory drugs is most effective (Mickley et al., 2015). The possibility of using drugs based on miRNA mimics and anti-miR is also being considered for these purposes (Schanza et al., 2017; Serocki et al., 2018).

As mentioned before, miRNAs contribute significantly to regulation of the main hypoxia response pathways. At least 40 miRNAs affect the expression of HIF alone in various tissues, including miR-687 in embryonic renal tissue and miR-429 and miR-19a in endothelial cells (Serocki et al., 2018). Upregulation of miR-106a and miR-106b and downregulation of their target gene *VHL* has been described for ccRCC (Oliveira et al., 2017).

VEGFR2 is the putative target of miR-221 in metastatic RCC (Khella H.W.Z. et al., 2015). Accordingly, poor survival outcome in patients treated with the receptor tyrosine kinase inhibitor sunitinib correlates with a high level of miR-221 accompanied by low levels of VEGFR2. The effect of miR-221 is different for endothelial and tumor cells; it lowers proliferation and angiogenesis in human umbilical vein endothelial cells

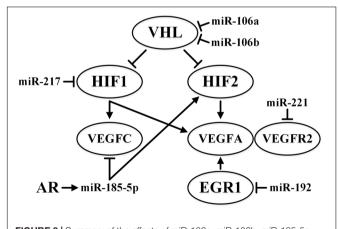
(HUVECs) but increases the proliferation of ACHN kidney cancer cells (Khella H.W.Z. et al., 2015).

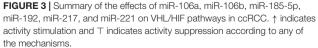
Responses to hypoxia are also influenced by lncRNA. Antisense transcripts of the 3' and 5' non-coding regions of *HIF1A* are 3'aHIF-1 α and 5'aHIF-1 α , respectively. In human lung epithelial cells, the expression of these lncRNAs is induced by hypoxia, and they can suppress the expression of *HIF1A* (Uchida et al., 2004). In addition, increased expression of the HOX transcript antisense intergenic RNA (HOTAIR) lncRNA is observed in RCC, and it correlates with the tumor progression. It has a direct inhibitory effect on miR-217, the target of which is *HIF1A*. Suppression of miR-217 reduces apoptosis, increases proliferation and migration of tumor cells, and stimulates EMT in RCC (Hong et al., 2017).

In RCC and ovarian cancer, miR-192 has anti-angiogenic properties due to the effect on its *EGR1* and *HOXB9* targets (Wu et al., 2016). In lung cancer, EGR1 binds to the *VEGFA* proximal promoter, activating its expression (Shimoyamada et al., 2010). The effect of HOXB9 on vascularization in breast cancer has been documented (Seki et al., 2012). Moreover, low levels of miR-192 in various malignant tumors are a predictor of poor prognosis (Wu et al., 2016).

The influence of some miRNAs on the VHL/HIF pathway is summarized in **Figure 3**.

In turn, the expression of certain miRNAs is affected by the proteins of the VHL-HIF-VEGF pathway (for example, Schanza et al., 2017). Thus, knockdown or genetic elimination of the *VHL* gene in RCC cells leads to an increase in miR-28-3p expression. The target of this miRNA is the gene for mitotic spindle checkpoint protein (Mad2), which blocks cells with unformed spindles from entering mitosis. Dysregulation of Mad2 expression leads to chromosomal instability (Hell et al., 2014). The expression of miR-204 in ccRCC cells is, on the contrary, induced by VHL. The target of miR-204 is *MAP1LC3B* (*LC3B*), which is associated with macro-autophagy (Mikhaylova et al., 2012). LC3B-mediated macro-autophagy is necessary for nutrient supply to the tumor during the period of metabolic





stress and is important for cell survival, tumor growth, and increased aggressiveness (Sato et al., 2007). At the same time, hypoxia in RCC suppresses miR-30c under the influence of HIF (Huang J. et al., 2013).

As a result of the interaction of different proteins of the VHL/HIF/VEGF pathway, along with the participation of miRNAs, positive and negative feedback loops may appear that prolongs the initial effect of hypoxia.

In mice, renal ischemia/reperfusion procedure was reported to reduce the concentration of 40 miRNAs by almost half, while the concentration of 36 other miRNAs more than doubled (Liu et al., 2012). Quantitative RT-PCR performed 4-12 h after the procedure confirmed an increase in miR-210 concentration. In HUVEC-12 cells, angiogenesis is stimulated by increases in the expression of VEGF and VEGFR2, which are triggered by an increased miR-210 concentration (Liu et al., 2012). Since the target of miR-210 in chondrocytes and hepatocellular carcinoma cells is HIF3A, it is highly likely that this miRNA may regulate the switch between acute hypoxia and chronic hypoxia responses in these cells (Serocki et al., 2018). HIF1A may regulate miR-210 and has been shown to enhance its expression in RCC (Nakada et al., 2011). HIF1A-mediated enhancement of miR-210 expression was also shown in breast cancer, while HIF2A showed no regulatory effect (Camps et al., 2008). However, in 786-O RCC cells, HIF2A also influenced miR-210 expression (Zhang Z. et al., 2009). Increased expression of miR-210 is most typical for ccRCC (Valera et al., 2011).

A feedback loop has been constructed for the cardiomyocytes. HIF1 increases transcription of the MIR-21 gene, and miR-21, in turn, induces the expression of HIF1A via the regulation of PTEN/AKT signaling pathway (Liu et al., 2014). The expression of miR-21 is also increased in oral squamous cell carcinoma (OSCC). It is highly concentrated in the exosomes of OSCC cells and enhances the migratory and invasive properties of tumor cells in a mechanism that is dependent on HIF1A and HIF2A (Li et al., 2016).

During hypoxia, exosomes containing miR-135b are formed in multiple myeloma cells. Suppression of the HIF1AN by miR-135b in HUVEC cells leads to increased HIF1 activity and angiogenesis (Umezu et al., 2014). This mechanism remains turned on even after the restoration of the normal supply of oxygen, which prolongs the antihypoxic response.

Enhanced miR-18a expression after 24 h of hypoxia was observed in human choroidal endothelial cells (Han et al., 2015). Since HIF1A mRNA is the direct target of this miRNA, this interaction may act as the switch between the responses to acute hypoxia and chronic hypoxia (Han et al., 2015).

Hypoxia suppresses the expression of DICER, which is associated with miRNA processing. However, it is not understood how this does not lead to the total disruption of miRNA activity (Serocki et al., 2018). Some miRNAs, such as the tumor suppressor miR-182-5p, are more sensitive to hypoxic suppression of DICER expression than others. In addition to the aforementioned effect on the AKT pathway, miR-182-5p acts as an inhibitor of its target HIF2A, even though it is primarily manifest in VHL-deficient ccRCC cells (Fan Y. et al., 2016). The miR-200 family is another example of miRNAs that are sensitive to disruption of DICER expression. DICER is the direct target of miR-122, a miRNA that is highly expressed in ccRCC, while the loss of DICER decreases the expression of the miR-200 family, leading to EMT (Fan Y. et al., 2018).

Most tumor cells respond to hypoxia by shifting their energy metabolism to glycolysis (the Warburg Effect). Many miRNAs participate in this shift, which is a characteristic of many solid tumors (Morais et al., 2017). In particular, in RCC, miR-1291 directly inhibits the expression of the *GLUT1* glucose transporter gene, and its levels in tumor tissue are reduced as compared to the levels in normal tissue (Yamasaki et al., 2013). In ccRCC, hypoxia-induced miR-101 expression can be a mechanism for activating glycolysis, as miR-101 inhibits the *TIGAR* (TP53-Induced Glycolysis and Apoptosis Regulator) gene, which is a member of the p53 pathway that is involved in mediating the exchange to the pentose phosphate pathway (Xu et al., 2017). However, TIGAR-mediated regulation of glycolysis has an anti-apoptotic effect, protecting against DNA damage.

lncRNAs also affect the transition to glycolysis (Serocki et al., 2018). In cell cultures obtained from solid tumors, HIF1A-inducible long intergenic non-coding (linc) RNA-p21 was shown to bind to HIF1A, disrupting its interaction with VHL and attenuating the ubiquitination of HIF1A. As a result, both HIF1A and lincRNA-p21 accumulate that causes a transition to glycolysis (Yang et al., 2014).

METASTASIS AND EMT IN KIDNEY CANCER: THE ROLE OF MIRNA

The incidence of metastasis is high in ccRCC (25–30%) and increases to over 50% after surgery (Rydzanicz et al., 2013). Metastatic ccRCC is extremely resistant to therapy, with responses to chemo-, radio-, and immunotherapy observed in no more than 10% of patients, with 5-year survival rates as low as 9% in the presence of distant metastases (Randall et al., 2014).

Metastasis of primary tumors occurs in several stages – from changes in biochemistry, morphology, and migration abilities of tumor cells to the appearance of surface receptors that provide directed migration to target organs, followed by the formation of specific microenvironment in the target organ, into which metastatic cells can enter and survive.

Epithelial-mesenchymal transition is an important stage in metastasis, during which changes occur in the properties of cancer cells. The changes contribute to metastasis. The process of EMT is characterized by a loss of cell polarity and intercellular bonds, as well as an increase in the migration activity of cells and their invasive properties. A critical event in EMT is the loss of E-cadherin and an increase in the activity of its transcriptional repressors, which include *ZEB1*, *ZEB2*, *TWIST*, *SNAIL*, and *SLUG* (Piva et al., 2016). EMT causes the development of sarcomatoid ccRCC (the most unfavorable form of cancer) due to the switched expression of E-cadherin to N-cadherin, dissociation of β -catenin from the membrane, and increased expression of *SNAIL* and *SPARC* (Conant et al., 2011).

An evaluation of the expression of 11 EMT markers in nephrectomized samples of RCC revealed that E-cadherin, clusterin, TWIST, and vimentin levels were significant predictors of recurrence (Harada et al., 2012).

Wilms' tumor, WT1, causes mesenchymal-to-epithelial transition (MET) in kidneys during their development. During development of other organs, such as the heart, this gene causes EMT. WT1 is absent in normal renal tissue but is expressed in ccRCC due to the decreased expression of VHL or expression of mutated VHL. WT1 expression in ccRCC in the presence of SNAIL and E-cadherin starts a "hybrid" process in which EMT and MET traits are simultaneously expressed (Sampson et al., 2014). The loss of VHL activity also stimulates EMT, thereby increasing the expression of HIF1A, which subsequently activates nuclear factor-kappa B (NF- κ B). The cytoplasmic expression of NF-kB has been correlated with the invasiveness of ccRCC (Kankaya et al., 2015).

Chronic oxidative stress-stimulated malignant transformation of kidney cells is accompanied by their acquisition of stem characteristics and EMT (Mahalingaiah et al., 2015). This leads to a significant increase in the expression of genes such as *BCL2, CCND1* (CyclinD1), *BIRC5* (Survivin), and *PCNA*, as well as *VIM* (Vimentin), *ACTA2* (α -Smooth muscle actin, α -SMA) and *SNAIL1*, as well as a significant decrease in the expression of *CDH1* (E-cadherin), cytokeratin, and *CTNNB1* (β -catenin). The suppression of *OCT4* and *SNAIL* with small interfering RNA (siRNA) can partially reverse these changes (Mahalingaiah et al., 2015). The effect of hypoxia on EMT can also be partially achieved by reducing the expression of miR-30c and increasing the expression of its target, *SLUG*, which also leads to a decrease in E-cadherin expression and stimulation of cell migration in RCC (Huang J. et al., 2013).

Downregulation of *FOXO3a* in ccRCC also leads to an increase in expression of *SNAIL* that stimulates EMT (Ni et al., 2014). In ccRCC, *FOXO3a* is a target of oncogenic miR-155, which increases proliferation, colony formation, migration, and invasion (Ji et al., 2017). *SNAIL1* in ccRCC is the target of miR-30e-3p (minor variant), which inhibits invasion and migration of tumor cells (Wang D. et al., 2017).

The level of miR-720 is elevated in RCC, while its downregulation suppresses tumor growth (Bhat et al., 2017). miR-720 in RCC targets *CTNNA1* (α E-catenin) and *CDH1* (E-cadherin), thereby stimulating cell migration and invasiveness.

ZEB2 suppresses the expression of E-cadherin. In RCC, ZEB2 enhances the migratory and invasive ability of cells and the expression of ZEB2 correlates with more advanced forms of the disease and worse survival rates (see the review by Piva et al., 2016). One of the targets of miR-141 (a member of the miR-200 family) in RCC is *ZEB2* and thus it can suppress EMT (Li W. et al., 2014). Reduced expression of miR-141 has been correlated with a poor response to sunitinib (Berkers et al., 2013). In addition, in RCC, miR-141 binds to the lncRNA HOTAIR causing its cleavage by the Argonaute 2 (Ago2) complex (Chiyomaru et al., 2014). As already mentioned, HOTAIR is also capable of stimulating EMT (Hong et al., 2017). The high expression of miR-122 in ccRCC stimulates EMT as a result of the targeting of *DICER* and *OCLN* (occludin), which plays an important role in tight junctions, which are characteristic of renal tissue. The loss of tight junctions leads to an increase in cell mobility (Jingushi et al., 2017). Downregulated expression of DICER results in the suppression of the maturation of the miR-200 family.

In RCC, three miRNAs – miR-192 (which has anti-angiogenic effects), miR-194, and miR-215 – target *ZEB2*, *MDM2*, and *TYMS* oncogenes (Khella H.W. et al., 2013). miR-30a is specifically downregulated in metastatic tumors compared to its expression in primary tumors (Butz et al., 2015). Other studies on ccRCC have implicated the *ZEB2* and glucose-regulated protein 78 (*GRP78*) genes as targets of miR-30a (Chen Z. et al., 2017; Wang C. et al., 2017).

Expression of the transcription factor SOX4, which is normally involved in embryogenesis, is limited to a small number of cell types in which the mature cells retain stem cell characteristics. Due to the influence of SOX4 on the Wnt/β-catenin, Notch1, and p53 pathways, as well as the components of the miRNA processing machinery (DICER, Argonaute 1, RNA helicase A), upregulation of the expression of SOX4 can cause the development of different types of cancer and stimulate EMT. In contrast, the high expression of SOX4 may be associated with a more favorable disease course in other types of cancer. In different cancer types, many miRNAs target SOX4. They usually act in an anti-oncogenic manner and are characterized by reduced expression in ccRCC (Geethadevi et al., 2018). In ccRCC, miR-138 has a tumor suppressor role via the targeting of SOX4. The effects include the inhibited proliferation, migration, and invasion of tumor cells, increased expression of E-cadherin, and decreased expression of vimentin (Liu F. et al., 2017).

In ccRCC, miR-210 targets *TWIST1*. When the expression of this miRNA is turned off, the cancer cells begin to display the EMT morphology and increased tumor growth is observed in xenografts. High TWIST1 expression along with low expression of miR-210 is associated with a low survival rate in ccRCC. The aforementioned association of miR-210 with a response to hypoxia and its angiogenic behavior is important. It cannot be ruled out that miR-210 may show varied roles at different stages in the development of ccRCC (Yoshino et al., 2017).

