

Article

Synergic Effect of Novel WS₂ Carriers Holding Spherical Cobalt Ferrite @cubic Fe₃O₄ (WS₂/s-CoFe₂O₄@c-Fe₃O₄) Nanocomposites in Magnetic Resonance Imaging and Photothermal Therapy for Ocular Treatments and Investigation of Corneal Endothelial Cell Migration

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Abstract: The design of novel materials to use simultaneously in an ocular system for driven therapeutics and wound healing is still challenging. Here, we produced nanocomposites of tungsten disulfide carriers with spherical cobalt ferrite nanoparticles (NPs) as core inside a cubic iron oxide NPs shell (WS₂/s-CoFe₂O₄@c-Fe₃O₄). Transmission electron microscopy (TEM) confirmed that 10 nm s-CoFe₂O₄@c-Fe₃O₄ NPs were attached on the WS₂ sheet surfaces. The cytotoxicity of the WS₂ sheets and nanocomposites were evaluated on bovine cornea endothelial cells (BCECs) using 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay for a duration of three days. The MTT assay results showed low toxicity of the WS₂ sheets on BCECs by 67% cell viability at 100 μ g/mL in 24 h, while the nanocomposites show 50% cell viability in the same conditions. The magnetic resonance imaging (MRI) of nanocomposites revealed the excellent T_2 -weighted imaging with an r_2 contrast of 108 mM⁻¹ S⁻¹. The in vitro photothermal therapy based on WS₂ sheets and WS₂/s-CoFe₂O₄ @c-Fe₃O₄ nanocomposites using 808 nm laser showed that the maximum thermal energy dispatched in medium at different applied power densities (1200 mw, 1800, 2200, 2600 mW) was for 0.1 mg/mL of the sample solution. The migration assay of BCECs showed that the wound healing was approximately 20% slower for the cell exposed by nanocomposites compared with the control (no exposed BCECs). We believe that WS₂/s-CoFe₂O₄@c-Fe₃O₄ nanocomposites have a synergic effect as photothermal therapy agents for eye diseases and could be a target in an ocular system using MRI.

Keywords: WS₂/s-CoFe₂O₄@c-Fe₃O₄ nanocomposites; MRI; BCECs; photo thermal therapy

1. Introduction

The new generation of two dimensional (2D) materials called transition metal dichalcogenide (TMDC) consists of hexagonal layers of metal atoms between two layers of the chalcogen atoms which have the ability to observed microwave and near infrared (NIR) wave lengths [1–3]. These novel two-dimensional sheets such as MoS₂, MoSe₂, WS₂, and WSe₂, exhibit large surface area and show a



boundary effect, resulting in notable electronics and photonics properties [4–6]. In addition, the TMDC semiconductors have direct band gaps in the visible and infrared regimes with giant light matter coupling properties and thus become a suitable candidate for the optoelectronics and optics applications [7–9]. Among the TMDCs, tungsten disulfide (WS_2) sheets warrant particular attention because of their non-toxicity, high thermal degradation and high resistance against oxidations [10,11]. The WS₂ sheets have high NIR absorption capability to use for photothermal treatments in cancer therapy [9,12]. The laser-assisted therapeutics for in vivo and in vitro treatments can be more effective while coupled with materials with high NIR absorbance [13–16]. In this regard, various types of nanoparticles (NPs) are found to assist laser treatments specially for treatment of eye diseases [17,18]. Sauvage et al. reported on applying laser to ablate the plasmonic NPs conjugated with the vitreous opacities as feasible treatments for eye diseases [19]. In other research, laser has been used to trigger graphene nanosheets to release the drugs inside the eyes after administrations [20–22]. To navigate therapeutics, in NPs movements inside the eyes, the use of non-invasive medical diagnostic tools like magnetic resonance imaging (MRI) is promising. To target the nanodevices by MRI, the novel design of nanocomposites for theragnostic applications in biomedicine is of interest. Hatamie et al. designed graphene (2D)/cobalt nano composites for both cancer therapy and MRI imaging [23], while Shahsavari Alavijeh et al. synthesized the molybdenum disulfide/cobalt ferrite (MoS₂/CoFe₂O₄) nanocomposites to use as contrast agents for MRI in cancer treatments [24]. To induce the magnetic properties into the 2D carriers (graphene, TMDCs), magnetic nanoparticles (MNPs) with better magnetic properties are required. Besides, iron oxide MNPs (Fe₃O₄) in magnetite phase have great potential in therapeutic and advanced diagnostics because of their high saturation magnetization [25–27]. Also, the spinel ferrimagnetic cobalt ferrite MNPs (CoFe₂O₄) have high curie temperature (\sim 400 °C), high magnetic anisotropy, and high coercivity compared to those of magnetite NPs [28–30]. Interestingly, the combination of MNPs named bi-magnetic core-shell NPs are used in multiferroic technologies and MRI bioimaging because of their unique properties such as high exchange bias, tunable coercivity, blocking temperatures, and large resonance fields compared to the single MNPs [31,32]. The magnetic properties of the core-shell nanostructures are reported to improve due to the diverse coupling interactions between the core and shell nano particles via interfacial defect acts as anisotropy in ferro-ferri and ferro-anti ferro magnetic structures [33–35], which make them great candidates to use in magnetic imaging [36–38]. In this study, we used the seed-growth technique to prepare spherical cobalt ferrite (s-CoFe₂O₄) as a magnetic core template to cover with the cubic iron oxide as shell (c-Fe₃O₄) to form (s-CoFe₂O₄@c-Fe₃O₄) core-shell NPs [39-42]. The WS₂ nanosheets were synthesized by a series of chemical methods followed by a non-oxygen annealing process. Furthermore, the nanocomposites of the WS₂/s-CoFe₂O₄@c-Fe₃O₄ are prepared with the facile bath sonication of as-synthesized materials. The morphology of the WS₂, core-shell NPs, and nanocomposites have been characterized using TEM. The cytotoxicity of the WS₂ and nanocomposites are evaluated using MTT assay on BCECs. The cell migration assay for wound healing was done on the BCECs exposed with nanocomposites. The in vitro photothermal measurements were done on nanocomposites and WS₂ for comparison. These WS₂/s-CoFe₂O₄@c-Fe₃O₄ nanocomposites have great potential for use in the ragnostic applications as the rapeutics and bioimaging material [43].

