

# Pulmonary vascular wall stiffness: An important contributor to the increased right ventricular afterload with pulmonary hypertension

Zhijie Wang and Naomi C. Chesler

Department of Biomedical Engineering, University of Wisconsin-Madison, Wisconsin, USA

## ABSTRACT

Pulmonary hypertension (PH) is associated with structural and mechanical changes in the pulmonary vascular bed that increase right ventricular (RV) afterload. These changes, characterized by narrowing and stiffening, occur in both proximal and distal pulmonary arteries (PAs). An important consequence of arterial narrowing is increased pulmonary vascular resistance (PVR). Arterial stiffening, which can occur in both the proximal and distal pulmonary arteries, is an important index of disease progression and is a significant contributor to increased RV afterload in PH. In particular, arterial narrowing and stiffening increase the RV afterload by increasing steady and oscillatory RV work, respectively. Here we review the current state of knowledge of the causes and consequences of pulmonary arterial stiffening in PH and its impact on RV function. We review direct and indirect techniques for measuring proximal and distal pulmonary arterial stiffness, measures of arterial stiffness including elastic modulus, incremental elastic modulus, stiffness coefficient  $\beta$  and others, the changes in cellular function and the extracellular matrix proteins that contribute to pulmonary arterial stiffening, the consequences of PA stiffening for RV function and the clinical implications of pulmonary vascular stiffening for PH progression. Future investigation of the relationship between PA stiffening and RV dysfunction may facilitate new therapies aimed at improving RV function and thus ultimately reducing mortality in PH.

**Key Words:** biomechanics, impedance, vascular-ventricular coupling, right ventricular dysfunction, hypertrophy

## INTRODUCTION

Pulmonary hypertension (PH) is a complex disorder that manifests as abnormally high blood pressure in the vasculature of the lungs. Based on the causes of PH, the World Health Organization (WHO) has classified the disease into five categories including Group I, pulmonary arterial hypertension (PAH) resulting from increased pulmonary vascular resistance, Group II, PH associated with left heart disease, Group III, PH associated with lung diseases and/or hypoxemia, Group IV, PH due to chronic thrombotic and/or embolic disease, and Group V, other miscellaneous causes of PH.<sup>[1]</sup> The chronic structural and

mechanical changes in the pulmonary vasculature that occur as a consequence of PH are referred to as pulmonary vascular remodeling. Structurally, these changes include smooth muscle cell (SMC) proliferation, changes in extracellular matrix (ECM) protein content, composition and cross-linking, and either dilation or constriction of vessels, depending on their location in the circulation. The mechanical consequences of these structural changes are increased resistance and decreased compliance. The increase in pulmonary vascular resistance (PVR) that occurs with pulmonary arterial hypertension (PAH) is well studied and characterized by narrowing of small arteries

### Address correspondence to:

Naomi C. Chesler, PhD

Associate Professor of Biomedical Engineering  
University of Wisconsin at Madison  
2146 ECB; 1550 Engineering Drive  
Madison WI 53706-1609 USA  
Phone: 608/265-8920  
Fax: 608/265-9239

### Access this article online

**Quick Response Code:**



**Website:** [www.pulmonarycirculation.org](http://www.pulmonarycirculation.org)

**DOI:** 10.4103/2045-8932.83453

**How to cite this article:** Wang Z, Chesler NC. Pulmonary vascular wall stiffness: An important contributor to the increased right ventricular afterload with pulmonary hypertension. *Pulm Circ* 2011;1:212-23.

and arterioles. However, the decrease in compliance (i.e. increase in stiffness) is also important and affects the entire vasculature. Both narrowing and stiffening contribute to increased right ventricular (RV) afterload.

RV afterload is a critical metric of PH progression because typically the cause of death in PH is right heart failure due to RV overload. Left untreated, the estimated median survival of PAH is 2.8 years.<sup>[2]</sup> Conventionally, mean pulmonary arterial pressure (mPAP) is used to diagnose and predict survival in PH. Because of the dominating influence of small artery narrowing on increases in mPAP, a traditional strategy for PH treatment has been to reduce or attempt to reverse this aspect of pulmonary vascular remodeling. However, it is becoming apparent that mPAP does not correlate with either the severity of symptoms or survival. More recently, parameters reflecting RV functional status, such as RV mass and size, mean right atrial pressure, ejection fraction and cardiac index, have been shown to be strong predictors of survival.<sup>[2,4-8]</sup> As a consequence, there is growing interest in therapies that act directly on the RV to restore function.<sup>[9]</sup> Importantly, RV function depends not only on the state of the heart muscle itself (oxygen supply and demand, metabolic status, etc.) but also the vascular bed to which it is coupled, which represents its afterload.

RV afterload results from a dynamic interplay between resistance, compliance and wave reflections. It can be measured by hydraulic load or hydraulic power, which is work per unit time generated by the heart to sustain forward blood flow.<sup>[10]</sup> The power provided by RV consists of two components: the steady power required to produce net forward flow and the oscillatory power required to produce zero-mean oscillations in flow.<sup>[11]</sup> Historically, the increase in RV afterload in PAH has been attributed to increased PVR, which only reflects the steady component of total RV power. However, over a third of the RV workload increase in PH is caused by large artery stiffening,<sup>[12]</sup> which mostly influences the oscillatory RV power. In clinical studies, an increase in PA stiffness was found to be an excellent predictor of mortality in patients with PAH,<sup>[13,14]</sup> which suggests an important role of PA stiffening in right heart failure. Thus, to obtain a comprehensive view of the contributions of the pulmonary vascular bed to RV function, one must consider the complete arterial tree and investigate both steady and oscillatory components of the RV power, which are determined by steady and oscillatory components of pulmonary hemodynamics. Pulmonary artery stiffness is an important determinant of the oscillatory component of pulmonary hemodynamics.

In this review, we will focus on the current state of knowledge of the impact of pulmonary arterial remodeling (narrowing and stiffening) on RV afterload during PH progression, with a particular focus on stiffening. To do so,

we review direct and indirect techniques for measuring proximal and distal pulmonary arterial stiffness, metrics of arterial stiffness including elastic modulus, incremental elastic modulus, stiffness coefficient  $\beta$  and others, the changes in cellular function and the extracellular matrix proteins that contribute to the pulmonary arterial stiffening, and the consequences of PA stiffening for RV function. We also discuss the coupling between RV and pulmonary circulation and interactions between the proximal and distal vascular beds. Finally, we recommend directions for future studies to better understand pulmonary vascular stiffening and its relationship to RV function, dysfunction and failure in PH.

## MEASUREMENTS OF PULMONARY ARTERIAL STIFFENING

### Direct measurements

Direct measurement of pulmonary vascular stiffness can be performed *in vivo* and *in vitro* by quantifying arterial diameter as a function of pressure to generate pressure-diameter relationships or PD curves. In large animals, large and small arteries can be tested,<sup>[15]</sup> in small animals, typically only large arteries are tested.<sup>[16-18]</sup> *In vivo*, usually either inner or outer diameter is measured as a function of pressure and wall thickness is either assumed<sup>[19]</sup> or ignored. *In vitro*, both inner and outer diameter can be measured as a function of pressure,<sup>[20]</sup> as well as factors that influence the mechanical behavior of the tissue.<sup>[20-23]</sup> The most direct technique to measure stiffness is an isolated vessel mechanical test<sup>[15-18,24]</sup> in which the vessel inner and outer diameter are measured over a range of physiological and pathological pressures. This type of test allows fine control of the test conditions such that the effect of single parameter (e.g., SMC tone, collagen and/or elastin content, drug treatment, etc.) on stiffness can be determined. Biaxial (in two directions) tests of arterial sections and uniaxial (in one direction) tests of tissue either in strips or rings can also be performed.<sup>[25,26]</sup> Most biaxial and strip test methods do not allow the effects of smooth muscle cell (SMC) tone to be investigated but isolated vessel and ring tests do.<sup>[17,18,27-29]</sup> Indirect measures of proximal and distal arterial stiffness also exist, which we will review in the next section.

