

# **Obesity-Mediated Immune Modulation: One Step Forward, (Th)2 Steps Back**

Viviane Schmidt<sup>1</sup>, Andrew E. Hogan<sup>2,3</sup>, Padraic G. Fallon<sup>4</sup> and Christian Schwartz<sup>1,5\*</sup>

<sup>1</sup> Mikrobiologisches Institut - Klinische Mikrobiologie, Immunologie und Hygiene, Universitätsklinikum Erlangen and Friedrich-Alexander-Universität (FAU) Erlangen-Nürnberg, Erlangen, Germany, <sup>2</sup> Kathleen Lonsdale Human Health Institute, Maynooth University, Maynooth, Ireland, <sup>3</sup> Obesity Immunology Research, St. Vincent's University Hospital and University College Dublin, Dublin, Ireland, <sup>4</sup> Trinity Biomedical Sciences Institute, School of Medicine, Trinity College Dublin, Dublin, Ireland, <sup>5</sup> Medical Immunology Campus Erlangen, Friedrich-Alexander-Universität (FAU) Erlangen-Nürnberg, Erlangen, Germany

Over the past decades, the relationship between the immune system and metabolism has become a major research focus. In this arena of immunometabolism the capacity of adipose tissue to secrete immunomodulatory molecules, including adipokines, within the underlying low-grade inflammation during obesity brought attention to the impact obesity has on the immune system. Adipokines, such as leptin and adiponectin, influence T cell differentiation into different T helper subsets and their activation during immune responses. Furthermore, within the cellular milieu of adipose tissue nutrient availability regulates differentiation and activation of T cells and changes in cellular metabolic pathways. Upon activation, T cells shift from oxidative phosphorylation to oxidative glycolysis, while the differential signaling of the kinase mammalian target of rapamycin (mTOR) and the nuclear receptor PPARy, amongst others, drive the subsequent T cell differentiation. While the mechanisms leading to a shift from the typical type 2-dominated milieu in lean people to a Th1-biased pro-inflammatory environment during obesity are the subject of extensive research, insights on its impact on peripheral Th2-dominated immune responses become more evident. In this review, we will summarize recent findings of how Th2 cells are metabolically regulated during obesity and malnutrition, and how these states affect local and systemic Th2-biased immune responses.

Keywords: obesity, Th2 (type-2) immune responses, T helper cell 2, metabolism, helminth, malnutrition, adipokine cytokines

# INTRODUCTION

Obesity has become a major health problem, especially in first world countries, affecting about one third of the population worldwide (1). Individuals with a body-mass-index (BMI) >30 kg/m<sup>2</sup> are classified as obese, and develop an immune response that, as part of the metabolic syndrome, increases the risk of non-communicable diseases such as type 2 diabetes, hypertension, cardiovascular disease, asthma and cancer (2). Reasons for the development of obesity are multifactorial. By far the most important factor is a chronically dysregulated energy balance – with more energy being taken up than being burnt and thus stored as triglycerides in adipocytes. This leads to the accumulation of triglycerides in adipocytes,

#### **OPEN ACCESS**

#### Edited by:

Nicholas Van Panhuys, Sidra Medical and Research Center, Qatar

#### Reviewed by:

Meritxell Espino Guarch, Sidra Medicine, Qatar Mingzhu Zheng, Southeast University, China

\*Correspondence: Christian Schwartz christian.schwartz@uk-erlangen.de

#### Specialty section:

This article was submitted to T Cell Biology, a section of the journal Frontiers in Immunology

Received: 30 April 2022 Accepted: 27 May 2022 Published: 30 June 2022

#### Citation:

Schmidt V, Hogan AE, Fallon PG and Schwartz C (2022) Obesity-Mediated Immune Modulation: One Step Forward, (Th)2 Steps Back. Front. Immunol. 13:932893. doi: 10.3389/fimmu.2022.932893

1

increasing the fat mass, which ultimately causes decreased blood flow and oxygen availability in the adipose tissue, cell death, and mechanical stress on the connective tissue. Together with the increased permeability of the gut and disseminated bacterial products, a low grade chronic inflammatory response develops, which leads to local and peripheral dysregulation of T cell polarization.

The impact of the inflammatory state that obesity elicits throughout the body was recently highlighted by the greater severity and poorer outcome following infection with severe acute respiratory syndrome coronavirus 2 (SARS–CoV–2) in individuals with obesity relative to lean counterparts (3). In the past decade, due to the increasing recognition of obesity and the resulting altered metabolism, research has increasingly focused on the influence of the individual's metabolism as well as cellular metabolism on the immune system and vice versa.

Cellular metabolism is a complex, dynamic process of consuming available nutrients in the cellular environment and producing new metabolites ensuring the proper function of the cell. Depending on the activation status of a cell, their metabolic requirements change. Quiescent cells generate energy using mitochondrial pathways, such as fatty acid oxidation or the tricarboxylic acid (TCA) cycle. These pathways are highly efficient in adenosine triphosphate (ATP) generation and allow for a constant energy production in long-lived cells. Upon cell activation, metabolic reprogramming occurs to adjust to the metabolic requirements. Immune responses evolved to be energetically costly even affecting maintenance programs, such as homeothermy, resulting in physiologic trade-offs (4). Activated cells rapidly produce high amounts of ATP using glycolytic pathways to energize the differentiation and proliferation of cells. With a production of two mole ATP per unit glucose, glycolysis is less efficient in energy production in comparison to mitochondrial catabolic pathways (5). However, this inefficiency is offset as glycolysis can be upregulated faster than mitochondrial metabolic pathways since it does not require mitochondrial growth. Additionally, glycolysis provides biosynthetic intermediates that can further be utilized for ribose synthesis and other essential pathways and therefore represents a dominant metabolic pathway in activated cells (6). Naïve immune cells in the steady state are characterized by a quiescent metabolic profile. Antigen contact or stimulating signals, such as inflammatory cytokines, lead to the activation and subsequent metabolic reprogramming of these cells. For example, type 1 inflammatory macrophages are characterized by energy production via glycolytic pathways, while the antiinflammatory M2 macrophages (M2) predominantly utilize fatty acid oxidation and oxidative phosphorylation. For fatty acid oxidation, fatty acids are first activated to fatty acid acyl-CoA in the cytosol, and then degraded via  $\beta$ -oxidation in the mitochondria producing acyl-CoA, NADH and FADH<sub>2</sub> that can then be used for ATP generation (5). Additionally, it was shown that inhibition of fatty acid oxidation promotes antimicrobial macrophage functions (7).

Cellular functions and metabolism are not only dependent on cell-intrinsic factors, such as activation status, but also on their local environment as well. A change in the composition or quantity of nutrients available within a cellular milieu can lead to differences in the type and magnitude of cellular pathways activated and thereby consequently changes the cell's function (8). Therefore, changes in the metabolic status of an organism, such as the development of obesity or underweight due to over- or undernutrition, respectively, can influence the metabolism of single cells. An oversupply of nutrients leads to the expansion of adipose tissue (AT) and the progression to overweight and obesity. Based on their distinct functions, AT can be divided into brown (BAT) and white (WAT) adipose tissue, as reviewed by Frigolet et al. (9). BAT is a thermogenic tissue characterized by a high number of mitochondria, while WAT functions as lipid storage. In recent years, adipose tissue has gained a new reputation beyond the previous view as a site of fat deposition for energy storage. AT is now considered a secondary immune organ, with an abundance of leukocytes that populate the tissue, which may act as a reservoir of immune cells and mediators during immune responses. Indeed, novel single-cell sequencing data of cells present in human and mouse adipose tissue underpin the differences of immune cells between the AT of lean and obese individuals (10, 11). These findings led to the conclusion that the immune system and the metabolic state of the organism are closely linked and regulate each other - making the AT one of the most important immunometabolic modulators within the body.

This dynamics of the prevailing immune cell repertoires within the AT is exemplified by the marked differences in the cell composition of AT in persons with or without obesity. In individuals with a lean bodyweight, populations of antiinflammatory cells such as regulatory T cells (Treg), T helper (Th) type 2 cells, group 2 innate lymphoid cells (ILC2), M2, and eosinophils dominate the AT milieu and may serve to counteract inflammation (12). Th2 cells are classically regarded as important effector cells that produce the hallmark cytokines IL-4 and IL-13, but also IL-3 and IL-5 that promote basophilia and eosinophilia, respectively. Following activation via the TCR and co-stimulation, in the presence of type 2 polarizing cytokines, naïve T cells differentiate into Th2 cells expressing the master transcription factor GATA3. These type 2 polarizing factors include the presence of IL-4, which activates the STAT6signaling pathway; activation of STAT5-signaling through cytokines, such as IL-2, IL-7 and TSLP; weak TCR signaling strength; certain costimulatory molecules and activation of Jag1/ Notch-interaction (reviewed in (13)). Downstream effects of Th2 cell activation and release of type 2 cytokines include the differentiation of M2 macrophages, basophilia, eosinophilia, B cell antibody isotype class-switching to IgG1 and IgE. This spectrum of type 2 responses is associated primarily with parasitic infections and allergies. Th2 responses also inhibit Th1 responses and vice versa. During obesity on the other hand, infiltration and/or expansion of pro-inflammatory immune cells, such as inflammatory macrophages, cytotoxic CD8<sup>+</sup> T cells and Th1 cells in white adipose tissue leads to a constant low-grade inflammation and consequently to metabolic dysfunctions underpinning the obesity-related metabolic syndrome. Mucosal associated invariant T (MAIT) cells are another subset of T cells implicated in obesity related

dysfunction (14–19). In people with obesity, MAIT cell frequencies are reduced and biased towards a Th17 phenotype and can directly disrupt metabolic processes such as insulinmediated glucose uptake (14, 16). Their contribution to obesityrelated metabolic dysfunction is further supported by data from murine models, where MAIT cell deficiency protects against metabolic dysfunction (17). MAIT cells from persons with obesity (PWO) display an exhausted phenotype (elevated PD-1) and elevated rates of apoptosis (14, 15). Interestingly, chronically stimulated MAIT cells increase their expression of GATA3 and production of Th2 cytokines such as IL-13 (20). But whether chronic activation in obesity drives Th2 like MAIT cells has not been investigated to date.

Moreover, obesity, and indeed malnutrition, not only affect adipose tissue inflammation but extend to peripheral tissues, where they interfere with systemic immune cell activation and functions. While obesity is often associated with a metabolically diseased state, metabolically healthy PWO constitute up to 50% of PWO (21). However, only few studies have addressed the differences in metabolically healthy and unhealthy PWO with regard to immune cell function. Generally, metabolically healthy PWO still harbor less pro-inflammatory cells within the AT, including M1 macrophages as well as Th17 and Th22 cells (22-24), while Th2 cells correlated with insulin sensitivity (25). The mechanisms causing progression from a metabolically healthy obese state to metabolic disease are still under investigation. Remarkably, mitochondria from type 2 diabetes patients appear altered in their function to promote Th17 cytokine production (26). Although much progress has been made over the past decade (27, 28), further studies stratifying metabolically healthy and diseased PWO are still required to elucidate the underlying processes.

The mutual influence of the immune system and metabolism is an important topic and immunometabolism is an arena of current active research. Here, we will review the current understanding of the metabolic regulation of T helper cells and, in particular, Th2 cells in obesity in contrast to expansion of pro-inflammatory Th subsets. Furthermore, we will summarize recent findings on the dysregulation of Th2-biased immune responses during obesity and malnutrition.

# T CELL METABOLISM

T cell metabolism is, as in other immune cells, dependent on the activation status of the cell. As quiescent cells with little need for the *de novo* synthesis of DNA, proteins and lipids, naïve T cells as well as memory T cells produce energy using oxidative phosphorylation (OXPHOS). Effector T cells, on the other hand, switch from mitochondrial pathways to glycolysis due to the rapidly increasing energy consumption following activation. After T cell receptor (TCR) stimulation, expression of genes involved in glycolysis and glutaminolysis are upregulated (29–31). Initially, glycolysis is the dominant metabolic pathway in effector T cells. Transcription factors including c-myc and the hypoxia inducible factor 1 alpha (HIF-1 $\alpha$ ) control upregulation of glucose. Recently, it was shown that the switch from the quiescent state in CD4<sup>+</sup> T cells is mediated

intracellularly *via* Akt and STAT5 signaling that increased both glycolysis and OXPHOS (32). Upon CD28 ligation, expression of the glucose transporter (Glut) 1 and glucose uptake of CD4<sup>+</sup> and CD8<sup>+</sup> effector T cells are increased *via* the PI3K-AKT axis (8, 33, 34). Downstream of PI3K, the kinases mTOR and AMPK as well as the nuclear hormone receptor PPAR $\gamma$  regulate the cell metabolism and differentially influence T cell differentiation. Importantly, Glut1 is essential for CD4<sup>+</sup> but not CD8<sup>+</sup> T cells (35). Without the co-stimulatory signal received through CD28-signaling, T cells enter the anergic state (8). Following activation of tumor necrosis factor receptor-associated factor 6 (TRAF6), effector T cells again start utilizing fatty acid oxidation over glycolysis for energy generation and develop into memory T cells (36).

In essence, glycolysis primarily supports the rapid generation of Th1 and Th17 inflammation and promotes IL-2 and IFN $\gamma$ production. While Th2 cells initially utilize glycolysis, lipid metabolism pathways, such as fatty acid oxidation, synthesis and uptake are upregulated and play an important role in late activation and tissue adaptation (37). Similar to Th2 cells, Tregs mainly generate energy using OXPHOS (38). Importantly, studies investigating differentiation into effector, memory or regulatory T cells, often focused on the use of etomoxir, a drug supposed to inhibit the central enzyme responsible for limitation of long chain fatty acid oxidation, Cpt1a (39–41). However, Raud and colleagues showed that Cpt1a is largely dispensable in this context, implying that etomoxir may exert its functions by alternative mechanisms involving mitochondrial respiration (42).

Environmental factors, such as nutrient availability or adipokines, signal molecules secreted by adipocytes, shape not only macrophage function but also the T cell response *via* modulation of metabolic pathways. The most abundant adipokines secreted by the adipose tissue are leptin and adiponectin. While the plasma concentration of leptin, which dampens hunger, increases proportionally with adipose tissue mass and promotes inflammation, adiponectin counters inflammation and supports the Th2 response (**Figure 1**). After we summarize intrinsic regulation of Th2 cell metabolism, we will discuss extrinsic factors that modulate Th2 cell metabolism and hence, function.