2. Result and Discussion

2.1. TEM of WS₂/s-CoFe₂O₄@c-Fe₃O₄ Nanocomposites

TEM images of the WS₂ sheets, s-CoFe₂O₄@c-Fe₃O₄ core-shell NPs, WS₂/s-CoFe₂O₄@c-Fe₃O₄ nanocomposites are shown in Figure 1. As seen in Figure 1a, the WS₂ sheets performed layered structures in micro size. The TEM images of core shell MNPs (Figure 1b) showed the cubic iron oxide shell. The size of the core shell NPs is evaluated to be ~10 nm. The spherical cobalt ferrites are expected to cover inside the core-shell structures. Here, the core-shell NPs were obtained by a seed mediation thermal decomposition routes, which was used as prepared CoFe₂O₄ NPs as template, then the cubic structured iron oxide NPs is grown as a shell. As seen in Figure 1b,d, the CoFe₂O₄ does

not appear in the TEM mode. This could be considered as an obscured signal because of the thick shell. This assumption can be supported by energy dispersive spectrometer (EDS), which confirmed the present of the cobalt and iron and oxygen element in the core-shell NPs (Figure 1d). The TEM image of nanocomposites (Figure 1c) confirmed the s-CoFe₂O₄@c-Fe₃O₄ NPs attachments in the WS₂ surfaces. In addition, its inset shows the selected area electron diffraction (SAED) pattern for the nanocomposites; lattice fringes corresponding to the WS₂ sheets and core-shell NPs could be observed [44].



Figure 1. Transmission Electron Microscopy (TEM) images of the (**a**) WS₂ sheets, (**b**) s-CoFe₂O₄@c-Fe₃O₄ NPs and (**c**) WS₂/s-CoFe₂O₄@c-Fe₃O₄ nanocomposites shows the layered structure of WS₂ and the size of the nanoparticles (NPs) is observed to be 10 nm, (**d**) the energy dispersive spectrometer (EDS) structural study of the s-CoFe₂O₄@c-Fe₃O₄ NPs. The inset of figure (**c**) showed its corresponding selected area electron diffraction (SAED) pattern. (**d**) The EDS structural study of the s-CoFe₂O₄@c-Fe₃O₄ NPs confirmed the presence of the cobalt, iron and oxygen elements in the core-shell magnetic nanoparticles (MNPs).

2.2. MTT Assay of WS₂ Sheets and WS₂/s-CoFe₂O₄@c-Fe₃O₄ Nanocomposites

To study the cytotoxicity of the WS₂ sheets and WS₂/s-CoFe₂O₄@c-Fe₃O₄ nanocomposites, the bovine cornea endothelial cells were utilized. MTT assay of the BCECs exposed to the different concentrations of WS₂ sheets and nanocomposites (range from 0 to 100 μ g/mL) with treatment times of 24, 48, 72 h are presented in Figure 2. The result of cell viability of WS₂ sheets on the BCECs (Figure 2a) shows low toxicity effects (~25% cell destruction) at the higher concentration of 100 μ g/mL for 72 h [45]. Teo et al. [46] also reported previously that the cytotoxicity of TMDs (MoS₂, WS₂, and WSe₂) was even lower than of graphene derivatives [47,48]. In their experiments, the MTT assays are evaluated on human lung carcinoma epithelial cells (A549) for 24 h of exposure, for which WS₂ sheets show higher cell viability of 90.6%. The cytotoxicity of the WS₂/s-CoFe₂O₄@c-Fe₃O₄ nanocomposites toward BCECs was shown in Figure 2b. As seen in the figure, BCECs cells exhibit different toxicity responses in different time and dosage of nanocomposites. The MTT assay result of nanocomposites revealed that toxicity is enhanced drastically in 100 μ g/mL compared with the WS₂ sheets. Moreover, cell destruction of ~50% was detected in 72 h of incubations. These results can be supported by research done by

Yang et al. [43] on WS₂@Fe₃O₄ nanocomposite coated with the mesoporous silica which showed no toxicity to the three types of tested cells (i.e., 4T1 murine breast cancer cells, HeLa human cervical cancer cells and 293T human embryonic kidney cells) even in 200 μ g/mL. However, WS₂ sheets and iron oxide nanoparticles are well known to be biocompatible materials and used often for in vivo cancer therapy. Then the increases of the cell death in nanocomposites could correspond to the trace amount of the cobalt content in the cobalt ferrites NPs [49,50]. In contrast, the WS₂ was used as a biocompatible 2D material here for offsetting the toxic nature of cobalt add-up in the WS₂/s-CoFe₂O₄@c-Fe₃O₄ nanocomposites [51].



Figure 2. 3-[4,5-dimethylthiazol-2yl]-2,5-diphenyl tetrazolium bromide (MTT) assay of the (**a**) WS₂ sheets and (**b**) WS₂/s-CoFe₂O₄@c-Fe₃O₄ nanocomposites on bovine cornea endothelial cells for duration of the 24, 48, and 72 h. The data were represented as mean \pm SD (n = 3).

2.3. WS₂/s-CoFe₂O₄@c-Fe₃O₄ Nanocomposites Effect on BCECs Migration Capacity

The cell migration response of the cornea endothelial cells is important for wound healing in eye surgery, ocular diseases, and angiogenesis [52,53]. The cell migration assay tested on BCECs was exposed with WS_2/s -CoFe₂O₄@c-Fe₃O₄ nanocomposites for 24 h. The scratch wound assay of the layers of BCECs used as control, and the cell closures were scanned by optical microscope. The presented photograph was procured at 0, 16 h, 24 h after the wound formation in the culture of exposed BCECs and control (Figure 3a). The nanocomposites-treated BCECs show moderate wound closures compared

to the control of ~33% closure and ~80% after 16 h and 24 h of incubation, respectively. Our results could be supported by previous studies on investigation the effect of nanoceria conjugated with heparin that suppressed the migrations of cornea endothelial cells [54]. The representative photograph of the BCECs used as control and cells contain nanocomposites shows that the BCECs loaded the gap by their migration and not by stretching the cell via increasing the cell size. Furthermore, the result shows that the WS₂/s-CoFe₂O₄@c-Fe₃O₄ nanocomposites-treated BCECs attenuate the migration in transwell migration assay [55].



Figure 3. (a) The cell migration scratching assay on bovine cornea endothelial cells (b) quantitative analysis of BCECs migration represent the relative wound width calculated in relation to initial time (0 h). Results are expressed as mean \pm SD repeated n = 3 experiments.

