



OPEN ACCESS

New horizons for newborn brain protection: enhancing endogenous neuroprotection

K Jane Hassell,¹ Mojgan Ezzati,¹ Daniel Alonso-Alconada,¹ Derek J Hausenloy,² Nicola J Robertson¹

¹Institute for Women's Health, University College London, London, UK

²The Hatter Cardiovascular Institute, Institute of Cardiovascular Science, NIHR University College London Hospitals Biomedical Research Centre, University College London Hospital & Medical School, London, UK

Correspondence to

Professor Nicola J Robertson, Institute for Women's Health, 74 Huntley Street, 2nd floor, room 239, University College London, London WC1E 6HX, UK; n.robertson@ucl.ac.uk

Received 23 October 2014
Revised 21 January 2015
Accepted 28 January 2015
Published Online First
10 June 2015

ABSTRACT

Intrapartum-related events are the third leading cause of childhood mortality worldwide and result in one million neurodisabled survivors each year. Infants exposed to a perinatal insult typically present with neonatal encephalopathy (NE). The contribution of pure hypoxia-ischaemia (HI) to NE has been debated; over the last decade, the sensitising effect of inflammation in the aetiology of NE and neurodisability is recognised. Therapeutic hypothermia is standard care for NE in high-income countries; however, its benefit in encephalopathic babies with sepsis or in those born following chorioamnionitis is unclear. It is now recognised that the phases of brain injury extend into a tertiary phase, which lasts for weeks to years after the initial insult and opens up new possibilities for therapy.

There has been a recent focus on understanding endogenous neuroprotection and how to boost it or to supplement its effectors therapeutically once damage to the brain has occurred as in NE. In this review, we focus on strategies that can augment the body's own endogenous neuroprotection. We discuss in particular remote ischaemic postconditioning whereby endogenous brain tolerance can be activated through hypoxia/reperfusion stimuli started immediately after the index hypoxic-ischaemic insult. Therapeutic hypothermia, melatonin, erythropoietin and cannabinoids are examples of ways we can supplement the endogenous response to HI to obtain its full neuroprotective potential. Achieving the correct balance of interventions at the correct time in relation to the nature and stage of injury will be a significant challenge in the next decade.

BACKGROUND

Intrapartum-related insults at full term such as hypoxia-ischaemia (HI) are the third leading cause of global child deaths.¹ Each year, over 0.7 million affected newborns die and 1.15 million develop acute disordered brain function known as neonatal encephalopathy (NE).² NE is the second commonest preventable cause of childhood neurodisability worldwide³ with profound psychosocial and economic consequences for families and society. Protecting the newborn brain from injury around the time of birth is a global health priority.^{4 5}

TERM NEWBORN BRAIN INJURY: CAUSES, PATHOGENESIS AND MANAGEMENT

NE is a descriptive term for neurological dysfunction in the newborn infant, manifested by symptoms including difficulty with initiating and maintaining respiration, depression of tone and reflexes, subnormal level of consciousness, poor feeding and seizures.⁶ NE has a complex and

multifactorial aetiology. For over two decades, perinatal neuroprotection research has focused on pure hypoxic-ischaemic brain injury; however, accumulating preclinical^{7 8} and clinical⁹ evidences suggest the critical importance of the sensitising effect of inflammation.

The clinical signs of NE progress after a latent period of hours to days. This timing of the evolution of clinical signs is thought to reflect brain energy levels and the cascade of neurochemical processes responsible for brain injury. These are summarised in [figure 1](#) and described in more detail below.

