Jundishapur J Nat Pharm Prod.2012;7(4):168-175.

Jundishapur Journal of Natural Pharmaceutical Products www.iinpb.com

The Antioxidant Capacity and Anti-diabetic Effect of *Boswellia serrata* Triana and Planch Aqueous Extract in Fertile Female Diabetic Rats and the Possible Effects on Reproduction and Histological Changes in the Liver and Kidneys

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## ARTICLE INFO

Article type: Research Article

*Article history:* Received: 11 Jun 2012 Revised: 03 Jul 2012 Accepted: 22 Jul 2012

Keywords: Boswellia Hypoglycemic Agents Diabetes Complications Rats Antioxidants Histology

#### ABSTRACT

**Background:** Boswellia serrata has been used in a wide variety of diseases, including diae betes mellitus and inflammatory diseases.

**Objectives:** This study focused on the effects of *Boswellia serrata* aqueous extract on blood glucose and the complications of diabetes in the liver and kidneys and examined the impact of plant on reproduction in diabetic rats.

**Materials and Methods:** The antioxidant capacity of plant extract was performed using FRAP assay. Diabetic and control rats were administered 200, 400, and 600 mg/kg *Boswellia serrata* extract. Vaginal plaque was mentioned as a positive sign of pregnancy ;and treatment started with extract or vehicle from 1th to 17th day of gestation by gastric gavage. Blood glucose was measured during 17 days.

**Results:** The Administration of *Boswellia serrata* in diabetic rats significantly decreased the level of blood glucose and HbA1c after 17th days ( $P \le 0.01$ ). In diabetic group that received no treatment, the abortion of fetus spontaneous was 19.14%. The percentage of absorptions significantly was elevated in vehicle-treated diabetic rats; in comparison with vehicle- treated healthy rats. In the diabetic group, separated necrosis of hepatocytes, anarchism of liver plates, and lymphocytic inflammation were improved. Diabetic complications were not seen and the severity of damage was reduced. These damages include: lymphocytic inflammation in the port areas, irregularities, apoptosis of liver cells, and dilatation of the sinusoids.

**Conclusions:** The results suggest that *Boswellia serrata* extract has the antidiabetic effects and can prevent the complications of diabetes in the kidneys and liver.

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▶ Implication for health policy/practice/research/medical education:

*Boswellia* extracts used in appropriate doses, while having the anti-hyperglycemia effect on blood sugar can prevent the complications of diabetes in the liver and kidneys.

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#### Please cite this paper as:

Namjoyan F, Azemi ME, Khodayar MJ, Ahmadpour F, Darvish Padok A, Panahi M. The Antioxidant Capacity and Anti-diabetic Effect of Boswellia serrata Triana and Planch Aqueous Extract in Fertile Female Diabetic Rats and the Possible Effects on Reproduction and Histological Changes in the Liver and Kidneys. *Jundishapur J Nat Pharm Prod*. 2012:**7**(4);168-175.

# 1. Background

Diabetes is a metabolic disorder resulting from insulin resistance or occurring impaired insulin secretion, therefore a diabetic patient will suffer from chronic hyperglycemia (1). Chronic high blood glucose in diabetes leads in advanced glycation end-products which will subsequently produce reactive oxygen species. The free radicals originating from damaged cell membranes and lipid peroxidation, causes irreversible damage to the liver, kidneys, eyes, nervous system, cardiovascular and other parts of the body (2). The liver is the organ that is severely damaged in diabetes.3 These damages and disorders include inflammation, necrosis, fibrosis of non-alcoholic fatty liver disease, cirrhosis, hepatocellular carcinoma, hepatitis, and acute liver failure (3). Impaired glucose and protein metabolism in diabetes can also lead to dysfunction and structure damages in kidney called diabetic nephropathy which causes the glomerular basement membrane thickening and increases the kidney interstitial tissue (4).

Diabetes is considered as an important reason for increasing malformations ratio (5) of fetus to 10 times and the risk of fetus death to 5 times (6). Malformations as a result of maternal diabetes occur at high rate including: spinal bifida, structural brain anomalies, anencephaly, hydrocephalus, neuropathic bowel and bladder, sexual dysfunction, skeletal deformation, endocrine disorder and other central nervous system defects (7). Good control of diabetes can reduce complications during pregnancy. The use of medicinal plants to treat a wide variety of diseases, including inflammatory diseases, diabetes mellitus, and many disorders in the liver and the kidneys are growing (8).