# Intrinsic Regulation mTOR

The serine/threonine kinase mechanistic target of rapamycin (mTOR) forms two distinct complexes depending on the scaffolding protein it associates with. The mTOR complex (mTORC) 1 associates with the regulatory-associated protein of mTOR (RAPTOR), while mTORC2 is associated with the rapamycin-insensitive companion of mTOR (RICTOR). The capability to form two distinct complexes allows mTOR to act as a metabolic switch that can exert different functions. mTORC1 integrates primarily signals that indicate favorable conditions for cell growth. In contrast to the sensing of nutrient-availability by mTORC1, mTORC2 can be stimulated by growth factors and cytokines (43). First evidence that mTOR affects T cell differentiation came from studies, which observed that rapamycin





treatment inhibited T effector cell differentiation, while allowing expansion of Tregs (44–46). mTOR is a downstream target of PI3K and acts as a crucial global regulator of cellular metabolism (47). PI3K is a kinase activated by factors involved in cell proliferation, such as the epidermal growth factor (48), and cellular metabolism, such as leptin (49). This enables T cells to meet the high energy demand of effector responses following activation (29) and upregulate catabolic pathways, such as glycolysis and lipolysis. mTOR coordinates T cell growth, proliferation, metabolism, and differentiation upon shifts in growth factor and nutrient availability. Activation of mTOR is an important condition for the differentiation of naïve T cells into Th1, Th2 or Th17 cells, while inhibition of mTOR results in primary differentiation into Tregs. Absence of mTOR signaling also leads to a decreased proliferation capacity of T cells (44). Activation of mTORC1 induces lipid synthesis pathways and glycolysis (50) and limits autophagy. mTORC2 regulates cytoskeleton organization in addition to induction of glycolysis (51) and lipolysis. Interestingly, during

obesity defective mTOR is observed in NK cells and mucosalassociated invariant T (MAIT) cells: O'Brien and colleagues described an inhibition of mTORC1 in MAIT cells dysregulating their cytokine profile during obesity (18). In contrast, Tobin et al. found an increase of mTORC1 activity in natural killer cells in the adipose tissue of obese children (52). However, these NK cells were reduced in numbers and - despite their increased expression of activation markers - less effective in cell lysis (52). Both, mTORC1 and mTORC2 activate the nuclear hormone receptor PPARy and are activated by IL-4 (53, 54). Signaling of mTORC1 is enhanced by insulin and influences Th1, Th2 and Th17 cell differentiation (44, 55). Hyperactivation of mTORC1 has been shown to limit invariant natural killer T cells (iNKT) (44, 56). NKT cells are a subset of innate-like T lymphocytes and share characteristics of T and natural killer cells. Once activated, NKT cells can release type 1 or type 2 inflammatory cytokines but the mechanisms of recruitment are not well elucidated yet. The role of NKT cells, which can also be potent producers of IL-4, in the context of immune modulation due to metabolic changes of the individual requires further investigation as these cells are often overlooked (57, 58). Th2 differentiation is more sensitive to graded reductions in mTORC1 activity and can develop in the absence of mTORC1 but not in the absence of mTORC2 (59, 60). While mTORC1 responds to signals including growth factors, oxygen and amino acid availability, mTORC2 primarily reacts to other stimuli. However, it has been shown that signaling of mTORC2 responds to nutrient fluctuations and only promotes Th2 but not Th1 or Th17 cell differentiation (59). These observations lead to the hypothesis that Th2 differentiation is subject to a more dynamic regulation through both nutrients and cytokines/growth factors and is less dependent on nutrient-sensing mTORC1 signaling. Upstream of the mTOR-signaling, activation of the PI3K pathway increases mTORC1 activity via AKT, while mTORC2 regulates AKT concentrations and thereby modulates mTORC1 function. Due to the increased nutrient uptake, glucose availability in obesity is increased and subsequently leads to an upregulation of glycolysis and promotion of Th1 differentiation of T cells.

#### AMPK

The AMP activated protein kinase AMPK reacts to the energy level of the cell by sensing the ATP : ADP ratio, regulates glycolysis accordingly and plays a regulating role in energy homeostasis of the cell and T cell activation (61). In a low energy state of the cell, characterized by low ATP levels, AMPK is activated and subsequently inhibits mTORC1 and downregulates glycolytic pathways and Th1 and Th17 inflammation (62, 63). In contrast, AMPK activates mTORC2 and thereby promotes fatty acid catabolism and Th2 and Treg responses (64). In order to generate energy, Th1 cells predominantly upregulate glycolysis, supported by mTORC1 signaling that can directly phosphorylate the Th1 transcription factor Tbet (65), while Th17 cells favor glutaminolysis and regulatory T cells oxidative phosphorylation. The mechanisms of regulation of Treg metabolism are less well elucidated. Compared to naïve T cells, the steady-state activity of mTORC1 is increased and thereby supports the suppressive Treg function partly by inhibiting the mTORC2 pathway in these cells (66). Nevertheless, the predominant metabolic pathways in

regulatory T cells are fatty acid oxidation pathways. HIF1 $\alpha$  reacts to oxygen stress and decreases Glut1 expression in Th17 cells and regulates c-Myc, a transcription factor that is essential for the metabolic reprogramming early after activation, leading to a decreased activation of the cells. HIF1 $\alpha$  signaling is controlled by mTOR. By regulating the Th1 transcription factor Tbet and Eomesodermin (67), a transcription factor amongst others responsible for steering effector T cells into memory T cells, mTOR also determines the T cell fate. Interestingly, Xiong and colleagues found that HIF1 $\alpha$  supports Th2 polarization *via* dendritic cells (DC) priming of naïve T cells (68). Taken together, AMPK inhibits type 1 inflammatory responses by inhibiting mTORC1 and supports type 2 inflammatory responses *via* mTORC2.

#### **PPARs**

The peroxisome proliferator activated receptors (PPAR) are nuclear hormone receptors acting as inducible transcription factors that play a pivotal role in glucose and lipid metabolism. The different isoforms PPAR $\alpha$ , PPAR $\beta/\delta$  and PPAR $\gamma$  are primarily activated by fatty acids and predominantly expressed in different tissues, with PPAR $\alpha$  being found especially in tissues with increased fatty acid oxidation, such as hepatocytes (69), and PPAR $\beta/\delta$  in the gastrointestinal tract (70). PPAR $\gamma$  is considered the master regulator of adipogenesis and highly expressed by adipocytes as well as macrophages and CD4<sup>+</sup> T cells within the adipose tissue (71, 72). PPARy acts as a sensor of the metabolic status of the cell and regulates glucose metabolism and lipid storage, as well as adipocyte differentiation (73). PPARy is prominently expressed in Th2 cells and has been shown to have an inhibiting effect on Th17 inflammation while strongly supporting Th2 and Treg responses in the adipose tissue (74, 75). Activation of PPARy by mTOR leads to an increase of fatty acid uptake, increased plasma concentrations of adiponectin (76) and an induction of adipogenesis, reviewed in (77). PPARy function is supported by the adipokine adiponectin, that is found in higher concentrations in individuals with a lean bodyweight compared to those with obesity. Consequently, a correlation between PPARy activation and weight loss can be observed (78). Importantly, in mice on HFD, IL-4 promoted lipolysis and weight loss through PPARy downregulation in adipocytes (79). PPARy has also been described as an important inducer of regulatory T cell generation (80).

Kopf and coworkers found that PPAR $\gamma$  plays a proinflammatory role in type 2 immunity and is an important mediator for DC-T cell interactions (81). While PPAR $\gamma$  is largely dispensable for the induction of IL-4 production, IL-4 and IL-33 promote the up-regulation of PPAR $\gamma$  in lung resident CD11b<sup>+</sup> DCs, which leads to an enhanced migration to the draining lymph nodes and Th2 priming capacity (81). Micossé et al. defined a phenotypically and functionally distinct Th9 phenotype of T cells that could be a subtype of Th2 cells that are defined by PPAR $\gamma$ expression (82). The cytokines IL-4 and TGF $\beta$  both induce PPAR $\gamma$ and leads to IL-9 production in Th2 cells. Meta-analytic approaches identified a possible contribution of PPAR $\gamma$  to a decreased susceptibility for type two diabetes mellitus in different ancestries (83).

Taken together, these results suggest that a metabolic profile characteristic of activated CD4<sup>+</sup> T cells leads to differentiation into type 1 inflammatory Th1 cells, while the expansion towards Th2 and Treg cells is supported by a metabolic profile characteristic of quiescent cells (Figure 1). As type 1 inflammatory responses are mostly directed to respond to acute viral or bacterial infections a rapid cell proliferation is necessary so that the host can react quickly to the pathogen. Consequently, metabolic pathways that can be upregulated quickly, such as glycolysis, are important in this context. In contrast, type 2 inflammatory responses are directed against continuous inflammatory insults, such as helminth parasite infections or exposure to allergens. In such chronic infections or prolonged exposure and allergen sensitization, no rapid response is necessary and mitochondrial pathways, such as the TCA cycle and oxidative phosphorylation, can be utilized. Additionally, oxidative phosphorylation produces reactive oxygen species that may negatively affect a fast T cell response.

### **Extrinsic Regulation**

Cell metabolism is regulated by various extrinsic factors. Hormones or cytokines influence the metabolic regulation as well as nutrient availability in the cell, such as glucose, by regulating transporter expression in the cell. In obesity, not only the nutritional status of the individual changes, but also concentrations of messenger molecules, such as leptin, adiponectin, resistin, visfatin, and others, which may directly affect Th2 cell function or indirectly inhibit Th2 cells by promoting Th1/Th17-biased inflammation. Adipose tissue produces the hormone leptin and with the expansion of adipose tissue in obesity, the leptin concentration increases. In contrast, the concentration of adiponectin, a hormone regulating the glucose metabolism, decreases following the onset of obesity. Adiponectin is secreted by adipocytes, as well as lymphocytes (84) and has anti-inflammatory properties, such as limiting IFN $\gamma$ production (85) or promoting IL-10 secretion by Tregs (86). An important regulator of the blood glucose concentration is the hormone insulin, produced by the  $\beta$  cells of the pancreas (87). By inducing glucose uptake into the tissue, insulin is an anabolic hormone whose effect on T cells is only started to be researched in detail. The underlying chronic Th1 inflammation in individuals with obesity leads to an increase of proinflammatory cytokines, such as IFNy, while anti-inflammatory cytokines, such as IL-10 are decreased. IL-33 is an alarmin that has been found to induce type 1 as well as type 2 responses. The cytokine produced by endothelial and epithelial cells has been shown to have an protective effect on obesity (88). Aside from an oversupply of nutrients that leads to obesity, undernutrition also has significant influence on cellular metabolism and function. The influence of the hormones leptin, adiponectin and insulin, the cytokine IL-33, as well as undernutrition and fasting on T cell function will be discussed below.

#### Leptin

The pro-inflammatory adipokine leptin is secreted by adipocytes and regulates energy consumption and conversion by regulating food intake as well as glucose metabolism (89). Leptin can pass the blood brain barrier and upon binding the leptin receptor in the brain induce various signaling cascades affecting food intake and energy balance. Impaired crossing of leptin of the blood brain barrier, dysfunctions in the subsequent pathways or decreased sensitivity of the leptin receptor lead to a decreased leptin signaling in the brain. These are possible reasons for leptin resistance and can subsequently lead to a dysregulation in the energy homeostasis (reviewed in (90)). Additionally, leptin has been shown to promote T cell survival and proliferation (91). Leptin has a strong positive association with obesity, is expressed at higher levels in metabolically unhealthy PWO and is negatively associated with being underweight and malnutrition (92, 93). The anorexigenic hormone promotes energy consumption by improving glucose metabolism, controlling the appetite and improving insulin sensitivity (reviewed in (89)). During obesity, the serum leptin concentration increases, which can result in a leptin resistance. The consequential increase in food intake, impaired nutrient absorption and inhibition of lipid and glucose metabolism (94) can lead to a further aggravation of the obese phenotype. Especially in individuals with obesity, leptin leads to an increased secretion on IFNy and suppression of Th2. Naïve T cells lack expression of the leptin receptor, but upregulate it upon activation. In activated CD4<sup>+</sup> T cells, leptin functions as an activator of mTOR (95), upregulates glucose uptake and metabolism and thereby leads to an increased Th1 (96) and Th17 (97) cellular response and a suppression of Treg cells (98). Indeed, another study showed that leptin enhanced Th1 cytokine production, while IL-4 production was decreased (99). Contrarily, Zhang et al. did not find an effect of leptin on Th2 differentiation but a promoting effect on Th2 survival (100). Moreover, Zeng et al. described an enhancing effect of recombinant leptin on ILC2 and Th2 cytokine expression via the PI3K-AKT axis (91, 101). In their study, leptin supported Th1 and Th2 proliferation and survival by activating the (JAK2-STAT3, MAPK and) mTOR pathway. Therefore, they propose the supporting function of leptin on distinct T cell subsets is dependent on the skewing conditions, leading to a leptindependent enhancement of a type 2 response in the context of allergy. Consequently, the increased serum leptin concentration could support T cell proliferation and the type 1 biased inflammatory environment presents skewing conditions to support a Th1 differentiation of T cells. Due to increased serum leptin levels in allergic rhinitis, they propose a possible connection of allergic rhinitis to obesity. Taken together, leptin affects Th2 cells indirectly by supporting the underlying Th1 inflammation in obesity.

#### Adiponectin

The protein hormone adiponectin regulates glucose and lipid metabolism. Mechanisms for this regulation include support of fatty acid oxidation and inhibition of gluconeogenesis *via* an activation of AMPK (102). Adiponectin possesses antiinflammatory properties, such as inducing IL-10 secretion in Tregs (86), and is negatively associated with BMI (103). Furthermore, metabolically healthy PWO often show higher levels of adiponectin similar to the levels of lean people (104). Additionally, two studies found a decrease in glycolysis in activated T cells leading to impaired Th1 or Th17 differentiation (85, 105). Research on the direct effect of adiponectin on Th2 cells is still lacking, but Li et al. could demonstrate a positive effect of adiponectin on IL-4 production that in turn can lead to an increased Th2 response (106). Interestingly, women have more adiponectin in their blood than men. Taking the immunomodulatory effects of adiponectin into account, this could lead to a stronger activation of a type 2 immune response in females (107), which could be relevant to sex differences in obesity and allergic disorders (108). Additionally, as adiponectin promotes insulin sensitivity it supports a potential important player in the context of obesity (109). Taken together, adiponectin is strongly linked to type 2 inflammatory responses, in which its anti-inflammatory effect is further increased (86). Ramos-Ramírez and colleagues demonstrated that adiponectin increased the ability of Treg cells to secrete IL-10 and this effect was further increased in a type 2 inflammatory environment (110). Activation of PPARy, which induces Th2 and Treg responses, leads to increased adiponectin levels in high fat diet (HFD) fed mice, while overexpression of adiponectin also increases the expression of PPAR $\gamma$  (110). In adipose tissue, adiponectin negatively regulates ILC2 function by activation of AMPK and the subsequent suppression of IL-33 signaling (111). Collectively, these data suggest a supporting role of adiponectin for Th2 responses by inhibiting type 1 inflammatory responses and amplifying IL-4 and IL-10 production.

#### Visfatin, Resistin, Apelin

A number of other adipocyte-derived mediators are increased during obesity, such as visfatin (112), resistin (113) and apelin (114). Visfatin leads to activation of T cells and promotes the release of IL-6, IL-1 $\beta$  and TNF but also IL-10 from monocytes leading to CD4<sup>+</sup> T cell activation (115). Resistin, which confers resistance to the action of insulin, is considered a pro-inflammatory molecule activating NF $\kappa$ B, TNF and IL-6 (113). However, it has also been shown that modulation of DCs by resistin leads to enhanced Treg expansion (116). Taken together, the direct effect of all three mediators on Th2 cells has not yet been studied in detail.

#### Growth Differentiation Factor 15 (GDF15)

GDF15, which has emerged as a putative target for treating obesity (117), is a stress-induced hormone produced by a variety of cells – including adipocytes - in the body. Recently, it was shown that the Th2 cytokines IL-4 and IL-13 induce GDF15 production by adipocytes in a STAT6-dependent manner (118). Whether GDF15 directly affects Th2 cells *via* its receptor GFRAL is currently unclear.

#### Insulin

The anabolic hormone insulin plays an important role in energy storage, glucose uptake and synthesis of glycogen and lipids. After food consumption, the insulin concentration increases to subsequently increase the glucose uptake from the blood stream into the cells *via* Glut4 (87). In obesity, this increase of insulin concentration is abated and the insulin sensitivity decreases, leading to higher blood glucose levels in the individual. Importantly, metabolically healthy PWO show greater insulin sensitivity than metabolically unhealthy PWO, whereas they are more insulin resistant than metabolically health lean persons (21). Upon activation, T cells begin to express the insulin receptor but contrarily to the anabolic function of insulin within tissue, in T cells insulin signaling supports T cell proliferation and effector function (119). Additionally, insulin influences the differentiation of regulatory T cells by inducing PPAR $\gamma$  (80). Li et al. described an insulin dependent pathway that activates HIF1 $\alpha$  and subsequently induces PPARy (80). Additionally, Jeschke and colleagues reported an anti-inflammatory role of insulin after severe trauma by decreasing type 1 and increasing type 2 inflammatory responses (120). As a possible mode of action, they propose an indirect effect of insulin by reducing of the blood glucose concentration and consequently decreasing glycolysis in effector T cells. Due to glycolysis being the prominent pathway for energy generation in type 1 inflammatory cells, this type of immune response is more severely inhibited. Another mode of action of insulin could be the upregulation of Glut1 on T cells and subsequently increasing the glucose uptake into the cells. Following the increased glucose availability, T cells are steered towards an upregulation of glycolysis and Th1 differentiation. By supporting Th1 differentiation of T cells, insulin could indirectly inhibit Th2 differentiation. Obesity and related pathologies such as type 2 diabetes mellitus (T2DM) increase the blood glucose concentration. Here, insulin resistance that characterizes T2DM could skew T cell differentiation to a Th1 phenotype in a graded manner by a gradual increase of blood glucose concentration and subsequently the nutrients for T cells to utilize glycolysis. Additionally, a strong body of literature shows an association of IL-17 signaling with insulin resistance and the development of obesity and T2DM. Nicholas and colleagues recently described a positive association of Th17 responses with T2DM (121), confirming the positive association of increased Th17 responses during obesity shown by Fabbrini et al. (122). IL-17 directly affects adipogenesis and glucose metabolism as well as impairing insulin sensitivity (123, 124). Thus, interfering with Th cell subset polarization may be a novel therapeutic approach to improve T2DM.