The most commonly used approach for assessing structural mechanical properties of arteries is PD curves. If wall thickness is measured optically or histologically, material properties can also be calculated. Because transmural pressure results in circumferential stretch of the arterial wall, most mechanical properties are characterized in the direction of circumference and sometimes derived from calculation of circumferential wall stress and strain. However, recent studies on systemic arteries have

considered changes in longitudinal stretch with disease as well.<sup>[30-32]</sup> Calculated parameters either represent the whole vessel stiffness but dependent on geometry, such as pressure-strain modulus ( $E_p$ ),<sup>[17,33-35]</sup> stiffness constant  $\beta$ ,<sup>[36,37]</sup> distensibility ( $D$ )<sup>[23,38-41]</sup> and compliance ( $C$ ),<sup>[23,38,42-44]</sup> or describe the material properties of the wall independent of geometry, such as elastic modulus ( $E$ )<sup>[34,45]</sup> and incremental elastic modulus ( $E_{inc}$ ).<sup>[21,45,46]</sup> Sometimes the geometric-dependent mechanical properties are termed “extrinsic” since they depend on the amount of wall material whereas the geometry-independent material properties are termed “intrinsic” since they depend only on the constitutive materials themselves. For example, two arteries made of the exact same material but different wall thicknesses would have the same intrinsic properties but different extrinsic properties. A summary of commonly used arterial intrinsic and extrinsic mechanical and material properties are presented in Table 1. Consistently, pulmonary artery wall stiffness is found to increase during PH progression.<sup>[16-18,26,47,48]</sup>

It is important to note that all of these parameters are linear approximations of non-linear relationships. That is, for sufficiently large changes in stress and strain (for  $E$ ) or volume and pressure (for  $D$ ), a single calculation (of  $E$  or  $D$ ) will be a poor approximation of the true material or mechanical behavior. In a physiological pressure range, most of the above metrics are good approximations but when testing is performed over a larger range, curve fits

may be applied to individual sections. The best example is elastic modulus,  $E$ , which for most arteries is low in a low strain range due to loading of elastin and high in a high strain range due to collagen engagement. Thus, low and high strain moduli ( $E_{low}$  and  $E_{high}$  respectively) are often calculated separately.<sup>[17,26,47]</sup>

Besides these structural and material properties that can be measured from PD and sometimes pressure-length curves, other parameters that require less information and thus are more feasible for clinical measurements have been introduced to investigate PA stiffening during PH progression. One example is the relative area change (RAC).<sup>[14]</sup> RAC is calculated as the relative cross-section area change ( $\Delta A/A$ ) of the proximal PA from systole to diastole and it is reduced significantly in PH patients.<sup>[14]</sup> RAC shows a moderate inverse curvilinear relationship with mPAP and predicts mortality better than area distensibility. Interestingly, RAC does not require knowledge of the distending pressure, so it is neither a material nor structural property; instead it is a geometrical property.

It is well known that blood vessels are not purely elastic but in fact viscoelastic materials, which means the elastic deformation under dynamic loads exhibits time-dependent behavior. A consequence of arterial viscoelasticity is pressure energy loss. When a purely viscous material is deformed, all the energy is dissipated

**Table 1: Summary of parameters commonly used to measure arterial elasticity**

Definition (unit)	Formula	Notes
Pressure-strain modulus $E_p$ (Pa)	$E_p = \frac{\Delta P}{\Delta R} R$	$R$ can also be replaced with diameter ( $D$ ). Extrinsic mechanical property
Elastic modulus $E$ (Pa)	$E = \frac{\sigma}{\epsilon}$	The slope of stress-strain ( $\sigma$ - $\epsilon$ ) curve, assuming linear homogeneous, incompressible wall material. Thin-wall or thick-wall assumptions lead to different calculations of $\sigma$ and $\epsilon$ . Intrinsic material property
Incremental elastic modulus <sup>1</sup> $E_{inc}$ (Pa)	$E_{inc} = \frac{\Delta P}{\Delta R_o} \frac{2(1-\nu^2)R_i^2 R_o}{(R_o^2 - R_i^2)}$	$R_o$ and $R_i$ are external and internal radii and can be replaced with corresponding diameters ( $D$ ). Assuming locally linear, homogenous, incompressible (if $\nu=0.5$ ) wall material. Thick wall assumption. Intrinsic material property
Incremental elastic modulus <sup>2</sup> $E_{inc}$ (Pa)	$E_{inc} = \frac{\Delta P}{\Delta D_o} \frac{2D_o^2 D_i}{(D_o^2 - D_i^2)} + \frac{2PD_o^2}{(D_o^2 - D_i^2)}$	Modified $E_{inc}$ , assuming orthotropic cylindrical tube. Intrinsic material property
Stiffness constant $\beta$ (dimensionless)	$\beta = \frac{\ln(P_s / P_d)}{(R_s / R_d - 1)}$	$P_s$ and $P_d$ are systolic and diastolic pressures. Radius $R$ can be replaced with diameter ( $D$ ). Assume homogeneous, incompressible, isotropic material. Extrinsic mechanical property
Distensibility $D$ (1/Pa)	$D = \frac{\Delta V}{V \cdot \Delta P}$	$V$ (volume) can be replaced with $A$ (area). Extrinsic mechanical property
Compliance $C$ (m <sup>3</sup> /Pa)	$C = \frac{\Delta V}{\Delta P}$	$V$ (volume) can be replaced with $A$ (area). Extrinsic mechanical property

**P:** pressure; **R:** radius; **D:** diameter; **V:** volume; **A:** area

and none is returned. When a purely elastic material is deformed, all the energy is returned, like a spring. A viscoelastic material is both viscous and elastic so some of the energy provided to the material during loading is dissipated during unloading and some is returned. Since most in vitro measurements do not perform dynamic loading-unloading tests such as occur in the intact animal or human due to physiological pulsatile pressure waveforms, mechanical properties obtained from in vivo or dynamic in vitro experiments are more realistic and physiological. For example, the compliance measured from static PD curves is less than that measured from the dynamic PD curves under the same pressures.<sup>[49]</sup> Although not well recognized, the characterization of arterial mechanical behavior including elasticity and viscosity may be of clinical importance in PH.

In systemic arteries, viscosity has been shown to change with aging, with atherosclerosis and systemic hypertension.<sup>[50-52]</sup> The effect of acute PH on arterial viscosity has been investigated by the group of Armentano.<sup>[53,54]</sup> Our group, to our knowledge, is the only one that has investigated changes in proximal PA viscoelasticity in chronic PH.<sup>[16,47,55]</sup> Traditionally, SMCs are considered to be the primary source of arterial viscosity<sup>[29,56]</sup> but our results suggest that SMCs may not be the only determinant. The damping capacity of large PAs increases after chronic hypoxia<sup>[16,47,55]</sup> and this increase correlates with the accumulation of collagen.<sup>[55]</sup> The clinical implications of increased PA viscosity during PH are unclear, but arterial viscosity likely increases circulatory energy loss<sup>[56]</sup> and affects wave reflections,<sup>[57]</sup> thus impairing dynamic interactions between heart and lung and increasing RV afterload.

### Indirect measurements

Both proximal and distal pulmonary arterial stiffness can be measured indirectly from pulmonary pressure-flow relationships in vivo<sup>[58-63]</sup> or ex vivo in isolated whole lungs.<sup>[63-69]</sup> Advantages of the isolated, perfused and ventilated lung preparation are that pressure-flow relationships are not affected by anesthesia,<sup>[60]</sup> volume status<sup>[59]</sup> and level of sympathetic nervous system activation.<sup>[58]</sup> In addition, the effect of drugs on the pulmonary vasculature can be investigated independent of the effects of those drugs on the systemic vasculature.<sup>[70]</sup> However, in most isolated, ventilated, perfused lung preparations, only steady pressure-flow relationships are obtained, from which only distal arterial stiffness can be derived. Our group has used pulsatile flow waveforms to obtain pulsatile pressure-flow relationships, which allow estimates of proximal artery stiffness to be made, although these are still difficult to compare to in vivo measurements obtained with physiological flow waveforms.<sup>[68,70-72]</sup> The metrics of proximal and distal pulmonary arterial stiffness

that can be obtained by in vivo and ex vivo methods are reviewed below.

### Steady pressure-flow relationships

Increases in mPAP that occur with increases in flow whether in vivo (due to exercise or drugs) or ex vivo (due to imposed flow waveforms) can provide insight into distal arterial stiffness. If one imagines the distal pulmonary arterial bed to be a parallel network of rigid tubes, then any increase in flow will lead to a proportional increase in pressure. If more rigid tubes open as a result of increased flow (i.e., are recruited), then the increase in pressure will be less at high flows. If these rigid tubes increase in diameter with increased flow (i.e., by the mechanism of flow-induced vasodilation), then the increase in pressure at high flows will be less still. Finally, if the tubes are not in fact rigid but can distend, then the pressure-flow curve may plateau at high flow. A theoretical approach to measuring this distal arterial distensibility, assuming a fully recruited and dilated pulmonary vasculature, was first described by Linehan et al.<sup>[73]</sup>:

$$mPAP = \frac{[(1 + \alpha P_v)^5 + 5\alpha R_0 (Hct) CO]^{1/5} - 1}{\alpha} \quad (1)$$

where  $P_v$  is pulmonary venous pressure, CO is cardiac output,  $R_0$  is the total pulmonary vascular resistance (mPAP/CO) at rest, Hct is the hematocrit, and  $\alpha$  is the pulmonary arterial distensibility, which is assumed to be constant throughout the pulmonary vascular bed.

In vivo<sup>[7,74]</sup> and ex vivo<sup>[75]</sup> studies have demonstrated that PH decreases  $\alpha$ . However, whether distal arterial stiffening impairs exercise capacity in PH by exacerbating flow-induced increases in mPAP remains unknown.

### Pulsatile pressure-flow relationships

From pulsatile pressure-flow relationships obtained either in vivo or ex vivo, indirect measurements of proximal arterial stiffness can be obtained. In addition, other characteristics of the pulmonary circulation can be determined such as PVR and wave reflections. Two approaches to analyze these pulsatile pressure-flow relationships are commonly used: a frequency domain method; and a time domain method.