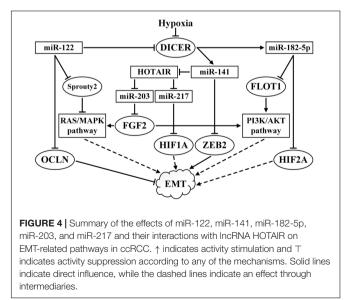
The reduced expression of miR-203 in ccRCC primary tumors correlates with poor prognosis and metastasis, while increased expression of miR-203 suppresses RCC cell lines growth and ccRCC metastasis (Xu M. et al., 2015). The authors attributed this to the action of miR-203 on fibroblast growth factor 2 (FGF2) as a target. The FGF2 protein is able to interact with the dimer of growth factor receptor-bound protein 2 (GRB2) involved in the PI3K/AKT and Ras/MAPK signaling pathways. Moreover, FGF2 is involved in the fine regulation of the Ras/MAPK pathway via negative feedback. FGF2 is overexpressed in many cancers (Ornitz and Itoh, 2015). The interaction between lncRNA HOTAIR and miR-203 leads to the suppression of miR-203 and stimulates EMT; when this interaction is eliminated, the expression of E-cadherin, claudin, PTEN, p21, and p27 increase, while vimentin expression is reduced (Hong et al., 2017). Methylation of MIR-203 is a specific characteristic of metastatic forms of ccRCC (see below and Pereyaslova et al., 2016). The influence of some miRNAs and lncRNA HOTAIR on EMT-related pathways is summarized in **Figure 4**.

The role of the cell environment, in particular the intercellular matrix, is important in metastasis. *CD44* is one of the important targets of miR-34a in the RCC lines. The CD44 protein is a cell surface glycoprotein that is involved in the intercellular interaction, cell adhesion, and migration. CD44 is a receptor for hyaluronic acid, osteopontin, collagen, and it can interact with matrix metalloproteinases. Ectopic expression of miR-34a stops cellular growth, migration, and invasion, and also significantly inhibits the growth of carcinoma in xenografts and metastasis in nude mice (Yu et al., 2014).

ADAM metallopeptidase domain 17 (*ADAM17*, also known as disintegrin and A metalloproteinase 17) is a target of miR-145 in RCC. The level of ADAM17 is significantly increased in RCC, whereas the level of miR-145 is reduced. ADAM17 stimulates cell proliferation and migration, and also negatively regulates miR-145 through tumor necrosis factor-alpha (TNF α). The result is the formation of a positive feedback loop. In healthy kidneys, this metalloproteinase is found in various types of cells capable of giving rise to RCC (Doberstein et al., 2013).

The expression of miR-186 is greatly reduced in RCC. This miRNA suppresses cell growth, colony formation, invasion, and arrests the cell cycle at the G0/G1 stage. It is believed that the effects of miR-186 are due to the action on *SENP1* (sentrin/SUMO-specific protease 1) as a target (Jiao et al., 2018). The SENP1 protein is the regulator of the SUMO pathway, which triggers the detachment of SUMO from homeodomain interacting protein kinase 2 (HIPK2, a transcriptional regulator), histone deacetylase 1 (HDAC1), and metastasis associated 1 (MTA1). The influence of the mimic miR-186 on *SENP1* can suppress the pathway of NF-kB, which reduces the expression of p-IkBa and p-p65, as well as the underlying cyclin D1 and MMP9, and increases the expression of p21 and BCL-2 (Jiao et al., 2018).

Reduced expressions of miR-149-5p and miR-149-3p are correlated in ccRCC with increased tumor aggressiveness and



metastases (Okato et al., 2017). The authors attributed this to the fact that the target of both miRNAs is forkhead box protein M1 (*FOXM1*), a transcriptional activator that regulates the expression of cyclins, such as cyclin B1 and cyclin D1.

ABERRANTLY METHYLATED MIRNA GENES IN CCRCC

One of the critical means of regulating the expression of miRNA genes is the methylation of the CpG island that is adjacent to or overlaps with the miRNA gene (Han et al., 2007; Lopez-Serra and Esteller, 2012). It is assumed that the percentage of miRNA genes that are aberrantly methylated in tumors is several times higher than the genes encoding proteins, which increases their prospects as biomarkers (Kunej et al., 2011; Baylin and Jones, 2016; Piletic and Kunej, 2016). Methylation profiles of miRNA genes have been constructed for epithelial tumors that have different localizations, including the colon, lung, breast, prostate, and ovaries. These are being considered as new potential markers and marker systems for the diagnosis and prognosis of these malignant cancers (Pronina et al., 2017b; Torres-Ferreira et al., 2017; Heller et al., 2018; Loginov et al., 2018a,b; Shi et al., 2018; Zare et al., 2018).

In contrast to the most common and widely studied types of cancer, such as colon, lung, breast, and prostate cancer, less is known about the methylation of the miRNA genes in ccRCC. What is known mainly concerns genes of the miR-9 and miR-34 families (Hildebrandt et al., 2010; Vogt et al., 2011; Schiffgen et al., 2013). These studies have shown that the promoter of the gene coding for miR-34a is significantly methylated in RCC cell lines and that the expression of this miRNA is downregulated. Treatment with 5-aza-2'-deoxycytidine increases the expression of miR-34a and reduces the expression of its target (Yu et al., 2014).

The last 5 years have seen a significant increase in the understanding of the role of hypermethylation of miRNA genes in the pathogenesis of ccRCC. In ccRCC, miR-10b is characterized by a highly methylated promoter and significantly reduced expression (He C. et al., 2015). Transfection by lentivirus carrying this miRNA or treatment with demethylating agents inhibits cell proliferation, migration, and invasiveness in cell culture. However, the target genes that are significant in RCC are unknown (He C. et al., 2015). In stomach cancer, miR-10b acts as a tumor suppressor and targets microtubule-associated protein RP/EB family member 1 (*MAPRE1*) (Kim et al., 2011). In non-small-cell lung cancer and bladder cancer, it acts as an oncogene by enhancing cellular migration and invasion. The putative targets associated with this effect are *KLF4* and *HOXD10* (Xiao et al., 2014).

A high miR-21/miR-10b ratio in metastatic ccRCC correlates with a poor prognosis (Fritz et al., 2014). Interestingly, an increased level of MIR-21 expression that correlates with a poor prognosis is associated with hypomethylation of its promoter (Cancer Genome Atlas Research, 2013).

The aforementioned miR-182-5p tumor suppressor miRNA can also be regulated by methylation. An upstream CpG

island (8–10 kb) contains a putative transcription start site. Its methylation level is significantly increased in RCC cell cultures and is slightly elevated in ccRCC samples compared to the normal tissue, which may explain the decreased expression of this miRNA. One of the targets of miR-182-5p is *MALAT-1*. This gene produces a precursor transcript from which a ncRNA is derived by RNase P cleavage of a tRNA-like small ncRNA (known as mascRNA) from its 3' end. Its ribonucleoprotein complexes may act as a transcriptional regulator for numerous genes, including some genes involved in cancer metastasis and cell migration. Downregulation of *MALAT-1* leads to upregulation of p53 and downregulation of CDC20 and AURKA, which are the drivers of the cell cycle mitotic phase (Kulkarni et al., 2018).

The gene encoding miR-766-3p is highly methylated in RCC tissues compared to the normal tissues. This miRNA behaves like a suppressor. Its direct target is SF2, whose repression also reduces the expression of AKT and ERK (Chen C. et al., 2017). The promoter of miR-145, which targets the metallopeptidase gene ADAM17, is also strongly methylated in RCC. Treatment with 5-aza-2'-deoxycytidine increases the expression of miR-145 and reduces the expression of ADAM17 (Doberstein et al., 2013). The promoter region of the MIR-200c gene is hypermethylated in RCC cell lines, which correlates with decreased expression, and it is not methylated in the normal renal cell line (Gao et al., 2014). The MIR-492 gene has a strongly methylated promoter in ccRCC and reduced expression. The use of 5-aza-2'-deoxycytidine or the histone deacetylase inhibitor 4-phenylbutyric acid increases the expression of miR-492 in ccRCC cell cultures, inhibits cell proliferation, and stimulates apoptosis and adhesion (Wu et al., 2015). The upstream promoter of MIR-106a gene is hypermethylated in RCC, which might be responsible for its downregulation (Pan et al., 2017).

Over the past 5 years, we have systematically analyzed the methylation of up to 20 miRNA genes in various cancers, including lung, breast, ovarian, and kidney cancer. These analyses have identified new markers and marker systems that are useful for the diagnosis and prognosis of these malignant diseases (Beresneva et al., 2013; Rykov et al., 2013; Pronina et al., 2017b; Braga et al., 2018; Loginov et al., 2018a,b; Varachev et al., 2018). Moreover, we have shown a correlation between the levels of expression and methylation, which has confirmed the functional role of methylation of a group of miRNA genes in the pathogenesis of breast, ovarian, and kidney cancer (Pronina et al., 2017b; Loginov et al., 2018a,b; Varachev et al., 2018).

We first found that the frequency of methylation of six miRNA genes (MIR-124-2, MIR-124-3, MIR-9-1, MIR-9-3, MIR-34b/c, and MIR-129-2) was significantly higher in malignant tumors of patients with ccRCC than in normal kidney tissues (Beresneva et al., 2013). Subsequent studies by our group established that 16 miRNA genes (MIR-124-1/-2/-3, -125b-1, MIR-129-2, MIR-132, MIR-137, MIR-193a, MIR-34b/c, MIR-375, MIR-203, MIR-9-1, MIR-9-3, MIR-107, MIR-130b, and MIR-1258) were hypermethylated and two miRNA genes (MIR-191, MIR-212) were hypomethylated in ccRCC (Pereyaslova et al., 2016; Loginov et al., 2017; Varachev et al., 2018). Moreover, seven miRNA genes (MIR-124-3, MIR-125b-1, MIR-129-2, MIR-137, MIR-34b/c, MIR-375, MIR-9-3) were downregulated and correlated with altered methylation. The findings provided evidence of the functional significance of methylation in the deregulation of miRNA genes and in the pathogenesis of ccRCC (Varachev et al., 2018).

Besides hypermethylation, some miRNA genes studied (MIR-124-2/-3, -34b/c, MIR-129-2, MIR-107, MIR-148a, MIR-203) have been associated with aspects of ccRCC progression that include advanced pathologic stage, tumor size, and differentiation grade. In particular, hypermethylation of six miRNA genes (MIR-125b-1, MIR-129-2, MIR-203, MIR-375, MIR-107, and MIR-1258) significantly correlated with the presence of metastasis (Pereyaslova et al., 2016; Varachev et al., 2018), and most significantly for MIR-375 and MIR-1258 (*p*-value $\sim 10^{-7}$ - 10^{-8}).

On the basis of hypermethylated miRNA genes, novel marker systems have been suggested for ccRCC diagnosis (MIR-125b-1, MIR-375, MIR-137, MIR-193a) and prediction of metastasis (MIR-125b-1, MIR-375, MIR-107, MIR-1258, MIR-203) (Loginov et al., 2017; Varachev et al., 2018).

A Russian team of researchers demonstrated the correlation of the methylation of the MIR-129-2 gene with increased expression of genes, such as *RARB(2)*, *RHOA*, *NKIRAS1*, and *CHL1* (Pronina et al., 2017a). Preliminary data from another study indicated a negative correlation between the expression levels of miR-375 and the pro-apoptotic gene *APAF1* (Varachev et al., 2018). The targets of most hypermethylated miRNAs, identified by us recently remain unclear and further research is needed.

However, the roles of some of the novel hypermethylated miRNAs in the pathogenesis of kidney cancer have been studied. For instance, the reduced expression of miR-124 is considered a highly significant factor in ccRCC genesis and may be an essential predictor of survival (Butz et al., 2015). Moreover, the miRNA and gene expression data from 458 ccRCC and 254 normal kidney specimens were used to construct the integrated miRNA-target interaction network, which revealed miR-124 as a key miRNA contributing to the aggressive behavior of ccRCC (Butz et al., 2015).

Table 1 presents examples of miRNAs encoded by genes that are hypermethylated and downregulated in ccRCC. Data on target genes and functions in RCC are also provided. The information presented pertains mainly to those genes whose role of hypermethylation was first explored by our group and data from other authors concerning the functional roles, target genes, and in some cases the signaling pathways involved.

For five miRNAs we identified (miR-124-3p, miR-129-3p, miR-137, miR-203, and miR-375), functional studies by other authors confirmed suppressor properties and/or association with metastasis. The tumor suppressor and anti-metastatic functions of the hypermethylated MIR-129-2 gene we discovered (Beresneva et al., 2013), are consistent with the data that miR-129-3p downregulates multiple metastasis-related genes in RCC cells, including *SOX4*, and also decreased phosphorylation of focal adhesion kinase and expression of *MMP-2/9* (Chen et al., 2014). In addition, miR-129-3p is involved in miR-129-3p/TRPM7/AKT/FOXO1 signaling

pathway (Zhao Z. et al., 2018). The collective data confirm the ability of miR-129-3p to restrain metastasis in ccRCC.

In ccRCC, the hypermethylated MIR-137 gene (Varachev et al., 2018) abrogates the tumor-promoting effect of overexpressed ncRNA Small Nucleolar RNA Host Gene 1 (SNHG1), which is associated with metastasis and poor prognosis of ccRCC patients (Zhao S. et al., 2018).

Data on the nature of the role of miR-203 in RCC, either as a tumor suppressor or an oncogene, is inconsistent (Hu et al., 2014). However, in other cancer types (hepatocellular carcinoma, multiple myeloma, lung cancer, esophageal cancer, bladder cancer), miR-203 acts as a suppressor (Hu et al., 2014). It is possible that this miRNA has different roles at different stages of the development of ccRCC and/or in its different variants. Overall, we observed significantly more frequent hypermethylation of the MIR-203 gene in tumors of patients with ccRCC with metastases. Moreover, our data on the anti-metastatic activity of miR-203 are consistent with data from three other papers (see **Table 1**), including the results on the role of miRNA-203 in inhibition of EMT and metastatic genes (Dasgupta et al., 2018).

Our data concerning the correlation of miR-375 hypermethylation with metastasis strengthens the evidence that the suppression of the tumor aggressive phenotypes of ccRCC mediated by this miRNA regulates the oncogene *YWHAZ*, which is its direct target (Zhang X. et al., 2018). As discussed earlier, aberrantly methylated miR-148a was suggested to function as a tumor suppressor in RCC by targeting *AKT2* (Cao et al., 2017). A recent paper (Reustle et al., 2018) implicated the role of aberrantly methylated miR-212-3p and miR-132-3p in ccRCC in the post-transcriptional regulation of *BCRP/ABCG2* (breast cancer resistance protein), which contributes to the multi-drug resistance seen in cancer. Hypermethylated miR-193a-3p can affect the PI3K/AKT pathway in RCC (Pan et al., 2018a).

However, our data on the frequent hypermethylation of the MIR-125b-1 gene and the hypermethylation linkages with progression and metastasis (Varachev et al., 2018) do not agree with the increased level of this miRNA and its role in promoting cell (Jin et al., 2017). The properties of this miRNA are also ambiguous in other types of cancer, such as breast cancer (Loginov et al., 2018a). Additional studies are required to determine the role and mechanism of miR-125b in RCC.

