### Pancreatic cancer tumour initiating cells: the molecular regulation and therapeutic values

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#### Introduction

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### Abstract

Pancreatic cancer is an aggressive solid tumour characterized by its local invasion, early metastasis and resistance to standard chemotherapy or radiation therapy. Tumour initiating cells (TICs) are not only capable of self-renewal and differentiation, but also play an important role in multi-drug resistance, and thus become a popular topic in cancer research especially in pancreatic cancer. In this review, we summarize the current progress of TICs in tumourigenesis, various newly identified surface markers of pancreatic TICs, and the signalling pathways such as epithelial-mesenchymal transition, sonic hedgehog and Notch that regulate TICs. We also discuss the role which microRNA plays in TICs as well as its application in TIC-targeted therapy along with other approaches.

Keywords: tumour initiating cells • microRNA • pancreatic cancer

### Introduction

Pancreatic cancer is known for its notorious mortality rate and the lowest overall survival among all cancers. In 2011, the incidence of pancreatic cancer has gradually increased with an estimated 44,030 newly diagnosed cases in the United States. Unfortunately, 37,660 of these patients are expected to succumb to this deadly disease, making pancreatic cancer the fourth leading cause of cancer death. Pancreatic cancer is an aggressive solid tumour with local invasion, early metastasis, and resistance to standard chemotherapy and radiation therapy [1,2]. In 1997, gemcitabine was approved by the FDA as the first-line chemotherapy drug for

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patients with locally advanced or metastatic pancreatic adenocarcinoma [3]. From then on, many studies have been done with the goal of improving clinical efficacy of chemotherapy. Unfortunately, little progress has been made and the overall survival has not improved up to now [4]. Drug resistance usually contributes to treatment failure and plays a significant role in high mortality in patients diagnosed with this cancer. Previous studies have indicated various mechanisms of drug resistance in pancreatic cancer, such as changes in individual genes or signalling pathways, and the influence of the tumour microenvironment [5].

MSB 3.000 Houston, TX 77030, USA. Tel: (713) 500-6491 Fax: (713) 500-6493 E-mail: Min.Li@uth.tmc.edu Current therapeutic strategies for patients with pancreatic cancer involve targeting and killing differentiated cancer cells as well as the quiescent tumour initiating cells (TICs) [6].

Tumour initiating cells are defined as a unique subpopulation of cells that possess the ability to initiate tumour growth and sustain self-renewal as well as metastatic potential from which they were isolated or identified [7]. The definition implies TICs' ability to induce tumourigenesis in xenotransplanted immunodeficient mice. Tumour initiating cells are also referred to as "cancer stem cells (CSCs)" or "tumourigenic cells" in many studies [8]. The first evidence regarding TICs was observed in a lymphoma study, sparkling the debate on the role of TIC in cancer progression [9] while the TICs were first identified by Park *et al.* when they observed extensive proliferation of a subset of cancer cells isolated from myeloma mice in 1971 [10]. These special sets of cells can self-renew and differentiate to develop the cellular and molecular heterogeneity of the originating tumour [11].

The lineage of TICs is still under great debate, however, many investigators have hypothesized that TICs arise from normal stem or progenitor cells after accumulation of genetic mutations [12]. In some cases, TICs may also arise from differentiated cells such as acinar cells that are once committed but re-acquire stem cell characteristics after mutations take place. As the lineage of pancreatic cancer still remains unclear, tracking down the origin of TICs in pancreas has posed great challenges.

The TICs' existence has now been validated in several studies on solid tumours, such as breast cancer [13], glioblastoma [14], colorectal cancer [15] and liver cancer [16]. The concept of TICs provides a distinct view of carcinogenesis and may give rise to novel therapeutic strategies for pancreatic cancer, preventing tumour recurrence. Therefore, elucidating the mechanisms underlying pancreatic tumourigenesis employing TICs is clinically significant to improve the treatment of pancreatic cancer [17].

### Surface markers of pancreatic TICs

Consistent to its role in other solid tumours, TICs are also responsible for tumour recurrence as well as tumour metastasis in pancreatic cancer. Li et al. identified a subpopulation of highly tumourigenic cancer cells expressing the cell surface marker CD44<sup>+</sup>, CD24<sup>+</sup> and epithelial-specific antigen (ESA<sup>+</sup>), and these CD44<sup>+</sup>CD24<sup>+</sup>ESA<sup>+</sup> cancer cells display stem cell-like characteristics such as self-renewing and the ability to produce differentiated cells as well as drive continuous malignant cell expansion in a invasive and metastatic manner. As few as hundreds of ESA<sup>+</sup>CD44<sup>+</sup>CD24<sup>+</sup> cells were able to generate a tumour in 50% of the animals compared with the cells negative for all three markers (CD44<sup>-</sup>CD24<sup>-</sup>ESA<sup>-</sup>) that would require 10<sup>4</sup> or more cells implanted to induce tumour formation [18]. In another study, Hermann et al. used CD133<sup>+</sup> as a marker to isolate pancreatic cancer cells with a significantly higher tumourigenic potential and found these CD133<sup>+</sup> expressing cancer cells are highly resistant