Boswellia serrata Triana & Planch (B.S) is a moderate-tolarge branching tree, were found in India, North Africa and the Middle East. Strips of B.S bark are peeled away, yielding a gummy oleo-resin (9). B.S contains oils and β-boswellic acid, 3-O-acetyl-β-boswellic acid, 11-keto-βboswellic acid and 3-O-acetyl-11-keto-β-boswellic acid (10). B.S has a variety of pharmacological effects, particularly anti-hyperglycemia effect (11), antioxidant effects, may have beneficial effects in peptic ulcer (12) and also antiinflammatory effects in asthma (13), inflammatory bowel disease (14), cancer (15), and osteoarthritis (16). The hexane and alcoholic extracts of B.S protect liver against toxicity and damage induced by carbon tetrachloride, paracetamol or thioacetamide (17). Preparations that contain B.S oleo gum resin reduce blood sugar in STZ-induced diabetic rats (11). In another study, the root and leaf extracts of Boswellia glabra also reduced blood glucose levels in diabetic rats with alloxan (18). These effects of B.S

made this plant a good candidate for studies on diabetes and its complications in the liver and the kidney.

# 2. Objectives

This study focused on the effects of *B*.*S* extract on diabetes complications in the liver and kidney and examined its impact on reproduction in diabetic animals.

## 3. Materials and Methods

B.S dried resin were purchased from herbal shop in 2011 and it was confirmed by pharmacognosy department school of pharmacy. Streptozotocin (STZ) from Sigmaaldrich corporation of America and TPTZ, Fecl<sub>3</sub>.6H<sub>2</sub>O, FeSO<sub>4</sub>.7H<sub>2</sub>O, HCL, Acetic acid, CH<sub>3</sub>COONa.3H<sub>2</sub>O and methanol were purchased from Merck.

All other reagents were of analytical grade.

#### 3.1. Preparation of Plant Extract

Dried B.S oleo gum resin was added to hot distilled water on the magnetic heater for 1 hr, and then the extract was filtered. Solvent was removed using freeze dryer (operon). Resulting porous powder was stored in a refrigerator at 4-8°C till use.

#### 3.2. Ferric-Reducing Antioxidant Power (FRAP) Assay

The antioxidant capacity of plant extracts was performed according to the method of Namjooyan *et al.* (19). The FRAP reagent included 10 mM TPTZ solution in 40 mM HCl, 20 mM FeCl<sub>3</sub> solution and 0.3 M acetate buffer (pH 3.6) in proportions of 1:1:10(v/v). Then, 50  $\mu$ L of each diluted extracts were mixed with 3 mL of freshly prepared FRAP reagent and the reaction mixtures were incubated at 37 °C for 4 min.

Absorbance at 593 nm was determined against distilled water blank. Aqueous solutions of ferrous sulfate (100–2000  $\mu$ M) were used for calibration. Triplicate measurements were taken and the FRAP values were expressed as mmol of Fe (II)/g dry weight of plant powder (20).

#### 3.3. Animals

Adult female Sprague-Dawley rats between 70-90 days of age with body weight ranging between 200-240 gm were used for the study. Animals were taken from experimental animal house of Ahvaz university of Medical Sciences. Animals were maintained under standard condition (temperature  $20 \pm 2$ , humidity 60 to 65 percent and in the 12-hour dark, lighting conditions) (7). Animals were fed with standard pellet diet and had free access to

water. All the ethical issues were considered based on the Ahvaz Medical University Ethical Protocols (AMUEP) on animal experiments.

## 3.4. Induction of Experimental Diabetes

The animals were kept hungry for 8 hours before the injection of STZ. STZ was dissolved in sodium citrate buffer (0.09 M and pH 4.8). Diabetes induced by a single intraperitoneal administration of freshly dose of STZ (45 mg/ kg body weight). One week after STZ injection, animals with blood glucose level above 250 mg/dl were taken as diabetic for the study. The blood sample was drawn from the lateral tail vein of rats and measured by portable glucometer (bionime model).