Tumor suppressor or/and anti-metastatic properties were found for some miRNAs, encoded by 16 aberrantly methylated genes that discovered by us, including miR-124, miR-129, miR-132, miR-137, miR-148a, miR-203, and miR-375, as well as for miR-9 and miR-34 genes. However, further analyses should explore the expression and functional properties of some other miRNAs, which genes were also found as hypermethylated (miR-107, miR-125b, miIR-1258, miR-130b, and miR-193a) or showed hypomethylation (miR-191, miR-212) in cancers of the kidney. Overall, our group has provided novel data on the contribution of methylation to the regulation of more than 10 miRNA genes in ccRCC (Beresneva et al., 2013; Loginov et al., 2017; Varachev et al., 2018). Further research and validation of target genes for these aberrantly methylated miRNAs are required. Moreover, hypermethylated

| miRNA | miRNA gene hypermethylation and downregulation | Target genes of miRNA and some pathways | Functions of miRNA in ccRCC | References |
|------------|--|--|---|---|
| miR-124-3p | Hypermethylation of the <i>miR-124-2</i> gene correlates with decrease of miR-124-3p levels in ccRCC tissues and cell lines | <i>CAV1, FLOT, FZD5</i> (miR-124-3p/FZD5/PKC pathway), IncRNA HOTAIR, <i>ST8SIA4</i> (HOTAIR/miR-124/ST8SIA4 pathway) | Hypermethylation of <i>miR-124-2</i> is strongly associated with advanced pathologic stage and grade of differentiation. Restoration miR-124-3p reduces migration, invasion, proliferation, metastasis; decreased cell cycle S phase; increases patient survival, drug resistance. By sponging miR-124 HOTAIR as ceRNA upregulated ST8SIA4 and promoted the proliferation and metastasis in RCC | Butz et al., 2015; Long et al., 2015; Pan et al., 2018b; Varachev et al., 2018 |
| miR-129-3p | Hypermethylation of the <i>miR-129-2</i> gene correlates with decrease of miR-129-3p levels in ccRCC tissues and cell lines | SOX-4, MMP-2/9, TRPM7 (miR-129- 3p/TRPM7/AKT/FOXO1 pathway) | Hypermethylation of <i>miR-129-2</i> is associated with advanced pathologic stage and metastasis in ccRCC; miR-129-3p inhibits migration, invasion, metastasis | Beresneva et al., 2013; Chen et al., 2014; Zhao Z. et al., 2018 |
| miR-137 | Downregulation of miR-137 in RCC tissues and cell lines correlates with hypermethylation of the <i>miR-137</i> gene | IncRNA SNHG1, oncogene RLIP76 | Decrease proliferation, migration, invasion, metastasis; promotes apoptosis; inhibits PI3K/PKB pathway; tumor suppressor function confirmed <i>in vitro</i> and <i>in vivo</i> | Zhang and Li, 2016; Varachev et al., 2018; Wang M. et al., 2018; Zhao S. et al., 2018 |
| miR-182-5p | Downregulation correlates with hypermethylation in RCC tissues and cell lines | HIF-2α, IncRNA MALAT-1, FLOT1 | Oncosuppressor; inhibited tumorigenicity <i>in vitro</i> and <i>in vivo</i> and induced mitotic cell cycle arrest and apoptosis | Xu et al., 2014; Fan Y. et al., 2016; Fang et al., 2018; Kulkarni et al., 2018 |
| miR-203 | Downregulation of miR-203 in RCC cell lines and ccRCC specimens; methylation of the <i>miR-203</i> gene is significantly increased only in metastatic tumors | <i>FGF2</i> , IncRNA SNHG14, N-WASP (SNHG14/miR-203/N-WASP pathway), IncRNA HOTAIR | <i>miR-203</i> methylation and low expression are associated with metastasis in ccRCC. SNHG14 sponging miR-203 and elevating N-WASP as ceRNA promotes ccRCC migration and invasion. Overexpression of miR-203 inhibited migration, invasion, and induced apoptosis and cell cycle arrest. miR-203-HOTAIR interaction inhibited EMT and metastatic genes | Xu M. et al., 2015; Liu G. et al., 2017; Dasgupta et al., 2018; Varachev et al., 2018 |
| miR-375 | Downregulation in RCC cell lines and ccRCC specimens; the <i>miR-375</i> gene is hypermethylated in ccRCC tissues | Oncogene YWHAZ | <i>miR-375</i> methylation and low expression are associated with metastasis ccRCC; according to functional studies miR-375 suppressed ccRCC cell proliferation, migration, and invasion | Varachev et al., 2018; Zhang X et al., 2018 |
| miR-766-3p | Downregulation caused by hypermethylation in RCC tissues | SF2 (miR-766-3p/SF2/P- AKT/P-ERK pathway) | Downregulation is associated with clinical stage and worse prognosis; upregulation attenuates cell cycle progression | Chen C. et al., 2017 |

TABLE 1 | Examples of the most studied miRNAs encoded by hypermethylated genes in ccRCC, focusing on the role of aberrant methylation in miRNA deregulation, target genes, and functions.

miRNAs could someday be used as convenient markers for ccRCC in the clinic, as has been suggested for non-invasive diagnosis and prognosis of prostate cancer using miR-193b, miR-129-2, and miR-34b/c methylation in tissue and urine samples (Torres-Ferreira et al., 2017).

In **Table 1**, it is interesting to note that among the seven most studied hypermethylated miRNAs, four have lncRNAs as one or more targets. For example, miR-124-3p interacts with the lncRNA HOTAIR, miR-137 with lncRNA SNHG1, miR-182-5p with lncRNA MALAT1, and miR-203 with the HOTAIR and SNHG14 lncRNAs. These data reinforce the role of lncRNAs in the regulatory functions of miRNA, apparently by acting primarily as ceRNA to reduce the content of specific miRNAs depending on other factors.

CLINICAL APPLICATION OF MIRNAS AS MARKERS FOR DIAGNOSIS AND PROGNOSIS

In this review, we included only the most convincing results obtained from studies featuring large sample sizes and detailed

statistical evaluations. Initially, we examined the features of miRNAs that are relevant concerning their diagnostic potential. These features are summarized in **Table 2**.

The diagnostic value of elevated miR-210 levels is well established. This may be due to chronic hypoxia observed in ccRCC (Zhang Z. et al., 2009). However, no correlation of miR-210 expression with tumor stage has been reported (Iwamoto et al., 2014). The expression of miR-1233 in RCC cell lines is also stimulated by hypoxia (Dias et al., 2017).

The target of miR-451 is an mRNA encoding the PSMB8 protein, which has a pro-inflammatory function, and which is presumably significant in RCC (Zhu et al., 2016). In RCC, miR-193a-3p affects the PI3K/AKT pathway (Liu L. et al., 2017; Pan et al., 2018a). miR-28-5p is associated with chromosomal instability (Hell et al., 2014). In addition, its target is Ras-related small GTP-binding oncoprotein RAP1B (Wang et al., 2016). miR-141 is involved in the suppression of EMT (Li W. et al., 2014).

Targets of miR-144 include *mTOR* (Xiang et al., 2016) and *MAP3K8* (Liu F. et al., 2016). Thus, miR-144 can suppress cellular proliferation, EMT, and metastasis. However, another study (Xiao et al., 2017) implicated miR-144 as having an oncogenic role, with involvements in the stimulation of proliferation, migration, invasion, and resistance to sunitinib. Furthermore, miR-144 acts as an upregulated marker in the plasma of ccRCC patients, particularly those with advanced pT stage, which is a characteristic of oncogenes (Lou et al., 2017). The inconsistent functional properties of miR-144 in ccRCC revealed to date highlight the need for further studies before its clinical application can be realized.

miR-378 stands apart from other miRNAs. In a study that used samples from patients at all stages of ccRCC (Redova et al., 2012),

the diagnostic value of the increased expression of miR-378 was found. However, when samples were used only from patients at the I-II stage, the decreased expression of miR-378 had diagnostic value (Wang et al., 2015). This dichotomy of miRNA - with an oncogenic role in some types of cancer or at certain stages of the same disease, while being a tumor suppressor in other cases - is quite common. In ccRCC, miR-378 could be a potential marker of the disease stage (Li H.C. et al., 2014; Fedorko et al., 2015). However, Hauser et al. (2012) rejected the correlation between its expression in serum and pathologic stage. The authors also did not find evidence of differential expression of miR-378 levels in diseased and healthy individuals. The expression of miR-378 is likely associated with angiogenesis (Li H.C. et al., 2014). Indeed, in the ccRCC cell line, the expression of miR-378 declines with prolonged hypoxia (Chen S.C. et al., 2017). However, the mechanisms of action of miR-378 in ccRCC have not yet been adequately studied.

The genes discussed above, including some with inconsistent features (miR-144 and miR-378), have been suggested for the diagnosis of ccRCC (**Table 2**). Thus, the methods of ccRCC diagnosis need further verification.

The level of some miRNAs can be valuable as prognostic markers of survival in ccRCC (**Table 3**). Reviews describing miRNA markers used to predict survival and metastasis in ccRCC, have been published (Fedorko et al., 2016; Ran et al., 2017; He Y.H. et al., 2018). In the present review, we present only the most recent studies that featured large sample sizes (**Table 3**). Unfortunately, the data obtained so far has not always been consistent. For example, an increase of miR-210, which serves as a diagnostic criterion for ccRCC, was associated with prolonged overall survival in one study (McCormick et al., 2013) and

| miRNA and level | Diagnostic value | Type and size of samples used | References |
|---|---|--|-----------------------|
| miR-378 ↑, miR-451 ↓ | AUC 0.86, Sn 81%, Sp 83% | 15 ccRCC + 90 RCC (73 ccRCC), 12+35 healthy controls | Redova et al., 2012 |
| miR-210 ↑ | AUC 0.81, Sn 74%, Sp 76% | Meta-analysis – 570 RCC patients | Chen et al., 2018 |
| miR-378 ↑, miR-210 ↑ | Sn 80%, Sp 78% | 157 ccRCC; 12 chromophobe RCC; 26 papillary RCC | Fedorko et al., 2015 |
| miR-193a-3p ↑, miR-362 ↑, miR-572 ↑, miR-28-5p ↓, miR-378 ↓ | AUC 0.81 – for training sample set; AUC 0.80 – for the validating sample set | 25+107 ccRCC patients | Wang et al., 2015 |
| miR-210 ↑ | Sn 70%, Sp 62% | 82 ccRCC patients, serum exosomes | Zhang W. et al., 2018 |
| miR-1233 ↑ | Sn 81%, Sp 76% | | |
| miR-34a ↓ | Sn 81%, Sp 80% | 30 ccRCC patients | Yadav et al., 2017 |
| miR-141 ↓ | Sn 75%, Sp 73% | | |
| miR-1233 ↑ | Sn 93%, Sp 100% | | |
| miR-141 ↓, miR-1233 ↑ | Sn 100%, Sp 73%. | | |
| miR-144-3p ↑ | AUC 0.91, Sn 87%, Sp 83% – ccRCC vs healthy donors; AUC 0.82, Sn 75%, Sp 72% – ccRCC vs renal angiomyolipomas | 106 ccRCC, 28 renal angiomyolipomas, 123 healthy donors | Lou et al., 2017 |

TABLE 2 | Non-invasive markers suggested for use in ccRCC diagnosis: miRNA level and diagnostic value

Note: \downarrow – decreased level of miRNA; \uparrow – increased level of miRNA; AUC – area under the receiver operating characteristic curve, Sn – sensitivity, Sp – specificity. Only the most convincing results were included, with sample sizes \geq 30 and detailed statistical evaluations: AUC, sensitivity, and specificity.

TABLE 3 | Prognostic markers for poor survival and metastasis in ccRCC: miRNA level and predictive value.

| miRNA and level | Predictive value | References |
|---|--|--------------------------|
| | Lower stage and grade, mir-210 \downarrow is associated with the worst OS | McCormick et al., 2013 |
| miR-21 ↑, miR-126 ↓ | Shorter CSS | Vergho et al., 2014 |
| (miR-21/miR-10b)* ↑ | Poor prognosis in M0 | Fritz et al., 2014 |
| miR-100 ↑ | Worse OS and CSS | Wang et al., 2013 |
| miR-155 ↓ | Poor prognosis in stage III and IV (43 from 57 patients) | Shinmei et al., 2013 |
| miR-217 ↓ | Higher stage and grade, worse OS | Li H. et al., 2013 |
| miR-497 ↓ | Shorter OS | Zhao et al., 2015 |
| miR-125b ↑ | Worse CSS, early recurrence | Fu et al., 2014 |
| miR-514 ↓ | Primary M1 and recurrent | Wotschofsky et al., 2013 |
| miR-204, miR-30c, miR-30a-3p, miR-30a-5p, miR-30e-3p, miR-30e-5p, miR-30c-2-3p ↓ | M1 and distant metastasis vs M0 | Heinzelmann et al., 2014 |
| miR-30c, miR-126, miR-451 ↑ | Shorter PFS and CSS | Heinzelmann et al., 2014 |
| miR-215 ↓ | Reduced DFS | Khella H.W. et al., 2013 |
| miR-30a↓ | Hematogenous metastasis | Huang Q.B. et al., 2013 |
| miR-630 ↑ | Worst OS, metastasis | Zhao et al., 2014 |
| miRNA-23b/27b cluster ↓ | Worst OS, higher chance of disease recurrence | lshihara et al., 2014 |
| miR-29b ↑ | Worst OS | Xu Y. et al., 2015 |
| miR-126 ↓ | Worst OS for large tumors (>4 cm) | Khella H.W. et al., 2015 |
| miR-203 ↓ | Worst OS; FGF2 – target (mitogenic and angiogenic activities) | Xu M. et al., 2015 |
| miR-21, miR-1260b, miR-210, miR-100, miR-125b, miR-221, miR-630, miR-497 ↑; miR-106b, miR-99a, miR-1826, miR-215, miR-217, miR-187, miR-129-3p, miR-23b, miR-27b, miR-126 ↓ | miR-21 \uparrow is associated with the worst OS, CSS, DFS; miR-126 \downarrow is associated with the worst OS, CSS, DFS | Gu et al., 2015 |
| miR-21 ↑, miR-210 ↑, miR-141 ↓, miR-200c ↓, miR-429 ↓ | Using these 5 miRNAs, a classificator of the risk of recurrence was constructed | Tang and Xu, 2015 |
| miR-210 ↑ | Worst OS, higher chance of disease recurrence | Samaan et al., 2015 |
| miR-194 ↓ | Worst OS and DFS | Nofech-Mozes et al., 201 |
| miR-429 ↓ | Association with metastases and shorter DFS and OS. Prevents a reduction in the level of E-cadherin in EMT, reduces motility | Machackova et al., 2016 |
| miR-10a-5p ↓ | Association with tumor progression. If in the prediction of survival by stage and degree data use miR-10a-5p, AUC increased from 0.942 up to 0.995 | Kowalik et al., 2017 |
| miR-10b ↓ | Level of miR-10b is significantly lower in primary ccRCC vs normal tissues; it is lower in primary ccRCC vs oncocytoma; it is lower in metastatic ccRCC vs primary ccRCC; it is lower in stages III/IV vs I/II. Decrease in expression is associated with smaller DFS and OS | Khella et al., 2017 |
| miR-224 ↑ | Correlation with significantly worse survival | Fujii et al., 2017 |
| miR-18a-5p ↑ | iR-18a-5p ↑ Correlation with significantly worse survival | |
| miR-122-5p \downarrow miR-206 \downarrow in ccRCC and in benign renal tumors | miR-122-5p \uparrow correlates with metastases and worse survival; miR-206 \uparrow correlates with pT stage and metastases and worse survival | Heinemann et al., 2018 |
| miR-21 ↑, miR-142-5p ↑, miR-194 ↓ | Correlation with metastases is characterized by 87% sensitivity and 82% specificity | Lokeshwar et al., 2018 |

Note: \downarrow – decreased level of miRNA; \uparrow – increased level of miRNA; OS – overall survival; DFS – disease-free survival; CSS – cancer-specific survival; PFS – progression-free survival, * – ratio. Only the most recent (since 2013) works in which sets of >35 ccRCC samples were included.

decreased overall survival in another study (Samaan et al., 2015). For most miRNAs, the nature of interactions in ccRCC is not well understood.

