

Cutaneous phaeohyphomycosis caused by *Exophiala xenobiotica*: A case report



Clarissa Mitri Espanhol^a, Júlia Kanaan Recuero^a, Danielle Machado Pagani^b,
Amanda Carvalho Ribeiro^c, Gerson Vettorato^a, Rodrigo Pereira Duquia^a, Laura Luzzatto^d,
Maria Lúcia Scroferneker^{d,*}

^a Department of Dermatology of the Hospital Santa Casa de Misericórdia de Porto Alegre, Rua Prof. Annes Dias, 285, Porto Alegre, CEP:90020-090, Brazil

^b Postgraduate Program in Agricultural and Environmental Microbiology, Federal University of Rio Grande do Sul, Rua Sarmento Leite, 500, Sala 325, Porto Alegre, Rio Grande do Sul, CEP: 90050-170, Brazil

^c Faculty of Pharmaceutical Sciences, Federal University of Rio Grande do Sul, Avenida Ipiranga, 2752, CEP: 90610-000, Brazil

^d Department of Microbiology, ICBS, Universidade Federal do Rio Grande do Sul, Rua Sarmento Leite, 500, Sala 325, Porto Alegre, Rio Grande do Sul, CEP: 90050-170, Brazil

ARTICLE INFO

Keywords:

Exophiala xenobiotica
Phaeohyphomycosis
Immunosuppressed
Renal transplantation

ABSTRACT

We report a case of phaeohyphomycosis that affected the leg of a 45-year-old Brazilian man, car mechanic and renal transplanted. The direct mycological examination evidenced dematiaceous septated hyphae. The pathogenic fungal species was identified as *Exophiala xenobiotica*. Antifungal activity *in vitro* revealed terbinafine as the best antifungal. For treatment, it was chosen surgical excision of the entire lesion and used systemic itraconazole. Phaeohyphomycosis caused by *Exophiala xenobiotica* is extremely rare and is closely related to transplant patients.

1. Introduction

Dematiaceous fungi are found in nature and one of their characteristics is the presence of melanin, responsible for the dark pigmentation of their spores and conidia and that seems to behave as a virulence factor. More than a hundred species and sixty genera of these fungi are connected to a broad spectrum of human infections [1]. The main clinical forms of presentation of the infections through these dematiaceous fungi include phaeohyphomycoses (cutaneous, subcutaneous and systemic), that affect both immunocompetent and immunocompromised individuals [2].

Phaeohyphomycosis is a term used to denominate opportunistic, cutaneous and systemic infections [3]. The mainly genus are *Wangiella*, *Alternaria* and *Exophiala*. There are more than 100 species of fungi associated with phaeohyphomycosis [4]. The most prevalent species are *E. jeanselmei* and *E. dermatitidis*. *Exophiala xenobiotica* is a rare species. This disease usually affects rural and tropical populations in Central and South America [5,6]. The clinical polymorphism presents in phaeohyphomycosis is exacerbated by iatrogenic immunosuppression, making the diagnosis extremely difficult. The treatment is not defined, and it is usually carried out empirically [7].

2. Case

A 45-year-old white man, resident in Porto Alegre, car mechanic, hypertensive, with hepatitis C and renal transplanted two years ago, using mycophenolic acid, tacrolimus, prednisone, sulfamethoxazole-trimethoprim, sofosbuvir, daclatasvir and anti-hypertensive drugs, was attended in the department of Dermatology of the Hospital Santa Casa de Porto Alegre. He referred pruritic and fibrotic nodule on the right leg three months ago, with no secretion (Fig. 1A and B).

An incisional biopsy was performed, in the first day that he consulted (day 0), with histopathologic features of leveduriform cells and brownish hyphae within a dermal infiltrate which was compatible with phaeohyphomycosis (Fig. 2A and B).

On the same day of the biopsy (day 0), direct and culture microscopic examinations were performed. The fungal isolate was grown in Sabouraud agar medium in 30 °C for 14 days. Mycelium was taken and the genomic DNA was extracted (day14) using Power Soil DNA isolation kit (Mobio, USA). Polymerase chain reaction was performed (day 14) targeting the ITS1-5.8S rDNA-ITS2 region using the ITS1 and ITS4 universal primers and the polymerase chain reaction (PCR) was performed under the following conditions: denaturation step at 94 °C for 5

* Corresponding author. Department of Microbiology, ICBS, Universidade Federal do Rio Grande do Sul, Rua Sarmento Leite, 500, Room, 325, Porto Alegre, CEP: 90050-170, Brazil.

E-mail address: scrofern@ufrgs.br (M.L. Scroferneker).