## 3.5. The Establishment of Pregnancy in Female Rats

For mating, the female diabetic and healthy male rats were placed together in the cage. Successful mating in rats is verified by the presence of a vaginal plug (VP) by microscope. Female rats were caged with syngeneic male rats and checked daily for the presence of vaginal plaque. Plaque positive female rat were selected and examined for the presence of sperm in vaginal smears. Only when both criteria of vaginal plaque and sperm presence were met, rats were considered to be at zero day of pregnancy (6, 21).

### 3.6. Experimental Procedure

VP positive rats were randomized into five groups of five rats each. Rats received vehicle or extract orally by gavage from the first pregnancy day until the seventeenth day as follows:

- Group I: Healthy untreated rats
- Group II: Diabetic control rats

Group III, IV and V: Diabetic rats given the doses of 200, 400 and 600 mg/kg of *B.S* extract, respectively.

Blood glucose was measured three times: including 7th days after STZ injection, after single dose administration of extract at the first day and at 17th day of study. Blood

glucose measured half an hour after the administration of the extract or vehicle. In the 17th day of pregnancy, animals were scarified under anesthesia with ether, blood sample obtained from heart and collected in heparinated tubes to determine blood glucose level and HbA1C factor. The liver, kidney and uterus were quickly removed and washed with ice-cold isotonic saline. The liver and kidneys were placed in 10% formalin and the paraffin blocks of tissue prepared then cut in sections (3-5  $\mu$ m thick). In order to evaluate possible morphological changes of liver and kidney, tissues of the different groups stained with hematoxylin and eosin (H and E). In the uterus the number of cicatricial plaques showing sites of previous implantation fetus were counted and the crown-rump length was measured (22).

## 3.7. Statistical Analysis

All the results were expressed as Mean± S.E for five animals in each group. Data were analyzed using analysis of variance (ANOVA) and post hoc test of Tukey's. Differr ences below the 0.05 level were considered as statistically significant.

## 4. Results

Antioxidant effect of B.S oleo gum resin extract were evaluated and compared with the standard control group. The FRAP value for B.S aqueous extract was 0.99 mmol Fe<sup>2+</sup>/L. The mean blood glucose values in all groups are shown in *Figure 1*. As presented in *Figure 1*, diabetic control rats showed a significant increase in blood glucose and HbA1c compared to control group ( $P \leq 0.001$ )

Table 1. HbA1c level in different groups					
	HbA1c				
Group I	$3.58\pm.302$				
Group II	$9.75 \pm .3$				
Group III	$5.6 \pm 0.05$				
Group IV	$7.7 \pm 0.002$				
Group V	$7.75 \pm 0.57$				

Table 2. Distribution of variables in the mother and fetus experiences extracts of B.S							
	Group I	Group II	Group III	Group IV	Group V		
The number of VP + at the day 0	5	5	5	5	5		
The number of pregnant rats	3	5	1	-	1		
The total number of embryos	21	47	12	-	9		
The average number of fetuses per pregnant rat	7	9.4	12	-	9		
The number of resorbed Embryos	0	9	0	-	0		
Resorbed/ total embryos Ratio	0	0.19 <sup>a</sup>	0	-	0		
The crown-rump length of embryos, cm	$1.9\pm0.09$	$1.54\pm0.125$	$0.5\pm\!.0001^{\mathrm{b}}$		$1.5\pm0.0001$		
the crown-rump length of embryos, cm/ The to- tal number of embryos Ratio	$0.27\pm0.03$	$0.23 \pm 0.08$	$0.04 \pm 0.001$	-	$0.16\pm0.025$		

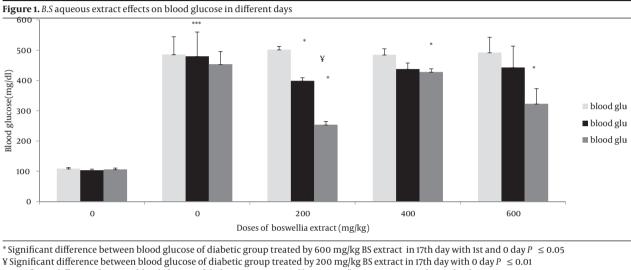
Comparison resorbtion/total embryos Ratio in the different groups:

 $^{a}$  Significant difference between diabetic group with control group and diabetic groups receiving 200 and 600 mg/ kg B.S extract.  $P \leq 0.001$ 

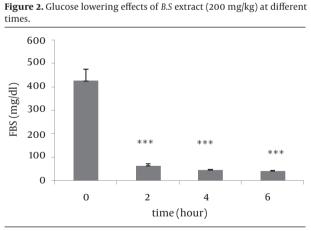
<sup>b</sup> Significant difference between the crown-rump length of embryos in diabetic group receiving 200mg/ kg B.S extract with control group and diabetic groups receiving 600 mg/kg B.S extract. *P* ≤ 0.01

(*Table 1*). Administration of *B.S* in diabetic rats significantly decreased the level of blood glucose and HbAic at day 17 of gestation ( $P \le 0.01$ ). Among the examined doses, the most effective dose of *B.S* extract was 200 mg/kg. In order to evaluate the time-course activity of 200 mg/kg dose, fasting blood glucose was measured at the 2, 4, and 6 hours after extract administration (*Figure 2*).

normal and hepatocytes are normal, although some are staining more vulnerable but the core of some hepatocytes such as vesicular cytoplasm is eosinophilic and cells are seen with two nuclei (*Figure 3*). In diabetic group, size of hepatocytes was small, in addition to endothelial cells around central vein, cells with multi nucleus are gathered around central vein that have normal staining. The classic lobules around central vein in the cumula-



\* Significant difference between blood glucose of diabetic group treated by 200 mg/kg BS extract in 17 day with 1 day  $P \le 0.01$ \* Significant difference between blood glucose of diabetic group treated by 200 mg/kg BS extract in 17 day with 1 day  $P \le 0.05$ \* Significant difference between blood glucose of diabetic group treated by 200 mg/kg BS extract in 17 day with 1 day  $P \le 0.05$ \* Significant difference between blood glucose of diabetic group treated by 200 mg/kg BS extract in 17 day with 0 day  $P \le 0.05$ \* Significant difference between blood glucose of diabetic group treated by 400 mg/kg BS extract in 17 day with 1 and 0 day of diabetic  $P \le 0.05$ \*\* Significant difference between HbAtc of diabetic group with that of control group  $P \le 0.001$ 



\*\*\* Significant difference between blood glucose of diabetic group treated by 200 mg/kg BS extract before and after 2, 4 and 6 hours.  $P \le 0.001$ 

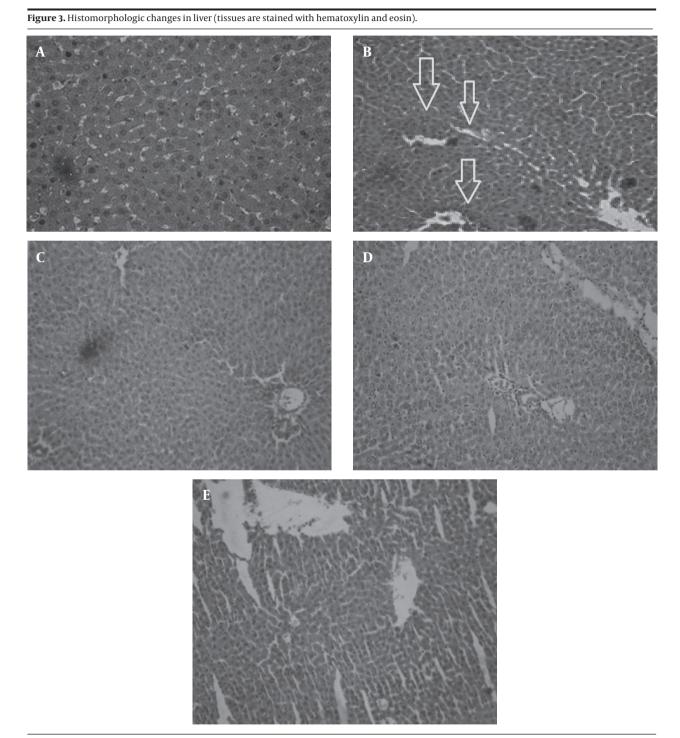
### 4.1. Histomorphologic Changes in Liver

