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Dawei Cao,^{1,2,3} Zhangpei Chen,¹ Leiyang Lv,¹ Huiying Zeng,³ Yong Peng,² and Chao-Jun Li^{1,4,*}

SUMMARY

Hydroxyl is widely found in organic molecules as functional group and its deprivation plays an inevitable role in organic synthesis. However, the direct cleavage of Csp^3 -O bond in alcohols with high selectivity and efficiency, especially without the assistance of metal catalyst, has been a formidable challenge because of its strong bond dissociation energy and unfavorable thermodynamics. Herein, an efficient metal-free strategy that enables direct deoxygenation of alcohols has been developed for the first time, with hydrazine as the reductant induced by light. This protocol features mild reaction conditions, excellent functional group tolerance, and abundant and easily available starting materials, rendering selective deoxygenation of a variety of 1° and 2° alcohols, vicinal diols, and β -1 and even β -O-4 models of natural wood lignin. This strategy is also highlighted by its "traceless" and non-toxic by-products N₂ and H₂, as readily escapable gases. Mechanistic studies demonstrated dimethyl sulfide being a key intermediate in this transformation.

INTRODUCTION

In view of the diminishing oil reserves and the ongoing climate change, improving the energy and utilization efficiency of natural resources is a key task to achieve future chemical sustainability (He et al., 2013; Li, 2016). Therefore, direct and selective transformation of naturally abundant functional groups is very important to increase efficiency in organic synthesis (Palacios et al., 2007; Veitch et al., 2007). In particular, the deoxygenation reaction provides an enabling tool for future biorefinery concepts through the cleavage of C-O bonds (Ruppert et al., 2012; Li et al., 2020; Schwob et al., 2019; Volkov et al., 2015; Wang et al., 2018), which allows the conversion of biomass-based readily available alcohols and polyols into platform chemicals and fuels (Bozell and Petersen, 2010; Corma et al., 2007; Dam and Hanefeld, 2011). The common methods for the deoxygenation of alcohols are divided into two categories: indirect and direct C-O bond cleavages. A well-known indirect protocol is the Barton-McCombie deoxygenation procedure, which involves first converting the alcohols into reactive xanthate intermediates that are subsequently reduced easily with stannane (Barton and McCombie, 1975; Crich and Quintero, 1989; Hartwig, 1983; Robins et al., 1983). Later, other active derivatives such as benzoyl ester and phosphite variants are also used in the radical deoxygenation process (Scheme 1, 1a) (Jordan and Miller, 2012; Lam and Markó, 2008, 2009, 2011; Saito et al., 1986; Zhang and Koreeda, 2004). Another indirect deoxygenation protocol is the ionic process that converts the hydroxyl group into more easily leaving groups, such as OTs, OMs, and halogens (Masamune et al., 1973, 1974; Nguyen et al., 2013) (Scheme 1, 1b). However, these methods suffered from multistep conversions that resulted in poor step efficiency and atom efficiency. The more desirable direct deoxygenation of aliphatic alcohols is a great challenge owing to the strong C-O bond dissociation energy (332.6-468.6 kJ/mol) (Haynes, 2012).

To overcome these issues, single-step direct deoxygenation strategies for alcohols have been developed, catalyzed by transition metals such as Ti, Pd, Ir, Ru, and Mn (Bauer et al., 2017; Ciszek and Fleischer, 2018; Dai and Li, 2016; Diéguez et al., 2010; Huang et al., 2013; Sawadjoon et al., 2013). (Scheme 1, **2a**) However, the requirement of relatively high temperature (>80°C) and the cost of some precious metal catalysts have greatly limited their broad applicability. Alternatively, Doyle developed a photoredox catalysis strategy that could enable the direct C–O bond cleavage of alcohols at room temperature via the phosphorus radical intermediates (Stache et al., 2018). Nevertheless, it still requires the use of expensive iridium

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(1) Indirect deoxygenation of alcohols

(a) Radical deoxygenation	protocol			
$R' = R^1C(S), R^1$	C(O), P(OR) ₂	R^0 R'	Bu₃SnH ►	_R ∕∧ _H
IX ON			or Sml ₂ , HMPA	
(b) Ionic deoxygenation pr	otocol			
R-OHLG = OTs, OM	s, X, etc.	R-LG	Hydrogenation	R=H
Direct deoxygenation of alco	hos			
(a) Metal-catalyzed deoxy	genation protoc	ol		
R-0H	Ti, Pd, Ir, Ru	ı, Mn 🛌	R = H	
(b) This work Metal-free	deoxygenatior	n protocol		
ОН	metal-free	•	H	
R ¹ R ²	hv	F	$R^1 R^2$	

Scheme 1. Strategies for the Deoxygenation of Alcohols

(2)

photoredox catalyst in this protocol. Therefore, the development of mild and efficient direct deoxygenation in a single step, especially without using precious metals, is still highly desirable.

Inspired by our previous work on using N_2H_4 as traceless mediator for homo- and cross-aryl couplings (Lv et al., 2018), and as non-metallic hydrogen-atom-transfer (HAT) reductant for pinacol couplings enabled by light (Qiu et al., 2019), we postulate to use N_2H_4 as clean reductant for the direct deoxygenation of alcohols. In this transformation, N_2 and H_2 are generated as gaseous nontoxic by-products, making the deoxygenation process dramatically clean and easy to handle. We herein disclose the first light-driven metal-free direct deoxygenation of alcohols with hydrazine through Csp³-O bond cleavage (Scheme 1, 2b).

RESULTS AND DISCUSSION

To validate our hypothesis, the direct deoxygenation of (3,4,5-trimethoxyphenyl)methanol (1L) with hydrazine (2) was initially carried out under hv (254 nm) irradiation with DMSO as solvent and KOH as base under air at room temperature. Gratifyingly, the desired product **3**I was obtained in 63% yield after 24 h (Table 1, entry 1). However, other conditions including blue light emitting diode (LED), compact fluorescent lamps (CFL), or without light showed poor activities for this conversion (Table 1, entries 2–4). Next, a variety of bases, including NaOH, *t*-BuOK, and K₂CO₃, were further evaluated (Table 1, entries 5–7). Disappointingly, the efficiency of this transformation did not improve. The reaction showed poor activity when using other solvents such as MeCN, H₂O, and 1,4-dioxane, which indicated that the solvent played an important role in the reaction (Table 1, entries 8–10). The influence of the amount of hydrazine was also investigated, and 4 equiv. of hydrazine was found to be the best choice (Table 1, entries 11–14). In addition, the effluence of reaction time was also studied (Table 1, entries 15–16). When the reaction time was reduced to 18 h, there was no obvious effect on the reaction, whereas the yield was significantly lower when the time was reduced to 12 h. Finally, only 20% of the product was obtained when the reaction (Table 1, entry 17).