Despite the relative simplicity of miRNA gene methylation studies, the approach has barely been used instead of the expression levels of miRNAs, to evaluate the prognosis of survival or predict metastasis in ccRCC. The only exception is a study (Gebauer et al., 2013) that described that the methylation of the gene encoding miR-124-3 was associated with a more advanced RCC stage, metastasis, and an increased risk of relapse in 111 RCC samples (80 ccRCC, with 77 paired with histologically normal tissue). Also, a recent review did not mention studies involving miRNA gene methylation in RCC (Ran et al., 2017). Studies we undertook identified novel biomarker systems by using hypermethylated miRNA genes as an indicator in ccRCC, both for the diagnosis (MIR-125b-1, MIR-375, MIR-137, and MIR-193a) and for the prediction of metastasis (MIR-125b-1, MIR-375, MIR-107, MIR-1258, and MIR-203) (Loginov et al., 2017; Varachev et al., 2018). The use of both biomarker systems in 70 patients was characterized by a high discrimination value [area under the curve (AUC) 0.93], sensitivity (86%), and specificity (95%).

Few studies have explored the use of the miRNA level as an indicator to predict the response to ccRCC treatment. Most of the studies have focused on predicting the success of the response to tyrosine kinase inhibitors that are the first-line treatment for metastatic ccRCC. However, approximately 20% of the patients who are treated rapidly develop resistance to these drugs. For example, for 74 patients with metastatic ccRCC who were treated with tyrosine kinase inhibitors, early progression of the disease was observed in 16 (Garcia-Donas et al., 2016). Selection of increased miR-1307-3p and miR-425-5p levels as the biomarker predicted the risk of poor outcome to treatment with an AUC value of 0.75, which is better than the risks predicted using parameters based on clinicopathological factors. This result was revalidated in a group of 64 patients (Garcia-Donas et al., 2016). The predictors of the outcome of sunitinib treatment were studied in 123 patients with ccRCC (the dataset included metastatic and non-metastatic ccRCC patients). Of the 123 patients, 97 were characterized by prolonged (>22 months) progression-free survival (PFS) and the remaining 26 showed rapid progression of the disease in the first 3 months (Puente et al., 2017). It must be noted that, along with the other characteristic clinical indices, the former group was distinguished by the presence of high levels of the transcription factors HEYL, HEY, and HES, accompanied by elevated levels of miR-27b, miR-23b, and miR-628-5p, although miR-23b is considered as a typical oncogene (Zaman et al., 2012). In another study, 56 sunitinib-treated patients with metastatic ccRCC (24 responsive patients with PFS > 18 months and 32 non-responsive patients with PFS < 6 months) (Kovacova et al., 2018). A good response to the drug was positively associated with miR-942 and miR-133 levels, and the model based on using their expression levels as indicators gave predictions with an AUC value of 0.81.

Another aspect of the use of miRNA as biomarkers is the predictive capabilities of miRNA polymorphisms (variations or polymorphisms in sequences of miRNA, target mRNA, miRNA genes, and target miRNA genes and their pathways). The polymorphism in the MIR-34b/c promoter region has been amply studied. This polymorphism reduces the expression of this miRNA (allele C for rs4938723) in the homozygotic condition, increases the risk of RCC in the Chinese population, especially in elderly, men, smokers, and alcohol abusers (Zhang S. et al., 2014). Polymorphisms in the miRNA-binding sites of 102 genes belonging to the VHL-HIF1a pathway and their effects on RCC risk have been analyzed (Wei et al., 2014). The most significant effect of two polymorphisms in the MAPK1 gene, and the presence of four of the five adverse alleles in the MAPK1, CDCP1, TFRC, and DEC1 gene set, was a more than twofold increase in the risk of RCC.

The genotype CC, a variant of the 3' untranslated region (UTR) of the SET domain containing lysine methyltransferase 8 (*SET8*) gene, significantly reduced the risk of ccRCC (the odds ratio = 0.318). This single nucleotide polymorphism, also designated rs16917496, is located in the miR-502 binding site within the 3'UTR of the *SET8* gene and is associated with

its downregulation in ccRCC. Knockdown of *SET8* inhibited proliferation, migration, and invasiveness of ccRCC cells in culture (Zhang et al., 2017). Many ccRCC-influencing genes are associated with remodeling, and *SET8* may be one of them (Joosten et al., 2018).

CONCLUSION

In recent years, the role of miRNA in the pathogenesis of ccRCC has been amply studied. In ccRCC, miRNAs involved in processes that include the response to hypoxia, EMT, and chromatin remodeling play significant roles. Nevertheless, the picture is far from complete. The search for new diagnostic and prognostic markers is still largely empirical and the associated pathways and processes are not clearly elucidated. For example, in a study involving prediction and correlation analyses, miRNA-mRNA pairs displayed a significant inverse relationship. miR-30b was one of these miRNAs, and its expression was negatively correlated with the highest number of genes in ccRCC (Liu et al., 2018). Among the latter, there are, for example, genes that are significant for tumor invasion, such as Integrin Subunit Alpha 5 (ITGA5). Nevertheless, almost nothing is known concerning the influence of miR-30b on ccRCC. The many potential biomarkers include some differentially expressed miRNAs or differentially methylated sites. However, only a few have been experimentally verified (Wang Z. et al., 2018).

For the diagnosis of ccRCC, the use of miRNAs like miR-144 has been proposed since their targets may be relevant for this cancer. However, this use is hampered by inconsistent findings, with both tumor suppressive and oncogenic activity reported in separate studies. Results of various studies for other miRNAs, including miR-106a, miR-125b, miR-203, and miR-378, have been contradictory. These discrepancies may be due to the varying roles of these miRNAs at different stages of cancer or in different forms of the disease or to their ability to interact with various targets depending on the cellular context.

Our recent contributions have revealed the potential for the use of the methylation profiles of miRNA genes in the search for reliable diagnostic and prognostic marker systems (i.e., high AUC). The results have suggested the need to expand the search for the selection of new miRNA genes regulated by methylation. Moreover, in-depth studies of the identified hypermethylated miRNA genes associated with metastasis with respect to their target genes and functions are necessary.

The recently discovered lncRNAs represent a novel class of potential prognostic biomarkers in ccRCC. Data on the function and clinical applications of lncRNA in ccRCC is accumulating, with 60 articles identified in PubMed as of January 2019. However, only a few of these studies (10%) explored the interactions of lncRNAs with miRNAs. Nevertheless, a detailed analysis of the molecular mechanisms of ccRCC in original articles allowed us to identify at least seven miRNAs (miR-122, miR-124-3p, miR-137, miR-141, miR-182-5p, miR-203, and miR-217) that interact with the lncRNAs (e.g., HOTAIR, MALAT-1, SNHG1, and SNHG14). Further research is needed to elucidate these interactions in ccRCC and to assess their clinical potential.

AUTHOR CONTRIBUTIONS

EB, MF, VL, AD, and SM wrote the manuscript. All the authors revised the work critically for important intellectual content, approved the version to be published, and agreed to be

REFERENCES

- Andorfer, C. A., Necela, B. M., Thompson, E. A., and Perez, E. A. (2011). MicroRNA signatures: clinical biomarkers for the diagnosis and treatment of breast cancer. *Trends Mol. Med.* 17, 313–319. doi: 10.1016/j.molmed.2011. 01.006
- Baylin, S. B., and Jones, P. A. (2016). Epigenetic determinants of cancer. Cold Spring Harb. Perspect. Biol. 8:a019505. doi: 10.1101/cshperspect.a019505
- Beresneva, E. V., Rykov, S. V., Khodyrev, D. S., Pronina, I. V., Ermilova, V. D., Kazubskaya, T. P., et al. (2013). Methylation profile of group of miRNA genes in clear cell renal cell carcinoma and their involvement in cancer progression. *Russ. J. Genet.* 49, 320–328. doi: 10.1134/S102279541303 0034
- Berkers, J., Govaere, O., Wolter, P., Beuselinck, B., Schoffski, P., van Kempen, L. C., et al. (2013). A possible role for microRNA-141 down-regulation in sunitinib resistant metastatic clear cell renal cell carcinoma through induction of epithelial-to-mesenchymal transition and hypoxia resistance. J. Urol. 189, 1930–1938. doi: 10.1016/j.juro.2012. 11.133
- Bhat, N. S., Colden, M., Dar, A. A., Saini, S., Arora, P., Shahryari, V., et al. (2017). MicroRNA-720 regulates E-cadherin-alphaE-catenin complex and promotes renal cell carcinoma. *Mol. Cancer Ther.* 16, 2840–2848.
- Braga, E. A., Khodyrev, D. S., Loginov, V. I., Pronina, I. V., Senchenko, V. N., Dmitriev, A. A., et al. (2015). Methylation in the regulation of the expression of chromosome 3 and microRNA genes in clear-cell renal cell carcinomas. *Russ. J. Genet.* 51, 566–581. doi: 10.1134/S1022795415050026
- Braga, E. A., Loginov, V. I., Burdennyi, A. M., Filippova, E. A., Pronina, I. V., Kurevlev, S. V., et al. (2018). Five hypermethylated microRNA genes as potential markers of ovarian cancer. *Bull. Exp. Biol. Med.* 164, 351–355. doi: 10.1007/ s10517-018-3988-y
- Butz, H., Szabo, P. M., Khella, H. W., Nofech-Mozes, R., Patocs, A., and Yousef, G. M. (2015). miRNA-target network reveals miR-124as a key miRNA contributing to clear cell renal cell carcinoma aggressive behaviour by targeting CAV1 and FLOT1. Oncotarget 6, 12543–12557. doi: 10.18632/oncotarget.3815
- Cairns, P. (2010). Renal cell carcinoma. Cancer Biomark. 9, 461–473. doi: 10.3233/ CBM-2011-0176
- Camps, C., Buffa, F. M., Colella, S., Moore, J., Sotiriou, C., Sheldon, H., et al. (2008). hsa-miR-210 Is induced by hypoxia and is an independent prognostic factor in breast cancer. *Clin. Cancer Res.* 14, 1340–1348. doi: 10.1158/1078-0432.CCR-07-1755
- Cancer Genome Atlas Research, N. (2013). Comprehensive molecular characterization of clear cell renal cell carcinoma. *Nature* 499, 43–49. doi: 10.1038/nature12222
- Cao, H., Liu, Z., Wang, R., Zhang, X., Yi, W., Nie, G., et al. (2017). miR-148a suppresses human renal cell carcinoma malignancy by targeting AKT2. Oncol. Rep. 37, 147–154. doi: 10.3892/or.2016.5257
- Chan, J. J., and Tay, Y. (2018). Noncoding RNA:RNA regulatory networks in cancer. *Int. J. Mol. Sci.* 19:E1310. doi: 10.3390/ijms19051310
- Chen, C., Xue, S., Zhang, J., Chen, W., Gong, D., Zheng, J., et al. (2017). DNAmethylation-mediated repression of miR-766-3p promotes cell proliferation via targeting SF2 expression in renal cell carcinoma. *Int. J. Cancer* 141, 1867–1878. doi: 10.1002/ijc.30853
- Chen, J., Gu, Y., and Shen, W. (2017). MicroRNA-21 functions as an oncogene and promotes cell proliferation and invasion via TIMP3 in renal cancer. *Eur. Rev. Med. Pharmacol. Sci.* 21, 4566–4576.

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- Chen, S. C., Chen, F. W., Hsu, Y. L., and Kuo, P. L. (2017). Systematic analysis of transcriptomic profile of renal cell carcinoma under long-term hypoxia using next-generation sequencing and bioinformatics. *Int. J. Mol. Sci.* 18:E2657. doi: 10.3390/ijms18122657
- Chen, X., Ruan, A., Wang, X., Han, W., Wang, R., Lou, N., et al. (2014). miR-129-3p, as a diagnostic and prognostic biomarker for renal cell carcinoma, attenuates cell migration and invasion via downregulating multiple metastasisrelated genes. J. Cancer Res. Clin. Oncol. 140, 1295–1304. doi: 10.1007/s00432-014-1690-7
- Chen, Y., Wang, X., Zhu, X., and Shao, S. (2018). Detection performance of circulating microRNA-210 for renal cell carcinoma: a meta-analysis. *Clin. Lab.* 64, 569–576. doi: 10.7754/Clin.Lab.2017.171103
- Chen, Z., Zhang, J., Zhang, Z., Feng, Z., Wei, J., Lu, J., et al. (2017). The putative tumor suppressor microRNA-30a-5p modulates clear cell renal cell carcinoma aggressiveness through repression of ZEB2. *Cell Death Dis.* 8:e2859. doi: 10. 1038/cddis.2017.252
- Chiyomaru, T., Fukuhara, S., Saini, S., Majid, S., Deng, G., Shahryari, V., et al. (2014). Long non-coding RNA HOTAIR is targeted and regulated by miR-141 in human cancer cells. *J. Biol. Chem.* 289, 12550–12565. doi: 10.1074/jbc.M113. 488593
- Conant, J. L., Peng, Z., Evans, M. F., Naud, S., and Cooper, K. (2011). Sarcomatoid renal cell carcinoma is an example of epithelial–mesenchymal transition. *J. Clin. Pathol.* 64, 1088–1092. doi: 10.1136/jclinpath-2011-200216
- Cora, D., Re, A., Caselle, M., and Bussolino, F. (2017). MicroRNA-mediated regulatory circuits: outlook and perspectives. *Phys. Biol.* 14:045001. doi: 10. 1088/1478-3975/aa6f21
- Cui, L., Zhou, H., Zhao, H., Zhou, Y., Xu, R., Xu, X., et al. (2012). MicroRNA-99a induces G1-phase cell cycle arrest and suppresses tumorigenicity in renal cell carcinoma. *BMC Cancer* 12:546. doi: 10.1186/1471-2407-12-546
- Cui, M., Liu, W., Zhang, L., Guo, F., Liu, Y., Chen, F., et al. (2017). Over-Expression of miR-21 and Lower PTEN Levels in Wilms' Tumor with aggressive behavior. *Tohoku J. Exp. Med.* 242, 43–52. doi: 10.1620/tjem.242.43
- Dasgupta, P., Kulkarni, P., Majid, S., Shahryari, V., Hashimoto, Y., Bhat, N. S., et al. (2018). MicroRNA-203 inhibits long noncoding RNA HOTAIR and regulates tumorigenesis through epithelial-to-mesenchymal transition pathway in renal cell carcinoma. *Mol. Cancer Ther.* 17, 1061–1069. doi: 10.1158/1535-7163. MCT-17-0925
- Dias, F., Teixeira, A. L., Ferreira, M., Adem, B., Bastos, N., Vieira, J., et al. (2017). Plasmatic miR-210, miR-221 and miR-1233 profile: potential liquid biopsies candidates for renal cell carcinoma. *Oncotarget* 8, 103315–103326. doi: 10. 18632/oncotarget.21733
- Ding, J., Yeh, C. R., Sun, Y., Lin, C., Chou, J., Ou, Z., et al. (2018). Estrogen receptor beta promotes renal cell carcinoma progression via regulating LncRNA HOTAIR-miR-138/200c/204/217 associated CeRNA network. Oncogene 37, 5037–5053. doi: 10.1038/s41388-018-0175-6
- Dmitriev, A. A., Rudenko, E. E., Kudryavtseva, A. V., Krasnov, G. S., Gordiyuk, V. V., Melnikova, N. V., et al. (2014). Epigenetic alterations of chromosome 3 revealed by NotI-microarrays in clear cell renal cell carcinoma. *Biomed. Res. Int.* 2014:735292. doi: 10.1155/2014/735292
- Doberstein, K., Steinmeyer, N., Hartmetz, A. K., Eberhardt, W., Mittelbronn, M., Harter, P. N., et al. (2013). MicroRNA-145 targets the metalloprotease ADAM17 and is suppressed in renal cell carcinoma patients. *Neoplasia* 15, 218–230.
- Dragomir, M., Mafra, A. C. P., Dias, S. M. G., Vasilescu, C., and Calin, G. A. (2018). Using microRNA Networks to Understand Cancer. *Int. J. Mol. Sci.* 19:E1871. doi: 10.3390/ijms19071871