<https://doi.org/10.1016/j.mmcr.2019.12.003>

Received 23 September 2019; Received in revised form 4 November 2019; Accepted 2 December 2019

Available online 26 December 2019

2211-7539/ © 2019 The Authors. Published by Elsevier B.V. on behalf of International Society for Human and Animal Mycology. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

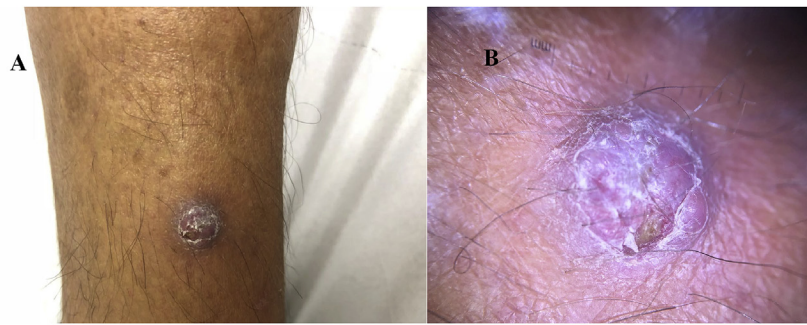


Fig. 1. (A) Fibrotic nodule in the right leg. (B) Fibrotic nodule - photo taken with dermlite 3gen DL4.

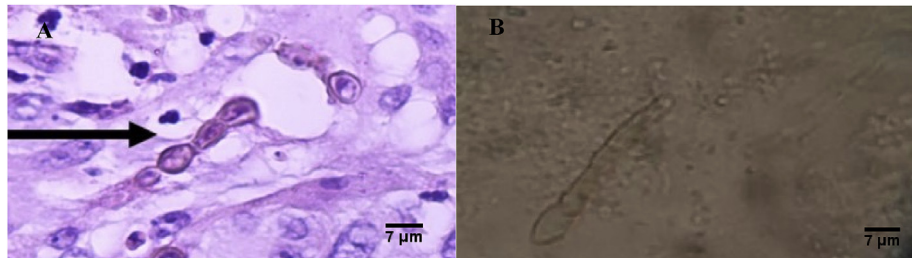


Fig. 2. (A) Hematoxylin Eosin Stain. Leveduriform cells and brownish hyphae within a dermal infiltrate (400X magnification). (B) Direct mycological examination - dematiaceous septated hyphae (400X magnification).

min, 30 cycles of denaturation at 94 °C for 1 min, annealing at 55 °C for 1 min, extension at 72 °C for 2 min, and final extension at 72 °C for 10 min [8].

The PCR product was purified using ExoSap-IT (Affumetrix, USA) and sequenced in the ABI-PRISM 300 Genetic Analyzer (Applied Biosystems) following the manufacturer's instruction. The obtained sequence was analyzed using Staden Package [9] and compared with sequence of type strain available in the GenBank database using the BLAST tool [10]. The sequence obtained had a coverage of 92% and 100% of identity with *Exophiala xenobiotica* (CBS118157) type strain. The sequence was deposited at GenBank database under the identification number MK411560. The strain susceptibility assay was performed (day 14) in triplicates against amphotericin B, itraconazole, ketoconazole, voriconazole, posaconazole and terbinafine in the final concentration range of 0.03–16 µg/mL, all drugs were from Sigma-Aldrich. Microdilution technique from M27-A3 standardized by the Clinical and Laboratory Standards Institute (CLSI) was followed [11]. The *Exophiala xenobiotica* was grown for 14 days at 30 °C and the inoculum was prepared with a 0.85% of sterile saline solution, filtered and the conidial-only presence was verified and counted in Neubauer chamber by microscopy, to a final concentration in the wells were 2.5×10^4 CFU/mL. The microplates were incubated in a temperature of 35 °C for up to 5 days, after, the minimal inhibitory concentrations (MICs) were determined by visualization of 100% of inhibition comparing the wells with the growth control. Antifungal activity *in vitro* revealed terbinafine as the best antifungal. *In vitro* antifungal activity MIC were: amphotericin B (16 µg/mL), itraconazole (16 µg/mL), ketoconazole (1 µg/mL), voriconazole (2 µg/mL), posaconazole (0,125 µg/mL) and terbinafine (> 0,5 µg/mL).

The patient returned 30 days after the biopsy with the renal transplant medical team that decided to treat the patient with itraconazole (day 30), that is available in the public health system. Itraconazol was given at a dose of 100 mg every 12 hours for three months (day 120). The patient could not do the treatment with terbinafine because it is not available in the public health system and he was not able to pay for medication. The dermatology team also chose to do a surgical excision of the entire lesion (day 33). The patient is in follow-up with no signs of relapse.

3. Discussion

We report a case of cutaneous phaeohyphomycosis caused by *E. xenobiotica* in a renal transplant patient treated with systemic itraconazole and surgery. Phaeohyphomycosis is an opportunistic disease. The incidence of phaeohyphomycosis in transplanted patients is increasing in the last decades due to the increased number of transplanted patients and the use of immunosuppressants, the majority of cases occur two years after transplantat. Calcineurin inhibitors appear to further increase susceptibility to fungal infections when compared to other immunosuppressants [5,12].

Surgical excision is the first choice for localized and well delimited lesions, it is preferred to use antifungal drugs before and after surgery, but there is no consensus about which drug to choose or how long the treatment will last. It is also recommended in severe or disseminated cases to reduce the doses of immunosuppressants [13–16]. Most melanized fungi are susceptible to azoles, which makes itraconazole and voriconazole the main drugs used, followed by amphotericin B [17].