With the optimized reaction conditions in hand, the substrate scope of primary benzyl alcohols was investigated as shown in Table 2. To our delight, different benzyl alcohols with electron-donating or electronwithdrawing groups were successfully deoxygenated to afford the corresponding aromatics (3a-3h) in moderate to good yields. 4-(Hydroxymethyl)phenol bearing free hydroxyl could also be smoothly deoxygenated to generate the desired product 3d in 40% yield. The NO₂ group is reduced during the deoxygenation process, as shown for (4-nitrophenyl)methanol, which was converted to (4-aminophenyl)methanol (3f) in 55% yield. Primary alcohols bearing multisubstituted phenyl group were also effective in this transformation to provide the products (3i-3m) in 62%–95% yields. Benzyl alcohols bearing polycyclic (hetero-) aromatic substituents such as indene, dioxole, naphthalene, phenanthrene, benzothiophene, pyridine, and quinolone units could be well tolerated in the reaction (3n–3u). Moreover, easy removal of both hydroxyl groups in o-dibenzyl alcohol was achieved, leading to 1,2,4-trimethylbenzene (3v) in 62% yield when hydrazine was increased to 0.6 mmol. However, benzyl alcohols bearing other electron-withdrawing groups (e.g., 4-CF₃, 4-F, 4-CN) were investigated, showing poor reactivity in this transformation (3w-3y). Various

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	MeO MeO 11	OH N₂H₄⁺H₂C base, solv <i>hv</i> , rt	o (2) Me	O O O Me 3I	
Entry ^a	hv	$N_2H_4 \cdot H_2O$	Base	Solvent	3l Yield ^b /%
1	hv (254 nm)	2	КОН	DMSO	63
2	Blue LED	2	КОН	DMSO	n.p.
3	CFL	2	КОН	DMSO	n.p.
4	Dark	2	КОН	DMSO	n.p.
5	hv (254 nm)	2	NaOH	DMSO	44
6	hv (254 nm)	2	t-BuOK	DMSO	48
7	hv (254 nm)	2	K ₂ CO ₃	DMSO	Trace
8	<i>hv</i> (254 nm)	2	КОН	MeCN	Trace
9	<i>hv</i> (254 nm)	2	КОН	H ₂ O	n.p.
10	<i>hv</i> (254 nm)	2	КОН	1,4-dioxane	Trace
11	hv (254 nm)	0	КОН	DMSO	8
12	hv (254 nm)	1	КОН	DMSO	33
13	hv (254 nm)	4	КОН	DMSO	93
14	hv (254 nm)	5	КОН	DMSO	92
15 ^c	<i>hv</i> (254 nm)	4	КОН	DMSO	93(89)
16 ^d	hv (254 nm)	4	КОН	DMSO	62
17 ^e	hv (254 nm)	4	КОН	DMSO	20

Table 1. Optimization of the Reaction Conditions

^aGeneral conditions: **11** (0.1 mmol), **2** (x equiv.), base (0.2 mmol), and solvent (1 mL) at rt for 24 h under air. ^bYields were determined by ¹HNMR with 1,3,5-trimethoxybenzene as internal standard and isolated yields in brackets. ^c18 h.

^eUnder Ar.

secondary benzyl alcohols were then examined to expand the generality of this system. 1-Phenylethanol and phenylethanols bearing methoxy at different positions all reacted smoothly with hydrazine to afford the corresponding products (**3z-3ab**). 1-Phenylheptan-1-ol and 1,2-diphenylethanol were also competent substrates, delivering the corresponding products (**3ac**, **3ad**) in excellent yields. Additionally, the deoxy-genation of various diaryl alcohols including symmetrical and unsymmetrical substrates proceeded well and gave compounds **3ae–3ag** smoothly. When a bromine-containing substrate was used, the product diphenylmethane (**3ae**) could be obtained by simultaneous cleavages of C–Br and C-O bonds, which was consistent with previous reported results (**Cao et al.**, 2019). The substrates with other aromatic (hetero-) rings such as indene, chroman, and xanthene also tolerated in this reaction to generate the desired products **3ah–3aj**.

The conversion of vicinal diols via C-O cleavage to realize the stoichiometric removal of all (or most) of the oxygen content is greatly significant in organic synthesis because of its prevalence in medicines, agrochemical chemicals, and biomass, especially carbohydrates and lignins. Up to now, a range of reductive deoxygenation methods such as hydrodeoxygenation, hydrogenolysis, decarbonylation, and decarboxylation have been developed (Dethlefsen and Fristrup, 2015). However, these approaches suffer from poor deoxygenation selectivity, resulting in the possibility of forming a mixture of products. To our delight, the

^d12 h.





	R OH + N ₂ H	H ₄ ·H ₂ O <u>hv (254 nm)</u> KOH, DMSO 2 air, rt, 18 h	→ R → H	
1° alcohols ^a				
Et	5(+)0 H	Eto H OEt	НО	MeO ₂ C
3 a 80% ^b	3b 86%	3c 92%	3d 40% ^c	3e 45%, 36 h
H ₂ N H	H OMe	H NH ₂	MeO OMe	MeO
3f 55% ^d	3g 68%	3h 92%	3i 70%	3 j 72%
OMe H OMe	MeO MeO OMe	Me H Me Me	H	O O H
3k 92%	3 I 89%	3m 62%	3n 86%	3o 91%
Н	H	H	K S S	H
3 p 82%	3q 61%	3r 60%	3s 95%	3t 96% ^b
H N Me	Me	F ₃ C	F	NC
3u 80%	3v 62%, 36 h ^e	3w trace 36 h	3x trace 36 h	3y n.p. 36 h
2° alcohols ^a				
Me	MeO	OMe H Me	H () ₄	H C
3z 50%, 36 h ^b	3aa 62%, 36 h	3ab 36%, 36 h	3ac 93%, 36 h	3ad 96%, 36 h
H	Me Me	H	H J J J J J J J J J J J J J J J J J J J	H H
3ae 52%, 36 h	3af 95%, 36 h	3ag 90%, 36 h	3ae 48%, 36 h ^f	3ah 57%, 36 h

Table 2. Substrate Scope of 1° and 2° Alcohols

(Continued on next page)





	R OH + 1	N ₂ H ₄ ·H ₂ O	<i>hv</i> (254 nm) KOH, DMSO air, rt, 18 h	R H 3	
H C O	H O O				
3ai 89%, 36 h	3aj 62%, 36 h				

Table 2. Continued

^aGeneral conditions: 1 (0.1 mmol), 2 (0.4 mmol), KOH (0.2 mmol) in DMSO (1 mL) at room temperature for 18 h under air and isolated yield based on 1. ^bYield was determined by GC-MS.

^cKOH (0.4 mmol).

^d(4-Nitrophenyl)methanol was used as substrate.

^e2 (0.6 mmol) and KOH (0.4 mmol) were used.

^fDebromination product **3ab** was obtained.

selective deoxygenation occurred at the benzyl position to give 2-phenylethanol (Table 3, 3ak) when 1-phenylethane-1,2-diol was tested. To our surprise, when diaryl pinacols were used as the substrates, the simultaneous C-O and C-C bonds both cleavage products (Table 3, 3al, 3am, 3ae) were obtained in good to high yields under the current catalytic system.