#### IL-33

The alarmin IL-33 is a versatile interleukin that can induce Th1 responses as well as Th2 responses. IL-33 supports IFNy secretion of CD8<sup>+</sup> T cells and NK cells (125, 126) as well as neutrophil activation (127), thereby supporting characteristic effector functions of a type 1 inflammatory immune response. However, IL-33 administration can also induce marked type 2 inflammation, with eosinophilia, increased production of the interleukins IL-4, IL-5 and IL-13 as well as mucus production and epithelium remodeling (128). Th2 cells, as well as ILC2, Treg cells and M2 macrophages, express the IL-33 receptor ST2 and are activated by IL-33 ligation to produce type 2 inflammatory cytokines (128). These effects are characteristic for inducing type 2 immune responses. Additionally, IL-33 has been shown to be protective in obesity. In obese mice, administration of IL-33 restores immunological and metabolic profiles of adipose tissue and exerts this effect through activation of adipose tissue ILC2. Treatment of obese mice with  $\omega_1$ , a helminth derived RNAse

that induces type 2 inflammatory responses, induced a release of IL-33 from adipocytes that subsequently can further support Th2 responses (129). Mahalakõiv et al. also described a protective effect of IL-33, produced by stromal cells, in diet-induced obesity in mice (130). In a study in humans Tang et al. found elevated levels of IL-33 in Chinese adults with a positive correlation between IL-33 and risk factors for metabolic syndrome (131).

#### IL-25, TSLP

In recent years, also other alarmins, such as IL-25 and thymic stromal lymphopoietin (TSLP), have been investigated and, for both, a supporting role on the Th2 response and protective effect against obesity could be shown. While IL-25 mainly protects from obesity by stimulating M2 macrophages and inducing lipolysis (132-134), TSLP acts trough activation of DCs to prime naïve T cells for differentiation into Th2 cells (135). Treatment of mice on HFD with TSLP protects from weight gain and glucose intolerance, and it further induces the loss of white AT mediated by T cells that upregulate sebum secretion (136). Although increased IL-4 production was observed in TSLP-treated mice, the polarization state of the CD4<sup>+</sup> T cells was not assessed (136). Thus, there could be a critical role for Th2 cell induction through TSLP-activation promoting an unexpected link to skin barrier maintenance and sebum secretion during obesity.

#### Fasting

While obesity is a well-known and acknowledged health burden, research on the effects of undernutrition on immune cell metabolism and function is sparse. Short-term effects of undernutrition can be assessed through fasting-mimicking diets or intermittent fasting, which has a beneficial effect on the host organisms (reviewed in (137)) and hematopoietic stem cells (138), B cells (139), monocytes (140) and memory T cells (141). Collins and colleagues described a collapse of circulating memory T cells after dietary restriction and an accumulation in bone marrow (141). This effect was even further enhanced by glucocorticoids, steroid hormones that support gluconeogenesis. The accumulation of memory T cells in the bone marrow protected the cells during dietary restriction. Due to the decrease in blood glucose levels following a period of fasting, AMPK is activated and subsequently, mTOR is inhibited to reduce the energy consumption of the cells. Consequently, processes such as autophagy are increased, while glucose uptake and glycolysis are decreased (142). Due to the increased utilization of energy conserving pathways, fasting has especially anti-inflammatory effects, while highly energy consuming pro-inflammatory responses are downregulated (139, 143). Consistent with this, Lenehan et al. found a decrease especially of Th1 and Th17 inflammation in wasting AT of tumor-bearing mice, while the Th2 response were maintained (144). Collectively, fasting has a supporting effect on Th2 responses, while the more energy consuming type 1 inflammatory responses are inhibited.

Collectively, studies indicate that Th2 cells are affected by their nutritional environment (**Figure 1**). While it is known that factors such as mTORC1 activation and leptin signaling inhibit Th2 cells, while AMPK and adiponectin signaling activate or promote Th2 cell function, more research is required to pinpoint the evident differences between the T helper cell subsets. Studies, mainly performed in mice, using different models of obesity and starvation as well as disease models highlight how Th2 cells are influenced by the hosts' metabolic status.

# TH2 CELL FUNCTION DURING OBESITY AND MALNUTRITION

The regulation of CD4<sup>+</sup> T cell metabolism on a cellular level is an important factor for the activation, differentiation and polarization of the cells. The impact of metabolic regulation also affects immune responses both locally within adipose tissue (**Figure 2**) and in the periphery (**Figure 3**). During obesity, T cells live in a nutrient-rich environment, in which fatty acids are abundantly present, whereas during malnutrition glucose availability is decreased. However, due to the systemic immune and hormonal alterations in an obese or fasted state it is difficult to disentangle effects of nutrients and cytokine/hormone activities and this area is only beginning to be studied in detail. Therefore, we will summarize the currently available literature on local – within AT- and peripheral Th2 cell function during obesity and fasting (**Figure 3**).

## Local Effect on Adipose Tissue Th2 Cell Function

Within the healthy adipose tissue, an anti-inflammatory environment predominates. Several cell types that are associated with a classical type 2 response have been identified in the adipose tissue, including ILC2, eosinophils and M2-polarized macrophages (145, 146). However, data concerning Th2 cells in adipose tissue are limited.

During obesity, Th1 cells increase within the adipose tissue, which could have an inhibitory effect on Th2 cells through the production of IFNy (147, 148). IFNy induces SOCS1 that inhibits IL-4R signaling (149, 150). Furthermore, the Th1 transcription factor Tbet directly interferes with GATA3 (151). Interestingly, the absolute number of Th2 cells remains relatively constant (152), suggesting that cell proliferation is inhibited. Other mechanisms than the modulators, including leptin and adiponectin, outlined above, may also play important roles in the suppression of Th2 cells during obesity. We have recently shown that PD-L1-upregulation occurs during late stages of obesity in mice and also in human visceral adipose tissue of obese individuals (153), which may block further Th2 proliferation via PD-1 engagement. Similarly, regulation of innate effectors, such as ILC2, through cellular interactions shape adaptive T cell responses (154). In essence, obesity disrupts the homeostatic environment in favor of a proinflammatory Th1 bias, the step forwards, while Th2 cells cause a step back from obesity (Figure 2).

# Systemic Effect of Obesity on Th2 Cell Function

In human subcutaneous and visceral adipose tissue, it was shown that Th2 cells negatively correlate with systemic inflammation and insulin resistance, suggesting Th2 cells have a protective role (25). In







FIGURE 3 | Effects of nutritional dysregulation on peripheral immune responses. Both obesity and malnutrition lead to dysregulation of Th2 cells, which can affect many different inflammatory conditions ranging from allergies to infections and autoimmunity.

mice, transfer of CD4<sup>+</sup> T cells into obese Rag1<sup>-/-</sup> recipients led to the differentiation towards Th2 cells and reversed enhanced weight gain and insulin resistance. These effects were further shown to be dependent on STAT6 (152). In mice on HFD, Th2 cell frequency significantly decreased in the adipose tissue as mice become obese (155). In human adipose tissue, Th2 cells inversely correlated with plasma CRP concentration, a marker indicating systemic inflammation (25). Inflammation has been linked to insulin resistance since the early 1990s, when adipose tissue TNF was shown to be increased during obesity and neutralization of TNF improved peripheral glucose uptake (156). Additionally, IFNy and IL-1 $\beta$  also modulate insulin signaling (reviewed in (157, 158)), whereas IL-4 and IL-10 were shown to promote insulin sensitivity (159). Given that T cells influence M1/M2-polarization, it is likely that T cell-derived cytokines play an important role for the generation of pro-inflammatory M1-polarized macrophages during obesity (160). Therefore, the decreased production of IL-4 by Th2 cells and IL-10 from Treg cells within the adipose tissue leads to a relative decline of anti-inflammatory M2 macrophages, while the concurrent increase of Th1 cells promotes TNFexpressing M1 macrophages and hence, insulin resistance. Importantly, frequency of Th2 cells in adipose tissue is associated with systemic insulin resistance (152).

#### Asthma

Obesity is associated with an increased risk to develop asthma – a chronic inflammatory disease of the lung. Asthma has a highly heterogeneous pathogenesis and a continuum of endotypes: from a Th2-driven (type 2) to a non-Th2-driven endotype (161). Th2-driven asthma includes early-onset allergic asthma, late onset eosinophilic asthma and exercise-induced asthma, and is characterized by classical type-2 associated factors, including Th2 cells, eosinophils and IgE (162). Non-Th2 asthma includes neutrophilic asthma and characterized by Th1, Th17 cells and neutrophils. However, Th2-driven asthma has been described in children and adults with obesity (163, 164) and asthma-obesity endotypes may be associated with age of onset (165).

Dysregulation of T cells may also be an important factor during obesity-induced asthma. Here, the expansion of Th1 cells contributes to the non-Th2-driven asthma pathology. As outlined above, Th2 cells decline with active suppression of Th2 cell differentiation and proliferation by Th1 cell IFNy and Tbet to counter-regulate Th2 responses. In contrast, it was also found that leptin increased Th2 cells during airway hyperreactivity in mice, in part through MAPK, STAT3 and ER stress response (100, 166). Similarly, in mice on HFD that are immunized and challenged with Ovalbumin (OVA) allergen to induce allergic asthma-like lung inflammation, a mixed inflammatory response developed with increased levels of TNF, IL-5 and IL-10, ultimately leading to an eosinophil-dominated allergic airway response (167). Further studies, have implicated ILC2 and ILC3 (168), or mast cells (169) in asthma pathogenesis, with the latter promoting a delayed Th1, Th2, Th17 profile. A recent study found that autophagy may also be involved in exacerbation of eosinophilic airway inflammation as mice deficient in Atg5 on HFD had increased Th2 cell numbers in

the inflamed lung (170). Interestingly, a 12-week HFD in female mice was protective against airway hyperreactivity through pulmonary DCs recruitment and decreased Th1/17 responses while leaving Th2 cells intact (171). Whereas mice do not develop asthma per se, only aspects of human asthma endotypes can be modelled and should be taken into consideration (161). Taken together, while human obesityassociated asthma is more Th1-driven, in mice – and in certain cases also in humans – a contribution of type 2 associated immune cells including Th2 cells may be important.

#### **Skin Inflammation**

Inflammatory skin diseases are widespread and have many etiologies. The two most important chronic diseases of the skin are atopic dermatitis (AD) and psoriasis, both of which have been associated with obesity. While AD is classically regarded as a Th2-biased chronic allergic inflammatory skin disease with contribution of eosinophils, basophils, mast cells and ILC2 to pathogenicity, psoriasis is driven by a Th1/Th17-biased immune response and includes neutrophils and ILC3 (172). Both AD and psoriasis have been associated with obesity (reviewed in (173–176)). However, Th2 cells are not directly involved in psoriasis, but a case could be made for diverting the Th1/Th17-response towards a less pathogenic Th2 response by treatment with IL-4 (177).

The overall risk for AD is only minimally increased for adults with obesity (OR=1.08) according to a recent study of patients in the UK (178) but AD is a disease that usually has an early onset during the first years of life. Indeed, several studies have linked childhood obesity to AD (173, 174, 179, 180). However, the pathomechanisms linking these diseases are still not fully understood. Th2 cells seem to be involved, as PPAR $\gamma$  in Th cells drives obesity-associated Th2-immunopathology in severe AD (preprint doi: https://doi.org/10.1101/825836). Furthermore, a recent study in atopic children observed an abnormal blood profile with higher cholesterol and triglycerides (181) – which are also increased during obesity. Whether the increased fatty acids directly influence Th2 polarization *via* PPAR signaling remains unclear (182).

An important factor promoting the development of AD is the breakdown of the skin barrier (172, 183), which can be measured by trans-epidermal water loss (TEWL). Importantly, TEWL is significantly increased in people with obesity (184, 185). The cardinal Th2 cytokines IL-4 and IL-13 may contribute to this breakdown as their homeostatic levels maintain skin barrier integrity (186). Thus, the pro-inflammatory T cell bias during obesity may interfere with homeostatic function or the production of IFN $\gamma$ , which is also found during chronic AD lesions, may alter skin integrity to disrupt barrier function through alteration of the fatty acid composition of ceramide (187) or downregulation of tight junction function (188). More studies are required to link obesity and the development of a pathogenic Th2 response in the skin.

#### Wound Healing

Obese patients are at risk of developing complications after surgery including wound infections and wound separation

(reviewed in (189)), which can be consequences of the impaired wound healing in obese patients (reviewed in Anderson 2015). Th2 cells are an important factor in mediating wound repair by induction of M2 macrophages and eosinophils that promote angiogenesis, myofibroblast activation and deposition of extracellular matrix, as well as by inhibition of proinflammatory Th1 and Th17 responses (190–192). Whether obesity directly interferes with the capacity of Th2 cells to promote wound healing is currently under investigation.

#### Rheumatoid Arthritis (RA)

A hallmark of chronic synovial inflammation is the sustained influx of immune cells into the joints. Innate effector cells, including neutrophils and pro-inflammatory macrophages, and effector T and B cells promote synovial inflammation. The anti-inflammatory cytokines IL-4, IL-13, IL-10 and TGFβ are downregulated during established RA (193). However, during early RA IL-4 and IL-13 are upregulated in synovial fluid suggesting an early modulatory or compensatory role, also highlighted by their anti-arthritic properties (194-197). In a mouse model of collagen-induced arthritis, Th2 cells were shown to be increased after repeated IL-33 administration and promoted amelioration of disease (198). People with obesity that additionally have RA, also have poorer quality of life and show higher disease activity (199). To date it is unclear how Th2 may be affected in RA patients with obesity. It is tempting to speculate that the pro-inflammatory state in obese individuals negatively affects RA pathogenesis and disease activity, which is ameliorated through the induction of Th2 cells.

#### Eosinophilic Oesophagitis (EoE)

EoE is an emerging disease, which is characterized by a deregulated type 2 response with high numbers of eosinophils in the oesophageal epithelial layer, leading to clinical symptoms that include dysphagia, feeding dysfunction and vomiting. In a model of OVA-induced EoE, obesity increased Th2 cytokine expression and reduced regulatory T cell responses (200). The altered immune response was not only limited to the oesophageal tissue but expanded to the spleen and mediastinal lymph nodes. Whereas Th2 cells were increased, OVA-specific IgE responses were reduced in obese allergic mice (200). Leptin levels were increased in obese allergic mice, which may affect T cell polarization as outlined above, but more experimentation is required to formally address this in the context of EoE. Whether the exacerbation of Th2 responses during obesity may be involved in other allergic settings, such as food allergies, and how these processes can be therapeutically exploited must be the focus of future studies.

#### Infectious Diseases

Obesity is associated with increased risk to contract infectious diseases including skin infections and infections of the lower respiratory tract (201). Importantly, differences between men and women were also noted. Similarly, obesity affects the outcome of infections (202). During the SARS-CoV-2-pandemic it also became evident that obesity is an important risk factor for severe disease and mortality (203). Remarkably, Th2-associated cytokines showed an upward trend over the

severe courses of Covid-19 (204). Similarly, IL-13 was found to be a driver of severe disease (205) promoting lung damage with participation of Th2 cells (206). However, it is currently unclear to what extent Th2 cells and the associated cytokines contribute to pathology in severe Covid-19.

