

# Peripheral pain mechanisms in osteoarthritis

Tonia L. Vincent

PAIN

#### Abstract

There is a well-established historical observation that structural joint damage by plain X-ray correlates poorly with symptomatic disease in osteoarthritis (OA). This is often attributed to the inability to visualise soft-tissue pathology within the joint and the recognition of heterogeneous patient factors that drive central pain sensitisation. A major issue is the relative paucity of mechanistic studies in which molecular pathogenesis of pain is interrogated in relation to tissue pathology. Nonetheless, in recent years, three broad approaches have been deployed to attempt to address this: correlative clinical studies of peripheral and central pain outcomes using magnetic resonance imaging, where soft-tissue processes can be visualised; molecular studies on tissue from patients with OA; and careful molecular targets such as nerve growth factor. Not only is the regulation of nerve growth factor within the joint being used to explore the relationship between tissue pathology and the origins of pain in OA, but it also provides a core model on which other molecules present within the joint can modulate the pain response. In this narrative review, how molecular and pathological tissue change relates to joint pain in OA will be discussed. Finally, a model for how tissue damage may lead to pain over the disease course will be proposed.

### 1. Introduction

The study of structural and symptomatic disease in osteoarthritis (OA) has been thwarted by having blunt tools on both sides.<sup>6</sup> Plain radiographs (X-rays) are only able to visualise 2-dimensional radiopaque structures, so are good for assessing gross changes in bone shape, osteophyte size, chondrocalcinosis, subchondral bone sclerosis, and bone cysts. From the space between the bone ends, one is able to extrapolate cartilage thickness, but the meniscus, synovium, and inflammatory processes are invisible. For patient pain outcomes, many tools have been developed and validated over the years to assess patient- and physicianreported measures of pain, but these are limited by their subjectivity and necessarily complicated when trying to capture the quality and pattern of different types of pain at different times. The contribution of central and peripheral components of pain also varies between and within individuals and there are few routinely applied methods for their assessment. Taken together, it is hardly surprising that our ability to relate clinical pain outcomes with structural change has been challenging.

Sponsorships or competing interests that may be relevant to content are disclosed at the end of this article.

NDORMS, Centre for OA Pathogenesis Versus Arthritis, Kennedy Institute for Rheumatology, University of Oxford, Oxford, United Kingdom

Corresponding author. Address: Centre for OA Pathogenesis Versus Arthritis, Kennedy Institute for Rheumatology, Roosevelt Drive, Oxford OX3 7FY, United Kingdom. Tel.: 0044 1865 612645. E-mail address: Tonia.vincent@kennedy.ox.ac.uk (T.L Vincent).

PAIN 161 (2020) S138-S146

http://dx.doi.org/10.1097/j.pain.0000000000001923

#### 2. The course of pain in human osteoarthritis

The course of pain in human OA is variable. Some individuals seem to present with painful joints with little in the way of change on a plain X-ray. This is often described as early OA especially if the individual has other risk factors eg, age >50 years, obesity. Evidence that these individuals necessarily go on to develop OA is modest. The classification criteria for defining early OA are actively being explored by Osteoarthritis Research Society International (OARSI) and other relevant OA groups.<sup>62</sup> Others can present seemingly for the first time after considerable damage has occurred,<sup>21</sup> although on the whole, most individuals presenting after the age of 50 years and with joint pain will have damage evident on an X-ray.<sup>56</sup> A few characteristics seem to be consistent: pain is usually mechanically induced in the early stages,<sup>64</sup> worsening with exercise and better at rest; the pain is typically burning in nature but can be associated with sharp shooting pains especially, eg, in the base of the thumb; pain often waxes and wanes but as the disease progresses, it becomes more frequent and can occur at rest or at night (reviewed in Ref. 38). The structural course thereafter for an individual is unpredictable with only 40% of individuals progressing on a plain X-ray over 10 years.<sup>59</sup> Almost all patients with a chronic peripheral drive of their pain will exhibit evidence of central sensitisation, but this may be particularly marked in some individuals where a more diffuse pain syndrome develops (reviewed in Refs. 2, 75). Other than qualitative pain questionnaires, quantitative sensory testing is the most commonly used clinical assessment tool for central sensitisation. Local and widespread sensory threshold changes after experimental stimulation eg, hot/cold, light touch, pressure, are common and may relate to patient outcome, for instance after joint replacement.<sup>80</sup> Other sensitisation features include evidence of temporal summation and increased spinal reflexes.<sup>60</sup> Patientrelated factors such as stress, adverse life experiences, sex, gut microbiome, and comorbidities predispose to the development of central sensitisation and contribute to the pain phenotype.<sup>7,24,93</sup>

Copyright © 2020 The Author(s). Published by Wolters Kluwer Health, Inc. on behalf of the International Association for the Study of Pain. This is an open access article distributed under the Creative Commons Attribution License 4.0 (CCBY), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

# 3. The course of pain-like behaviour in rodent osteoarthritis

In rodents, pain cannot be measured directly but is inferred from "pain-like behaviour." This can be through evoked behavioural responses (von Frey hair induced mechanical allodynia, hot/cold hypersensitivity, for instance) or by studying spontaneous behavioural responses (activity levels or by distribution of weight through the diseased compared with nondiseased joint). A number of different induced and spontaneous models in small and large animals have been used to study pain behaviour in OA (comprehensively reviewed in Ref. 69). In the past 10 years, these have largely been in rodents and broadly divide into those induced by monosodium iodoacetate (MIA) and those that are induced by surgical destabilisation of the joint (destabilisation of the medial meniscus [DMM], partial or complete meniscectomy, and anterior cruciate ligament transection, alone or in combination with ligamentous injury). A small minority of studies have used spontaneous rodent models of OA: in genetically susceptible strains, by ageing animals and by dietary modification, eg, highfat diet. Monosodium iodoacetate has been favoured historically by pain biologists because it produces rapid, robust changes in pain-like behaviour, in line with the rapidly destructive

inflammatory changes in the joint. Although it was recognised that features of MIA-induced disease may be relevant to some aspects of human OA, in 2012, the Arthritis Research UK animal models working group recognised that surgical models were generally becoming favoured for pathogenesis studies because these produce insidious, progressive, relatively noninflammatory changes in the cartilage, bone, and synovium, similar to the human condition<sup>99</sup> (Fig. 1A). Whether surgically induced joint destabilisation is representative of all types of OA is subject to intense debate. Despite attempts to categorise OA into different phenotypes (metabolic, post-traumatic, inflammatory, for instance), there is little evidence thus far to support a clinical benefit of doing so. In the case of synovitis, this is discussed further below. Conversely, there is a growing body of evidence showing that target discovery in surgical models of OA has clinical relevance in human nontraumatic disease; thus, these models seem to be generalisable.<sup>23,55,67,108</sup>

Focusing solely on surgical models, 2 distinct pain-like behaviours can be discerned: an initial postoperative phase that occurs in both OA and sham-operated animals and a late phase that, in our hands, starts at around 11 weeks after DMM, and 8 to 9 weeks after partial meniscectomy (PMX)<sup>26,39,100</sup> (**Fig. 1B**). Evoked and nonevoked measures, at the level of the joint or distal



Figure 1. Temporal relationship between pain-like behavior, pathology, and molecular expression in murine osteoarthritis. (A) Pathological changes in the joint over the course of surgically induced OA. Initial postoperative synovitis that is modest thereafter; progressive chondropathy; rapid early osteophyte formation that grows slowly over time. (B) Pain-like behavior after surgically induced OA (red) after either DMM or PMX or sham (gray) surgery. Postoperative behavioral change in joint destabilized groups is modest compared with sham. Some groups show postoperative mechanical allodynia that persists in the destabilized group compared with sham for 12 weeks (not shown). (C) Two key pain-sensitizing molecules with proven analgesic effects in vivo and where temporal data are available on their expression, nerve growth factor and CCL2. X represents actual time points tested. Taken from Refs. 12, 26, 67, 70, 72, 100. DMM, destabilisation of the medial mensions.