2.4. Photothermal Effect of WS₂ Sheets and WS₂/s-CoFe₂O₄@c-Fe₃O₄ Nanocomposites

To investigate the photothermal response of the WS₂ sheets and WS₂/s-CoFe₂O₄@c-Fe₃O₄ nanocomposites, their NIR absorption was evaluated using 808 nm laser. The laser ablation evaluations of three different concentrations of the WS₂ and WS₂/CoFe₂O₄ @Fe₃O₄ nanocomposites (1, 0.5 and 0.1 mg/mL) for four different laser power densities of 1200, 1800, 2200 and 2600 mW were shown in Figures 4 and 5, respectively. The results show an increasing trend of temperature via time in each laser when densities for all nanocomposites suspensions are applied. In other words, for concentration

of 1 mg/mL of the WS₂ sheets and WS₂/CoFe₂O₄ @Fe₃O₄ nanocomposites suspension in DI water, in the same power density of 1200 mW at 2 min of laser irradiation, the temperature was found to be 17.4 °C and 15.6 °C, respectively. The increasing of the temperature rate is proved to be dependent on samples concentration, laser irradiation time, and power density. As reported previously, WS₂ sheets and their composites exhibited an absorption in NIR regions (750–850 nm) which was higher than that for graphene in the same region [56,57].

The reason for the decreases of temperature rate in nanocomposites compared with the WS₂ sheets could be due to the partial coverage of the WS₂ sheet surfaces with the core-shell NPs. Also, when the magnetic core-shell NPs are attached electrostatically on the WS₂ sheet surfaces, with proper distance between them, it avoids their aggregations via dipole-dipole interactions, which is a great benefit to manage the appropriate distance between the WS₂ sheets for better NIR absorption when the WS₂ sheets are multilayered. The experiments were repeated for three cycles under laser irradiation for each sample to confirm the equal temperature rate. Thermal stability of the samples shows no significant reduction in optical absorbance in each cycle and the temperature of the water under laser irradiation of reported power densities in two minutes shows elevation below 1 °C.



Figure 4. The photothermal evaluations of laser absorption of WS_2 sheets in various laser power densities of (a) 1200, (b) 1800, (c) 2200 and (d) 2600 mW at concentration 1, 0.5 and 0.1 mg/mL.



Figure 5. The photothermal evaluations of laser absorption of $WS_2/CoFe_2O_4$ @Fe₃O₄ nanocomposites in various laser power densities of (a) 1200, (b) 1800, (c) 2200 and (d) 2600 mW at concentration 1, 0.5, and 0.1 mg/mL.

2.5. MRI Study

MRI is a strong medical imaging technique that can provide great anatomical detail. It works by interaction of protons with the tissue's surrounding molecules [58,59]. To improve the resolution and specificity of MRI, the use of the effective contrast agents is important. Recently, research attentions have been devoted to the design and synthesis of novel MNPs with higher saturation magnetization values that have better MRI contrast agents and shorten T₂ relaxation times. However, superparamagnetic iron oxide NPs (SPIONs) have been used in clinics as negative contrast agents. Harrison et al. injected SPIONs coated polymers and tags with the fluorophore in the animal optical nerve for in vivo MR imaging [60]. Here, the nanocomposites of 2D/iron-based contrast agents in the core-shell form are designed to use as in vitro MRI contrast agents of the ocular system. Figure 6 shows the relaxivity measurements of WS₂/s-CoFe₂O₄@c-Fe₃O₄ nanocomposites for various concentrations. The T₁-weighted and T₂-weighted images show concentration-dependent contrast. The results show which nanocomposites could act as a T₂-weighted contrast agent. Figure 6b shows the 1/T₁ and 1/T₂ relaxation diagram versus the nanocomposite's concentrations. The longitudinal and transverse nuclear relaxivities generated from the slopes of $1/T_1$ and $1/T_2$ plots are approximately $r_1 = 0.73$ mM⁻¹ S^{-1} ($R^2 = 0.84$), and $r_2 = 108 \text{ mM}^{-1} \text{ S}^{-1}$ ($R^2 = 0.98$). The high r_2 revealed the darkening effect of nanocomposites via different concentrations. The results show that the nanocomposites have the potential to be used as T₂ MRI contrast agents in a diagnostic probe.



Figure 6. (a) T_1 and T_2 -weighted MR images at five different concentrations of nanocomposites. (b) The linear fitting of relaxation rates $(1/T_2)$ of versus nanocomposites concentrations. (c) The linear fitting of relaxation rates $(1/T_1)$ of versus nanocomposites concentrations. The relaxivity values of r_2 and r_1 were obtained from the slopes.

3. Materials and Methods

3.1. Materials

Iron acetylacetonate (Fe(acac)₃), cobalt acetylacetonate (Co(acac)₂), phenyl ether, benzyl ether, oleic acid (OA), and absolute ethanol were obtained from (ACROS ORGANIC, Morris Plains, NJ, USA), Tungsten hexachloride (WCl₆) and 1-octadecene were purchased from (Alfa Aesar Inc., Haverhill, MA, USA), Oleylamine (OAm), hexane, ethylene glycol, 3-(3,4-dihydroxy)hydrocinnamic acid (DHCA), tetrahydrofuran (THF), NaOH, and phosphate buffered saline (PBS, concentration of PO₄^{2–} ions (0.0067 M) were acquired from Tokyo Chemical Industry (TCI Co, Tokyo, Japan), (Fisher chemicals, Pittsburgh, PA, USA), (J.T.Baker, Phillipsburg, NJ, USA), (Sigma Aldrich, St. Louis, MO, USA) (ECHO CHEMICAL CO., LTD., Taipei, Taiwan), (SHOWA Chemical, Tokyo, Japan), and HyCloneTM, Marlborough, MA, USA) respectively. The above chemicals were used without further purification. The 3-[4,5-dimethylthiazol-2yl]-2,5-diphenyl tetrazolium bromide (MTT) and Dulbecco's Phosphate Buffered Saline (DPBS) were purchased from (Sigma Aldrich, St. Louis, MO, USA).

3.2. Synthesis of the Core-Shell NPs

3.2.1. Cobalt Ferrite (s-CoFe₂O₄) Seed Nanoparticle Preparation

A quantity of 2.655 g Fe(acac)₃ and 1.26 g Co(acac)₂ were mixed in the presence of 60 mL phenyl ether, followed by adding 60 mL OAm. The mixed solution was added into the nitrogen-injected three-necked round-bottom flask and heated at 100 °C for 1 h, in order to remove the moisture. After 1 h, the solution was heated to the temperature 260 °C at rate 3 °C/min and heated to reflux at 260 °C for 2 h. Then, the solution was cooled down by the removal of the heat resource to the room temperature. The as-synthesized mixture was washed by ethanol and collected by the magnetic decantation twice for each sample to remove the residues of the unreacted chemicals. The black precipitation was redispersed in 30 mL hexane. The obtained product was labeled as spherical OAm-CoFe₂O₄.