Acute HI

During the acute hypoxic-ischaemic insult, some cells undergo primary cell death, the magnitude of which will depend on the severity and duration of HI. In the absence of substrates (oxygen, glucose), the neuron's supply of high-energy metabolites such as ATP falls below a critical threshold. The Na⁺/K⁺ ATP-dependent pump begins to fail, neuronal depolarisation occurs and the synaptic cleft floods with glutamate, which activates the N-methyl-D-aspartate (NMDA) receptor. Toxic cytoplasmic Ca²⁺ concentrations arise through several mechanisms, including overactivation of glutamate receptors (NMDA, α -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid (AMPA)), other channels and transporters, or through release from internal stores through physical damage to mitochondria and endoplasmic reticulum¹⁰; the increased Ca²⁺ triggers many downstream neurotoxic cascades. As well as generating an osmotic gradient that leads to oedema and lysis of cells, Ca²⁺ activates nitric oxide synthase, which in turn generates high levels of the toxic reactive oxygen species nitric oxide (NO•). At high concentrations, NO• reacts with superoxide (O^{•-}) to produce peroxynitrite (ONOO⁻), which damages mitochondria via peroxidation and nitrosylation of membrane lipids. Consequently, mitochondrial dysfunction and membrane depolarisation develop with further release of O^{•-} and decline in endogenous anti-oxidants such as glutathione. Ca²⁺ also triggers the activation of cytosolic phospholipases, which increase eicosanoid release leading to inflammation.

Latent phase

After reperfusion, the initial hypoxia-induced cytotoxic oedema and accumulation of excitatory amino acids partially resolve in 30–60 min, with apparent recovery of cerebral oxidative metabolism. It is thought that the neurotoxic cascade is largely inhibited during the latent phase, when there is



Open Access
Scan to access more
free content



CrossMark

To cite: Hassell KJ, Ezzati M, Alonso-Alconada D, et al. *Arch Dis Child Fetal Neonatal Ed* 2015;**100**:F541–F552.