to the joint, seem to mirror one another when they are performed in the same study.<sup>26,48,90</sup> Differences in the model used to induce OA, the periodicity and length of time of follow-up (many looking only 2-4 weeks after surgery), and the behavioural tests deployed make it hard to reach a consensus on the pattern of pain-like behaviour after joint destabilisation; some groups reproducing our observed biphasic behavioural response,<sup>26,48,72,90,100</sup> and others detecting mechanical allodynia early after surgery, which fails to resolve over time.<sup>8,31,43,70,107</sup> Regardless, the ability to map pain-like behaviour in a temporal fashion is important because it provides an opportunity to relate this to pathological tissue changes occurring in the joint and associated dorsal root ganglia (L2-4). Studies are further enhanced by being able to perform molecular analyses on microdissected tissues of the joint, and to do such studies in genetically modified mice.<sup>12,70,72</sup>

# 4. Nerve growth factor as a pain target in human and murine osteoarthritis

In 2010, in a candidate molecule approach, we identified nerve growth factor (NGF) as a pain target in murine OA after DMM.<sup>67</sup> Ngf mRNA was upregulated in the whole joint during the 2 phases of pain behaviour (postoperative and late OA pain). The drivers of NGF in each case were shown to be different; postoperative pain being inhibited by neutralising tumour necrosis factor (TNF), and the late phase seemingly independent of inflammation. Soluble tropomyosin receptor kinase A, the receptor for NGF, was able to suppress both postoperative and late OA pain-like behaviour.<sup>67</sup> More recently, we also showed suppression of pain-like behaviour in mice following PMX after vaccination with murine Ngf.<sup>100</sup> Other groups have also demonstrated NGF-dependent pain-like behaviour in rats including by showing that intra-articular injection of Ngf exacerbates pain-like behavioural change,<sup>3</sup> and suppression of early pain-like behaviour with anti-Ngf<sup>53</sup> or by inhibition of Trka<sup>77</sup> after meniscal injury.

The first clinical Phase II trial of anti-NGF in patients with knee OA was published in 2010.<sup>55</sup> Since then, a number of studies using neutralising antibodies have demonstrated clinical efficacy validating NGF as a target in OA pain.<sup>86,91</sup> Concerns about the risk of rapidly progressive OA may yet limit its clinical utility (reviewed in Ref. 54). The FDA is yet to pronounce whether this class of drug, which was given "fast-track designation" in 2017, will be approved for use in patients in the United States. Two studies using TrkA inhibitors have also been published recently. One showed limited success after intra-articular delivery,<sup>51</sup> and a second oral agent study failed to reach its primary endpoint.<sup>104</sup>

The success of anti-NGF in the treatment of painful OA raises several questions: Where is NGF made? What are the drivers of NGF in the OA joint? And how does this change over the course of disease? Several developments in the past few years have helped to unravel this relationship in OA: (1) use of improved imaging modalities, particularly magnetic resonance imaging (MRI), to visualise the soft tissues of the joint, (2) agnostic molecular profiling of human OA tissues, and (3) careful dissection of molecular processes occurring over the course of rodent OA. In the next section, pathological changes in each of the main tissues of the joint (synovium, meniscus, bone, and cartilage) will be considered.

#### 5. The role of the synovium in osteoarthritis pain

The synovium has always been a good candidate tissue as a generator of pain responses in the OA joint; it is highly innervated, remodels in a dynamic fashion, and has a key role in driving both pain and joint damage in rheumatoid arthritis. Rheumatologists recognise joint space tenderness, joint effusion, and synovial thickening in individuals with OA, and most patients, at least anecdotally, gain benefit from intra-articular steroid. In recent years, visualisation by enhanced and nonenhanced MRI and ultrasound has enabled correlative studies with patient symptoms to be performed. Contrast-enhanced MRI is superior over nonenhanced imaging, and demonstrates that around 90% of individuals with symptomatic disease will show evidence of synovial enhancement indicative of hypertrophy and/or synovitis.<sup>85</sup> Most of these studies report a positive correlation, albeit often weak, between patient symptoms and severity of synovitis.<sup>36,109,112</sup> Synovitis has been shown to be predictive of disease progression in hand OA,<sup>35,49</sup> and a change in volume of the synovium is associated with a clinical improvement in patients given intra-articular steroid.<sup>78</sup> However, these studies have recognised limitations. They have frequently looked at synovitis separately from other pathologies in the joint, and where other pathologies have been examined, these usually co-correlate making it difficult to exclude whether synovitis is simply a biomarker of OA severity.<sup>110</sup> The incorrect conflation of correlation with causation and the assumption that synovial volume necessarily means "inflammation" further complicates interpretation. After the success of disease-modifying drugs in rheumatoid arthritis, it was assumed that the same antiinflammatory drugs would work in OA also. The recent lack of clinical success with a number of agents including anti-IL1, anti-TNF, and hydroxychloroguine indicate that this was not correct.<sup>14,15,45–47,57,96</sup> Other studies including those targeting IL6 or using methotrexate have been conducted but are vet to report.

Glucocorticoids, either delivered intra-articularly or orally, are assumed to target the synovium primarily, although it is worth remembering that several tissues of the joint express glucocorticoid receptors, including chondrocytes.<sup>92</sup> Randomised controlled trials (RCTs) using intra-articular or oral steroid show that the clinical response is mixed. Recent studies show that a longacting intra-articular steroid preparation, FX0006, did demonstrate statistically significant superiority over placebo, although the Phase II study failed to meet its primary endpoint (improve-ment over placebo at 12 weeks).<sup>16</sup> An RCT of intra-articular steroid in hand OA, in which lignocaine alone was used as a placebo, met 2 out of 11 coprimary endpoints.<sup>88</sup> A recent RCT of 10-mg daily oral prednisolone did show significant improvement over placebo for the duration of treatment, reverting back to pretreatment levels of pain within 3 weeks.<sup>50</sup> Few studies have examined the effect of repeated intra-articular steroid in patients with OA. In the study by McAlindon et al., 3 monthly triamcinolone intra-articular injections to individuals with knee OA over 2 years not only failed to register symptomatic improvement over placebo at any of the 3 monthly assessments, but also showed a small but significant increase in cartilage degradation in repeat steroid injected joints. This study may have missed a transient analgesic response to the steroid injection occurring within the 3-month follow-up period. Similar data have been observed in the Osteoarthritis Initiative, collectively highlighting the potential risks associated with this commonly delivered treatment. 65,111 Most of the above studies also had imaging outcomes that showed inconsistent responses when examining synovial volume and vascularity.

