



Draft Genome Sequence of *Streptomyces* sp. Strain NL15-2K, a Degrader of Lignin-Derived Aromatic Compounds, Isolated from Forest Soil

Motohiro Nishimura,^a Susumu Kawakami,^a Hideaki Otsuka^a

^aDepartment of Pharmaceutical Chemistry, Faculty of Pharmacy, Yasuda Women's University, Hiroshima, Japan

ABSTRACT *Streptomyces* sp. strain NL15-2K is a degrader of lignin-derived aromatic compounds and was isolated from a forest soil sample. Here, we report the draft genome sequence of this strain and its annotation. This genome of 12,072,023 bp exhibits a GC content of 70.32% and encodes 10,874 predicted proteins and 75 RNAs.

Most bacteria of the genus *Streptomyces* are found in the soil, where they play a critical role in the global carbon cycle (1). This role can be implemented because streptomycetes have evolved complex and efficient enzymatic systems that catabolize diverse organic substances, such as lignin and lignin-derived aromatic compounds (2, 3). Therefore, *Streptomyces* species and their enzymes are promising as biocatalysts in the production of commercially valuable compounds, such as vanillin, from inexpensive plant constituents (4). Strain NL15-2K was isolated from a forest soil sample on the campus of the University of British Columbia, Vancouver, Canada, by screening for bacteria capable of catabolizing lignin-derived aromatic compounds (5). This strain was identified as a *Streptomyces* species by 16S rRNA gene analysis (5).

Streptomyces sp. strain NL15-2K was cultivated for 2 days in yeast extract-malt extract (YEME) medium (6) supplemented with 17% sucrose and 0.5% glycine at 30°C. Genomic DNA was extracted and purified using the Genomic-tip 100/G kit (Qiagen), according to the manufacturer's protocol. DNA library preparation (paired-end 2×100 -bp reads) and sequencing were performed on the Illumina HiSeq 2500 (CASAVA version 1.8.2) sequencing platform by Hokkaido System Science Co., Ltd. (Hokkaido, Japan). Shotgun sequencing generated 24,114,726 high-quality paired-end reads. All reads were cleaned up using cutadapt version 1.1 (7) and Trimmomatic version 0.32 (8) by trimming adapter sequences and removing low-quality reads, respectively. The resulting 22,713,654 reads with a mean size of 98 bp were assembled into the genome sequence using Velvet version 1.2.08 (9), and the gaps were closed using Platanus version 1.2.4 (10). Scaffolding was performed using MeDuSa version 1.6 (11), with the Streptomyces lincolnensis NRRL 2936 genome (GenBank accession number CP016438) used as a guide for alignment. The draft genome was annotated using the RAST server (http://rast.nmpdr.org/) (12), with an additional annotation being conducted using antiSMASH version 4.1.0 (13).

The genome size of *Streptomyces* sp. NL15-2K was 12,072,023 bp, with a GC content of 70.3%, and it comprised 292 scaffolds with an N_{50} value of 103,610 bp. Gene prediction and annotation revealed that the *Streptomyces* sp. NL15-2K genome comprises 10,874 protein-coding sequences and 75 RNA-coding sequences, including 4 rRNAs and 71 tRNAs. The antiSMASH algorithm predicted gene clusters for the biosynthesis of coelichelin, alkylresorcinol, gamma-butyrolactone, albaflavenone, desferrioxamine B, ectoine, endophenazines, indigoidine, and GE37468. Moreover, the genome contained genes involved in the catabolism of lignin-derived aromatic compounds, including *pcaHG* (14) and *catA* (15), which encode protocatechuate 3,4-dioxygenase

Citation Nishimura M, Kawakami S, Otsuka H. 2019. Draft genome sequence of *Streptomyces* sp. strain NL15-2K, a degrader of lignin-derived aromatic compounds, isolated from forest soil. Microbiol Resour Announc 8:e01456-18. https://doi.org/10.1128/MRA.01456-18.

Editor David Rasko, University of Maryland School of Medicine

Copyright © 2019 Nishimura et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Motohiro Nishimura, m-nishi@yasuda-u.ac.jp.

Received 22 October 2018 Accepted 7 February 2019 Published 7 March 2019 and catechol 1,2-dioxygenase, respectively, and play a role in cleavage of the aromatic ring. Thus, this study provides valuable genetic information required to understand the catabolism of lignin-derived aromatic compounds in strain NL15-2K and to develop biocatalysts for producing valuable compounds from inexpensive plant constituents.

Data availability. The draft genome sequence of *Streptomyces* sp. NL15-2K has been deposited in DDBJ/ENA/GenBank under accession numbers BHXA01000001 to BHXA01000292. The raw sequencing reads have been submitted to the DDBJ/Sequence Read Archive under the accession number DRA007948.