The area around the port, hepatocytes, sinusoids, space around sinusoids, interstitial area, and areas around the central veins were observed in liver sections. In each slide ten fields were examined. In control group, endothelial cells around central vein are appropriately regular and their origins are not clear whether endothelial cells or hepatocytes. Some parts of the sinusoids are swollen. Sinusoid spaces are guite inflated and dilated which could be due to decreased glycogen stores, it is not observed in all the tissues, but in some parts of this expansion is well evident. Fat accumulation is seen in some cells that can result in diabetes (Figure 3). In treated group by 200 mg/ kg B.S extract: Hepatocyte and central vein are normal, the inflammation is not in sinusoid and nuclear accumulation is not seen around central vein, hepatocytes are small (Figure 3). In treated group by 400 mg/kg B.S extract: Cells are less eosinophilic, glycogen stores are less, central vein and cells are normal. There is not seen any inflammation, nuclear accumulation is not seen around central vein, but inflation is seen around some of them (Figure 3). In treated group by 600 mg/kg B.S extract: Inflammation is present in sinusoid cells around central vein and sinusoids are seen fully dilate (Figure 3).

tive core abnormality that occurs in the nucleus, which

### 4.2. Histomorphologic Changes in Kidney

Distal and proximal tubules, Bowman capsule, Urinary tubules, glomeruli Bulb and <u>malpighi</u> body were examined in kidney. In control group, Cortexes and Bowman capsule are healthy, collecting tubes in the medullar appear normal (*Figure 4*). In diabetic group, Cell lyses occurred in the tubules of the kidney and cortex has been damaged, the distal and proximal tubules were destructed. The cortex of is not normal intact but Bowman's capsule is not injured. Urinary space dilated, and the Malpighi body is wrinkled in some parts (*Figure 4*). In treated group by 200 mg/kg B.S extract: Cortex, Bowman capsule and collecting tubes in the medullar are normal (*Figure* 



A: Photomicrograph of liver from healthy animals receiving vehicle. (X = 100).

B: Photomicrograph of liver from diabetic animals receiving vehicle. (X = 100).

C: Photomicrograph of liver from diabetic rats treated by 200 mg/kg BS extract dosage (X = 100)

D: Photomicrograph of liver from diabetic rats treated by 400 mg/kg BS extract dosage (X = 100)

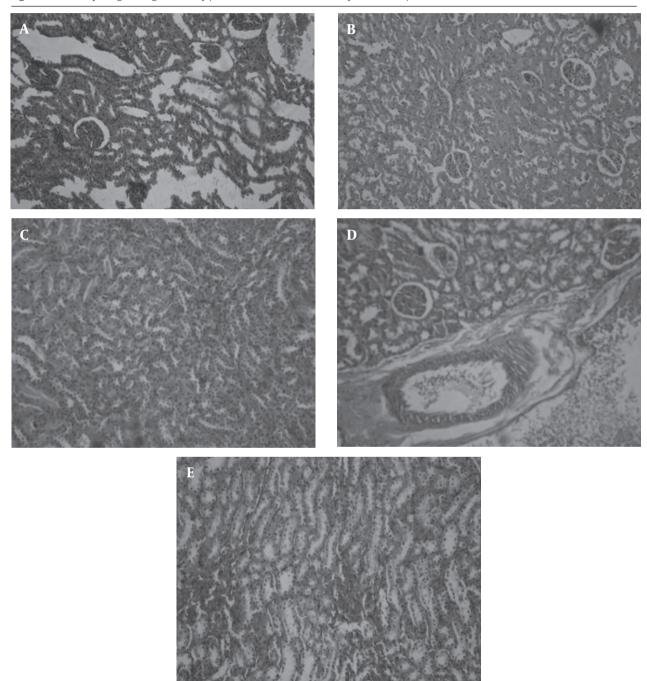
E: Photomicrograph of liver from diabetic rats treated by 600 mg/kg BS extract) dosage (X = 100)

4c).In treated group by 400 mg/kg B.S extract, some glomeruli are damaged and wrinkled. Dilation can be seen in the space between the tubes and in the urine lumen in the cortex and medulla (*Figure 4d*). In treated group by 600 mg/kg B.S extract: Slightly dilatation can be seen in medulla, cortical veins are dilated, glomeruli shrinkage affected cortex (*Figure 4e*).