In conclusion, obesity is an important factor regulating Th2 functionality extending beyond local tissue responses. However, to date studies investigating Th2 cells in other inflammatory settings are scarce and such studies may reveal unexpected novel regulatory functions of Th2 cells.

# Systemic Effect of Fasting/Malnutrition on Th2 Cells

Obesity is a consequence of overnutrition and its high prevalence in industrialized countries led to a heightened research interest in this area, whereas the study of the effects of malnutrition on the immune system is less widespread. Immunity in the context of chronic malnutrition is difficult to study and according to a 2014 systematic review yields varying results (207). However, starvation and cachexia are linked to immune dysfunction in humans and mice (208). After two days of starvation, splenic CD4<sup>+</sup> T cells in mice declined by 50% (209), which can also be observed in malnourished children (210). Importantly, not only numbers of Th cells were reduced in the blood of starved mice and also malnourished children, but their capacity to produce IFNy and IL-2 (209, 211). Instead, malnourished children had increased concentrations of IL-4 and IL-10 in their blood (212). In the absence of glucose, activation of Th cells via Glut1 is impaired (8), but whether this also impairs Th2 cell function, which seems to be intact although glycolysis cannot be used (213), remains to be determined. Interestingly, a recent study showed that during cancer-associated adipose tissue wasting the Th2-environment in adipose tissue is maintained both in mice and humans (144).

Intermittent fasting, which has become a widespread strategy to lose weight, is another approach to investigate the effects of short-term caloric restriction on immune cells. In a mouse model of eosinophilic asthma - induced by intranasal administration of IL-33 and OVA - a two-day fasting period reduced the accumulation of Th2 cells, IL-13, generation of OVA-specific IgG1, and eosinophils in the lungs compared to non-fasted mice (142). Whether this effect is solely attributable to Th2 cells or mediated by ILC2s remains to be determined. During a model of autoimmunity, fasting reduced accumulation of CD4<sup>+</sup> T cells and IFN $\gamma$ -producing cells, while it increased IL-17A production (214). Taken together, while Th2 cells, and indeed other T helper subsets, are affected by malnutrition well-controlled studies that address the functional impact on immunity are required.

# Helminth-Mediated Induction of Th2 Cells During Obesity

Taking into account that Th2 cells are beneficial for the homeostatic environment in adipose tissue of lean individuals, it is only logical that the question arises whether induction of Th2 cells during obesity has an ameliorating effect. Several studies have addressed this question using helminth infections to evoke a Th2 and regulatory state that may impact the severity of unrelated inflammatory conditions and disease states. There are extensive studies in mouse models, as well as human clinical trials using live helminth infections for the treatment of several inflammatory diseases (reviewed in (215)).

The impact of the induction of Th2 cells by helminth infection on the low-grade chronic inflammation during obesity has been addressed in a number of studies. Studies in helminth-infected humans observed improved metabolic health (216-223). In mice, acute helminth infection with Nippostrongylus brasiliensis also improved metabolic health and decreased weight gain (146, 224). Hussaarts and colleagues revealed that chronic Schistosoma mansoni infection (>12 weeks) significantly ameliorated diet-induced obesity in mice compared to non-infected controls (225). Unsurprisingly, infection with the gastrointestinal helminth Heligmosomoides polygyrus also led to ameliorated weight gain and improved metabolic function (226). Su et al. brought more complexity to these findings as they show that Th2 cells were critical in the regulation of the helminthinduced changes to the microbiome that subsequently affected nutrient uptake and weight gain (227). Studies with Strongyloides stercoralis-infected patients that were obese revealed that they had also higher type 2-associated cytokines in their circulation (228). Interestingly, these changes were reversed upon antihelminthic treatment to remove the worm infection.

While these infection models rely on infection with live helminths, the side effects may outweigh the benefits and thus single helminth-derived products may have a better safety profile for human application. Omega-1, derived from *S. mansoni* eggs, that is a potent inducer of Th2 cells in mice, was shown to ameliorate HFD-induced obesity (129, 229). More recently, ES-62, a glycoprotein of *Acanthocheilonema vitae*, was also shown to improve metabolic health (230). Importantly, small-molecule analogues still possess anti-inflammatory properties (231). With regard to the variety of helminth-derived products and their immunomodulatory functions (reviewed in (232)), many of these may be able to modulate Th2 cell function during obesity and thus may improve diabetes or even counteract weight gain.

In contrast to these hypothetical experimental interventions, there is also potential to target T cell metabolism using 2-deoxyglucose (2-DG), which inhibits glycolysis and blocks CD4<sup>+</sup> T cell proliferation, with 2-DG used in numerous clinical trials focusing mostly on cancer but to date none have investigated a possible impact on AT T cells and obesity. Leptin has also been used in clinical trials exploring therapeutic potential in obesity (233). It is clear that further insight on the regulation of different T cell subsets

### REFERENCES

- Cautivo KM, Molofsky AB. Regulation of Metabolic Health and Adipose Tissue Function by Group 2 Innate Lymphoid Cells. *Eur J Immunol* (2016) 46(6):1315–25. doi: 10.1002/eji.201545562
- Stolarczyk E. Adipose Tissue Inflammation in Obesity: A Metabolic or Immune Response? *Curr Opin Pharmacol* (2017) 37:35–40. doi: 10.1016/ j.coph.2017.08.006
- Kompaniyets L, Goodman AB, Belay B, Freedman DS, Sucosky MS, Lange SJ, et al. Body Mass Index and Risk for Covid-19-Related Hospitalization, Intensive Care Unit Admission, Invasive Mechanical Ventilation, and Death

in the context of obesity and, or, the metabolic syndrome are required to inform on the development of new T cell mediated therapies for obesity.

# CONCLUSION

In recent years, the importance of adipose tissue as an immune organ has become increasingly appreciated. It has been shown that changes in the metabolic status of an individual subsequently lead to changes in the immune balance. Overall, obesity primarily impairs Th2 responses, while reinforcing type 1 inflammatory responses. Starvation and malnutrition, on the other hand lead to a type 2 biased immune response. However, more research on the metabolic pathways promoting T cell polarization towards Th2 is still required, especially with regard to the temporal regulation of the utilization of glycolysis and OXPHOS during different immune responses. Further investigation into the importance of IL-4-producing iNKT cells will also help to develop further therapeutic options, such as treatment with  $\alpha$ GalCer for the activation of iNKT cells (234), IL-25, TSLP or IL-33 to activate ILC2 and Th2 cells (136, 235-237), or IL-4 or helminth products to promote Th2 differentiation (238). In the ongoing immunometabolism dance, it is one step forward and (Th)2 steps back.

## AUTHOR CONTRIBUTIONS

VS, AH, PF, and CS wrote the manuscript. PF and CS conceived the manuscript. All authors contributed to the article and approved the submitted version.

### FUNDING

PF was supported by the National Children's Research Centre and Science Foundation Ireland (10/IN.1/B3004). CS is supported by the Interdisciplinary Center for Clinical Research (IZKF) at the University Hospital of the University of Erlangen-Nuremberg (J79), the Else Kröner-Fresenius-Stiftung (2019\_A181) and the Federal Ministry of Education and Research (BMBF 01KI2109).

- United States, March-December 2020. MMWR Morb Mortal Wkly Rep (2021) 70(10):355-61. doi: 10.15585/mmwr.mm7010e4

- Ganeshan K, Nikkanen J, Man K, Leong YA, Sogawa Y, Maschek JA, et al. Energetic Trade-Offs and Hypometabolic States Promote Disease Tolerance. *Cell* (2019) 177(2):399–413.e12. doi: 10.1016/j.cell.2019.01.050
- O'Neill LA, Kishton RJ, Rathmell J. A Guide to Immunometabolism for Immunologists. Nat Rev Immunol (2016) 16(9):553-65. doi: 10.1038/ nri.2016.70
- Lochner M, Berod L, Sparwasser T. Fatty Acid Metabolism in the Regulation of T Cell Function. *Trends Immunol* (2015) 36(2):81–91. doi: 10.1016/ j.it.2014.12.005

- Chandra P, He L, Zimmerman M, Yang G, Koster S, Ouimet M, et al. Inhibition of Fatty Acid Oxidation Promotes Macrophage Control of Mycobacterium Tuberculosis. *mBio* (2020) 11(4):e01139-20. doi: 10.1128/ mBio.01139-20
- Jacobs SR, Herman CE, Maciver NJ, Wofford JA, Wieman HL, Hammen JJ, et al. Glucose Uptake Is Limiting in T Cell Activation and Requires Cd28-Mediated Akt-Dependent and Independent Pathways. J Immunol (2008) 180(7):4476–86. doi: 10.4049/jimmunol.180.7.4476
- Frigolet ME, Gutierrez-Aguilar R. The Colors of Adipose Tissue. Gac Med Mex (2020) 156(2):142–9. doi: 10.24875/GMM.M20000356
- Emont MP, Jacobs C, Essene AL, Pant D, Tenen D, Colleluori G, et al. A Single-Cell Atlas of Human and Mouse White Adipose Tissue. *Nature* (2022) 603(7903):926–33. doi: 10.1038/s41586-022-04518-2
- Hildreth AD, Ma F, Wong YY, Sun R, Pellegrini M, O'Sullivan TE. Single-Cell Sequencing of Human White Adipose Tissue Identifies New Cell States in Health and Obesity. *Nat Immunol* (2021) 22(5):639–53. doi: 10.1038/ s41590-021-00922-4
- Kohlgruber A, Lynch L. Adipose Tissue Inflammation in the Pathogenesis of Type 2 Diabetes. *Curr Diabetes Rep* (2015) 15(11):92. doi: 10.1007/s11892-015-0670-x
- Zhu J. T Helper 2 (Th2) Cell Differentiation, Type 2 Innate Lymphoid Cell (Ilc2) Development and Regulation of Interleukin-4 (Il-4) and Il-13 Production. *Cytokine* (2015) 75(1):14–24. doi: 10.1016/j.cyto.2015.05.010
- Magalhaes I, Pingris K, Poitou C, Bessoles S, Venteclef N, Kiaf B, et al. Mucosal-Associated Invariant T Cell Alterations in Obese and Type 2 Diabetic Patients. J Clin Invest (2015) 125(4):1752–62. doi: 10.1172/ JCI78941
- Carolan E, Tobin LM, Mangan BA, Corrigan M, Gaoatswe G, Byrne G, et al. Altered Distribution and Increased Il-17 Production by Mucosal-Associated Invariant T Cells in Adult and Childhood Obesity. *J Immunol* (2015) 194 (12):5775–80. doi: 10.4049/jimmunol.1402945
- Bergin R, Kinlen D, Kedia-Mehta N, Hayes E, Cassidy FC, Cody D, et al. Mucosal-Associated Invariant T Cells Are Associated With Insulin Resistance in Childhood Obesity, and Disrupt Insulin Signalling *Via* Il-17. *Diabetologia* (2022) 65(6):10122–7. doi: 10.1007/s00125-022-05682-w
- Toubal A, Kiaf B, Beaudoin L, Cagninacci L, Rhimi M, Fruchet B, et al. Mucosal-Associated Invariant T Cells Promote Inflammation and Intestinal Dysbiosis Leading to Metabolic Dysfunction During Obesity. *Nat Commun* (2020) 11(1):3755. doi: 10.1038/s41467-020-17307-0
- O'Brien A, Loftus RM, Pisarska MM, Tobin LM, Bergin R, Wood NAW, et al. Obesity Reduces Mtorc1 Activity in Mucosal-Associated Invariant T Cells, Driving Defective Metabolic and Functional Responses. *J Immunol* (2019) 202(12):3404–11. doi: 10.4049/jimmunol.1801600
- Brien AO, Kedia-Mehta N, Tobin L, Veerapen N, Besra GS, Shea DO, et al. Targeting Mitochondrial Dysfunction in Mait Cells Limits Il-17 Production in Obesity. *Cell Mol Immunol* (2020) 17(11):1193–5. doi: 10.1038/s41423-020-0375-1
- Kelly J, Minoda Y, Meredith T, Cameron G, Philipp MS, Pellicci DG, et al. Chronically Stimulated Human Mait Cells Are Unexpectedly Potent Il-13 Producers. *Immunol Cell Biol* (2019) 97(8):689–99. doi: 10.1111/imcb.12281
- Smith GI, Mittendorfer B, Klein S. Metabolically Healthy Obesity: Facts and Fantasies. J Clin Invest (2019) 129(10):3978–89. doi: 10.1172/JCI129186
- 22. Esser N, L'Homme L, De Roover A, Kohnen L, Scheen AJ, Moutschen M, et al. Obesity Phenotype Is Related to Nlrp3 Inflammasome Activity and Immunological Profile of Visceral Adipose Tissue. *Diabetologia* (2013) 56 (11):2487–97. doi: 10.1007/s00125-013-3023-9
- Wentworth JM, Naselli G, Brown WA, Doyle L, Phipson B, Smyth GK, et al. Pro-Inflammatory Cd11c+Cd206+ Adipose Tissue Macrophages Are Associated With Insulin Resistance in Human Obesity. *Diabetes* (2010) 59 (7):1648–56. doi: 10.2337/db09-0287
- Pandolfi JB, Ferraro AA, Sananez I, Gancedo MC, Baz P, Billordo LA, et al. Atp-Induced Inflammation Drives Tissue-Resident Th17 Cells in Metabolically Unhealthy Obesity. J Immunol (2016) 196(8):3287–96. doi: 10.4049/jimmunol.1502506
- McLaughlin T, Liu LF, Lamendola C, Shen L, Morton J, Rivas H, et al. T-Cell Profile in Adipose Tissue Is Associated With Insulin Resistance and Systemic Inflammation in Humans. *Arterioscler Thromb Vasc Biol* (2014) 34 (12):2637–43. doi: 10.1161/ATVBAHA.114.304636