ACKNOWLEDGMENTS

We are grateful to Julian Davies, Department of Microbiology and Immunology, University of British Columbia, Canada, for the generous gift of *Streptomyces* sp. NL15-2K.

The genome sequencing of *Streptomyces* sp. NL15-2K was supported by the Hokkaido System Science Co., Ltd. (Sapporo, Hokkaido, Japan). This study was also supported by Yasuda Women's University.

REFERENCES

- Book AJ, Lewin GR, McDonald BR, Takasuka TE, Doering DT, Adams AS, Blodgett JAV, Clardy J, Raffa KF, Fox BG, Currie CR. 2014. Cellulolytic *Streptomyces* strains associated with herbivorous insects share a phylogenetically linked capacity to degrade lignocellulose. Appl Environ Microbiol 80:4692–4701. https://doi.org/10.1128/AEM.01133-14.
- Crawford DL. 1978. Lignocellulose decomposition by selected Streptomyces strains. Appl Environ Microbiol 35:1041–1045.
- 3. Zimmermann W. 1990. Degradation of lignin by bacteria. J Biotechnol 13:119–130. https://doi.org/10.1016/0168-1656(90)90098-V.
- Kashiwagi N, Ogino C, Kondo A. 2017. Production of chemicals and proteins using biomass-derived substrates from a *Streptomyces* host. Bioresour Technol 245:1655–1663. https://doi.org/10.1016/j.biortech.2017.06.001.
- Nishimura M, Ooi O, Davies J. 2006. Isolation and characterization of *Streptomyces* sp. NL15-2K capable of degrading lignin-related aromatic compounds. J Biosci Bioeng 102:124–127. https://doi.org/10.1263/jbb .102.124.
- Hopwood DA, Bibb MJ, Chater KF, Kieser T, Bruton CJ, Kieser HM, Lydiate DJ, Smith CP, Ward JM, Schrempf H. 1985. Genetic manipulation of Streptomyces: a laboratory manual. The John Innes Foundation, Norwich, UK.
- Martin M. 2011. Cutadapt removes adapter sequences from highthroughput sequencing reads. EMBnet J 17:10. https://doi.org/10.14806/ ej.17.1.200.
- Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. Bioinformatics 30:2114–2120. https://doi.org/10 .1093/bioinformatics/btu170.
- Zerbino DR, Birney E. 2008. Velvet: algorithms for *de novo* short read assembly using de Bruijn graphs. Genome Res 18:821–829. https://doi .org/10.1101/gr.074492.107.

- Kajitani R, Toshimoto K, Noguchi H, Toyoda A, Ogura Y, Okuno M, Yabana M, Harada M, Nagayasu E, Maruyama H, Kohara Y, Fujiyama A, Hayashi T, Itoh T. 2014. Efficient *de novo* assembly of highly heterozygous genomes from whole-genome shotgun short reads. Genome Res 24:1384–1395. https://doi.org/10.1101/gr.170720.113.
- Bosi E, Donati B, Galardini M, Brunetti S, Sagot M-F, Lió P, Crescenzi P, Fani R, Fondi M. 2015. MeDuSa: a multi-draft based scaffolder. Bioinformatics 31:2443–2451. https://doi.org/10.1093/bioinformatics/btv171.
- Overbeek R, Olson R, Pusch GD, Olsen GJ, Davis JJ, Disz T, Edwards RA, Gerdes S, Parrello B, Shukla M, Vonstein V, Wattam AR, Xia F, Stevens R. 2014. The SEED and the Rapid Annotation of microbial genomes using Subsystems Technology (RAST). Nucleic Acids Res 42:D206–D214. https:// doi.org/10.1093/nar/gkt1226.
- Blin K, Wolf T, Chevrette MG, Lu X, Schwalen CJ, Kautsar SA, Suarez Duran HG, de Los Santos ELC, Kim HU, Nave M, Dickschat JS, Mitchell DA, Shelest E, Breitling R, Takano E, Lee SY, Weber T, Medema MH. 2017. antiSMASH 4.0—improvements in chemistry prediction and gene cluster boundary identification. Nucleic Acids Res 45:W36–W41. https://doi .org/10.1093/nar/gkx319.
- Iwagami SG, Yang K, Davies J. 2000. Characterization of the protocatechuic acid catabolic gene cluster from *Streptomyces* sp. strain 2065. Appl Environ Microbiol 66:1499–1508. https://doi.org/10.1128/AEM.66.4.1499 -1508.2000.
- An H-R, Park H-J, Kim E-S. 2001. Cloning and expression of thermophilic catechol 1, 2-dioxygenase gene (*catA*) from *Streptomyces setonii*. FEMS Microbiol Lett 195:17–22. https://doi.org/10.1111/j.1574 -6968.2001.tb10491.x.