

Synthesis of σ Receptor Ligands with a Spirocyclic System Connected with a Tetrahydroisoquinoline Moiety via Different Linkers

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With the aim to develop new σ_2 receptor ligands, spirocyclic piperidines or cyclohexanamines with 2-benzopyran and 2-benzofuran scaffolds were connected to the 6,7-dimethoxy-1,2,3,4-tetrahydroisoquinoline moiety by variable linkers. In addition to flexible alkyl chains, linkers containing an amide as functional group were synthesized. The 2-benzopyran and 2-benzofuran scaffold of the spirocyclic compounds were synthesized from 2-bromobenzaldehyde. The amide linkers were constructed by acylation of amines with chloroacetyl chloride and subsequent nucleophilic substitution, the alkyl linkers were obtained by LiAlH₄ reduction of the corresponding amides. For the development of σ_2 receptor ligands, the spirocyclic 2-

1. Introduction

 σ Receptors, initially classified as class of opioid receptors, are well established as unique class of receptors without any homology to opioid receptors or NMDA receptors. ^[1] Based on the results of comprehensive radioligand binding studies and biochemical analysis, the class of σ receptors was further divided into two distinct subtypes, which were termed σ_1 and σ_2 receptor. ^[2]

The σ_1 receptor has been cloned from different species, including human, rat, mouse, and guinea pig. The crystal structure of the human σ_1 receptor was recently reported by Kruse et al.^[3,4] In contrast to the σ_1 receptor, details concerning the σ_2 receptor have been rather vague for many years. As a result from photoaffinity labeling studies a molecular weight of 21.5 kDa was postulated for the σ_2 receptor.^[5] Xu and coworkers utilized a photoaffinity probe to label σ_2 receptors in rat liver and proposed that the σ_2 receptor binding site resides within the progesterone receptor membrane component 1 (PGRMC1) complex.^[6] During the following years, the correlation between the σ_2 receptor and PGRMC1 protein complex was considered controversial.^[7] In 2017, the σ_2 receptor was isolated

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© 2020 The Authors. ChemMedChem published by Wiley-VCH GmbH. This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited. benzopyran scaffold is more favorable than the ring-contracted 2-benzofuran system. Compounds bearing an alkyl chain as linker generally show higher σ affinity than acyl linkers containing an amide as functional group. A higher σ_1 affinity for the *cis*-configured cyclohexanamines than for the *trans*-configured derivatives was found. The highest σ_2 affinity was observed for *cis*-configured spiro[[2]benzopyran-1,1'-cyclohexan]-4'-amine connected to the tetrahydroisoquinoline system by an ethylene spacer (*cis*-**31**, K_i (σ_2) = 200 nM; the highest σ_1 affinity was recorded for the corresponding 2-benzofuran derivative with a CH₂C=O linker (*cis*-**29**, K_i (σ_1) = 129 nM).

from calf liver tissue and identified as the endoplasmic reticulum (ER)-resident membrane protein TMEM97, which is also described as MAC30 (meningioma-associated protein 30). Subsequent molecular cloning and binding experiments confirmed this result. Mutagenesis studies identified two aspartate residues as crucial for binding of [3H]DTG, a radioligand frequently used in σ_2 receptor binding assays. Furthermore, it was demonstrated that the TMEM97 ligands elacridar (1; Figure 1) and Ro 48-8071 showed the same K_i values towards cell membranes from Sf9 cells overexpressing the TMEM97 protein and σ_2 receptor overexpressing MCF-7 cells.^[8] According to these findings, the σ_2 receptor is now often termed σ_2 receptor/TMEM97. In 2018, Riad et al. demonstrated that the σ_2 receptor/TMEM97 protein, the PGRMC1 protein and the LDL receptor form a ternary complex, which is necessary for the rapid internalization of LDL.^[9] In contrast to the σ_1 receptor, no crystal structure of the σ_2 receptor protein has been published so far.



Figure 1. Elacridar and some prototypical σ_2 receptor ligands.



For a variety of tumor cells an overexpression of σ_2 receptors was demonstrated, including breast cancer, lung cancer, colon cancer, leukemia and prostate cancer.^[10-15] It was also shown that σ_2 receptor agonists are capable of killing tumor cells via apoptotic and non-apoptotic mechanisms. For example, several derivatives of the high affinity σ_2 receptor agonist PB28 (3; Figure 1) are able to inhibit the growth of pancreatic cancer cells and the neuroblastoma SK-N-SH cell line.^[16,17] Very recently, it has been found that the potent and selective σ_2 receptor ligand PB221 inhibits the proliferation of brain tumor murine astrocytoma cells (ALTS1C1).^[18] Haloperidol and its homopiperazine analog SYA013 exhibit high σ_2 affinity and, furthermore, antiproliferative effects on different tumor cell lines, including Panc-1.^[19] Therefore, the development of σ_2 receptor ligands is a very promising goal. However, very recently it was reported that σ_2 receptor ligands could also induce cytotoxic effects in σ_2 /TMEM97 knock out and σ_2 / TMEM97 and PGRMC1 double knock out cell lines. It was concluded that the cytotoxic effects of these σ_2 ligands could not be mediated by the σ_2 receptor, but other mechanisms have to be responsible for these cytotoxic effects.^[20]

In Figure 1, some prototypical σ_2 receptor ligands are shown. The spirocyclic benzofuran siramesine (2) displays a considerable selectivity for the σ_2 receptor ($K_i = 0.12$ nM) over the σ_1 receptor ($K_i = 17$ nM).^[21] PB28 (3) with the 4-(cyclohexyl) piperazine substructure is also a potent σ_2 ligand ($K_i = 0.68$ nM), but exhibits even higher affinity towards the σ_1 subtype ($K_i = 0.38$ nM).^[22] In the group of bicyclic compounds some morphans (e.g., CB184, 4) and granatanes (e.g., SV119, 5) display high σ_2 receptor affinity and high selectivity over the σ_1 receptor.^[23,24]

The 6,7-dimethoxy-1,2,3,4-tetrahydroisoquinoline residue is a pharmacophoric element present in several σ_2 receptor ligands.^[25-33] (Figure 2) Mach and co-workers published a series of benzamides connected to the 6,7-dimethoxy-1,2,3,4-tetrahydroisoquinoline residue by linkers of different chain lengths. Among this series, **7** and **8** (ISO-1^[32]) with an ethylene and tetramethylene linker, respectively, showed high σ_2 affinity and



Figure 2. Lead compounds with a tetrahydroisoquinoline ring system showing a preference for σ_2 receptors over σ_1 receptors.

selectivity over the σ_1 receptor.^[33] The same isoquinoline ring system is also a structural element of the σ_2 ligand 1 (Figure 1).

In a previous study, we have reported that the spirocyclic 2benzopyran derivatives trans-6 and cis-6 bearing the 6,7dimethoxy-1,2,3,4-tetrahydroisoguinoline residue without linker show medium to high affinity to the σ_2 receptor.^[34] (Figure 2) However, the selectivity over the σ_1 subtype is moderate and has room for improvement. Therefore, it was envisaged to synthesize a new set of σ_2 selective ligands by introducing a linker between the spirocyclic 2-benzopyran scaffold and the isoquinoline ring system. To exploit further structure affinity relationships, not only alkyl chains were planned as linkers, but also amides with variable chain lengths and different positions of the carbonyl group were designed. Moreover, a ring contraction of the spirocyclic 2-benzopyran to the spirocyclic 2benzofuran ring system was planned as this compound class is also known for its high σ affinity from previous studies. An overview of the structure modifications is presented in Figure 3.

2. Results and Discussion

2.1. Synthesis

For the synthesis of the designed σ ligands, spirocyclic 2benzopyrans and 2-benzofurans **10–13** with endocyclic and exocyclic amino moiety were synthesized starting from 2bromobenzaldehyde (Scheme 1).

The spirocyclic piperidines **10** and **12** were prepared by addition of an aryllithium intermediate at *N*-benzyl-protected piperidin-4-one as previously described.^[35,36] The exocyclic primary amines *trans*-**11** and *cis*-**11** were obtained as reported in ref. [34]. Transfer hydrogenolysis using NH₄ HCO₂ in the presence of Pd/C converted *trans*- and *cis*-configured benzyl-amines *trans*-**9** and *cis*-**9**^[37,38] into the diastereomeric primary



Figure 3. Overview of planned σ_2 receptor ligands with various linkers.

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Scheme 1. Outline of the synthesis of spirocyclic piperidines $10^{(35)}$ and 12.⁽³⁶⁾ and cyclohexanamines $11^{(34)}$ and 13 with 2-benzopyran and 2-benzofuran scaffold. a) 5 steps;⁽³⁵⁾ b) 7 steps;⁽³⁴⁾ c) 4 steps;⁽³⁶⁾ d) 4 steps;^(37,38) e) NH₄ HCO₂, Pd/C, CH₃OH, 17–21 h, 65 °C; *trans*-13, 86 %, *cis*-13, 66 %.

amines *trans*-**13** and *cis*-**13**. The secondary amine 6,7-dimethoxy-1,2,3,4-tetrahydroisoquinoline HCI (**14** HCI) was commercially available (Scheme 1).

The amines **10–14** were acylated with α -chloroacetyl chloride affording chloroacetamides **15**^[39]–**17** and **19–20**. The homologous 4-chlorobutyryl derivative *cis*-**18** was prepared by reaction of *cis*-**11** with 4-chlorobutyryl chloride. The amides **15–20** were obtained in yields of 56–89% (Scheme 2). Acylation of tetrahydroisoquinoline **14** with 3-chloropropionyl chloride did not lead to the desired 3-chloropropinamide.

The final compounds were obtained by nucleophilic substitution of the terminal chloride in the side chain of the amides **15–20**. The acylated spirocyclic 2-benzopyrans **16–18** and 2benzofurans **19** and **20** were reacted with the tetrahydroisoquinoline **14**, while the acylated isoquinoline **15** underwent a nucleophilic substitution with the spirocyclic amines **10–13**. In Table 1, the products and yields of these transformations are summarized.

The nucleophilic substitution of the 2-chloroacetylated isoquinoline derivative **15** with spirocyclic amines **10–13** in DMF with TBAI as catalyst resulted in the formation of the desired compounds **21–24** in satisfactory yields. Due to purification problems, the benzopyran-based spirocyclic compound *trans-***22** could not be isolated in pure form for testing. $S_N 2$ reaction of spirocyclic chloroacetamides **16**, **17** and **20** with the tetrahydroisoquinoline **14** provided the amides **25**, **26** and **29** in 62–86% yields. The pure spirocyclic benzofuran **28** was obtained in only 44% yield, due to purification problems. While the nucleophilic substitution of the 2-chloroacetylated compounds **16**, **17**, **19**, and **20** with tetrahydroisoquinoline **14** led to clean conversions, he corresponding 4-chlorobutyramide **18** reacted slower to produce *cis-***27**, which was isolated in only **19%** yield (Table 1).

During the reaction to obtain the secondary amines *trans*-**22**, *cis*-**22**, *trans*-**24** and *cis*-**24**, formation of tertiary amines as side-products was observed (double nucleophilic substitution). The R_f values of the tertiary amines was almost identical to the R_f value of the secondary amines, rendering the fc purification



Scheme 2. Acylation of amines with chloroacyl chlorides. *The spirocyclic piperidines 16 and 19 were not isolated, but directly used for subsequent nucleophilic substitution with 14.

of the desired products very difficult. Although the isolation and purification of the secondary amines *cis*-22, *trans*-24 and *cis*-24 was successful, *trans*-22 could not be isolated in sufficient purity.

As not only linkers bearing a carbonyl group were planned, derivatives **30**, *trans*-**31** and *cis*-**31** with an ethylene linker between the amino moiety of the spirocyclic benzopyran and the tetrahydroisoquinoline were synthesized. (Scheme 3) This type of compounds features two basic amino moieties instead of one and can therefore adopt different orientations within the

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Table 1. Nucleophilic substitution at chloroamides 15–20. ^[a]								
Chloroamide	Amine	Product	Yield [%]					
15	10	21	22					
15	cis-11	cis- 22	60					
15 15	12 trans-13	23 trans-24	36 54					
15	cis-13	cis-24	43					
trans-17	14	trans-26	83					
cis-17 cis-18	14 14	cis- 26 cis- 27	62 19					
19 trans- 20	14 14	28 trans- 29	44 66					
cis- 20	14	cis- 29	86					

[a] For structures, see Scheme 2 and Table 2. *The product could not be isolated.



Scheme 3. Synthesis of isoquinolines 30, trans-31 und cis-31 with an ethylene linker. a) LiAlH₄, THF, 2–22 h, 70 °C, 63 % (30), 86 % (trans-31), 76 % (cis-31).

binding pocket of both σ receptor subtypes. Additionally, the effect of the carbonyl moiety on the binding affinity and selectivity can be studied.

At first, a direct alkylation of the tetrahydroisoquinoline 14 was envisaged. For this purpose, 2-bromoethanol was oxidized with Dess-Martin perioidinane to afford 2-bromoacetaldehyde. The aldehyde should be attached to the isoquinoline 14 in a reductive alkylation with NaBH(OAc)₃. Unfortunately, after 4 h reaction time the alkylated isoquinoline could not be isolated. Next, a nucleophilic substitution with 1,2-dibromoethane and K₂CO₃ in CH₃CN was performed. But even after a reaction time of 18 h the desired product could not be obtained. The reaction conditions which led to a successful acylation of the isoquinoline 14 (DMF, Et₃N and TBAI) also didn't lead to the formation of the alkylated product. Finally, the desired alkylated amines 30, trans-31 and cis-31 were synthesized by reduction of the corresponding amides 25, trans-26 and cis-26 with LiAlH₄. (Scheme 3) The piperidine derivative 30 was obtained in 63% yield after 2 h heating to reflux. trans-31 and cis-31 were isolated after 22 h in 86 and 76% yield, respectively.

2.2. σ_1 and σ_2 receptor affinity

Competitive binding assays with tritiated radioligands were utilized to determine the σ_1 and σ_2 receptor affinity of the synthesized compounds. In the σ_1 binding assay, [³H]-(+)-pent-

azocine was used as radioligand and homogenates of guinea pig brains served as receptor material. The σ_2 assay was performed with the radioligand [³H]-di(o-tolyl)guanidine ([³H] DTG) and homogenates of rat liver were used as receptor material. The nonselective properties of DTG was compensated by masking σ_1 receptors with an excess of non-tritiated (+)-pentazocine.^[40]

In Table 2, the receptor affinities of the synthesized compounds are summarized. In comparison to the lead compounds *trans*-**7** and *cis*-**7**, the 2-benzopyran derivatives with an acetyl linker generally show a lower σ_2 affinity. The highest σ_2 affinity was observed for *cis*-**26** with a K_i value of 371 nM. In this compound, the acyl group is located at the spirocyclic ring system. When the acyl moiety is located at the isoquinoline ring system (*cis*-**22**), the σ_2 affinity is reduced (11% inhibition of radioligand binding). A similar trend was observed for the corresponding piperidine derivatives **25** and **21** with K_i values of 534 nM and 19% inhibition of radioligand binding, respectively.

The σ_1 affinity of the piperidine derivative **21** is higher than that of the corresponding cyclohexanamine derivative *cis*-**22**. A general observation is that the σ_1 affinity is higher for the compounds bearing the acyl group at the isoquinoline ring. For the development of σ_1 ligands with the 2-benzoypran scaffold, it can therefore be concluded that the basic center at the spirocyclic ring system should be retained.

For the derivatives with the 2-benzofuran scaffold similar observations were made in terms of σ_2 affinity. The introduction of an acetyl spacer led to loss of σ_2 affinity, independent of the position of the acyl moiety (e.g., *trans*-**24**, *cis*-**24**, **28**). In contrast to the 2-benzopyrans, the σ_1 affinity of the piperidine derivatives of the spirocyclic 2-benzofurans was not higher than the respective cyclohexanamines. A notable exception is *cis*-**29** with a K_i value of 129 nM at the σ_1 receptor. This compound even represents a σ_1 receptor selective ligand despite the 6,7-dimethoxy-1,2,3,4-tetrahydroisoquinoline structural element.

The elongation of the acyl linker also led to a decrease in σ_2 affinity, while the σ_1 affinity was slightly increased. For the butyramide *cis*-**27** a K_i value of 2100 nM at the σ_2 receptor and 712 nM at the σ_1 receptor was observed.

For the derivatives **30**, *cis*-**31** and *trans*-**31** with an ethylene linker an increased σ_2 affinity in comparison to the corresponding amides (e.g., **21**, *cis*-**22**) was found. The σ_1 receptor affinity of the cyclohexanamines *cis*-**31** and *trans*-**31** was also increased, resulting in a loss of σ_2 preference of *cis*-**31**. The piperidine **30** shows a slight preference for the σ_2 receptor (K_i values of 348 nM and 608 nM, respectively).