to standard chemotherapy. In addition, the authors also found CD133<sup>+</sup>CXCR4<sup>+</sup> expressing cancers cells to be essential for tumour metastasis [19]. Moreover, Shah et al. showed that gemcitabine-resistant pancreatic cancer cells have increased expression of CD24<sup>+</sup>, CD44<sup>+</sup> and ESA<sup>+</sup> while possess the morphological and biochemical properties of epithelial-mesenchymal transition (EMT) [20]. In a more recent study, Rasheed et al. identified aldehyde dehydrogenase positive (ALDH<sup>+</sup>) pancreatic cancer cells with stem cell-like features, clonogenic potential and characteristics of EMT [21]. They also suggested that ALDH<sup>+</sup> pancreatic cancer cells can negatively affect the overall survival of cancer patients. A recent study by Kim et al. [22] demonstrated cell populations with high ALDH<sup>+</sup> activity alone are sufficient for efficient tumour-initiation and recapitulating the phenotype of original tumour in NOD/SCID mice regardless of the level of CD133<sup>+</sup> expression, indicating ALDH<sup>+</sup> might possibly be a more ideal marker to identify novel therapeutic targets for pancreatic cancer.

### Signalling pathways in pancreatic TICs

Conventionally, EMT is recognized as a pathological mechanism during the progression of various diseases including inflammation. fibrosis and cancer [23]. In recent years, emerging evidence has implicated that EMT plays a critical role in the molecular mechanism of TICs in pancreatic cancer. Mani et al. [24] found that EMT generates stem cell-like cells when they induced EMT in non-tumourigenic human mammary epithelial cells (HMLEs) and later identified those EMT-induced cells displaying CD44<sup>high</sup>/CD24<sup>low</sup> pattern, associated with both human breast CSCs and normal mammary epithelial stem cells. Their findings suggested cells undergo EMT share many markers and properties with tumour-initiating cells, indicating a possible mechanism involved in both EMT and self-renewal. Thus, the inhibition of EMT along with other pluripotency maintaining factors in pancreatic TICs isolated from Kras<sup>G12D</sup> mice by anticancer drug resveratrol indicated that resveratrol can be used to target TICs and suppress their metastatic potential [25]. Other than inhibiting EMT markers, the authors also found resveratrol's ability to inhibit the self-renewal capacity of pancreatic TICs by preventing the formation of primary and secondary spheroid, implicating its role in pancreatic TICs management.

In addition to EMT signalling, other important signalling pathways associated TICs such as sonic hedgehog (SHH) was elucidated by several studies. It has been shown that down-regulation of SHH by cyclopamine, an inhibitor of SHH, can reduce the growth and viability of pancreatic cancer cells [26]. Li *et al.* assessed the expression levels of SHH by real-time reverse transcriptase polymerase chain reaction (RT-PCR) and found a 46-fold increase in the expression levels of SHH in pancreatic cancer TICs and a four-fold increase in CD44<sup>-</sup>CD24<sup>-</sup>ESA<sup>-</sup> pancreatic cancer cells compared with normal pancreatic epithelial cells [18]. The SHH inhibitor cyclopamine is administered as a part of the dual-compartment therapeutic approach to further reduce tumour growth and

decreased both static and dynamic pancreatic TIC markers such as CD24 and ALDH [27]. These studies suggest that SHH represents a promising target for therapeutic interventions which may facilitate in eliminating pancreatic TICs.

The Notch signalling pathway is typically involved in cellular development through regulation of cell proliferation and apoptosis [28]. Notch gene is abnormally activated in many human malignancies such as lung cancer [29], prostate cancer [30] and pancreatic cancer [31]. Four Notch proteins have been identified in a variety of stem cells [32]. Down-regulation of Notch-1 using specific small interfering RNA (siRNA) was correlated with decreased proliferative rates, increased apoptosis, reduced migration and decreased invasive properties of pancreatic cancer cells. Notch signalling could induce the activity of its downstream target NF-KB in pancreatic cancer [33], and forced over-expression of Notch-1 could lead to overgrowth, clonogenicity, migration and invasion of AsPC-1 cells. The over-expression of Notch-1 can also lead to the induction of EMT phenotype and up-regulation of TIC markers such as CD44<sup>+</sup> and ESA<sup>+</sup>, suggesting that targeting Notch-1 pathway may be an effective therapeutic strategy to treat pancreatic TICs [34].