3.2.2. s-Cobalt Ferrite@c-iron Oxide (s-CoFe2O4@c-Fe3O4) Core-Shell Preparation

A quantity of 1 g Fe(acac)₃, 3 mL seed solution (12.6 mg CoFe₂O₄), 20 mL OA, 20 mL OAm, 20 mL benzyl ether, and 1.75 mL ethylene glycol were mixed uniformly, followed by being added to the three-necked round-bottom flask, which was preheated at 100 °C under nitrogen atmosphere. After preheating for 1 h, the solution was heated to the temperature 290 °C at rate 6 °C/min and refluxing for 30 min. Due to the presence of ethylene glycol as mentioned above, as the temperature reached 240 °C, a vigorous evaporation would be expected. Thus, the nitrogen flow rate was adjusted to increase the flow rate to remove the vaporous ethylene glycol from flask. After refluxing, the solution was cooled down to room temperature and further washed with the absolute ethanol and magnetic decantation twice for each. The obtained powders were suspended in 10 mL hexane for storage, and labeled as cubic OA-CoFe₂O₄@Fe₃O₄.

3.2.3. Ligand Exchange of Core-Shell Nanoparticles

A volume of 2 mL cubic OA-CoFe₂O₄@Fe₃O₄ (solvent: hexane) was added into a flask, which contained 3-(3,4-dihydroxy) hydrocinnamic acid/tetrahydrofuran solution (DHCA/THF, 0.125 g/15 mL). The solution was stirred with magnetic stirrer at 60 °C for 4 h. After the ligand exchange finished, the products were washed with ethanol containing a small amount of NaOH twice. The precipitation was further redispersed in 2 mL DI water. The samples were labeled as cubic DHCA-CoFe₂O₄@Fe₃O₄ [61].

3.2.4. Preparation of WS₂/Core-Shell Nanocomposites

WS₂ Preparation

A quantity of 0.3966 g WCl₆ was mixed with 20 mL OAm and 10 mL 1-octadecene at room temperature [45]. After 1 h of preheating and nitrogen injection at 100 °C, the mixture was heated to 300 °C and reflux for 30 min, followed by adding 0.4 M of sulfur/OAm solution. The reaction at 300 °C was further carried out for 1 h to form WS₂ sheets. The solution was cooled down and washed by conducting the addition of absolute ethanol and centrifugation at rate 4000 rpm twice. The resultant solution was dried and annealed at 500 °C in the oven under nitrogen atmosphere for 2 h. Finally, the WS₂ sheets could be collected to use in further work.

WS₂/Core-Shell Nanocomposite Preparation

To decorate the cubic DHCA-CoFe₂O₄@Fe₃O₄ between the layers of WS₂ sheets, the few-layer WS₂ sheets were sonicated in a bath-type sonicator before capping the cubic core/shell structured MNPs. The as-made few-layer WS₂ sheets with phosphate buffered saline (PBS) (1 mg mL⁻¹) were added to cubic DHCA-CoFe₂O₄@Fe₃O₄ in the ratio of 1:5 in weight. The mixture was stirred at room temperature for 24 h. The products were collected by magnetic decantation. The nanocomposites were labeled as WS₂/spherical CoFe₂O₄@cubicFe₃O₄ (see Scheme 1).



Scheme 1. Shows the synthesis process of WS_2 /s-CoFe₂O₄@c-Fe₃O₄ nanocomposites.

3.3. Characterization Techniques

3.3.1. Transmission Electron Microscopy (TEM)

The morphology, selected area electron diffraction (SAED) and energy dispersive spectrometer (EDS) nanocomposites was obtained with the TEM (operating accelerating voltage: 200 kV, Philips field-emission, Tecnai F20, electron gun of ZrO/W(100) Schottky type, resolution \leq 0.23 nm (Philips/FEI Corporation, Eindhoven, The Netherlands). The aqueous samples suspended in DI-water were dropped on the carbon-coated copper grids (200 mesh), followed by dried at 80 °C for a few hours before entering the TEM chamber.

3.3.2. Derivation of Bovine Cornea Endothelial Cells (BCECs)

The bovine eyes were collected from butcher (Taipei, Taiwan) and cleaned and fumigated by iodine solution. Then the eyes were washed with Dulbecco's Phosphate Buffered Saline (DPBS) and the cornea were detached from the eyes followed by removal of the Descemet membranes. The 10 mL of trypsin were added to the membranes and incubated at 37 °C with 5% of CO_2 flow for 45 min. The BCECs were collected using centrifuge for five minutes of 1000 rpm and dispersed in the 6 cm culture dish. The cells grew and increased during a two-week incubation.

3.3.3. Cytotoxicity Measurements

The cytotoxicity of the WS₂ and the WS₂/s-CoFe₂O₄@c-Fe₃O₄ nanocomposites on the BCECs were assessed using 3-[4,5-dimethylthiazol-2yl]-2,5-diphenyl tetrazolium bromide (MTT) assay. The cells were cultured in the 96 well plates (10^4 cells/well) and incubated in 37 °C with 5% of Co₂ for 24 h.

Then the various concentrations of the WS₂ and nanocomposites (10, 30, 50, 70, 100 μ g/mL) were dispersed in the PBS and added to the cells. The MTT assay was done on BCECs incubated with the samples after 24, 48 and 72 h. Finally, the 10 μ L of the MTT solution was added to each well and incubated under 5% CO₂ at 37 °C for 4 h. The cells were washed by PBS followed by adding the dimethyl sulfoxide (DMSO) in each plate and shake for 10 min. The optical density was collected by inserting the plates in microplate reader (Bio-Rad S/N 21648, Pleasanton, CA, USA) with excitation wave length of 595 nm. [62]. Data were presented as mean standard deviation (±SD) of three experiments. The following equation was used to calculate the survival BCECs from the absorbance collected by microplate:

MTT assay(cell viability%) = $(\text{sample abs 595 nm})/(\text{control abs 595 nm}) \times 100$

3.3.4. BCECs Migration Assay

In vitro BCECs scratch assay or migration assay was performed in 6 cm round culture plates. The procedure was started with the seeding of the 10^6 BCECs in each dish and incubated for 24 h. Then 100 µL of the WS₂/s-CoFe₂O₄@c-Fe₃O₄ nanocomposites was dispersed in PBS and added to the culture dish. The cells and nanocomposites were incubated for 24 h until the nanocomposites were taken up by the BCECs. Straight lines were scratched in the cultured dishes (control and dish with the nanocomposites) by a sterile pipet tip of 100 µL. The cell migrations were targeted for 24 h. The collected images of the scratches were achieved at 100× magnification followed by further analysis by Image J software. Each experiment was repeated in triplicate.