- Exposito-Villen, A., E Aránega, A., and Franco, D. (2018). Functional role of noncoding RNAs during epithelial-to-mesenchymal transition. *Noncoding RNA* 4:E14. doi: 10.3390/ncrna4020014
- Fan, W., Huang, J., Xiao, H., and Liang, Z. (2016). MicroRNA-22 is downregulated in clear cell renal cell carcinoma, and inhibits cell growth, migration and invasion by targeting PTEN. *Mol. Med. Rep.* 13, 4800–4806. doi: 10.3892/mmr. 2016.5101
- Fan, Y., Li, H., Ma, X., Gao, Y., Bao, X., Du, Q., et al. (2016). Dicer suppresses the malignant phenotype in VHL-deficient clear cell renal cell carcinoma by inhibiting HIF-2alpha. Oncotarget 7, 18280–18294. doi: 10.18632/oncotarget. 7807
- Fan, Y., Ma, X., Li, H., Gao, Y., Huang, Q., Zhang, Y., et al. (2018). miR-122 promotes metastasis of clear-cell renal cell carcinoma by downregulating Dicer. *Int. J. Cancer* 142, 547–560. doi: 10.1002/ijc.31050
- Fang, X., Sun, R., Hu, Y., Wang, H., Guo, Y., Yang, B., et al. (2018). miRNA-182-5p, via HIF2alpha, contributes to arsenic carcinogenesis: evidence from human renal epithelial cells. *Metallomics* 10, 1607–1617. doi: 10.1039/c8mt 00251g
- Fedorko, M., Pacik, D., Wasserbauer, R., Juracek, J., Varga, G., Ghazal, M., et al. (2016). MicroRNAs in the pathogenesis of renal cell carcinoma and their diagnostic and prognostic utility as cancer biomarkers. *Int. J. Biol. Markers* 31, e26–e37. doi: 10.5301/jbm.5000174
- Fedorko, M., Stanik, M., Iliev, R., Redova-Lojova, M., Machackova, T., Svoboda, M., et al. (2015). Combination of MiR-378 and MiR-210 serum levels enables sensitive detection of renal cell carcinoma. *Int. J. Mol. Sci.* 16, 23382–23389. doi: 10.3390/ijms161023382
- Fritz, H. K., Lindgren, D., Ljungberg, B., Axelson, H., and Dahlback, B. (2014). The miR(21/10b) ratio as a prognostic marker in clear cell renal cell carcinoma. *Eur. J. Cancer* 50, 1758–1765. doi: 10.1016/j.ejca.2014.03.281
- Fu, Q., Liu, Z., Pan, D., Zhang, W., Xu, L., Zhu, Y., et al. (2014). Tumor miR-125b predicts recurrence and survival of patients with clear-cell renal cell carcinoma after surgical resection. *Cancer Sci.* 105, 1427–1434. doi: 10.1111/cas. 12507
- Fujii, N., Hirata, H., Ueno, K., Mori, J., Oka, S., Shimizu, K., et al. (2017). Extracellular miR-224 as a prognostic marker for clear cell renal cell carcinoma. *Oncotarget* 8, 109877–109888. doi: 10.18632/oncotarget.22436
- Gao, C., Peng, F. H., and Peng, L. K. (2014). MiR-200c sensitizes clear-cell renal cell carcinoma cells to sorafenib and imatinib by targeting heme oxygenase-1. *Neoplasma* 61, 680–689. doi: 10.4149/neo_2014_083
- Garcia-Donas, J., Beuselinck, B., Inglada-Perez, L., Grana, O., Schoffski, P., Wozniak, A., et al. (2016). Deep sequencing reveals microRNAs predictive of antiangiogenic drug response. *JCI Insight* 1:e86051. doi: 10.1172/jci.insight. 86051
- Gattolliat, C. H., Couve, S., Meurice, G., Orear, C., Droin, N., Chiquet, M., et al. (2018). Integrative analysis of dysregulated microRNAs and mRNAs in multiple recurrent synchronized renal tumors from patients with von Hippel-Lindau disease. *Int. J. Oncol.* 53, 1455–1468. doi: 10.3892/ijo.2018.4490
- Gebauer, K., Peters, I., Dubrowinskaja, N., Hennenlotter, J., Abbas, M., Scherer, R., et al. (2013). Hsa-mir-124-3 CpG island methylation is associated with advanced tumours and disease recurrence of patients with clear cell renal cell carcinoma. *Br. J. Cancer* 108, 131–138. doi: 10.1038/bjc.2012.537
- Geethadevi, A., Sharma, A., Sharma, M. K., and Parashar, D. (2018). An interplay between microRNA and SOX4 in the regulation of epithelial-mesenchymal transition and cancer progression. *Cancer Transl. Med.* 4, 17–27. doi: 10.4103/ ctm.ctm_4_18
- Goradel, N. H., Mohammadi, N., Haghi-Aminjan, H., Farhood, B., Negahdari, B., and Sahebkar, A. (2018). Regulation of tumor angiogenesis by microRNAs: State of the art. J. Cell. Physiol. 234, 1099–1110. doi: 10.1002/jcp.27051
- Gu, L., Li, H., Chen, L., Ma, X., Gao, Y., Li, X., et al. (2015). MicroRNAs as prognostic molecular signatures in renal cell carcinoma: a systematic review and meta-analysis. *Oncotarget* 6, 32545–32560. doi: 10.18632/oncotarget.5324
- Han, F., Wu, Y., and Jiang, W. (2015). MicroRNA-18a decreases choroidal endothelial cell proliferation and migration by inhibiting HIF1A expression. *Med. Sci. Monit.* 21, 1642–1647. doi: 10.12659/MSM.893068
- Han, L., Witmer, P. D., Casey, E., Valle, D., and Sukumar, S. (2007). DNA methylation regulates MicroRNA expression. *Cancer Biol. Ther.* 6, 1284–1288.
- Harada, K., Miyake, H., Kusuda, Y., and Fujisawa, M. (2012). Expression of epithelial-mesenchymal transition markers in renal cell carcinoma: impact on

prognostic outcomes in patients undergoing radical nephrectomy. *BJU Int.* 110(11 Pt C), E1131–E1137. doi: 10.1111/j.1464-410X.2012.11297.x

- Hauser, S., Wulfken, L. M., Holdenrieder, S., Moritz, R., Ohlmann, C. H., Jung, V., et al. (2012). Analysis of serum microRNAs (miR-26a-2*, miR-191, miR-337-3p and miR-378) as potential biomarkers in renal cell carcinoma. *Cancer Epidemiol.* 36, 391–394. doi: 10.1016/j.canep.2012.04.001
- He, C., Zhao, X., Jiang, H., Zhong, Z., and Xu, R. (2015). Demethylation of miR-10b plays a suppressive role in ccRCC cells. *Int. J. Clin. Exp. Pathol.* 8, 10595–10604.
- He, H., Dai, J., Zhuo, R., Zhao, J., Wang, H., Sun, F., et al. (2018). Study on the mechanism behind lncRNA MEG3 affecting clear cell renal cell carcinoma by regulating miR-7/RASL11B signaling. J. Cell. Physiol. 233, 9503–9515. doi: 10.1002/jcp.26849
- He, H., Wang, L., Zhou, W., Zhang, Z., Wang, L., Xu, S., et al. (2015). MicroRNA expression profiling in clear cell renal cell carcinoma: identification and functional validation of key miRNAs. *PLoS One* 10:e0125672. doi: 10.1371/ journal.pone.0125672
- He, Y. H., Chen, C., and Shi, Z. (2018). The biological roles and clinical implications of microRNAs in clear cell renal cell carcinoma. J. Cell. Physiol. 233, 4458–4465. doi: 10.1002/jcp.26347
- Heinemann, F. G., Tolkach, Y., Deng, M., Schmidt, D., Perner, S., Kristiansen, G., et al. (2018). Serum miR-122-5p and miR-206 expression: non-invasive prognostic biomarkers for renal cell carcinoma. *Clin. Epigenetics* 10:11. doi: 10.1186/s13148-018-0444-9
- Heinzelmann, J., Unrein, A., Wickmann, U., Baumgart, S., Stapf, M., Szendroi, A., et al. (2014). MicroRNAs with prognostic potential for metastasis in clear cell renal cell carcinoma: a comparison of primary tumors and distant metastases. *Ann. Surg. Oncol.* 21, 1046–1054. doi: 10.1245/s10434-013-3361-3
- Hell, M. P., Thoma, C. R., Fankhauser, N., Christinat, Y., Weber, T. C., and Krek, W. (2014). miR-28-5p promotes chromosomal instability in VHLassociated cancers by inhibiting Mad2 translation. *Cancer Res.* 74, 2432–2443. doi: 10.1158/0008-5472.CAN-13-2041
- Heller, G., Altenberger, C., Steiner, I., Topakian, T., Ziegler, B., Tomasich, E., et al. (2018). DNA methylation of microRNA-coding genes in non-small-cell lung cancer patients. J. Pathol. 245, 387–398. doi: 10.1002/path.5079
- Hildebrandt, M. A., Gu, J., Lin, J., Ye, Y., Tan, W., Tamboli, P., et al. (2010). HsamiR-9 methylation status is associated with cancer development and metastatic recurrence in patients with clear cell renal cell carcinoma. *Oncogene* 29, 5724–5728. doi: 10.1038/onc.2010.305
- Hirata, H., Ueno, K., Nakajima, K., Tabatabai, Z. L., Hinoda, Y., Ishii, N., et al. (2013). Genistein downregulates onco-miR-1260b and inhibits Wnt-signalling in renal cancer cells. *Br. J. Cancer* 108, 2070–2078. doi: 10.1038/bjc.2013.173
- Hong, Q., Li, O., Zheng, W., Xiao, W. Z., Zhang, L., Wu, D., et al. (2017). LncRNA HOTAIR regulates HIF-1alpha/AXL signaling through inhibition of miR-217 in renal cell carcinoma. *Cell Death Dis.* 8:e2772. doi: 10.1038/cddis. 2017.181
- Hu, G., Lai, P., Liu, M., Xu, L., Guo, Z., Liu, H., et al. (2014). miR-203a regulates proliferation, migration, and apoptosis by targeting glycogen synthase kinase-3beta in human renal cell carcinoma. *Tumour Biol.* 35, 11443–11453. doi: 10. 1007/s13277-014-2476-x
- Huang, J., Yao, X., Zhang, J., Dong, B., Chen, Q., Xue, W., et al. (2013). Hypoxiainduced downregulation of miR-30c promotes epithelial-mesenchymal transition in human renal cell carcinoma. *Cancer Sci.* 104, 1609–1617. doi: 10.1111/cas.12291
- Huang, Q., Sun, Y., Ma, X., Gao, Y., Li, X., Niu, Y., et al. (2017). Androgen receptor increases hematogenous metastasis yet decreases lymphatic metastasis of renal cell carcinoma. *Nat. Commun.* 8:918.
- Huang, Q. B., Ma, X., Zhang, X., Liu, S. W., Ai, Q., Shi, T. P., et al. (2013). Downregulated miR-30a in clear cell renal cell carcinoma correlated with tumor hematogenous metastasis by targeting angiogenesis-specific DLL4. *PLoS One* 8:e67294. doi: 10.1371/journal.pone.0067294
- Huang, X., Huang, M., Kong, L., and Li, Y. (2015). miR-372 suppresses tumour proliferation and invasion by targeting IGF2BP1 in renal cell carcinoma. *Cell Prolif.* 48, 593–599. doi: 10.1111/cpr.12207
- Ishihara, T., Seki, N., Inoguchi, S., Yoshino, H., Tatarano, S., Yamada, Y., et al. (2014). Expression of the tumor suppressive miRNA-23b/27b cluster is a good prognostic marker in clear cell renal cell carcinoma. *J. Urol.* 192, 1822–1830. doi: 10.1016/j.juro.2014.07.001