## 4.3. The Fetus Data

The pregnancy rate in group I to V was 60, 100, 20, 0, and

Figure 4. Histomorphologic changes in kidney (tissues are stained with hematoxylin and eosin).



A: Photomicrograph of renal from healthy animals receiving vehicle (X = 100). B: Photomicrograph of renal from diabetic animals receiving vehicle (X = 100). C: Photomicrograph of renal from diabetic rats treated by of 200 mg/kg BS extract dosage (X = 100) D: Photomicrograph of renal of diabetic animals treated by 400 mg/kg BS extract dosage (X = 100) E: Photomicrograph of renal of diabetic animals treated by 600 mg/kg BS extract dosage (X = 100) 20% respectively. *B.S* probably could decreases pregnancy rate. Based on the results of diabetic rats received extract were not sufficient for argument the group I and II were used for discussion of fetus data. The average number of fetuses per pregnant rat in groups I and II was 7 and 9.4 respectively. The mean crown-rump length of fetus in groups I and II was 1.9 and 1.54 cm respectively. The fetus resorption in groups I and II was 0 and 0.19 respectively.

## 5. Discussion

Oxidative stress recently as one of the mechanisms in diabetes mellitus, that carbohydrate metabolism, lipids and proteins are affected, has been proposed.diabetes is characterized by increase oxidative stress and hyperglycemia was shown to directly induce oxidative stress by depleting natural anti-oxidants and facilitating the production of reactive oxygen species (ROS) under diabetic conditions ROS are produced through glycation reaction, which occurs in various tissues (25). Several changes and features are oxidative nature in people with diabetes or that are associated with the increased oxidative stress (26). Glycation of some compounds (27), and false -hypoxia induced by hyperglycemia (28), can be a position in the oxide and Reducing imbalances within cells to cause liver and kidney tissues.

It has been reported that the anti-oxidants vitamin C or vitamin E, protect beta cells from destruction in diabetic animal models (25). The present study shows that B.S extract could reduce blood sugar and HbAtc level in diabetic rats. B.S extracts at the dose of 200 mg/kg showed the highest effect compared with other doses. Our study shows that there reduction two hours after administration of the extract reaches its highest and up to six hours after administration of the extract is continuing.

Chronic high blood glucose increases free radicals production and oxidative stress which causes damage in other organs including the liver and kidneys. One of the most damaging effects of free radical initiated lipid peroxidation, is destruction of cell membranes. Histopathological changes in the liver caused by diabetes are including inflammation around the portal vein, fibrosis and vein enlargement, the hyperama in sinusoid, granular destruction, cell necrosis, the formation of macrovascular and microvascular vacuoles (2). In the histopathological study of liver in diabetic group, lymphocytic inflammation in the port areas, irregularities, apoptosis of liver cells, and dilatation of the sinuosoids were observed (*Figure 3*).

Glomerular hypertrophy occurs in diabetic nephropathy, hyper filtration occurs due to the enlargement of the glomeruli. The dimensions and size of renal glomeruli, glomerular area, mesenchymal matrix cells and glomerular basement membrane increases in diabetes, and a mild decrease occurs in size parenchymal matrix (1). Microscopic slides of the liver and kidney tissues demonstrates that B.S with has the effects anti-oxidant and antihyperglycemia Protective effects against damage caused by diabetes. B.S extract in doses of 200, and 400 mg/kg can largely reduce the harmful effects of diabetes, that including lymphocytic inflammation in the port areas, irregularities, apoptosis of liver cells, and dilatation of the sinuosoids were not observed. B.S extract at the dose of 600 mg/ml did not show a protective effect on liver and also cause cell death, inflammation and vasodilatation in sinuosoids space (*Figure 3*).