## Function of microRNAs in TICs regulation, self-renewal and differentiation

MicroRNAs (miRNAs) are small conserved non-coding RNAs with the length of 18-25 nucleotides that act as translational regulators of genes involved in many cellular processes, such as development, differentiation, cell proliferation, survival and death [35]. These regulatory molecules can directly interfere differentiation in both normal stem cells [36] and TICs [37]. Recently, miRNAs have become a spotlight in cancer genetics because of their ability to regulate gene expression. Previous studies found that the expression of miRNAs is altered in haematologic [38] and solid tumours [39] when compared with their normal counterparts. Meanwhile, emerging evidence has shown that several miRNAs may play an important role in the development and progression of diverse tumours, such as breast [40], ovarian [41], endometrial [42], hepatocellular [43], colon [44], esophageal [45], lung [46] and pancreatic cancer [47]. To date, the associations between miRNAs and TICs are still not completely understood. Ji et al. found that miR-34 restoration inhibits the CD44<sup>+</sup>/CD133<sup>+</sup> MIA PaCa-2 cells, accompanied by significant inhibition of tumour sphere growth in vitro and tumour formation in vivo. Furthermore, miR-34 is involved in pancreatic TICs' self-renewal, potentially via the direct modulation of downstream targets Notch and Bcl-2 [48]. Interestingly, in another study of prostate TICs, Liu et al. found that systemically delivered miR-34a inhibited prostate cancer metastasis as well as extended survival of tumour-bearing mice. Moreover, CD44<sup>+</sup> was identified as a direct and functional target of miR-34a, and the CD44<sup>+</sup> knockdown mimicked the effect of miR-34a over-expression in inhibiting prostate cancer regeneration and

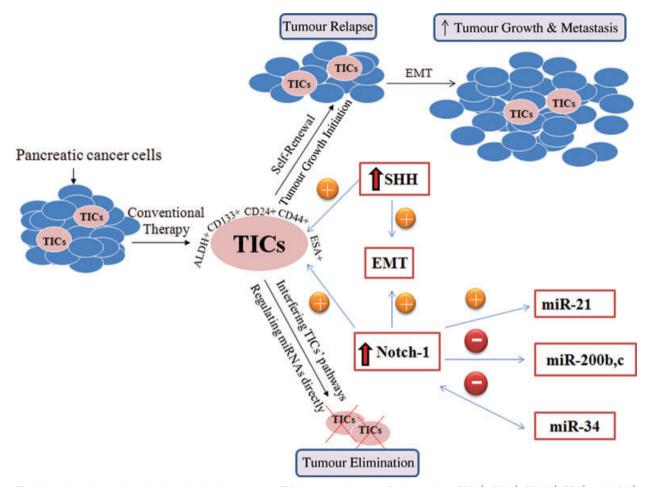
metastasis [49]. These studies strongly suggest that miR-34a is a negative regulator of TICs and can serve as a novel therapeutic target in cancer treatment.

Another TIC-regulating miRNA complex in pancreatic cancer is the miR-200 family which is reciprocally linked to zinc-finger enhancer binding (ZEB)/miR-200 feedback loop, whereas ZEB transcription factors are crucial to EMT activators [50]. According to their seed sequences, the members of miR-200 family can be further divided into two subaroups which showed slight difference in their target gene sets. Subgroup A contains miR-141 and miR-200a while subgroup B contains miR-200b, miR-200c and miR-429. The most prominent targets of the miR-200 family are two E box binding transcription factors, ZEB1 and ZEB2, and C. elegans Sma and Drosophila Mad (SMAD) interacting protein 1 (SIP1) [51-56]. Transient over-expression of miR-200c decreased the sphere-forming capacity of wild-type Panc-1 and MIA PaCa-2 cells. Conversely, specific inhibition of endogenous miR-200c led to an increase in sphere numbers in the differentiated pancreatic cancer cell line BxPC-3 cells [57]. The result is consistent with another study which found that the expression of miR-200b, miR-200c, let-7b. let-7c. let-7d and let-7e was significantly down-regulated in gemcitabine-resistant cells with TICs' characteristics [58].