Molecular studies in OA synovium come from synovial biopsies performed in individuals with OA discordant for pain. In one study, individuals with Kellgren and Lawrence (K&L) grades of greater or

equal to 3 were stratified by high or low pain scores (by visual analogue scale). A microarray study was performed on synovial biopsies (from 5 individuals in each group), which revealed a number of dysregulated genes.<sup>10</sup> Of the regulated genes, candidate molecules of interest that have been implicated in OA pain included *TRKB*, the receptor for brain-derived growth factor, angiotensinogen, and netrin-1. Blockade of the angiotensin type Il receptor is analgesic in individuals with postherpetic neuralgia and in murine OA (see below), and netrin-1 has been shown to drive axonal growth in the subchondral bone of murine OA.<sup>33,83,113</sup> There were few classic inflammatory genes regulated apart from CCL14 and ADAMTS15. Notably, NGF was not identified as a regulated gene. Two other published studies have looked at synovial biopsies from symptomatic and asymptomatic groups that were matched for chondropathy score. It is worth noting that in this study, the asymptomatic group was taken from postmortem samples where there was no apparent history of OA pain from first-degree relatives or patient notes. Tissue morphology was examined and candidate molecules (NGF and CD68) were examined by immunohistochemistry. This study showed that there was increased synovial hypertrophy, cartilage damage, and both NGF and CD68 immunostaining in diseased tissue from those with higher pain.<sup>89</sup> A second study, by the same group, examined mRNA for 96 genes associated with pain in discovery and replication cohorts. Validated, regulated genes were further explored by protein assays. Few genes were consistent across groups, most likely reflecting the heterogeneity of patient samples, and there was a notable lack of inflammatory cytokines and NGF regulation even when compared with nonarthritis control tissue (presented in supplementary data).<sup>105</sup> Nonetheless, some molecules showed strong or consistent regulation at mRNA and protein level including angiotensin-converting enzyme (ACE) (increased), IL1R1 (decreased), CCL2, CCL8 (increased), and MMP1 (increased). Of these, ACE and CCL2 may be of particular interest. Angiotensin-converting enzyme is involved in the conversion of angiotensinogen to angiotensin II, which can activate the type II receptor, present on pain fibres, to reduce the threshold for firing.<sup>1,34</sup> Our studies show that ATII blockade is analgesic in murine OA (Vincent et al., manuscript in preparation). CCL2 has also been implicated in pain sensitisation in murine OA, causing increased macrophage infiltration in the dorsal root ganglia and delaying pain-like behaviour development.<sup>70,72</sup> The notion that synovitis may drive inflammatory priming of pain, contributing to pain severity is attractive (Fig. 1C). Of interest, synovitis correlates with sensitisation, measured by quantitative sensory testing in patients<sup>76</sup> and these predict response to nonsteroidal anti-inflammatory drugs.81

The role of an activated innate immune system in OA is strong from a variety of descriptive studies (some agnostic),<sup>17,71,73</sup> but there have been surprisingly few studies that have identified immune cells specifically driving this process and contributing to pain. One exception to this is the established role of mast cells in OA pain. Mast cells are increased in murine and human OA joints and contribute to structural degradation.<sup>11,22,103</sup> Mast cells also express TRKA, are responsive to NGF, and secrete NGF,<sup>37,58</sup> and hence have a potentially important role in structural and symptomatic OA.

# 6. The role of the meniscus, ligaments, and other soft tissues of the joint

The meniscus and ligaments (and extra-articular muscle) are critical for maintaining stability of the joint, and damage to any one of these increases the risk of OA.<sup>25</sup> This is seen upon acute ligamentous injuries such as anterior cruciate ligament rupture;

around 50% of such individuals develop OA within 10 years.<sup>61</sup> Extrusion of the meniscus, with or without evidence of previous meniscal tear, is also a common feature in age-related OA, suggesting that meniscus-dependent mechanical joint instability may contribute to OA risk in the elderly.<sup>27,29,63</sup> Significant pathology and molecular change has been documented in human meniscus in OA or after injury,<sup>82</sup> and also after joint destabilisation in rats.<sup>28</sup>

Much less is known about the role of the meniscus, ligaments, and other soft tissues in OA pain. Normal anatomical studies reveal that these tissues are highly innervated and this innervation can be deranged after development of disease.<sup>4,44,66</sup> In the mouse, 16 weeks after destabilisation of the meniscus, there is a marked increase in calcitonin gene related peptide-positive nerve fibres associating with the medial meniscus and synovium, in addition to the subchondral bone (discussed further below),<sup>79</sup> perhaps indicating that these tissues contribute to pain in OA. More studies are required in this area to be able to make more informed conclusions.

#### 7. The role of the bone in osteoarthritis pain

The bone is a very mechanoresponsive tissue and multiple changes occur during OA. New bone formation (osteophytosis) forms initially as a chondrophyte, and occurs at various sites in the joint, probably as a mechanoadaptive response.32,94 Osteophytes, often having a cartilaginous cap, change the overall shape of the articulating surface and have been shown to change in a consistent manner over time in age-related and posttraumatic human OA.9 In the mouse after joint destabilisation, the osteophyte has the appearance of extending the articulating surface (Fig. 2). Osteophytes in certain anatomical sites may be painful, for instance in the spine where they can restrict the exit of the nerve root.<sup>68</sup> Whether they are painful without inducing nerve stretch or compression is less clear despite the fact that they are highly innervated (unlike the articular cartilage). Osteophytosis forms one component of the K&L score. In a carefully controlled study in which only patients with unilateral OA were examined so that the noninvolved joint could act as an intrapatient control, painful episodes did correlate with K&L score, and this was shown to be upheld when just joint space narrowing was considered but was lost when the bone changes were examined on their own.74 Three extrapolations can be drawn from this study: first, X-ray bone changes (osteophytosis, subchondral bone sclerosis, and cysts) are not principal drivers of OA pain; second, person-specific factors, such as central sensitisation, contribute greatly to the patient pain experience; and third, joint space narrowing (cartilage loss) does show an association with OA pain once these patient factors are controlled for.

Sclerosis of the subchondral bone plate occurs immediately below the articular cartilage especially on the damaged or most loaded side of the joint and this is also where bone marrow (BM) oedema develops. Bone marrow oedema is not visible on the plain X-ray but is visualised by MRI. Similar to results in the synovium, BM oedema shows weak but significant correlation with pain severity<sup>20</sup> and there is also some evidence that prospective change in BM oedema correlates with a change in pain<sup>30,84</sup> and the development of bone cysts.<sup>13</sup>

A few molecular studies have tried to examine these pathological processes and link them to pain. Kuttapitiya et al.<sup>52</sup> performed an analysis in which they analysed regions of BM oedema (by MRI), taken from individuals undergoing joint replacement for severe symptomatic OA, and compared these with normal subchondral bone controls. A microarray analysis



Figure 2. Murine osteophyte 8 weeks after DMM showing medial compartment of the knee joint in coronal view. Note clear demarcation between old lateral boarder of epiphysis and new bone (osteophytes circled). Osteophytes have cartilaginous caps that seem to "extend" the articulating surface of the joint. Moderate cartilage damage is seen in the midportion of the weightbearing region of the joint particularly on the tibial surface (×40). DMM, destabilisation of the medial meniscus.

identified a number of molecules and pathways that were regulated specifically in the oedematous regions. By pathway analysis, these included molecules involved in neurogenesis, angiogenesis, and connective tissue remodelling. No evidence of *NGF* was found, but one of the most strongly regulated molecules was stathmin 2, known to be involved in regulating responsive-ness to NGF.<sup>42</sup>

In mice, osteophytes form extremely rapidly (within 2 weeks) upon joint destabilisation and continue to grow slowly over time.<sup>18</sup> The formation of the osteophyte does not seem to correlate with the onset of spontaneous pain-like behaviour (Fig. 1A). In one of our studies, we examined the differential expression of 67 molecules known to regulate pain. These included inflammatory cytokines, chemokines, neurotrophins, and their receptors. We microdissected the tissues from the mouse knee joint at the start of late OA pain behaviour and compared the expression levels with sham-operated controls.<sup>26</sup> Only 8 of the 67 molecules were regulated 8 weeks after partial meniscectomy compared with sham. When considering the whole joint RNA, these included Ngf, as we had seen previously after DMM, bradykinin receptors (Bdkr) B1 and B2, tachykinin (Tac1), tachykinin receptor (Tacr1), Tnf, Trpv4, and Vegf (the latter 3 being <1.5-fold). When microdissected tissues were analysed separately, NGF, Bdkrb1, Tac1, and Tacr1 were regulated in the articular cartilage. Bdkrb2 was regulated in the epiphysis (containing subchondral bone and osteophyte combined). Due to technical challenges, it was not possible at this time to exclude regulation of molecules in the synovium.<sup>26</sup> We validated the results by repeating the study at the time of pain-like behaviour development in a different surgical OA model, DMM. The same molecules were regulated in the joint at the time of pain with some minor differences. Importantly, in both studies, Ngf was exclusively upregulated in the articular cartilage.<sup>26</sup>