3. Conclusion

The introduction of a spacer between the spirocyclic 2benzopyran and 2-benzofuran scaffold and the tetrahydroisoquinoline system was envisaged to study structure affinity relationships and evaluate possibilities to optimize selectivity of the lead compounds *trans*-7 and *cis*-7. A set of compounds with amide and alkyl spacers was synthesized and pharmacologically

Table 2. σ 1 and σ 2 receptor affinities of synthesized compounds.									
$H_{3}CO$ H_{3									
	21, 2	23, 25, 28, 30	cis-/trans-22, 24,	26, 27, 29, 31					
Compound	x	Υ	n	$m{\mathcal{K}_{i}}\left[\mathbf{nM} ight] \pm SEM^{\left[a ight] }$ σ_{1}	σ_2				
trans-6	-	-	1	639	58+27				
cis- 6	_	_	1	1200	105 ± 8				
21	C=O	CH	1	319	19%				
cis- 22	C=O	CH ₂	1	740	11%				
23	C=O	CH	0	0%	0%				
trans-24	C=O	CH	0	15%	0%				
cis- 24	C=O	CH	0	1600	9%				
25	CH ₂	C=0	1	518	534				
trans-26	CH ₂	C=0	1	7%	4%				
cis- 26	CH ₂	C=O	1	1000	371				
cis- 27	(CH ₂)3	C=O	1	712	2100				
28	CH	C=O	0	19%	3400				
trans- 29	CH ₂	C=O	0	1300	1900				
cis- 29	CH ₂	C=O	0	129	0%				
30	CH ₂	CH ₂	1	608	348				
trans-31	CH ₂	CH ₂	1	1400	499				
cis- 31	CH ₂	CH ₂	1	251	200				
[a] <i>K</i> _i values are given a compound.	as means of 3 different expe	eriments; percentage value	s indicate inhibition of th	e radioligand at a concentra	tion of 1 μ M of the test				

evaluated in competitive binding assays. Although the introduction of the linker generally resulted in a loss of σ affinity in comparison to the lead compounds 7 without linker, some interesting observations could be made. Compounds containing the 2-benzopyran scaffold showed a higher affinity than the corresponding 2-benzofurans. Compounds 30, trans-31 and cis-31 with an ethylene linker displayed higher affinity than compounds with an amide in the side chain. The introduction of the linker in compounds 21 and cis-29 resulted in an unexpected selectivity for the σ_1 receptor. In conclusion, the combination of wo promising σ_2 pharmacophoric elements, that is, the connection of an O-containing spirocyclic system with the tetrahydroisoquinoline moiety by different spacers, did not provide high-affinity σ_2 selective ligands. However, the synthesized σ ligands allow an interesting insight into the limitations of acyl chains as linker between the two pharmacophoric elements. cis-31 and trans-31 could serve as a starting point for further structural modifications resulting in higher σ_2 affinity and selectivity.

Experimental Section

Chemistry, General

Unless otherwise noted, moisture sensitive reactions were conducted under dry nitrogen. CH_2Cl_2 was distilled over CaH_2 . THF was distilled over sodium/benzophenone. Thin layer chromatography (tlc): Silica gel 60 F254 plates (Merck). Flash chromatography (fc): Silica gel 60, 40–64 µm (Merck); parentheses include: diameter of the column (*d*), length of the stationary phase (*l*), fraction size (*V*), eluent. Melting point: Melting point apparatus Mettler Toledo MP50 melting point system, uncorrected. MS: microTOF-Q II (Bruker Daltonics); APCI, atmospheric pressure chemical ionization; micro-Tof mass spectrometer (Bruker Daltonics); ESI, electrospray ionization. IR: FTIR spectrophotometer MIRacle 10 (Shimadzu) equipped with ATR technique. Nuclear magnetic resonance (NMR) spectra were recorded on Agilent 600-MR (600 MHz for ¹H, 151 MHz for ¹³C) or Agilent 400-MR spectrometer (400 MHz for ¹H, 101 MHz for ¹³C); δ in ppm related to tetramethylsilane and measured referring to CHCl₃ (δ = 7.26 ppm (¹H NMR) and δ = 77.2 ppm (¹³C NMR)) and CHD₂OD (δ = 3.31 ppm (¹H NMR) and δ = 49.0 ppm (¹³C NMR)); coupling constants are given with 0.5 Hz resolution; the assignments of ¹³C and ¹H NMR signals were supported by 2-D NMR techniques where necessary (data not shown); multiplicities of the signals are abbreviated as follows: s = singlet, d = doublet, t =triplet, q=quartet; dd=doublet of doublets, m=multiplet. HPLC: pump: LPG-3400SD, degasser: DG-1210, autosampler: ACC-3000T, UV-detector: VWD-3400RS, interface: DIONEX UltiMate 3000, data acquisition: Chromeleon 7 (Thermo Fisher Scientific); column: LiChrospher® 60 RP-select B (5 µm), LiChroCART® 250-4 mm cartridge; guard column: LiChrospher[®] 60 RP-select B (5 μm), LiChroCART® 4-4 mm cartridge (no.: 1.50963.0001), manu-CART® NT cartridge holder; flow rate: 1.0 mL/min; injection volume: 5.0 µL; detection at $\lambda = 210$ nm; solvents: A: water with 0.05% (v/v) trifluoroacetic acid; B: acetonitrile with 0.05% (v/v) trifluoroacetic acid: gradient elution: (A%): 0-4 min: 90%, 4-29 min: 90-0%, 29-31 min: 0%, 31–31.5 min: 0→90%, 31.5–40 min: 90%. The purity of all compounds was determined by this method. Unless otherwise mentioned, the purity of all test compounds is higher than 95%.

Synthetic procedures

The synthesis of the spirocyclic piperidines 10 and 12 has been reported in the literature.^[35,36] The synthesis of exocyclic primary amines *trans*-11 and *cis*-11 was described in ref. [34]. The synthesis



of *trans*- and *cis*-configured benzylamines *trans*-**9** and *cis*-**9** was reported in ref. [37] and [38].

trans-3-Methoxy-3H-spiro[[2] benzofuran-1,1'-cyclohexan]-4'-amine (*trans*-13)



A solution of benzylamine trans-9 (248 mg, 0.76 mmol), ammonium formate (255 mg, 4.05 mmol, 5.3 equiv) and 10% Pd/C (35 mg, 0.03 mmol, 4 mol-%) in CH₃OH (15 mL) was heated to reflux for 17 h. The mixture was filtered through Celite, washed with CH₂Cl₂ (150 mL) and concentrated in vacuo. 1 M NaOH (15 mL) was added and the aqueous layer was extracted with CH_2CI_2 (3×15 mL). The combined organic layers were dried (Na₂SO₄), filtered and concentrated in vacuo. Yellow oil, yield 153 mg (86%). C14H19NO2 (233.3 g/ mol). TLC: $R_f = 0.03$ (cyclohexane/ethyl acetate 67:33 + 1% N,Ndimethylethanamine). HRMS (APCI, method 1): m/z 234.1477 (calcd. 234.1489 for $C_{14}H_{20}NO_2$ [*M*H⁺]). ¹H NMR (600 MHz, CD₃OD): $\delta =$ 1.58-1.64 (m, 1H, 2'-H), 1.65-1.76 (m, 3H, 3'-H, 5'-H, 6'-H), 2.01-2.09 (m, 4H, 2'-H, 3'-H, 5'-H, 6'-H), 3.11-3.15 (m, 1H, 4'-H_{equ}), 3.47 (s, 3H, OCH3), 6.04 (s, 1H, 3-H), 7.34-7.38 (m, 2H, 4-H, 5-H), 7.38-7.42 (m, 1H, 6-H), 7.49 ppm (d, J=7.7 Hz, 1H, 7-H). A signal for the NH₂ protons is not observed in the spectrum. ¹³C NMR (151 MHz, CD₃OD): δ = 30.5 (1 C, C-3' or C-5'), 30.7 (1 C, C-3' or C-5'), 34.0 (1 C, C-2'), 35.2 (1 C, C-6'), 47.2 (1 C, C-4'), 54.8 (1 C, OCH₃), 88.0 (1 C, C-1), 106.9 (1 C, C-3), 122.6 (1 C, C-7), 124.2 (1 C, C-4), 128.9 (1 C, C-5), 130.3 (1 C, C-6), 138.8 (1 C, C-3a), 148.8 ppm (1 C, C-7a). FTIR (neat): ν [cm⁻¹]=3364 (N–H), 2924, 2855 (C–H_{alkyl}), 1435, 1366 (C=C_{arom}). Purity (HPLC): 93.9%, $t_{\rm R} = 8.7$ min.

cis-3-Methoxy-3*H*-spiro[[2] benzofuran-1,1'-cyclohexan]-4'-amine (*cis*-13)



A solution of benzylamine cis-9 (414 mg, 1.28 mmol), ammonium formate (406 mg, 6.44 mmol, 5.0 equiv) and 10% Pd/C (55 mg, 0.05 mmol, 4 mol-%) in CH₃OH (25 mL) was heated to reflux for 21 h. The mixture was filtered through Celite, washed with CH₂Cl₂ (200 mL) and concentrated in vacuo. 1 M NaOH (15 mL) was added and the aqueous phase was extracted with CH_2CI_2 (3×15 mL). The combined organic layers were dried (Na₂SO₄), filtered and concentrated in vacuo. Yellow oil, yield 198 mg (66%). C14H19NO2 (233.3 g/ mol). TLC: R_f=0.02 (cyclohexane/ethyl acetate 67:33+1% N,Ndimethylethanamine). HRMS (APCI, method 1): m/z 234.1479 (calcd. 234.1489 for $C_{14}H_{20}NO_2$ [*M*H⁺]). ¹H NMR (600 MHz, CD₃OD): $\delta =$ 1.63-1.68 (m, 1H, 2'-H), 1.68-1.76 (m, 2H, 3'-H, 5'-H), 1.82-1.89 (m, 4H, 3'-H, 5'-H, 6'-H), 1.93 (td, J=13.5/4.1 Hz, 1H, 2'-H), 2.82 (tt, J= 11.5/3.8 Hz, 1H, 4'-H_{ax}), 3.49 (s, 3H, OCH₃), 6.05 (s, 1H, 3-H), 7.23 (d, J=7.4 Hz, 1H, 7-H), 7.32-7.41 ppm (m, 3H, 4-H, 5-H, 6-H). A signal for the NH₂ protons is not observed in the spectrum. ¹³C NMR (151 MHz, CD₃OD): δ = 32.5 (1 C, C-3' or C-5'), 32.9 (1 C, C-3' or C-5'), 37.6 (1 C, C-2'), 38.9 (1 C, C-6'), 50.5 (1 C, C-4'), 54.9 (1 C, OCH₃), 87.1 (1 C, C-1), 107.1 (1 C, C-3), 121.7 (1 C, C-7), 124.2 (1 C, C-4), 128.9 (1 C, C-5), 130.4 (1 C, C-6), 138.7 (1 C, C-3a), 148.6 ppm (1 C, C-7a). FTIR (neat): ν [cm⁻¹]=3356 (N–H), 2928, 2859 (C–H_{alkyl}), 1458, 1439 (C=C_{arom}). Purity (HPLC): 99.3 %, t_R = 10.5 min.

2-Chloro-1-(6,7-dimethoxy-3,4-dihydroisoquinolin-2(1*H*)-yl) ethan-1-one (15)



The compound was synthesized according to the literature.^[39] 2-Chloroacetyl chloride (0.12 mL, 1.51 mmol, 1.2 equiv) was slowly added to a solution of isoquinoline 14·HCl (281 mg, 1.22 mmol) and Et_3N (0.42 mL, 3.03 mmol, 2.5 equiv) in CH_2Cl_2 (30 mL) under N_2 at 0 °C. After stirring for 4.5 h at RT, $\rm H_2O$ (50 mL) was added and the aqueous layer was extracted with CH_2Cl_2 (3 \times 50 mL). The combined organic layers were dried (Na₂SO₄), filtered, concentrated in vacuo and the residue was purified by fc (d=2.5 cm, I=20 cm, V=10 mL, cyclohexane/ethyl acetate 67:33). Pale yellow solid, m.p. 109°C, yield 247 mg (75%). C₁₃H₁₆CINO₃ (269.7 g/mol). R_f=0.26 (cyclohexane/ethyl acetate 50:50). HRMS (APCI): m/z 270.0864 (calcd. 270.0891 for C₁₃H₁₇³⁵CINO₃ [*M*H⁺]). ¹H NMR (400 MHz, CDCl₃): δ = 2.80 (t, J = 6.0 Hz, 0.8H, 4-H), 2.89 (t, J = 5.9 Hz, 1.2H, 4-H), 3.73 (t, J=5.9 Hz, 1.2H, 3-H), 3.81-3.88 (m, 6.8H, 3-H, 6-OCH₃, 7-OCH₃), 4.15 (s, 1.2H, COCH2CI), 4.16 (s, 0.8H, COCH2CI), 4.62 (s, 0.8H, 1-H), 4.67 (s, 1.2H, 1-H), 6.59–6.65 ppm (m, 2H, 5-H, 8-H). ¹³C NMR (101 MHz, CDCl₃): $\delta = 27.9$ (0.4 C, C-4), 29.1 (0.6 C, C-4), 40.7 (0.4 C, C-3), 41.3 (0.6 C, COCH2CI), 41.6 (0.4 C, COCH2CI), 44.1 (0.6 C, C-3), 44.6 (0.6 C, C-1), 47.7 (0.4 C, C-1), 56.13 (1.2 C, 6-OCH₃, 7-OCH₃), 56.18 (0.8 C, 6-OCH₃, 7-OCH₃), 109.0 (0.4 C, C-8), 109.5 (0.6 C, C-8), 111.4 (0.6 C, C-5), 111.7 (0.4 C, C-5), 123.7 (0.4 C, C-8a), 124.6 (0.6 C, C-8a), 125.6 (0.6 C, C-4a), 126.7 (0.4 C, C-4a), 148.0 (1.2 C, C-6, C-7), 148.2 (0.8 C, C-6, C-7), 165.5 (0.4 C, C=O), 165.6 ppm (0.4 C, C=O). FTIR (neat): ν $[cm^{-1}] = 2974$, 2932 (C-H_{alkvl}), 1651 (C=O), 1520, 1443 (C=C_{arom}). Purity (HPLC): 99.7 %, $t_{\rm R} = 15.9$ min.

trans-2-Chloro-*N*-(3-methoxy-3,4-dihydrospiro[[2] benzopyran-1,1'-cyclohexan]-4'-yl)acetamide (*trans*-17)



2-Chloroacetyl chloride (19 µL, 0.24 mmol, 1.2 equiv) was slowly added to a solution of amine trans-11 (50 mg, 0.20 mmol) and Et₃N (0.07 mL, 0.50 mmol, 2.5 equiv) in CH_2CI_2 (5 mL) under N_2 at 0 °C. After stirring for 6.5 h at RT, H₂O (30 mL) was added and the aqueous layer was extracted with CH_2CI_2 (3×30 mL). The combined organic layers were dried (Na₂SO₄), filtered, concentrated in vacuo and the residue was purified by fc (d=2 cm, l=16 cm, V=5 mL, cyclohexane/ethyl acetate 50:50). Pale yellow solid, m.p. 144°C, yield 36 mg (56%). $C_{17}H_{22}CINO_3$ (323.8 g/mol). $R_f = 0.22$ (cyclohexane/ethyl acetate 67:33). HRMS (APCI): m/z 292.1080 (calcd. 292.1099 for $C_{16}H_{19}^{35}CINO_2$ [*M*-CH₃OH + H⁺]). ¹H NMR (600 MHz, CD₃OD): δ = 1.68-1.72 (m, 1H, 2'-H), 1.76-1.81 (m, 2H, 3'-H, 5'-H), 1.88 (td, J=14.1/3.8 Hz, 1H, 6'-H), 1.92–1.97 (m, 1H, 6'-H), 2.10-2.25 (m, 3H, 2'-H, 3'-H, 5'-H), 2.81 (dd, J=15.6/7.4 Hz, 1H, 4-H), 2.94 (dd, J=15.6/3.1 Hz, 1H, 4-H), 3.55 (s, 3H, OCH₃), 4.12-4.16 (m, 1H, 4'-H_{equ}), 4.13 (s, 2H, COCH₂Cl), 4.92 (dd, J=7.4/3.1 Hz, 1H, 3-H), 7.10 (dd, J=7.6/1.3 Hz, 1H, 5-H), 7.17 (td, J=7.4/1.3 Hz, 1H, 6-H), 7.20-7.23 (m, 1H, 7-H), 7.29 ppm (dd, J=7.8/1.3 Hz, 1H, 8-H). A signal for the NH proton is not observed in the spectrum. ¹³C NMR (151 MHz, CD₃OD): $\delta = 26.2$ (1 C, C-3'), 26.3 (1 C, C-5'), 32.0 (1 C, C-6'), 34.4 (1 C, C-2'), 36.1 (1 C, C-4), 43.4 (1 C, COCH2CI), 45.8 (1 C, C-