In the present study in groups treated by 200 and 400 mg/kg extract any inflammation and accumulation in the liver tissue was observed. All of symptoms that mentioned were improved in B.S treatment groups. Administration of 600 mg/kg of extract in the diabetic group had increased symptoms. The use of B.S extract in 200,400 doses could help prevent complications of diabetes in the kidneys (*Figures 4 c and d*). These complications were including: cell lyses occurred in the tubules of the kidney and cortex has been damaged, the distal and proximal tubules were destructed. The cortex of is not normal intact but Bowman's capsule is not injured. Urinary space dilated, and the Malpighi body is wrinkled in some parts. (*Figures 1 and 2 b*)

B.S extracts used in appropriate doses, while having anti-hyperglycemia effect on blood sugar can be prevented complications of diabetes in liver and kidney. Boswellia serrata could probably decrease the pregnancy rate. This data suggests that diabetes increases the number of fetus, significantly fetus resorption, and decreases the fetus length. The crown-to-rump length is likely a reflection of a general environmental stress factor during embryonic growth (23). In previous studies conducted in this field has also been shown that fetuses of diabetic mothers give birth to smaller ones. In the diabetic groups a decrease was seen in the crown-rump length of embryos (24). Studies show that the rate of spontaneous abortion, miscarriage and perinatal mortality in diabetic mothers was higher in the poor glycemic control groups (5).

# Acknowledgments

This work was extracted from pharm. D. thesis and financially supported by Jundishapur University of Medical Sciences, Ahvaz, Iran

## **Financial Disclosure**

None declared.

## **Funding/Support**

None declared.

### **References:**

 Masjedi F, Gol A, Dabiri S, Javadi A. Preventive Effect of Garlic on Histopathology of Liver and Markers of Hepatic Injury in Streptozotocin-Induced Diabetic Rats. Iran J Endocrin Metab. 2009;11(4):433-41.

- Khaki A, Khaki A, Nouri M, Ahmadi-Ashtiani H, Rastegar H, Rezazadeh S, et al. Evaluation effects of quercetin on liver apoptosis in streptozotocin-induced diabetic rat. J Med Plan. 2009;8(Supplement 5):70-8.
- Guven A, Yavuz O, Cam M, Ercan F, Bukan N, Comunoglu C, et al. Effects of melatonin on streptozotocin-induced diabetic liver injury in rats. Acta Histochem. 2006;108(2):85-93.
- 4. Rashki KM, Gol A, Dabiri S. Preventive Effects of Garlic Juice on Renal Damages Induced by Diabetes Mellitus in Rats. *Iran J Endocrin Metab.* 2009;**3**:331-9.
- Inkster ME, Fahey TP, Donnan PT, Leese GP, Mires GJ, Murphy DJ. Poor glycated haemoglobin control and adverse pregnancy outcomes in type 1 and type 2 diabetes mellitus: systematic review of observational studies. *BMC Pregnancy Childbirth*. 2006;6:30.
- Goldman AS, Baker L, Piddington R, Marx B, Herold R, Egler J. Hyperglycemia-induced teratogenesis is mediated by a functional deficiency of arachidonic acid. *Proc Natl Acad Sci U S A*. 1985;82(23):8227-31.
- Wentzel P, Welsh N, Eriksson UJ. Developmental damage, increased lipid peroxidation, diminished cyclooxygenase-2 gene expression, and lowered prostaglandin E2 levels in rat embryos exposed to a diabetic environment. *Diabetes*. 1999;48(4):813-20.
- Eriksson UJ, Wentzel P, Minhas HS, Thornalley PJ. Teratogenicity of 3-deoxyglucosone and diabetic embryopathy. *Diabetes*. 1998;47(12):1960-6.
- 9. Vitamin D. Alternative Medicine Review. 2008;13(2):153-64.
- Krohn K, Rao MS, Raman NV, Khalilullah M. High-performance thin layer chromatographic analysis of anti-inflammatory triterpenoids from Boswellia serrata Roxb. *Phytochem Anal.* 2001;12(6):374-6.
- al-Awadi F, Fatania H, Shamte U. The effect of a plants mixture extract on liver gluconeogenesis in streptozotocin induced diabetic rats. *Diabetes Res.* 1991;18(4):163-8.
- Borrelli F, Capasso F, Capasso R, Ascione V, Aviello G, Longo R, et al. Effect of Boswellia serrata on intestinal motility in rodents: inhibition of diarrhoea without constipation. Br J Pharmacol. 2006;148(4):553-60.
- 13. Gupta I, Gupta V, Parihar A, Gupta S, Ludtke R, Safayhi H, *et al.* Effects of Boswellia serrata gum resin in patients with bronchial asthma: results of a double-blind, placebo-controlled, 6-week clinical study. *Eur J Med Res.* 1998;3(11):511-4.
- Gupta I, Parihar A, Malhotra P, Singh GB, Ludtke R, Safayhi H, et al. Effects of Boswellia serrata gum resin in patients with ulcerative colitis. Eur J Med Res. 1997;2(1):37-43.