- Nicholas DA, Proctor EA, Agrawal M, Belkina AC, Van Nostrand SC, Panneerseelan-Bharath L, et al. Fatty Acid Metabolites Combine With Reduced Beta Oxidation to Activate Th17 Inflammation in Human Type 2 Diabetes. *Cell Metab* (2019) 30(3):447–61.e5. doi: 10.1016/ j.cmet.2019.07.004
- Agrawal M, Kern PA, Nikolajczyk BS. The Immune System in Obesity: Developing Paradigms Amidst Inconvenient Truths. *Curr Diabetes Rep* (2017) 17(10):87. doi: 10.1007/s11892-017-0917-9
- SantaCruz-Calvo S, Bharath L, Pugh G, SantaCruz-Calvo L, Lenin RR, Lutshumba J, et al. Adaptive Immune Cells Shape Obesity-Associated Type 2 Diabetes Mellitus and Less Prominent Comorbidities. *Nat Rev* Endocrinol (2022) 18(1):23–42. doi: 10.1038/s41574-021-00575-1
- 29. Macintyre AN, Gerriets VA, Nichols AG, Michalek RD, Rudolph MC, Deoliveira D, et al. The Glucose Transporter Glut1 Is Selectively Essential for Cd4 T Cell Activation and Effector Function. *Cell Metab* (2014) 20 (1):61–72. doi: 10.1016/j.cmet.2014.05.004
- Wang R, Dillon CP, Shi LZ, Milasta S, Carter R, Finkelstein D, et al. The Transcription Factor Myc Controls Metabolic Reprogramming Upon T Lymphocyte Activation. *Immunity* (2011) 35(6):871–82. doi: 10.1016/ j.immuni.2011.09.021
- Michalek RD, Gerriets VA, Jacobs SR, Macintyre AN, MacIver NJ, Mason EF, et al. Cutting Edge: Distinct Glycolytic and Lipid Oxidative Metabolic Programs Are Essential for Effector and Regulatory Cd4+ T Cell Subsets. *J Immunol* (2011) 186(6):3299–303. doi: 10.4049/jimmunol.1003613
- Jones N, Vincent EE, Cronin JG, Panetti S, Chambers M, Holm SR, et al. Akt and Stat5 Mediate Naive Human Cd4+ T-Cell Early Metabolic Response to Tcr Stimulation. *Nat Commun* (2019) 10(1):2042. doi: 10.1038/s41467-019-10023-4
- Frauwirth KA, Riley JL, Harris MH, Parry RV, Rathmell JC, David RP, et al. The Cd28 Signaling Pathway Regulates Glucose Metabolism. *Immunity* (2002) 16:769–77. doi: 10.1016/S1074-7613(02)00323-0
- Wieman HL, Wofford JA, Rathmell JC. Cytokine Stimulation Promotes Glucose Uptake Via Phosphatidylinositol-3 Kinase/Akt Regulation of Glut1 Activity and Trafficking. Mol Biol Cell (2007) 18(4):1437–46. doi: 10.1091/ mbc.e06-07-0593
- 35. Cretenet G, Clerc I, Matias M, Loisel S, Craveiro M, Oburoglu L, et al. Cell Surface Glut1 Levels Distinguish Human Cd4 and Cd8 T Lymphocyte Subsets With Distinct Effector Functions. *Sci Rep* (2016) 6:24129. doi: 10.1038/srep24129
- Pearce EL, Walsh MC, Cejas PJ, Harms GM, Shen H, Wang LS, et al. Enhancing Cd8 T-Cell Memory by Modulating Fatty Acid Metabolism. *Nature* (2009) 460(7251):103–7. doi: 10.1038/nature08097
- 37. Tibbitt CA, Stark JM, Martens L, Ma J, Mold JE, Deswarte K, et al. Single-Cell Rna Sequencing of the T Helper Cell Response to House Dust Mites Defines a Distinct Gene Expression Signature in Airway Th2 Cells. *Immunity* (2019) 51(1):169–84.e5. doi: 10.1016/j.immuni.2019.05.014
- Almeida L, Lochner M, Berod L, Sparwasser T. Metabolic Pathways in T Cell Activation and Lineage Differentiation. *Semin Immunol* (2016) 28(5):514– 24. doi: 10.1016/j.smim.2016.10.009
- Declercq PE, Falck JR, Kuwajima M, Tyminski H, Foster DW, McGarry JD. Characterization of the Mitochondrial Carnitine Palmitoyltransferase Enzyme System. I. Use of Inhibitors. J Biol Chem (1987) 262(20):9812–21.
- van der Windt GJ, Everts B, Chang CH, Curtis JD, Freitas TC, Amiel E, et al. Mitochondrial Respiratory Capacity Is a Critical Regulator of Cd8+ T Cell Memory Development. *Immunity* (2012) 36(1):68–78. doi: 10.1016/ j.immuni.2011.12.007
- Huang SC, Everts B, Ivanova Y, O'Sullivan D, Nascimento M, Smith AM, et al. Cell-Intrinsic Lysosomal Lipolysis Is Essential for Alternative Activation of Macrophages. *Nat Immunol* (2014) 15(9):846–55. doi: 10.1038/ni.2956
- Raud B, Roy DG, Divakaruni AS, Tarasenko TN, Franke R, Ma EH, et al. Etomoxir Actions on Regulatory and Memory T Cells Are Independent of Cpt1a-Mediated Fatty Acid Oxidation. *Cell Metab* (2018) 28(3):504–15.e7. doi: 10.1016/j.cmet.2018.06.002
- Laplante M, Sabatini DM. Mtor Signaling in Growth Control and Disease. Cell (2012) 149(2):274–93. doi: 10.1016/j.cell.2012.03.017
- 44. Delgoffe GM, Kole TP, Zheng Y, Zarek PE, Matthews KL, Xiao B, et al. The Mtor Kinase Differentially Regulates Effector and Regulatory T Cell Lineage

Commitment. Immunity (2009) 30(6):832-44. doi: 10.1016/ j.immuni.2009.04.014

- 45. Battaglia M, Stabilini A, Migliavacca B, Horejs-Hoeck J, Kaupper T, Roncarolo MG. Rapamycin Promotes Expansion of Functional Cd4+Cd25+Foxp3+ Regulatory T Cells of Both Healthy Subjects and Type 1 Diabetic Patients. J Immunol (2006) 177(12):8338-47. doi: 10.4049/jimmunol.177.12.8338
- Battaglia M, Stabilini A, Roncarolo MG. Rapamycin Selectively Expands Cd4+Cd25+Foxp3+ Regulatory T Cells. *Blood* (2005) 105(12):4743–8. doi: 10.1182/blood-2004-10-3932
- 47. Tan H, Yang K, Li Y, Shaw TI, Wang Y, Blanco DB, et al. Integrative Proteomics and Phosphoproteomics Profiling Reveals Dynamic Signaling Networks and Bioenergetics Pathways Underlying T Cell Activation. *Immunity* (2017) 46(3):488–503. doi: 10.1016/j.immuni.2017.02.010
- Ojeda L, Gao J, Hooten KG, Wang E, Thonhoff JR, Dunn TJ, et al. Critical Role of Pi3k/Akt/Gsk3beta in Motoneuron Specification From Human Neural Stem Cells in Response to Fgf2 and Egf. *PLoS One* (2011) 6(8): e23414. doi: 10.1371/journal.pone.0023414
- Garcia-Galiano D, Borges BC, Allen SJ, Elias CF. Pi3k Signalling in Leptin Receptor Cells: Role in Growth and Reproduction. *J Neuroendocrinol* (2019) 31(5):e12685. doi: 10.1111/jne.12685
- Yang K, Shrestha S, Zeng H, Karmaus PW, Neale G, Vogel P, et al. T Cell Exit From Quiescence and Differentiation Into Th2 Cells Depend on Raptor-Mtorc1-Mediated Metabolic Reprogramming. *Immunity* (2013) 39(6):1043– 56. doi: 10.1016/j.immuni.2013.09.015
- Ouyang X, Han Y, Qu G, Li M, Wu N, Liu H, et al. Metabolic Regulation of T Cell Development by Sin1-Mtorc2 Is Mediated by Pyruvate Kinase M2. *J Mol Cell Biol* (2019) 11(2):93–106. doi: 10.1093/jmcb/mjy065
- Tobin LM, Mavinkurve M, Carolan E, Kinlen D, O'Brien EC, Little MA, et al. Nk Cells in Childhood Obesity Are Activated, Metabolically Stressed, and Functionally Deficient. *JCI Insight* (2017) 2(24):e94939. doi: 10.1172/ jci.insight.94939
- Ray JP, Staron MM, Shyer JA, Ho PC, Marshall HD, Gray SM, et al. The Interleukin-2-Mtorc1 Kinase Axis Defines the Signaling, Differentiation, and Metabolism of T Helper 1 and Follicular B Helper T Cells. *Immunity* (2015) 43(4):690–702. doi: 10.1016/j.immuni.2015.08.017
- Angela M, Endo Y, Asou HK, Yamamoto T, Tumes DJ, Tokuyama H, et al. Fatty Acid Metabolic Reprogramming *Via* Mtor-Mediated Inductions of Ppargamma Directs Early Activation of T Cells. *Nat Commun* (2016) 7:13683. doi: 10.1038/ncomms13683
- Hopkins BD, Pauli C, Du X, Wang DG, Li X, Wu D, et al. Suppression of Insulin Feedback Enhances the Efficacy of Pi3k Inhibitors. *Nature* (2018) 560(7719):499–503. doi: 10.1038/s41586-018-0343-4
- Wu J, Yang J, Yang K, Wang H, Gorentla B, Shin J, et al. Inkt Cells Require Tsc1 for Terminal Maturation and Effector Lineage Fate Decisions. J Clin Invest (2014) 124(4):1685–98. doi: 10.1172/JCI69780
- 57. Lynch L, Nowak M, Varghese B, Clark J, Hogan AE, Toxavidis V, et al. Adipose Tissue Invariant Nkt Cells Protect Against Diet-Induced Obesity and Metabolic Disorder Through Regulatory Cytokine Production. *Immunity* (2012) 37(3):574–87. doi: 10.1016/j.immuni.2012.06.016
- LaMarche NM, Kane H, Kohlgruber AC, Dong H, Lynch L, Brenner MB. Distinct Inkt Cell Populations Use Ifngamma or Er Stress-Induced Il-10 to Control Adipose Tissue Homeostasis. *Cell Metab* (2020) 32(2):243–58.e6. doi: 10.1016/j.cmet.2020.05.017
- Delgoffe GM, Pollizzi KN, Waickman AT, Heikamp E, Meyers DJ, Horton MR, et al. The Kinase Mtor Regulates the Differentiation of Helper T Cells Through the Selective Activation of Signaling by Mtorc1 and Mtorc2. *Nat Immunol* (2011) 12(4):295–303. doi: 10.1038/ni.2005
- 60. Lee K, Gudapati P, Dragovic S, Spencer C, Joyce S, Killeen N, et al. Mammalian Target of Rapamycin Protein Complex 2 Regulates Differentiation of Th1 and Th2 Cell Subsets Via Distinct Signaling Pathways. Immunity (2010) 32(6):743–53. doi: 10.1016/j.immuni.2010.06.002
- Blagih J, Coulombe F, Vincent EE, Dupuy F, Galicia-Vazquez G, Yurchenko E, et al. The Energy Sensor Ampk Regulates T Cell Metabolic Adaptation and Effector Responses in Vivo. *Immunity* (2015) 42(1):41–54. doi: 10.1016/ j.immuni.2014.12.030
- Dunlop EA, Hunt DK, Acosta-Jaquez HA, Fingar DC, Tee AR. Ulk1 Inhibits Mtorc1 Signaling, Promotes Multisite Raptor Phosphorylation and Hinders

Substrate Binding. Autophagy (2011) 7(7):737-47. doi: 10.4161/ auto.7.7.15491

- Gwinn DM, Shackelford DB, Egan DF, Mihaylova MM, Mery A, Vasquez DS, et al. Ampk Phosphorylation of Raptor Mediates a Metabolic Checkpoint. *Mol Cell* (2008) 30(2):214–26. doi: 10.1016/j.molcel.2008.03.003
- Kazyken D, Magnuson B, Bodur C, Acosta-Jaquez H, Zhang D, Tong X, et al. Ampk Directly Activates Mtorc2 to Promote Cell Survival During Acute Energetic Stress. Sci Signal (2019) 12(585):eaav3249. doi: 10.1126/ scisignal.aav3249
- Chornoguz O, Hagan RS, Haile A, Arwood ML, Gamper CJ, Banerjee A, et al. Mtorc1 Promotes T-Bet Phosphorylation to Regulate Th1 Differentiation. J Immunol (2017) 198(10):3939-48. doi: 10.4049/ jimmunol.1601078
- Zeng H, Yang K, Cloer C, Neale G, Vogel P, Chi H. Mtorc1 Couples Immune Signals and Metabolic Programming to Establish T(Reg)-Cell Function. *Nature* (2013) 499(7459):485–90. doi: 10.1038/nature12297
- Reiser J, Sadashivaiah K, Furusawa A, Banerjee A, Singh N. Eomesodermin Driven Il-10 Production in Effector Cd8(+) T Cells Promotes a Memory Phenotype. Cell Immunol (2019) 335:93–102. doi: 10.1016/ j.cellimm.2018.11.008
- Xiong Y, Lingrel JB, Wuthrich M, Klein BS, Vasudevan NT, Jain MK, et al. Transcription Factor Klf2 in Dendritic Cells Downregulates Th2 Programming Via the Hif-1alpha/Jagged2/Notch Axis. *mBio* (2016) 7(3): e00436–16. doi: 10.1128/mBio.00436-16
- 69. Braissant O, Foufelle F, Scotto C, Dauca M, Wahli W. Differential Expression of Peroxisome Proliferator-Activated Receptors (Ppars): Tissue Distribution of Ppar-A,-B, and -Y in the Adult Rat. *Endocrinology* (1995) 137:354–66. doi: 10.1210/endo.137.1.8536636
- 70. Han L, Shen W, Bittner S, Kraemer FB, Azhar S. Ppars: Regulators of Metabolism and as Therapeutic Targets in Cardiovascular Disease. Part Ii: Ppar-B/ $\Delta$  and Ppar- $\Gamma$ . *Future Cardiol* (2017), 13(3):279–96. doi: 10.2217/fca-2017-0019.
- Forman BM, Tontonoz P, Chen J, Brun RP, Spiegelman BM, Evans RM. 15-Deoxy-Delta 12, 14-Prostaglandin J2 Is a Ligand for the Adipocyte Determination Factor Ppar Gamma. *Cell* (1995) 83(5):803-12. doi: 10.1016/0092-8674(95)90193-0
- Kliewer SA, Lenhard JM, Willson TM, Patel I, Morris DC, Lehmann JM. A Prostaglandin J2 Metabolite Binds Peroxisome Proliferator-Activated Receptor Gamma and Promotes Adipocyte Differentiation. *Cell* (1995) 83 (5):813–9. doi: 10.1016/0092-8674(95)90194-9
- Poulsen L, Siersbaek M, Mandrup S. Ppars: Fatty Acid Sensors Controlling Metabolism. Semin Cell Dev Biol (2012) 23(6):631–9. doi: 10.1016/ j.semcdb.2012.01.003
- 74. Chen T, Tibbitt CA, Feng X, Stark JM, Rohrbeck L, Rausch L, et al. Ppar-γ Promotes Type 2 Immune Responses in Allergy and Nematode Infection. *Sci Immunol* (2017) 2(9):eaal5196. doi: 10.1126/sciimmunol.aal5196
- Klotz L, Burgdorf S, Dani I, Saijo K, Flossdorf J, Hucke S, et al. The Nuclear Receptor Ppar Gamma Selectively Inhibits Th17 Differentiation in a T Cell-Intrinsic Fashion and Suppresses Cns Autoimmunity. *J Exp Med* (2009) 206 (10):2079–89. doi: 10.1084/jem.20082771
- 76. Yamauchi T, Kamon J, Waki H, Terauchi Y, Kubota N, Hara K, et al. The Fat-Derived Hormone Adiponectin Reverses Insulin Resistance Associated With Both Lipoatrophy and Obesity. *Nat Med* (2001) 7:941–6. doi: 10.1038/ 90984
- 77. Xu C, Wang J, Zhu T, Shen Y, Tang X, Fang L, et al. Cross-Talking Between Ppar and Wnt Signaling and Its Regulation in Mesenchymal Stem Cell Differentiation. *Curr Stem Cell Res Ther* (2016) 11:247–54. doi: 10.2174/ 1574888X10666150723145707
- Torres JL, Usategui-Martin R, Hernandez-Cosido L, Bernardo E, Manzanedo-Bueno L, Hernandez-Garcia I, et al. Ppar-Gamma Gene Expression in Human Adipose Tissue Is Associated With Weight Loss After Sleeve Gastrectomy. J Gastrointest Surg (2021) 26(2):286–97. doi: 10.1007/s11605-021-05216-6
- Tsao CH, Shiau MY, Chuang PH, Chang YH, Hwang J. Interleukin-4 Regulates Lipid Metabolism by Inhibiting Adipogenesis and Promoting Lipolysis. J Lipid Res (2014) 55(3):385–97. doi: 10.1194/jlr.M041392
- Li Y, Lu Y, Lin SH, Li N, Han Y, Huang Q, et al. Insulin Signaling Establishes a Developmental Trajectory of Adipose Regulatory T Cells. *Nat Immunol* (2021) 22(9):1175–85. doi: 10.1038/s41590-021-01010-3