The conversion of bio-renewable lignin into sustainable aromatic compounds has attracted widespread attention owing to its natural abundance and the rapid reduction of non-renewable fossil resources (Ragauskas et al., 2014; Singh et al., 2015; Verma et al., 2016; Xu et al., 2014). To evaluate the application potentials of direct deoxygenation reaction, β -1 and β -O-4 model compounds—as important family of natural wood lignin—were tested under the standard reaction conditions (Table 4). Aromatic products including 1-methoxy-4-methylbenzene (**3a**) and 1,2-dimethoxy-4-methylbenzene (**3j**) can be obtained in moderate yields through both C-O and C-C bonds cleavage of β -1 lignin model substrates. Interestingly, the corresponding aromatic (**3z**, **3an**, **3a**) and phenol (**4a**, **4b**) compounds were obtained when β -O-4 lignin model compounds were used as substrates.

In order to gain the mechanistic insights of this transformation, some control experiments were performed (Scheme 2). First, considering that the deoxygenation reaction could only occur with DMSO as the solvent and also an unpleasant odor was sensed when opening the reaction tube, we suspected that dimethyl sulfide gas, generated during the reaction, might have a certain effect on the reaction. Therefore, we used dimethyl sulfide instead of N_2H_2 as the reducing agent for this deoxygenation reaction, and the desired product was obtained in 81% yield. The result indicated that dimethyl sulfide intermediate may be produced from DMSO in the presence of hydrazine, which played a crucial role in the reduction process. Second, we performed the reaction with N_2D_4 + D_2O under the standard conditions, and the deoxygenation product *d*-3I was obtained in 90% yield with about 65% deuteration at benzyl position. Third, free radical trap experiments were performed: when 1 or 3 equiv. of TEMPO (2,2,6,6-tetramethyl-1-piperidinyloxy) were added into the reaction products, demonstrating an unlikely radical pathway in this transformation. Finally, with 3,4,5-trimethoxybenzaldehyde as raw material, only a small amount of the corresponding product was obtained in the presence of hydrazine or dimethyl sulfide, whereas by-products 8 and 9 were generated in 65% and 78% yields, showing that aldehyde may not be an intermediate in this reaction.

Based on the mechanistic studies above, we proposed a plausible reaction mechanism as shown in Scheme 3. Initially, dimethyl sulfide is *in situ* generated from the reaction of DMSO with hydrazine under the promotion of light, meanwhile releasing N₂, H₂, and H₂O (Kim and Lee, 2018; Smit et al., 2008). Considering the importance of O₂ in this reaction and in view of previous studies (Ishiguro et al., 1996; Liang et al., 1983), dimethyl sulfide then interacts with light-excited O₂ to form intermediate **A**, which reacts with alcohols **1** in the presence of base to provide the species **B**. The release of O₂ and DMSO from **B** results in the



Table 3. Substrate Scope of Vicinal Diols

^aGeneral conditions: 1 (0.1 mmol), 2 (0.8 mmol), KOH (0.4 mmol) in DMSO (1 mL) at room temperature for 36 h under air and isolated yield based on 1.

^bSymmetrical substrate, total yield doubles.

carbanion intermediate C. Finally, hydrogen proton abstraction from H_2O by carbanion C gives the deoxygenation product 3.

It is particularly noteworthy for the unprecedented role of dimethylsulfide in this novel alcohol deoxygenation reaction. On the other hand, the amount of dimethyl sulfide generated from the oceans and microorganisms is about 38–40 million metric tons per annum (Chasteen and Bentley, 2004). Moreover, dimethyl sulfide is a by-product of many reactions such as the Kornblum oxidation, the Moffatt oxidation, the Parikh-Doering oxidation, and the Swern oxidation. Thus, we wondered the possibility of using dimethyl sulfide for alcohol deoxygenation. Indeed, we found that a simple and generally applicable methodology for the direct deoxygenation of alcohols (0.1 mmol) can be achieved with dimethyl sulfide (2 equiv.) in DMSO (1 mL) enabled by light at room temperature under air. Different alcohols including primary and secondary benzyl alcohols (Table 5) all gave the products of Csp³-O bond cleavage in good yields. The reactions of dimethyl sulfide as reducing agent are ongoing in our laboratory and will be reported in due course.

Conclusions

In conclusion, we described a photo-induced metal-free direct deoxygenation of alcohols with hydrazine at room temperature under air atmosphere. The fundamental innovation of this strategy is that traceless non-toxic N₂ and H₂ are generated as by-products, making the direct deoxygenation dramatically clean. This protocol is functional-group tolerant and selective for 1°, 2° alcohols and vicinal diols. Importantly, β -1 and β -O-4 lignin model compounds displayed exceptional reactivity, producing the corresponding aromatics and phenols through C-O/C-C bonds cleavage, which provides a potential method for converting lignin and its model compounds into useful chemicals. Moreover, we successfully used dimethyl sulfide

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Table 4. Substrate Scope of $\beta\textsc{--}1$ and $\beta\textsc{-}O\textsc{--}4$ Lignin Model Compounds

^aGeneral conditions: **1** (0.1 mmol), **2** (0.8 mmol), KOH (0.4 mmol) in DMSO (1 mL) at room temperature for 36 h under air and isolated yield based on **1**. ^bSubstrate with the same substituent on both aromatic rings, double product yield. ^cYield was determined by GC-MS.

instead of hydrazine as the reducing agent for such alcohol deoxygenation reaction. Further studies on the mechanism and synthetic applications of this protocol are undergoing in our laboratory.

Limitations of the Study

Our direct deoxygenation of alcohols works well for most of the tested substrates; however, primary benzyl alcohols bearing electron-withdrawing groups (e.g., 4-CF₃, 4-F, 4-CN) showed poor reactivity in this transformation. In addition, the detailed mechanism of simultaneous C-O and C-C bonds cleavage process is still not clear.

Resource Availability

Lead Contact

Further information and requests for resources and reagents should be directed to and will be fulfilled by the Lead Contact, Chao-Jun Li (cj.li@mcgill.ca).

Materials Availability

All unique/stable reagents generated in this study are available from the Lead Contact without restriction.







Scheme 2. Control Experiments

Data and Code Availability

All relevant data supporting the findings of this study are available within the paper and its Supplemental Information files. Additional data are provided upon reasonable request.

METHODS

All methods can be found in the accompanying Transparent Methods supplemental file.

SUPPLEMENTAL INFORMATION

Supplemental Information can be found online at https://doi.org/10.1016/j.isci.2020.101419.

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Scheme 3. The Plausible Mechanism for the Direct Dehydroxylation of Alcohols

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Table 5. Substrate Scope of Alcohols with Dimethyl Sulfide

General conditions: 1 (0.1 mmol), (CH₃)₂S (0.2 mmol), KOH (0.2 mmol) in DMSO (1 mL) at room temperature for 24 h under air. Isolated yield based on 1.