Similar to the miR-34 and miR-200 families, most miRNAs were down-regulated in many types of tumours [39, 59] except for miR-21, which was dramatically up-regulated in the analysed tumours of breast, colon, lung, stomach, prostate and pancreas [59]. The human miR-21 gene is well characterized and mapped to chromosome 17q23.2, where it overlaps with the protein-coding gene VMP1, a human homologue of rat vacuole membrane protein [60,61]. Meng et al. found that miR-21 is a downstream effecter of PI3-kinase signalling pathway in human cholangiocarcinoma cells [62]. It has been shown that aberrant expression of miR-21 may contribute to the growth of hepatocellular carcinoma and regulate the expression of the tumour suppressor phosphatase and tensin (PTEN) homologue [63]. Similar results were found in another study of pancreatic cancer cell lines by Bao et al. [64]. In addition, expression of miR-21 is found to be up-regulated in gemcitabine-resistant pancreatic cancer cells with TIC markers (CD44<sup>+</sup> and ESA<sup>+</sup>). Furthermore, suppressing the expression of miR-21 could lead to re-expression of PTEN and miR-200, reversing the EMT phenotype in the cells. Those studies indicate that aberrant expression of miRNAs certainly contribute to the molecular regulation of pancreatic TICs and may play important roles in TIC-targeted cancer therapy.

# New therapies targeting the deregulated pathways and miRNAs to treat TICs in pancreatic cancer

Because of its self-renewal capability and high resistance to chemotherapy, targeted killing of pancreatic TICs may become an



**Fig. 1** New therapies targeting the deregulated pathways to treat TICs in pancreatic cancer. Surface markers CD24<sup>+</sup>, CD44<sup>+</sup>, CD133<sup>+</sup>, ESA<sup>+</sup> and ALDH<sup>+</sup> have been identified to help isolate TICs. Conventional therapy can reduce the tumour size by exerting effect on cancer cells; however, it does not eradicate TICs. The remaining TICs possess self-renewal ability and can re-initiate tumour growth, increasing the risk of tumour recurrence as well as promoting tumour metastasis. With the use of TIC-targeting therapy that eliminates TICs by either interfering with the signalling pathway or regulating the activity of miRNAs, complete tumour eradication is possible. The signalling pathways associated with TICs include the SHH pathway and the Notch-1 pathway that both up-regulate the activity of EMT and TIC itself. The Notch-1 pathway can also influence the expression of miRNA such as miR21, miR200b, miR200c and miR34 that all play a role in TIC regulation.

effective strategy for preventing pancreatic cancer recurrence as well as overcoming drug resistance. Therefore, one of the most promising approaches to target TICs is to inhibit TIC-associated pathways (Fig. 1). An *in vitro* and *in vivo* model of pancreatic cancer has been used to examine the effects of cyclopamine SHH inhibition and mammalian target of rapamycin (mTOR) blockade on the TICs population. It was found that the eradication of pancreatic TICs only occurred when combined blockade of SHH and mTOR signalling were used together with standard chemotherapy [65]. Curcumin, a well-used dietary polyphenol [22] was demonstrated to down-regulate Notch-1 mRNA level in pancreatic cancer cells. This study has shown that curcumin-induced inactivation of NF- $\kappa$ B DNA-binding activity was potentially mediated by Notch-1 signalling pathway [66]. Recently, difluorinated-curcumin (CDF), a novel synthetic analogue of curcumin, had been proved to be more

potent. Difluorinated-curcumin can inhibit the sphere-forming ability of drug-resistant pancreatic cancer cells, which is consistent with inactivation of TIC biomarkers such as CD44 and ESA. The anti-tumour activity was showed in both CDF treatment alone and in combinational therapy of CDF with gemcitabine. Difluorinated-curcumin treatment significantly inhibited tumour growth in a xenograft mouse model of MIA PaCa-2 by decreasing the DNA binding activity of NF- $\kappa$ B, down-regulating the expression of COX-2 and miR-21 as well as increasing PTEN and miR-200 expression in tumour remnants [64]. Interestingly, sulforaphane (SF), another plant compound isothiocyanate enriched in broccoli and other cruciferous vegetables, also demonstrated the ability to eliminate MIA PaCa-2 pancreatic cancer cells with TIC characteristics such as clonogenicity, spheroidal growth and ALDH1 activity. Nevertheless, co-treatment with SF and gemcitabine had a synergistic effect on the inhibition of tumour growth compared to the treatment with gemcitabine alone [67]. Another study also demonstrated that combining quercetin may increase the efficacy of SF to eliminate pancreatic TICs [68]. Moreover, resveratrol, a polyphenol derived from a wide variety of plants such as grapes. berries, plums and peanuts was also found to be a member of natural dietary compounds which can exert effect on pancreatic TICs by inhibiting the EMT markers [25]. As induction of EMT can lead to invasion of surrounding stroma and colonization of distant sites, inhibiting EMT in TICs will cause decreased metastasis. Meanwhile, Genistein, one of the most active soy isoflavones, was found to display similar effect [34]. Blocking the hypoxia, angiogenesis and inflammation pathways in TICs is also a promising therapeutic strategy to inhibit the interaction of the TICs with the surrounding stromal and vascular endothelial cells to prevent tumour metastasis.