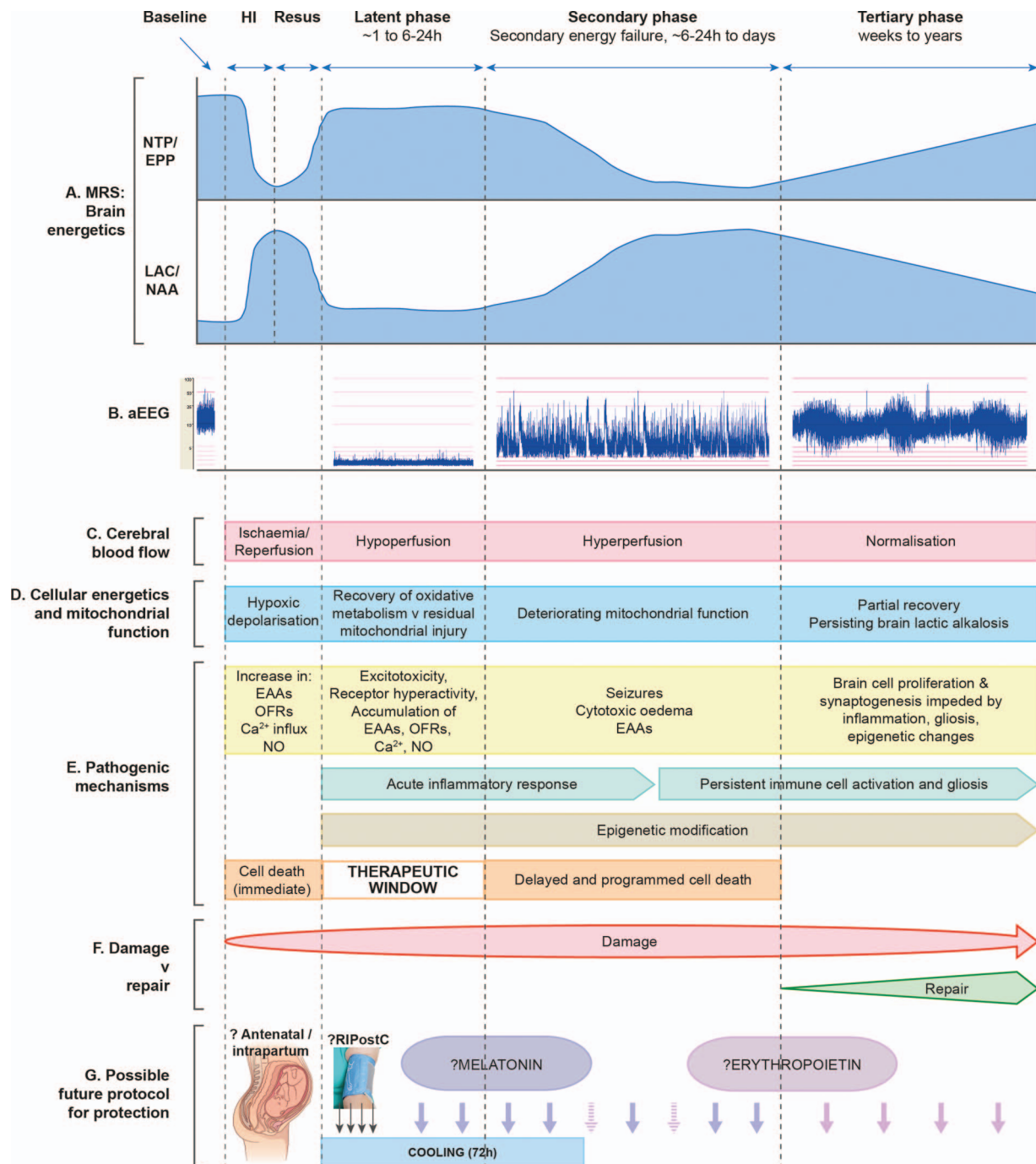


Figure 1 Schematic diagram illustrating the different pathological phases of cerebral injury after cerebral HI. The primary phase (acute HI), latent phase, secondary energy failure phase and tertiary brain injury phase are shown. (A) Magnetic resonance spectra showing the biphasic pattern of NTP/EPP decline and lactate/NAA increase during primary and secondary phases following HI insult. Persisting lactic alkalosis is shown in tertiary phase. (B) Amplitude-integrated EEG showing normal trace at baseline, flat tract following HI, burst-suppression pattern in latent phase, emergence of seizures in secondary phase and normalisation with sleep–wake cycling in tertiary phase. (C) Following HI, there is a period of hypoperfusion associated with hypometabolism during latent phase, followed by relative hyperperfusion in secondary phase. (D) Cellular energetics and mitochondrial function are reflected in the biphasic response shown on magnetic resonance spectroscopy (A), with a period of recovery in latent phase followed by deterioration in secondary phase. There is partial recovery in tertiary phase. (E) The most important pathogenic changes are shown for each phase (see main text for description), including generation of toxic free radical species, accumulation of EAAs, cytotoxic oedema, seizures and inflammation. Cell lysis occurs immediately following HI, while programmed cell death occurs in secondary phase; latent phase provides a therapeutic window. Persisting inflammation and epigenetic changes impede long-term repair. (F) Damage is maximal in the secondary phase, but persists into the tertiary phase as inflammation and gliosis evolve. (G) In the future, neuroprotective treatments are likely to involve a ‘cocktail’ of therapies to be administered intrapartum, in the latent phase to prevent secondary energy failure and through secondary and tertiary phases to offset evolving damage. HI, hypoxia-ischaemia; EAAs, excitatory amino acids; EPP, exchangeable phosphate pool; NAA, N-acetylaspartate; NO, nitric oxide; NTP, nucleoside triphosphate (this is mainly ATP); OFRs, oxygen free radicals; RIPostC, remote ischaemic postconditioning.

endogenous inhibition of oxidative metabolism and increased tissue oxygenation.¹¹ The ‘therapeutic window’ is believed to span this period. Much of our understanding of cerebral metabolism following HI has evolved through magnetic resonance spectroscopy (MRS) through which we have shown that latent phase duration is inversely related to insult severity.¹² In the early recovery period (2–8 h after HI), MRS may provide an early marker of injury severity; an overshoot of phosphocreatine (PCr; donates phosphate to ADP to generate ATP) is associated with favourable outcome¹³ and raised cerebral lactate or inorganic phosphate (Pi) at 2 h is indicative of adverse outcome.¹⁴