### 8. The role of cartilage in osteoarthritis pain

Of all the tissues considered by rheumatologists, connective tissue experts, and pain biologists over the years, the articular

cartilage has been at the bottom of the list as a candidate tissue for driving pain because it is aneural and insensate. Our results in the mouse were provocative and unexpected but seemed to suggest that damaged articular cartilage was capable of making pain-inducing molecules. This result is corroborated by the demonstration that NGF represents of 1 of 7 chondrocyte subsets recently identified by single-cell RNA sequencing of human OA articular cartilage<sup>40</sup> and was identified in early microarrays studies of severely damaged OA cartilage.<sup>87</sup> Simple mechanical injury of cartilage that activates inflammatory signalling within the chondrocyte by a process we have termed "mechanoflammation"<sup>97</sup> also strongly induced Ngf (300-fold within 8 hours of injury).<sup>26</sup> Injury was also able to upregulate Tac1, Bdkr1, and Bdkr2. Presented in a graphical abstract, we proposed a model by which cartilage damage, particularly towards the basal layer, causes upregulation and release of Ngf and other paininducing molecules, to sensitise local pain fibres and induce new sprouting and neoinnervation of the tissue.<sup>98</sup> This theory fits with the observed neoinnervation of the osteochondral junction documented by Walsh in late human OA, which relates to pain severity,<sup>5,101</sup> and supported by recent studies in murine OA in which new nerves develop in the subchondral bone late in disease, localised to the region below the most damaged cartilage.<sup>79</sup> Moreover, in a recent article by Zhu et al., pain driven by new nerve growth within the subchondral bone is also supported. Here, the authors demonstrate that axonal elongation is dependent on netrin-1, secreted by osteoclasts, which facilitates the passage of the nerve as it extends through the bone. Deletion of netrin-1 in osteoclasts or early treatment with a bisphosphonate was able to prevent pain-like behaviour in OA mice.<sup>113</sup> It is tempting to speculate that the osteoclast-dependent process is primarily concerned with the process of neoinnervation of the tissue but not beyond this period. Once the new nerve is established, osteoclast inhibition is less likely to modulate pain and this might explain why bisphosphonate trials in human OA seem to have at best a modest effect on symptoms and no effect on structural outcomes.<sup>95,106</sup> NGF is possibly the directional cue for these new nerves, as has been demonstrated in tumours,<sup>41</sup> and once formed it is also acts to sensitise them, thus explaining rapid clinical success in anti-NGF trials. New nerves require nutrients from new blood vessels and this would fit with the strong angiogenic signature found in BM lesions by microarray.52

# 9. Summary—the origin of peripheral osteoarthritis pain—a new model emerging

It has been fashionable in recent years to describe OA as a whole joint disease—this is hard to refute if the protagonists mean that most of the tissues in the joint are changed in disease. It has perhaps been less clear whether symptomatic disease, specifically pain considered here, is also driven by multiple tissue processes occurring in the joint at the same or different times and contributing to the pain experience. Nor has it been clear whether different joint pathologies could give rise to distinct pain phenotypes. The emerging literature would tend to suggest this, although there remain significant gaps in our knowledge. To date, the strongest evidence seems to lie within the subchondral bone where new blood vessels provide nutrient supply to axonal growth and neoinnervation of the osteochondral junction, likely driven by NGF released from basal articular chondrocytes (**Fig. 3**). Blockade



**Figure 3.** Proposed model for the molecular pathogenesis of pain in OA. Neoinnervation of the osteochondral junction is driven by NGF release from damaged cartilage and facilitated by osteoclast mediator netrin-1 within the oedematous bone marrow. Further sensitization within the joint is facilitated by CCL2, and likely other inflammatory mediators, eg, those released from mast cells. Angiotensinogen, processed by angiotensin-converting enzyme (ACE) to angiotensin (AT) II in the synovium, sensitises neurons through ATII type 2 receptor (AT2).

of angiogenesis or axonal extension suppresses pain in experimental models consistent with this proposed process.<sup>19,113</sup> The role of the synovium in driving pain is less clear. Despite strong correlative evidence from epidemiological studies and the experience in other synovitic diseases where targeting inflammation is efficacious, the synovium does not seem to be the principal pain driver in OA. The synovium is making some inflammatory molecules such as the chemokine CCL2, which leads to inflammatory response in the dorsal root ganglia of OA mice.<sup>70</sup> Whether the synovitic component somehow primes and thereby augments the strong NGF-driven process, or whether it acts independently is as yet unknown. Either way, measurements of such molecules could help to phenotype patients and provide a rationale for stratification in the management of OA pain.

#### **Conflict of interest statement**

In the past 3 years, T.V. has been on Advisory Boards for UCB, GSK, and Mundipharma. She directs the STEpUP OA Consortium, which receives financial project support (no personal gain) from Galapagos, Fidia, and Samumed. She has no other conflicts of interest to declare.

#### Acknowledgements

T.L. Vincent directs the Centre for Osteoarthritis Pathogenesis, Versus Arthritis, funded by Versus Arthritis, grant nos. 20205 and 21621. Work relating to pain studies mentioned in this review received additional support from a Kennedy Trust for Rheumatology Research DPhil studentship. Particular thanks to Miss Laura Collins for help generating the figures.