The early clinical suspicion of phaeohyphomycosis in transplant patients is extremely important, because immunosuppression is the main risk factor for infection and responsible for the systemic spread of the disease, culminating in severe and fatal conditions. In transplant patients, the therapeutic is challenging due to the antifungals high doses required, besides the possibility of drug interactions. Based on our case and in the literature, it is extremely important to use the molecular identification to discover the causative agent, as well as the evaluation of *in vitro* antifungal activity to use the best treatment for the patient [18].

Declaration of competing interest

There are none.

References

- [1] S.G. Revankar, Dematiaceous fungi, *Mycoses* 50 (2007) 91–101.
- [2] H. Nishikawa, Y. Taniguchi, Phaeohyphomycosis due to *Exophiala oligosperma* in an immunocompromised host, *J. Rheumatol.* 46 (2019) 6.
- [3] S.R. Jinkala, D. Basu, S. Neelaiah, N. Stephen, S.B. Hanuman, R. Singh,

- Subcutaneous phaeohyphomycosis: a clinical mimic of skin and soft tissue neoplasms—a descriptive study from India, *World J. Surg.* 42 (2018) 3861–3866.
- [4] F. Silveira, M. Nucci, Emergence of Black moulds in fungal disease: epidemiology and therapy, *Curr. Opin. Infect. Dis.* 14 (6) (2001) 679–684.
- [5] W.R.P. Oliveira, M.F.L. Borsato, C. Festa Neto, L.A. Rocha, R.S. Nunes, Feohifomicose em transplante renal: relato de dois casos, *An. Bras. Dermatol.* 91 (1) (2016) 93–96.
- [6] A.L. Rossetto, R.A. Pérsio, R.C.B. Cruz, G. Dellatorre, J.C.M. Romeiro, Feo-hifomicose subcutânea por *Exophialajeanselmei* localizada na bolsa escrotal - Relato de caso, *An. Bras. Dermatol.* 85 (4) (2010) 517–520.
- [7] A. Singal, D. Pandhi, S.N. Bhattacharya, S. Das, S. Aggarwal, K. Mishra, Pheohyphomycosis caused by *Exophiala spinifera*: arareoccurrence, *Int. J. Dermatol.* 47 (2008) 44–47.
- [8] D. Heidrich, G.M. González, D.M. Pagani, M. Ramírez-Castrillón, M.L. Scroferneker, Chromoblastomycosis caused by *Rhinocladiella similis*: case report, *Med. Mycol. Case Rep.* 16 (2017) 25–27.
- [9] R. Staden, The staden sequence analysis package, *Mol. Biotechnol.* 5 (1996) 233–241.
- [10] S.F. Altschul, W. Gish, W. Miller, E.W. Myers, D.J. Lipman, Basic local alignment search tool, *J. Mol. Biol.* 215 (1990) 403–410.
- [11] CLSI, Reference method for broth dilution antifungal susceptibility testing of yeasts, Clinical and Laboratory Standards Institute (CLSI), Approved Standard M27- A3, third ed., 2008 Wayne, PA.
- [12] C.C. Hoffmann, I.P. Danuclov, K.S.M. Purim, F. Queiroz-Telles, Infecções causadas por fungos demácios e suas correlações anátomo-clínicas, *An. Bras. Dermatol.* 86 (1) (2011) 138–141.
- [13] M.A. Ocampo, J. Kanitakis, A.L. Bienvenu, C. Chauvet, S. Euvrard, Phaeohyphomycosis caused by *Pyrenochaetaaromeroi* mimicking a plantar wart in a kidney transplant recipient, *Transpl. Infect. Dis.* 14 (2012) E173–E174.
- [14] S.E. Vermeire, H. de Jonge, K. Lagrou, D.R. Kuypers, Cutaneous phaeohyphomycosis in renal allograft recipients: report of 2 cases and review of the literature, *Diagn. Microbiol. Infect. Dis.* 68 (2010) 177–180.
- [15] D. Cunha, C. Amaro, M.R. Vieira, L. Martins Mda, A.P. Maduro, J. Inácio, et al., Phaeohyphomycosis caused by *Alternaria* infectoria presenting as multiple vegetating lesions in a renal transplant patient, *Rev. Iberoam. De. Micol.* 29 (2012) 44–46.
- [16] N.C. Fernandes, D. Nacif, T. Akiti, T. Cuzzi, Subcutaneous phaeohyphomycosis caused by *Cladophialophora* sp.: a case report, São Paulo, *Rev. do Inst. Med. Trop. São Paulo* 49 (2) (Mar./Apr. 2007).
- [17] R. Isa-Isa, C. García, M. Isa, R. Arenas, Subcutaneous phaeohyphomycosis (mycotic cyst), *Clin. Dermatol.* 30 (2012) 425–431.
- [18] T.C. Daboit, R.P. Duquia, C.M. Magagnin, S.D.C. Mendes, M. Ramírez-Castrillón, R. Steglich, et al., A case of *Exophiala spinifera* infection in Southern Brazil: molecular identification and antifungal susceptibility, *Med. Mycol. Case Rep.* 1 (2012) 72–75.