### Impedance (Frequency domain analysis)

The relationship between pulsatile blood pressure and flow can be quantified by the impedance.<sup>[76]</sup> Its calculation requires synchronized pulmonary arterial pressure and flow measurements, which are typically obtained with a right heart catheterization and either ultrasound<sup>[77,78]</sup> or a catheter-based flow sensor<sup>[79,80]</sup> in vivo or by direct recording of pressure and flow ex vivo. From these data, a comprehensive measurement of RV afterload is

given by pulmonary arterial input impedance (PVZ). The calculation of PVZ requires a spectral analysis of the pulmonary arterial pressure waveform  $P$  and flow waveform  $Q$  and a mathematical elaboration (Fourier analysis) to derive a PVZ spectrum, which is expressed as the ratio of  $P$  to  $Q$  moduli and a phase angle ( $\theta$ ), both as a function of frequency<sup>[76]</sup>:

$$PVZ(\omega) = \frac{|P(\omega)|}{|Q(\omega)|} \quad (2)$$

$$\theta(\omega) = \Phi(\omega) - \phi(\omega) \quad (3)$$

where  $\omega$  is the frequency,  $\Phi$  is the pressure phase and  $\phi$  is the flow phase. This approach has been adopted in PH animal models using *in vivo* and *ex vivo* techniques and in patients.<sup>[19,68,78,81-83]</sup> Details of how to calculate impedance for the pulmonary circulation have been reviewed previously.<sup>[23,84,85]</sup>

The impedance spectra in systemic and pulmonary circulations share a similar, classic pattern of a high 0-Hz value ( $Z_0$ ) followed by a local minimum and oscillations at high frequencies (Fig. 1).  $Z_0$  increases during PH progression and this parameter is equal to total PVR. Like PVR,  $Z_0$  is essentially as a measure of distal pulmonary resistance; it is the input impedance in the absence of flow oscillations. The average of the impedance modulus at higher harmonics is used as an estimation of characteristic impedance ( $Z_c$ ), which is the input impedance in the absence of wave reflection.  $Z_c$  is determined principally by the ratio of stiffness of the proximal arteries to fluid inertia.<sup>[76,86]</sup> It is dependent on both the size and material properties of the proximal PAs. For example, an increase

in proximal PA radius decreases  $Z_c$  whereas an increase in stiffness has the opposite effect. These relationships are described theoretically by:

$$Z_c = \sqrt{\frac{\rho E h}{2\pi^2 r^5}} \quad (4)$$

where  $E$ ,  $h$ , and  $r$  are proximal PA elastic modulus, wall thickness and luminal radius, respectively, and  $\rho$  is the density of blood.<sup>[87,88]</sup>  $Z_c$  is usually found to remain constant in subjects with PH,<sup>[68,76,89]</sup> which may be a combined result of increased proximal PA stiffness<sup>[16,17,26,47]</sup> and diameter.<sup>[14,16]</sup> This observation suggests that during PH, the RV and pulmonary vasculature adapt to maintain the mechanical load on the RV and conserve energy.<sup>[76,89]</sup> However,  $Z_c$  has also been found to change (increase or decrease) with PH depending on the animal model used or different pathological mechanisms of PH.<sup>[61,62,81,82,88,90,91]</sup>

A third parameter that can be derived from the impedance spectrum is pulse wave reflection ( $\Gamma$ ),<sup>[76,92]</sup> which is calculated by

$$\Gamma = \frac{Z_0 - Z_c}{Z_0 + Z_c} \quad (5)$$

The wave reflection increases significantly with PH,<sup>[68,93]</sup> which suggests a bigger impedance mismatch between the proximal and distal pulmonary vasculature and may have detrimental effects on the RV function as reported to occur in the systemic circulation.<sup>[10]</sup>

Overall, PVZ has the potential to be a better prognostic indicator than PVR alone because it captures both static and dynamic characteristics of the opposition to flow in the pulmonary circulation. PVR is typically a single-value measurement assuming a linear relationship between the pressure difference  $\Delta P$  ( $\Delta P$  = Pulmonary arterial pressure - left atrial pressure) and the flow ( $Q$ ). When left atrial pressure is normal, total PVR can be measured by the ratio of mean pulmonary pressure and mean pulmonary flow (mPAP/ $Q$ ). Both PVR and total PVR characterize the steady, time-averaged hemodynamics and represent only the static component of RV afterload. It is well known that progressive pulmonary artery remodeling during PH increases PVR. However, PVR is likely limited as a prognostic parameter because PVR alone is not sufficient to measure total RV afterload.<sup>[94]</sup> In fact, more and more evidence suggests that a global stiffness measurement<sup>[13,14,83]</sup> that includes both static and dynamic components is a better representative and potentially prognostic parameter for PH progression.

### Time domain analysis

Although the rapid development of new technologies

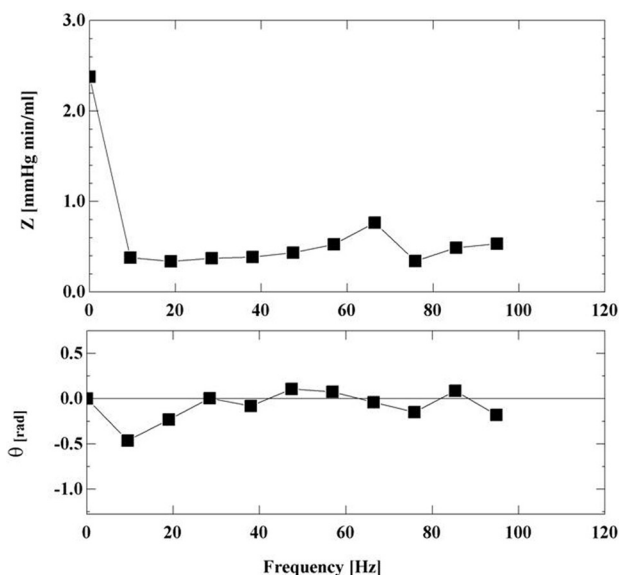


Figure 1: Representative pulmonary vascular impedance (magnitude  $Z$  and phase  $\theta$ ) spectra in a healthy mouse.

and instrumentation may allow more widespread use of impedance analysis in the frequency domain, current clinical applications are limited because of the complexity of performing and interpreting the frequency domain analysis and results, respectively. Instead, the time domain analysis of the pulse pressure waveform is a useful surrogate for assessing  $Z_0$ ,  $Z_c$  and other clinical indices of pulsatile pulmonary hemodynamics.<sup>[84,95,96]</sup>

In the time domain analysis,  $Z_0$  is calculated as mean pressure divided by mean flow, which is the same as in the frequency domain, and  $Z_c$  is calculated as

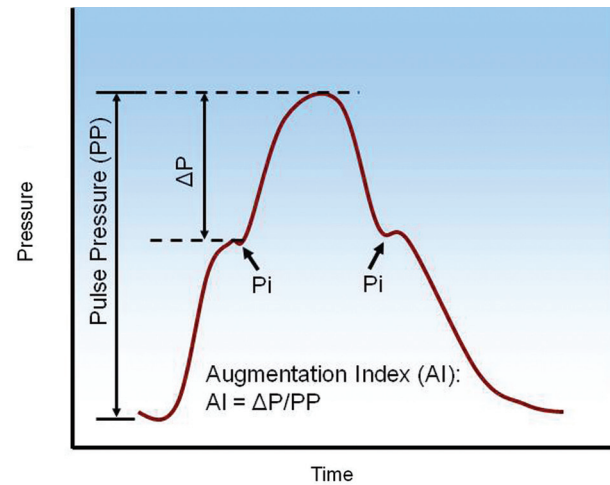
$$Z_c = \frac{dP}{dQ} \quad (6)$$

where  $dP$  and  $dQ$  are pressure and flow increases taken prior to when flow reaches 95% of its maximal value during one cardiac cycle.<sup>[97,98]</sup> Because of the rapid upstroke of the waveforms at this early ejection phase, the reflected waves do not have time to return to the proximal bed and thus the system is reasonably assumed to be free from wave reflections.<sup>[97]</sup> From  $Z_c$ , the instantaneous pressure waveform is decomposed into forward ( $P_f$ ) and backward ( $P_b$ ) traveling waves using the linear wave separation method.<sup>[99]</sup> The global wave reflection index ( $R_0$ ) is then calculated as the ratio of the amplitude of  $P_b$  to  $P_f$ . In contrast to the wave reflection ( $\Gamma$ ) that is most applicable for simple systems of one or two tubes<sup>[100]</sup> and thus characterizes mismatch between proximal and distal beds,  $R_0$  captures the wave reflections of the whole vascular bed.<sup>[101]</sup>

Correlation between impedance measurements performed using frequency and time domain analyses requires further investigation. Our preliminary results have shown good agreement between the two methods.<sup>[102]</sup>

Pulse pressure (PP) is often measured in the time domain analysis of pulmonary hemodynamics and has been found to be a useful indicator of heart health.<sup>[103,104]</sup> PP is defined as the difference between systolic and diastolic pressure (Fig. 2) and is determined by the characteristics of ventricular ejection and proximal arterial stiffness. In both systemic and pulmonary circulations, PP is elevated when large arteries stiffen, even without changes in the distal vasculature.<sup>[105-108]</sup>

Another parameter derived from the pressure waveform is the augmentation index (AI, Fig. 2), defined as the ratio of the height from the inflection point ( $P_i$ ) to peak systolic pressure ( $\Delta P$ ), which is an estimate of the magnitude of the reflected pressure wave to the PP,<sup>[96]</sup> which may correlate with proximal PA stiffness. In normal subjects there is very little wave reflection, and PH increases wave



**Figure 2:** An illustrative example of pressure waveform and the derivation of pulse pressure (PP) and augmentation index (AI).  $P_i$  is the inflection point, which may present either in the systolic or diastolic phase.

reflection.<sup>[79,89,93,96,109]</sup> AI can differentiate different types of PH, i.e., chronic pulmonary thromboembolism (CPTe) and primary pulmonary hypertension (PPH)<sup>[95,96]</sup> but it may be more sensitive to the proximal arteries than the whole vascular bed and can be confounded by other factors such as heart rate.<sup>[100,101]</sup> Finally, the relationship between AI and the severity of PH remains unknown.