### Article history:

Received 3 March 2020 Received in revised form 6 May 2020 Accepted 7 May 2020

#### References

- Anand U, Yiangou Y, Sinisi M, Fox M, MacQuillan A, Quick T, Korchev YE, Bountra C, McCarthy T, Anand P. Mechanisms underlying clinical efficacy of Angiotensin II type 2 receptor (AT2R) antagonist EMA401 in neuropathic pain: clinical tissue and in vitro studies. Mol pain 2015;11: 38.
- [2] Arendt-Nielsen L. Pain sensitisation in osteoarthritis. Clin Exp Rheumatol 2017;35(suppl 107):68–74.
- [3] Ashraf S, Mapp PI, Burston J, Bennett AJ, Chapman V, Walsh DA. Augmented pain behavioural responses to intra-articular injection of nerve growth factor in two animal models of osteoarthritis. Ann Rheum Dis 2014;73:1710–8.
- [4] Ashraf S, Wibberley H, Mapp PI, Hill R, Wilson D, Walsh DA. Increased vascular penetration and nerve growth in the meniscus: a potential source of pain in osteoarthritis. Ann Rheum Dis 2011;70:523–9.
- [5] Aso K, Shahtaheri SM, Hill R, Wilson D, McWilliams DF, Walsh DA. Associations of symptomatic knee osteoarthritis with histopathologic features in subchondral bone. Arthritis Rheuma 2019;71:916–24.
- [6] Bedson J, Croft PR. The discordance between clinical and radiographic knee osteoarthritis: a systematic search and summary of the literature. BMC Musculoskel Disord 2008;9:116.
- [7] Boer CG, Radjabzadeh D, Medina-Gomez C, Garmaeva S, Schiphof D, Arp P, Koet T, Kurilshikov A, Fu J, Ikram MA, Bierma-Zeinstra S, Uitterlinden AG, Kraaij R, Zhernakova A, van Meurs JBJ. Intestinal microbiome composition and its relation to joint pain and inflammation. Nat Commun 2019;10:4881.
- [8] Bove SE, Laemont KD, Brooker RM, Osborn MN, Sanchez BM, Guzman RE, Hook KE, Juneau PL, Connor JR, Kilgore KS. Surgically induced osteoarthritis in the rat results in the development of both osteoarthritislike joint pain and secondary hyperalgesia. Osteoarthritis Cartilage 2006; 14:1041–8.
- [9] Bowes MA, Lohmander LS, Wolstenholme CBH, Vincent GR, Conaghan PG, Frobell RB. Marked and rapid change of bone shape in acutely ACL injured knees—an exploratory analysis of the Kanon trial. Osteoarthritis Cartilage 2019;27:638–45.
- [10] Bratus-Neuenschwander A, Castro-Giner F, Frank-Bertoncelj M, Aluri S, Fucentese SF, Schlapbach R, Sprott H. Pain-associated transcriptome changes in synovium of knee osteoarthritis patients. Genes (Basel) 2018;9:338.
- [11] Buckley MG, Gallagher PJ, Walls AF. Mast cell subpopulations in the synovial tissue of patients with osteoarthritis: selective increase in numbers of tryptase-positive, chymase-negative mast cells. J Pathol 1998;186:67–74.
- [12] Burleigh A, Chanalaris A, Gardiner MD, Driscoll C, Boruc O, Saklatvala J, Vincent TL. Joint immobilization prevents murine osteoarthritis and reveals the highly mechanosensitive nature of protease expression in vivo. Arthritis Rheum 2012;64:2278–88.
- [13] Carrino JA, Blum J, Parellada JA, Schweitzer ME, Morrison WB. MRI of bone marrow edema-like signal in the pathogenesis of subchondral cysts. Osteoarthritis Cartilage 2006;14:1081–5.
- [14] Chevalier X, Goupille P, Beaulieu AD, Burch FX, Bensen WG, Conrozier T, Loeuille D, Kivitz AJ, Silver D, Appleton BE. Intraarticular injection of anakinra in osteoarthritis of the knee: a multicenter, randomized, double-blind, placebo-controlled study. Arthritis Rheum 2009;61: 344–52.
- [15] Chevalier X, Ravaud P, Maheu E, Baron G, Rialland A, Vergnaud P, Roux C, Maugars Y, Mulleman D, Lukas C, Wendling D, Lafforgue P, Loeuille D, Foltz V, Richette P; Fso osteoarthritis. Adalimumab in patients with hand osteoarthritis refractory to analgesics and NSAIDs: a randomised, multicentre, double-blind, placebo-controlled trial. Ann Rheum Dis 2015;74:1697–705.
- [16] Conaghan PG, Cohen SB, Berenbaum F, Lufkin F, Johnson F, Bodick N. Brief report: a phase IIb trial of a novel extended-release microsphere formulation of triamcinolone acetonide for intraarticular injection in knee osteoarthritis. Arthritis Rheumatol 2018;70:204–11.
- [17] Daghestani HN, Pieper CF, Kraus VB. Soluble macrophage biomarkers indicate inflammatory phenotypes in patients with knee osteoarthritis. Arthritis Rheumatol 2015;67:956–65.

- [18] Das Neves Borges P, Vincent TL, Marenzana M. Automated assessment of bone changes in cross-sectional micro-CT studies of murine experimental osteoarthritis. PLoS One 2017;12: e0174294.
- [19] Das V, Kc R, Li X, O-Sullivan I, van Wijnen AJ, Kroin JS, Pytowski B, Applegate DT, Votta-Velis G, Ripper RL, Park TJ, Im HJ. Blockade of vascular endothelial growth factor receptor-1 (Flt-1), reveals a novel analgesic for osteoarthritis-induced joint pain. Gene Rep 2018;11: 94–100.
- [20] Daugaard CL, Riis RG, Bandak E, Gudbergsen H, Henriksen M, Bliddal H, Boesen M. Perfusion in bone marrow lesions assessed on DCE-MRI and its association with pain in knee osteoarthritis: a cross-sectional study. Skeletal Radiol 2019;393:1745–8.
- [21] Davis MA, Ettinger WH, Neuhaus JM, Barclay JD, Segal MR. Correlates of knee pain among US adults with and without radiographic knee osteoarthritis. J Rheumatol 1992;19:1943–9.
- [22] de Lange-Brokaar BJ, Kloppenburg M, Andersen SN, Dorjee AL, Yusuf E, Herb-van Toorn L, Kroon HM, Zuurmond AM, Stojanovic-Susulic V, Bloem JL, Nelissen RG, Toes RE, Ioan-Facsinay A. Characterization of synovial mast cells in knee osteoarthritis: association with clinical parameters. Osteoarthritis Cartilage 2016;24:664–71.
- [23] Deshmukh V, Hu H, Barroga C, Bossard C, Kc S, Dellamary L, Stewart J, Chiu K, Ibanez M, Pedraza M, Seo T, Do L, Cho S, Cahiwat J, Tam B, Tambiah JRS, Hood J, Lane NE, Yazici Y. A small-molecule inhibitor of the Wnt pathway (SM04690) as a potential disease modifying agent for the treatment of osteoarthritis of the knee. Osteoarthritis Cartilage 2018; 26:18–27.
- [24] Dieppe PA, Lohmander LS. Pathogenesis and management of pain in osteoarthritis. Lancet 2005;365:965–73.
- [25] Donahue TL, Fisher MB, Maher SA. Meniscus mechanics and mechanobiology. J Biomech 2015;48:1341–2.
- [26] Driscoll C, Chanalaris A, Knights C, Ismail H, Sacitharan PK, Gentry C, Bevan S, Vincent TL. Nociceptive sensitizers are regulated in damaged joint tissues, including articular cartilage, when osteoarthritic mice display pain behavior. Arthritis Rheumatol 2016;68:857–67.
- [27] Emmanuel K, Quinn E, Niu J, Guermazi A, Roemer F, Wirth W, Eckstein F, Felson D. Quantitative measures of meniscus extrusion predict incident radiographic knee osteoarthritis—data from the Osteoarthritis Initiative. Osteoarthritis Cartilage 2016;24:262–9.
- [28] Endo J, Sasho T, Akagi R, Muramatsu Y, Watanabe A, Akatsu Y, Fukawa T, Tahara M, Yamaguchi S. Comparative analysis of gene expression between cartilage and menisci in early-phase osteoarthritis of the knee-an animal model study. J Knee Surg 2018;31:664–9.
- [29] Englund M, Haugen IK, Guermazi A, Roemer FW, Niu J, Neogi T, Aliabadi P, Felson DT. Evidence that meniscus damage may be a component of osteoarthritis: the Framingham study. Osteoarthritis Cartilage 2016;24:270–3.
- [30] Felson DT, Niu J, Guermazi A, Roemer F, Aliabadi P, Clancy M, Torner J, Lewis CE, Nevitt MC. Correlation of the development of knee pain with enlarging bone marrow lesions on magnetic resonance imaging. Arthritis Rheum 2007;56:2986–92.
- [31] Fernihough J, Gentry C, Malcangio M, Fox A, Rediske J, Pellas T, Kidd B, Bevan S, Winter J. Pain related behaviour in two models of osteoarthritis in the rat knee. PAIN 2004;112:83–93.
- [32] Gelse K, Söder S, Eger W, Diemtar T, Aigner T. Osteophyte development—molecular characterization of differentiation stages. YJOCA 2003;11:141–8.
- [33] Gowler PRW, Li L, Woodhams SG, Bennett AJ, Suzuki R, Walsh DA, Chapman V. Peripheral brain-derived neurotrophic factor contributes to chronic osteoarthritis joint pain. PAIN 2020;161:61–73.
- [34] Hashikawa-Hobara N, Hashikawa N. Angiotensin II AT2 receptors regulate NGF-mediated neurite outgrowth via the NO-cGMP pathway. Biochem Biophys Res Commun 2016;478:970–5.
- [35] Haugen IK, Slatkowsky-Christensen B, Bøyesen P, Sesseng S, van der Heijde D, Kvien TK. MRI findings predict radiographic progression and development of erosions in hand osteoarthritis. Ann Rheum Dis 2016; 75:117–23.
- [36] Hill CL, Hunter DJ, Niu J, Clancy M, Guermazi A, Genant H, Gale D, Grainger A, Conaghan P, Felson DT. Synovitis detected on magnetic resonance imaging and its relation to pain and cartilage loss in knee osteoarthritis. Ann Rheum Dis 2007;66:1599–603.
- [37] Horigome K, Pryor JC, Bullock ED, Johnson EM Jr. Mediator release from mast cells by nerve growth factor. Neurotrophin specificity and receptor mediation. J Biol Chem 1993;268:14881–7.
- [38] Hunter DJ, McDougall JJ, Keefe FJ. The symptoms of osteoarthritis and the genesis of pain. Rheum Dis Clin North Am 2008;34:623–43.
- [39] Inglis JJ, McNamee KE, Chia SL, Essex D, Feldmann M, Williams RO, Hunt SP, Vincent T. Regulation of pain sensitivity in experimental

osteoarthritis by the endogenous peripheral opioid system. Arthritis Rheum 2008;58:3110–19.