4'), 56.4 (1 C, OCH₃), 77.4 (1 C, C-1), 97.9 (1 C, C-3), 125.7 (1 C, C-8), 127.5 (1 C, C-7), 127.7 (1 C, C-6), 130.1 (1 C, C-5), 132.5 (1 C, C-4a), 143.0 (1 C, C-8a), 169.2 ppm (1 C, C=O). FTIR (neat): ν [cm⁻¹]=3321 (N–H), 2928, 2851 (C–H_{alkyl}), 1636 (C=O), 1547, 1447 (C=C_{arom}). Purity (HPLC): 94.8%, $t_{\rm R}$ =18.9 min.

cis-2-Chloro-*N*-(3-methoxy-3,4-dihydrospiro[[2] benzopyran-1,1'-cyclohexan]-4'-yl)acetamide (*cis*-17)



2-Chloroacetyl chloride (19 µL, 0.24 mmol, 1.2 equiv) was slowly added to a solution of amine cis-11 (50 mg, 0.20 mmol) and Et₃N (0.07 mL, 0.50 mmol, 2.5 equiv) in CH_2Cl_2 (5 mL) under N₂ at 0 °C. After stirring for 6 h at RT, H₂O (30 mL) was added and the aqueous layer was extracted with CH_2Cl_2 (3 \times 30 mL). The combined organic layers were dried (Na₂SO₄), filtered, concentrated in vacuo and the residue was purified by fc (d=2 cm, l=20 cm, V=10 mL, cyclohexane/ethyl acetate 67:33). Colorless solid, m.p. 148 °C, yield 40 mg (60%). C₁₇H₂₂CINO₃ (323.8 g/mol). R_f=0.44 (cyclohexane/ ethyl acetate 50:50). HRMS (APCI): m/z 292.1072 (calcd. 292.1099 for $C_{16}H_{19}^{35}CINO_2$ [*M*-CH₃OH + H⁺]). ¹H NMR (600 MHz, CD₃OD): $\delta =$ 1.77–1.91 (m, 5H, 2'-H_{ax}, 3'-H, 5'-H, 6'-H_{equ}), 1.92–1.97 (m, 1H, 5'-H), 2.08 (td, J=13.2/3.9 Hz, 1H, 6'-H_{ax}), 2.13 (dq, J=14.0/2.9 Hz, 1H, 2'-H_{enu}), 2.81 (dd, J=15.6/7.5 Hz, 1H, 4-H), 2.94 (dd, J=15.8/2.9 Hz, 1H, 4-H), 3.58 (s, 3H, OCH₃), 3.88 (tt, J = 11.3/4.7 Hz, 1H, 4'-H_{ax}), 4.03 (s, 2H, COCH₂Cl), 4.92 (dd, J=7.5/3.1 Hz, 1H, 3-H), 7.09 (d, J=7.3 Hz, 1H, 5-H), 7.16 (td, J=7.2/1.9 Hz, 1H, 6-H), 7.18-7.24 ppm (m, 2H, 7-H, 8-H). A signal for the NH proton is not observed in the spectrum. ¹³C NMR (151 MHz, CD₃OD): $\delta = 28.7$ (1 C, C-5'), 28.8 (1 C, C-3'), 36.1 (1 C, C-4), 36.5 (1 C, C-2'), 39.1 (1 C, C-6'), 43.3 (1 C, COCH2CI), 49.7 (1 C, C-4'), 56.4 (1 C, OCH₃), 76.9 (1 C, C-1), 97.9 (1 C, C-3), 125.7 (1 C, C-8), 127.6 (1 C, C-7), 127.7 (1 C, C-6), 130.1 (1 C, C-5), 132.6 (1 C, C-4a), 142.5 (1 C, C-8a), 168.6 ppm (1 C, C=O). FTIR (neat): ν [cm⁻¹]= 3302 (N–H), 2932, 2862 (C–H_{alkyl}), 1643 (C=O), 1447 (C=C_{arom}). Purity (HPLC): 97.1 %, *t*_R = 18.6 min.

cis-4-Chloro-*N*-(3-methoxy-3,4-dihydrospiro[[2] benzopyran-1,1'-cyclohexan]-4'-yl)butanamide (*cis*-18)



4-Chlorobutyryl chloride (27.4 µL, 0.24 mmol, 1.1 equiv) was slowly added to a solution of amine *cis.***11** (54 mg, 0.22 mmol) and Et₃N (0.07 mL, 0.50 mmol, 2.3 equiv.) in CH₂Cl₂ (15 mL) under N₂ at 0 °C. After stirring for 4 h at RT, H₂O (30 mL) was added and the aqueous layer was extracted with CH₂Cl₂ (3×30 mL). The combined organic layers were dried (Na₂SO₄), filtered and concentrated in vacuo. Colorless solid, m.p. 126 °C, yield 69 mg (89%). C₁₉H₂₆ClNO₃ (351.9 g/mol). R_f=0.37 (cyclohexane/ethyl acetate 50:50). HRMS (ESI): *m/z* 374.1504 (calcd. 374.1493 for C₁₉H₂₆³⁵ClNO₃Na [*M*Na⁺]). ¹H NMR (400 MHz, CD₃OD): δ =1.78–1.94 (m, 6H, 2'-H, 3'-H, 6'-H), 2.02–2.17 (m, 4H, 2'-H, 6'-H, COCH₂CH₂CL), 2.39 (t, *J*=7.3 Hz, 2H, COCH₂CH₂CH₂CL), 2.82 (dd, *J*=15.7/7.5 Hz, 1H, 4-H), 2.94 (dd, *J*= 15.7/3.2 Hz, 1H, 4-H), 3.60 (s, 3H, OCH₃), 3.63 (t, *J*=6.5 Hz, 2H,

COCH₂CH₂CH₂CH), 3.81–3.91 (m, 1H, 4'-H_{ax}), 4.93 (dd, J=7.4/3.2 Hz, 1H, 3-H), 7.10 (d, J=7.0 Hz, 1H, 5-H), 7.14–7.26 ppm (m, 3H, 6-H, 7-H, 8-H). A signal for the NH proton is not observed in the spectrum. ¹³C NMR (151 MHz, CD₃OD): δ =28.9 (1 C, C-3' or C-5'), 29.1 (1 C, C-3' or C-5'), 30.0 (1 C, COCH₂CH₂CH₂CI), 34.2 (1 C, COCH₂CH₂CH₂CI), 36.1 (1 C, C-4), 36.5 (1 C, C-2'), 39.1 (1 C, C-6'), 45.1 (1 C, COCH₂CH₂CH₂CI), 49.1 (1 C, C-4'), 56.4 (1 C, OCH₃), 77.0 (1 C, C-1), 97.8 (1 C, C-3), 125.7 (1 C, C-8), 127.5 (1 C, C-6 or C-7), 127.7 (1 C, C-6 or C-7), 130.1 (1 C, C-5), 132.6 (1 C, C-4a), 142.5 (1 C, C-8a), 174.1 ppm (1 C, C=O). FTIR (neat): ν [cm⁻¹]=3306 (N–H), 2928, 2862 (C–H_{alkyl}), 1636 (C=O), 1543, 1443 (C=C_{arom}). Purity (HPLC): 72.3 %, t_{R} =19.6 min.

trans-2-Chloro-*N*-(3-methoxy-3*H*-spiro[[2] benzopyran-1,1'-cyclohexan]-4'-yl)acetamide (*trans*-20)



2-Chloroacetyl chloride (20 µL, 0.25 mmol, 1.2 equiv) was slowly added to a solution of amine trans-13 (49 mg, 0.21 mmol) and Et₃N (0.07 mL, 0.50 mmol, 2.4 equiv) in CH_2CI_2 (10 mL) under N_2 at 0 °C. After stirring for 6 h at RT, H₂O (30 mL) was added and the aqueous layer was extracted with CH_2CI_2 (3×30 mL). The combined organic layers were dried (Na₂SO₄), filtered, concentrated in vacuo and the residue was purified by fc (d=2 cm, l=20 cm, V=5 mL, cyclohexane/ethyl acetate 67:33). Colorless solid, m.p. 123°C, yield 45 mg (69%). C₁₆H₂₀CINO₃ (309.8 g/mol). R_f=0.50 (cyclohexane/ ethyl acetate 50:50). HRMS (ESI): m/z 332.1037 (calcd. 332.1024 for $C_{16}H_{20}^{35}$ CINO₃Na [*M*Na⁺]). ¹H NMR (600 MHz, CD₃OD): δ = 1.61–1.66 (m, 1H, 2'-H), 1.78 (dddd, J=13.6/5.7/4.1/1.9 Hz, 1H, 6'-H_{enu}), 1.83-1.92 (m, 2H, 3'-H, 5'-H), 1.97–2.15 (m, 4H, 2'-H, 3'-H, 5'-H, 6'-H_{ax}), 3.48 (s, 3H, OCH₃), 4.07–4.14 (m, 1H, 4'- H_{equ}), 4.11 (s, 2H, COCH₂Cl), 6.05 (s, 1H, 3-H), 7.35–7.38 (m, 2H, 4-H, 5-H), 7.38–7.43 ppm (m, 2H, 6-H, 7-H). A signal for the NH proton is not observed in the spectrum. ¹³C NMR (151 MHz, CD₃OD): $\delta = 27.4$ (1 C, C-3' or C-5'), 27.6 (1 C, C-3' or C-5'), 34.0 (1 C, C-2'), 35.3 (1 C, C-6'), 43.4 (1 C, COCH2CI), 46.6 (1 C, C-4'), 55.0 $(1 C, OCH_3)$, 87.3 (1 C, C-1), 107.0 (1 C, C-3), 122.1 (1 C, C-7), 124.3 (1 C, C-4), 129.0 (1 C, C-5), 130.4 (1 C, C-6), 138.9 (1 C, C-3a), 148.5 (1 C, C-7a), 169.0 ppm (1 C, C=O). FTIR (neat): ν $[cm^{-1}] = 3302$ (N–H), 2932 (C–H_{alkyl}), 1643 (C=O), 1543, 1443 (C=C_{arom}). Purity (HPLC): 97.6 %, t_R = 14.6 min.

cis-2-Chloro-*N*-(3-methoxy-3*H*-spiro[[2] benzopyran-1,1'-cyclohexan]-4'-yl)acetamide (*cis*-20)



2-Chloroacetyl chloride (20 µL, 0.25 mmol, 1.2 equiv) was slowly added to a solution of amine *cis*-**13** (50 mg, 0.21 mmol) and Et₃N (0.07 mL, 0.50 mmol, 2.4 equiv) in CH₂Cl₂ (8 mL) under N₂ at 0 °C. After stirring for 6 h at RT, H₂O (30 mL) was added and the aqueous layer was extracted with CH₂Cl₂ (3×30 mL). The combined organic layers were dried (Na₂SO₄), filtered, concentrated in vacuo and the residue was purified by fc (d=2 cm, I=18 cm, V=10 mL, cyclohexane/ethyl acetate 33:67). Colorless solid, m.p. 165 °C, yield 51 mg (78%). C₁₆H₂₀ClNO₃ (309.8 g/mol). $R_{\rm f}=0.48$ (cyclohexane/ethyl acetate 50:50). HRMS (ESI): m/z 332.1014 (calcd. 332.1024 for



C₁₆H₂₀³⁵CINO₃Na [*M*Na⁺]). ¹H NMR (600 MHz, CD₃OD): δ = 1.69 (dq, *J* = 13.6/3.0 Hz, 1H, 2'-H_{equ}), 1.82–1.94 (m, 6H, 3'-H, 5'-H, 6'-H), 2.00 (td, *J* = 13.5/4.1 Hz, 1H, 2'-H_{ax}), 3.49 (s, 3H, OCH₃), 3.87 (tt, *J* = 11.1/ 3.9 Hz, 1H, 4'-H_{ax}), 4.03 (s, 2H, COCH₂Cl), 6.07 (s, 1H, 3-H), 7.26 (d, *J* = 7.5 Hz, 1H, 7-H), 7.33–7.38 (m, 2H, 4-H, 5-H), 7.39–7.42 ppm (m, 1H, 6-H). A signal for the NH proton is not observed in the spectrum. ¹³C NMR (151 MHz, CD₃OD): δ = 29.3 (1 C, C-3'), 29.7 (1 C, C-5'), 37.5 (1 C, C-2'), 38.7 (1 C, C-6'), 43.3 (1 C, COCH₂Cl), 49.5 (1 C, C-4'), 54.9 (1 C, OCH₃), 86.8 (1 C, C-1), 107.2 (1 C, C-3), 121.8 (1 C, C-7), 124.2 (1 C, C-4), 129.0 (1 C, C-5), 130.5 (1 C, C-6), 138.6 (1 C, C-3a), 148.3 (1 C, C-7a), 168.6 ppm (1 C, C=O). FTIR (neat): ν [cm⁻¹]=3271 (N–H), 2978, 2940, 2866 (C–H_{alkyl}), 1651 (C=O), 1555, 1462, 1431 (C=C_{arom}). Purity (HPLC): 98.1%, t_R=15.1 min.

1-(6,7-Dimethoxy-3,4-dihydroisoquinolin-2(1*H*)-yl)-2-{3-methoxy-3,4-dihydrospiro[[2]benzopyran-1,4'-piperidin]-1'-yl} ethan-1-one (21)



A solution of piperidine 10 (43 mg, 0.18 mmol), chloroacetamide 15 (58 mg, 0.22 mmol, 1.2 equiv), Et₃N (0.09 mL, 0.65 mmol, 3.6 equiv) and TBAI (9 mg, 0.02 mmol, 0.1 equiv) in DMF (4 mL) was stirred at RT for 18 h. H₂O (70 mL) was added and the aqueous layer was extracted with CH_2Cl_2 (3×60 mL). The combined organic layers were dried (Na₂SO₄), filtered, concentrated in vacuo and the residue was purified twice by fc (d=2 cm, l=25 cm, V=10 mL, $CH_2CI_2/$ CH₃OH 99:1+1% N,N-dimethylethanamine; d = 1 cm, l = 20 cm, V =3 mL, cyclohexane/ethyl acetate 67:33+1% *N*,*N*-dimethylethanamine \rightarrow 1:1+1% *N*,*N*-dimethylethanamine). Pale yellow oil, yield 31 mg (22%). $C_{27}H_{34}N_2O_5$ (466.6 g/mol). $R_f = 0.46$ (CH₂Cl₂/ CH₃OH 99:1+1% N,N-dimethylethanamine). HRMS (APCI): m/z 467.2531 (calcd. 467.2540 for $C_{27}H_{35}N_2O_5$ [*M*H⁺]). ¹H NMR (400 MHz, CD₃OD): $\delta = 1.70$ (dq, J = 13.5/2.8 Hz, 0.5H, 3'- H_{equ}), 1.75–1.85 (m, 1H, 3'-H, 5'-H), 1.95 (dq, J=13.9/2.6 Hz, 0.5H, 5'-H_{equ}), 2.00-2.09 (m, 1.5H, 3'-H, 5'-H), 2.29 (td, J=13.1/4.5 Hz, 0.5H, 3'-H_{ax}), 2.57-2.73 (m, 2H, 2'-H, 6'-H), 2.73-3.00 (m, 6H, 4-H, 2'-H, 6'-H, 4-H_{isoquinoline}), 3.38-3.52 (m, 2H, COCH₂N), 3.54 (s, 1.5H, 3-OCH₃), 3.57 (s, 1.5H, 3-OCH₃), 3.81-3.90 (m, 8H, 3-H_{isoquinoline}, 6-OCH₃, 7-OCH₃), 4.66 (s, 1H, 1- $H_{isoquinoline}$), 4.80 (s, 1H, 1- $H_{isoquinoline}$), 4.90 (dd, J=7.2/3.2 Hz, 0.5H, 3-H), 4.95 (dd, J=7.2/3.2 Hz, 0.5H, 3-H), 6.76-6.83 (m, 1.5H, 5-Hisoquinoline, 8-Hisoquinoline), 6.86 (s, 0.5H, 8-Hisoquinoline), 7.01-7.05 (m, 0.5H, 8-H), 7.06-7.14 (m, 1H, 5-H) 7.14-7.26 ppm (m, 2.5H, 6-H, 7-H, 8-H). ¹³C NMR (101 MHz, CD₃OD): $\delta = 28.6$ (0.5 C, C-4_{isoquinoline}), 29.7 (0.5 C, C-4_{isoquinoline}), 35.86 (0.5 C, C-4), 35.89 (0.5 C, C-4), 37.18 (0.5 C, C-5'), 37.20 (0.5 C, C-5'), 39.6 (0.5 C, C-3'), 39.7 (0.5 C, C-3'), 41.7 (0.5 C, C-3_{isoquinoline}), 44.4 (0.5 C, C-3_{isoquinoline}), 44.9 (0.5 C, C-1_{isoquinoline}), 48.2 (0.5 C, C-1_{isoquinoline}), 50.2–50.7 (m, 2 C, C-2', C-6'), 56.2–56.4 (m, 3 C, 3-OCH₃, 6-OCH₃, 7-OCH₃), 61.68 (0.5 C, COCH₂N), 61.74 (0.5 C, COCH₂N), 75.2 (0.5 C, C-1), 75.4 (0.5 C, C-1), 97.55 (0.5 C, C-3), 97.59 (0.5 C, C-3), 110.6 (0.5 C, C-8_{isoquinoline}), 110.9 (0.5 C, C-8_{isoquinoline}), 112.9 $(0.5 \text{ C}, \text{ C-5}_{isoquinoline}), 113.1 (0.5 \text{ C}, \text{ C-5}_{isoquinoline}), 125.6 (1 \text{ C}, \text{ C-8}), 126.1$ (1 C, C-8a_{isoquinoline}), 126.9 (1 C, C-4a_{isoquinoline}), 127.37 (0.5 C, C-7), 127.39 (0.5 C, C-7), 127.56 (0.5 C, C-6), 127.62 (0.5 C, C-6), 129.9 (0.5 C, C-5), 130.0 (0.5 C, C-5), 132.4 (0.5 C, C-4a), 132.5 (0.5 C, C-4a), 141.89 (0.5 C, C-8a), 141.92 (0.5 C, C-8a), 149.0-149.3 (m, 2 C, C-6_{isoquinoline}, C-7_{isoquinoline}), 170.7 (0.5 C, C=O), 170.8 ppm (0.5 C, C=O). FTIR (neat): v [cm⁻¹]=2978, 2835 (C–H_{alkyl}), 1624 (C=O), 1516, 1454 (C= C_{arom}). Purity (HPLC): 92.1%, $t_{R} = 17.4$ min.