3.3.5. Magnetic Resonance Imaging of Nanocomposites

To achieve the WS₂/s-CoFe₂O₄@c-Fe₃O₄ nanocomposites MRI relaxivity, a 7 Tesla (Bruker BioS pec 70/30 US, Billerica, MA, USA) scanner was used. The phantoms are prepared by the various concentration of the nanocomposites (0.07, 0.05, 0.03, 0.02, 0.01 mg/mL) dispersed in deionized water (DW) by adding 1% of agarose gel in the 0.5 mL plastic container. The relaxation times of hydrogen protons in the aqueous solution (T₁ and T₂-weighted) were measured at repetition time TR: 4000 ms; TE: 18 ms. To calculate the data, the obtained T₁ and T₂ maps were analysed presuming a mono-exponential signal decay. The maps were recognized using six-spin echo (SE) images of TE and TR. The T1 and T₂-weighted images were analyzed using a non-linear least-square curve using pixel intensities basis (Levenberg–Marquardt fit) using MATLAB (MathWorks inc. Natick, MA, USA) [59]. The r₁ and r₂ relaxivities were calculated from the slop of 1/T₁ and 1/T₂ (i.e., reciprocal for T₁ and T₂ relaxation times) versus the nano composite's concentrations. The images were acquired by designing a sequence of slice thickness of 1 mm and a matrix size of 128 × 128 over FOV of 6 × 6 cm².

3.3.6. Near Infrared (NIR) Experiment

The WS₂ sheets and WS₂/s-CoFe₂O₄@c-Fe₃O₄ nanocomposites with concentrations of 0.1, 0.5, 1 mg/mL were dispersed in 1 mL DW and ultrasonicated until complete dispersion in the solution was achieved. The suspensions were irradiated by 808 nm NIR laser system (Arno Electro-Optics Ltd. Taipei, Taiwan) under four various power densities of 1200, 1800, 2200, 2600 mW. The temperature of the samples' suspension in the DW were monitored using k-type thermocouple (TM-947SD) (Lutron, Taipei, Taiwan).

4. Conclusions

The WS₂/s-CoFe₂O₄@c-Fe₃O₄ nanocomposites were synthesized via chemical routes. The TEM showed that the core-shell NPs with the size of the 10 nm were pinned up on the WS₂ sheet surfaces. The MTT cell viability assay of the nanocomposites on the BCECs showed the 50% cell viability in 100 μ g/mL at 72 h of incubation. The cell migration studies of the nanocomposites exposed to the BCECs showed wound closure of 80% compared to the control. The photothermal studies of the WS₂/s-CoFe₂O₄@c-Fe₃O₄ nanocomposites showed temperature elevated trend in each power density and the temperature rates were maximized for the lower concentration of 0.1 mg/mL. In the MRI of WS₂/s-CoFe₂O₄@c-Fe₃O₄ nanocomposites the r₂ relaxivity value was calculated to be 108 mM⁻¹ S⁻¹. The results showed that nanocomposites have the potential for use in laser treatments and as a T₂-weighted MRI contrast agent for ocular systems.

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References

- 1. Levin, T.; Sade, H.; Binyamini, R.B.-S.; Pour, M.; Nachman, I.; Lellouche, J.-P. Tungsten disulfide-based nanocomposites for photothermal therapy. *Beilstein J. Nanotechnol.* **2019**, *10*, 811–822. [CrossRef]
- Nandi, S.; Bhunia, S.K.; Zeiri, L.; Pour, M.; Nachman, I.; Raichman, D.; Lellouche, J.-P.; Jelinek, R. Bifunctional Carbon-Dot-WS2Nanorods for Photothermal Therapy and Cell Imaging. *Chem.-A Eur. J.* 2017, 23, 963–969. [CrossRef] [PubMed]
- 3. Gu, Z.; Zhou, L.; Gu, Z.; Yan, L.; Tian, G.; Zheng, X.; Liu, X.; Zhang, X.; Shi, J.; Cong, W.; et al. WS2 nanosheet as a new photosensitizer carrier for combined photodynamic and photothermal therapy of cancer cells. *Nanoscale* **2014**, *6*, 10394–10403. [CrossRef]
- 4. Mao, X.; Xu, Y.; Xue, Q.; Wang, W.; Gao, D. Ferromagnetism in exfoliated tungsten disulfide nanosheets. *Nanoscale Res. Lett.* **2013**, *8*, 430. [CrossRef] [PubMed]
- Gao, Y.; Liu, Z.; Sun, D.-M.; Huang, L.; Ma, L.-P.; Yin, L.-C.; Ma, T.; Zhang, Z.; Ma, X.-L.; Peng, L.-M.; et al. Large-area synthesis of high-quality and uniform monolayer WS2 on reusable Au foils. *Nat. Commun.* 2015, 6, 8569. [CrossRef] [PubMed]
- Yin, W.; Yu, J.; Lv, F.; Yan, L.; Zheng, L.R.; Gu, Z.; Zhao, Y. Functionalized Nano-MoS2with Peroxidase Catalytic and Near-Infrared Photothermal Activities for Safe and Synergetic Wound Antibacterial Applications. ACS Nano 2016, 10, 11000–11011. [CrossRef] [PubMed]
- Liu, Q.; Sun, C.; He, Q.; Khalil, A.; Xiang, T.; Liu, D.; Zhou, Y.; Wang, J.; Song, L. Stable metallic 1T-WS2 ultrathin nanosheets as a promising agent for near-infrared photothermal ablation cancer therapy. *Nano Res.* 2015, *8*, 3982–3991. [CrossRef]