- Nobs SP, Natali S, Pohlmeier L, Okreglicka K, Schneider C, Kurrer M, et al. Ppargamma in Dendritic Cells and T Cells Drives Pathogenic Type-2 Effector Responses in Lung Inflammation. J Exp Med (2017) 214 (10):3015–35. doi: 10.1084/jem.20162069
- Micossé C, von Meyenn L, Steck O, Kipfer E, Adam C, Simillion C, et al. Human "T H 9" Cells Are a Subpopulation of Ppar-γ + T H 2 Cells. Sci Immunol (2019) 4(31):eaat5943. doi: 10.1126/sciimmunol.aat5943
- Sarhangi N, Sharifi F, Hashemian L, Hassani Doabsari M, Heshmatzad K, Rahbaran M, et al. Pparg (Pro12ala) Genetic Variant and Risk of T2dm: A Systematic Review and Meta-Analysis. Sci Rep (2020) 10(1):12764. doi: 10.1038/s41598-020-69363-7
- Crawford LJ, Peake R, Price S, Morris TC, Irvine AE. Adiponectin Is Produced by Lymphocytes and Is a Negative Regulator of Granulopoiesis. *J Leukoc Biol* (2010) 88(4):807–11. doi: 10.1189/jlb.1109723
- Surendar J, Frohberger SJ, Karunakaran I, Schmitt V, Stamminger W, Neumann A-L, et al. Adiponectin Limits Ifn-γ and Il-17 Producing Cd4 T Cells in Obesity by Restraining Cell Intrinsic Glycolysis. *Front Immunol* (2019) 10. doi: 10.3389/fimmu.2019.02555
- Ramos-Ramirez P, Malmhall C, Tliba O, Radinger M, Bossios A. Adiponectin/Adipor1 Axis Promotes Il-10 Release by Human Regulatory T Cells. Front Immunol (2021) 12. doi: 10.3389/fimmu.2021.677550
- Niswender KD. Basal Insulin: Physiology, Pharmacology, and Clinical Implications. *Postgrad Med* (2011) 123(4):17–26. doi: 10.3810/pgm.2011. 07.2300
- Moussion C, Ortega N, Girard JP. The Il-1-Like Cytokine Il-33 Is Constitutively Expressed in the Nucleus of Endothelial Cells and Epithelial Cells in Vivo: A Novel 'Alarmin'? *PLoS One* (2008) 3(10):e3331. doi: 10.1371/journal.pone.0003331
- Amitani M, Asakawa A, Amitani H, Inui A. The Role of Leptin in the Control of Insulin-Glucose Axis. *Front Neurosci* (2013) 7. doi: 10.3389/ fnins.2013.00051
- Singla P, Bardoloi A, Parkash AA. Metabolic Effects of Obesity: A Review. World J Diabetes (2010) 1(3):76–88. doi: 10.4239/wjd.v1.i3.76
- Zeng Q, Luo X, Tang Y, Liu W, Luo R. Leptin Regulated Ilc2 Cell Through the Pi3k/Akt Pathway in Allergic Rhinitis. *Mediators Inflamm* (2020) 2020:4176082. doi: 10.1155/2020/4176082
- 92. Wahyuni S, van Dorst M, Amaruddin AI, Muhammad M, Yazdanbakhsh M, Hamid F, et al. The Relationship Between Malnutrition and Th 2 Immune Markers: A Study in School-Aged Children of Different Socio-Economic Backgrounds in Makassar, Indonesia. *Trop Med Int Health* (2021) 26 (2):195–203. doi: 10.1111/tmi.13513
- 93. Jamar G, Caranti DA, de Cassia Cesar H, Masquio DCL, Bandoni DH, Pisani LP. Leptin as a Cardiovascular Risk Marker in Metabolically Healthy Obese: Hyperleptinemia in Metabolically Healthy Obese. *Appetite* (2017) 108:477– 82. doi: 10.1016/j.appet.2016.11.013
- Sáinz N, Barrenetxe J, Moreno-Aliaga MJ, Martínez JA. Leptin Resistance and Diet-Induced Obesity: Central and Peripheral Actions of Leptin. *Metabolism* (2015) 64(1):35–46. doi: 10.1016/j.metabol.2014.10.015
- Procaccini C, De Rosa V, Galgani M, Carbone F, Cassano S, Greco D, et al. Leptin-Induced Mtor Activation Defines a Specific Molecular and Transcriptional Signature Controlling Cd4+ Effector T Cell Responses. *J Immunol* (2012) 189(6):2941–53. doi: 10.4049/jimmunol.1200935
- Mattioli B, Straface E, Quaranta MG, Giordani L, Viora M. Leptin Promotes Differentiation and Survival of Human Dendritic Cells and Licenses Them for Th1 Priming. *J Immunol* (2005) 174(11):6820–8. doi: 10.4049/ jimmunol.174.11.6820
- Orlova EG, Shirshev SV. Role of Leptin and Ghrelin in Induction of Differentiation of Il-17-Producing and T-Regulatory Cells. *Bull Exp Biol Med* (2013) 156(12):786–90. doi: 10.1007/s10517-014-2459-3
- Francisco V, Pino J, Campos-Cabaleiro V, Ruiz-Fernández C, Mera A, Gonzalez-Gay MA, et al. Obesity, Fat Mass and Immune System: Role for Leptin. *Front Physiol* (2018) 9. doi: 10.3389/fphys.2018.00640
- Lord GM, Matarese G, Howard JK, Baker RJ, Bloom SR, Lechler RI. Leptin Modulates the T-Cell Immune Response and Reverses Starvation-Induced Immunosuppression. *Nature* (1998) 394(6696):897–901. doi: 10.1038/29795
- 100. Zheng H, Zhang X, Castillo EF, Luo Y, Liu M, Yang XO. Leptin Enhances Th2 and Ilc2 Responses in Allergic Airway Disease. J Biol Chem (2016) 291 (42):22043–52. doi: 10.1074/jbc.M116.743187

- 101. Zeng Q, Luo X, Han M, Liu W, Li H. Leptin/Osteopontin Axis Regulated Type 2t Helper Cell Response in Allergic Rhinitis With Obesity. *EBioMedicine* (2018) 32:43–9. doi: 10.1016/j.ebiom.2018.05.037
- 102. Yamauchi T, Kamon J, Minokoshi Y, Ito Y, Waki H, Uchida S, et al. Adiponectin Stimulates Glucose Utilization and Fatty-Acid Oxidation by Activating Amp-Activated Protein Kinase. *Nat Med* (2002) 8(11):1288–95. doi: 10.1038/nm788
- 103. Hotta K, Funahashi T, Arita Y, Takahashi M, Matsuda M, Okamoto Y, et al. Plasma Concentrations of a Novel, Adipose-Specific Protein, Adiponectin, in Type 2 Diabetic Patients. *Arteriosclerosis Thrombosis Vasc Biol* (2000) 20 (6):1595–9. doi: 10.1161/01.ATV.20.6.1595
- 104. Aguilar-Salinas CA, Garcia EG, Robles L, Riano D, Ruiz-Gomez DG, Garcia-Ulloa AC, et al. High Adiponectin Concentrations Are Associated With the Metabolically Healthy Obese Phenotype. J Clin Endocrinol Metab (2008) 93 (10):4075–9. doi: 10.1210/jc.2007-2724
- 105. Zhang Q, Wang L, Jiang J, Lin S, Luo A, Zhao P, et al. Critical Role of Adipor1 in Regulating Th17 Cell Differentiation Through Modulation of Hif-1alpha-Dependent Glycolysis. *Front Immunol* (2020) 11. doi: 10.3389/ fmmu.2020.02040
- 106. Li D, Tsang JY, Peng J, Ho DH, Chan YK, Zhu J, et al. Adiponectin Mediated Mhc Class Ii Mismatched Cardiac Graft Rejection in Mice Is Il-4 Dependent. *PLoS One* (2012) 7(11):e48893. doi: 10.1371/journal.pone.0048893
- 107. Ryo M, Nakamura T, Kihara S, Kumada M, Shibazaki S, Takahashi M, et al. Adiponectin as a Biomarker of the Metabolic Syndrome. *Circ J* (2004) 68:975–81. doi: 10.1253/circj.68.975
- Otelea MR, Arghir OC, Zugravu C, Rascu A. Adiponectin and Asthma: Knowns, Unknowns and Controversies. *Int J Mol Sci* (2021) 22(16):8971. doi: 10.3390/ijms22168971
- 109. Li X, Zhang D, Vatner DF, Goedeke L, Hirabara SM, Zhang Y, et al. Mechanisms by Which Adiponectin Reverses High Fat Diet-Induced Insulin Resistance in Mice. *Proc Natl Acad Sci USA* (2020) 117(51):32584– 93. doi: 10.1073/pnas.1922169117
- 110. Yang W, Yang C, Luo J, Wei Y, Wang W, Zhong Y. Adiponectin Promotes Preadipocyte Differentiation *Via* the Ppargamma Pathway. *Mol Med Rep* (2018) 17(1):428–35. doi: 10.3892/mmr.2017.7881
- 111. Wang L, Luo Y, Luo L, Wu D, Ding X, Zheng H, et al. Adiponectin Restrains Ilc2 Activation by Ampk-Mediated Feedback Inhibition of Il-33 Signaling. *J Exp Med* (2021) 218(2):e20191054. doi: 10.1084/jem.20191054
- 112. Fukuhara A, Matsuda M, Nishizawa M, Segawa K, Tanaka M, Kishimoto K, et al. Visfatin: A Protein Secreted by Visceral Fat That Mimics the Effects of Insulin. *Science* (2005) 307(5708):426–30. doi: 10.1126/science.1097243
- 113. Steppan CM, Bailey ST, Bhat S, Brown EJ, Banerjee RR, Wright CM, et al. The Hormone Resistin Links Obesity to Diabetes. *Nature* (2001) 409 (6818):307–12. doi: 10.1038/35053000
- 114. Boucher J, Masri B, Daviaud D, Gesta S, Guigne C, Mazzucotelli A, et al. Apelin, a Newly Identified Adipokine Up-Regulated by Insulin and Obesity. *Endocrinology* (2005) 146(4):1764–71. doi: 10.1210/en.2004-1427
- 115. Moschen AR, Kaser A, Enrich B, Mosheimer B, Theurl M, Niederegger H, et al. Visfatin, an Adipocytokine With Proinflammatory and Immunomodulating Properties. J Immunol (2007) 178(3):1748–58. doi: 10.4049/jimmunol.178.3.1748
- 116. Son YM, Ahn SM, Kim GR, Moon YS, Kim SH, Park YM, et al. Resistin Enhances the Expansion of Regulatory T Cells Through Modulation of Dendritic Cells. *BMC Immunol* (2010) 11:33. doi: 10.1186/1471-2172-11-33
- 117. Hale C, Veniant MM. Growth Differentiation Factor 15 as a Potential Therapeutic for Treating Obesity. *Mol Metab* (2021) 46:101117. doi: 10.1016/j.molmet.2020.101117
- 118. Lee SE, Kang SG, Choi MJ, Jung SB, Ryu MJ, Chung HK, et al. Growth Differentiation Factor 15 Mediates Systemic Glucose Regulatory Action of T-Helper Type 2 Cytokines. *Diabetes* (2017) 66(11):2774–88. doi: 10.2337/ db17-0333
- 119. Tsai S, Clemente-Casares X, Zhou AC, Lei H, Ahn JJ, Chan YT, et al. Insulin Receptor-Mediated Stimulation Boosts T Cell Immunity During Inflammation and Infection. *Cell Metab* (2018) 28(6):922-34.e4. doi: 10.1016/j.cmet.2018.08.003
- Jeschke MG, Klein D, Herndon DN. Insulin Treatment Improves the Systemic Inflammatory Reaction to Severe Trauma. *Ann Surg* (2004) 239 (4):553–60. doi: 10.1097/01.sla.0000118569.10289.ad

- 121. Nicholas DA, Proctor EA, Agrawal M, Belkina AC, Van Nostrand SC, Panneerseelan-Bharath L, et al. Fatty Acid Metabolites Combine With Reduced B Oxidation to Activate Th17 Inflammation in Human Type 2 Diabetes. *Cell Metab* (2019) 30(3):447–61. doi: 10.1016/j.cmet.2019.07.004
- 122. Fabbrini E, Cella M, McCartney SA, Fuchs A, Abumrad NA, Pietka TA, et al. Association Between Specific Adipose Tissue Cd4+ T-Cell Populations and Insulin Resistance in Obese Individuals. *Gastroenterology* (2013) 145(2):366– 74.e1-3. doi: 10.1053/j.gastro.2013.04.010
- 123. Teijeiro A, Garrido A, Ferre A, Perna C, Djouder N. Inhibition of the Il-17a Axis in Adipocytes Suppresses Diet-Induced Obesity and Metabolic Disorders in Mice. Nat Metab (2021) 3(4):496–512. doi: 10.1038/s42255-021-00371-1
- 124. Zuniga LA, Shen WJ, Joyce-Shaikh B, Pyatnova EA, Richards AG, Thom C, et al. Il-17 Regulates Adipogenesis, Glucose Homeostasis, and Obesity. *J Immunol* (2010) 185(11):6947–59. doi: 10.4049/jimmunol.1001269
- 125. Bonilla WV, Fröhlich A, Senn K, Kallert S, Fernandez M, Johnson S, et al. The Alarmin Interleukin-33 Drives Protective Antiviral Cd8+ T Cell Responses. Sci Rep (2012) 335:984–9. doi: 10.1126/science.1215418
- 126. Sesti-Costa R, Silva GK, Proenca-Modena JL, Carlos D, Silva ML, Alves-Filho JC, et al. The Il-33/St2 Pathway Controls Coxsackievirus B5-Induced Experimental Pancreatitis. *J Immunol* (2013) 191(1):283–92. doi: 10.4049/jimmunol.1202806
- 127. Lan F, Yuan B, Liu T, Luo X, Huang P, Liu Y, et al. Interleukin-33 Facilitates Neutrophil Recruitment and Bacterial Clearance in s. Aureus-Caused Peritonitis Mol Immunol (2016) 72:74-80. doi: 10.1016/ j.molimm.2016.03.004
- 128. Schmitz J, Owyang A, Oldham E, Song Y, Murphy E, McClanahan TK, et al. Il-33, an Interleukin-1-Like Cytokine That Signals *Via* the Il-1 Receptor-Related Protein St2 and Induces T Helper Type 2-Associated Cytokines. *Immunity* (2005) 23(5):479–90. doi: 10.1016/j.immuni.2005.09.015
- 129. Hams E, Bermingham R, Wurlod FA, Hogan AE, O'Shea D, Preston RJ, et al. The Helminth T2 Rnase Omega1 Promotes Metabolic Homeostasis in an Il-33- and Group 2 Innate Lymphoid Cell-Dependent Mechanism. *FASEB J* (2016) 30(2):824–35. doi: 10.1096/fj.15-277822
- 130. Mahlakoiv T, Flamar AL, Johnston LK, Moriyama S, Putzel GG, Bryce PJ, et al. Stromal Cells Maintain Immune Cell Homeostasis in Adipose Tissue Via Production of Interleukin-33. Sci Immunol (2019) 4(35):eaax0416. doi: 10.1126/sciimmunol.aax0416
- 131. Tang H, Liu N, Feng X, Yang Y, Fang Y, Zhuang S, et al. Circulating Levels of Il-33 Are Elevated by Obesity and Positively Correlated With Metabolic Disorders in Chinese Adults. J Transl Med (2021) 19(1):52. doi: 10.1186/ s12967-021-02711-x
- 132. Wang H, Jiang Y, Wang H, Luo Z, Wang Y, Guan X. Il-25 Promotes Th2-Type Reactions and Correlates With Disease Severity in the Pathogenesis of Oral Lichen Planus. Arch Oral Biol (2019) 98:115–21. doi: 10.1016/ j.archoralbio.2018.11.015
- Hong HY, Chen FH, Sun YQ, Hu XT, Wei Y, Fan YP, et al. Local Il-25 Contributes to Th2-Biased Inflammatory Profiles in Nasal Polyps. *Allergy* (2018) 73(2):459–69. doi: 10.1111/all.13267
- 134. Feng J, Li L, Ou Z, Li Q, Gong B, Zhao Z, et al. Il-25 Stimulates M2 Macrophage Polarization and Thereby Promotes Mitochondrial Respiratory Capacity and Lipolysis in Adipose Tissues Against Obesity. *Cell Mol Immunol* (2018) 15(5):493–505. doi: 10.1038/cmi.2016.71
- 135. Ito T, Wang YH, Duramad O, Hori T, Delespesse GJ, Watanabe N, et al. Tslp-Activated Dendritic Cells Induce an Inflammatory T Helper Type 2 Cell Response Through Ox40 Ligand. J Exp Med (2005) 202(9):1213–23. doi: 10.1084/jem.20051135
- 136. Choa R, Tohyama J, Wada S, Meng H, Hu J, Okumura M, et al. Thymic Stromal Lymphopoietin Induces Adipose Loss Through Sebum Hypersecretion. *Science* (2021) 373(6554):eabd2893. doi: 10.1126/ science.abd2893
- 137. Patterson RE, Laughlin GA, LaCroix AZ, Hartman SJ, Natarajan L, Senger CM, et al. Intermittent Fasting and Human Metabolic Health. J Acad Nutr Diet (2015) 115(8):1203–12. doi: 10.1016/j.jand.2015.02.018
- Mihaylova MM, Cheng CW, Cao AQ, Tripathi S, Mana MD, Bauer-Rowe KE, et al. Fasting Activates Fatty Acid Oxidation to Enhance Intestinal Stem Cell Function During Homeostasis and Aging. *Cell Stem Cell* (2018) 22 (5):769–78.e4. doi: 10.1016/j.stem.2018.04.001