cis-1-(6,7-Dimethoxy-3,4-dihydroisoquinolin-2(1*H*)-yl)-2-[*N*-(3-methoxy-3,4-dihydrospiro[[2]benzopyran-1,1'-cyclohexan]-4'-yl) amino]ethan-1-one (*cis*-22)



A solution of chloroacetamide 15 (32 mg, 0.12 mmol), amine cis-11 (36 mg, 0.15 mmol, 1.2 equiv), Et₃N (0.06 mL, 0.43 mmol, 2.9 equiv) and TBAI (5 mg, 0.01 mmol, 0.1 equiv) in DMF (4 mL) was stirred at RT for 7 d. H₂O (80 mL) was added and the aqueous layer was extracted with CH_2CI_2 (3×60 mL). The combined organic layers were dried (Na₂SO₄), filtered, concentrated in vacuo and the residue was purified by fc (d=2 cm, l=29 cm, V=10 mL, CH₂Cl₂/CH₃OH 99:1+1% N,N-dimethylethanamine). Pale yellow oil, yield 35 mg (60%). $C_{28}H_{36}N_2O_5$ (480.6 g/mol). $R_f = 0.29$ (CH₂Cl₂/CH₃OH 95:5+1%) N,N-dimethylethanamine). HRMS (ESI): m/z 481.2700 (calcd. 481.2697 for $\rm C_{28}H_{37}N_2O_5$ [MH^+]). $^1\rm H$ NMR (600 MHz, CD_3OD): $\delta\!=\!$ 1.64-1.78 (m, 2H, 2'-H, 3'-H), 1.78-1.92 (m, 4H, 3'-H, 5'-H, 6'-H), 1.92-2.02 (m, 1H, 6'-H), 2.07-2.13 (m, 1H, 2'-H), 2.63-2.72 (m, 1H, 4'-H_{ax}), 2.77–2.82 (m, 2H, 4-H, 4- $H_{isoquinoline}$), 2.87 (t, J=6.0 Hz, 1H, 4-H_{isoquinoline}), 2.89–2.95 (m, 1H, 4-H), 3.56 (s, 1.5H, OCH₃), 3.57 (s, 1.5H, 3-OCH₃), 3.65 (s, 2H, COCH₂NH), 3.70 (t, J=5.9 Hz, 1H, 3-H_{isoquinoline}), 3.80-3.83 (m, 1H, 3-H_{isoquinoline}), 3.82 (s, 6H, 6-OCH₃, 7-OCH₃), 4.61 (s, 1H, 1-H_{isoauinoline}), 4.66 (s, 1H, 1-H_{isoauinoline}), 4.89-4.92 (m, 1H, 3-H), 6.76-6.79 (m, 1.5H, 5-H_{isoquinoline}, 8-H_{isoquinoline}), 6.80 (s, 0.5H, 8-H_{isoquinoline}), 7.06–7.09 (m, 1H, 5-H), 7.12–7.20 ppm (m, 3H, 6-H, 7-H, 8-H). A signal for the NH proton is not observed in the spectrum. ^{13}C NMR (151 MHz, CD₃OD): δ = 28.8 (0.5 C, C-4_{isoquinoline}), 29.1 (1 C, C-3') or C-5'), 29.3 (1 C, C-3' or C-5'), 29.6 (0.5 C, C-4_{isoquinoline}), 36.2 (1 C, C-4), 36.4 (1 C, C-2'), 38.9 (1 C, C-6'), 41.6 (0.5 C, C-3_{isoquinoline}), 43.6 $(0.5 \text{ C}, \text{ C-3}_{isoquinoline}), 45.2 (0.5 \text{ C}, \text{ C-1}_{isoquinoline}), 46.9 (0.5 \text{ C}, \text{ C-1}_{isoquinoline}),$ 48.1 (0.5 C, COCH₂NH), 48.3 (0.5 C, COCH₂NH), 56.4–56.6 (m, 3 C, 3-OCH₃, 6-OCH₃, 7-OCH₃), 57.47 (0.5 C, C-4'), 57.49 (0.5 C, C-4'), 77.37 (0.5 C, C-1), 77.39 (0.5 C, C-1), 97.8 (1 C, C-3), 110.9 (0.5 C, C-8_{isoquinoline}), 111.0 (0.5 C, C-8_{isoquinoline}), 113.0 (0.5 C, C-5_{isoquinoline}), 113.1 (0.5 C, C-5_{isoquinoline}), 125.6 (0.5 C, C-8), 125.7 (0.5 C, C-8), 125.8 (0.5 C, C-8a_{isoquinoline}), 126.3 (0.5 C, C-8a_{isoquinoline}), 127.5 (1 C, C-7), 127.67 (1 C, C-6), 127.74 (0.5 C, C-4a_{isoquinoline}), 128.2 (0.5 C, C-4a_{isoquinoline}), 130.1 (1 C, C-5), 132.6 (1 C, C-4a), 142.61 (0.5 C, C-8a), 142.62 (0.5 C, C-8a), 149.2–149.6 (m, 2 C, C-6 $_{isoquinoline'}$ C-7 $_{isoquinoline}$), 171.4 (0.5 C, C=O), 171.5 ppm (0.5 C, C=O). FTIR (neat): ν [cm $^{-1}$]=3375 (N–H), 2978, 2936, 2909 (C-H_{alkyl}), 1640 (C=O), 1516, 1435 (C=C_{arom}). Purity (HPLC): 98.0 %, $t_{\rm R} = 17.7$ min.

1-(6,7-Dimethoxy-3,4-dihydroisoquinolin-2(1*H*)-yl)-2-{3-methoxy-3H-spiro[[2]benzofuran-1,4'-piperidin]-1'-yl}ethan-1-one (23)



A solution of piperidine **12** (40 mg, 0.18 mmol), chloroacetamide **15** (50 mg, 0.19 mmol, 1.0 equiv), Et₃N (0.09 mL, 0.65 mmol, 3.6 equiv) and TBAI (7 mg, 0.02 mmol, 0.1 equiv) in DMF (5 mL) was stirred at



RT for 19 h. H_2O (70 mL) was added, and the aqueous layer was extracted with CH_2Cl_2 (3×60 mL). The combined organic layers were dried (Na₂SO₄), filtered, and concentrated in vacuo, and the residue was purified twice by fc (d=2 cm, l=25 cm, V=10 mL, CH₂Cl₂/CH₃OH 95:5; d=2 cm, l=20 cm, V=10 mL, cyclohexane/ ethyl acetate 50:50+1% N,N-dimethylethanamine). Pale yellow solid, m.p. 103 °C, yield 30 mg (36 %). $C_{26}H_{32}N_2O_5$ (452.6 g/mol). R_f = 0.30 (CH₂Cl₂/CH₃OH 95:5+1% N,N-dimethylethanamine). HRMS (APCI): *m*/*z* 453.2406 (calcd. 453.2384 for C₂₆H₃₃N₂O₅ [*M*H⁺]). ¹H NMR (600 MHz, CD₃OD): $\delta = 1.52$ (dq, J = 13.5/2.7 Hz, 0.5H, 3'- H_{equ}), 1.61 (dq, J = 13.6/2.8 Hz, 0.5H, 3'- H_{equ}), 1.70 (dq, J = 13.5/2.7 Hz, 0.5H, 5'-H_{eau}), 1.78 (dq, J=13.6/2.8 Hz, 0.5H, 5'-H_{eau}), 1.88-2.00 (m, 1H, 3'- H_{ax} , 5'- H_{ax}), 2.09 (td, J=13.1/4.5 Hz, 0.5H, 5'- H_{ax}), 2.17 (td, J= 13.2/4.5 Hz, 0.5H, 3'- H_{ax}), 2.55–2.66 (m, 2H, 2'-H, 6'-H), 2.81 (t, J =6.1 Hz, 1H, 4-H_{isoquinoline}), 2.84–2.89 (m, 1H, 2'-H or 6'-H), 2.90–2.96 (m, 2H, 2'-H or 6'-H, 4- $H_{isoquinoline}$), 3.40–3.43 (m, 2H, COCH₂N), 3.47 (s, 1.5H, 3-OCH₃), 3.49 (s, 1.5H, 3-OCH₃), 3.79-3.87 (m, 8H, 3-H_{isoquinoline}, 6-OCH₃, 7-OCH₃), 4.65 (s, 1H, 1-H_{isoquinoline}), 4.76 (d, J=15.9 Hz, 0.5H, $1-H_{isoquinoline}$), 4.79 (d, J = 15.9 Hz, 0.5H, $1-H_{isoquinoline}$), 6.04 (s, 0.5H, 3-H), 6.06 (s, 0.5H, 3-H), 6.77 (s, 1H, 5-H_{isoquinoline}, 8-H_{isoquinoline}), 6.79 (s, 0.5H, 5-H_{isoquinoline}), 6.83 (s, 0.5H, 8-H_{isoquinoline}), 7.15 (d, J=7.6 Hz, 0.5H, 4-H), 7.29 (d, J=7.6 Hz, 0.5H, 4-H), 7.32-7.43 ppm (m, 3H, 5-H, 6-H, 7-H). ¹³C NMR (151 MHz, CD₃OD): $\delta = 28.8$ (0.5 C, C-4_{isoquinoline}), 30.0 (0.5 C, C-4_{isoquinoline}), 38.07 (0.5 C, C-3'), 38.14 (0.5 C, C-3'), 39.60 (0.5 C, C-5'), 39.64 (0.5 C, C-5'), 41.8 (0.5 C, C-3_{isoquinoline}), 44.6 (0.5 C, C-3_{isoquinoline}), 45.2 (0.5 C, C-1_{isoquinoline}), 48.3 (0.5 C, C-1_{isoquinoline}), 50.9-51.7 (m, 2 C, C-2', C-6'), 54.98 (0.5 C, 3-OCH₃), 55.01 (0.5 C, 3-OCH₃), 56.5-56.6 (m, 2 C, 6-OCH₃, 7-OCH₃), 61.89 (0.5 C, COCH₂N), 61.92 (0.5 C, COCH₂N), 85.5 (1 C, C-1), 107.1 (0.5 C, C-3), 107.2 (0.5 C, C-3), 110.9 (0.5 C, C-8_{isoquinoline}), 111.0 (0.5 C, C-8_{isoquinoline}), 113.1 (0.5 C, C-5_{isoquinoline}), 113.3 (0.5 C, C-5_{isoquinoline}), 121.7 (0.5 C, C-4), 121.8 (0.5 C, C-4), 124.21 (0.5 C, C-7), 124.25 (0.5 C, C-7), 126.3 (0.5 C, C- $8a_{isoquinoline}), \ 126.9 \ (0.5 \ C, \ C-4a_{isoquinoline}), \ 127.8 \ (0.5 \ C, \ C-8a_{isoquinoline}),$ 128.0 (0.5 C, C-4a_{isoquinoline}), 129.08 (0.5 C, C-6), 129.12 (0.5 C, C-6), 130.49 (0.5 C, C-5), 130.51 (0.5 C, C-5), 138.86 (0.5 C, C-7a), 138.92 (0.5 C, C-7a), 148.01 (0.5 C, C-3a), 148.02 (0.5 C, C-3a), 149.2-149.5 (m, 2 C, C-6_{isoquinoline'} C-7_{isoquinoline}), 170.9 (0.5 C, C=O), 171.0 ppm (0.5 C, C=O). FTIR (neat): ν [cm⁻¹]=2913, 2735 (C-H_{alkvl}), 1636 (C=O), 1516, 1447 (C=C_{arom}). Purity (HPLC): 99.7%, t_R=14.9-17.0 min.

trans-1-(6,7-Dimethoxy-3,4-dihydroisoquinolin-2(1*H*)-yl)-2-[*N*-(3-methoxy-3*H*-spiro[[2]benzofuran-1,1'-cyclohexan]-4'-yl)amino] ethan-1-one (*trans*-24)



A solution of chloroacetamide **15** (30 mg, 0.11 mmol), amine *trans*-**13** (26 mg, 0.11 mmol, 1.0 equiv), Et₃N (0.05 mL, 0.36 mmol, 3.3 equiv.) and TBAI (5 mg, 0.01 mmol, 0.1 equiv) in DMF (4 mL) was stirred at RT for 3 d. H₂O (80 mL) was added and the aqueous layer was extracted with CH₂Cl₂ (3×60 mL). The combined organic layers were dried (Na₂SO₄), filtered, concentrated in vacuo and the residue was purified twice by fc (d=2 cm, I=18 cm, V=10 mL, cyclohexane/ethyl acetate 33:67+1% *N*,*N*-dimethylethanamine \rightarrow 20:80+1% *N*,*N*-dimethylethanamine; d=2 cm, I=31 cm, V=10 mL, CH₂Cl₂/CH₃OH 95:5+1% *N*,*N*-dimethylethanamine). Colorless solid, m.p. 66 °C, yield 28 mg (54%). C₂₇H₃₄N₂O₅ (466.6 g/mol). *R*_f=0.33 (CH₂Cl₂/CH₃OH 95:5+1% *N*,*N*-dimethylethanamine).