- [40] Ji Q, Zheng Y, Zhang G, Hu Y, Fan X, Hou Y, Wen L, Li L, Xu Y, Wang Y, Tang F. Single-cell RNA-seq analysis reveals the progression of human osteoarthritis. Ann Rheum Dis 2019:78:100–10.
- [41] Jimenez-Andrade JM, Ghilardi JR, Castaneda-Corral G, Kuskowski MA, Mantyh PW. Preventive or late administration of anti-NGF therapy attenuates tumor-induced nerve sprouting, neuroma formation, and cancer pain. PAIN 2011;152:2564–74.
- [42] Jin K, Mao XO, Cottrell B, Schilling B, Xie L, Row RH, Sun Y, Peel A, Childs J, Gendeh G, Gibson BW, Greenberg DA. Proteomic and immunochemical characterization of a role for stathmin in adult neurogenesis. FASEB J 2004;18:287–99.
- [43] Kc R, Li X, Kroin JS, Liu Z, Chen D, Xiao G, Levine B, Li J, Hamilton JL, van Wijnen AJ, Piel M, Shelly DA, Brass D, Kolb E, Im HJ. PKCδ null mutations in a mouse model of osteoarthritis alter osteoarthritic pain independently of joint pathology by augmenting NGF/TrkA-induced axonal outgrowth. Ann Rheum Dis 2016;75:2133–41.
- [44] Kellgren JH, Samuel EP. The sensitivity and innervation of the articular capsule. J Bone Joint Surg 1950;32:84–92.
- [45] Kingsbury SR, Tharmanathan P, Keding A, Ronaldson SJ, Grainger A, Wakefield RJ, Arundel C, Birrell F, Doherty M, Vincent T, Watt FE, Dziedzic K, O'Neill TW, Arden NK, Scott DL, Dickson J, Garrood T, Green M, Menon A, Sheeran T, Torgerson D, Conaghan PG. Hydroxychloroquine effectiveness in reducing symptoms of hand osteoarthritis: a randomized trial. Ann Intern Med 2018;168:385–95.
- [46] Kloppenburg M, Peterfy C, Haugen IK, Kroon F, Chen S, Wang L, Liu W, Levy G, Fleischmann RM, Berenbaum F, van der Heijde D, Bansal P, Wittoek R, Feng S, Fang Y, Saltarelli M, Medema JK, Levesque MC. Phase Ila, placebo-controlled, randomised study of lutikizumab, an antiinterleukin-1α and anti-interleukin-1β dual variable domain immunoglobulin, in patients with erosive hand osteoarthritis. Ann Rheum Dis 2019;78:413–20.
- [47] Kloppenburg M, Ramonda R, Bobacz K, Kwok WY, Elewaut D, Huizinga TWJ, Kroon FPB, Punzi L, Smolen JS, Vander Cruyssen B, Wolterbeek R, Verbruggen G, Wittoek R. Etanercept in patients with inflammatory hand osteoarthritis (EHOA): a multicentre, randomised, double-blind, placebo-controlled trial. Ann Rheum Dis 2018;77:1757–64.
- [48] Knights CB, Gentry C, Bevan S. Partial medial meniscectomy produces osteoarthritis pain-related behaviour in female C57BL/6 mice. PAIN 2012;153:281–92.
- [49] Kortekaas MC, Kwok WY, Reijnierse M, Kloppenburg M. Inflammatory ultrasound features show independent associations with progression of structural damage after over 2 years of follow-up in patients with hand osteoarthritis. Ann Rheum Dis 2015;74:1720–4.
- [50] Kroon FPB, Kortekaas MC, Boonen A, Bohringer S, Reijnierse M, Rosendaal FR, Riyazi N, Starmans M, Turkstra F, van Zeben J, Allaart CF, Kloppenburg M. Results of a 6-week treatment with 10 mg prednisolone in patients with hand osteoarthritis (HOPE): a double-blind, randomised, placebo-controlled trial. Lancet 2019;394:1993–2001.
- [51] Krupka E, Jiang GL, Jan C. Efficacy and safety of intra-articular injection of tropomyosin receptor kinase A inhibitor in painful knee osteoarthritis: a randomized, double-blind and placebo-controlled study. Osteoarthritis Cartilage 2019;27:1599–607.
- [52] Kuttapitiya A, Assi L, Laing K, Hing C, Mitchell P, Whitley G, Harrison A, Howe FA, Ejindu V, Heron C, Sofat N. Microarray analysis of bone marrow lesions in osteoarthritis demonstrates upregulation of genes implicated in osteochondral turnover, neurogenesis and inflammation. Ann Rheum Dis 2017;76:1764–73.
- [53] LaBranche TP, Bendele AM, Omura BC, Gropp KE, Hurst SI, Bagi CM, Cummings TR, Grantham LE, Shelton DL, Zorbas MA. Nerve growth factor inhibition with tanezumab influences weight-bearing and subsequent cartilage damage in the rat medial meniscal tear model. Ann Rheum Dis 2017;76:295–302.
- [54] Lane NE, Corr M. Osteoarthritis in 2016: anti-NGF treatments for pain—two steps forward, one step back? Nat Rev Rheumatol 2017;13: 76–8.
- [55] Lane NE, Schnitzer TJ, Birbara CA, Mokhtarani M, Shelton DL, Smith MD, Brown MT. Tanezumab for the treatment of pain from osteoarthritis of the knee. N Engl J Med 2010;363:1521–31.
- [56] Lawrence JS, Bremner JM, Bier F. Osteo-arthrosis. Prevalence in the population and relationship between symptoms and x-ray changes. Ann Rheum Dis 1966;25:1–24.
- [57] Lee W, Ruijgrok L, Boxma-de Klerk B, Kok MR, Kloppenburg M, Gerards A, Huisman M, Hazes M, de Sonnaville P, Grillet B, Weel A, Basoski N. Efficacy of hydroxychloroquine in hand osteoarthritis: a randomized, double-blind, placebo-controlled trial. Arthritis Care Res (Hoboken) 2018;70:1320–5.