AUTHOR CONTRIBUTIONS

D.C., Y.P., and C.-J.L. conceived and designed the project. D.C. conducted the experiments, analyzed the data, and composed the manuscript. Z.C., L.L., H.Z., and Y.P. discussed the experimental results and commented on the manuscript. C.-J.L. conducted general guidance, project directing, and manuscript revisions.

DECLARATION OF INTERESTS

The authors declare no competing interests.

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Supplemental Information

Light-Driven Metal-Free Direct

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Light-Driven Metal-Free Direct Deoxygenation of Alcohols under Mild Conditions

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Supplemental Figures for NMR spectra:

Figure S1. ¹H NMR spectrum of compound 1-(hexyloxy)-4-methylbenzene (3b), related to Table 2, Table 5.



Figure S2. ¹³C NMR spectrum of compound 1-(hexyloxy)-4-methylbenzene (3b), related to Table 2, Table 5.



Figure S3. ¹H NMR spectrum of compound **1-(diethoxymethyl)-4-methylbenzene (3c),** related to **Table 2**, **Table 5**.



Figure S4. ¹³C NMR spectrum of compound 1-(diethoxymethyl)-4-methylbenzene (3c), related to Table 2, Table 5.

-137.85 -136.12 -136.12 -128.75 -126.48 -101.51 -101.51 -101.51 -101.51 -101.51 -101.51

-21.11 -15.14







Figure S6. ¹³C NMR spectrum of compound *p*-cresol (3d), related to Table 2.





Figure S7. ¹H NMR spectrum of compound methyl 4-methylbenzoate (3e) , related to Table 2.





Figure S11. ¹H NMR spectrum of compound **1-methoxy-2-methylbenzene (3g)**, related to **Table 2**.



Figure S14. ¹³C NMR spectrum of compound *o*-toluidine (3h), related to Table 2.



Figure S13. ¹H NMR spectrum of compound *o*-toluidine (3h), related to Table 2.

Figure S15. ¹H NMR spectrum of compound **1,3-dimethoxy-5-methylbenzene (3i)**, related to **Table 2**.



Figure S17. ¹H NMR spectrum of compound **1,2-dimethoxy-4-methylbenzene (3j)**, related to **Table 2**.



-10 200 190 170 160 150 140

3.81 -2.24 -7.28 6.79 6.77 6.77 6.72 6.72 6.70 6.70 H₃C _CH3 CH-2.02 3.04 3.00-9.5 9.0 8.5 8.0 7.5 7.0 6.5 6.0 5.5 5.0 4.5 4.0 3.5 3.0 2.5 2.0 1.5 1.0 0.5 0.0 -0.5 12.5 11.5 10.5 Figure S20. ¹³C NMR spectrum of compound **1,4-dimethoxy-2-methylbenzene (3k),** related to **Table 2**. -127.82 153.34 76.75 55.90 -16.38 H₃C CH-CH. 210 -10 200 190 40 180 170 160 150 140 130 120 110 100 60 50 30 20 10 0

Figure S19. ¹H NMR spectrum of compound **1,4-dimethoxy-2-methylbenzene (3k)**, related to **Table 2**.

Figure S21. ¹H NMR spectrum of compound **1,2,3-trimethoxy-5-methylbenzene (3I)**, related to **Table 1**, **Table 2**, **Table 5**.



Figure S22. ¹³C NMR spectrum of compound **1,2,3-trimethoxy-5-methylbenzene (3I)**, related to **Table 1**, **Table 2**, **Table 5**.



Figure S23. ¹H NMR spectrum of compound **1,2,3,5-tetramethylbenzene (3m)**, related to Table 2.





H₃C



Figure S25. ¹H NMR spectrum of compound 5-methyl-2,3-dihydro-1*H*-indene (3n), related to Table 2.

Figure S27. ¹H NMR spectrum of compound 5-methylbenzo[*d*][1,3]dioxole (30), related to Table 2, Table 5.



Figure S28. ¹³C NMR spectrum of compound 5-methylbenzo[*d*][1,3]dioxole (3o), related to Table 2, Table 5.









Figure S30. ¹³C NMR spectrum of compound 2-methylnaphthalene (3p), related to Table 2.





-10 200 190 180 170 160 150 140 130 120

Figure S33. ¹H NMR spectrum of compound 9-methylphenanthrene (3r), related to Table 2.



Figure S34. ¹³C NMR spectrum of compound 9-methylphenanthrene (3r), related to Table 2.





Figure S36. ¹³C NMR spectrum of compound 2-methylbenzo[*b*]thiophene (3s), related to Table 2, Table 5.



Figure S35. ¹H NMR spectrum of compound 2-methylbenzo[*b*]thiophene (3s), related to Table 2, Table 5.







Figure S39. ¹H NMR spectrum of compound **1,2,4-trimethylbenzene (3v)**, related to **Table 2**.





Figure S41. ¹H NMR spectrum of compound 1-ethyl-3-methoxybenzene (3aa), related to Table 2.





Figure S45. ¹H NMR spectrum of compound hexylbenzene (3ac), related to Table 2, Table 5.



 200 190 180 170 160 150 140 130 120 110 100

Figure S47. ¹H NMR spectrum of compound 1,2-diphenylethane (3ad), related to Table 2.





-10

200 190 180 170 160 150 140 130 120 110 100

7.14 7.14 7.13 7.13 7.13 7.11 7.11 -3.95 -2.36 H₃C CH₂ 8.07-2.04 - € 6.00-12.5 9.5 9.0 8.5 8.0 7.5 7.0 6.5 6.0 5.5 5.0 4.5 4.0 3.5 3.0 2.5 2.0 1.5 1.0 0.5 0.0 -0.5 11.5 10.5 Figure S52. ¹³C NMR spectrum of compound di-*p*-tolylmethane (3af), related to Table 2, Table 5. -138.34-135.40 $\angle 129.09$ $\angle 128.72$ 77.25 77.00 76.75 -41.06 -20.99 CH3 210 -10 200 190 180 170 160 150 140 130 120 110 100 20 0

10

Figure S51. ¹H NMR spectrum of compound di-*p*-tolylmethane (3af), related to Table 2, Table 5.















Figure S63. ¹H NMR spectrum of compound **1-methoxy-4-methylbenzene (3al)**, related to **Table 3**, **Table 4**.



Figure S64. ¹³C NMR spectrum of compound 1-methoxy-4-methylbenzene (3al), related to Table 3, Table 4.