As miRNAs are critical regulators for TICs in different cancers, many miRNA-based therapies have been developed lately to treat pancreatic TICs [69]. Knockdown of doublecortin and Ca<sup>2+</sup>/calmodulin-dependent kinase-like-1 (DCAMKL-1) in human pancreatic cancer cells by siRNA induces miR-200a but down-regulates ZEB1, ZEB2, Snail, Slug and Twist. Furthermore, DCAMKL-1 knockdown resulted in down-regulation of c-Myc and Kras through a let-7a miRNA-dependent mechanism as well as downregulation of Notch-1 through a miR-144 miRNA-dependent mechanism. These findings suggest the direct regulatory links between DCAMKL-1. miRNAs and EMT in pancreatic cancer. These results also justify that DCAMKL-1 can serve as a potential therapeutic target for eradicating pancreatic TICs [70]. In a recent study, Pramanik et al. synthesized a lipid-based nanoparticle for systemic delivery of miRNA expression vectors with a diameter of 100 nm. Systemic intravenous delivery with either miR-34a or miR-143/145 nanovectors inhibited the tumour growth in both MIA PaCa-2 subcutaneous and orthotopic xenograft mice. MicroRNA restitution was confirmed in treated xenografts by significant up-regulation of the corresponding miRNA while dramatically decreases in miR-34a targets such as SIRT1, CD44 and ALDH and miR-143/145 targets such as KRAS2 and RREB1 [71]. Different from other TICs, pancreatic TICs have unique miRNA expression patterns. Recent data demonstrated that drug resistance is regulated not only by genetic and epigenetic changes, but also by miRNAs [72]. At present, several important tumour suppressor miRNAs, oncogenic miRNAs and their molecular targets have been identified in pancreatic cancer. Thus, utilizing miRNAs as one of the therapeutic targets offers a promising treatment strategy to eradicate pancreatic TICs. However, more in-depth studies are needed to better understand the molecular mechanism and design more effective therapies [73].

### Conclusion

Although there have been some improvements in the diagnostic and surgical modalities as well as chemotherapeutic efficacy, lack of early diagnosis and successfully curative resection still poses great challenges for treating pancreatic cancers over the last decades [74]. Tumour recurrence and chemoresistance due to TICs also hinder the survival rate of pancreatic cancer patients. Emerging evidence has gradually revealed an intertwined connection between TICs, signalling pathways and miRNAs, shedding new lights in tailoring successful therapy against pancreatic TICs that can potentially benefit the survival of this deadly disease. In addition to their therapeutic value, circulating TICs can be utilized in assessing the risk of recurrence and prognosis in pancreatic cancer as it has been used in patients who suffer from melanoma [75] or colon cancers [76]. Various surface markers have helped isolate TICs from solid tumour; however, a more refined technique is needed for precise isolation of these tumourigenic cells in upcoming investigation on TICs both in vitro and in vivo. The validity of these critical TIC markers in TIC identification also has to be justified with larger patient sample size. There is no doubt that pancreatic TICs, their associated signalling pathways as well as miRNAs have come to light as promising targets for cancer therapy, revolutionizing the way of traditional cancer therapy is performed, however, further studies are needed to unravel the detailed mechanism of TICs in pancreatic cancer.

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### **Conflict of interest**

The authors declare no conflict of interest.

### References

- Jemal A, Siegel R, Xu J, et al. Cancer statistics, 2010. CA Cancer J Clin. 2010; 60: 277–300.
- 2. Siegel R, Ward E, Brawley O, et al. Cancer statistics, 2011: the impact of

eliminating socioeconomic and racial disparities on premature cancer deaths. *CA Cancer J Clin.* 2011; 61: 212–36. **Burris HA, 3rd, Moore MJ, Andersen J, et al.** Improvements in survival

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and clinical benefit with gemcitabine as first-line therapy for patients with advanced pancreas cancer: a randomized trial. *J Clin Oncol.* 1997; 15: 2403–13.