- [58] Leon A, Buriani A, Dal Toso R, Fabris M, Romanello S, Aloe L, Levi-Montalcini R. Mast cells synthesize, store, and release nerve growth factor. Proc Natl Acad Sci U S A 1994;91:3739–43.
- [59] Leyland KM, Hart D, Javaid MK, Judge A, Kiran A, Soni A, Goulston LM, Cooper C, Spector TD, Arden NK. The natural history of radiographic knee osteoarthritis: a fourteen year population-based cohort study. Arthritis Rheum 2012;64:2243–51.
- [60] Lluch E, Torres R, Nijs J, Van Oosterwijck J. Evidence for central sensitization in patients with osteoarthritis pain: a systematic literature review. Eur J Pain 2014;18:1367–75.
- [61] Lohmander LS, Englund PM, Dahl LL, Roos EM. The long-term consequence of anterior cruciate ligament and meniscus injuries: osteoarthritis. Am J Sports Med 2007;35:1756–69.
- [62] Luyten FP, Bierma-Zeinstra S, Dell'Accio F, Kraus VB, Nakata K, Sekiya I, Arden NK, Lohmander LS. Toward classification criteria for early osteoarthritis of the knee. Semin Arthritis Rheum 2018;47:457–63.
- [63] Madan-Sharma R, Kloppenburg M, Kornaat PR, Botha-Scheepers SA, Le Graverand MP, Bloem JL, Watt I. Do MRI features at baseline predict radiographic joint space narrowing in the medial compartment of the osteoarthritic knee 2 years later? Skeletal Radiol 2008;37:805–11.
- [64] Marriott KA, Birmingham TB, Leitch KM, Pinto R, Giffin JR. Strong independent associations between gait biomechanics and pain in patients with knee osteoarthritis. J Biomech 2019;94:123–9.
- [65] McAlindon TE, LaValley MP, Harvey WF, Price LL, Driban JB, Zhang M, Ward RJ. Effect of intra-articular triamcinolone vs saline on knee cartilage volume and pain in patients with knee osteoarthritis: a randomized clinical trial. JAMA 2017;317:1967–75.
- [66] McDougall JJ, Bray RC, Sharkey KA. Morphological and immunohistochemical examination of nerves in normal and injured collateral ligaments of rat, rabbit, and human knee joints. Anat Rec 1997; 248:29–39.
- [67] McNamee KE, Burleigh A, Gompels LL, Feldmann M, Allen SJ, Williams RO, Dawbarn D, Vincent TL, Inglis JJ. Treatment of murine osteoarthritis with TrkAd5 reveals a pivotal role for nerve growth factor in noninflammatory joint pain. PAIN 2010;149:386–92.
- [68] Menkes CJ, Lane NE. Are osteophytes good or bad? Osteoarthritis Cartilage 2004;12(suppl A):S53–54.
- [69] Miller RE, Malfait AM. Osteoarthritis pain: what are we learning from animal models? Best Pract Res Clin Rheumatol 2017;31:676–87.
- [70] Miller RE, Tran PB, Das R, Ghoreishi-Haack N, Ren D, Miller RJ, Malfait AM. CCR2 chemokine receptor signaling mediates pain in experimental osteoarthritis. Proc Natl Acad Sci USA 2012;109:20602–7.
- [71] Miller RE, Tran PB, Ishihara S, Syx D, Ren D, Miller RJ, Valdes AM, Malfait AM. Microarray analyses of the dorsal root ganglia support a role for innate neuro-immune pathways in persistent pain in experimental osteoarthritis. Osteoarthritis Cartilage 2020;28:581–92.
- [72] Miotla Zarebska J, Chanalaris A, Driscoll C, Burleigh A, Miller RE, Malfait AM, Stott B, Vincent TL. CCL2 and CCR2 regulate pain-related behaviour and early gene expression in post-traumatic murine osteoarthritis but contribute little to chondropathy. Osteoarthritis Cartilage 2017;25:406–12.
- [73] Nees TA, Rosshirt N, Zhang JA, Reiner T, Sorbi R, Tripel E, Walker T, Schiltenwolf M, Hagmann S, Moradi B. Synovial cytokines significantly correlate with osteoarthritis-related knee pain and disability: inflammatory mediators of potential clinical relevance. J Clin Med 2019;8:1343.
- [74] Neogi T, Felson D, Niu J, Nevitt M, Lewis CE, Aliabadi P, Sack B, Torner J, Bradley L, Zhang Y. Association between radiographic features of knee osteoarthritis and pain: results from two cohort studies. BMJ 2009; 339:b2844.
- [75] Neogi T, Frey-Law L, Scholz J, Niu J, Arendt-Nielsen L, Woolf C, Nevitt M, Bradley L, Felson DT; Multicenter Osteoarthritis S. Sensitivity and sensitisation in relation to pain severity in knee osteoarthritis: trait or state? Ann Rheum Dis 2015;74:682–8.
- [76] Neogi T, Guermazi A, Roemer F, Nevitt MC, Scholz J, Arendt-Nielsen L, Woolf C, Niu J, Bradley LA, Quinn E, Law LF. Association of joint inflammation with pain sensitization in knee osteoarthritis: the multicenter osteoarthritis study. Arthritis Rheumatol 2016;68:654–61.
- [77] Nwosu LN, Mapp PI, Chapman V, Walsh DA. Blocking the tropomyosin receptor kinase A (TrkA) receptor inhibits pain behaviour in two rat models of osteoarthritis. Ann Rheum Dis 2016;75:1246–54.
- [78] O'Neill TW, Parkes MJ, Maricar N, Marjanovic EJ, Hodgson R, Gait AD, Cootes TF, Hutchinson CE, Felson DT. Synovial tissue volume: a treatment target in knee osteoarthritis (OA). Ann Rheum Dis 2016; 75:84–90.
- [79] Obeidat AM, Miller RE, Miller RJ, Malfait AM. The nociceptive innervation of the normal and osteoarthritic mouse knee. Osteoarthritis Cartilage 2019;27:1669–79.