- 8. Heising, J.; Kanatzidis, M.G. Exfoliated and Restacked MoS2and WS2: Ionic or Neutral Species? Encapsulation and Ordering of Hard Electropositive Cations. J. Am. Chem. Soc. **1999**, 121, 11720–11732. [CrossRef]
- 9. Feng, W.; Chen, L.; Qin, M.; Zhou, X.; Zhang, Q.; Miao, Y.; Qiu, K.; Zhang, Y.; He, C. Flower-like PEGylated MoS2 nanoflakes for near-infrared photothermal cancer therapy. *Sci. Rep.* **2015**, *5*, 17422. [CrossRef]
- 10. Nazari, H.; Heirani-Tabasi, A.; Hajiabbas, M.; Khalili, M.; Alavijeh, M.S.; Hatamie, S.; Gorabi, A.M.; Esmaeili, E.; Tafti, S.H.A. Incorporation of two-dimensional nanomaterials into silk fibroin nanofibers for cardiac tissue engineering. *Polym. Adv. Technol.* **2020**, *31*, 248–259. [CrossRef]
- 11. Cao, H.; De Hosson, J.T.; Pei, Y.T. Self-healing of a pre-notched WS2/a-C coating. *Mater. Res. Lett.* **2018**, *7*, 103–109. [CrossRef]
- 12. Shirshahi, V.; Tabatabaei, S.N.; Hatamie, S.; Saber, R. Photothermal enhancement in sensitivity of lateral flow assays for detection of E-coli O157:H7. *Colloids Surfaces B Biointerfaces* **2020**, *186*, 110721. [CrossRef]
- 13. Espinosa, A.; Di Corato, R.; Kolosnjaj-Tabi, J.; Flaud, P.; Pellegrino, T.; Wilhelm, C. Duality of Iron Oxide Nanoparticles in Cancer Therapy: Amplification of Heating Efficiency by Magnetic Hyperthermia and Photothermal Bimodal Treatment. *ACS Nano* **2016**, *10*, 2436–2446. [CrossRef] [PubMed]
- 14. Sidman, R.L.; Li, J.; Lawrence, M.; Hu, W.; Musso, G.F.; Giordano, R.J.; Cardó-Vila, M.; Pasqualini, R.; Arap, W. The peptidomimetic Vasotide targets two retinal VEGF receptors and reduces pathological angiogenesis in murine and nonhuman primate models of retinal disease. *Sci. Transl. Med.* 2015, *7*, 309ra165. [CrossRef] [PubMed]
- 15. Kamiya, K.; Takahashi, M.; Igarashi, A.; Shoji, N. Visual Performance in Eyes Undergoing Femtosecond Laser-Assisted Keratoplasty for Advanced Keratoconus. *Sci. Rep.* **2019**, *9*, 6442. [CrossRef] [PubMed]
- 16. Kim, H.; Robinson, S.B.; Csaky, K.G. Investigating the Movement of Intravitreal Human Serum Albumin Nanoparticles in the Vitreous and Retina. *Pharm. Res.* **2009**, *26*, 329–337. [CrossRef] [PubMed]
- 17. Li, Y.-J.; Luo, L.-J.; Harroun, S.G.; Wei, S.-C.; Unnikrishnan, B.; Chang, H.-T.; Huang, Y.-F.; Lai, J.-Y.; Huang, C.-C. Synergistically dual-functional nano eye-drops for simultaneous anti-inflammatory and anti-oxidative treatment of dry eye disease. *Nanoscale* **2019**, *11*, 5580–5594. [CrossRef]
- Wang, M.; Yang, Q.; Li, M.; Zou, H.; Wang, Z.; Ran, H.; Zheng, Y.; Jian, J.; Zhou, Y.; Luo, Y.; et al. Multifunctional Nanoparticles for Multimodal Imaging-Guided Low-Intensity Focused Ultrasound/Immunosynergistic Retinoblastoma Therapy. ACS Appl. Mater. Interfaces 2020, 12, 5642–5657. [CrossRef]
- Sauvage, F.; Fraire, J.; Remaut, K.; Sebag, J.; Peynshaert, K.; Harrington, M.; Van De Velde, F.J.; Xiong, R.; Tassignon, M.-J.; Brans, T.; et al. Photoablation of Human Vitreous Opacities by Light-Induced Vapor Nanobubbles. *ACS Nano* 2019, *13*, 8401–8416. [CrossRef]
- 20. Yu, X.; Cheng, H.; Zhang, M.; Zhao, Y.; Qu, L.; Shi, G. Graphene-based smart materials. *Nat. Rev. Mater.* **2017**, *2*, 1–14. [CrossRef]
- 21. Machado, M.; Silva, G.A.; Bitoque, D.B.; Ferreira, J.; Pinto, L.A.; Morgado, J.; Ferreira, Q. Self-Assembled Multilayer Films for Time-Controlled Ocular Drug Delivery. *ACS Appl. Bio Mater.* **2019**, *2*, 4173–4180. [CrossRef]
- Khan, M.S.; Misra, S.K.; Schwartz-Duval, A.S.; Daza, E.; Ostadhossein, F.; Bowman, M.; Jain, A.; Taylor, G.; McDonagh, D.; Labriola, L.T.; et al. Real-Time Monitoring of Post-Surgical and Post-Traumatic Eye Injuries Using Multilayered Electrical Biosensor Chip. ACS Appl. Mater. Interfaces 2017, 9, 8609–8622. [CrossRef] [PubMed]
- 23. Hatamie, S.; Ahadian, M.M.; Ghiass, M.A.; Zad, A.I.; Saber, R.; Parseh, B.; Oghabian, M.A.; Zadeh, S.S. Graphene/cobalt nanocarrier for hyperthermia therapy and MRI diagnosis. *Colloids Surfaces B Biointerfaces* **2016**, *146*, 271–279. [CrossRef] [PubMed]
- 24. Alavijeh, M.S.; Maghsoudpour, A.; Khayat, M.; Rad, I.; Hatamie, S. Distribution of "molybdenum disulfide/cobalt ferrite" nanocomposite in animal model of breast cancer, following injection via differential infusion flow rates. *J. Pharm. Investig.* **2020**, *50*, 583–592. [CrossRef]
- Talebi, M.; Balasi, Z.M.; Ahadian, M.M.; Hatamie, S.; Alavijeh, M.H.S.; Ghafuri, H. Biocompatibility and Hyperthermia Efficiency of Sonochemically Synthesized Magnetic Nanoparticles. SPIN 2019, 9, 1940006. [CrossRef]
- Béalle, G.; Di Corato, R.; Kolosnjaj-Tabi, J.; Dupuis, V.; Clément, O.; Gazeau, F.; Wilhelm, C.; Ménager, C. Ultra Magnetic Liposomes for MR Imaging, Targeting, and Hyperthermia. *Langmuir* 2012, 28, 11834–11842. [CrossRef]
- 27. Bani, M.S.; Hatamie, S.; Haghpanahi, M. Biocompatibility and hyperthermia cancer therapy of casein-coated iron oxide nanoparticles in mice. *Polym. Adv. Technol.* **2020**, *31*, 1544–1552. [CrossRef]