Secondary phase

Both preclinical¹⁵ and baby studies¹⁶ using phosphorus-31 (³¹P) MRS have demonstrated the deterioration in cerebral oxidative metabolism 6–24 h after HI (termed secondary energy failure) (figure 1). Despite adequate oxygenation and circulation, PCr and nucleotide triphosphate (NTP—mainly ATP) fell and Pi increased. Low cerebral PCr/Pi, NTP/total mobile phosphates,^{16 17} increased brain lactate¹⁸ and an alkaline intracellular pH (pHi)¹⁹ in the first few days after birth were associated with neurodevelopmental impairment and increased mortality.

This secondary phase is marked by the onset of seizures, secondary cytotoxic oedema, accumulation of cytokines and mitochondrial failure (figure 1). Mitochondrial failure is a key step leading to delayed cell death. The degree of energy failure influences the type of neuronal death during early and delayed stages,^{20 21} and the degree of trophic support influences the angiogenesis and neurogenesis during the recovery phase after HI.

Tertiary phase

There is evidence that active pathological processes occur for weeks, months and years after a hypoxic-ischaemic insult; this has been termed tertiary brain injury.²² Indeed, a persisting cerebral lactic alkalosis has been observed using MRS over the first year after birth in those infants with adverse neurodevelopmental outcomes.¹⁸ Mechanisms of this persisting damage involve gliosis, persistent inflammatory receptor activation and epigenetic changes.

Endogenous neuroprotection

Brain damage and lasting functional impairment after NE are the results of a balance between injurious mechanisms (cell death, persistent inflammation) and endogenous protection (acute response, recovery, repair). Optimal therapy will demand exploitation of multiple pathways that prevent brain cell death and promote repair.²³ Much neonatal neuroprotection research has emphasised immediate cytotoxic mechanisms; however, the brain also mounts a potent, though only partially successful, defensive response against many of the deleterious secondary mechanisms of injury.²⁴ Therapies to boost the endogenous neuroprotective response are particularly attractive; they are less likely to disrupt physiological neurotransmission, so may offer more effective treatments with fewer unwanted side effects.

We discuss five interventions whose actions include the augmentation of the endogenous neuroprotective response. Birth asphyxiated babies have an endogenous cooling response;²⁵ therapeutic hypothermia is already the standard clinical care for babies with moderate or severe NE. Remote ischaemic postconditioning (RIPostC) is a novel therapy, which has enormous promise as an intervention that harnesses the body’s neuroprotective ‘conditioning’ mechanisms. Melatonin is known for its role in entraining the circadian rhythm;²⁶ however, endogenous levels of melatonin increase after HI and exogenously

administered melatonin confers brain protection.²⁷ Endogenous endocannabinoids and erythropoietin (Epo), likewise increased following HI, also have a role in neuroprotection. There is expanding evidence to show that Epo confers protection that extends to the tertiary phase of injury, promoting repair.

THERAPEUTIC HYPOTHERMIA

Background

For over 50 years, it has been known that babies with birth depression have an endogenous cooling response.²⁵ We observed this phenomenon in our pilot cooling study in a low-resource setting.²⁸ After two decades of laboratory studies,^{29 30} clinical trials³¹ and endorsement from regulatory bodies (<http://www.nice.org.uk/guidance/ipg347>), therapeutic hypothermia is now standard clinical care for moderate-to-severe NE in the UK and high-income countries.⁵

Mechanism

Pathways underpinning hypothermic neuroprotection are covered in detail in recent reviews by Wassink *et al*³² and Edwards *et al*³³ In brief, these pathways include a decrease in metabolic rate with parallel decreases in O₂ consumption and CO₂ production, reduced loss of high-energy phosphates during HI and during secondary cerebral energy failure, reduced excitotoxicity, reduced reactive oxygen species production, protein synthesis preservation, decreased oedema, modulation of the inflammatory cascade and a change in pro-apoptotic and anti-apoptotic signalling.^{34–36}