HRMS (ESI): *m/z* 467.2533 (calcd. 467.2540 for C₂₇H₃₅N₂O₅ [*M*H⁺]). ¹H NMR (600 MHz, CD₃OD): $\delta = 1.52 - 1.60$ (m, 1H, 2'-H), 1.66-1.74 (m, 1H, 6'-H), 1.77-1.86 (m, 2H, 3'-H, 5'-H), 2.01-2.13 (m, 4H, 2'-H, 3'-H, 5'-H, 6'-H), 2.81 (t, J=6.0 Hz, 1H, 4-H_{isoquinoline}), 2.81 (t, J=6.0 Hz, 1H, 4-H_{isoquinoline}), 2.90–2.94 (m, 1H, 4'-H_{equ}), 3.46 (s, 1.5H, 3-OCH₃), 3.47 (s, 1.5H, 3-OCH₃), 3.65 (s, 2H, COCH₂NH), 3.73 (t, J=5.9 Hz, 1H, 3- $H_{isoquinoline}$), 3.80–3.86 (m, 7H, 3- $H_{isoquinoline}$, 6-OCH₃, 7-OCH₃), 4.67 (s, 1H, 1- $H_{isoquinoline}$), 4.68 (s, 1H, 1- $H_{isoquinoline}$), 6.04 (s, 0.5H, 3-H), 6.05 (s, 0.5H, 3-H), 6.77 (s, 0.5H, 5-H_{isoquinoline}), 6.78 (s, 0.5H, 5-H_{isoquinoline}), 6.79 (s, 0.5H, 8-H_{isoquinoline}), 6.81 (s, 0.5H, 8-H_{isoquinoline}), 7.30-7.41 (m, 3.5H, 4-H, 5-H, 6-H, 7-H), 7.44 ppm (d, J=7.5 Hz, 0.5H, 7-H). A signal for the NH proton is not observed in the spectrum. ¹³C NMR (151 MHz, CD₃OD): δ = 27.9 (0.5 C, C-3' or C-5'), 28.0 (0.5 C, C-3' or C-5'), 28.11 (0.5 C, C-3' or C-5'), 28.14 (0.5 C, C-3' or C-5'), 28.8 (0.5 C, C-4_{isoquinoline}), 29.6 (0.5 C, C-4_{isoquinoline}), 34.0 (0.5 C, C-2'), 34.1 (0.5 C, C-2'), 35.2 (0.5 C, C-6'), 35.3 (0.5 C, C-6'), 41.5 (0.5 C, C-3_{isoquinoline}), 43.7 $(0.5 \text{ C}, \text{ C-3}_{isoquinoline}), 45.2 (0.5 \text{ C}, \text{ C-1}_{isoquinoline}), 46.9 (0.5 \text{ C}, \text{ C-1}_{isoquinoline}),$ 49.7 (1 C, COCH₂NH), 54.2 (0.5 C, C-4'), 54.3 (0.5 C, C-4'), 54.8 (1 C, 3-OCH_3), 56.47 (0.5 C, 6-OCH_3 or 7-OCH_3), 56.50 (0.5 C, 6-OCH_3 or 7-OCH₃), 56.5 (0.5 C, 6-OCH₃ or 7-OCH₃), 56.6 (0.5 C, 6-OCH₃ or 7-OCH₃), 88.1 (1 C, C-1), 106.9 (1 C, C-3), 110.9 (0.5 C, C-8_{isoquinoline}), 111.0 (0.5 C, C-8_{isoquinoline}), 113.0 (0.5 C, C-5_{isoquinoline}), 113.1 (0.5 C, C-5_{isoquinoline}), 122.4 (0.5 C, C-7), 122.5 (0,5 C, C-7), 124.16 (0.5 C, C-6), 124.19 (0.5 C, C-6), 125.8 (0.5 C, C-8a_{isoquinoline}), 126.3 (0.5 C, C-8a_{isoquinoline}), 127.8 (0.5 C, C-4a_{isoquinoline}), 128.2 (0.5 C, C-4a_{isoquinoline}), 128.88 (0.5 C, C-5), 128.90 (0.5 C, C-5), 130.30 (0.5 C, C-4), 130.31 (0.5 C, C-4), 138.7 (0.5 C, C-3a), 138.8 (0.5 C, C-3a), 148.7 (1 C, C-7a), 149.3 (0.5 C, C-6 $_{\rm isoquinoline}$ or C-7 $_{\rm isoquinoline}$), 149.38 (0.5 C, C-6 $_{\rm isoquinoline}$ or C-7_{isoquinoline}), 149.41 (0.5 C, C-6_{isoquinoline} or C-7_{isoquinoline}), 149.5 (0.5 C, C-6_{isoquinoline} or C-7_{isoquinoline}), 171.7 (0.5 C, C=O), 171.8 ppm (0.5 C, C=O). FTIR (neat): ν [cm⁻¹]=3321 (N–H), 2978, 2928 (C–H_{alkyl}), 1643 (C=O), 1516, 1435 (C=C_{arom}). Purity (HPLC): 95.3%, t_R=14.5 min.

cis-1-(6,7-Dimeth-

oxy-3,4-dihydroisoquinolin-2(1*H*)-yl)-2-[*N*-(3-methoxy-3H-spiro[[2]benzofuran-1,1'-cyclohexan]-4'-yl)amino] ethan-1-one (*cis*-24)



A solution of chloroacetamide 15 (47 mg, 0.17 mmol, 1.3 equiv), amine cis-13 (32 mg, 0.14 mmol), Et₃N (0.06 mL, 0.43 mmol, 3.1 equiv.) and TBAI (6 mg, 0.02 mmol, 0.1 equiv) in DMF (4 mL) was stirred at RT for 6 d. H₂O (80 mL) was added and the aqueous layer was extracted with CH_2CI_2 (3×60 mL). The combined organic layers were dried (Na₂SO₄), filtered, concentrated in vacuo and the residue was purified by fc (d=2 cm, l=29 cm, V=10 mL, CH₂Cl₂/CH₃OH 99:1+1% N,N-dimethylethanamine). Yellow oil, yield 28 mg (43%). $C_{27}H_{34}N_2O_5$ (466.6 g/mol). $R_f = 0.23$ (CH₂Cl₂/CH₃OH 95:5+1% N,Ndimethylethanamine). HRMS (ESI): m/z 467.2534 (calcd. 467.2540 for $C_{27}H_{35}N_2O_5$ [*M*H⁺]). ¹H NMR (600 MHz, CD₃OD): $\delta = 1.64-1.70$ (m, 1H, 2'-H), 1.72-1.83 (m, 3H, 3'-H, 5'-H, 6'-H), 1.83-1.93 (m, 2H, 2'-H, 6'-H), 1.93–2.00 (m, 2H, 3'-H, 5'-H), 2.64–2.72 (m, 1H, 4'- H_{ax}), 2.81 (t, J= 6.2 Hz, 1H, 4-H_{isoquinoline}), 2.87 (t, J=6.0 Hz, 1H, 4-H_{isoquinoline}), 3.49 (s, 1.5H, 3-OCH₃), 3.49 (s, 1.5H, 3-OCH₃), 3.67 (s, 2H, COCH₂N), 3.70 (t, J=6.0 Hz, 1H, 3-H_{isoquinoline}), 3.80-3.85 (m, 7H, 3-H_{isoquinoline}, 6-OCH₃, 7-OCH₃), 4.61 (s, 1H, 1-H_{isoquinoline}), 4.66 (s, 1H, 1-H_{isoquinoline}), 6.04 (s, 0.5H, 3-H), 6.05 (s, 0.5H, 3-H), 6.76-6.79 (m, 1.5H, 5-H_{isoquinoline}, 8-H_{isoquinoline}), 6.80 (s, 0.5H, 8-H_{isoquinoline}), 7.18-7.25 (m, 1H, 7-H), 7.31-7.40 ppm (m,



3H, 4-*H*, 5-*H*, 6-*H*). A signal for the NH proton is not observed in the spectrum. ¹³C NMR (151 MHz, CD₃OD): δ = 28.8 (0.5 C, C-4_{isoquinoline}), 29.6 (0.5 C, C-4_{isoquinoline}), 29.7 (1 C, C-3' or C-5'), 30.0 (1 C, C-3' or C-5'), 37.3 (1 C, C-2'), 38.7 (1 C, C-6'), 41.6 (0.5 C, C-3_{isoquinoline}), 43.6 (0.5 C, C-3_{isoquinoline}), 45.2 (0.5 C, C-1_{isoquinoline}), 46.8 (0.5 C, C-1_{isoquinoline}), 48.0 (0.5 C, COCH₂N), 48.2 (0.5 C, COCH₂N), 55.0 (1 C, 3-OCH₃), 56.5–56.6 (m, 2 C, 6-OCH₃, 7-OCH₃), 57.2 (1 C, C-4'), 87.30 (0.5 C, C-1), 87.32 (0.5 C, C-1), 107.1 (1 C, C-3), 110.9 (0.5 C, C-8_{isoquinoline}), 111.0 (0.5 C, C-8_{isoquinoline}), 113.0 (0.5 C, C-5_{isoquinoline}), 113.1 (0.5 C, C-5_{isoquinoline}), 121.68 (0.5 C, C-7), 121.70 (0.5 C, C-7), 124.2 (1 C, C-4), 125.7 (0.5 C, C-8a_{isoquinoline}), 126.3 (0.5 C, C-8a_{isoquinoline}), 127.7 (0.5 C, C-4a_{isoquinoline}), 128.2 (0.5 C, C-4a_{isoquinoline}), 129.0 (1 C, C-5), 130.4 (1 C, C-6), 138.8 (1 C, C-3a), 148.5 (1 C, C-7a), 149.2–149.6 (m, 2 C, C-6_{isoquinoline}), 171.2 (0.5 C, C = O), 171.3 ppm (0.5 C, C = O). FTIR (neat): ν [cm⁻¹] = 3402 (N–H), 2928, 2855 (C–H_{alkyl}), 1643 (C=O), 1516, 1435 (C=C_{arom}). Purity (HPLC): 98.1%, t_R = 15.1 min.

2-(6,7-Dimethoxy-3,4-dihydroisoquinolin-2(1*H*)-yl)-1-(3-methoxy-3,4-dihydrospiro[[2]benzopyran-1,4'-piperidin]-1'-yl) ethan-1-one (25)



2-Chloroacetyl chloride (19 µL, 0.24 mmol, 1.2 equiv) was slowly added to a solution of piperidine **10** (46 mg, 0.20 mmol) and Et₃N (0.07 mL, 0.50 mmol, 2.5 equiv) in CH₂Cl₂ (4 mL) under N₂ at 0 °C. After 5 h of stirring at RT, H₂O (10 mL) was added, and the aqueous layer was extracted with CH₂Cl₂ (3×10 mL). The combined organic layers were dried (Na₂SO₄), filtered, and concentrated in vacuo, and the residue was purified by fc (d=2 cm, I=18 cm, V=10 mL, cyclohexane/ethyl acetate 67:33 \rightarrow 50:50). Chloroacetamide **16**: Pale yellow oil, yield 25 mg (40%). C₁₆H₂₀ClNO₃ (309.8 g/mol).

A solution of chloroacetamide 16 (25 mg, 0.08 mmol), isoquinoline 14·HCl (20 mg, 0.09 mmol, 1.1 equiv), Et₃N (0.03 mL, 0.22 mmol, 2.8 equiv) and TBAI (5 mg, 0.01 mmol, 0.1 equiv) in DMF (4 mL) was stirred at RT for 63 h. H₂O (80 mL) was added and the aqueous layer was extracted with CH_2CI_2 (3×60 mL). The combined organic layers were dried (Na₂SO₄), filtered, concentrated in vacuo and the residue was purified by fc (d=2 cm, l=20 cm, V=10 mL, cyclohexane/ethyl acetate 50:50+1% N,N-dimethylethanamine). Pale yellow solid, m.p. 165 °C, yield 30 mg (75%). $C_{27}H_{34}N_2O_5$ (466.6 g/mol). $R_f = 0.26$ (cyclohexane/ethyl acetate 50:50+1% *N*,*N*-dimethylethanamine). HRMS (APCI): *m/z* 467.2521 (calcd. 467.2540 for C₂₇H₃₅N₂O₅ [*M*H⁺]). ¹H NMR (600 MHz, CD₃OD): δ = 1.76–1.86 (m, 1.5H, 3'-H, 5'-H), 1.90– 1.96 (m, 0.5H, 5'-H), 1.99–2.10 (m, 1.5H, 3'-H, 5'-H), 2.17 (td, J=13.3/ 4.6 Hz, 0.5H, 5'-H), 2.79–2.93 (m, 5H, 4-H, 3-H_{isoauinoline}, 4-H_{isoauinoline}), 2.93-2.98 (m, 1H, 4-H), 3.13-3.23 (m, 1H, 2'-H), 3.37 (d, J=14.0 Hz, 1H, COCH₂N), 3.54 (s, 1.5H, 3-OCH₃), 3.55 (s, 1.5H, 3-OCH₃), 3.56-3.66 (m, 3H, 6'-H, COCH₂N, 1-H_{isoquinoline}), 3.67–3.72 (m, 1H, 1-H_{isoquinoline}), 3.79-3.82 (m, 6H, 6-OCH₃, 7-OCH₃), 4.09-4.14 (m, 1H, 6'-H), 4.50-4.57 (m, 1H, 2'-H), 4.96 (dd, J=6.9/3.2 Hz, 0.5H, 3-H), 4.98 (dd, J= 7.0/3.2 Hz, 0.5H, 3-H), 6.69 (s, 0.5H, 8-H_{isoquinoline}), 6.69 (s, 0.5H, 8-H_{isoquinoline}), 6.70 (s, 0.5H, 5-H_{isoquinoline}), 6.72 (s, 0.5H, 5-H_{isoquinoline}), 6.94-6.97 (m, 0.5H, 8-H), 7.00 (dd, J=7.5/1.6 Hz, 0.5H, 8-H), 7.08-7.17 ppm (m, 3H, 5-H, 6-H, 7-H). 13 C NMR (151 MHz, CD₃OD): $\delta =$ 29.5 (0.5 C, C-4_{isoquinoline}), 29.6 (0.5 C, C-4_{isoquinoline}), 36.0 (1 C, C-4), 37.6 (0.5 C, C-3'), 38.2 (0.5 C, C-5'), 39.6 (0.5 C, C-2'), 39.7 (0.5 C, C-2'), 39.8 (0.5 C, C-3'), 40.4 (0.5 C, C-5'), 43.48 (0.5 C, C-6'), 43.50 (0.5 C, C-6'), 52.2 (0.5 C, C-3 $_{\rm isoquinoline}$), 52.3 (0.5 C, C-3 $_{\rm isoquinoline}$), 56.4–56.6 (m, 4 C, C-1_{isoquinoline}, 3-OCH₃, 6-OCH₃, 7-OCH₃), 61.28 (0.5 C, COCH₂N), 61.30 (0.5 C, COCH₂N), 76.08 (0.5 C, C-1), 76.11 (0.5 C, C-1), 98.15 (0.5 C, C-3), 98.22 (0.5 C, C-3), 111.1 (1 C, C-8_{isoquinoline}), 113.1 (0.5 C, C-5_{isoquinoline}), 125.7 (1 C, C-8), 127.40 (1 C, C-4a_{sioquinoline}), 127.44 (1 C, C-4a_{sioquinoline}), 127.6 (0.5 C, C-7), 127.7 (0.5 C, C-7), 128.0 (1 C, C-6), 130.2 (1 C, C-5), 132.6 (1 C, C-4a), 141.32 (0.5 C, C-8a), 141.34 (0.5 C, C-8a), 148.8 (1 C, C-6_{sioquinoline}), 147.24 (1 C, C-6_{sioquinoline}), 127.44 (0.5 C, C-8a), 148.8 (1 C, C-6_{sioquinoline}), 149.2 (1 C, C-6_{sioquinoline}), 127.44 (0.5 C, C-8a), 148.8 (1 C, C-6_{sioquinoline}), 149.2 (1 C, C-6_{sioquinoline}), 127.44 (0.5 C, C-8a), 148.8 (1 C, C-6_{sioquinoline}), 1639 (C=O), 1516, 1443 (C=C_{arom}). Purity (HPLC): 99.4%, $t_{\rm R} = 17.4$ min.

trans-2-(6,7-Dimeth-

oxy-3,4-dihydroisoquinolin-2(1*H*)-yl)-*N*-(3-methoxy-3,4-dihydrospiro[[2]benzopyran-1,1'-cyclohexan]-4'-yl) acetamide (*trans*-26)