- Shao Y, Ho CT, Chin CK, Badmaev V, Ma W, Huang MT. Inhibitory activity of boswellic acids from Boswellia serrata against human leukemia HL-60 cells in culture. *Planta Med.* 1998;64(4):328-31.
- Kulkarni RR, Patki PS, Jog VP, Gandage SG, Patwardhan B. Treatment of osteoarthritis with a herbomineral formulation: a double-blind, placebo-controlled, cross-over study. *J Ethnopharmacol*. 1991;33(1-2):91-5.
- Y J, Kamath JV, Asad M. Effect of hexane extract of Boswellia serrata oleo-gum resin on chemically induced liver damage. *Pak J Pharm Sci.* 2006;**19**(2):129-33.
- Kavitha JV, Rosario JF, Chandran J, Anbu P, Bakkiyanathan. Hypoglycemic and other related effects of Boswellia glabra in alloxaninduced diabetic rats. *Indian J Physiol Pharmacol.* 2007;**51**(1):29-39.
- Namjooyan F, Azemi M, Rahmanian V. Investigation of antioxidant activity and total phenolic content of various fractions of aerial parts of pimpinella barbata (dc.) Boiss. Jundishapur J Nat Pharm Prod. 2012;5(1):1-5.
- Heinonen IM, Lehtonen PJ, Hopia AI. Antioxidant Activity of Berry and Fruit Wines and Liquors. J Agric Food Chem. 1998;46(1):25-31.
- Hemmati S, Jeddi-Tehrani M, Torkabadi E, Ghassemi J, Akhondi MM, Zarnani AH. Indoleamine 2, 3-dioxygenase (IDO) is expressed at feto-placental unit throughout mouse gestation: An immunohistochemical study. J Reprod Infertil. 2009;10(3):177-83.
- 22. Mohamed RA, Ramadan RS, Ahmed LA. Effect of substituting pumpkin seed protein isolate for casein on serum liver enzymes, lipid profile and antioxidant enzymes in CCl4-intoxicated rats. *Advanc in Biol Res.* 2009;**3**(1-2):9-15.
- Eriksson UJ, den Bieman M, Prins JB, van Zutphen LFM, editors. Differences in susceptibility for diabetes induced malformations in separated rat colonies of common origin. Proc 4th FELA-SA Symp; 1990; Lyon, France.
- 24. Wiznitzer A, Ayalon N, Hershkovitz R, Khamaisi M, Reece EA, Trischler H, *et al.* Lipoic acid prevention of neural tube defects in offspring of rats with streptozocin-induced diabetes. *Am J Obstet Gynecol.* 1999;**180**(1 Pt 1):188-93.
- Koren-Gluzer M, Aviram M, Meilin E, Hayek T. The antioxidant HDL-associated paraoxonase-1 (PON1) attenuates diabetes development and stimulates beta-cell insulin release. *Atherosclerosis*. 2011;**219**(2):510-8.
- Baynes JW. Role of oxidative stress in development of complications in diabetes. *Diabetes*. 1991;40(4):405-12.
- 27. Vlassara H, Bucala R, Striker L. Pathogenic effects of advanced glycosylation: biochemical, biologic, and clinical implications for diabetes and aging. *Lab Invest.* 1994;**70**(2):138-51.
- Williamson JR, Kilo C. Capillary basement membranes in diabetes. *Diabetes*. 1983;32 Suppl 2:96-100.