- Moore MJ, Goldstein D, Hamm J, et al. Erlotinib plus gemcitabine compared with gemcitabine alone in patients with advanced pancreatic cancer: a phase III trial of the National Cancer Institute of Canada Clinical Trials Group. J Clin Oncol. 2007; 25: 1960–6.
- Long J, Zhang Y, Yu X, et al. Overcoming drug resistance in pancreatic cancer. Expert Opin Ther Targets. 2011; 15: 817–28.
- Yu X, Zhang Y, Chen C, et al. Targeted drug delivery in pancreatic cancer. *Biochim Biophys Acta*. 2010; 1805: 97–104.
- Takaishi S, Okumura T, Wang TC. Gastric cancer stem cells. J Clin Oncol. 2008; 26: 2876–82.
- Saikawa Y, Fukuda K, Takahashi T, et al. Gastric carcinogenesis and the cancer stem cell hypothesis. Gastric Cancer. 2010; 13: 11–24.
- Bruce WR, Van Der Gaag H. A quantitative assay for the number of murine lymphoma cells capable of proliferation *in vivo*. *Nature*. 1963; 199: 79–80.
- Park CH, Bergsagel DE, McCulloch EA. Mouse myeloma tumour stem cells: a primary cell culture assay. J Natl Cancer Inst. 1971; 46: 411–22.
- Lapidot T, Sirard C, Vormoor J, et al. A cell initiating human acute myeloid leukaemia after transplantation into SCID mice. Nature. 1994; 367: 645–8.
- Dorado J, Lonardo E, Miranda-Lorenzo I, et al. Pancreatic cancer stem cells: new insights and perspectives. J Gastroenterol. 2011; 46: 966–73.
- AI-Hajj M, Wicha MS, Benito-Hernandez A, et al. Prospective identification of tumourigenic breast cancer cells. Proc Natl Acad Sci USA. 2003; 100: 3983–8.
- Singh SK, Hawkins C, Clarke ID, et al. Identification of human brain tumour initiating cells. *Nature*. 2004; 432: 396–401.
- Ricci-Vitiani L, Lombardi DG, Pilozzi E, et al. Identification and expansion of human colon-cancer-initiating cells. *Nature*. 2007; 445: 111–5.
- Ma S, Chan KW, Hu L, et al. Identification and characterization of tumourigenic liver cancer stem/progenitor cells. Gastroenterology. 2007; 132: 2542–56.
- Frank NY, Schatton T, Frank MH. The therapeutic promise of the cancer stem cell concept. J Clin Invest. 2010; 120: 41–50.
- Li C, Heidt DG, Dalerba P, et al. Identification of pancreatic cancer stem cells. *Cancer Res.* 2007; 67: 1030–7.
- Hermann PC, Huber SL, Herrler T, et al. Distinct populations of cancer stem cells determine tumour growth and metastatic

activity in human pancreatic cancer. *Cell Stem Cell.* 2007; 1: 313–23.

- Shah AN, Summy JM, Zhang J, et al. Development and characterization of gemcitabine-resistant pancreatic tumour cells. Ann Surg Oncol. 2007; 14: 3629–37.
- Rasheed ZA, Yang J, Wang Q, et al. Prognostic significance of tumourigenic cells with mesenchymal features in pancreatic adenocarcinoma. J Natl Cancer Inst. 2010; 102; 340–51.
- Kim MP, Fleming JB, Wang H, et al. ALDH activity selectively defines an enhanced tumour-initiating cell population relative to CD133 expression in human pancreatic adenocarcinoma. *PLoS One.* 2011; doi:10.1371/journal. pone.0020636.
- Dvorak HF. Tumours: wounds that do not heal. Similarities between tumour stroma generation and wound healing. N Engl J Med. 1986; 315: 1650–9.
- Mani SA, Guo W, Liao MJ, et al. The epithelial-mesenchymal transition generates cells with properties of stem cells. *Cell*. 2008; 133: 704–15.
- Shankar S, Nall D, Tang SN, et al. Resveratrol inhibits pancreatic cancer stem cell characteristics in human and KrasG12D transgenic mice by inhibiting pluripotency maintaining factors and epithelial-mesenchymal transition. *PLoS One.* 2011; doi:10.1371/journal.pone. 0016530.
- Berman DM, Karhadkar SS, Maitra A, et al. Widespread requirement for Hedgehog ligand stimulation in growth of digestive tract tumours. *Nature*. 2003; 425: 846–51.
- Jimeno A, Feldmann G, Suarez-Gauthier A, et al. A direct pancreatic cancer xenograft model as a platform for cancer stem cell therapeutic development. Mol Cancer Ther. 2009; 8: 310–4.
- Ohishi K, Katayama N, Shiku H, et al. Notch signalling in hematopoiesis. Semin Cell Dev Biol. 2003; 14: 143–50.
- Sriuranpong V, Borges MW, Ravi RK, et al. Notch signaling induces cell cycle arrest in small cell lung cancer cells. *Cancer Res.* 2001; 61: 3200–5.
- Shou J, Ross S, Koeppen H, et al. Dynamics of notch expression during murine prostate development and tumourigenesis. *Cancer Res.* 2001; 61: 7291–7.
- Miyamoto Y, Maitra A, Ghosh B, et al. Notch mediates TGF alpha-induced changes in epithelial differentiation during pancreatic tumourigenesis. Cancer Cell. 2003; 3: 565–76.