A solution of chloroacetamide trans-17 (29 mg, 0.09 mmol), isoquinoline 14·HCl (23 mg, 0.10 mmol, 1.1 equiv), Et₃N (0.04 mL, 0.29 mmol, 3.2 equiv) and TBAI (3 mg, 0.01 mmol, 0.1 equiv) in DMF (4 mL) was stirred at RT for 68 h. H₂O (80 mL) was added and the aqueous layer was extracted with CH_2CI_2 (3×60 mL). The combined organic layers were dried (Na₂SO₄), filtered, concentrated in vacuo and the residue was purified twice by fc (d=2 cm, l=18 cm, V=cyclohexane/ethyl acetate 50:50+1% N,N-dimeth-10 mL, ylethanamine; d = 2 cm, l = 20 cm, V = 10 mL, cyclohexane/ethyl acetate 50:50+1% N,N-dimethylethanamine). Colorless oil, yield 36 mg (83%). C₂₈H₃₆N₂O₅ (480.6 g/mol). R_f=0.11 (cyclohexane/ethyl acetate 50:50+1% *N.N*-dimethylethanamine). HRMS (ESI): m/z481.2695 (calcd. 481.2697 for C₂₈H₃₇N₂O₅ [*M*H⁺]). ¹H NMR (600 MHz, CD₃OD): δ = 1.66−1.72 (m, 2H, 2'-H, 6'-H), 1.73−1.78 (m, 2H, 3'-H, 5'-H), 1.90-1.95 (m, 1H, 6'-H), 1.96-2.00 (m, 1H, 2'-H), 2.10-2.17 (m, 1H, 5'-H), 2.17-2.24 (m, 1H, 3'-H), 2.77 (dd, J=15.7/7.3 Hz, 1H, 4-H), 2.90 (dd, J = 15.7/3.1 Hz, 1H, 4-H), 2.92–2.95 (m, 2H, 3- $H_{isoquinoline}$), 2.98 (t, J=5.6 Hz, 2H, 4-H_{isoquinoline}), 3.30 (s, 2H, COCH₂N), 3.53 (s, 3H, 3- OCH_3), 3.72 (s, 5H, 1- $H_{isoquinoline'}$ 7- OCH_3), 3.85 (s, 3H, 6- OCH_3 ,), 4.19 (quint, J = 3.3 Hz, 1H, 4'- \dot{H}_{equ}), 4.88 (dd, J = 7.4/3.1 Hz, 1H, 3-H), 6.68 (s, 1H, 8-H_{isoquinoline}), 6.71 (dd, J=7.9/1.2 Hz, 1H, 8-H), 6.80 (s, 1H, 5-H_{isoquinoline}), 6.89–6.92 (m, 1H, 7-H), 7.05 (dd, J=7.6/1.3 Hz, 1H, 5-H), 7.10 ppm (td, J=7.4/1.2 Hz, 1H, 6-H). A signal for the NH proton is not observed in the spectrum. ¹³C NMR (151 MHz, CD₃OD): δ = 26.5 (1 C, C-3'), 26.6 (1 C, C-5'), 30.1 (1 C, C-4_{isoquinoline}), 32.2 (1 C, C-6'), 34.6 (1 C, C-2'), 36.1 (1 C, C-4), 44.6 (1 C, C-4'), 52.7 (1 C, C-3_{isoquinoline}), 56.35 (1 C, 3-OCH₃), 56.44 (1 C, 7-OCH₃), 56.5 (1 C, 6-OCH₃), 56.8 (1 C, C-1_{isoquinoline}), 62.3 (1 C, COCH₂N), 77.1 (1 C, C-1), 97.9 (1 C, C-3), 111.1 (1 C, C-8_{isoquinoline}), 113.2 (1 C, C-5_{isoquinoline}), 125.4 (1 C, C-8), 127.1 (1 C, C-4a_{isoquinoline}), 127.4 (1 C, C-8a_{isoquinoline}), 127.5 (1 C, C-7), 127.7 (1 C, C-6), 130.1 (1 C, C-5), 132.4 (1 C, C-4a), 142.6 (1 C, C-8a), 148.9 (1 C, C-7_{isoquinoline}), 149.4 (1 C, C-6_{isoquinoline}), 172.1 ppm (1 C, C=O). FTIR (neat): ν [cm⁻¹]=3341 (N–H), 2928, 2832 (C–H_{alkvl}), 1674 (C=O), 1516, 1447 (C= C_{arom}). Purity (HPLC): 90.6 %, $t_{B} = 17.0$ min.



cis-2-(6,7-Dimethoxy-3,4-dihydroisoquinolin-2(1*H*)-yl)-*N*-(3-methoxy-3,4-dihydrospiro[[2]benzopyran-1,1'-cyclohexan]-4'-yl) acetamide (*cis*-26)



A solution of chloroacetamide cis-17 (34 mg, 0.10 mmol), isoquinoline 14·HCl (26 mg, 0.11 mmol, 1.1 equiv), Et₃N (0.04 mL, 0.29 mmol, 2.9 equiv) and TBAI (4 mg, 0.01 mmol, 0.1 equiv) in DMF (4 mL) was stirred at RT for 66 h. H₂O (80 mL) was added and the aqueous layer was extracted with CH_2CI_2 (3×60 mL). The combined organic layers were dried (Na₂SO₄), filtered, concentrated in vacuo and the residue was purified by fc (d = 2 cm, l = 20 cm, V = 10 mL, cyclohexane/ethyl acetate 50:50+1% N,N-dimethylethanamine). Colorless solid, m.p. 176 °C, yield 31 mg (62%). $C_{28}H_{36}N_2O_5$ (480.6 g/mol). $R_f = 0.08$ (cyclohexane/ethyl acetate 50:50+1% N,N-dimethylethanamine). HRMS (APCI): m/z 481.2720 (calcd. 481.2697 for $C_{28}H_{37}N_2O_5$ [MH^+]). ¹H NMR (600 MHz, CD₃OD): δ = 1.77–1.95 (m, 6H, 2'-H, 3'-H, 5'-H, 6'-H), 2.05–2.14 (m, 2H, 2'-H, 6'-H), 2.77–2.84 (m, 3H, 4-H, 3-H_{isoquinoline}), 2.86-2.90 (m, 2H, 4-H_{isoquinoline}), 2.93 (dd, J=15.7/3.1 Hz, 1H, 4-H), 3.22 (s, 2H, COCH₂Cl), 3.54 (s, 3H, 3-OCH₃), 3.67 (s, 2H, 1-H_{isoquinoline}), 3.80 (s, 3H, 7-OCH₃), 3.81 (s, 3H, 6-OCH₃), 3.88-3.95 (m, 1H, 4'-H_{ax}), 4.91 (dd, J=7.4/3.1 Hz, 1H, 3-H), 6.65 (s, 1H, 8-H_{isoquinoline}), 6.72 (s, 1H, 5-H_{isoquinoline}), 7.09 (d, J=7.4 Hz, 1H, 5-H), 7.16 (td, J=7.2/1.8 Hz, 1H, 6-H), 7.18–7.24 ppm (m, 2H, 7-H, 8-H). A signal for the NH proton is not observed in the spectrum. ¹³C NMR (151 MHz, CD₃OD): $\delta = 29.0$ (1 C, C-3' or C-5'), 29.2 (1 C, C-3' or C-5'), 29.4 (1 C, C-4_{isoquinoline}), 36.1 (1 C, C-4), 36.6 (1 C, C-2'), 39.1 (1 C, C-6'), 49.0 (1 C, C-4'), 52.3 (1 C, C-3_{isoquinoline}), 56.4 (2 C, C-1_{isoquinoline}, 3-OCH₃), 56.47 (1 C, 6-OCH₃), 56.51 (1 C, 7-OCH₃), 62.0 (1 C, COCH₂N), 76.9 (1 C, C-1), 97.9 (1 C, C-3), 111.1 (1 C, C-8_{isoquinoline}), 113.1 (1 C, C-5_{isoquinoline}), 125.7 (1 C, C-8), 127.2 (1 C, C-4a_{isoquinoline}), 127.5 (1 C, C-8a_{isoquinoline}), 127.6 (1 C, C-7), 127.7 (1 C, C-6), 130.1 (1 C, C-5), 132.6 (1 C, C-4a), 142.5 (1 C, C-8a), 148.9 (1 C, C-7_{isoquinoline}), 149.2 (1 C, C-6_{isoquinoline}), 171.9 ppm (1 C, C= O). FTIR (neat): ν [cm⁻¹]=3298 (N–H), 2978, 2924, 2835 (C–H_{alkyl}), 1639 (C=O), 1512, 1443 (C=C_{arom}). Purity (HPLC): 98.1%, $t_R = 17.5$ min.

cis-4-(6,7-Dimeth-

oxy-3,4-dihydroisoquinolin-2(1*H*)-yl)-*N*-(3-methoxy-3,4-dihydrospiro[[2]benzopyran-1,1'-cyclohexan]-4'-yl) butanamide (*cis*-27)



A solution of isoquinoline **14**·HCl (27 mg, 0.12 mmol), chlorobutyramide *cis*-**18** (45 mg, 0.13 mmol, 1.1 equiv), Et₃N (0.05 mL, 0.36 mmol, 3.0 equiv) and TBAl (5 mg, 0.01 mmol, 0.1 equiv) in DMF (4 mL) was stirred at RT for 6 d. H₂O (80 mL) was added and the aqueous layer was extracted with CH₂Cl₂ (3×60 mL). The combined organic layers were dried (Na₂SO₄), filtered, concentrated in vacuo and the residue was purified twice by fc (d=2 cm, l=32 cm, V=10 mL, CH_2CI_2/CH_3OH 99:1+1% *N*,*N*-dimethylethanamine; d =1 cm, I = 25 cm, V = 3 mL, CH₂Cl₂/CH₃OH 95:5). Pale yellow oil, yield 11 mg (19%). $C_{30}H_{40}N_2O_5$ (508.7 g/mol). $R_f = 0.19$ (CH₂Cl₂/CH₃OH 95:5). HRMS (ESI): *m/z* 509.3024 (calcd. 509.3010 for C₃₀H₄₁N₂O₅ $[MH^+]$). ¹H NMR (400 MHz, CD₃OD): $\delta = 1.74-1.91$ (m, 6H, 2'-H, 3'-H, 5'-H, 6'-H), 1.97 (quint, J=7.5 Hz, 2H, COCH₂CH₂CH₂N), 2.02-2.09 (m, 1H, 6'-H), 2.09–2.15 (m, 1H, 2'-H), 2.31 (t, J=7.3 Hz, 2H, COCH₂CH₂CH₂N), 2.65 (t, J=7.8 Hz, 2H, COCH₂CH₂CH₂N), 2.78-2.88 (m, 3H, 4-H, 3-H_{isoquinoline}), 2.88–2.92 (m, 2H, 4-H_{isoquinoline}), 2.95 (dd, J= 15.7/3.2 Hz, 1H, 4-H), 3.58 (s, 3H, 3-OCH₃), 3.68 (s, 2H, 1-H_{isoquinoline}), 3.79-3.90 (m, 1H, 4'-H_{ax}), 3.817 (s, 3H, 6-OCH₃ or 7-OCH₃), 3.818 (s, 3H, 6-OCH₃ or 7-OCH₃), 4.93 (dd, J=7.5/3.2 Hz, 1H, 3-H), 6.69 (s, 1H, 8-H_{isoquinoline}), 6.73 (s, 1H, 5-H_{isoquinoline}), 7.08-7.13 (m, 1H, 5-H), 7.15-7.20 (m, 1H, 6-H), 7.20-7.24 ppm (m, 2H, 7-H, 8-H). A signal for the NH proton is not observed in the spectrum. $^{13}\mbox{C}$ NMR (101 MHz, CD₃OD): $\delta = 23.8$ (1 C, COCH₂CH₂CH₂N), 28.8 (1 C, C-4_{isoquinoline}), 29.0 (1 C, C-3' or C-5'), 29.1 (1 C, C-3' or C-5'), 35.0 (1 C, COCH₂CH₂CH₂N), 36.2 (1 C, C-4), 36.6 (1 C, C-2'), 39.1 (1 C, C-6'), 49.1 (1 C, C-4'), 52.0 (1 C, C-3_{isoquinoline}), 56.4 (1 C, C-1_{isoquinoline}), 56.4 (1 C, 3-OCH₃), 56.47 (1 C, 6-OCH₃ or 7-OCH₃), 56.53 47 (1 C, 6-OCH₃ or 7-OCH₃), 58.4 (1 C, COCH₂CH₂CH₂N), 77.0 (1 C, C-1), 97.9 (1 C, C-3), 111.2 (1 C, C-8_{isoquinoline}), 113.0 (1 C, C-5_{isoquinoline}), 125.7 (1 C, C-8), 126.9 (1 C, C-8a_{isoquinoline}), 127.1 (1 C, C-4a_{isoquinoline}), 127.6 (1 C, C-7), 127.7 (1 C, C-6), 130.1 (1 C, C-5), 132.6 (1 C, C-4a), 142.5 (1 C, C-8a), 149.0 (1 C, C-7_{isoquinoline}), 149.4 (1 C, C-6_{isoquinoline}), 174.8 ppm (1 C, C=O). FTIR (neat): ν [cm⁻¹]=3275 (N–H), 2924, 2855 (C–H_{alkyl}), 1639 (C=O), 1516, 1447 (C=C_{arom}). Purity (HPLC): 93.5 %, t_R = 17.9 min.

2-(6,7-Dimethoxy-3,4-dihydroisoquinolin-2(1*H*)-yl)-1-{3-methoxy-3*H*-spiro[[2]benzofuran-1,4'-piperidin]-1'-yl}ethan-1-one (28)



2-Chloroacetyl chloride (36 μ L, 0.45 mmol, 1.2 equiv) was slowly added to a solution of piperidine **12** (83 mg, 0.38 mmol) and Et₃N (0.12 mL, 0.87 mmol, 2.3 equiv) in CH₂Cl₂ (10 mL) under N₂ at 0 °C. After stirring for 6 h at RT, H₂O (10 mL) was added and the aqueous layer was extracted with CH₂Cl₂ (3×10 mL). The combined organic layers were dried (Na₂SO₄), filtered, concentrated in vacuo and the residue was purified by fc (d=2 cm, I=19 cm, V=10 mL, cyclohexane/ethyl acetate 67:33). Chloroacetamide **19**: Pale yellow oil, yield 50 mg (44%). C₁₅H₁₈CINO₃ (295.8 g/mol).

A solution of chloroacetamide **19** (50 mg, 0.17 mmol, 1.2 equiv), isoquinoline **14**·HCl (33 mg, 0.14 mmol), Et₃N (0.07 mL, 0.50 mmol, 3.6 equiv) and TBAI (6 mg, 0.02 mmol, 0.1 equiv) in DMF (6 mL) was stirred at RT for 5 d. H₂O (70 mL) was added and the aqueous layer was extracted with CH₂Cl₂ (4×30 mL). The combined organic layers were dried (Na₂SO₄), filtered, concentrated in vacuo and the residue was purified twice by fc (d=2 cm, I=25 cm, V=10 mL, CH₂Cl₂/CH₃OH 95:5; d=2 cm, I=20 cm, V=10 mL, cyclohexane/ethyl acetate 50:50 + 1% *N*,*N*-dimethylethanamine). Pale yellow oil, yield 31 mg (49%). C₂₆H₃₂N₂O₅ (452.6 g/mol). *R*_f = 0.16 (cyclohexane/ethyl acetate 50:50 + 1% *N*,*N*-dimethylethanamine). HRMS (APCI): *m*/z 453.2403 (calcd. 453.2384 for C₂₆H₃₃N₂O₅ [*M*H⁺]). ¹H NMR (600 MHz, CD₃OD): δ = 1.59–1.67 (m, 1H, 3'-*H*, 5'-*H*), 1.77–1.84 (m, 1H, 3'-*H*, 5'-*H*), 1.88–1.94 (m, 0.5H, 3'-*H*), 2.82–2.85 (m, 2H, 3-*H*_{isoquinoline}), 2.86–2.90



(m, 2H, 4-H_{isoquinoline}), 3.13–3.19 (m, 1H, 2'-H), 3.40 (d, J=8.3 Hz, 0.5H, $COCH_2N$), 3.42 (d, J = 8.3 Hz, 0.5H, $COCH_2N$), 3.50 (s, 1.5H, 3- OCH_3), 3.51 (s, 1.5H, 3-OCH₃), 3.53-3.61 (m, 2H, 6'-H, COCH₂N), 3.66 (s, 1H, 1-H_{isoquinoline}), 3.67 (s, 1H, 1-H_{isoquinoline}), 3.79-3.82 (m, 6H, 6-OCH₃, 7-OCH3), 4.16-4.21 (m, 1H, 6'-H), 4.56-4.62 (m, 1H, 2'-H), 6.09 (s, 0.5H, 3-H), 6.10 (s, 0.5H, 3-H), 6.67 (s, 0.5H, 8-H_{isoquinoline}), 6.68 (s, 0.5H, 8-H_{isoquinoline}), 6.71 (s, 0.5H, 5-H_{isoquinoline}), 6.72 (s, 0.5H, 5-H_{isoquinoline}), 7.11-7.13 (m, 0.5H, 4-H), 7.15-7.17 (m, 0.5H, 4-H), 7.34-7.40 ppm (m, 3H, 5-H, 6-H, 7-H). ^{13}C NMR (151 MHz, CD_3OD): $\delta\!=\!29.47$ (0.5 C, C- $4_{\rm isoquinoline}),~29.51$ (0.5 C, C-4 $_{\rm isoquinoline}),~38.0$ (0.5 C, C-3'), 38.7 (0.5 C, C-5'), 39.5 (0.5 C, C-3'), 40.1 (1 C, C-2', C-5'), 40.5 (0.5 C, C-2'), 43.9 (0.5 C, C-6'), 44.2 (0.5 C, C-6'), 52.2 (1 C, C-3_{isoquinoline}), 55.2 (1 C, 3-OCH₃), 56.4–56.6 (m, 3 C, C-1_{isoquinoline}, 6-OCH₃, 7-OCH₃), 61.1 (0.5 C, COCH₂N), 61.2 (0.5 C, COCH₂N), 85.9 (1 C, C-1), 107.4 (1 C, C-3), 111.1 (1 C, C-8_{isoquinoline}), 113.11 (0.5 C, C-5_{isoquinoline}), 113.13 (0.5 C, C-5_{isoquinoline}), 121.75 (0.5 C, C-4), 121.77 (0.5 C, C-4), 124.4 (1 C, C-7), 127.38 (0.5 C, C-4a_{isoquinoline}), 127.40 (0.5 C, C-4a_{isoquinoline}), 127.6 (0.5 C, C-8a_{isoquinoline}), 127.7 (0.5 C, C-8a_{isoquinoline}), 129.3 (1 C, C-6), 130.58 (0.5 C, C-5), 130.60 (0.5 C, C-5), 138.9 (1 C, C-7a), 147.3 (1 C, C-3a), 148.82 (0.5 C, C-7_{isoauinoline}), 148.83 (0.5 C, C-7_{isoauinoline}), 149.16 (0.5 C, C-6_{isoquinoline}), 149.17 (0.5 C, C-6_{isoquinoline}), 170.39 (0.5 C, C=O), 170.40 ppm (0.5 C, C=O). FTIR (neat): ν [cm⁻¹] = 2913, 2735 (C-H_{alkvl}), 1636 (C=O), 1516, 1447 (C=C_{arom}). Purity (HPLC): 98.7%, t_R=14.3-16.7 min.