- Iwamoto, H., Kanda, Y., Sejima, T., Osaki, M., Okada, F., and Takenaka, A. (2014). Serum miR-210 as a potential biomarker of early clear cell renal cell carcinoma. *Int. J. Oncol.* 44, 53–58. doi: 10.3892/ijo.2013.2169
- Jasinski-Bergner, S., Stoehr, C., Bukur, J., Massa, C., Braun, J., Huttelmaier, S., et al. (2015). Clinical relevance of miR-mediated HLA-G regulation and the associated immune cell infiltration in renal cell carcinoma. *Oncoimmunology* 4:e1008805. doi: 10.1080/2162402X.2015.1008805
- Ji, H., Tian, D., Zhang, B., Zhang, Y., Yan, D., and Wu, S. (2017). Overexpression of miR-155 in clear-cell renal cell carcinoma and its oncogenic effect through targeting FOXO3a. *Exp. Ther. Med.* 13, 2286–2292. doi: 10.3892/etm.2017. 4263
- Jiao, D., Wu, M., Ji, L., Liu, F., and Liu, Y. (2018). MicroRNA-186 suppresses cell proliferation and metastasis through targeting sentrin-specific protease 1 in renal cell carcinoma. *Oncol. Res.* 26, 249–259. doi: 10.3727/ 096504017X14953948675430
- Jin, L., Zhang, Z., Li, Y., He, T., Hu, J., Liu, J., et al. (2017). miR-125b is associated with renal cell carcinoma cell migration, invasion and apoptosis. Oncol. Lett. 13, 4512–4520. doi: 10.3892/ol.2017.5985
- Jingushi, K., Kashiwagi, Y., Ueda, Y., Kitae, K., Hase, H., Nakata, W., et al. (2017). High miR-122 expression promotes malignant phenotypes in ccRCC by targeting occludin. *Int. J. Oncol.* 51, 289–297. doi: 10.3892/ijo.2017.4016
- Joosten, S. C., Smits, K. M., Aarts, M. J., Melotte, V., Koch, A., Tjan-Heijnen, V. C., et al. (2018). Epigenetics in renal cell cancer: mechanisms and clinical applications. *Nat. Rev. Urol.* 15, 430–451. doi: 10.1038/s41585-018-0023-z
- Jung, M., Mollenkopf, H. J., Grimm, C., Wagner, I., Albrecht, M., Waller, T., et al. (2009). MicroRNA profiling of clear cell renal cell cancer identifies a robust signature to define renal malignancy. *J. Cell. Mol. Med.* 13, 3918–3928. doi: 10.1111/j.1582-4934.2009.00705.x
- Kankaya, D., Kiremitci, S., Tulunay, O., and Baltaci, S. (2015). Gelsolin, NFkappaB, and p53 expression in clear cell renal cell carcinoma: impact on outcome. *Pathol. Res. Pract.* 211, 505–512. doi: 10.1016/j.prp.2015.03.002
- Kashyap, D., Tuli, H. S., Garg, V. K., Goel, N., and Bishayee, A. (2018). Oncogenic and tumor-suppressive roles of MicroRNAs with special reference to apoptosis: molecular mechanisms and therapeutic potential. *Mol. Diagn. Ther.* 22, 179–201. doi: 10.1007/s40291-018-0316-1
- Kawakami, K., Enokida, H., Chiyomaru, T., Tatarano, S., Yoshino, H., Kagara, I., et al. (2012). The functional significance of miR-1 and miR-133a in renal cell carcinoma. *Eur. J. Cancer* 48, 827–836. doi: 10.1016/j.ejca.2011. 06.030
- Khella, H. W., Bakhet, M., Allo, G., Jewett, M. A., Girgis, A. H., Latif, A., et al. (2013). miR-192, miR-194 and miR-215: a convergent microRNA network suppressing tumor progression in renal cell carcinoma. *Carcinogenesis* 34, 2231–2239. doi: 10.1093/carcin/bgt184
- Khella, H. W., Scorilas, A., Mozes, R., Mirham, L., Lianidou, E., Krylov, S. N., et al. (2015). Low expression of miR-126 is a prognostic marker for metastatic clear cell renal cell carcinoma. *Am. J. Pathol.* 185, 693–703. doi: 10.1016/j.ajpath.2014. 11.017
- Khella, H. W. Z., Butz, H., Ding, Q., Rotondo, F., Evans, K. R., Kupchak, P., et al. (2015). miR-221/222 are involved in response to sunitinib treatment in metastatic renal cell carcinoma. *Mol. Ther.* 23, 1748–1758. doi: 10.1038/mt. 2015.129
- Khella, H. W. Z., Daniel, N., Youssef, L., Scorilas, A., Nofech-Mozes, R., Mirham, L., et al. (2017). miR-10b is a prognostic marker in clear cell renal cell carcinoma. *J. Clin. Pathol.* 70, 854–859. doi: 10.1136/jclinpath-2017-204341
- Kim, K., Lee, H. C., Park, J. L., Kim, M., Kim, S. Y., Noh, S. M., et al. (2011). Epigenetic regulation of microRNA-10b and targeting of oncogenic MAPRE1 in gastric cancer. *Epigenetics* 6, 740–751.
- Kovacova, J., Juracek, J., Poprach, A., Buchler, T., Kopecky, J., Fiala, O., et al. (2018). Candidate MicroRNA biomarkers of therapeutic response to sunitinib in metastatic renal cell carcinoma: a validation study in patients with extremely good and poor response. *Anticancer Res.* 38, 2961–2965. doi: 10.21873/ anticanres.12546
- Kowalik, C. G., Palmer, D. A., Sullivan, T. B., Teebagy, P. A., Dugan, J. M., Libertino, J. A., et al. (2017). Profiling microRNA from nephrectomy and biopsy specimens: predictors of progression and survival in clear cell renal cell carcinoma. *BJU Int.* 120, 428–440. doi: 10.1111/bju.13886
- Kulkarni, P., Dasgupta, P., Bhat, N. S., Shahryari, V., Shiina, M., Hashimoto, Y., et al. (2018). Elevated miR-182-5p associates with renal cancer cell mitotic arrest

through diminished MALAT-1 expression. *Mol. Cancer Res.* 16, 1750–1760. doi: 10.1158/1541-7786.MCR-17-0762

- Kunej, T., Godnic, I., Ferdin, J., Horvat, S., Dovc, P., and Calin, G. A. (2011). Epigenetic regulation of microRNAs in cancer: an integrated review of literature. *Mutat. Res.* 717, 77–84. doi: 10.1016/j.mrfmmm.2011. 03.008
- Li, H., Zhao, J., Zhang, J. W., Huang, Q. Y., Huang, J. Z., Chi, L. S., et al. (2013). MicroRNA-217, down-regulated in clear cell renal cell carcinoma and associated with lower survival, suppresses cell proliferation and migration. *Neoplasma* 60, 511–515. doi: 10.4149/neo_2013_066
- Li, H. C., Li, J. P., Wang, Z. M., Fu, D. L., Li, Z. L., Zhang, D., et al. (2014). Identification of angiogenesis-related miRNAs in a population of patients with renal clear cell carcinoma. *Oncol. Rep.* 32, 2061–2069. doi: 10.3892/or.2014.3403
- Li, L., Li, C., Wang, S., Wang, Z., Jiang, J., Wang, W., et al. (2016). Exosomes derived from hypoxic oral squamous cell carcinoma cells deliver miR-21 to normoxic cells to elicit a prometastatic phenotype. *Cancer Res.* 76, 1770–1780. doi: 10.1158/0008-5472.CAN-15-1625
- Li, M., Wang, Y., Cheng, L., Niu, W., Zhao, G., Raju, J. K., et al. (2017). Long non-coding RNAs in renal cell carcinoma: a systematic review and clinical implications. *Oncotarget* 8, 48424–48435. doi: 10.18632/oncotarget.17053
- Li, M., Wang, Y., Song, Y., Bu, R., Yin, B., Fei, X., et al. (2015). MicroRNAs in renal cell carcinoma: a systematic review of clinical implications (Review). Oncol. Rep. 33, 1571–1578. doi: 10.3892/or.2015.3799
- Li, W., Wang, Q., Su, Q., Ma, D., An, C., Ma, L., et al. (2014). Honokiol suppresses renal cancer cells' metastasis via dual-blocking epithelial-mesenchymal transition and cancer stem cell properties through modulating miR-141/ZEB2 signaling. Mol. Cells 37, 383–388. doi: 10.14348/molcells.2014.0009
- Li, X., Xin, S., He, Z., Che, X., Wang, J., Xiao, X., et al. (2014). MicroRNA-21 (miR-21) post-transcriptionally downregulates tumor suppressor PDCD4 and promotes cell transformation, proliferation, and metastasis in renal cell carcinoma. *Cell Physiol. Biochem.* 33, 1631–1642. doi: 10.1159/000362946
- Lian, J. H., Wang, W. H., Wang, J. Q., Zhang, Y. H., and Li, Y. (2013). MicroRNA-122 promotes proliferation, invasion and migration of renal cell carcinoma cells through the PI3K/Akt signaling pathway. *Asian Pac. J. Cancer Prev.* 14, 5017–5021.
- Liang, B., Zhao, J., and Wang, X. (2017). A three-microRNA signature as a diagnostic and prognostic marker in clear cell renal cancer: an *in silico* analysis. *PLoS One* 12:e0180660. doi: 10.1371/journal.pone.0180660
- Liu, F., Chen, N., Xiao, R., Wang, W., and Pan, Z. (2016). miR-144-3p serves as a tumor suppressor for renal cell carcinoma and inhibits its invasion and metastasis by targeting MAP3K8. *Biochem. Biophys. Res. Commun.* 480, 87–93. doi: 10.1016/j.bbrc.2016.10.004
- Liu, F., Lou, Y. L., Wu, J., Ruan, Q. F., Xie, A., Guo, F., et al. (2012). Upregulation of microRNA-210 regulates renal angiogenesis mediated by activation of VEGF signaling pathway under ischemia/perfusion injury in vivo and in vitro. *Kidney Blood Press Res.* 35, 182–191. doi: 10.1159/000331054
- Liu, F., Wu, L., Wang, A., Xu, Y., Luo, X., Liu, X., et al. (2017). MicroRNA-138 attenuates epithelial-to-mesenchymal transition by targeting SOX4 in clear cell renal cell carcinoma. *Am. J. Transl. Res.* 9, 3611–3622.
- Liu, G., Ye, Z., Zhao, X., and Ji, Z. (2017). SP1-induced up-regulation of lncRNA SNHG14 as a ceRNA promotes migration and invasion of clear cell renal cell carcinoma by regulating N-WASP. Am. J. Cancer Res. 7, 2515–2525.
- Liu, J., Liu, B., Guo, Y., Chen, Z., Sun, W., Gao, W., et al. (2018). Key miRNAs and target genes played roles in the development of clear cell renal cell carcinoma. *Cancer Biomark*. 23, 279–290. doi: 10.3233/CBM-181558
- Liu, L., Li, Y., Liu, S., Duan, Q., Chen, L., Wu, T., et al. (2017). Downregulation of miR-193a-3p inhibits cell growth and migration in renal cell carcinoma by targeting PTEN. *Tumour Biol.* 39:1010428317711951. doi: 10.1177/ 1010428317711951
- Liu, Y., Nie, H., Zhang, K., Ma, D., Yang, G., Zheng, Z., et al. (2014). A feedback regulatory loop between HIF-1alpha and miR-21 in response to hypoxia in cardiomyocytes. *FEBS Lett.* 588, 3137–3146. doi: 10.1016/j.febslet.2014. 05.067
- Loginov, V. I., Beresneva, E. V., Kazubskaya, T. P., Braga, E. A., and Karpukhin, A. V. (2017). Methylation of 10 miRNA genes in clear cell renal cell carcinoma and their diagnostic value. *Cancer Urol.* 13, 27–33. doi: 10.17650/1726-9776-2017-13-3-27-33

- Loginov, V. I., Filippova, E. A., Kurevlev, S. V., Fridman, M. V., Burdennyy, A. M., and Braga, E. A. (2018a). Suppressive and Hypermethylated MicroRNAs in the Pathogenesis of Breast Cancer. *Russ. J. Genet.* 54, 770–787. doi: 10.1134/ S1022795418070086
- Loginov, V. I., Pronina, I. V., Burdennyy, A. M., Filippova, E. A., Kazubskaya, T. P., Kushlinsky, D. N., et al. (2018b). Novel miRNA genes deregulated by aberrant methylation in ovarian carcinoma are involved in metastasis. *Gene* 662, 28–36. doi: 10.1016/j.gene.2018.04.005
- Loginov, V. I., Rykov, S. V., Fridman, M. V., and Braga, E. A. (2015). Methylation of miRNA genes and oncogenesis. *Biochemistry* 80, 145–162. doi: 10.1134/ S0006297915020029
- Lokeshwar, S. D., Talukder, A., Yates, T. J., Hennig, M. J. P., Garcia-Roig, M., Lahorewala, S. S., et al. (2018). Molecular characterization of renal cell carcinoma: a potential three-MicroRNA prognostic signature. *Cancer Epidemiol. Biomarkers Prev.* 27, 464–472. doi: 10.1158/1055-9965.EPI-17-0700
- Long, Q. Z., Du, Y. F., Liu, X. G., Li, X., and He, D. L. (2015). miR-124 represses FZD5 to attenuate P-glycoprotein-mediated chemo-resistance in renal cell carcinoma. *Tumour Biol.* 36, 7017–7026. doi: 10.1007/s13277-015-3369-3
- Lopez-Serra, P., and Esteller, M. (2012). DNA methylation-associated silencing of tumor-suppressor microRNAs in cancer. Oncogene 31, 1609–1622. doi: 10. 1038/onc.2011.354
- Lou, N., Ruan, A. M., Qiu, B., Bao, L., Xu, Y. C., Zhao, Y., et al. (2017). miR-144-3p as a novel plasma diagnostic biomarker for clear cell renal cell carcinoma. *Urol. Oncol.* 35, 36.e–36.e14. doi: 10.1016/j.urolonc.2016.07.012.
- Lu, J., Wei, J. H., Feng, Z. H., Chen, Z. H., Wang, Y. Q., Huang, Y., et al. (2017). miR-106b-5p promotes renal cell carcinoma aggressiveness and stemcell-like phenotype by activating Wnt/beta-catenin signalling. *Oncotarget* 8, 21461–21471. doi: 10.18632/oncotarget.15591
- Machackova, T., Mlcochova, H., Stanik, M., Dolezel, J., Fedorko, M., Pacik, D., et al. (2016). MiR-429 is linked to metastasis and poor prognosis in renal cell carcinoma by affecting epithelial-mesenchymal transition. *Tumour Biol.* 37, 14653–14658. doi: 10.1007/s13277-016-5310-9
- Mahalingaiah, P. K., Ponnusamy, L., and Singh, K. P. (2015). Chronic oxidative stress leads to malignant transformation along with acquisition of stem cell characteristics, and epithelial to mesenchymal transition in human renal epithelial cells. J. Cell. Physiol. 230, 1916–1928. doi: 10.1002/jcp.24922
- Malouf, G. G., Flippot, R., and Khayat, D. (2016). Therapeutic strategies for patients with metastatic renal cell carcinoma in whom first-line vascular endothelial growth factor receptor-directed therapies fail. J. Oncol. Pract. 12, 412–420. doi: 10.1200/JOP.2016.011809
- McCormick, R. I., Blick, C., Ragoussis, J., Schoedel, J., Mole, D. R., Young, A. C., et al. (2013). miR-210 is a target of hypoxia-inducible factors 1 and 2 in renal cancer, regulates ISCU and correlates with good prognosis. *Br. J. Cancer* 108, 1133–1142. doi: 10.1038/bjc.2013.56
- Mens, M. M. J., and Ghanbari, M. (2018). Cell Cycle Regulation of Stem Cells by MicroRNAs. Stem Cell Rev. 14, 309–322. doi: 10.1007/s12015-018-9808-y
- Mickley, A., Kovaleva, O., Kzhyshkowska, J., and Gratchev, A. (2015). Molecular and immunologic markers of kidney cancer-potential applications in predictive, preventive and personalized medicine. *EPMA J.* 6:20. doi: 10.1186/s13167-015-0042-2
- Mikhaylova, O., Stratton, Y., Hall, D., Kellner, E., Ehmer, B., Drew, A. F., et al. (2012). VHL-regulated MiR-204 suppresses tumor growth through inhibition of LC3B-mediated autophagy in renal clear cell carcinoma. *Cancer Cell* 21, 532–546. doi: 10.1016/j.ccr.2012.02.019
- Morais, M., Dias, F., Teixeira, A. L., and Medeiros, R. (2017). MicroRNAs and altered metabolism of clear cell renal cell carcinoma: potential role as aerobic glycolysis biomarkers. *Biochim. Biophys. Acta Gen. Subj.* 1861, 2175–2185. doi: 10.1016/j.bbagen.2017.05.028
- Morris, M. R., and Latif, F. (2017). The epigenetic landscape of renal cancer. *Nat. Rev. Nephrol.* 13, 47–60. doi: 10.1038/nrneph.2016.168
- Müller, S., and Nowak, K. (2014). Exploring the miRNA-mRNA regulatory network in clear cell renal cell carcinomas by next-generation sequencing expression profiles. *Biomed. Res. Int.* 2014:948408. doi: 10.1155/2014/ 948408
- Nakada, C., Tsukamoto, Y., Matsuura, K., Nguyen, T. L., Hijiya, N., Uchida, T., et al. (2011). Overexpression of miR-210, a downstream target of HIF1alpha, causes centrosome amplification in renal carcinoma cells. *J. Pathol.* 224, 280–288. doi: 10.1002/path.2860