- Mumm JS, Kopan R. Notch signaling: from the outside in. *Dev Biol.* 2000; 228: 151–65.
- Wang Z, Zhang Y, Li Y, *et al.* Down-regulation of Notch-1 contributes to cell growth inhibition and apoptosis in pancreatic cancer cells. *Mol Cancer Ther.* 2006; 5: 483–93.
- Bao B, Wang Z, Ali S, et al. Notch-1 induces epithelial-mesenchymal transition consistent with cancer stem cell phenotype in pancreatic cancer cells. Cancer Lett. 2011; 307: 26–36.
- Garzon R, Marcucci G, Croce CM. Targeting microRNAs in cancer: rationale, strategies and challenges. *Nat Rev Drug Discov.* 2010; 9: 775–89.
- Hatfield SD, Shcherbata HR, Fischer KA, et al. Stem cell division is regulated by the microRNA pathway. *Nature*. 2005; 435: 974–8.
- He L, Thomson JM, Hemann MT, et al. A microRNA polycistron as a potential human oncogene. Nature. 2005; 435: 828–33.
- Lu J, Getz G, Miska EA, et al. MicroRNA expression profiles classify human cancers. *Nature*. 2005; 435: 834–8.
- Volinia S, Calin GA, Liu CG, et al. A microRNA expression signature of human solid tumours defines cancer gene targets. *Proc Natl Acad Sci USA*. 2006; 103: 2257–61.
- Iorio MV, Ferracin M, Liu CG, et al. MicroRNA gene expression deregulation in human breast cancer. *Cancer Res.* 2005; 65: 7065–70.
- Iorio MV, Visone R, Di Leva G, et al. MicroRNA signatures in human ovarian cancer. Cancer Res. 2007; 67: 8699–707.
- Boren T, Xiong Y, Hakam A, et al. MicroRNAs and their target messenger RNAs associated with endometrial carcinogenesis. *Gynecol Oncol.* 2008; 110: 206–15.
- Murakami Y, Yasuda T, Saigo K, et al. Comprehensive analysis of microRNA expression patterns in hepatocellular carcinoma and non-tumourous tissues. Oncogene. 2006; 25: 2537–45.
- Michael MZ, SM OC, van Holst Pellekaan NG, et al. Reduced accumulation of specific microRNAs in colorectal neoplasia. *Mol Cancer Res.* 2003; 1: 882–91.
- Nguyen GH, Schetter AJ, Chou DB, et al. Inflammatory and microRNA gene expression as prognostic classifier of Barrett's-associated esophageal adenocarcinoma. *Clin Cancer Res.* 2010; 16: 5824–34.

- Dacic S, Kelly L, Shuai Y, *et al.* miRNA expression profiling of lung adenocarcinomas: correlation with mutational status. *Mod Pathol.* 2010; 23: 1577–82.
- Bloomston M, Frankel WL, Petrocca F, et al. MicroRNA expression patterns to differentiate pancreatic adenocarcinoma from normal pancreas and chronic pancreatitis. JAMA. 2007; 297: 1901–8.
- Ji Q, Hao X, Zhang M, et al. MicroRNA miR-34 inhibits human pancreatic cancer tumour-initiating cells. *PLoS One*. 2009; 4: e6816. doi: 10.1371/journal.pone.0006816.
- Liu C, Kelnar K, Liu B, et al. The microRNA miR-34a inhibits prostate cancer stem cells and metastasis by directly repressing CD44. Nat Med. 2011; 17: 211–5.
- Brabletz S, Brabletz T. The ZEB/miR-200 feedback loop-a motor of cellular plasticity in development and cancer? *EMBO Rep.* 2010: 11: 670–7.
- Christoffersen NR, Silahtaroglu A, Orom UA, et al. miR-200b mediates post-transcriptional repression of ZFHX1B. RNA. 2007; 13: 1172–8.
- Burk U, Schubert J, Wellner U, et al. A reciprocal repression between ZEB1 and members of the miR-200 family promotes EMT and invasion in cancer cells. EMBO Rep. 2008; 9: 582–9.
- Gregory PA, Bert AG, Paterson EL, et al. The miR-200 family and miR-205 regulate epithelial to mesenchymal transition by targeting ZEB1 and SIP1. Nat Cell Biol. 2008; 10: 593–601.
- Hurteau GJ, Carlson JA, Spivack SD, et al. Overexpression of the microRNA hsa-miR-200c leads to reduced expression of transcription factor 8 and increased expression of E-cadherin. Cancer Res. 2007; 67: 7972–6.
- Korpal M, Lee ES, Hu G, et al. The miR-200 family inhibits epithelial-mesenchymal transition and cancer cell migration by direct targeting of E-cadherin transcriptional repressors ZEB1 and ZEB2. J Biol Chem. 2008; 283: 14910–4.
- 56. Park SM, Gaur AB, Lengyel E, et al. The miR-200 family determines the epithelial phenotype of cancer cells by targeting the

E-cadherin repressors ZEB1 and ZEB2. *Genes Dev.* 2008; 22: 894–907.