Clinical application

In intensive care settings, clinical trials have included whole body cooling with core temperature reduced to 33.5°C for 72 h³⁷ and selective head cooling with core temperatures reduced to 34.5°C.³⁸ Some studies suggest less severe brain MRI findings in babies who have had whole body cooling versus selective head cooling; other studies suggest equal benefit from both cooling methods.^{39 40}

There is clear evidence that therapeutic hypothermia as a therapy for moderate-to-severe NE reduces adverse outcome (mortality and neurodevelopmental disability) at 18 months of age (typical relative risk 0.75, 95% CI 0.68 to 0.83)³¹; this improvement persists into childhood⁴¹ and there is widespread benefit to society, individuals and the economy (UK >£125 million benefit).⁴² However, therapeutic hypothermia offers only an 11% reduction in risk of death or disability, from 58% to 47%.⁴ Moreover, effective cooling treatment requires a high level of neonatal intensive care support, which is not available in many lower resource settings. There is an urgent need to develop additional simple, safe and effective neuroprotective treatment strategies.

Caveats of hypothermia

Recently, therapeutic hypothermia has been shown to be ineffective and even harmful in the presence of infection/inflammation in adult clinical studies.⁴³ In a preclinical neonatal rodent study, cooling was not neuroprotective in inflammation-sensitised HI.⁴⁴ In a small prospective study of placental histology relative to MRI in babies, therapeutic hypothermia was less protective in babies whose placenta showed chorioamnionitis.⁴⁵ We reported an unexpectedly high mortality in NE cases cooled to 33.5°C in a small pilot therapeutic hypothermia feasibility (not efficacy) study in sub-Saharan Africa²⁸; this may have been related in part to higher rates of intercurrent infection/inflammation in affected infants.

Brain injury and hypothermia both alter immune responsiveness. Following HI, a bidirectional communication between the injured brain and the peripheral innate and adaptive immune system regulates the progression of both ischaemic pathology and tissue repair (for an in-depth review of the dualistic role of inflammation, see An *et al.*,⁴⁶). HI acutely triggers the release of cytokines and chemokines from neurons, astrocytes and microglia. These signals induce microglial activation, trigger further release of pro-inflammatory cytokines such as tumour necrosis factor α and interleukin 6 (IL-6) and recruit white blood cells (WBCs). The infiltration of macrophages is both detrimental in ischaemic injury and protective against haemorrhage. Similarly, while early elevation of circulating neutrophils after HI may augment brain injury,⁴⁷ prolonged immunosuppression and T cell lymphopenia is associated with immune paralysis and worse outcome in animal models of stroke,⁴⁸ traumatic brain injury⁴⁹ and human adult stroke.⁵⁰ Thus, inflammation following HI has both helpful and harmful effects, which may affect the response to neuroprotective treatments.

A key mechanism of action of therapeutic hypothermia is the inhibition of the pro-inflammatory cascade⁵¹; and hypothermia may therefore inhibit both protective and damaging responses.⁵² A recent study investigated the effect of therapeutic hypothermia on modulating the peripheral immune response over the first 72 h after birth in 65 infants with NE.⁵³ Hypothermia lowered absolute neutrophil and lymphocyte counts compared with normothermic infants. In the hypothermic group, those patients who did not have a recovery of their WBC counts after rewarming had poor outcomes, whereas those who had better recovery of WBC counts had a better long-term outcome.⁵³ This may indicate immune paralysis in the adverse outcome group. It is possible, therefore, that hypothermic immune suppression has a negative influence on infants with infection-sensitised brain injury. Several adult studies of hypothermia have found higher infection rates in cooled groups.⁵⁴ This has not been shown in neonatal studies of hypothermia treatment; however, blood culture-positive neonatal sepsis rates are low (5–12%) and much larger trials would be needed to detect any increase. A better understanding is crucial for achieving optimal neuroprotection in NE.