- Neal, C. S., Michael, M. Z., Rawlings, L. H., Van der Hoek, M. B., and Gleadle, J. M. (2010). The VHL-dependent regulation of microRNAs in renal cancer. *BMC Med.* 8:64. doi: 10.1186/1741-7015-8-64
- Ni, D., Ma, X., Li, H. Z., Gao, Y., Li, X. T., Zhang, Y., et al. (2014). Downregulation of FOXO3a promotes tumor metastasis and is associated with metastasis-free survival of patients with clear cell renal cell carcinoma. *Clin. Cancer Res.* 20, 1779–1790. doi: 10.1158/1078-0432.CCR-13-1687
- Nofech-Mozes, R., Khella, H. W., Scorilas, A., Youssef, L., Krylov, S. N., Lianidou, E., et al. (2016). MicroRNA-194 is a marker for good prognosis in clear cell renal cell carcinoma. *Cancer Med.* 5, 656–664. doi: 10.1002/ cam4.631
- Okato, A., Arai, T., Yamada, Y., Sugawara, S., Koshizuka, K., Fujimura, L., et al. (2017). Dual strands of Pre-miR-149 inhibit cancer cell migration and invasion through targeting FOXM1 in renal cell carcinoma. *Int. J. Mol. Sci.* 18:E1969. doi: 10.3390/ijms18091969
- Oliveira, R. C., Ivanovic, R. F., Leite, K. R. M., Viana, N. I., Pimenta, R. C. A., Junior, J. P., et al. (2017). Expression of micro-RNAs and genes related to angiogenesis in ccRCC and associations with tumor characteristics. *BMC Urol.* 17:113. doi: 10.1186/s12894-017-0306-3
- Ornitz, D. M., and Itoh, N. (2015). The Fibroblast Growth Factor signaling pathway. Wiley Interdiscip. Rev. Dev. Biol. 4, 215–266. doi: 10.1002/wdev.176
- Pan, Y., Hu, J., Ma, J., Qi, X., Zhou, H., Miao, X., et al. (2018a). MiR-193a-3p and miR-224 mediate renal cell carcinoma progression by targeting alpha-2,3sialyltransferase IV and the phosphatidylinositol 3 kinase/Akt pathway. *Mol. Carcinog.* 57, 1067–1077. doi: 10.1002/mc.22826
- Pan, Y., Wu, Y., Hu, J., Shan, Y., Ma, J., Ma, H., et al. (2018b). Long noncoding RNA HOTAIR promotes renal cell carcinoma malignancy through alpha-2, 8-sialyltransferase 4 by sponging microRNA-124. *Cell Prolif.* 51:e12507. doi: 10.1111/cpr.12507
- Pan, Y. J., Wei, L. L., Wu, X. J., Huo, F. C., Mou, J., and Pei, D. S. (2017). MiR-106a-5p inhibits the cell migration and invasion of renal cell carcinoma through targeting PAK5. *Cell Death Dis.* 8:e3155. doi: 10.1038/cddis.2017.561
- Pereyaslova, E., Loginov, V., Pronina, I., Dmitriev, A., Burdennyy, A., Kazubskaya, T. P., et al. (2016). Hypermethylation of MIR-129-2, MIR-203, and MIR-107 microRNA genes is associated with metastasis of clear cell renal cell cancer. *FEBS J.* 283, 251–251.
- Piletic, K., and Kunej, T. (2016). MicroRNA epigenetic signatures in human disease. Arch. Toxicol. 90, 2405–2419.
- Piva, F., Giulietti, M., Santoni, M., Occhipinti, G., Scarpelli, M., Lopez-Beltran, A., et al. (2016). Epithelial to mesenchymal transition in renal cell carcinoma: implications for cancer therapy. *Mol. Diagn. Ther.* 20, 111–117. doi: 10.1007/ s40291-016-0192-5
- Pronina, I. V., Klimov, E. A., Burdennyy, A. M., Beresneva, E. V., Fridman, M. V., Ermilova, V. D., et al. (2017a). Methylation of the genes for the MicroRNAs miR-129-2 and miR-9-1, changes in their expression, and activation of their potential target genes in clear cell renal cell carcinoma. *Mol. Biol.* 51, 61–71. doi: 10.1134/S0026893316060169
- Pronina, I. V., Loginov, V. I., Burdennyy, A. M., Fridman, M. V., Senchenko, V. N., Kazubskaya, T. P., et al. (2017b). DNA methylation contributes to deregulation of 12 cancer-associated microRNAs and breast cancer progression. *Gene* 604, 1–8. doi: 10.1016/j.gene.2016.12.018
- Puente, J., Lainez, N., Duenas, M., Mendez-Vidal, M. J., Esteban, E., Castellano, D., et al. (2017). Novel potential predictive markers of sunitinib outcomes in longterm responders versus primary refractory patients with metastatic clear-cell renal cell carcinoma. *Oncotarget* 8, 30410–30421. doi: 10.18632/oncotarget. 16494
- Qin, Z., Wei, X., Jin, N., Wang, Y., Zhao, R., Hu, Y., et al. (2018). MiR-199a targeting ROCK1 to affect kidney cell proliferation, invasion and apoptosis. *Artif. Cells Nanomed. Biotechnol.* 46, 1920–1925. doi: 10.1080/21691401.2017. 1396224
- Ran, L., Liang, J., Deng, X., and Wu, J. (2017). miRNAs in Prediction of Prognosis in Clear Cell Renal Cell Carcinoma. *Biomed. Res. Int.* 2017:4832931. doi: 10. 1155/2017/4832931
- Randall, J. M., Millard, F., and Kurzrock, R. (2014). Molecular aberrations, targeted therapy, and renal cell carcinoma: current state-of-the-art. *Cancer Metastasis Rev.* 33, 1109–1124.
- Reddy, K. B. (2015). MicroRNA (miRNA) in cancer. Cancer Cell Int. 15:38. doi: 10.1186/s12935-015-0185-1

Braga et al.

- Redova, M., Poprach, A., Nekvindova, J., Iliev, R., Radova, L., Lakomy, R., et al. (2012). Circulating miR-378 and miR-451 in serum are potential biomarkers for renal cell carcinoma. *J. Transl. Med.* 10:55. doi: 10.1186/1479-5876-10-55
- Redova, M., Svoboda, M., and Slaby, O. (2011). MicroRNAs and their target gene networks in renal cell carcinoma. *Biochem. Biophys. Res. Commun.* 405, 153–156. doi: 10.1016/j.bbrc.2011.01.019
- Reustle, A., Fisel, P., Renner, O., Buttner, F., Winter, S., Rausch, S., et al. (2018). Characterization of the breast cancer resistance protein (BCRP/ABCG2) in clear cell renal cell carcinoma. *Int. J. Cancer.* 143, 3181–3193. doi: 10.1002/ijc. 31741
- Roskoski, R. Jr. (2017). Vascular endothelial growth factor (VEGF) and VEGF receptor inhibitors in the treatment of renal cell carcinomas. *Pharmacol. Res.* 120, 116–132. doi: 10.1016/j.phrs.2017.03.010
- Rydzanicz, M., Wrzesinski, T., Bluyssen, H. A., and Wesoly, J. (2013). Genomics and epigenomics of clear cell renal cell carcinoma: recent developments and potential applications. *Cancer Lett.* 341, 111–126. doi: 10.1016/j.canlet.2013. 08.006
- Rykov, S. V., Khodyrev, D. S., Pronina, I. V., Kazubskaya, T. P., Loginov, V. I., and Braga, E. A. (2013). Novel miRNA genes methylated in lung tumors. *Russ. J. Genet.* 49, 782–786. doi: 10.1134/S1022795413070119
- Samaan, S., Khella, H. W., Girgis, A., Scorilas, A., Lianidou, E., Gabril, M., et al. (2015). miR-210 is a prognostic marker in clear cell renal cell carcinoma. J. Mol. Diagn. 17, 136–144. doi: 10.1016/j.jmoldx.2014.10.005
- Sampson, V. B., David, J. M., Puig, I., Patil, P. U., de Herreros, A. G., Thomas, G. V., et al. (2014). Wilms' tumor protein induces an epithelial-mesenchymal hybrid differentiation state in clear cell renal cell carcinoma. *PLoS One* 9:e102041. doi: 10.1371/journal.pone.0102041
- Sato, K., Tsuchihara, K., Fujii, S., Sugiyama, M., Goya, T., Atomi, Y., et al. (2007). Autophagy is activated in colorectal cancer cells and contributes to the tolerance to nutrient deprivation. *Cancer Res.* 67, 9677–9684. doi: 10.1158/0008-5472. CAN-07-1462
- Schanza, L. M., Seles, M., Stotz, M., Fosselteder, J., Hutterer, G. C., Pichler, M., et al. (2017). MicroRNAs associated with von hippel-lindau pathway in renal cell carcinoma: a comprehensive review. *Int. J. Mol. Sci.* 18:E2495. doi: 10.3390/ ijms18112495
- Schiffgen, M., Schmidt, D. H., von Rucker, A., Muller, S. C., and Ellinger, J. (2013). Epigenetic regulation of microRNA expression in renal cell carcinoma. *Biochem. Biophys. Res. Commun.* 436, 79–84. doi: 10.1016/j.bbrc.2013.05.061
- Seki, H., Hayashida, T., Jinno, H., Hirose, S., Sakata, M., Takahashi, M., et al. (2012). HOXB9 expression promoting tumor cell proliferation and angiogenesis is associated with clinical outcomes in breast cancer patients. *Ann. Surg. Oncol.* 19, 1831–1840. doi: 10.1245/s10434-012-2295-5
- Serocki, M., Bartoszewska, S., Janaszak-Jasiecka, A., Ochocka, R. J., Collawn, J. F., and Bartoszewski, R. (2018). miRNAs regulate the HIF switch during hypoxia: a novel therapeutic target. *Angiogenesis* 21, 183–202. doi: 10.1007/s10456-018-9600-2
- Shi, L., Li, X., Wu, Z., Li, X., Nie, J., Guo, M., et al. (2018). DNA methylation-mediated repression of miR-181a/135a/302c expression promotes the microsatellite-unstable colorectal cancer development and 5-FU resistance via targeting PLAG1. *J. Genet. Genomics* 45, 205–214. doi: 10.1016/j.jgg.2018. 04.003
- Shimoyamada, H., Yazawa, T., Sato, H., Okudela, K., Ishii, J., Sakaeda, M., et al. (2010). Early growth response-1 induces and enhances vascular endothelial growth factor-A expression in lung cancer cells. *Am J. Pathol.* 177, 70–83. doi: 10.2353/ajpath.2010.091164
- Shinmei, S., Sakamoto, N., Goto, K., Sentani, K., Anami, K., Hayashi, T., et al. (2013). MicroRNA-155 is a predictive marker for survival in patients with clear cell renal cell carcinoma. *Int. J. Urol.* 20, 468–477. doi: 10.1111/j.1442-2042. 2012.03182.x
- Song, T., Zhang, X., Wang, C., Wu, Y., Cai, W., Gao, J., et al. (2011). MiR-138 suppresses expression of hypoxia-inducible factor 1alpha (HIF-1alpha) in clear cell renal cell carcinoma 786-O cells. *Asian Pac. J. Cancer Prev.* 12, 1307–1311.
- Tang, K., and Xu, H. (2015). Prognostic value of meta-signature miRNAs in renal cell carcinoma: an integrated miRNA expression profiling analysis. *Sci. Rep.* 5:10272. doi: 10.1038/srep10272
- Toraih, E. A., Ibrahiem, A. T., Fawzy, M. S., Hussein, M. H., Al-Qahtani, S. A. M., and Shaalan, A. A. M. (2017). MicroRNA-34a: a key regulator in the hallmarks

of renal cell carcinoma. Oxid. Med. Cell. Longev. 2017:3269379. doi: 10.1155/2017/3269379

- Torres-Ferreira, J., Ramalho-Carvalho, J., Gomez, A., Menezes, F. D., Freitas, R., Oliveira, J., et al. (2017). MiR-193b promoter methylation accurately detects prostate cancer in urine sediments and miR-34b/c or miR-129-2 promoter methylation define subsets of clinically aggressive tumors. *Mol. Cancer* 16:26. doi: 10.1186/s12943-017-0604-0
- Uchida, T., Rossignol, F., Matthay, M. A., Mounier, R., Couette, S., Clottes, E., et al. (2004). Prolonged hypoxia differentially regulates hypoxia-inducible factor (HIF)-1alpha and HIF-2alpha expression in lung epithelial cells: implication of natural antisense HIF-1alpha. *J. Biol. Chem.* 279, 14871–14878. doi: 10.1074/jbc. M400461200
- Umezu, T., Tadokoro, H., Azuma, K., Yoshizawa, S., Ohyashiki, K., and Ohyashiki, J. H. (2014). Exosomal miR-135b shed from hypoxic multiple myeloma cells enhances angiogenesis by targeting factor-inhibiting HIF-1. *Blood* 124, 3748–3757. doi: 10.1182/blood-2014-05-576116
- Valera, V. A., Walter, B. A., Linehan, W. M., and Merino, M. J. (2011). Regulatory effects of microRNA-92 (miR-92) on VHL gene expression and the hypoxic activation of miR-210 in clear cell renal cell carcinoma. J. Cancer 2, 515–526.
- Varachev, V., Loginov, V., Pronina, I., Khodyrev, D., Kazubskaya, T., Morozov, A., et al. (2018). Hypermethylated tumor suppressor microRNAs as novel markers of clear cell renal cell carcinoma. *FEBS Open Bio* 8, 304–305.
- Vasudev, N. S., Selby, P. J., and Banks, R. E. (2012). Renal cancer biomarkers: the promise of personalized care. *BMC Med.* 10:112. doi: 10.1186/1741-7015-10-112
- Vergho, D., Kneitz, S., Rosenwald, A., Scherer, C., Spahn, M., Burger, M., et al. (2014). Combination of expression levels of miR-21 and miR-126 is associated with cancer-specific survival in clear-cell renal cell carcinoma. *BMC Cancer* 14:25. doi: 10.1186/1471-2407-14-25
- Vogt, M., Munding, J., Gruner, M., Liffers, S. T., Verdoodt, B., Hauk, J., et al. (2011). Frequent concomitant inactivation of miR-34a and miR-34b/c by CpG methylation in colorectal, pancreatic, mammary, ovarian, urothelial, and renal cell carcinomas and soft tissue sarcomas. *Virchows Arch.* 458, 313–322. doi: 10.1007/s00428-010-1030-5
- Wang, C., Cai, L., Liu, J., Wang, G., Li, H., Wang, X., et al. (2017). MicroRNA-30a-5p inhibits the growth of renal cell carcinoma by modulating GRP78 expression. *Cell Physiol. Biochem.* 43, 2405–2419. doi: 10.1159/00048 4394
- Wang, C., Hu, J., Lu, M., Gu, H., Zhou, X., Chen, X., et al. (2015). A panel of five serum miRNAs as a potential diagnostic tool for early-stage renal cell carcinoma. *Sci. Rep.* 5:7610. doi: 10.1038/srep07610
- Wang, C., Wu, C., Yang, Q., Ding, M., Zhong, J., Zhang, C. Y., et al. (2016). miR-28-5p acts as a tumor suppressor in renal cell carcinoma for multiple antitumor effects by targeting RAP1B. *Oncotarget* 7, 73888–73902. doi: 10. 18632/oncotarget.12516
- Wang, D., Zhu, C., Zhang, Y., Zheng, Y., Ma, F., Su, L., et al. (2017). MicroRNA-30e-3p inhibits cell invasion and migration in clear cell renal cell carcinoma by targeting Snail1. Oncol. Lett. 13, 2053–2058. doi: 10.3892/ol.2017. 5690
- Wang, G., Chen, L., Meng, J., Chen, M., Zhuang, L., and Zhang, L. (2013). Overexpression of microRNA-100 predicts an unfavorable prognosis in renal cell carcinoma. *Int. Urol. Nephrol.* 45, 373–379. doi: 10.1007/s11255-012-0374-y
- Wang, M., Gao, H., Qu, H., Li, J., Liu, K., and Han, Z. (2018). MiR-137 suppresses tumor growth and metastasis in clear cell renal cell carcinoma. *Pharmacol. Rep.* 70, 963–971. doi: 10.1016/j.pharep.2018.04.006
- Wang, Z., Qin, C., Zhang, J., Han, Z., Tao, J., Cao, Q., et al. (2017). MiR-122 promotes renal cancer cell proliferation by targeting Sprouty2. *Tumour Biol.* 39:1010428317691184. doi: 10.1177/1010428317691184
- Wang, Z., Zhang, Z., Zhang, C., and Xu, Y. (2018). Identification of potential pathogenic biomarkers in clear cell renal cell carcinoma. *Oncol. Lett.* 15, 8491–8499. doi: 10.3892/ol.2018.8398
- Wei, H., Ke, H. L., Lin, J., Shete, S., Wood, C. G., and Hildebrandt, M. A. (2014). MicroRNA target site polymorphisms in the VHL-HIF1alpha pathway predict renal cell carcinoma risk. *Mol. Carcinog.* 53, 1–7. doi: 10.1002/mc.21917
- Wotschofsky, Z., Busch, J., Jung, M., Kempkensteffen, C., Weikert, S., Schaser, K. D., et al. (2013). Diagnostic and prognostic potential of differentially expressed miRNAs between metastatic and non-metastatic renal cell carcinoma