- 139. Shushimita S, de Bruijn MJ, de Bruin RW, JN IJ, Hendriks RW, Dor FJ. Dietary Restriction and Fasting Arrest B and T Cell Development and Increase Mature B and T Cell Numbers in Bone Marrow. *PloS One* (2014) 9(2):e87772. doi: 10.1371/journal.pone.0087772
- 140. Jordan S, Tung N, Casanova-Acebes M, Chang C, Cantoni C, Zhang D, et al. Dietary Intake Regulates the Circulating Inflammatory Monocyte Pool. *Cell* (2019) 178(5):1102–14.e17. doi: 10.1016/j.cell.2019.07.050
- 141. Collins N, Han SJ, Enamorado M, Link VM, Huang B, Moseman EA, et al. The Bone Marrow Protects and Optimizes Immunological Memory During Dietary Restriction. *Cell* (2019) 178(5):1088–101.e15. doi: 10.1016/ j.cell.2019.07.049
- 142. Suzuki Y, Hayashi T, Yokoyama R, Nakagawa F, Inoue J, Higashi T, et al. Fasting Impairs Type 2 Helper T Cell Infiltration in the Lung of an Eosinophilic Asthma Mouse Model. *FEBS Open Bio* (2021) 11(9):2619–30. doi: 10.1002/2211-5463.13268
- 143. Faris MA, Kacimi S, Al-Kurd RA, Fararjeh MA, Bustanji YK, Mohammad MK, et al. Intermittent Fasting During Ramadan Attenuates Proinflammatory Cytokines and Immune Cells in Healthy Subjects. *Nutr Res* (2012) 32(12):947–55. doi: 10.1016/j.nutres.2012.06.021
- 144. Lenehan PJ, Cirella A, Uchida AM, Crowley SJ, Sharova T, Boland G, et al. Type 2 Immunity Is Maintained During Cancer-Associated Adipose Tissue Wasting. *Immunother Adv* (2021) 1(1):ltab011. doi: 10.1093/immadv/ltab011
- 145. Molofsky AB, Nussbaum JC, Liang HE, Van Dyken SJ, Cheng LE, Mohapatra A, et al. Innate Lymphoid Type 2 Cells Sustain Visceral Adipose Tissue Eosinophils and Alternatively Activated Macrophages. J Exp Med (2013) 210 (3):535–49. doi: 10.1084/jem.20121964
- 146. Wu D, Molofsky AB, Liang HE, Ricardo-Gonzalez RR, Jouihan HA, Bando JK, et al. Eosinophils Sustain Adipose Alternatively Activated Macrophages Associated With Glucose Homeostasis. *Science* (2011) 332(6026):243–7. doi: 10.1126/science.1201475
- 147. Oriss TB, McCarthy SA, Morel BF, Campana MA, Morel PA. Crossregulation Between T Helper Cell (Th)1 and Th2: Inhibition of Th2 Proliferation by Ifn-Gamma Involves Interference With Il-1. *J Immunol* (1997) 158(8):3666–72.
- 148. Gajewski TF, Fitch FW. Anti-Proliferative Effect of Ifn-Gamma in Immune Regulation. I. Ifn-Gamma Inhibits the Proliferation of Th2 But Not Th1 Murine Helper T Lymphocyte Clones. J Immunol (1988) 140(12):4245–52.
- 149. Naka T, Tsutsui H, Fujimoto M, Kawazoe Y, Kohzaki H, Morita Y, et al. Socs-1/Ssi-1-Deficient Nkt Cells Participate in Severe Hepatitis Through Dysregulated Cross-Talk Inhibition of Ifn-Gamma and Il-4 Signaling in Vivo. *Immunity* (2001) 14(5):535–45. doi: 10.1016/s1074-7613(01)00132-7
- 150. Yu CR, Mahdi RM, Ebong S, Vistica BP, Chen J, Guo Y, et al. Cell Proliferation and Stat6 Pathways Are Negatively Regulated in T Cells by Stat1 and Suppressors of Cytokine Signaling. J Immunol (2004) 173(2):737– 46. doi: 10.4049/jimmunol.173.2.737
- 151. Hwang ES, Szabo SJ, Schwartzberg PL, Glimcher LH. T Helper Cell Fate Specified by Kinase-Mediated Interaction of T-Bet With Gata-3. *Science* (2005) 307(5708):430–3. doi: 10.1126/science.1103336
- 152. Winer S, Chan Y, Paltser G, Truong D, Tsui H, Bahrami J, et al. Normalization of Obesity-Associated Insulin Resistance Through Immunotherapy. *Nat Med* (2009) 15(8):921–9. doi: 10.1038/nm.2001
- 153. Schwartz C, Schmidt V, Deinzer A, Hawerkamp HC, Hams E, Bayerlein J, et al. Innate Pd-L1 Limits T Cell-Mediated Adipose Tissue Inflammation and Ameliorates Diet-Induced Obesity. *Sci Transl Med* (2022) 14(635):eabj6879. doi: 10.1126/scitranslmed.abj6879
- 154. Oldenhove G, Boucquey E, Taquin A, Acolty V, Bonetti L, Ryffel B, et al. Pd-1 Is Involved in the Dysregulation of Type 2 Innate Lymphoid Cells in a Murine Model of Obesity. *Cell Rep* (2018) 25(8):2053–60.e4. doi: 10.1016/ j.celrep.2018.10.091
- 155. Deiuliis J, Shah Z, Shah N, Needleman B, Mikami D, Narula V, et al. Visceral Adipose Inflammation in Obesity Is Associated With Critical Alterations in Tregulatory Cell Numbers. *PLoS One* (2011) 6(1):e16376. doi: 10.1371/ journal.pone.0016376
- 156. Hotamisligil GS, Shargill NS, Spiegelman BM. Adipose Expression of Tumor Necrosis Factor-Alpha: Direct Role in Obesity-Linked Insulin Resistance. *Science* (1993) 259(5091):87–91. doi: 10.1126/science.7678183
- Mathis D. Immunological Goings-On in Visceral Adipose Tissue. Cell Metab (2013) 17(6):851–9. doi: 10.1016/j.cmet.2013.05.008

- Ouchi N, Parker JL, Lugus JJ, Walsh K. Adipokines in Inflammation and Metabolic Disease. Nat Rev Immunol (2011) 11(2):85–97. doi: 10.1038/ nri2921
- 159. Odegaard JI, Ricardo-Gonzalez RR, Goforth MH, Morel CR, Subramanian V, Mukundan L, et al. Macrophage-Specific Ppargamma Controls Alternative Activation and Improves Insulin Resistance. *Nature* (2007) 447 (7148):1116–20. doi: 10.1038/nature05894
- 160. Lumeng CN, Bodzin JL, Saltiel AR. Obesity Induces a Phenotypic Switch in Adipose Tissue Macrophage Polarization. J Clin Invest (2007) 117(1):175–84. doi: 10.1172/JCI29881
- 161. Fallon PG, Schwartz C. The High and Lows of Type 2 Asthma and Mouse Models. J Allergy Clin Immunol (2020) 145(2):496–8. doi: 10.1016/ j.jaci.2019.11.031
- 162. Miethe S, Guarino M, Alhamdan F, Simon HU, Renz H, Dufour JF, et al. Effects of Obesity on Asthma: Immunometabolic Links. *Pol Arch Intern Med* (2018) 128(7-8):469–77. doi: 10.20452/pamw.4304
- Wenzel SE. Asthma Phenotypes: The Evolution From Clinical to Molecular Approaches. Nat Med (2012) 18(5):716–25. doi: 10.1038/nm.2678
- 164. Lang JE. Obesity and Asthma in Children: Current and Future Therapeutic Options. *Paediatr Drugs* (2014) 16(3):179–88. doi: 10.1007/s40272-014-0069-1
- 165. Holguin F, Bleecker ER, Busse WW, Calhoun WJ, Castro M, Erzurum SC, et al. Obesity and Asthma: An Association Modified by Age of Asthma Onset. J Allergy Clin Immunol (2011) 127(6):1486–93.e2. doi: 10.1016/ j.jaci.2011.03.036
- 166. Zheng H, Wu D, Wu X, Zhang X, Zhou Q, Luo Y, et al. Leptin Promotes Allergic Airway Inflammation Through Targeting the Unfolded Protein Response Pathway. Sci Rep (2018) 8(1):8905. doi: 10.1038/s41598-018-27278-4
- 167. Calixto MC, Lintomen L, Schenka A, Saad MJ, Zanesco A, Antunes E. Obesity Enhances Eosinophilic Inflammation in a Murine Model of Allergic Asthma. Br J Pharmacol (2010) 159(3):617–25. doi: 10.1111/j.1476-5381.2009.00560.x
- 168. Everaere L, Ait-Yahia S, Molendi-Coste O, Vorng H, Quemener S, LeVu P, et al. Innate Lymphoid Cells Contribute to Allergic Airway Disease Exacerbation by Obesity. J Allergy Clin Immunol (2016) 138(5):1309– 18.e11. doi: 10.1016/j.jaci.2016.03.019
- 169. Silva FMC, Oliveira EE, Gouveia ACC, Brugiolo ASS, Alves CC, Correa JOA, et al. Obesity Promotes Prolonged Ovalbumin-Induced Airway Inflammation Modulating T Helper Type 1 (Th1), Th2 and Th17 Immune Responses in Balb/C Mice. *Clin Exp Immunol* (2017) 189(1):47–59. doi: 10.1111/cei.12958
- 170. Suzuki Y, Aono Y, Akiyama N, Horiike Y, Naoi H, Horiguchi R, et al. Involvement of Autophagy in Exacerbation of Eosinophilic Airway Inflammation in a Murine Model of Obese Asthma. *Autophagy* (2022) 1– 13. doi: 10.1080/15548627.2022.2025571
- 171. Schroder T, Wiese AV, Ender F, Quell KM, Vollbrandt T, Duhn J, et al. Short-Term High-Fat Diet Feeding Protects From the Development of Experimental Allergic Asthma in Mice. *Clin Exp Allergy* (2019) 49 (9):1245–57. doi: 10.1111/cea.13454
- 172. Hawerkamp HC, Fahy CMR, Fallon PG, Schwartz C. Break on Through: The Role of Innate Immunity and Barrier Defence in Atopic Dermatitis and Psoriasis. *Skin Health Dis* (2022) 2:e99. doi: 10.1002/ski2.99
- 173. Ali Z, Suppli Ulrik C, Agner T, Thomsen SF. Is Atopic Dermatitis Associated With Obesity? A Systematic Review of Observational Studies. J Eur Acad Dermatol Venereol (2018) 32(8):1246–55. doi: 10.1111/jdv.14879
- 174. Zhang A, Silverberg JI. Association of Atopic Dermatitis With Being Overweight and Obese: A Systematic Review and Metaanalysis. J Am Acad Dermatol (2015) 72(4):606–16.e4. doi: 10.1016/j.jaad.2014.12.013
- Jensen P, Skov L. Psoriasis and Obesity. Dermatology (2016) 232(6):633–9. doi: 10.1159/000455840
- 176. Snekvik I, Nilsen TIL, Romundstad PR, Saunes M. Metabolic Syndrome and Risk of Incident Psoriasis: Prospective Data From the Hunt Study, Norway. *Br J Dermatol* (2019) 180(1):94–9. doi: 10.1111/bjd.16885
- Hahn M, Ghoreschi K. The Role of Il-4 in Psoriasis. *Expert Rev Clin Immunol* (2017) 13(3):171–3. doi: 10.1080/1744666X.2017.1279054
- Ascott A, Mansfield KE, Schonmann Y, Mulick A, Abuabara K, Roberts A, et al. Atopic Eczema and Obesity: A Population-Based Study. *Br J Dermatol* (2021) 184(5):871–9. doi: 10.1111/bjd.19597

- 179. Huang AH, Roh YS, Sutaria N, Choi J, Williams KA, Canner JK, et al. Real-World Comorbidities of Atopic Dermatitis in the Pediatric Ambulatory Population in the United States. J Am Acad Dermatol (2021) 85(4):893–900. doi: 10.1016/j.jaad.2021.03.016
- 180. Silverberg JI, Kleiman E, Lev-Tov H, Silverberg NB, Durkin HG, Joks R, et al. Association Between Obesity and Atopic Dermatitis in Childhood: A Case-Control Study. J Allergy Clin Immunol (2011) 127(5):1180–6.e1. doi: 10.1016/j.jaci.2011.01.063
- 181. Kim JH, Lee SW, Yon DK, Ha EK, Jee HM, Sung M, et al. Association of Serum Lipid Parameters With the Scorad Index and Onset of Atopic Dermatitis in Children. *Pediatr Allergy Immunol* (2021) 32(2):322–30. doi: 10.1111/pai.13391
- 182. Gocke AR, Hussain RZ, Yang Y, Peng H, Weiner J, Ben LH, et al. Transcriptional Modulation of the Immune Response by Peroxisome Proliferator-Activated Receptor-{Alpha} Agonists in Autoimmune Disease. *J Immunol* (2009) 182(7):4479–87. doi: 10.4049/jimmunol.0713927
- 183. Weidinger S, Beck LA, Bieber T, Kabashima K, Irvine AD. Atopic Dermatitis. Nat Rev Dis Primers (2018) 4(1):1. doi: 10.1038/s41572-018-0001-z
- 184. Nino M, Franzese A, Ruggiero Perrino N, Balato N. The Effect of Obesity on Skin Disease and Epidermal Permeability Barrier Status in Children. *Pediatr Dermatol* (2012) 29(5):567–70. doi: 10.1111/j.1525-1470.2012.01738.x
- 185. Aoki M, Murase T. Obesity-Associated Insulin Resistance Adversely Affects Skin Function. *PLoS One* (2019) 14(10):e0223528. doi: 10.1371/ journal.pone.0223528
- 186. Strid J, McLean WHI, Irvine AD. Too Much, Too Little or Just Enough: A Goldilocks Effect for Il-13 and Skin Barrier Regulation? J Invest Dermatol (2016) 136(3):561–4. doi: 10.1016/j.jid.2015.12.025
- 187. Kanoh H, Ishitsuka A, Fujine E, Matsuhaba S, Nakamura M, Ito H, et al. Ifn-Gamma Reduces Epidermal Barrier Function by Affecting Fatty Acid Composition of Ceramide in a Mouse Atopic Dermatitis Model. *J Immunol Res* (2019) 2019:3030268. doi: 10.1155/2019/3030268
- Mizutani Y, Takagi N, Nagata H, Inoue S. Interferon-Gamma Downregulates Tight Junction Function, Which Is Rescued by Interleukin-17a. *Exp Dermatol* (2021) 30(12):1754–63. doi: 10.1111/exd.14425
- Pierpont YN, Dinh TP, Salas RE, Johnson EL, Wright TG, Robson MC, et al. Obesity and Surgical Wound Healing: A Current Review. *ISRN Obes* (2014) 2014:638936. doi: 10.1155/2014/638936
- 190. Gause WC, Wynn TA, Allen JE. Type 2 Immunity and Wound Healing: Evolutionary Refinement of Adaptive Immunity by Helminths. *Nat Rev Immunol* (2013) 13(8):607–14. doi: 10.1038/nri3476
- 191. Gieseck RL3rd, Wilson MS, Wynn TA. Type 2 Immunity in Tissue Repair and Fibrosis. Nat Rev Immunol (2018) 18(1):62–76. doi: 10.1038/nri.2017.90
- 192. Farhadihosseinabadi B, Gholipourmalekabadi M, Salimi M, Abdollahifar MA, Bagheri M, Samadikuchaksaraei A, et al. The in Vivo Effect of Lacto-N-Neotetraose (Lnnt) on the Expression of Type 2 Immune Response Involved Genes in the Wound Healing Process. *Sci Rep* (2020) 10(1):997. doi: 10.1038/s41598-020-57860-8
- 193. Murray PJ, Allen JE, Biswas SK, Fisher EA, Gilroy DW, Goerdt S, et al. Macrophage Activation and Polarization: Nomenclature and Experimental Guidelines. *Immunity* (2014) 41(1):14–20. doi: 10.1016/j.immuni. 2014.06.008
- 194. Raza K, Falciani F, Curnow SJ, Ross EJ, Lee CY, Akbar AN, et al. Early Rheumatoid Arthritis Is Characterized by a Distinct and Transient Synovial Fluid Cytokine Profile of T Cell and Stromal Cell Origin. *Arthritis Res Ther* (2005) 7(4):R784–95. doi: 10.1186/ar1733
- 195. Omata Y, Frech M, Primbs T, Lucas S, Andreev D, Scholtysek C, et al. Group 2 Innate Lymphoid Cells Attenuate Inflammatory Arthritis and Protect From Bone Destruction in Mice. *Cell Rep* (2018) 24(1):169–80. doi: 10.1016/ j.celrep.2018.06.005
- 196. Bessis N, Boissier MC, Ferrara P, Blankenstein T, Fradelizi D, Fournier C. Attenuation of Collagen-Induced Arthritis in Mice by Treatment With Vector Cells Engineered to Secrete Interleukin-13. *Eur J Immunol* (1996) 26(10):2399–403. doi: 10.1002/eji.1830261020
- 197. Bessis N, Chiocchia G, Kollias G, Minty A, Fournier C, Fradelizi D, et al. Modulation of Proinflammatory Cytokine Production in Tumour Necrosis Factor-Alpha (Tnf-Alpha)-Transgenic Mice by Treatment With Cells Engineered to Secrete Il-4, Il-10 or Il-13. *Clin Exp Immunol* (1998) 111 (2):391–6. doi: 10.1046/j.1365-2249.1998.00500.x