- 57. Wellner U, Schubert J, Burk UC, et al. The EMT-activator ZEB1 promotes tumourigenicity by repressing stemnessinhibiting microRNAs. *Nat Cell Biol.* 2009; 11: 1487–95.
- Li Y, VandenBoom TG, 2nd, Kong D, et al. Up-regulation of miR-200 and let-7 by natural agents leads to the reversal of epithelial-to-mesenchymal transition in gemcitabine-resistant pancreatic cancer cells. Cancer Res. 2009; 69: 6704–12.
- Krichevsky AM, Gabriely G. miR-21: a small multi-faceted RNA. J Cell Mol Med. 2009; 13: 39–53.
- Cai X, Hagedorn CH, Cullen BR. Human microRNAs are processed from capped, polyadenylated transcripts that can also function as mRNAs. *RNA*. 2004; 10: 1957–66.
- Fujita S, Ito T, Mizutani T, *et al.* miR-21 Gene expression triggered by AP-1 is sustained through a double-negative feedback mechanism. *J Mol Biol.* 2008; 378: 492–504.
- Meng F, Henson R, Wehbe-Janek H, et al. MicroRNA-21 regulates expression of the PTEN tumour suppressor gene in human hepatocellular cancer. *Gastroenterology*. 2007; 133: 647–58.
- Meng F, Henson R, Lang M, et al. Involvement of human micro-RNA in growth and response to chemotherapy in human cholangiocarcinoma cell lines. *Gastroenterology*. 2006; 130: 2113–29.
- Bao B, Ali S, Kong D, *et al.* Anti-tumour activity of a novel compound-CDF is mediated by regulating miR-21, miR-200, and PTEN in pancreatic cancer. *PLoS One.* 2011; doi:10.1371/journal.pone.0017850.
- Mueller MT, Hermann PC, Witthauer J, et al. Combined targeted treatment to eliminate tumourigenic cancer stem cells in human pancreatic cancer. *Gastroenterology*. 2009; 137: 1102–13.
- Wang Z, Zhang Y, Banerjee S, et al. Notch-1 down-regulation by curcumin is associated with the inhibition of cell growth and the induction of apoptosis in pancreatic cancer cells. *Cancer*. 2006; 106: 2503–13.

- Kallifatidis G, Labsch S, Rausch V, et al. Sulforaphane increases drug-mediated cytotoxicity toward cancer stem-like cells of pancreas and prostate. *Mol Ther.* 2011; 19: 188–95.
- Srivastava RK, Tang SN, Zhu W, et al. Sulforaphane synergizes with quercetin to inhibit self-renewal capacity of pancreatic cancer stem cells. Front Biosci (Elite Ed). 2011; 3: 515–28.
- Zimmerman AL, Wu S. MicroRNAs, cancer and cancer stem cells. *Cancer Lett.* 2011; 300: 10–9.
- Sureban SM, May R, Lightfoot SA, et al. DCAMKL-1 regulates epithelial-mesenchymal transition in human pancreatic cells through a miR-200a-dependent mechanism. Cancer Res. 2011; 71: 2328–38.
- Pramanik D, Campbell NR, Karikari C, et al. Restitution of tumour suppressor microRNAs using a systemic nanovector inhibits pancreatic cancer growth in mice. *Mol Cancer Ther.* 2011; 10: 1470–80.
- Fojo T. Multiple paths to a drug resistance phenotype: mutations, translocations, deletions and amplification of coding genes or promoter regions, epigenetic changes and microRNAs. *Drug Resist Updat.* 2007; 10: 59–67.
- Giovannetti E, Erozenci A, Smit J, et al. Molecular mechanisms underlying the role of microRNAs (miRNAs) in anticancer drug resistance and implications for clinical practice. *Crit Rev Oncol Hematol.* 2011; doi:10.1016/j.critrevonc. 2011.03.010.
- David M, Lepage C, Jouve JL, et al. Management and prognosis of pancreatic cancer over a 30-year period. Br J Cancer. 2009; 101: 215–8.
- Fusi A, Reichelt U, Busse A, et al. Expression of the stem cell markers nestin and CD133 on circulating melanoma cells. *J Invest Dermatol.* 2011; 131: 487–94.
- Iinuma H, Watanabe T, Mimori K, et al. Clinical significance of circulating tumour cells, including cancer stem-like cells, in peripheral blood for recurrence and prognosis in patients with Dukes' stage B and C colorectal cancer. J Clin Oncol. 2011; 29: 1547–55.