- Zhao, L.; Zhang, H.; Xing, Y.; Song, S.-Y.; Yu, S.; Shi, W.; Guo, X.; Yang, J.; Lei, Y.; Cao, F. Studies on the magnetism of cobalt ferrite nanocrystals synthesized by hydrothermal method. *J. Solid State Chem.* 2008, 181, 245–252. [CrossRef]
- Hatamie, S.; Parseh, B.; Ahadian, M.M.; Naghdabadi, F.; Saber, R.; Soleimani, M. Heat transfer of PEGylated cobalt ferrite nanofluids for magnetic fluid hyperthermia therapy: In vitro cellular study. *J. Magn. Magn. Mater.* 2018, 462, 185–194. [CrossRef]
- 30. Nanoparticles, C.O.M.; Pauline, S.; Amaliya, A.P. Size and Shape Control Evaluation of Cobalt (Co) and Cobalt Ferrite. *Arch. Phys. Res.* **2012**, *3*, 78–83.
- 31. Carvalho, A.; Domingues, I.; Gonçalves, M.C. Core–shell superparamagnetic nanoparticles with interesting properties as contrast agents for MRI. *Mater. Chem. Phys.* **2015**, *168*, 42–49. [CrossRef]
- Martínez-Banderas, A.I.; Aires, A.; Plaza-García, S.; Colás, L.; Moreno, J.A.; Ravasi, T.; Merzaban, J.S.; Cabrer, P.R.; Cortajarena, A.L.; Kosel, J. Magnetic core–shell nanowires as MRI contrast agents for cell tracking. *J. Nanobiotechnol.* 2020, *18*, 1–12. [CrossRef] [PubMed]
- 33. Habib, A.H.; Ondeck, C.L.; Chaudhary, P.; Bockstaller, M.R.; McHenry, M.E. Evaluation of iron-cobalt/ferrite core-shell nanoparticles for cancer thermotherapy. *J. Appl. Phys.* **2008**, *103*, 07A307. [CrossRef]
- 34. Londoño-calderón, C.L.; Moscoso-londoño, O. Synthesis and magnetic properties of cobalt- iron/cobalt-ferrite soft/hard magnetic core/shell nanowires. *Nanotechnology* **2017**, *28*, 245605. [CrossRef] [PubMed]
- 35. Wang, K.; An, L.; Tian, Q.; Lin, J.; Yang, H. Gadolinium-labelled iron/iron oxide core/shell nanoparticles as T1–T2 contrast agent for magnetic resonance imaging. *RSC Adv.* **2018**, *8*, 26764–26770. [CrossRef]
- Chai, Y.; Feng, F.; Li, Q.; Yu, C.; Feng, X. One-pot synthesis of high-quality bimagnetic core/shell nanocrystals with diverse exchange coupling 1 State Key Laboratory for Structural Chemistry of Unstable and Stable Species, Institute of Chemistry, Chinese Academy of Sciences (ICCAS) Zhongguan. J. Am. Chem. Soc. 2019, 141, 3366–3370. [CrossRef] [PubMed]
- Soleymani, M.; Velashjerdi, M.; Shaterabadi, Z.; Barati, A. One-pot preparation of hyaluronic acid-coated iron oxide nanoparticles for magnetic hyperthermia therapy and targeting CD44-overexpressing cancer cells. *Carbohydr. Polym.* 2020, 237, 116130. [CrossRef] [PubMed]
- Sharifi, I.; Shokrollahi, H.; Amiri, S. Ferrite-based magnetic nanofluids used in hyperthermia applications. J. Magn. Magn. Mater. 2012, 324, 903–915. [CrossRef]
- Esmaeili, E.; Soleimani, M.; Ghiass, M.A.; Hatamie, S.; Vakilian, S.; Zomorrod, M.S.; Sadeghzadeh, N.; Vossoughi, M.; Hosseinzadeh, S. Magnetoelectric nanocomposite scaffold for high yield differentiation of mesenchymal stem cells to neural-like cells. *J. Cell. Physiol.* 2019, 234, 13617–13628. [CrossRef]
- 40. Hatamie, S.; Shih, P.-J.; Zomorod, M.S.; Heravi, P.; Ahadian, M.M.; Hatami, N. Hyperthermia response of PEGylated magnetic graphene nanocomposites for heating applications and accelerate antibacterial activity using magnetic fluid hyperthermia. *Appl. Phys. A* **2020**, *126*, 1–10. [CrossRef]
- 41. Hatamie, S.; Mohamadyar-Toupkanlou, F.; Mirzaei, S.; Ahadian, M.M.; Hosseinzadeh, S.; Soleimani, M.; Sheu, W.-J.; Wei, Z.H.; Hsieh, T.-F.; Chang, W.-C.; et al. Cellulose Acetate/Magnetic Graphene Nanofiber in Enhanced Human Mesenchymal Stem Cells Osteogenic Differentiation Under Alternative Current Magnetic Field. *SPIN* **2019**, *9*, 1–10. [CrossRef]
- 42. Hatamie, S.; Ahadian, M.M.; Rashidi, A.; Karimi, A.; Akhavan, O. Novel synthesis of cobalt/poly vinyl alcohol/gamma alumina nanocomposite for catalytic application. *Appl. Phys. A* **2017**, *123*, 750. [CrossRef]
- Yang, G.; Gong, H.; Liu, T.; Sun, X.; Cheng, L.; Liu, Z. Two-dimensional magnetic WS2@Fe3O4 nanocomposite with mesoporous silica coating for drug delivery and imaging-guided therapy of cancer. *Biomaterials* 2015, 60, 62–71. [CrossRef] [PubMed]
- 44. Robles, J.; Das, R.; Glassell, M.; Phan, M.H.; Srikanth, H. Exchange-coupled Fe3O4/CoFe2O4 nanoparticles for advanced magnetic hyperthermia. *AIP Adv.* **2018**, *8*, 056719. [CrossRef]
- 45. Wang, S.; Zhao, J.; Yang, H.; Wu, C.; Hu, F.; Chang, H.; Li, G.; Ma, D.; Zou, D.; Huang, M. Bottom-up synthesis of WS 2 nanosheets with synchronous surface modification for imaging guided tumor regression. *Acta Biomater.* **2017**, *58*, 442–454. [CrossRef]
- 46. Teo, W.Z.; Chng, E.L.K.; Sofer, Z.; Pumera, M. Cytotoxicity of Exfoliated Transition-Metal Dichalcogenides (MoS2, WS2, and WSe2) is Lower Than That of Graphene and its Analogues. *Chem.-A Eur. J.* **2014**, *20*, 9627–9632. [CrossRef]