- [80] Petersen KK, Graven-Nielsen T, Simonsen O, Laursen MB, Arendt-Nielsen L. Preoperative pain mechanisms assessed by cuff algometry are associated with chronic postoperative pain relief after total knee replacement. PAIN 2016;157:1400–6.
- [81] Petersen KK, Simonsen O, Olesen AE, Morch CD, Arendt-Nielsen L. Pain inhibitory mechanisms and response to weak analgesics in patients with knee osteoarthritis. Eur J Pain 2019;23:1904–12.
- [82] Rai MF, Patra D, Sandell LJ, Brophy RH. Transcriptome analysis of injured human meniscus reveals a distinct phenotype of meniscus degeneration with aging. Arthritis Rheum 2013;65:2090–101.
- [83] Rice ASC, Dworkin RH, McCarthy TD, Anand P, Bountra C, McCloud PI, Hill J, Cutter G, Kitson G, Desem N, Raff M. EMA401, an orally administered highly selective angiotensin II type 2 receptor antagonist, as a novel treatment for postherpetic neuralgia: a randomised, doubleblind, placebo-controlled phase 2 clinical trial. Lancet 2014;383: 1637–47.
- [84] Roemer FW, Guermazi A, Javaid MK, Lynch JA, Niu J, Zhang Y, Felson DT, Lewis CE, Torner J, Nevitt MC, investigators MS. Change in MRIdetected subchondral bone marrow lesions is associated with cartilage loss: the MOST Study. A longitudinal multicentre study of knee osteoarthritis. Ann Rheum Dis 2009;68:1461–5.
- [85] Roemer FW, Kassim Javaid M, Guermazi A, Thomas M, Kiran A, Keen R, King L, Arden NK. Anatomical distribution of synovitis in knee osteoarthritis and its association with joint effusion assessed on nonenhanced and contrast-enhanced MRI. Osteoarthritis Cartilage 2010; 18:1269–74.
- [86] Sanga P, Katz N, Polverejan E, Wang S, Kelly KM, Haeussler J, Thipphawong J. Long-Term safety and efficacy of fulranumab in patients with moderate-to-severe osteoarthritis pain: a phase II randomized, double-blind, placebo-controlled extension study. Arthritis Rheum 2017;69:763–73.
- [87] Sato T, Konomi K, Yamasaki S, Aratani S, Tsuchimochi K, Yokouchi M, Masuko-Hongo K, Yagishita N, Nakamura H, Komiya S, Beppu M, Aoki H, Nishioka K, Nakajima T. Comparative analysis of gene expression profiles in intact and damaged regions of human osteoarthritic cartilage. Arthritis Rheum 2006;54:808–17.
- [88] Spolidoro Paschoal Nde O, Natour J, Machado FS, de Oliveira HAV, Furtado RNV. Effectiveness of triamcinolone hexacetonide intraarticular injection in interphalangeal joints: a 12-week randomized controlled trial in patients with hand osteoarthritis. J Rheumatol 2015;42:1869–77.
- [89] Stoppiello LA, Mapp PI, Wilson D, Hill R, Scammell BE, Walsh DA. Structural associations of symptomatic knee osteoarthritis. Arthritis Rheumatol 2014;66:3018–27.
- [90] Temp J, Labuz D, Negrete R, Sunkara V, Machelska H. Pain and knee damage in male and female mice in the medial meniscal transectioninduced osteoarthritis. Osteoarthritis Cartilage 2020;28:475–85.
- [91] Tive L, Bello AE, Radin D, Schnitzer TJ, Nguyen H, Brown MT, West CR. Pooled analysis of tanezumab efficacy and safety with subgroup analyses of phase III clinical trials in patients with osteoarthritis pain of the knee or hip. J Pain Res 2019;12:975–95.
- [92] Tu J, Stoner S, Fromm PD, Wang T, Chen D, Tuckermann J, Cooper MS, Seibel MJ, Zhou H. Endogenous glucocorticoid signaling in chondrocytes attenuates joint inflammation and damage. FASEB J 2018;32:478–87.
- [93] Valdes AM. Metabolic syndrome and osteoarthritis pain: common molecular mechanisms and potential therapeutic implications. Osteoarthritis Cartilage 2020;28:7–9.
- [94] Van Der Kraan PM, van den Berg WB. Osteophytes: relevance and biology. YJOCA 2007;15:237–44.
- [95] Vaysbrot EE, Osani MC, Musetti MC, McAlindon TE, Bannuru RR. Are bisphosphonates efficacious in knee osteoarthritis? A meta-analysis of randomized controlled trials. Osteoarthritis Cartilage 2018;26: 154–64.
- [96] Verbruggen G, Wittoek R, Vander Cruyssen B, Elewaut D. Tumour necrosis factor blockade for the treatment of erosive osteoarthritis of the interphalangeal finger joints: a double blind, randomised trial on structure modification. Ann Rheum Dis 2012;71:891–8.
- [97] Vincent TL. Mechanoflammation in osteoarthritis pathogenesis elsevier enhanced reader. Semin Arthritis Rheum 2019;49:S36–38.
- [98] Vincent TL. Nociceptive sensitizers are regulated in damaged joint tissue, including articular cartilage, when osteoarthritic mice display pain behavior. Arthritis Rheum 2019;68:A18.
- [99] Vincent TL, Williams RO, Maciewicz R, Silman A, Garside P; Arthritis Research UKamwg. Mapping pathogenesis of arthritis through small animal models. Rheumatology (Oxford) 2012;51:1931–41.
- [100] von Loga IS, El-Turabi A, Jostins L, Miotla-Zarebska J, Mackay-Alderson J, Zeltins A, Parisi I, Bachmann MF, Vincent TL. Active immunisation targeting nerve growth factor attenuates chronic

pain behaviour in murine osteoarthritis. Ann Rheum Dis 2019;78: 672–5.

- [101] Walsh DA, McWilliams DF, Turley MJ, Dixon MR, Fransès RE, Mapp PI, Wilson D. Angiogenesis and nerve growth factor at the osteochondral junction in rheumatoid arthritis and osteoarthritis. Rheumatology (Oxford) 2010;49:1852–61.
- [102] Walsh DA, Neogi T. A tale of two TrkA inhibitor trials: same target, divergent results. Osteoarthritis Cartilage 2019;27:1575–7.
- [103] Wang Q, Lepus CM, Raghu H, Reber LL, Tsai MM, Wong HH, von Kaeppler E, Lingampalli N, Bloom MS, Hu N, Elliott EE, Oliviero F, Punzi L, Giori NJ, Goodman SB, Chu CR, Sokolove J, Fukuoka Y, Schwartz LB, Galli SJ, Robinson WH. IgE-mediated mast cell activation promotes inflammation and cartilage destruction in osteoarthritis. Elife 2019;8: 390.
- [104] Watt FE, Blauwet MB, Fakhoury A, Jacobs H, Smulders R, Lane NE. Tropomyosin-related kinase A (TrkA) inhibition for the treatment of painful knee osteoarthritis: results from a randomized controlled phase 2a trial. Osteoarthritis Cartilage 2019;27:1590–8.
- [105] Wyatt LA, Nwosu LN, Wilson D, Hill R, Spendlove I, Bennett AJ, Scammell BE, Walsh DA. Molecular expression patterns in the synovium and their association with advanced symptomatic knee osteoarthritis. Osteoarthritis Cartilage 2019;27:667–75.
- [106] Xing RL, Zhao LR, Wang PM. Bisphosphonates therapy for osteoarthritis: a meta-analysis of randomized controlled trials. Springerplus 2016;5:1704.
- [107] Yang PY, Tang CC, Chang YC, Huang SY, Hsieh SP, Fan SS, Lee HP, Lin SC, Chen WF, Wen ZH, Jean YH. Effects of tibolone on osteoarthritis in ovariectomized rats: association with nociceptive pain behaviour. Eur J Pain 2014;18:680–90.

- [108] Yazici Y, McAlindon TE, Fleischmann R, Gibofsky A, Lane NE, Kivitz AJ, Skrepnik N, Armas E, Swearingen CJ, DiFrancesco A, Tambiah JRS, Hood J, Hochberg MC. A novel Wnt pathway inhibitor, SM04690, for the treatment of moderate to severe osteoarthritis of the knee: results of a 24-week, randomized, controlled, phase 1 study. Osteoarthritis Cartilage 2017;25:1598–606.
- [109] Yusuf E, Kortekaas MC, Watt I, Huizinga TW, Kloppenburg M. Do knee abnormalities visualised on MRI explain knee pain in knee osteoarthritis? A systematic review. Ann Rheum Dis 2011;70:60–7.
- [110] Yusup A, Kaneko H, Liu L, Ning L, Sadatsuki R, Hada S, Kamagata K, Kinoshita M, Futami I, Shimura Y, Tsuchiya M, Saita Y, Takazawa Y, Ikeda H, Aoki S, Kaneko K, Ishijima M. Bone marrow lesions, subchondral bone cysts and subchondral bone attrition are associated with histological synovitis in patients with end-stage knee osteoarthritis: a cross-sectional study. Osteoarthritis Cartilage 2015;23: 1858–64.
- [111] Zeng C, Lane NE, Hunter D, Wei J, Choi H, McAlindon T, Li H, Lu N, Lei G, Zhang Y. Intra-articular corticosteroids and the risk of knee osteoarthritis progression: results from the Osteoarthritis Initiative. Osteoarthritis and cartilage 2019;27:855–62.
- [112] Zhang Y, Nevitt M, Niu J, Lewis C, Torner J, Guermazi A, Roemer F, McCulloch C, Felson DT. Fluctuation of knee pain and changes in bone marrow lesions, effusions, and synovitis on magnetic resonance imaging. Arthritis Rheum 2011;63:691–9.
- [113] Zhu S, Zhu J, Zhen G, Hu Y, An S, Li Y, Zheng Q, Chen Z, Yang Y, Wan M, Skolasky RL, Cao Y, Wu T, Gao B, Yang M, Gao M, Kuliwaba J, Ni S, Wang L, Wu C, Findlay D, Eltzschig HK, Ouyang HW, Crane J, Zhou FQ, Guan Y, Dong X, Cao X. Subchondral bone osteoclasts induce sensory innervation and osteoarthritis pain. J Clin Invest 2019;129:1076–93.