7.13 7.13 7.12 6.77 6.75 2.96 -2.33 ,CH3 H₃C ĊH. 2.00-€ 2.00-€ 6.03-3.05-= 9.5 9.0 8.5 8.0 7.5 7.0 6.5 6.0 5.5 5.0 4.5 4.0 3.5 3.0 2.5 2.0 1.5 1.0 0.5 0.0 -0.5 12.5 11.5 10.5 Figure S66. ¹³C NMR spectrum of compound *N*,*N*,4-trimethylaniline (3am), related to Table 3. -126.04-126.04-113.18-113.1877.25777.2576.75-41.03 -20.21







Figure S67. ¹H NMR spectrum of compound 1-ethyl-4-methoxybenzene (3an), related to Table 4.







Figure S73. UV reactor, related to Table 1-5 and Scheme 1-3.



Transparent methods:

(1) General methods

All reagents and solvents were purchased from commercial sources (Alfa, Acros, Aldrich, TCI and Combi-Blocks) and used without further purification unless otherwise stated. ¹H, ¹⁹F and ¹³C NMR spectra were taken on Bruker 400 or 500 MHz spectrometer. Chemical shifts of ¹H NMR spectra were reported using either residual solvent signal of CDCl₃ (δ = 7.26 ppm) or TMS (δ = 0.00 ppm) as internal standard. Chemical shifts of ¹³C NMR spectra were reported using residual solvent signal of CDCl₃ (δ = 7.26 ppm) or TMS (δ = 0.00 ppm) as internal standard. Chemical shifts of ¹³C NMR spectra were reported using residual solvent signal of CDCl₃ (δ = 77.16 ppm) as internal standard. The peak patterns are indicated as follows: s, singlet; d, doublet; dd, doublet of doublet; t, triplet; q, quartet; m, multiplet. The coupling constants, J, are reported in Hertz (Hz). All reactions were monitored by thin-layer chromatography (TLC). Column chromatography was performed on silica gel (200-300 mesh) and visualized with ultraviolet light. Hydrazine hydrate-d6 was purchased from Toronto Research Chemicals. EI-MS was obtained from the Agilent GC-MS system. All solvents were purified and dried by standard techniques.

(2) General experimental procedure and spectroscopic data of products

In 15 mL quartz tube was charged with a magnetic stir-bar, were added sequentially alcohol (0.1 mmol, 1 equiv), N₂H₄•H₂O (0.4 mmol, 4 equiv) and DMSO (1 mL) under air. Then the tube was placed in a UV reactor (**Figure S73**) (Qiu et al., 2019) at room temperature and the mixture was stirred for 24 or 36 h. 10 mL water was added to quench reaction, and the mixture was extracted with EtOAc (5 mL × 4). The combined organic solvent was washed with brine, dried with anhydrous Na₂SO₄, and then concentrated under reduced pressure. The residues were purified by preparative TLC on silica gel eluting with hexane: EtOAc (300:1-20:1) to afford the product. Due to the volatility of most deoxygenated alkanes (**3a,3t,3w**), 1,3,5-trimethoxybenzene (16.8 mg, 0.1 mmol) was directly added into the reaction mixture as an internal standard upon completion of the reaction. A small amount of reaction mixture (roughly 20 μ L) was filtered through a short plug made of neutral Al₂O₃ and anhydrous Na₂SO₄, flushed by CDCl₃ (0.7 mL), analyzed by GC-MS, and subjected to ¹H NMR for yield determination. (**Note**: Considering safety issues, the reaction can also be conducted as following: a short 1 mm diameter syringe needle pierced through the rubber cap of the reaction tube to keep the reaction mixture in contact with the atmosphere and prevent the rapid evaporation of the volatile compounds simultaneously. Thus, the reaction could proceed smoothly without safety problems).



1-(Hexyloxy)-4-methylbenzene (3b) (CAS: 57792-40-2) (Sutter et al., 2013) Yield: 86% (16.5 mg) using hydrazine as reducing reagent;

Yield: 83% (16.0 mg) using dimethyl sulfide as reducing reagent.

¹H NMR (CDCl₃, 500 MHz) δ : 7.10 (d, J = 8.2 Hz, 2H), 6.82 (d, J = 8.6 Hz, 2H), 3.95 (t, J = 6.6 Hz, 2H), 2.31 (s, 3H), 1.79 (dd, J = 8.2, 7.0 Hz, 2H), 1.48 (dd, J = 10.0, 4.9 Hz, 2H), 1.38 – 1.35 (m, 4H), 0.93 (dd, J = 9.2, 4.8 Hz, 3H).

¹³C NMR (CDCl₃, 126 MHz) δ: 157.0, 129.8, 129.6, 114.3, 68.1, 31.6, 29.3, 25.7, 22.6, 20.4, 14.0.



1-(Diethoxymethyl)-4-methylbenzene (3c) (CAS: 2403-59-0) (Ugarte et al., 2017) **Yield: 92% (17.9 mg) using hydrazine as reducing reagent; Yield: 85% (16.5 mg) using dimethyl sulfide as reducing reagent.** ¹**H NMR (CDCl₃, 500 MHz)** δ: 7.41 (d, J = 8.0 Hz, 2H), 7.21 (d, J = 7.9 Hz, 2H), 5.52 (s, 1H), 3.66 (dq, J = 9.4, 7.1 Hz, 2H), 3.60 – 3.53 (m, 2H), 2.39 (s, 3H), 1.28 (t, J = 7.1 Hz, 6H). ¹³C NMR (CDCl₃, 126 MHz) δ: 137.9, 136.1, 128.8, 126.5, 101.5, 60.8, 21.1, 15.1.



p-Cresol (3d) (CAS: 106-44-5) (Wang et al., 2013)

Yield: 40% (4.3 mg).

¹H NMR (CDCl₃, 500 MHz) δ: 7.07 (d, J = 7.9 Hz, 2H), 6.77 (d, J = 8.4 Hz, 2H), 4.92 (s, 1H), 2.31 (s, 3H); ¹³C NMR (CDCl₃, 126 MHz) δ: 153.1, 130.1, 130.0, 115.1, 20.4.



Methyl 4-methylbenzoate (3e) (CAS: 99-75-2) (Diéguez et al., 2010)

Yield: 45% (6.8 mg).

¹H NMR (CDCl₃, 500 MHz) δ: 7.94 (d, J = 8.2 Hz, 2H), 7.23 (dd, J = 7.9, 0.4 Hz, 2H), 3.90 (s, 3H), 2.39 (s, 3H).

¹³C NMR (CDCl₃, **126** MHz) δ: 167.0, 143.4, 129.4, 128.9, 127.3, 51.7, 21.4.



p-Toluidine (3f) (CAS: 106-49-0) (Yan et al., 2020)

Yield: 55% (5.9 mg).

¹H NMR (CDCl₃, 500 MHz) δ: 7.00 (d, J = 8.0 Hz, 2H), 6.64 (d, J = 8.3 Hz, 2H), 3.55 (s, 2H), 2.28 (s, 3H). ¹³C NMR (CDCl₃, 126 MHz) δ: 143.8, 129.7, 127.7, 115.2, 20.4.