A potentially more useful parameter is compliance, which conceptually is the inverse of stiffness. Compliance (C) is calculated as the ratio of stroke volume (SV) to PP.<sup>[110,111]</sup> Similar parameters measured in the systemic circulation have been shown to be related to conduit artery stiffening and correlated with mortality in patients with left ventricular dysfunction.<sup>[112]</sup> In PAH patients, compliance, which is sometimes called capacitance, has been shown to have prognostic value as well.<sup>[113,113]</sup> It is important to point out that unlike in the systemic circulation where arterial compliance is mainly localized to the aorta, arterial compliance in pulmonary circulation is distributed over the entire vascular bed.<sup>[111]</sup> An interesting relationship between compliance and resistance (PVR) has recently been observed in the pulmonary circulation: the product of PVR and C is constant.<sup>[23,91,111,114]</sup> In particular, Lankhaar et al. have shown that the product of PVR and C is a constant for patients with and without PH<sup>[91]</sup> and that treatment does not change this time constant<sup>[114]</sup> Importantly, these results explain why changes in PVR alone do not reflect the clinical outcomes of PH patients. PH leads to increased PVR and decreased C; but for a similar decrease of PVR due to therapy, the patients with higher starting PVR (more severe PH) will have a smaller increase in C than the patients with lower starting PVR (milder PH). Thus the former group of patients has improvement

mainly in steady RV power requirements (due to a decrease in PVR) whereas the latter group of patients has improvement in both steady and oscillatory RV power requirements (due to comparable decrease in PVR and increase in C).

## CELLULAR AND MOLECULAR CONTRIBUTORS TO PULMONARY ARTERIAL STIFFENING

The arterial remodeling that occurs in the progression of PH involves all three layers of the arterial wall. Histological evidence of remodeling consists of intimal fibrosis, increased medial thickness, accumulation of extracellular matrix proteins such as collagen, increased adventitial thickness, pulmonary arteriolar occlusion and plexiform lesions. The process is characterized by proliferative and obstructive changes that involve cell types such as endothelial, smooth muscle and fibroblast.<sup>[12,115]</sup> Detailed reviews on the cellular and molecular mechanisms of PA remodeling including genetic factors have been reported previously.<sup>[12,116]</sup>

Stiffening of large PAs is linked with accumulation of collagen<sup>[16,47,48,117,118]</sup> and, in some studies, elastin.<sup>[16,26,47]</sup> However, if collagen and elastin concomitantly increase during PH, one cannot discern which is more responsible for arterial stiffening. A recent study suggests elastin remodeling contributes to PA stiffening in response to hypoxia-induced pulmonary hypertension (HPH) in neonatal calves,<sup>[26]</sup> but discrepant observations are also reported in other species in adults. For example, there was no change in elastin content in rodent large PAs after chronic hypoxia.<sup>[48,119]</sup> Our latest studies using a novel transgenic mouse model (Col1a1<sup>tm1Jae</sup>) suggests changes in collagen rather than elastin track large PA stiffness during HPH progression and recovery.<sup>[17]</sup> Furthermore, we specifically examined the effects of collagen content vs. cross-linking in the passive, dynamic mechanical behavior of large PAs in chronic PH and found that collagen content is critical to large PA stiffening.<sup>[55]</sup>

Differential impacts of elastin and collagen metabolism on PH progression may exist and can be attributed to the age and type of PH developed. There is evidence suggesting that elastin may play a more important role in PH in newborns whereas collagen may have a larger impact in PH developed in adults. Studying the biomechanics of newborn calf extralobar arteries, Lammers et al.<sup>[26]</sup> showed a significant increase in low-strain elastic modulus with chronic hypoxia that was dependent on elastin. However, it is well known that newborn animals develop more severe pulmonary hypertension than adults with more dramatic vascular changes.<sup>[120-122]</sup> Since

the neonatal period is associated with significant elastin production in the pulmonary trunk even in normoxic conditions,<sup>[123]</sup> elastin synthesis may be particularly sensitive to modulation by hypoxia during this time of rapid growth. Another study using a set of transgenic and knockout mice with graded elastin insufficiency found that reduced elastin content in large PAs predisposed the mice to elevated RV pressures and RV hypertrophy.<sup>[124]</sup> With the genetic deficiency in elastin, peripheral PAs developed muscularization and proximal PAs exhibited increased overall stiffness, but the  $E_{inc}$  remained constant over a wide range of pressure change (0-90 mmHg). However, the extent of pathological remodeling of heart and pulmonary vasculature was limited compared to those with similar pressure elevation induced by hypoxia, which suggests an adaptation occurred during the development from fetus to adult. Therefore, it is likely that different mechanisms (elastin vs. collagen) may dominate the development of PH, depending on the type of disease.

The SMC tone changes significantly during PH development. This is especially prominent in the distal vasculature. It is well accepted that the acute phase of PH is manifested by vasoconstriction. In the chronic phase of PAH, remodeling occurs with endothelial layer thickening (and formation of plexiform lesions in severe PAH) and smooth muscle hypertrophy, which leads to reduced smooth muscle contractility. The proximal PAs seem to have limited changes in SMC tone,<sup>[17,18,125]</sup> but discrepant results are also reported.<sup>[27,28,126]</sup> The impact of SMC tone on proximal PA mechanical behavior in large animals or humans is an important area of future research.

## CONSEQUENCES/CLINICAL RELEVANCE OF PULMONARY ARTERIAL STIFFENING

### Right ventricular overload and heart failure

Right heart function is closely tied to survival in PH. For instance, RV function assessed via right atrial pressure and cardiac output is associated with mortality in addition to mPAP in patients with PAH.<sup>[2]</sup> Previously, much attention has been paid to the pulmonary circulation and basic mechanisms underlying pulmonary arterial dysfunction; however, little is known about the RV in health and diseased states. It is recognized now that effective treatment for PH should not only impact the pulmonary vasculature but also improve RV function.

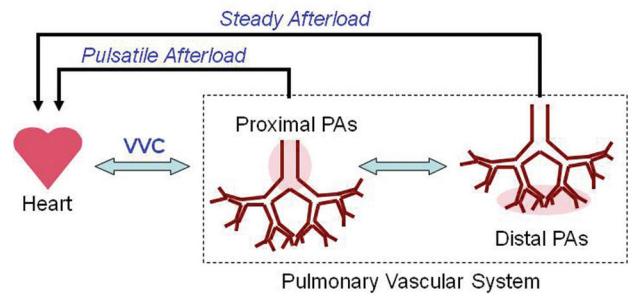
Stiffening of the pulmonary vascular bed can increase RV workload, which may be caused by impaired conduit and buffering function of the proximal PAs and early wave reflections to the RV (Fig. 3). Under normal circumstances, the oscillatory RV power is about 25% of the total

(oscillatory + steady) RV power, which is higher than the proportion in the systemic circulation (~10%).<sup>[76,127]</sup> A recent study on the RV pulsatile hydraulic load in PAH patients has shown that while both oscillatory RV work and total RV work increase with PAH severity, the proportion remains constant (~23%).<sup>[111]</sup> The oscillatory RV work correlates with pulse pressure, which increases with proximal PA stiffening.<sup>[127,128]</sup> Therefore, PA stiffness increases oscillatory RV work; however, the direct impact on RV function remains undefined. The effects of distal PA stiffening (loss of distensibility  $\alpha$ ) on RV afterload are also unknown.