- 47. Mahsa, S.; Hatamie, S.; Naderi, A.; Parivar, K. Asian Journal of Nanoscience and Orginal Research Article Reduced graphene oxide/nanohydroxy Apatite-Bismuth nanocomposites for osteogenic differentiation of human mesenchymal stem cells. *Asian J. Nanosci. Mater.* **2020**, *3*, 330–339. [CrossRef]
- 48. Almasi, N.; Hosseinzadeh, S.; Hatamie, S.; Sangsari, G.T. Stable conductive and biocompatible scaffold development using graphene oxide (GO) doped polyaniline (PANi). *Int. J. Polym. Mater.* **2019**, *69*, 896–906. [CrossRef]
- 49. Hatamie, S.; Nouri, M.; Karandikar, S.; Kulkarni, A.H.; Dhole, S.; Phase, D.; Kale, S.N. Complexes of cobalt nanoparticles and polyfunctional curcumin as antimicrobial agents. *Mater. Sci. Eng. C* 2012, 32, 92–97. [CrossRef]
- 50. Hatamie, S.; Ahadian, M.M.; Zomorod, M.S.; Torabi, S.; Babaie, A.; Hosseinzadeh, S.; Soleimani, M.; Hatami, N.; Wei, Z.-H. Antibacterial properties of nanoporous graphene oxide/cobalt metal organic framework. *Mater. Sci. Eng. C* 2019, 104, 109862. [CrossRef]
- Fardindoost, S.; Hatamie, S.; Zad, A.I.; Astaraei, F.R. Hydrogen sensing properties of nanocomposite graphene oxide/Co-based metal organic frameworks (Co-MOFs@GO). *Nanotechnology* 2017, 29, 15501. [CrossRef] [PubMed]
- 52. Ho, W.-T.; Chang, J.-S.; Chou, S.-F.; Hwang, W.-L.; Shih, P.-J.; Chang, S.-W.; Yang, M.-H.; Jou, T.; Wang, I.-J. Targeting non-muscle myosin II promotes corneal endothelial migration through regulating lamellipodial dynamics. *J. Mol. Med.* **2019**, *97*, 1345–1357. [CrossRef] [PubMed]
- 53. Farjo, K.M.; Ma, J.-X. The potential of nanomedicine therapies to treat neovascular disease in the retina. *J. Angiogenesis Res.* **2010**, *2*, 21. [CrossRef] [PubMed]
- 54. Choi, S.W.; Cha, B.G.; Kim, J. Therapeutic Contact Lens for Scavenging Excessive Reactive Oxygen Species on the Ocular Surface. *ACS Nano* **2020**, *14*, 2483–2496. [CrossRef]
- 55. Tang, M.; Ji, X.; Xu, H.; Zhang, L.; Jiang, A.; Song, B.; Su, Y.; He, Y. Photostable and Biocompatible Fluorescent Silicon Nanoparticles-Based Theranostic Probes for Simultaneous Imaging and Treatment of Ocular Neovascularization. *Anal. Chem.* **2018**, *90*, 8188–8195. [CrossRef]
- 56. Ghafary, S.M.; Nikkhah, M.; Hatamie, S.; Hosseinkhani, S. Simultaneous Gene Delivery and Tracking through Preparation of Photo-Luminescent Nanoparticles Based on Graphene Quantum Dots and Chimeric Peptides. *Sci. Rep.* **2017**, *7*, 1–14. [CrossRef]
- 57. Ghafary, S.M.; Hatamie, S.; Nikkhah, M.; Hosseinkhani, S. The effect of graphite sources on preparation of photoluminescent graphene nano-sheets for biomedical imaging. *Nanomed. J.* **2017**, *4*, 164–169. [CrossRef]
- Thapa, B.; Diaz-Diestra, D.; Beltran-Huarac, J.; Weiner, B.R.; Morell, G. Enhanced MRI T 2 Relaxivity in Contrast-Probed Anchor-Free PEGylated Iron Oxide Nanoparticles. *Nanoscale Res. Lett.* 2017, 12, 1–13. [CrossRef]
- 59. Tzameret, A.; Ketter-Katz, H.; Edelshtain, V.; Sher, I.; Corem-Salkmon, E.; Levy, I.; Last, D.; Guez, D.; Mardor, Y.; Margel, S.; et al. In vivo MRI assessment of bioactive magnetic iron oxide/human serum albumin nanoparticle delivery into the posterior segment of the eye in a rat model of retinal degeneration. *J. Nanobiotechnol.* 2019, *17*, 3. [CrossRef]
- 60. Harrison, J.; Bartlett, C.A.; Cowin, G.; Nicholls, P.; Evans, C.W.; Clemons, T.D.; Zdyrko, B.; Luzinov, I.; Harvey, A.R.; Iyer, K.S.; et al. In vivo Imaging and Biodistribution of Multimodal Polymeric Nanoparticles Delivered to the Optic Nerve. *Small* **2012**, *8*, 1579–1589. [CrossRef]
- 61. Daniel, P.; Shylin, S.I.; Lu, H.; Tahir, M.N.; Panthöfer, M.; Weidner, T.; Möller, A.; Ksenofontov, V.; Tremel, W. The surface chemistry of iron oxide nanocrystals: Surface reduction of? -Fe 2 O 3 to Fe 3 O 4 by redox-active catechol surface ligands. *J. Mater. Chem. C* 2018, *6*, 326–333. [CrossRef]
- 62. Hatamie, S.; Akhavan, O.; Sadrnezhaad, S.K.; Ahadian, M.M.; Shirolkar, M.M.; Wang, H.Q. Curcumin-reduced graphene oxide sheets and their effects on human breast cancer cells. *Mater. Sci. Eng. C* 2015, *55*, 482–489. [CrossRef] [PubMed]

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