1-Methoxy-2-methylbenzene (3g) (CAS: 578-58-5) (Huang et al., 2013) **Yield: 68% (8.3 mg).** ¹H NMR (CDCl₃, 500 MHz) δ: 7.23 – 7.16 (m, 2H), 6.90 (ddd, J = 13.7, 10.1, 4.5 Hz, 2H), 3.87 (s, 3H),

2.27 (s, 3H).

¹³C NMR (CDCl₃, 126 MHz) δ: 157.7, 130.6, 126.8, 126.6, 120.2, 109.9, 55.2, 16.2.



o-Toluidine (3h) (CAS: 95-53-4) (Yan et al., 2020)

Yield: 92% (9.9 mg).

¹H NMR (CDCl₃, 500 MHz) δ: 7.13 (t, J = 7.7 Hz, 2H), 6.80 (t, J = 7.4 Hz, 1H), 6.75 (d, J = 7.7 Hz, 1H), 3.64 (s, 2H), 2.25 (s, 3H).

 $^{13}\text{C}\,\text{NMR}\,\text{(CDCl}_3\text{, 126 MHz)}\,\delta\text{:}\,144.5\text{,}\,130.3\text{,}\,126.8\text{,}\,122.2\text{,}\,118.5\text{,}\,114.8\text{,}\,17.2\text{.}$



1,3-Dimethoxy-5-methylbenzene (3i) (CAS: 4179-19-5) (Xi et al., 2018)
Yield: 70% (10.7 mg).
¹H NMR (CDCl₃, 500 MHz) δ: 6.36 (s, 2H), 6.31 (s, 1H), 3.80 (s, 6H), 2.33 (s, 3H).
¹³C NMR (CDCl₃, 126 MHz) δ: 160.7, 140.2, 107.1, 97.5, 55.2, 21.8.



1,2-Dimethoxy-4-methylbenzene (3j) (CAS: 494-99-5) (Huang et al., 2013)

Yield: 72% (11.0 mg) using (3,4-dimethoxyphenyl)methanol as substrate;

Yield: 30% (4.6 mg) using 2-(3,4-dimethoxyphenyl)-1-(4-methoxyphenyl)propane-1,3-diol as substrate.

¹H NMR (CDCl₃, 500 MHz) δ: 6.79 (d, J = 8.5 Hz, 1H), 6.75 – 6.71 (m, 2H), 3.88 (s, 3H), 3.87 (s, 3H), 2.33 (s, 3H).

¹³C NMR (CDCl₃, 126 MHz) δ: 148.6, 146.8, 130.3, 120.7, 112.3, 111.2, 55.8, 55.6, 20.9.



1,4-Dimethoxy-2-methylbenzene (3k) (CAS: 24599-58-4) (Jiang et al., 2014) **Yield: 92% (14.0 mg).**

¹H NMR (CDCl₃, 500 MHz) δ: 6.77 (t, J = 5.9 Hz, 2H), 6.71 (dd, J = 8.7, 3.0 Hz, 1H), 3.81 (s, 3H), 3.79 (s, 3H), 2.24 (s, 3H).

¹³C NMR (CDCl₃, 126 MHz) δ: 153.3, 152.0, 127.8, 117.0, 110. 9, 110.7, 55.9, 55.7, 16.4.



1,2,3-Trimethoxy-5-methylbenzene (3l) (CAS: 6443-69-2) (Diéguez et al., 2010)
 Yield: 89% (16.2 mg) using hydrazine as reducing reagent;
 Yield: 81% (14.8 mg) using dimethyl sulfide as reducing reagent.
 ¹H NMR (CDCl₃, 500 MHz) δ: 6.42 (s, 2H), 3.87 (s, 6H), 3.84 (s, 3H), 2.34 (s, 3H).
 ¹³C NMR (CDCl₃, 126 MHz) δ: 153.0, 135.8, 133.6, 105.9, 60.9, 56.0, 21.8.



1,2,3,5-Tetramethylbenzene (3m) (CAS: 527-53-7) (Ding et al., 2019) Yield: 62% (8.3 mg).

¹H NMR (CDCl₃, 500 MHz) δ: 6.86 (s, 2H), 2.28 (s, 9H), 2.16 (s, 3H).
 ¹³C NMR (CDCl₃, 126 MHz) δ: 136.2, 134.5, 131.8, 128.3, 20.8, 20.4, 14.9.



5-Methyl-2,3-dihydro-1*H*-indene (3n) (CAS: 874-35-1) (Adamczyk et al., 1984) Yield: 86% (11.4 mg).

¹H NMR (CDCl₃, 500 MHz) δ: 7.15 (d, J = 7.5 Hz, 1H), 7.09 (s, 1H), 6.98 (d, J = 7.5 Hz, 1H), 2.90 (t, J = 7.3 Hz, 4H), 2.35 (s, 3H), 2.11 – 2.07 (m, 2H).

¹³C NMR (CDCl₃, 126 MHz) δ: 144.3, 141.1, 135.5, 126.7, 125.1, 124.0, 32.8, 32.4, 25.6, 21.2.



5-Methylbenzo[d][1,3]dioxole (3o) (CAS: 7145-99-5) (Huang et al., 2013) Yield: 91% (12.4 mg) using hydrazine as reducing reagent; Yield: 84% (11.4 mg) using dimethyl sulfide as reducing reagent.

¹H NMR (CDCl₃, 500 MHz) δ: 6.73 (d, J = 7.9 Hz, 1H), 6.69 (s, 1H), 6.64 (d, J = 7.9 Hz, 1H), 5.93 (s, 2H), 2.30 (s, 3H).

¹³C NMR (CDCl₃, **126** MHz) δ: 147.4, 145.2, 131.5, 121.5, 109.6, 108.0, 100.7, 21.2.



2-Methylnaphthalene (3p) (CAS: 91-57-6) (Zhu et al., 2018)

Yield: 82% (11.7 mg).

¹H NMR (CDCl₃, 500 MHz) δ: 7.82 (d, J = 7.8 Hz, 1H), 7.78 (dd, J = 8.0, 4.8 Hz, 2H), 7.64 (s, 1H), 7.45 (dtd, J = 14.6, 6.9, 1.3 Hz, 2H), 7.34 (dd, J = 8.4, 1.6 Hz, 1H), 2.54 (s, 3H).

¹³C NMR (CDCl₃, 126 MHz) δ: 135.4, 133.6, 131.7, 128.1, 127.7, 127.6, 127.2, 126.8, 125.8, 124.9, 21.7.



1-Methylnaphthalene (3q) (CAS: 90-12-0) (Zhu et al., 2018)

Yield: 61% (8.7 mg).

¹H NMR (CDCl₃, 500 MHz) δ: 8.09 – 8.04 (m, 1H), 7.91 (dd, J = 8.3, 1.0 Hz, 1H), 7.78 (d, J = 8.1 Hz, 1H), 7.57 (dddd, J = 18.1, 8.0, 6.8, 1.4 Hz, 2H), 7.47 – 7.41 (m, 1H), 7.39 (d, J = 6.9 Hz, 1H), 2.77 (s, 3H).
 ¹³C NMR (CDCl₃, 126 MHz) δ: 134.2, 133.5, 132.6, 128.5, 126.5, 126.3, 125.7, 125.5 (2C), 124.1, 19.6.