- 198. Biton J, Khaleghparast Athari S, Thiolat A, Santinon F, Lemeiter D, Herve R, et al. *In Vivo* Expansion of Activated Foxp3+ Regulatory T Cells and Establishment of a Type 2 Immune Response Upon II-33 Treatment Protect Against Experimental Arthritis. *J Immunol* (2016) 197(5):1708–19. doi: 10.4049/jimmunol.1502124
- 199. Withrow D, Alter DA. The Economic Burden of Obesity Worldwide: A Systematic Review of the Direct Costs of Obesity. Obes Rev (2011) 12(2):131– 41. doi: 10.1111/j.1467-789X.2009.00712.x
- 200. Silva F, Oliveira EE, Ambrosio MGE, Ayupe MC, Souza VP, Gameiro J, et al. High-Fat Diet-Induced Obesity Worsens Th2 Immune Response and Immunopathologic Characteristics in Murine Model of Eosinophilic Oesophagitis. *Clin Exp Allergy* (2020) 50(2):244–55. doi: 10.1111/cea.13533
- Ghilotti F, Bellocco R, Ye W, Adami HO, Trolle Lagerros Y. Obesity and Risk of Infections: Results From Men and Women in the Swedish National March Cohort. Int J Epidemiol (2019) 48(6):1783–94. doi: 10.1093/ije/dyz129
- Huttunen R, Syrjanen J. Obesity and the Risk and Outcome of Infection. Int J Obes (Lond) (2013) 37(3):333–40. doi: 10.1038/ijo.2012.62
- 203. Cai Z, Yang Y, Zhang J. Obesity Is Associated With Severe Disease and Mortality in Patients With Coronavirus Disease 2019 (Covid-19): A Meta-Analysis. BMC Public Health (2021) 21(1):1505. doi: 10.1186/s12889-021-11546-6
- 204. Lucas C, Wong P, Klein J, Castro TBR, Silva J, Sundaram M, et al. Longitudinal Analyses Reveal Immunological Misfiring in Severe Covid-19. Nature (2020) 584(7821):463–9. doi: 10.1038/s41586-020-2588-y
- 205. Donlan AN, Sutherland TE, Marie C, Preissner S, Bradley BT, Carpenter RM, et al. Il-13 Is a Driver of Covid-19 Severity. *JCI Insight* (2021) 6(15): e150107. doi: 10.1172/jci.insight.150107
- 206. Vaz de Paula CB, de Azevedo MLV, Nagashima S, Martins APC, Malaquias MAS, Miggiolaro A, et al. 11-4/II-13 Remodeling Pathway of Covid-19 Lung Injury. *Sci Rep* (2020) 10(1):18689. doi: 10.1038/s41598-020-75659-5
- 207. Rytter MJ, Kolte L, Briend A, Friis H, Christensen VB. The Immune System in Children With Malnutrition–a Systematic Review. *PLoS One* (2014) 9(8): e105017. doi: 10.1371/journal.pone.0105017
- Gerriets VA, MacIver NJ. Role of T Cells in Malnutrition and Obesity. Front Immunol (2014) 5. doi: 10.3389/fimmu.2014.00379
- Saucillo DC, Gerriets VA, Sheng J, Rathmell JC, Maciver NJ. Leptin Metabolically Licenses T Cells for Activation to Link Nutrition and Immunity. J Immunol (2014) 192(1):136–44. doi: 10.4049/jimmunol.1301158
- 210. Najera O, Gonzalez C, Toledo G, Lopez L, Ortiz R. Flow Cytometry Study of Lymphocyte Subsets in Malnourished and Well-Nourished Children With Bacterial Infections. *Clin Diagn Lab Immunol* (2004) 11(3):577–80. doi: 10.1128/CDLI.11.3.577-580.2004
- 211. Gonzalez-Torres C, Gonzalez-Martinez H, Miliar A, Najera O, Graniel J, Firo V, et al. Effect of Malnutrition on the Expression of Cytokines Involved in Th1 Cell Differentiation. *Nutrients* (2013) 5(2):579–93. doi: 10.3390/nu5020579
- 212. Gonzalez-Martinez H, Rodriguez L, Najera O, Cruz D, Miliar A, Dominguez A, et al. Expression of Cytokine Mrna in Lymphocytes of Malnourished Children. J Clin Immunol (2008) 28(5):593–9. doi: 10.1007/s10875-008-9204-5
- Dreau D, Morton DS, Foster M, Fowler N, Sonnenfeld G. Effects of 2-Deoxy-D-Glucose Administration on Cytokine Production in Bdf1 Mice. J Interferon Cytokine Res (2000) 20(2):247–55. doi: 10.1089/107999000312667
- 214. Bai M, Wang Y, Han R, Xu L, Huang M, Zhao J, et al. Intermittent Caloric Restriction With a Modified Fasting-Mimicking Diet Ameliorates Autoimmunity and Promotes Recovery in a Mouse Model of Multiple Sclerosis. *J Nutr Biochem* (2021) 87:108493. doi: 10.1016/j.jnutbio.2020.108493
- 215. Ryan SM, Eichenberger RM, Ruscher R, Giacomin PR, Loukas A. Harnessing Helminth-Driven Immunoregulation in the Search for Novel Therapeutic Modalities. *PLoS Pathog* (2020) 16(5):e1008508. doi: 10.1371/ journal.ppat.1008508
- 216. Aravindhan V, Mohan V, Surendar J, Muralidhara Rao M, Pavankumar N, Deepa M, et al. Decreased Prevalence of Lymphatic Filariasis Among Diabetic Subjects Associated With a Diminished Pro-Inflammatory Cytokine Response (Cures 83). *PLoS Negl Trop Dis* (2010) 4(6):e707. doi: 10.1371/journal.pntd.0000707
- 217. Chen Y, Lu J, Huang Y, Wang T, Xu Y, Xu M, et al. Association of Previous Schistosome Infection With Diabetes and Metabolic Syndrome: A Cross-

Sectional Study in Rural China. J Clin Endocrinol Metab (2013) 98(2):E283–7. doi: 10.1210/jc.2012-2517

- 218. Shen SW, Lu Y, Li F, Shen ZH, Xu M, Yao WF, et al. The Potential Long-Term Effect of Previous Schistosome Infection Reduces the Risk of Metabolic Syndrome Among Chinese Men. *Parasit Immunol* (2015) 37(7):333–9. doi: 10.1111/pim.12187
- Wiria AE, Hamid F, Wammes LJ, Prasetyani MA, Dekkers OM, May L, et al. Infection With Soil-Transmitted Helminths Is Associated With Increased Insulin Sensitivity. *PLoS One* (2015) 10(6):e0127746. doi: 10.1371/journal.pone.0127746
- 220. Hays R, Esterman A, Giacomin P, Loukas A, McDermott R. Does Strongyloides Stercoralis Infection Protect Against Type 2 Diabetes in Humans? Evidence From Australian Aboriginal Adults. *Diabetes Res Clin Pract* (2015) 107(3):355–61. doi: 10.1016/j.diabres.2015.01.012
- 221. Rajamanickam A, Munisankar S, Bhootra Y, Dolla C, Thiruvengadam K, Nutman TB, et al. Metabolic Consequences of Concomitant Strongyloides Stercoralis Infection in Patients With Type 2 Diabetes Mellitus. *Clin Infect Dis* (2019) 69(4):697–704. doi: 10.1093/cid/ciy935
- 222. Tahapary DL, de Ruiter K, Martin I, Brienen EAT, van Lieshout L, Cobbaert CM, et al. Effect of Anthelmintic Treatment on Insulin Resistance: A Cluster-Randomized, Placebo-Controlled Trial in Indonesia. *Clin Infect Dis* (2017) 65(5):764–71. doi: 10.1093/cid/cix416
- 223. Sanya RE, Webb EL, Zziwa C, Kizindo R, Sewankambo M, Tumusiime J, et al. The Effect of Helminth Infections and Their Treatment on Metabolic Outcomes: Results of a Cluster-Randomized Trial. *Clin Infect Dis* (2020) 71 (3):601–13. doi: 10.1093/cid/ciz859
- 224. Yang Z, Grinchuk V, Smith A, Qin B, Bohl JA, Sun R, et al. Parasitic Nematode-Induced Modulation of Body Weight and Associated Metabolic Dysfunction in Mouse Models of Obesity. *Infect Immun* (2013) 81(6):1905– 14. doi: 10.1128/IAI.00053-13
- 225. Hussaarts L, Garcia-Tardon N, van Beek L, Heemskerk MM, Haeberlein S, van der Zon GC, et al. Chronic Helminth Infection and Helminth-Derived Egg Antigens Promote Adipose Tissue M2 Macrophages and Improve Insulin Sensitivity in Obese Mice. *FASEB J* (2015) 29(7):3027–39. doi: 10.1096/fj.14-266239
- 226. Su CW, Chen CY, Li Y, Long SR, Massey W, Kumar DV, et al. Helminth Infection Protects Against High Fat Diet-Induced Obesity Via Induction of Alternatively Activated Macrophages. Sci Rep (2018) 8(1):4607. doi: 10.1038/ s41598-018-22920-7
- 227. Su CW, Chen CY, Jiao L, Long SR, Mao T, Ji Q, et al. Helminth-Induced and Th2-Dependent Alterations of the Gut Microbiota Attenuate Obesity Caused by High-Fat Diet. *Cell Mol Gastroenterol Hepatol* (2020) 10(4):763–78. doi: 10.1016/j.jcmgh.2020.06.010
- 228. Rajamanickam A, Munisankar S, Thiruvengadam K, Menon PA, Dolla C, Nutman TB, et al. Impact of Helminth Infection on Metabolic and Immune Homeostasis in Non-Diabetic Obesity. *Front Immunol* (2020) 11. doi: 10.3389/fimmu.2020.02195
- 229. van der Zande HJP, Gonzalez MA, de Ruiter K, Wilbers RHP, Garcia-Tardon N, van Huizen M, et al. The Helminth Glycoprotein Omega-1 Improves Metabolic Homeostasis in Obese Mice Through Type 2 Immunity-Independent Inhibition of Food Intake. *FASEB J* (2021) 35(2):e21331. doi: 10.1096/fj.202001973R
- 230. Crowe J, Lumb FE, Doonan J, Broussard M, Tarafdar A, Pineda MA, et al. The Parasitic Worm Product Es-62 Promotes Health- and Life-Span in a High Calorie Diet-Accelerated Mouse Model of Ageing. *PLoS Pathog* (2020) 16(3):e1008391. doi: 10.1371/journal.ppat.1008391
- 231. Lumb FE, Crowe J, Doonan J, Suckling CJ, Selman C, Harnett MM, et al. Synthetic Small Molecule Analogues of the Immunomodulatory Acanthocheilonema Viteae Product Es-62 Promote Metabolic Homeostasis During Obesity in a Mouse Model. *Mol Biochem Parasitol* (2019) 234:111232. doi: 10.1016/j.molbiopara.2019.111232
- Schwartz C, Hams E, Fallon PG. Helminth Modulation of Lung Inflammation. Trends Parasitol (2018) 34(5):388–403. doi: 10.1016/j.pt.2017.12.007
- Obradovic M, Sudar-Milovanovic E, Soskic S, Essack M, Arya S, Stewart AJ, et al. Leptin and Obesity: Role and Clinical Implication. *Front Endocrinol* (*Lausanne*) (2021) 12. doi: 10.3389/fendo.2021.585887
- 234. Lynch L, Hogan AE, Duquette D, Lester C, Banks A, LeClair K, et al. Inkt Cells Induce Fgf21 for Thermogenesis and Are Required for Maximal Weight Loss in Glp1 Therapy. *Cell Metab* (2016) 24(3):510–9. doi: 10.1016/j.cmet.2016.08.003

- 235. Smith AD, Fan A, Qin B, Desai N, Zhao A, Shea-Donohue T. Il-25 Treatment Improves Metabolic Syndrome in High-Fat Diet and Genetic Models of Obesity. *Diabetes Metab Syndr Obes* (2021) 14:4875–87. doi: 10.2147/DMSO.S335761
- 236. Miller AM, Asquith DL, Hueber AJ, Anderson LA, Holmes WM, McKenzie AN, et al. Interleukin-33 Induces Protective Effects in Adipose Tissue Inflammation During Obesity in Mice. *Circ Res* (2010) 107(5):650–8. doi: 10.1161/CIRCRESAHA.110.218867
- 237. Vasanthakumar A, Moro K, Xin A, Liao Y, Gloury R, Kawamoto S, et al. The Transcriptional Regulators Irf4, Batf and Il-33 Orchestrate Development and Maintenance of Adipose Tissue-Resident Regulatory T Cells. *Nat Immunol* (2015) 16(3):276–85. doi: 10.1038/ni.3085
- Ricardo-Gonzalez RR, Red Eagle A, Odegaard JI, Jouihan H, Morel CR, Heredia JE, et al. Il-4/Stat6 Immune Axis Regulates Peripheral Nutrient Metabolism and Insulin Sensitivity. *Proc Natl Acad Sci USA* (2010) 107 (52):22617–22. doi: 10.1073/pnas.1009152108

**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

**Publisher's Note:** All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2022 Schmidt, Hogan, Fallon and Schwartz. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.