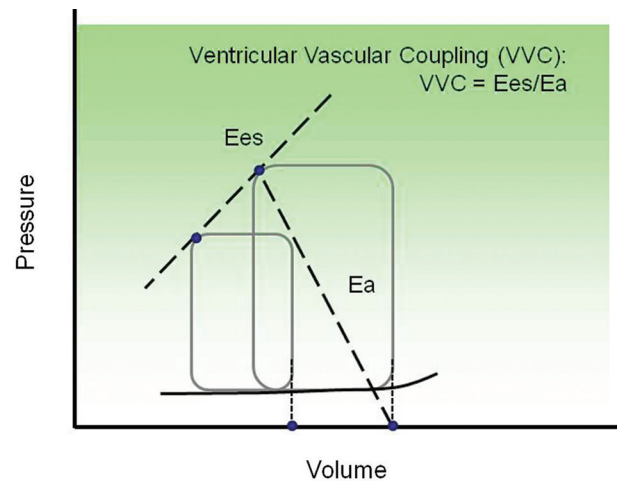
Increased RV afterload leads to augmented myocardial oxygen demand and the RV adapts primarily by hypertrophy. Hypertrophy serves to decrease the ventricular wall stress and maintain cardiac output initially and thus could be a healthy response. However, during PH progression, severe hypertrophy reduces cardiac output and cardiac index, which are the hallmarks of right heart dysfunction and failure.<sup>[9]</sup> The indicators of the transition from healthy adaptive hypertrophy to pathological maladaptive remodeling remain unknown. To understand the molecular pathways that differentiate adaptive from maladaptive ventricular hypertrophy is an important area of current research.<sup>[129]</sup>

### Ventricular vascular coupling (VVC) efficiency

To understand how pulmonary vascular remodeling affects ventricular output during PH progression, investigations into ventricular function alone or vascular function alone are not sufficient. Ideally, vascular function is efficiently matched to ventricular function, and vice versa. Ventricular pressure-volume (PV) loop analysis was first developed by Sagawa et al.<sup>[130]</sup> to describe the coupling efficiency between the left ventricle and systemic circulation, i.e., vascular-ventricular coupling (VVC). In this approach, the ventricular and arterial dynamic behaviors are quantified by ventricular end-systolic elastance ( $E_{es}$ , which represents contractility of the ventricle) and arterial effective elastance ( $E_a$ , which represents ventricular afterload), respectively, which are derived from the pressure-volume loops at different levels of preload (Fig. 4). The ratio of these two parameters ( $E_{es}/E_a$ ) yields a direct assessment of VVC efficiency. When the ventricle and vasculature are efficiently coupled, minimal energy is wasted in the pulse pressure and maximal energy is transmitted in the mean pressure. The optimal coupling efficiency ( $E_{es}/E_a$ ) has been identified and is comparable for both sides of the heart.<sup>[131,132]</sup> Recently, there is increasing interest in PV loop analysis for PH because RV contractility is a more reliable way to differentiate the effect of therapies than mPAP.<sup>[84]</sup> In addition, the linkage between RV function and vascular function, which is characterized by coupling efficiency



**Figure 3:** Hemodynamic interactions between the right ventricle and proximal and distal pulmonary arteries. PA: pulmonary artery. VVC: ventricular-vascular coupling.



**Figure 4:** An illustrative example of vascular-ventricular coupling analysis from pressure-volume (PV) loops. Ventricular end-systolic elastance ( $E_{es}$ ) and arterial effective elastance ( $E_a$ ) are calculated from multiple loops by varying the preload.

( $E_{es}/E_a$ ), may reveal the mechanisms that differentiate physiological and pathological ventricular remodeling. During acute PH, the optimal ventricular-arterial coupling appears to be maintained regardless of species and the way in which PH is induced.<sup>[82]</sup> As effective afterload ( $E_a$ ) increases, the ventricle increases contractility ( $E_{es}$ ) such that optimal coupling ( $E_{es}/E_a$ ) is maintained. However, such a balance may not exist in chronic PH due to both pulmonary vascular and right ventricular remodeling. Our recent study in mice demonstrated decreased pulmonary vascular-right ventricular coupling efficiency after chronic hypoxia.<sup>[132]</sup>

A potentially important consequence of PA wall stiffening may be impaired VVC efficiency via increased  $E_a$ . Optimal coupling between the RV and the pulmonary circulation in a healthy cardiopulmonary system is likely important to the adaptive response to acute stresses such as hypoxia or exercise. In conditions of impaired VVC, however, the RV may not be able to maintain cardiac output sufficiently to meet these challenges, leading to dysfunction and failure.



Because very few studies have examined ventricular-vascular coupling and pulmonary arterial stiffening concurrently, the linkage between changes in vascular and ventricular function is not established. For example, the arterial effective elastance ( $E_a$ ) is thought to represent the RV afterload presented by the pulmonary vasculature. However, the influences of proximal arterial stiffness, distal arterial stiffness and distal arterial narrowing on  $E_a$  are not clear. Understanding the relationship between right heart function and pulmonary vascular remodeling will aid in discerning critical factors that are relevant to clinical outcomes of PH.

### Interactions between proximal and distal pulmonary arteries

Finally, while there are different initiating mechanisms of PH (e.g., proximal occlusion and distal embolism), abnormalities in the upstream (proximal) and downstream (distal) PAs can affect each other via hemodynamics and result in a vicious cycle of remodeling (Fig. 3). In particular, large, conduit PA stiffening increases distal arterial cyclic strain damage,<sup>[133]</sup> which promotes SMC proliferation and narrowing. Increased flow pulsatility at distal arteries has been shown to induce inflammatory gene expression, leukocyte adhesion and cell proliferation in endothelial cells.<sup>[133]</sup> A similar mechanism has been demonstrated in the systemic circulation in which aortic stiffening causes damage to renal arterioles.<sup>[134,135]</sup> In turn, distal arterial narrowing increases mean pulmonary artery pressure, which dilates the proximal arteries, increasing circumferential stress and promoting SMC-mediated wall thickening,<sup>[12,136]</sup> which increase stiffness.<sup>[16-18,47]</sup>

### SUMMARY AND FUTURE DIRECTIONS

It is vital to examine the structure-function relationship of pulmonary vascular system in the context of RV function, which predicts clinical outcomes and ultimately determines mortality in PH. Recently, pulmonary arterial stiffness has gained increasing recognition due to its clinical relevance to PH outcomes; however, studies that examine local properties such as stiffness typically do not quantify global cardiopulmonary function so the relationship between the two remains poorly defined. Studies that quantify impedance, as a more comprehensive metric of RV afterload, and vascular-ventricular coupling, to assess the efficiency of the whole cardiopulmonary system, are required to address this gap in our knowledge. Impedance provides insight into steady (distal) and pulsatile (proximal) components of the pulmonary hemodynamics; VVC measurements demonstrate the impact of pulmonary hemodynamics on RV function and vice versa. In addition, the role of arterial viscoelasticity in VVC efficiency remains to be determined. Furthermore,

at the cellular and molecular level, it would be useful to identify the cells and cellular signaling pathways that control PA stiffening and should be targets of treatment. Finally, a better understanding of the impact of pulmonary hemodynamics on RV function and vice versa may allow more accurate prognoses of RV failure. That is, the hemodynamic indicators of the transition from adaptive ventricular remodeling to maladaptive ventricular remodeling, the precursor of ventricular failure, may lead to better care and treatment for patients with PH.

### REFERENCES

- McLaughlin VV, Archer SL, Badesch DB, Barst RJ, Farber HW, Lindner JR, et al. ACCF/AHA 2009 expert consensus document on pulmonary hypertension: A report of the American College of Cardiology Foundation Task Force on Expert Consensus Documents and the American Heart Association: Developed in collaboration with the American College of Chest Physicians, American Thoracic Society, Inc., and the Pulmonary Hypertension Association. *Circulation* 2009;119:2250-94.
- D'Alonzo GE, Barst RJ, Ayres SM, Bergofsky EH, Brundage BH, Detre KM, et al. Survival in patients with primary pulmonary hypertension. Results from a national prospective registry. *Ann Intern Med* 1991;115:343-9.
- Schannwell CM, Steiner S, Strauer BE. Diagnostics in pulmonary hypertension. *J Physiol Pharmacol* 2007;58 Suppl 5:591-602.
- Ghio S, Gavazzi A, Campana C, Inserra C, Klersy C, Sebastiani R, et al. Independent and additive prognostic value of right ventricular systolic function and pulmonary artery pressure in patients with chronic heart failure. *J Am Coll Cardiol* 2001;37:183-8.
- Forfia PR, Fisher MR, Mathai SC, Houston-Harris T, Hemnes AR, Borlaug BA, et al. Tricuspid annular displacement predicts survival in pulmonary hypertension. *Am J Respir Crit Care Med* 2006;174:1034-41.
- Voelkel NF, Quaife RA, Leinwand LA, Barst RJ, McGoon MD, Meldrum DR, et al. Right ventricular function and failure: Report of a National Heart, Lung, and Blood Institute working group on cellular and molecular mechanisms of right heart failure. *Circulation* 2006;114:1883-91.
- Blyth KG, Syeed R, Chalmers J, Foster JE, Saba T, Naeije R, et al. Pulmonary arterial pulse pressure and mortality in pulmonary arterial hypertension. *Respir Med* 2007;101:2495-501.
- Van Wolferen SA, Marcus JT, Boonstra A, Marques KM, Bronzwaer JG, Spreeuwenberg MD, et al. Prognostic value of right ventricular mass, volume, and function in idiopathic pulmonary arterial hypertension. *Eur Heart J* 2007;28:1250-7.
- Hemnes AR, Champion HC. Right heart function and haemodynamics in pulmonary hypertension. *Int J Clin Pract Suppl* 2008:11-9.
- Nichols WW, O'Rourke MF. McDonald's Blood Flow in Arteries: Theoretical, Experimental and Clinical Principles. 5th ed. London, UK: A Hodder Arnold Publication; 2005.
- Milnor WR, Bergel DH, Bargainer JD. Hydraulic power associated with pulmonary blood flow and its relation to heart rate. *Circ Res* 1966;19:467-80.
- Stenmark KR, Fagan KA, Frid MG. Hypoxia-induced pulmonary vascular remodeling: Cellular and molecular mechanisms. *Circ Res* 2006;99:675-91.
- Mahapatra S, Nishimura RA, Sorajja P, Cha S, McGoon MD. Relationship of pulmonary arterial capacitance and mortality in idiopathic pulmonary arterial hypertension. *J Am Coll Cardiol* 2006;47:799-803.
- Gan CT, Lankhaar JW, Westerhof N, Marcus JT, Becker A, Twisk JW, et al. Noninvasively assessed pulmonary artery stiffness predicts mortality in pulmonary arterial hypertension. *Chest* 2007;132:1906-12.
- Shimoda LA, Norins NA, Madden JA. Flow-induced responses in cat isolated pulmonary arteries. *J Appl Physiol* 1997;83:1617-22.
- Kobs RW, Muvarak NE, Eickhoff JC, Chesler NC. Linked mechanical and biological aspects of remodeling in mouse pulmonary arteries with hypoxia-induced hypertension. *Am J Physiol Heart Circ Physiol* 2005;288:H1209-17.
- Ooi CY, Wang Z, Tabima DM, Eickhoff JC, Chesler NC. The role of collagen in extralobar pulmonary artery stiffening in response to hypoxia-induced pulmonary hypertension. *Am J Physiol Heart Circ Physiol* 2010;299:H1823-31.
- Tabima DM, Chesler NC. The effects of vasoactivity and hypoxic