at the time of nephrectomy. *Clin. Chim. Acta* 416, 5–10. doi: 10.1016/j.cca.2012. 11.010

- Wotschofsky, Z., Gummlich, L., Liep, J., Stephan, C., Kilic, E., Jung, K., et al. (2016). Integrated microRNA and mRNA signature associated with the transition from the locally confined to the metastasized clear cell renal cell carcinoma exemplified by miR-146-5p. *PLoS One* 11:e0148746. doi: 10.1371/journal.pone. 0148746
- Wu, A., Wu, K., Li, M., Bao, L., Shen, X., Li, S., et al. (2015). Upregulation of microRNA-492 induced by epigenetic drug treatment inhibits the malignant phenotype of clear cell renal cell carcinoma in vitro. *Mol. Med. Rep.* 12, 1413–1420. doi: 10.3892/mmr.2015.3550
- Wu, C., Jin, B., Chen, L., Zhuo, D., Zhang, Z., Gong, K., et al. (2013). MiR-30d induces apoptosis and is regulated by the Akt/FOXO pathway in renal cell carcinoma. *Cell Signal.* 25, 1212–1221. doi: 10.1016/j.cellsig.2013. 01.028
- Wu, D., Pan, H., Zhou, Y., Zhou, J., Fan, Y., and Qu, P. (2014). microRNA-133b downregulation and inhibition of cell proliferation, migration and invasion by targeting matrix metallopeptidase-9 in renal cell carcinoma. *Mol. Med. Rep.* 9, 2491–2498. doi: 10.3892/mmr.2014.2116
- Wu, S. Y., Rupaimoole, R., Shen, F., Pradeep, S., Pecot, C. V., Ivan, C., et al. (2016). A miR-192-EGR1-HOXB9 regulatory network controls the angiogenic switch in cancer. *Nat. Commun.* 7:11169. doi: 10.1038/ncomms 11169
- Xiang, C., Cui, S. P., and Ke, Y. (2016). MiR-144 inhibits cell proliferation of renal cell carcinoma by targeting MTOR. J. Huazhong Univ. Sci. Technolog. Med. Sci. 36, 186–192. doi: 10.1007/s11596-016-1564-0
- Xiang, W., He, J., Huang, C., Chen, L., Tao, D., Wu, X., et al. (2015). miR-106b-5p targets tumor suppressor gene SETD2 to inactive its function in clear cell renal cell carcinoma. *Oncotarget* 6, 4066–4079. doi: 10.18632/oncotarget. 2926
- Xiao, H., Li, H., Yu, G., Xiao, W., Hu, J., Tang, K., et al. (2014). MicroRNA-10b promotes migration and invasion through KLF4 and HOXD10 in human bladder cancer. Oncol. Rep. 31, 1832–1838. doi: 10.3892/or.2014. 3048
- Xiao, W., Lou, N., Ruan, H., Bao, L., Xiong, Z., Yuan, C., et al. (2017). Mir-144-3p promotes cell proliferation, metastasis, sunitinib resistance in clear cell renal cell carcinoma by downregulating ARID1A. *Cell. Physiol. Biochem.* 43, 2420–2433. doi: 10.1159/000484395
- Xiao, X., Tang, C., Xiao, S., Fu, C., and Yu, P. (2013). Enhancement of proliferation and invasion by MicroRNA-590-5p via targeting PBRM1 in clear cell renal carcinoma cells. Oncol. Res. 20, 537–544. doi: 10.3727/ 096504013X13775486749335
- Xing, T., and He, H. (2016). Epigenomics of clear cell renal cell carcinoma: mechanisms and potential use in molecular pathology. *Chin. J. Cancer Res.* 28, 80–91. doi: 10.3978/j.issn.1000-9604.2016.02.09
- Xu, M., Gu, M., Zhang, K., Zhou, J., Wang, Z., and Da, J. (2015). miR-203 inhibition of renal cancer cell proliferation, migration and invasion by targeting of FGF2. *Diagn. Pathol.* 10:24. doi: 10.1186/s13000-015-0255-7
- Xu, X., Liu, C., and Bao, J. (2017). Hypoxia-induced hsa-miR-101 promotes glycolysis by targeting TIGAR mRNA in clear cell renal cell carcinoma. *Mol. Med. Rep.* 15, 1373–1378. doi: 10.3892/mmr.2017.6139
- Xu, X., Wu, J., Li, S., Hu, Z., Xu, X., Zhu, Y., et al. (2014). Downregulation of microRNA-182-5p contributes to renal cell carcinoma proliferation via activating the AKT/FOXO3a signaling pathway. *Mol. Cancer* 13:109. doi: 10. 1186/1476-4598-13-109
- Xu, X. W., Li, S., Yin, F., and Qin, L. L. (2018). Expression of miR-205 in renal cell carcinoma and its association with clinicopathological features and prognosis. *Eur. Rev. Med. Pharmacol. Sci.* 22, 662–670. doi: 10.26355/eurrev_201802_ 14291
- Xu, Y., Zhu, J., Lei, Z., Wan, L., Zhu, X., Ye, F., et al. (2015). Expression and functional role of miR-29b in renal cell carcinoma. *Int. J. Clin. Exp. Pathol.* 8, 14161–14170.
- Yadav, S., Khandelwal, M., Seth, A., Saini, A. K., Dogra, P. N., and Sharma, A. (2017). Serum microRNA expression profiling: potential diagnostic implications of a panel of serum microRNAs for clear cell renal cell cancer. *Urology* 104, 64–69. doi: 10.1016/j.urology.2017.03.013
- Yamamura, S., Saini, S., Majid, S., Hirata, H., Ueno, K., Chang, I., et al. (2012). MicroRNA-34a suppresses malignant transformation by targeting

c-Myc transcriptional complexes in human renal cell carcinoma. *Carcinogenesis* 33, 294–300. doi: 10.1093/carcin/bgr286

- Yamasaki, T., Seki, N., Yoshino, H., Itesako, T., Yamada, Y., Tatarano, S., et al. (2013). Tumor-suppressive microRNA-1291 directly regulates glucose transporter 1 in renal cell carcinoma. *Cancer Sci.* 104, 1411–1419. doi: 10.1111/ cas.12240
- Yang, F., Zhang, H., Mei, Y., and Wu, M. (2014). Reciprocal regulation of HIFlalpha and lincRNA-p21 modulates the Warburg effect. *Mol. Cell* 53, 88–100. doi: 10.1016/j.molcel.2013.11.004
- Yin, H., Wang, X., Zhang, X., Wang, Y., Zeng, Y., Xiong, Y., et al. (2018). Integrated analysis of long noncoding RNA associated-competing endogenous RNA as prognostic biomarkers in clear cell renal carcinoma. *Cancer Sci.* 109, 3336–3349. doi: 10.1111/cas.13778
- Yoshino, H., Enokida, H., Itesako, T., Kojima, S., Kinoshita, T., Tatarano, S., et al. (2013). Tumor-suppressive microRNA-143/145 cluster targets hexokinase-2 in renal cell carcinoma. *Cancer Sci.* 104, 1567–1574. doi: 10.1111/cas.12280
- Yoshino, H., Yonemori, M., Miyamoto, K., Tatarano, S., Kofuji, S., Nohata, N., et al. (2017). microRNA-210-3p depletion by CRISPR/Cas9 promoted tumorigenesis through revival of TWIST1 in renal cell carcinoma. *Oncotarget* 8, 20881–20894. doi: 10.18632/oncotarget.14930
- Yu, G., Li, H., Wang, J., Gumireddy, K., Li, A., Yao, W., et al. (2014). miRNA-34a suppresses cell proliferation and metastasis by targeting CD44 in human renal carcinoma cells. J. Urol. 192, 1229–1237. doi: 10.1016/j.juro.2014.05.094
- Zaman, M. S., Thamminana, S., Shahryari, V., Chiyomaru, T., Deng, G., Saini, S., et al. (2012). Inhibition of PTEN gene expression by oncogenic miR-23b-3p in renal cancer. *PLoS One* 7:e50203. doi: 10.1371/journal.pone.0050203
- Zare, M., Bastami, M., Solali, S., and Alivand, M. R. (2018). Aberrant miRNA promoter methylation and EMT-involving miRNAs in breast cancer metastasis: Diagnosis and therapeutic implications. *J. Cell. Physiol.* 233, 3729–3744. doi: 10.1002/jcp.26116
- Zhang, C., Mo, R., Yin, B., Zhou, L., Liu, Y., and Fan, J. (2014). Tumor suppressor microRNA-34a inhibits cell proliferation by targeting Notch1 in renal cell carcinoma. Oncol. Lett. 7, 1689–1694. doi: 10.3892/ol.2014.1931
- Zhang, H., and Li, H. (2016). miR-137 inhibits renal cell carcinoma growth in vitro and in vivo. *Oncol. Lett.* 12, 715–720. doi: 10.3892/ol.2016.4616
- Zhang, S., Guo, Z., Xu, J., Wang, J., Zhang, J., Cui, L., et al. (2017). miR-502mediated histone methyltransferase SET8 expression is associated with clear cell renal cell carcinoma risk. Oncol. Lett. 14, 7131–7138. doi: 10.3892/ol.2017.7115
- Zhang, S., Qian, J., Cao, Q., Li, P., Wang, M., Wang, J., et al. (2014). A potentially functional polymorphism in the promoter region of miR-34b/c is associated with renal cell cancer risk in a Chinese population. *Mutagenesis* 29, 149–154. doi: 10.1093/mutage/geu001
- Zhang, W., Ni, M., Su, Y., Wang, H., Zhu, S., Zhao, A., et al. (2018). MicroRNAs in serum exosomes as potential biomarkers in clear-cell renal cell carcinoma. *Eur. Urol. Focus* 4, 412–419. doi: 10.1016/j.euf.2016.09.007
- Zhang, X., Xing, N. D., Lai, C. J., Liu, R., Jiao, W., Wang, J., et al. (2018). MicroRNA-375 suppresses the tumor aggressive phenotypes of clear cell renal cell carcinomas through regulating YWHAZ. *Chin. Med. J.* 131, 1944–1950. doi: 10.4103/0366-6999.238153
- Zhang, Z., Sun, H., Dai, H., Walsh, R. M., Imakura, M., Schelter, J., et al. (2009). MicroRNA miR-210 modulates cellular response to hypoxia through the MYC antagonist MNT. *Cell Cycle* 8, 2756–2768. doi: 10.4161/cc.8.17.9387
- Zhao, J., Lei, T., Xu, C., Li, H., Ma, W., Yang, Y., et al. (2013). MicroRNA-187, down-regulated in clear cell renal cell carcinoma and associated with lower survival, inhibits cell growth and migration though targeting B7-H3. *Biochem. Biophys. Res. Commun.* 438, 439–444. doi: 10.1016/j.bbrc.2013. 07.095
- Zhao, J. J., Chen, P. J., Duan, R. Q., Li, K. J., Wang, Y. Z., and Li, Y. (2014). Upregulation of miR-630 in clear cell renal cell carcinoma is associated with lower overall survival. *Int. J. Clin. Exp. Pathol.* 7, 3318–3323.
- Zhao, S., Wang, Y., Luo, M., Cui, W., Zhou, X., and Miao, L. (2018). Long noncoding RNA small nucleolar RNA host gene 1 (SNHG1) promotes renal cell carcinoma progression and metastasis by negatively regulating miR-137. *Med. Sci. Monit.* 24, 3824–3831. doi: 10.12659/MSM. 910866
- Zhao, X., Zhao, Z., Xu, W., Hou, J., and Du, X. (2015). Down-regulation of miR-497 is associated with poor prognosis in renal cancer. *Int. J. Clin. Exp. Pathol.* 8, 758–764.

- Zhao, Z., Zhang, M., Duan, X., Chen, Y., Li, E., Luo, L., et al. (2018). TRPM7 regulates AKT/FOXO1-dependent tumor growth and is an independent prognostic indicator in renal cell carcinoma. *Mol. Cancer Res.* 16, 1013–1023. doi: 10.1158/1541-7786.MCR-17-0767
- Zhou, L., Li, Z., Pan, X., Lai, Y., Quan, J., Zhao, L., et al. (2018). Identification of miR-18a-5p as an oncogene and prognostic biomarker in RCC. Am. J. Transl. Res. 10, 1874–1886.
- Zhu, S., Huang, Y., and Su, X. (2016). Mir-451 correlates with prognosis of renal cell carcinoma patients and inhibits cellular proliferation of renal cell carcinoma. *Med. Sci. Monit.* 22, 183–190.

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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