trans-2-(6,7-Dimeth-

oxy-3,4-dihydroisoquinolin-2(1*H*)-yl)-*N*-(3-methoxy-3*H*-spiro [[2]benzofuran-1,1'-cyclohexan]-4'-yl)acetamide (*trans*-29)



A solution of chloroacetamide trans-20 (40 mg, 0.13 mmol), isoquinoline 14·HCl (33 mg, 0.14 mmol, 1.1 equiv), Et₃N (0.05 mL, 0.36 mmol, 2.8 equiv.) and TBAI (48 mg, 0.13 mmol, 1.0 equiv) in DMF (4 mL) was stirred at RT for 66 h. H₂O (80 mL) was added and the aqueous layer was extracted with CH_2CI_2 (3×60 mL). The combined organic layers were dried (Na2SO4), filtered, concentrated in vacuo and the residue was purified twice by fc (d=2 cm, l=20 cm, V = 10 mL, cyclohexane/ethyl acetate 50:50 + 1% N,Ndimethylethanamine; d=2 cm, l=18 cm, V=10 mL, cyclohexane/ ethyl acetate 50:50+1% N,N-dimethylethanamine). Colorless solid, m.p. 107 °C, yield 40 mg (66%). $C_{27}H_{34}N_2O_5$ (466.6 g/mol). $R_f = 0.11$ (cyclohexane/ethyl acetate 50:50+1% *N*,*N*-dimethylethanamine). HRMS (ESI): *m/z* 467.2549 (calcd. 467.2540 for C₂₇H₃₅N₂O₅ [*M*H⁺]). ¹H NMR (600 MHz, CD₃OD): $\delta = 1.55 - 1.61$ (m, 1H, 2'-H), 1.71-1.77 (m, 1H, 6'-H), 1.79-1.90 (m, 4H, 2'-H, 3'-H, 5'-H, 6'-H), 2.06-2.16 (m, 2H, 3'-H, 5'-H), 2.92 (t, J = 5.9 Hz, 2H, 3- $H_{isoquinoline}$), 2.97 (t, J = 5.9 Hz, 2H, 4-Hisoquinoline), 3.29 (s, 2H, COCH2N), 3.47 (s, 3H, 3-OCH3), 3.70 (s, 2H, 1- $H_{\text{isoquinoline}}$), 3.74 (s, 3H, 7-OCH₃), 3.85 (s, 3H, 6-OCH₃), 4.17 (quint, J= 3.7 Hz, 1H, 4'-H_{equ}), 6.03 (s, 1H, 3-H), 6.68 (s, 1H, 8-H_{isoquinoline}), 6.75 (d, J=7.5 Hz, 1H, 7-H), 6.80 (s, 1H, 5-H_{isoquinoline}), 7.24 (t, J=7.4 Hz, 1H, 6-H), 7.29-7.36 ppm (m, 2H, 4-H, 5-H). A signal for the NH proton is not observed in the spectrum. ¹³C NMR (151 MHz, CD₃OD): $\delta = 27.4$ (1 C, C-3' or C-5'), 27.7 (1 C, C-3' or C-5'), 30.1 (1 C, C-4_{isoquinoline}), 33.8 (1 C, C-2'), 35.1 (1 C, C-6'), 45.1 (1 C, C-4'), 52.6 (1 C, C-3_{isoquinoline}), 55.0 (1 C, 3-OCH₃), 56.4 (1 C, 7-OCH₃), 56.5 (1 C, 6-OCH₃), 56.7 (1 C, C-1_{isoquinoline}), 62.1 (1 C, COCH₂N), 87.0 (1 C, C-1), 107.1 (1 C, C-3), 111.1 (1 C, C-8_{isoquinoline}), 113.1 (1 C, C-5_{isoquinoline}), 121.8 (1 C, C-7), 124.2 (1 C, C-4), 127.2 (1 C, C-4a_{isoquinoline}), 127.5 (1 C, C-8a_{isoquinoline}), 129.0 (1 C, C-

5), 130.4 (1 C, C-6), 138.7 (1 C, C-3a), 148.2 (1 C, C-7a), 149.0 (1 C, C-7_{isoquinoline}), 149.4 (1 C, C-6_{isoquinoline}), 172.1 ppm (1 C, C=O). FTIR (neat): ν [cm⁻¹]=3341 (N–H), 2932, 2832 (C–H_{alkyl}), 1674 (C=O), 1516, 1463, 1451 (C=C_{arom}). Purity (HPLC): 92.4 %, $t_{\rm R}$ =14.0 min.

cis-2-(6,7-Dimeth-

oxy-3,4-dihydroisoquinolin-2(1*H*)-yl)-*N*-(3-methoxy-3*H*-spiro [[2]benzofuran-1,1'-cyclohexan]-4'-yl)acetamide (*cis*-29)



A solution of chloroacetamide cis-20 (35 mg, 0.11 mmol), isoquinoline 14·HCl (30 mg, 0.13 mmol, 1.2 equiv), Et₃N (0.05 mL, 0.36 mmol, 3.3 equiv) and TBAI (4 mg, 0.01 mmol, 0.1 equiv) in DMF (4 mL) was stirred at RT for 6 d. H₂O (80 mL) was added and the aqueous layer was extracted with CH_2CI_2 (3×60 mL). The combined organic layers were dried (Na₂SO₄), filtered, concentrated in vacuo and the residue was purified twice by fc (d=2 cm, l=29 cm, V=10 mL, $CH_2CI_2/$ CH₃OH 99:1+1% N,N-dimethylethanamine; d = 2 cm, l = 31 cm, V =10 mL, CH₂Cl₂/CH₃OH 199:1+1% *N*,*N*-dimethylethanamine). Yellow oil, yield 45 mg (86%). $C_{27}H_{34}N_2O_5$ (466.6 g/mol). $R_f = 0.31$ (CH₂Cl₂/ CH₃OH 95:5+1% N,N-dimethylethanamine). HRMS (ESI): m/z 467.2531 (calcd. 467.2540 for $C_{27}H_{35}N_2O_5$ [MH^+]). $^1\!H$ NMR (400 MHz, CD₃OD): $\delta = 1.70$ (dq, J = 13.3/2.9 Hz, 1H, 2'- H_{eau}), 1.78–1.97 (m, 6H, 3'-H, 5'-H, 6'-H), 2.01 (td, J = 13.4/4.1 Hz, 1H, 2'-H_{ax}), 2.81–2.86 (m, 2H, 3-H_{isoquinoline}), 2.87-2.92 (m, 2H, 4-H_{isoquinoline}), 3.24 (s, 2H, COCH₂N), 3.49 (s, 3H, 3-OCH₃), 3.69 (s, 2H, 1-H_{isoquinoline}), 3.81 (s, 3H, 7-OCH₃), 3.83 (s, 3H, 6-OCH₃), 3.92 (tt, J=11.4/4.1 Hz, 1H, 4'-H_{ax}), 6.07 (s, 1H, 3-H), 6.67 (s, 1H, 8- $H_{isoquinoline}$), 6.74 (s, 1H, 5- $H_{isoquinoline}$), 7.28 (d, J= 7.5 Hz, 1H, 7-H), 7.32-7.45 ppm (m, 3H, 4-H, 5-H, 6-H). A signal for the NH proton is not observed in the spectrum. $^{13}\mathrm{C}$ NMR (101 MHz, CD₃OD): δ = 29.4 (1 C, C-4_{isoquinoline}), 29.6 (1 C, C-3'), 30.0 (1 C, C-5'), 37.5 (1 C, C-2'), 38.8 (1 C, C-6'), 48.8 (1 C, C-4'), 52.3 (1 C, C-3_{isoquinoline}), 54.9 (1 C, 3-OCH₃), 56.4 (1 C, C-1_{isoquinoline}), 56.48 (1 C, 6-OCH₃ or 7-OCH₃), 56.52 (1 C, 6-OCH₃ or 7-OCH₃), 62.0 (1 C, COCH₂N), 86.9 (1 C, C-1), 107.2 (1 C, C-3), 111.1 (1 C, C-8_{isoquinoline}), 113.2 (1 C, C-5_{isoquinoline}), 121.8 (1 C, C-7), 124.2 (1 C, C-4), 127.2 (1 C, C-4a_{isoquinoline}), 127.5 (1 C, C-8a_{isoquinoline}), 129.0 (1 C, C-5), 130.5 (1 C, C-6), 138.6 (1 C, C-3a), 148.3 (1 C, C-7a), 148.9 (1 C, C-7 $_{isoquinoline}$), 149.2 (1 C, C-6 $_{isoquinoline}$), 172.0 ppm (1 C, C=O). FTIR (neat): ν [cm⁻¹]=3333 (N–H), 2932, 2909, 2832 (C–H_{alkyl}), 1667 (C=O), 1516, 1439 (C=C_{arom}). Purity (HPLC): 88.0 %, $t_{\rm R} = 14.4$ min.

1'-[2-(6,7-Dimethoxy-3,4-dihydroisoquinolin-2(1*H*)-yl) ethyl]-3-methoxy-3,4-dihydrospiro[[2] benzopyran-1,4'-piperidine] (30)



LiAlH₄ (7 mg, 0.19 mmol, 6.4 equiv) was added to a solution of amide **25** (16 mg, 0.03 mmol) in THF (3 mL) under N₂. The mixture was heated to reflux for 2 h. After cooling to RT, H₂O (10 mL) was added, the precipitate was filtered off, and the aqueous phase was



extracted with CH_2CI_2 (3×10 mL). The combined organic layers were dried (Na₂SO₄), filtered, concentrated in vacuo and the residue was purified twice by fc (d=1 cm, l=21 cm, V=3 mL, $CH_2CI_2/$ CH₃OH 97:3+1% *N*,*N*-dimethylethanamine; d = 1 cm, l = 21 cm, V =3 mL, CH₂Cl₂/CH₃OH 99:1+1% *N*,*N*-dimethylethanamine). Yellow oil, yield 10 mg (63%). C₂₇H₃₆N₂O₄ (452.6 g/mol). R_f=0.24 (CH₂Cl₂/ CH₃OH 95:5+1% N,N-dimethylethanamine). HRMS (ESI): m/z453.2754 (calcd. 453.2748 for C₂₇H₃₇N₂O₄ [*M*H⁺]). ¹H NMR (600 MHz, CD₃OD): $\delta = 1.82$ (dq, J = 13.8/2.6 Hz, 1H, 3'- H_{equ}), 2.00 (ddd, J =14.2/12.5/4.3 Hz, 1H, 5'- H_{ax}), 2.06 (dq, J=14.3/2.8 Hz, 1H, 5'- H_{equ}), 2.26 (td, J=13.4/4.3 Hz, 1H, 3'-H_{ax}), 2.64-2.74 (m, 2H, 2'-H, 6'-H), 2.75-2.86 (m, 7H, 4-H, 3-H_{isoquinoline}, NCH₂CH₂N), 2.88 (t, J=5.7 Hz, 2H, 4-H_{isoquinoline}), 2.91–2.97 (m, 3H, 4-H, 2'-H, 6'-H), 3.55 (s, 3H, 3-OCH₃), 3.67 (s, 2H, 1-H_{isoquinoline}), 3.80 (s, 3H, 7-OCH₃), 3.81 (s, 3H, 6-OCH₃), 4.94 (dd, J=7.2/3.2 Hz, 1H, 3-H), 6.68 (s, 1H, 8-H_{isoquinoline}), 6.71 (s, 1H, 5-H_{isoquinoline}), 7.10 (d, J=7.5 Hz, 1H, 5-H), 7.18 (ddd, J=7.6/6.2/ 2.4 Hz, 1H, 6-H), 7.19-7.24 ppm (m, 2H, 7-H, 8-H). ¹³C NMR (151 MHz, CD₃OD): $\delta = 29.1$ (1 C, C-4_{isoquinoline}), 36.1 (1 C, C-4), 37.1 (1 C, C-5'), 39.5 (1 C, C-3'), 50.9 (1 C, C-2'), 51.0 (1 C, C-6'), 52.5 (1 C, C-3_{isoquinoline}), 56.1 (1 C, N_{isoquinoline}CH₂CH₂N), 56.46 (2 C, 3-OCH₃, 6-OCH₃ or 7-OCH₃), 56.51 (1 C, $N_{isoquinoline}CH_2CH_2N$), 56.7 (1 C, 6-OCH₃ or 7-OCH₃), 56.9 (1 C, C-1_{isoquinoline}), 75.6 (1 C, C-1), 97.9 (1 C, C-3), 111.2 (1 C, C-8_{isoquinoline}), 113.0 (1 C, C-5_{isoquinoline}), 125.7 (1 C, C-8), 127.3 (1 C, C-8a_{isoquinoline}), 127.4 (1 C, C-4a_{isoquinoline}), 127.6 (1 C, C-7), 127.9 (1 C, C-6), 130.2 (1 C, C-5), 132.7 (1 C, C-4a), 141.9 (1 C, C-8a), 148.9 (1 C, C- $7_{isoquinoline}$), 149.3 ppm (1 C, C- $6_{isoquinoline}$). FTIR (neat): ν [cm⁻¹] = 2947, 2820, 2778 (C-H_{alkyl}), 1516, 1466, 1454 (C=C_{arom}). Purity (HPLC): 91.2%, $t_{\rm R}$ = 14.0 min.

trans-N-[2-(6,7-Dimethoxy-3,4-dihydroisoquinolin-2(1*H*)-yl) ethyl]-3-methoxy-3,4-dihydrospiro[[2] benzopyran-1,1'-cyclohexan]-4'-amine (*trans*-31)