- pulmonary hypertension on extralobar pulmonary artery biomechanics. *J Biomech* 2010;43:1864-9.
19. Hunter KS, Albietsz JA, Lee PF, Lanning CJ, Lammers SR, Hofmeister SH, *et al*. In vivo measurement of proximal pulmonary artery elastic modulus in the neonatal calf model of pulmonary hypertension: Development and ex vivo validation. *J Appl Physiol* 2010;108:968-75.
  20. Faury G, Maher GM, Li DY, Keating MT, Mecham RP, Boyle WA. Relation between outer and luminal diameter in cannulated arteries. *Am J Physiol* 1999;277:H1745-53.
  21. Hudetz AG. Incremental elastic modulus for orthotropic incompressible arteries. *J Biomech* 1979;12:651-5.
  22. Schulze-Bauer CA, Holzapfel GA. Determination of constitutive equations for human arteries from clinical data. *J Biomech* 2003;36:165-9.
  23. Yuan JG, Hales CA, Rich S, Archer SL, West JB, editors. *Textbook of Pulmonary Vascular Disease*. New York: Springer; 2011.
  24. Herrera EA, Pulgar VM, Riquelme RA, Sanhueza EM, Reyes RV, Ebensperger G, *et al*. High-altitude chronic hypoxia during gestation and after birth modifies cardiovascular responses in newborn sheep. *Am J Physiol Regul Integr Comp Physiol* 2007;292:R2234-40.
  25. Lally C, Reid AJ, Prendergast PJ. Elastic behavior of porcine coronary artery tissue under uniaxial and equibiaxial tension. *Ann Biomed Eng* 2004;32:1355-64.
  26. Lammers SR, Kao PH, Qi HJ, Hunter K, Lanning C, Albietsz J, *et al*. Changes in the structure-function relationship of elastin and its impact on the proximal pulmonary arterial mechanics of hypertensive calves. *Am J Physiol Heart Circ Physiol* 2008;295:H1451-9.
  27. Griffith SL, Rhoades RA, Packer CS. Pulmonary arterial smooth muscle contractility in hypoxia-induced pulmonary hypertension. *J Appl Physiol* 1994;77:406-14.
  28. Packer CS, Roepke JE, Oberlies NH, Rhoades RA. Myosin isoform shifts and decreased reactivity in hypoxia-induced hypertensive pulmonary arterial muscle. *Am J Physiol* 1998;274:L775-85.
  29. Boutouyrie P, Boumaza S, Challande P, Lacolley P, Laurent S. Smooth muscle tone and arterial wall viscosity: An in vivo/in vitro study. *Hypertension* 1998;32:360-4.
  30. Jackson ZS, Gotlieb AI, Langille BL. Wall tissue remodeling regulates longitudinal tension in arteries. *Circ Res* 2002;90:918-25.
  31. Dye WW, Gleason RL, Wilson E, Humphrey JD. Altered biomechanical properties of carotid arteries in two mouse models of muscular dystrophy. *J Appl Physiol* 2007;103:664-72.
  32. Humphrey JD, Eberth JF, Dye WW, Gleason RL. Fundamental role of axial stress in compensatory adaptations by arteries. *J Biomech* 2009;42:1-8.
  33. Feigl EO, Peterson LH, Jones AW. Mechanical and chemical properties of arteries in experimental hypertension. *J Clin Invest* 1963;42:1640-7.
  34. Silver FH, Christiansen DL, Buntin CM. Mechanical properties of the aorta: A review. *Crit Rev Biomed Eng* 1989;17:323-58.
  35. Weinberg CE, Hertzberg JR, Shandas R. Use of intravascular ultrasound to measure local compliance of the pediatric pulmonary artery: In vitro studies. *J Am Soc Echocardiogr* 2002;15:1507-14.
  36. Hayashi K, Handa H, Nagasawa S, Okumura A, Moritake K. Stiffness and elastic behavior of human intracranial and extracranial arteries. *J Biomech* 1980;13:175-84.
  37. Kawasaki T, Sasayama S, Yagi S, Asakawa T, Hirai T. Non-invasive assessment of the age related changes in stiffness of major branches of the human arteries. *Cardiovasc Res* 1987;21:678-87.
  38. Hayoz D, Rutschmann B, Perret F, Niederberger M, Tardy Y, Mooser V, *et al*. Conduit artery compliance and distensibility are not necessarily reduced in hypertension. *Hypertension* 1992;20:1-6.
  39. Laurent S, Caviezel B, Beck L, Girerd X, Billaud E, Boutouyrie P, *et al*. Carotid artery distensibility and distending pressure in hypertensive humans. *Hypertension* 1994;23:878-83.
  40. Berger RM, Cromme-Dijkhuis AH, Hop WC, Kruit MN, Hess J. Pulmonary arterial wall distensibility assessed by intravascular ultrasound in children with congenital heart disease: An indicator for pulmonary vascular disease? *Chest* 2002;122:549-57.
  41. Zheng D, Murray A. Peripheral arterial volume distensibility: Significant differences with age and blood pressure measured using an applied external pressure. *Physiol Meas* 2011;32:499-512.
  42. Tardy Y, Meister JJ, Perret F, Brunner HR, Arditi M. Non-invasive estimate of the mechanical properties of peripheral arteries from ultrasonic and photoplethysmographic measurements. *Clin Phys Physiol Meas* 1991;12:39-54.
  43. Chemla D, Hébert JL, Coirault C, Zamani K, Suard I, Colin P, *et al*. Total arterial compliance estimated by stroke volume-to-aortic pulse pressure ratio in humans. *Am J Physiol* 1998;274:H500-5.
  44. Hayashi K, Naiki T. Adaptation and remodeling of vascular wall: Biomechanical response to hypertension. *J Mech Behav Biomed Mater* 2009;2:3-19.
  45. Bergel DH. The static elastic properties of the arterial wall. *J Physiol* 1961;156:445-57.
  46. Chesler NC, Thompson-Figueroa J, Millburne K. Measurements of mouse pulmonary artery biomechanics. *J Biomech Eng* 2004;126:309-14.
  47. Kobs RW, Chesler NC. The mechanobiology of pulmonary vascular remodeling in the congenital absence of eNOS. *Biomech Model Mechanobiol* 2006;5:217-25.
  48. Drexler ES, Bischoff JE, Slifka AJ, McCowan CN, Quinn TP, Shandas R, *et al*. Stiffening of the extrapulmonary arteries from rats in chronic hypoxic pulmonary hypertension. *J Res Natl Inst Stand Technol* 2008;113:239-49.
  49. Glaser E, Lacolley P, Boutouyrie P, Sacunha R, Lucet B, Safar ME, *et al*. Dynamic versus static compliance of the carotid artery in living Wistar-Kyoto rats. *J Vasc Res* 1995;32:254-65.
  50. LeROYD BM, Taylor MG. Alterations with age in the viscoelastic properties of human arterial walls. *Circ Res* 1966;18:278-92.
  51. Armentano R, Megnien JL, Simon A, Bellenfant F, Barra J, Levenson J. Effects of hypertension on viscoelasticity of carotid and femoral arteries in humans. *Hypertension* 1995;26:48-54.
  52. Armentano RL, Graf S, Barra JG, Velikovskiy G, Baglivo H, Sánchez R, *et al*. Carotid wall viscosity increase is related to intima-media thickening in hypertensive patients. *Hypertension* 1998;31:534-9.
  53. Bia D, Grignola JC, Armentano RL, Gines FF. Improved pulmonary artery buffering function during phenylephrine-induced pulmonary hypertension. *Mol Cell Biochem* 2003;246:19-24.
  54. Grignola JC, Bia D, Gines F, Armentano RL. Acute pulmonary hypertension: Protective role of vascular smooth muscle activation. *Rev Esp Cardiol* 2003;56:1077-84.
  55. Wang Z, Chesler NC. Role of collagen content and cross-linking in large pulmonary arterial stiffening after chronic hypoxia. *Biomech Model Mechanobiol* 2011. [In press\*\*\*].
  56. Armentano RL, Barra JG, Pessana FM, Craiem DO, Graf S, Santana DB, *et al*. Smart smooth muscle spring-dampers. Smooth muscle smart filtering helps to more efficiently protect the arterial wall. *IEEE Eng Med Biol Mag* 2007;26:62-70.
  57. Taylor MG. Wave transmission through an assembly of randomly branching elastic tubes. *Biophys J* 1966;6:697-716.
  58. Pace JB. Sympathetic control of pulmonary vascular impedance in anesthetized dogs. *Circ Res* 1971;29:555-68.
  59. Dujardin JP, Stone DN, Forcino CD, Paul LT, Pieper HP. Effects of blood volume changes on characteristic impedance of the pulmonary artery. *Am J Physiol* 1982;242:H197-202.
  60. Ewalenko P, Stefanidis C, Holoye A, Brimiouille S, Naeije R. Pulmonary vascular impedance vs. resistance in hypoxic and hyperoxic dogs: Effects of propofol and isoflurane. *J Appl Physiol* 1993;74:2188-93.
  61. Ewalenko P, Brimiouille S, Delcroix M, Lejeune P, Naeije R. Comparison of the effects of isoflurane with those of propofol on pulmonary vascular impedance in experimental embolic pulmonary hypertension. *Br J Anaesth* 1997;79:625-30.
  62. Maggiorini M, Brimiouille S, De Canniere D, Delcroix M, Naeije R. Effects of pulmonary embolism on pulmonary vascular impedance in dogs and minipigs. *J Appl Physiol* 1998;84:815-21.
  63. Zhao L, Mason NA, Morrell NW, Kojonazarov B, Sadykov A, Maripov A, *et al*. Sildenafil inhibits hypoxia-induced pulmonary hypertension. *Circulation* 2001;104:424-8.
  64. Zhao L, Crawley DE, Hughes JM, Evans TW, Winter RJ. Endothelium-derived relaxing factor activity in rat lung during hypoxic pulmonary vascular remodeling. *J Appl Physiol* 1993;74:1061-5.
  65. Nossaman BD, Feng CJ, Kadowitz PJ. Analysis of responses to bradykinin and influence of HOE 140 in the isolated perfused rat lung. *Am J Physiol* 1994;266:H2452-61.
  66. Berkenbosch JW, Baribeau J, Perreault T. Decreased synthesis and vasodilation to nitric oxide in piglets with hypoxia-induced pulmonary hypertension. *Am J Physiol Lung Cell Mol Physiol* 2000;278:L276-83.
  67. Fagan KA, Oka M, Bauer NR, Gebb SA, Ivy DD, Morris KG, *et al*. Attenuation of acute hypoxic pulmonary vasoconstriction and hypoxic pulmonary hypertension in mice by inhibition of Rho-kinase. *Am J Physiol Lung Cell Mol Physiol* 2004;287:L656-64.
  68. Tuschcherer HA, Vanderpool RR, Chesler NC. Pulmonary vascular remodeling in isolated mouse lungs: Effects on pulsatile pressure-flow relationships. *J Biomech* 2007;40:993-1001.