9-Methylphenanthrene (3r) (CAS: 883-20-5) (Zhu et al., 2018)

Yield: 60% (11.5 mg).

¹H NMR (CDCl₃, 500 MHz) δ: 8.78 – 8.75 (m, 1H), 8.69 (d, J = 8.0 Hz, 1H), 8.10 (dd, J = 7.0, 2.4 Hz, 1H), 7.87 – 7.82 (m, 1H), 7.72 – 7.66 (m, 2H), 7.66 – 7.59 (m, 3H), 2.78 (s, 3H).

¹³C NMR (CDCl₃, **126** MHz) δ: 132.5, 132.1, 132.0, 130.4, 129.7, 127.8, 126.7, 126.6, 126.5, 126.2, 125.8, 124.7, 123.0, 122.4, 20.0.



2-Methylbenzo[b]thiophene (3s) (CAS: 1195-14-8) (Urban et al., 2012) Yield: 95% (14.1 mg) using hydrazine as reducing reagent; Yield: 88% (13.0 mg) using dimethyl sulfide as reducing reagent.

¹H NMR (CDCl₃, 500 MHz) δ: 7.77 (d, J = 8.0 Hz, 1H), 7.67 (d, J = 7.8 Hz, 1H), 7.32 (td, J = 7.6, 1.1 Hz, 1H), 7.29 – 7.25 (m, 1H), 7.00 (s, 1H), 2.62 (s, 3H).

¹³C NMR (CDCl₃, 126 MHz) δ: 140.8, 140.4, 139.7, 124.0, 123.3, 122.5, 122.0, 121.6, 16.1.



2,4-Dimethylquinoline (3u) (CAS: 1198-37-4) (Jin et al., 2015)

Yield: 96% (15.1 mg) using hydrazine as reducing reagent;

Yield: 76% (11.9 mg) using dimethyl sulfide as reducing reagent.

¹H NMR (CDCl₃, 500 MHz) δ: 8.04 (d, J = 8.4 Hz, 1H), 7.97 (d, J = 8.3 Hz, 1H), 7.71 – 7.67 (m, 1H), 7.54 – 7.50 (m, 1H), 7.16 (s, 1H), 2.72 (s, 3H), 2.69 (s, 3H).

¹³C NMR (CDCl₃, 126 MHz) δ: 158.6, 147.7, 144.1, 129.1, 129.1, 126.5, 125.4, 123.6, 122.7, 25.2, 18.6.



1,2,4-Trimethylbenzene (3v) (CAS: 95-63-6) (Dalling et al., 1977)

Yield: 62% (7.5 mg).

¹H NMR (CDCl₃, 500 MHz) δ: 7.07 (d, J = 7.6 Hz, 1H), 7.01 (s, 1H), 6.96 (d, J = 7.6 Hz, 1H), 2.34 (s, 3H), 2.28 (d, J = 3.9 Hz, 6H).

¹³C NMR (CDCl₃, 126 MHz) δ: 136.3, 135.1, 133.3, 130.4, 129.5, 126.3, 20.9, 19.7, 19.2.



1-Ethyl-3-methoxybenzene (3aa) (CAS: 10568-38-4) (Tietze et al., 2009)

Yield: 62% (8.4 mg).

¹H NMR (CDCl₃, 500 MHz) δ: 7.23 (t, J = 7.8 Hz, 1H), 6.82 (d, J = 7.5 Hz, 1H), 6.79 – 6.74 (m, 2H), 3.83 (s, 3H), 2.66 (q, J = 7.6 Hz, 2H), 1.26 (t, J = 7.6 Hz, 3H).

¹³C NMR (CDCl₃, 126 MHz) δ: 159.6, 145.9, 129.2, 120.3, 113.7, 110.8, 55.1, 28.9, 15.5.



1-Ethyl-2-methoxybenzene (3ab) (CAS: 14804-32-1) (Wang et al., 2013) Yield: 36% (4.9 mg).

¹H NMR (CDCl₃, 500 MHz) δ: 7.22 – 7.16 (m, 2H), 6.93 (td, J = 7.4, 0.8 Hz, 1H), 6.87 (d, J = 8.1 Hz, 1H), 3.86 (s, 3H), 2.67 (d, J = 7.5 Hz, 2H), 1.22 (t, J = 7.5 Hz, 3H).

¹³C NMR (CDCl₃, 126 MHz) δ: 157.3, 132.6, 128.9, 126.7, 120.4, 110.1, 55.2, 23.2, 14.1.



Hexylbenzene (3ac) (CAS: 1077-16-3) (Hu et al., 2019)

Yield: 93% (15.1 mg) using hydrazine as reducing reagent;

Yield: 81% (13.1 mg) using dimethyl sulfide as reducing reagent.

¹H NMR (CDCl₃, 500 MHz) δ: 7.33 – 7.28 (m, 2H), 7.21 – 7.18 (m, 3H), 2.63 (d, J = 10.2, 2H), 1.67 – 1.60 (m, 2H), 1.38 – 1.29 (m, 6H), 0.91 (t, J = 6.8 Hz, 3H).

¹³C NMR (CDCl₃, 126 MHz) δ: 142.9, 128.4, 128.2, 125.5, 36.0, 31.7, 31.5, 29.0, 22.6, 14.1.



1,2-Diphenylethane (3ad) (CAS: 103-29-7) (Hu et al., 2019) **Yield: 96% (17.5 mg).**

¹H NMR (CDCl₃, 500 MHz) δ: 7.33 (t, J = 7.5 Hz, 4H), 7.24 (t, J = 7.2 Hz, 6H), 2.97 (s, 4H). ¹³C NMR (CDCl₃, 126 MHz) δ: 141.8, 128.4, 128.3, 125.9, 37.9.



Diphenylmethane (3ae) (CAS: 101-81-5) (Wang et al., 2013) Yield: 52% (8.7 mg) using diphenylmethanol as substrate; Yield: 48% (8.1 mg) using (4-bromophenyl)(phenyl)methanol as substrate; Yield: 160% (26.9 mg) using 1,1,2,2-tetraphenylethane-1,2-diol as substrate. ¹H NMR (CDCl₃, 500 MHz) δ: 7.35 – 7.30 (m, 4H), 7.24 (t, J = 6.7 Hz, 6H), 4.03 (s, 2H). ¹³C NMR (CDCl₃, 126 MHz) δ: 141.1, 128.9, 128.4, 126.0, 41.9.