LiAlH₄ (4 mg, 0.12 mmol, 5.3 equiv) was added to a solution of amide trans-26 (11 mg, 0.02 mmol) in THF (3 mL) under N22. The mixture was heated to reflux for 22 h. After cooling to RT, $\mathrm{H_2O}$ (10 mL) was added, the precipitate was filtered off, and the aqueous phase was extracted with CH_2CI_2 (5×10 mL). The combined organic layers were dried (Na $_2\mathrm{SO}_4),$ filtered, concentrated in vacuo and the residue was purified twice by fc (d=1 cm, l=25 cm, V = 3 mL, CH_2CI_2/CH_3OH 99:1+1% *N*,*N*-dimethylethanamine; d =1 cm, l = 25 cm, V = 3 mL, CH_2CI_2/CH_3OH 99:1 \rightarrow 97:3+1% N,Ndimethylethanamine). Yellow solid, m.p. 73°C, yield 9 mg (86%). $C_{28}H_{38}N_2O_4$ (466.6 g/mol). $R_f = 0.33$ (CH₂Cl₂/CH₃OH 90:10+1% N,Ndimethylethanamine). HRMS (APCI): m/z 467.2895 (calcd. 467.2904 for $C_{28}H_{39}N_2O_4$ [*M*H⁺]). ¹H NMR (600 MHz, CD₃OD): $\delta = 1.60$ (dt, *J*= 9.5/2.0 Hz, 1H, 2'-H), 1.78-1.83 (m, 2H, 3'-H, 5'-H), 1.84-1.89 (m, 2H, 6'-H), 2.06-2.13 (m, 1H, 5'-H), 2.13-2.17 (m, 2H, 2'-H, 3'-H), 2.73-2.85 (m, 5H, 4-H, N_{isoquinoline}CH₂CH₂N, 3-H_{isoquinoline}), 2.86-2.95 (m, 5H, 4-H, $N_{isoquinoline}CH_2CH_2N$, 4- $H_{isoquinoline}$), 3.02 (quint, J=3.0 Hz, 1H, 4'- H_{equ}), 3.55 (s, 3H, 3-OCH₃), 3.66 (s, 2H, 1-H_{isoquinoline}), 3.78 (s, 3H, 7-OCH₃), 3.82 (s, 3H, 6-OCH₃), 4.90 (dd, J=7.5/3.1 Hz, 1H, 3-H), 6.68 (s, 1H, 8-H_{isoquinoline}), 6.73 (s, 1H, 5-H_{isoquinoline}), 7.01–7.08 (m, 2H, 5-H, 7-H), 7.09– 7.13 (m, 1H, 6-H), 7.17 ppm (d, J=7.8 Hz, 1H, 8-H). A signal for the NH proton is not observed in the spectrum. ¹³C NMR (151 MHz, CD₃OD): $\delta = 25.9$ (1 C, C-3'), 26.2 (1 C, C-5'), 29.5 (1 C, C-4_{isoquinoline}), 31.5 (1 C, C-6'), 34.1 (1 C, C-2'), 36.2 (1 C, C-4), 44.7 (1 C, $\begin{array}{l} \mathsf{N}_{\mathsf{isoquinoline}}\mathsf{CH}_2\mathsf{CH}_2\mathsf{N}), \quad 52.5 \quad (1\ \mathsf{C}, \ C-4'), \quad 52.5 \quad (1\ \mathsf{C}, \ C-3_{\mathsf{isoquinoline}} \ \text{or} \\ \mathsf{N}_{\mathsf{isoquinoline}}\mathsf{CH}_2\mathsf{CH}_2\mathsf{N}), \quad 56.3 \quad (1\ \mathsf{C}, \ 3-\mathsf{OCH}_3), \quad 56.4-56.5 \quad (\mathsf{m}, \ 2\ \mathsf{C}, \ 6-\mathsf{OCH}_3, \ 7-\mathsf{OCH}_3), \quad 56.8 \quad (1\ \mathsf{C}, \ C-1_{\mathsf{isoquinoline}}), \quad 58.0 \quad (1\ \mathsf{C}, \ C-3_{\mathsf{isoquinoline}} \ \text{or} \\ \mathsf{N}_{\mathsf{isoquinoline}}\mathsf{CH}_2\mathsf{CH}_2\mathsf{N}), \quad 77.8 \quad (1\ \mathsf{C}, \ C-1), \quad 97.9 \quad (1\ \mathsf{C}, \ C-3), \quad 111.3 \quad (1\ \mathsf{C}, \ C-8_{\mathsf{isoquinoline}}), \quad 125.9 \quad (1\ \mathsf{C}, \ C-3), \quad 111.3 \quad (1\ \mathsf{C}, \ C-8_{\mathsf{isoquinoline}}), \quad 127.5 \quad (1\ \mathsf{C}, \ C-7), \quad 127.55 \quad (1\ \mathsf{C}, \ C-6), \quad 127.62 \quad (1\ \mathsf{C}, \ C-8a_{\mathsf{isoquinoline}}), \quad 130.0 \quad (1\ \mathsf{C}, \ C-5), \quad 132.4 \quad (1\ \mathsf{C}, \ C-4a_{\mathsf{i},\mathsf{143.2}} \quad (1\ \mathsf{C}, \ C-8a_{\mathsf{i},\mathsf{148.9}}) \\ (1\ \mathsf{C}, \ C-7_{\mathsf{isoquinoline}}), \quad 149.3 \quad \mathsf{ppm} \quad (1\ \mathsf{C}, \ C-6_{\mathsf{isoquinoline}}), \quad \mathsf{FTIR} \quad (\mathsf{neat}): \quad \nu \\ [\mathsf{cm}^{-1}] = 3368 \quad (\mathsf{N}-\mathsf{H}), \quad 2924, \quad 2832 \quad (\mathsf{C}-\mathsf{H}_{\mathsf{alkyl}}), \quad 1516, \quad 1447 \quad (\mathsf{C}=\!\!\mathsf{C}_{\mathsf{arom}}). \\ \mathsf{Purity} \ (\mathsf{HPLC}): \ 61.5 \ \%, \ t_{\mathsf{R}} = 14.7 \ \mathsf{min}. \end{array}$

cis-N-[2-(6,7-Dimethoxy-3,4-dihydroisoquinolin-2(1*H*)-yl) ethyl]-3-methoxy-3,4-dihydrospiro[[2] benzopyran-1,1'-cyclohexan]-4'-amine (*cis*-31)



LiAlH₄ (8 mg, 0.22 mmol, 6.0 equiv.) was added to a solution of amide cis-26 (17 mg, 0.04 mmol) in THF (3 mL) under N₂. The mixture was heated to reflux for 19 h. After cooling to RT, H₂O (10 mL) was added, the precipitate was filtered off, and the aqueous phase was extracted with CH_2CI_2 (4×10 mL). The combined organic layers were dried (Na₂SO₄), filtered, concentrated in vacuo and the residue was purified by fc (d=1 cm, l=25 cm, V=3 mL, CH_2CI_2/CH_3OH 98:2 \rightarrow 98:2+1% *N,N*-dimethylethanamine). Yellow solid, m.p. 70 °C, yield 13 mg (76%). C28H38N2O4 (466.6 g/ mol). $R_{\rm f} = 0.26$ (CH₂Cl₂/CH₃OH 90:10+1% N,N-dimethylethanamine). HRMS (APCI): m/z 467.2921 (calcd. 467.2904 for $C_{28}H_{39}N_2O_4$ [*M*H⁺]). ¹H NMR (400 MHz, CD₃OD): δ = 1.65–1.82 (m, 3H, 2'-H, 3'-H, 5'-H), 1.82-1.96 (m, 3H, 3'-H, 5'-H, 6'-H), 2.03 (td, J=13.5/ 4.2 Hz, 1H, 6'-H), 2.10–2.18 (m, 1H, 2'-H), 2.71–2.78 (m, 3H, 4'-H_{ax}, N_{isoquinoline}CH₂CH₂N), 2.78-2.85 (m, 3H, 4-H, 3-H_{isoquinoline}), 2.88 (t, J= 5.9 Hz, 2H, 4-H_{isoquinoline}), 2.91-2.98 (m, 3H, 4-H, N_{isoquinoline}CH₂CH₂N), 3.58 (s, 3H, 3-OCH₃), 3.64 (s, 2H, 1-H_{isoquinoline}), 3.82 (s, 3H, 6-OCH₃ or 7-OCH₃), 3.82 (s, 3H, 6-OCH₃ or 7-OCH₃), 4.92 (dd, J=7.4/3.2 Hz, 1H, 3-H), 6.69 (s, 1H, 8-H_{isoquinoline}), 6.73 (s, 1H, 5-H_{isoquinoline}), 7.10 (d, J= 7.2 Hz, 1H, 5-H), 7.14-7.22 ppm (m, 3H, 6-H, 7-H, 8-H). A signal for the NH proton was not observed. ¹³C NMR (101 MHz, CD₃OD): $\delta =$ 29.0 (1 C, C-3' or C-5'), 29.1 (1 C, C-3' or C-5'), 29.2 (1 C, C-4_{isoquinoline}), 36.2 (1 C, C-4), 36.5 (1 C, C-2'), 39.0 (1 C, C-6'), 44.3 (1 C, $N_{isoquinoline}CH_2CH_2N$), 52.4 (1 C, C-3_{isoquinoline}), 56.4 (1 C, 3-OCH₃), 56.47 (1 C, 6-OCH₃ or 7-OCH₃), 56.53 (1 C, 6-OCH₃ or 7-OCH₃), 56.8 (1 C, C-1_{isoquinoline}), 57.4 (1 C, C-4'), 58.2 (1 C, N_{isoquinoline}CH₂CH₂N), 77.4 (1 C, C-1), 97.8 (1 C, C-3), 111.2 (1 C, C-8_{isoquinoline}), 113.0 (1 C, C-5_{isoquinoline}), 125.7 (1 C, C-8), 127.4 (1 C, C-4a_{isoquinoline}), 127.50 (1 C, C-7), 127.53 (1 C, C-8a_{isoquinoline}), 127.7 (1 C, C-6), 130.1 (1 C, C-5), 132.6 (1 C, C-4a), 142.6 (1 C, C-8a), 148.9 (1 C, C-7_{isoquinoline}), 149.3 ppm (1 C, C- $6_{isoquinoline}$). FTIR (neat): ν [cm⁻¹] = 3402 (N–H), 2928, 2832 (C–H_{alkyl}), 1516, 1447 (C=C_{arom}). Purity (HPLC): 69.2 %, t_R = 14.5 min.

Receptor binding studies

The σ_1 and σ_2 affinities were recorded as described in ref. [40]. Details of the assays are given in the Supporting Information.



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Conflict of Interest

The authors declare no conflict of interest.

Keywords: acyl linkers • alkyl linkers • sigma receptor affinity • spirocyclic sigma ligands • structure-affinity relationships • tetrahydroisoquinoline

- M. Hanner, F. F. Moebius, A. Flandorfer, H. G. Knaus, J. Striessnig, E. Kempner, H. Glossmann, Proc. Natl. Acad. Sci. USA 1996, 93, 8072–8077.
- [2] R. Quirion, W. D. Bowen, Y. Itzhak, J. L. Junien, J. M. Musacchio, R. B. Rothman, T. P. Su, S. W. Tam, D. P. Taylor, *Trends Pharmacol. Sci.* 1992, 13, 85–86.
- [3] H. R. Schmidt, R. M. Betz, R. O. Dror, A. C. Kruse, Nat. Struct. Mol. Biol. 2018, 25, 981–987.
- [4] H. R. Schmidt, S. D. Zheng, E. Gurpinar, A. Koehl, A. Manglik, A. C. Kruse, *Nature* 2016, *532*, 527–530.
- [5] S. B. Hellewell, A. Bruce, G. Feinstein, J. Orringer, W. Williams, W. D. Bowen, *Eur. J. Pharmacol.* **1994**, *268*, 9–18.
- [6] J. B. Xu, C. B. Zeng, W. H. Chu, F. H. Pan, J. M. Rothfuss, F. J. Zhang, Z. D. Tu, D. Zhou, D. X. Zeng, S. Vangveravong, F. Johnston, D. Spitzer, K. C. Chang, R. S. Hotchkiss, W. G. Hawkins, K. T. Wheeler, R. H. Mach, *Nat. Commun.* 2011, 2, 380.
- [7] C. Abate, M. Niso, V. Infantino, A. Menga, F. Berardi, Eur. J. Pharmacol. 2015, 758, 16–23.
- [8] A. Alon, H. R. Schmidt, M. D. Wood, J. J. Sahn, S. F. Martin, A. C. Kruse, Proc. Natl. Acad. Sci. USA, 2017, 114, 7160–7165.
- [9] A. Riad, C. Zeng, C. C. Weng, H. Winters, K. Xu, M. Makvandi, T. Metz, S. Carlin, R. H. Mach, *Sci. Rep.* 2018, *8*, 16845.
- [10] W. D. Bowen, Exp. Dermatol. 2004, 13, 570–570.
- [11] K. W. Crawford, W. D. Bowen, *Cancer Res.* **2002**, *62*, 313–322.
- [12] C. S. John, W. D. Bowen, V. M. Varma, J. G. Mcafee, T. W. Moody, *Life Sci.* 1995, 56, 2385–2392.
- [13] Z. Y. Liu, H. E. Nicholson, W. D. Bowen, *Cancer Res.* 2015, 75.
- [14] J. M. Walker, W. D. Bowen, F. O. Walker, R. R. Matsumoto, B. Decosta, K. C. Rice, *Pharmacol. Rev.* **1990**, *42*, 355–402.
- [15] X. Wang, W. Bowen, Cancer Res. 2009, 69, 422.
- [16] G. Cassano, G. Gasparre, M. Contino, M. Niso, F. Berardi, R. Perrone, N. A. Colabufo, *Cell Calcium*, 2006, 40, 23–28.
- [17] A. Azzariti, N. A. Colabufo, F. Berardi, L. Porcelli, M. Niso, G. M. Simone, R. Perrone, A. Paradiso, mol. *Cancer Ther.* 2006, *5*, 1807–1816.

- [18] C. C. Liu, C. F. Yu, S. C. Wang, H. Y. Li, C. M. Lin, H. H. Wang, C. Abate, C. S. Chiang, *BMC Cancer*, **2019**, *19*, 473.
- [19] G. Asong, X. Y. Zhu, B. Bricker, T. Andey, F. Amissah, N. Lamango, S. Y. Ablordeppey, *Bioorg. Med. Chem.* 2019, *27*, 2629–2636.
- [20] C. Zeng, C. C. Weng, M. E. Schneider Jr, L. Puentes, A. Riad, K. Xu, M. Makvandi, L. Jin, W. G. Hawkins, R. H. Mach, *Cell Death Discov.* 2019, 5, 58.
- [21] J. Perregaard, E. K. moltzen, E. Meier, C. Sanchez, J. Med. Chem. 1995, 38, 1998–2008.
- [22] F. Berardi, C. Abate, S. Ferorelli, N. Colabufo, R. Perrone, Cent. Nerv. Syst. Agents Med. Chem. 2009, 9, 205–219.
- [23] C. Abate, M. Niso, F. Berardi, *Future Med. Chem.* 2018, *10*, 1997–2019.
 [24] B. E. Blass, J. P. Rogers, *Expert Opin. Ther. Pat.* 2018, *28*, 655–663.
- [24] B. E. Blass, J. P. Rogers, Expert Opin. Iner. Pat. 2018, 28, 655–663.
- [25] C. Abate, S. V. Selivanova, A. Muller, S. D. Kramer, R. Schibli, R. Marottoli, R. Perrone, F. Berardi, M. Niso, S. M. Ametamey, *Eur. J. Med. Chem.* 2013, 69, 920–930.
- [26] R. H. Mach, Y. S. Huang, R. A. Freeman, L. Wu, S. Vangveravong, R. R. Luedtke, *Bioorg. Med. Chem. Lett.* 2004, 14, 195–202.
- [27] M. Niso, C. Riganti, M. L. Pati, D. Ghigo, F. Berardi, C. Abate, *ChemBioChem* 2015, *16*, 1078–1083.
- [28] Y. T. Sun, G. F. Wang, Y. Q. Yang, F. J. Jin, Y. F. Wang, X. Y. Xie, R. H. Mach, Y. S. Huang, *Eur. J. Med. Chem.* 2018,147, 227–237.
- [29] D. Z. Yang, A. Comeau, W. D. Bowen, R. H. Mach, B. D. Ross, H. Hong, M. E. Van Dort, *Pharm.* 2017, 14, 770–780.
- [30] R. Gitto, M. L. Barreca, L. De Luca, G. De Sarro, G. Ferreri, S. Quartarone, E. Russo, A. Constanti, A. Chimirri, J. Med. Chem. 2003, 46, 197–200.
- [31] R. Gitto, R. Caruso, B. Pagano, L. De Luca, R. Citraro, E. Russo, G. De Sarro, A. Chimirri, J. Med. Chem. 2006, 49, 5618–5622.
- [32] R. H. Mach, K. T. Wheeler, Cent. Nerv. Syst. Agents Med. Chem. 2009, 9, 230–245.
- [33] I. Lee, B. P. Lieberman, S. H. Li, C. Hou, M. Makvandi, R. H. Mach, Nucl. Med. Biol. 2016, 43, 721–731.
- [34] M. Bergkemper, E. Kronenberg, S. Thum, F. Börgel, C. Daniliuc, D. Schepmann, F. R. Nieto, P. Brust, R. F. Reinoso, I. Alvarez, B. Wünsch, J. Med. Chem. 2018, 61, 9666–9690.
- [35] C. A. Maier, B. Wünsch, J. Med. Chem. 2002, 45, 438-448.
- [36] E. Große Maestrup, C. Wiese, D. Schepmann, A. Hiller, S. Fischer, M. Scheunemann, P. Brust, B. Wünsch, *Bioorg. Med. Chem.* 2009, 17, 3630–3641.
- [37] E. Kronenberg, F. Weber, S. Brune, D. Schepmann, C. Almansa, K. Friedland, E. Laurini, S. Pricl, B. Wünsch, J. Med. Chem. 2019, 62, 4204– 4217.
- [38] E. Rack, R. Fröhlich, D. Schepmann, B. Wünsch, Bioorg. Med. Chem. 2011, 19, 3141–3151.
- [39] S. Chander, P. Ashok, A. Singh, S. Murugesan, Chem. Cent. J. 2015, 9, 33– 36.
- [40] P. Hasebein, B. Frehland, K. Lehmkuhl, R. Fröhlich, D. Schepmann, B. Wünsch, B. Org. Biomol. Chem. 2014, 12, 5407–5426.

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