69. Vanderpool RR, Kim AR, Molthen R, Chesler NC. Effects of acute Rho kinase inhibition on chronic hypoxia-induced changes in proximal and distal pulmonary arterial structure and function. *J Appl Physiol* 2010;110:188-98.
70. Vanderpool RR, Naeije R, Chesler NC. Impedance in isolated mouse lungs for the determination of site of action of vasoactive agents and disease. *Ann Biomed Eng* 2011;38:1854-61.
71. Vanderpool RR, Chesler NC. Characterization of the isolated, ventilated, and instrumented mouse lung perfused with pulsatile flow. *J Vis Exp* 2011;pii:2690.
72. Vanderpool RR, Kim AR, Molthen R, Chesler NC. Effects of acute Rho kinase inhibition on chronic hypoxia-induced changes in proximal and distal pulmonary arterial structure and function. *J Appl Physiol* 2011;110:188-98.
73. Linehan JH, Haworth ST, Nelin LD, Krenz GS, Dawson CA. A simple distensible vessel model for interpreting pulmonary vascular pressure-flow curves. *J Appl Physiol* 1992;73:987-94.
74. Reeves JT, Linehan JH, Stenmark KR. Distensibility of the normal human lung circulation during exercise. *Am J Physiol Lung Cell Mol Physiol* 2005;288:L419-25.
75. Chesler NC, Argiento P, Vanderpool R, D'Alto M, Naeije R. How to measure peripheral pulmonary vascular mechanics. *Conf Proc IEEE Eng Med Biol Soc* 2009;2009:173-6.
76. O'Rourke MF. Vascular impedance in studies of arterial and cardiac function. *Physiol Rev* 1982;62:570-623.
77. Nakayama Y, Nakanishi N, Sugimachi M, Takaki H, Kyotani S, Satoh T, et al. Characteristics of pulmonary artery pressure waveform for differential diagnosis of chronic pulmonary thromboembolism and primary pulmonary hypertension. *J Am Coll Cardiol* 1997;29:1311-6.
78. Huez S, Brimiouille S, Naeije R, Vachiery JL. Feasibility of routine pulmonary arterial impedance measurements in pulmonary hypertension. *Chest* 2004;125:2121-8.
79. Laskey WK, Ferrari VA, Palevsky HI, Kussmaul WG. Pulmonary artery hemodynamics in primary pulmonary hypertension. *J Am Coll Cardiol* 1993;21:406-12.
80. Syeed R, Reeves JT, Welsh D, Raeside D, Johnson MK, Peacock AJ. The relationship between the components of pulmonary artery pressure remains constant under all conditions in both health and disease. *Chest* 2008;133:633-9.
81. Naeije R, Maarek JM, Chang HK. Pulmonary vascular impedance in microembolic pulmonary hypertension: Effects of synchronous high-frequency jet ventilation. *Respir Physiol* 1990;79:205-17.
82. Wauthy P, Pagnamenta A, Vassalli F, Naeije R, Brimiouille S. Right ventricular adaptation to pulmonary hypertension: An interspecies comparison. *Am J Physiol Heart Circ Physiol* 2004;286:H1441-7.
83. Hunter KS, Lee PF, Lanning CJ, Ivy DD, Kirby KS, Clausen LR, et al. Pulmonary vascular input impedance is a combined measure of pulmonary vascular resistance and stiffness and predicts clinical outcomes better than pulmonary vascular resistance alone in pediatric patients with pulmonary hypertension. *Am Heart J* 2008;155:166-74.
84. Champion HC, Michelakis ED, Hassoun PM. Comprehensive invasive and noninvasive approach to the right ventricle-pulmonary circulation unit: State of the art and clinical and research implications. *Circulation* 2009;120:992-1007.
85. Chesler NC, Roldan A, Vanderpool RR, Naeije R. How to measure pulmonary vascular and right ventricular function. *Conf Proc IEEE Eng Med Biol Soc* 2009;2009:177-80.
86. Milnor WR. *Hemodynamics*. Baltimore, MD: Williams & Wilkins; 1989.
87. Milnor WR, Conti CF, Lewis KB, O'Rourke MF. Pulmonary arterial pulse wave velocity and impedance in man. *Circ Res* 1969;25:637-49.
88. Vanderpool RR, Naeije R, Chesler NC. Impedance in isolated mouse lungs for the determination of site of action of vasoactive agents and disease. *Ann Biomed Eng* 2010;38:1854-61.
89. Zuckerman BD, Orton EC, Latham LP, Barbieri CC, Stenmark KR, Reeves JT. Pulmonary vascular impedance and wave reflections in the hypoxic calf. *J Appl Physiol* 1992;72:2118-27.
90. Pagnamenta A, Fesler P, Vandinivit A, Brimiouille S, Naeije R. Pulmonary vascular effects of dobutamine in experimental pulmonary hypertension. *Crit Care Med* 2003;31:1140-6.
91. Lankhaar JW, Westerhof N, Faes TJ, Marques KM, Marcus JT, Postmus PE, et al. Quantification of right ventricular afterload in patients with and without pulmonary hypertension. *Am J Physiol Heart Circ Physiol* 2006;291:H1731-7.
92. Milnor WR, Conti CR, Lewis KB, O'Rourke MF. Pulmonary arterial pulse wave velocity and impedance in man. *Circ Res* 1969;25:637-49.
93. Weinberg CE, Hertzberg JR, Ivy DD, Kirby KS, Chan KC, Valdes-Cruz L, et al. Extraction of pulmonary vascular compliance, pulmonary vascular resistance, and right ventricular work from single-pressure and Doppler flow measurements in children with pulmonary hypertension: A new method for evaluating reactivity: In vitro and clinical studies. *Circulation* 2004;110:2609-17.
94. Naeije R, Vizza D. Current perspectives modern hemodynamic evaluation of the pulmonary circulation. Application to pulmonary arterial hypertension and embolic pulmonary hypertension. *Ital Heart J* 2005;6:784-8.
95. Nakayama Y, Nakanishi N, Hayashi T, Nagaya N, Sakamaki F, Satoh N, et al. Pulmonary artery reflection for differentially diagnosing primary pulmonary hypertension and chronic pulmonary thromboembolism. *J Am Coll Cardiol* 2001;38:214-8.
96. Castelain V, Herve P, Lecarpentier Y, Duroux P, Simonneau G, Chemla D. Pulmonary artery pulse pressure and wave reflection in chronic pulmonary thromboembolism and primary pulmonary hypertension. *J Am Coll Cardiol* 2001;37:1085-92.
97. Mitchell GF, Pfeffer MA, Westerhof N, Pfeffer JM. Measurement of aortic input impedance in rats. *Am J Physiol* 1994;267:H1907-15.
98. Mitchell GF. Clinical achievements of impedance analysis. *Med Biol Eng Comput* 2009;47:153-63.
99. Westerhof N, Sipkema P, van den Bos GC, Elzinga G. Forward and backward waves in the arterial system. *Cardiovasc Res* 1972;6:648-56.
100. Zamir M. *The physics of pulsatile flow*. New York: Springer; 2000.
101. Segers P, De Backer J, Devos D, Rabben SI, Gillebert TC, Van Bortel LM, et al. Aortic reflection coefficients and their association with global indexes of wave reflection in healthy controls and patients with Marfan's syndrome. *Am J Physiol Heart Circ Physiol* 2006;290:H2385-92.
102. Tabima DM, Vanderpool RR, Chesler NC. Comparing pulmonary input and characteristic impedance determined in the time and frequency domains: The effects of chronic hypoxia. In: American Thoracic Society (ATS) 2010 International Conference; 2010: Am J Respir Crit Care Med; 2010. p. 181:A6271.
103. Benetos A, Safar M, Rudnichi A, Smulyan H, Richard JL, Ducimetière P, et al. Pulse pressure: A predictor of long-term cardiovascular mortality in a French male population. *Hypertension* 1997;30:1410-5.
104. Mitchell GF, Moyé LA, Braunwald E, Rouleau JL, Bernstein V, Geltman EM, et al. Sphygmomanometrically determined pulse pressure is a powerful independent predictor of recurrent events after myocardial infarction in patients with impaired left ventricular function. SAVE investigators. Survival and Ventricular Enlargement. *Circulation* 1997;96:4254-60.
105. Grant BJ, Paradowski LJ, Fitzpatrick JM. Effect of perivascular electromagnetic flow probes on pulmonary hemodynamics. *J Appl Physiol* 1988;65:1885-90.
106. Morita S, Kuboyama I, Asou T, Tokunaga K, Nose Y, Nakamura M, et al. The effect of extraanatomic bypass on aortic input impedance studied in open chest dogs. Should the vascular prosthesis be compliant to unload the left ventricle? *J Thorac Cardiovasc Surg* 1991;102:774-83.
107. Grant BJ, Lieber BB. Clinical significance of pulmonary arterial input impedance. *Eur Respir J* 1996;9:2196-9.
108. Domanski MJ, Davis BR, Pfeffer MA, Kastantin M, Mitchell GF. Isolated systolic hypertension: Prognostic information provided by pulse pressure. *Hypertension* 1999;34:375-80.
109. Furuno Y, Nagamoto Y, Fujita M, Kaku T, Sakurai S, Kuroiwa A. Reflection as a cause of mid-systolic deceleration of pulmonary flow wave in dogs with acute pulmonary hypertension: Comparison of pulmonary artery constriction with pulmonary embolisation. *Cardiovasc Res* 1991;25:118-24.
110. Remington JW, Noback CR. Volume elasticity characteristics of the human aorta and prediction of the stroke volume from the pressure pulse. *Am J Physiol* 1948;153:298-308.
111. Saouti N, Westerhof N, Postmus PE, Vonk-Noordegraaf A. The arterial load in pulmonary hypertension. *Eur Respir Rev* 2010;19:197-203.
112. Domanski MJ, Mitchell GF, Norman JE, Exner DV, Pitt B, Pfeffer MA. Independent prognostic information provided by sphygmomanometrically determined pulse pressure and mean arterial pressure in patients with left ventricular dysfunction. *J Am Coll Cardiol* 1999;33:951-8.
113. Mahapatra S, Nishimura RA, Oh JK, McGoon MD. The prognostic value of pulmonary vascular capacitance determined by Doppler echocardiography in patients with pulmonary arterial hypertension. *J Am Soc Echocardiogr* 2006;19:1045-50.
114. Lankhaar JW, Westerhof N, Faes TJ, Gan CT, Marques KM, Boonstra A, et al. Pulmonary vascular resistance and compliance stay inversely related during treatment of pulmonary hypertension. *Eur Heart J* 2008;29:1688-95.