Di-*p*-tolylmethane (3af) (CAS: 4957-14-6) (Wang et al., 2013) Yield: 95% (18.6 mg) using hydrazine as reducing reagent; Yield: 83% (16.3 mg) using dimethyl sulfide as reducing reagent. ¹H NMR (CDCl₃, 500 MHz) δ: 7.15 – 7.09 (m, 8H), 3.95 (s, 2H), 2.36 (s, 6H). ¹³C NMR (CDCl₃, 126 MHz) δ: 138.3, 135.4, 129.1, 128.7, 41.1, 21.0.



1-Benzyl-4-methylbenzene (3ag) (CAS: 620-83-7) (Guan et al., 2014)

Yield: 90% (16.4 mg).

¹H NMR (CDCl₃, 500 MHz) δ: 7.27 (d, J = 7.6 Hz, 2H), 7.18 (d, J = 7.5 Hz, 3H), 7.09 (s, 4H), 3.95 (s, 2H), 2.32 (s, 3H).

¹³C NMR (CDCl₃, 126 MHz) δ: 141.4, 138.0, 135.5, 129.1, 128.8, 128.8, 128.4, 126.0, 41.5, 21.0.



2,3-Dihydro-1*H***-indene (3ah) (CAS: 496-11-7)** (Huang et al., 2013) **Yield: 57% (6.7 mg).** ¹**H NMR (CDCh: 500 MHz)** δ: 7 30 – 7 26 (m. 2H), 7 18 (dd. L = 5 5, 3 2 H

¹H NMR (CDCl₃, 500 MHz) δ: 7.30 – 7.26 (m, 2H), 7.18 (dd, J = 5.5, 3.2 Hz, 2H), 2.96 (t, J = 7.4 Hz, 4H), 2.15 – 2.08 (m, 2H).

¹³C NMR (CDCl₃, 126 MHz) δ: 144.1, 125.9, 124.3, 32.9, 25.3.



Chroman (3ai) (CAS: 493-08-3) (Meng et al., 2019) Yield: 89% (11.9 mg) using hydrazine as reducing reagent; Yield: 80% (10.7 mg) using dimethyl sulfide as reducing reagent. ¹H NMR (CDCl₃, 500 MHz) δ: 7.19 – 7.13 (m, 1H), 7.11 (d, J = 7.5 Hz, 1H), 6.90 (ddd, J = 12.4, 9.4, 4.6 Hz, 2H), 4.30 – 4.21 (m, 2H), 2.86 (t, J = 6.5 Hz, 2H), 2.11 – 2.02 (m, 2H).
 ¹³C NMR (CDCl₃, 126 MHz) δ: 154.8, 129.7, 127.1, 122.1, 120.0, 116.6, 66.3, 24.8, 22.3.



9H-xanthene (3aj) (CAS: 92-83-1) (Sousa et al., 2016) Yield: 62% (11.3 mg).

¹H NMR (CDCl₃, 500 MHz) δ: 7.22 (dd, J = 16.5, 7.8 Hz, 4H), 7.07 (dd, J = 15.7, 7.9 Hz, 4H), 4.09 (s, 2H). ¹³C NMR (CDCl₃, 126 MHz) δ: 151.9, 128.9, 127.6, 122.9, 120.6, 116.4, 27.9.



2-Phenylethanol (3ak) (CAS: 60-12-8) (Behera et al., 2020)

Yield: 76% (9.3 mg).

¹H NMR (CDCl₃, 500 MHz) δ: 7.37 – 7.32 (m, 2H), 7.27 (t, J = 6.3 Hz, 3H), 3.89 (t, J = 6.6 Hz, 2H), 2.90 (t, J = 6.6 Hz, 2H), 1.60 (s, 1H).

¹³C NMR (CDCl₃, 126 MHz) δ: 138.4, 129.0, 128.6, 126.5, 63.7, 39.2.



1-Methoxy-4-methylbenzene (3al) (CAS: 104-93-8) (Huang et al., 2013)

Yield: 126% (15.4 mg) using 1,2-bis(4-methoxyphenyl)ethane-1,2-diol as substrate.

Yield: 86% (10.5 mg) using (1R,2R)-1,2-bis(4-methoxyphenyl)propane-1,3-diol as substrate;

Yield: 45% (5.5 mg) using (1R,2R)-2-(3,4-dimethoxyphenyl)-1-(4-methoxyphenyl)propane-1,3-diol as substrate;

Yield: 32% (3.9 mg) using 2-(4-methoxyphenoxy)-1-(4-methoxyphenyl)propane-1,3-diol as substrate. ¹H NMR (CDCl₃, 500 MHz) δ: 7.15 (dd, J = 8.0, 0.5 Hz, 2H), 6.87 (dd, J = 8.0, 0.5 Hz, 2H), 3.84 (s, 3H), 2.36 (s, 3H).

¹³C NMR (CDCl₃, **126** MHz) δ: 157.4, 129.8, 129.8, 113.7, 55.2, 20.4.



N,N,4-Trimethylaniline (3am) (CAS: 99-97-8) (Yang et al., 2018)

Yield: 122% (16.5 mg).

¹H NMR (CDCl₃, 500 MHz) δ: 7.12 (d, J = 8.1 Hz, 2H), 6.76 (d, J = 8.4 Hz, 2H), 2.96 (d, J = 0.7 Hz, 6H), 2.33 (s, 3H).

¹³C NMR (CDCl₃, 126 MHz) δ: 148.8, 129.5, 126.0, 113.2, 41.0, 20.2.



1-Ethyl-4-methoxybenzene (3an) (CAS: 1515-95-3) (Huang et al., 2013) Yield: 43% (5.9 mg).

¹H NMR (CDCl₃, 500 MHz) δ : 7.15 (d, J = 8.7 Hz, 2H), 6.86 (d, J = 8.6 Hz, 2H), 3.82 (s, 3H), 2.62 (q, J = 7.6 Hz, 2H), 1.24 (t, J = 7.6 Hz, 3H).

¹³C NMR (CDCl₃, **126** MHz) δ: 157.6, 136.4, 128.7, 113.7, 55.3, 28.0, 15.9.



Phenol (4a) (CAS: 108-95-2) (Sun et al., 2019)
Yield: 46% (4.3 mg) using 2-phenoxy-1-phenylethanol as substrate;
Yield: 47% (4.4 mg) using 1-(4-methoxyphenyl)-2-phenoxyethanol as substrate.
¹H NMR (CDCl₃, 500 MHz) δ: 7.31 – 7.25 (m, 2H), 6.97 (t, J = 7.4 Hz, 1H), 6.87 (d, J = 7.6 Hz, 2H), 4.81 (s, 1H).

¹³C NMR (CDCl₃, 126 MHz) δ: 155.4, 129.7, 120.8, 115.3.



4-Methoxyphenol (4b) (CAS: 150-76-5) (Sun et al., 2019) **Yield: 44% (5.5 mg).** ¹H NMR (CDCl₃, 500 MHz) δ: 6.84 – 6.77 (m, 4H), 4.57 (s, 1H), 3.79 (s, 3H). ¹³C NMR (CDCl₃, 126 MHz) δ: 153.8, 149.4, 116.0, 114.8, 55.8.

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