115. Humbert M, Morrell NW, Archer SL, Stenmark KR, MacLean MR, Lang IM, et al. Cellular and molecular pathobiology of pulmonary arterial hypertension. *J Am Coll Cardiol* 2004;43 Suppl 12:13S-24.
116. Rabinovitch M. Molecular pathogenesis of pulmonary arterial hypertension. *J Clin Invest* 2008;118:2372-9.
117. Poiani GJ, Tozzi CA, Yohn SE, Pierce RA, Belsky SA, Berg RA, et al. Collagen and elastin metabolism in hypertensive pulmonary arteries of rats. *Circ Res* 1990;66:968-78.
118. Tozzi CA, Christiansen DL, Poiani GJ, Riley DJ. Excess collagen in hypertensive pulmonary arteries decreases vascular distensibility. *Am J Respir Crit Care Med* 1994;149:1317-26.
119. Merklinger SL, Wagner RA, Spiekerkoetter E, Hinek A, Knutsen RH, Kabir MG, et al. Increased fibulin-5 and elastin in S100A4/Mts1 mice with pulmonary hypertension. *Circ Res* 2005;97:596-604.
120. Peterson LH, Jensen RE, Parnell J. Mechanical properties of arteries in vivo. *Circ Res* 1960;8:622-39.
121. Tucker A, Migally N, Wright ML, Greenlees KJ. Pulmonary vascular changes in young and aging rats exposed to 5,486 m altitude. *Respiration* 1984;46:246-57.
122. Poiani GJ, Tozzi CA, Choe JK, Yohn SE, Riley DJ. An antifibrotic agent reduces blood pressure in established pulmonary hypertension in the rat. *J Appl Physiol* 1990;68:1542-7.
123. Leung DY, Glagov S, Mathews MB. Elastin and collagen accumulation in rabbit ascending aorta and pulmonary trunk during postnatal growth. Correlation of cellular synthetic response with medial tension. *Circ Res* 1977;41:316-23.
124. Shifren A, Durmowicz AG, Knutsen RH, Faury G, Mecham RP. Elastin insufficiency predisposes to elevated pulmonary circulatory pressures through changes in elastic artery structure. *J Appl Physiol* 2008;105:1610-9.
125. Madden JA, Vadula MS, Kurup VP. Effects of hypoxia and other vasoactive agents on pulmonary and cerebral artery smooth muscle cells. *Am J Physiol* 1992;263:L384-93.
126. Nagaoka T, Morio Y, Casanova N, Bauer N, Gebb S, McMurtry I, et al. Rho/Rho kinase signaling mediates increased basal pulmonary vascular tone in chronically hypoxic rats. *Am J Physiol Lung Cell Mol Physiol* 2004;287:L665-72.
127. Saouti N, Westerhof N, Helderma F, Marcus JT, Boonstra A, Postmus PE, et al. Right ventricular oscillatory power is a constant fraction of total power irrespective of pulmonary artery pressure. *Am J Respir Crit Care Med* 2010;182:1315-20.
128. Mitchell GF. Pulse pressure, arterial compliance and cardiovascular morbidity and mortality. *Curr Opin Nephrol Hypertens* 1999;8:335-42.
129. Bernardo BC, Weeks KL, Pretorius L, McMullen JR. Molecular distinction between physiological and pathological cardiac hypertrophy: Experimental findings and therapeutic strategies. *Pharmacol Ther* 2010;128:191-227.
130. Sagawa K, Maughan L, Suga H, Sunagawa K. *Cardiac Contraction and the Pressure-Volume Relationship*: Oxford University Press; 1988.
131. Segers P, Georgakopoulos D, Afanasyeva M, Champion HC, Judge DP, Millar HD, et al. Conductance catheter-based assessment of arterial input impedance, arterial function, and ventricular-vascular interaction in mice. *Am J Physiol Heart Circ Physiol* 2005;288:H1157-64.
132. Tabima DM, Hacker TA, Chesler NC. Measuring right ventricular function in the normal and hypertensive mouse hearts using admittance-derived pressure-volume loops. *Am J Physiol Heart Circ Physiol* 2010;299:H2069-75.
133. Li M, Scott DE, Shandas R, Stenmark KR, Tan W. High pulsatility flow induces adhesion molecule and cytokine mRNA expression in distal pulmonary artery endothelial cells. *Ann Biomed Eng* 2009;37:1082-92.
134. O'Rourke MF, Safar ME. Relationship between aortic stiffening and microvascular disease in brain and kidney: Cause and logic of therapy. *Hypertension* 2005;46:200-4.
135. O'Rourke MF. Arterial aging: Pathophysiological principles. *Vasc Med* 2007;12:329-41.
136. Humphrey JD. Mechanisms of arterial remodeling in hypertension: Coupled roles of wall shear and intramural stress. *Hypertension* 2008;52:195-200.

**Source of Support:** This work is in part supported by a grant from the National Institutes of Health (R01HL86939 to N.C.C.) and a Postdoctoral Fellowship from the American Heart Association Midwest Affiliation (10POST2640148 to Z.W.). **Conflict of Interest:** None declared.