REMOTE ISCHAEMIC POSTCONDITIONING

Background

'Conditioning' describes an adaptive process of endogenous protection that occurs in all mammalian species, in which small, sublethal doses of a harmful agent protect an organism against a lethal dose of the same agent. Conditioning paradigms include toxins, substrate deprivation and infection/inflammation.²⁴ One conditioning agent may also confer protection against a different insult.^{55–56}

Ischaemic *preconditioning* describes brief non-lethal episodes of ischaemia that confer protection against subsequent cell-lethal ischaemia, as has been observed in clinical studies of transient ischaemic attack^{57–58} and angina.⁵⁹ It is likely that contractions during labour also represent a preconditioning stimulus. Ischaemic *postconditioning* evolved from this concept,^{60–61} and is defined as intermittent sublethal interruptions to blood flow *after* the cell-lethal ischaemia.⁶² Postconditioning is effective if performed on a non-vital organ, such as a limb, remote to the affected organ⁶³—called *remote* ischaemic postconditioning (RIPostC). Use of a remote limb makes RIPostC a feasible clinical treatment strategy for NE.

Mechanism

RIPostC has been shown to protect the adult and neonatal brain in rodent models of stroke. The protective mechanisms of

RIPostC are incompletely understood, but are thought to involve three intimately inter-related pathways initiated by the release of a number of endogenous autocooids (including adenosine, bradykinin, opioids) from the ischaemic skeletal muscle. These pathways are (i) the neuronal pathway; (ii) the humoral pathway and (iii) the systemic response (figure 2).⁶⁴ Animal models have shown that interruption of any one of these pathways abrogates the neuroprotection conferred by RIPostC.

In brief, the neuronal pathway describes the autocooid-mediated stimulation of local afferent nerves that effect remote protection via efferent nerves, including the autonomic nervous system.^{65–66} Both limb ischaemia and efferent nerve activation trigger the release of a number of bloodborne protective factors that are transported in the circulation and mediate protection in the brain—the humoral pathway.^{67–68} The systemic pathway describes the impact of RIPostC throughout the body, including immune effects (such as reduced neutrophil activation) and reduced expression of apoptotic and inflammatory genes.⁶⁹

Following remote ischaemic stimulus, these three pathways converge in the brain to increase cerebral blood flow, attenuate neuroinflammation and at a cellular level to activate pro-survival signalling cascades, including genetic and epigenetic modulation. Ultimately these processes protect mitochondrial integrity, reduce energy demands, increase cell survival and promote repair mechanisms^{70–72} (figure 2).

In neonatal⁷³ and adult⁷⁴ small animal models, RIPostC reduces infarct size in focal and global ischaemia. Moreover, these studies have shown an extended therapeutic window for the application of RIPostC following hypoxic-ischaemic brain injury⁶³ and application of RIPostC up to 24 h after insult was associated with improved long-term motor outcomes.⁷⁵

In our large animal (piglet) model of perinatal asphyxia, we found that four 10 min cycles of ischaemia/reperfusion of both lower limbs, starting immediately following resuscitation, provided protection in the white matter,⁷⁶ with decreased cell death and inflammation. MRS data in our study showed that RIPostC mitigated the rise in white matter lactate/N-acetyl aspartate and increased whole-brain ATP, findings that predict better long-term outcome in clinical studies in human newborns.^{16–77}

Hurdles to clear before clinical translation

RIPostC has been explored in clinical settings for conditions, including cardiac disease and stroke.⁷⁸ A meta-analysis of 23 randomised clinical trials of limb conditioning in adults undergoing cardiac surgery found reduced incidence of myocardial infarction in the limb-conditioned groups, regardless of timing. A randomised control trial of RIPostC for children undergoing cardiac bypass also showed cardioprotection.⁷⁹ In adult stroke, a recent study of 443 adults who underwent prehospital remote ischaemic preconditioning as an adjunct to thrombolysis for acute ischaemic stroke found a reduced risk of tissue infarction in the treatment group.⁸⁰ In all studies, remote ischaemic conditioning was safe and well tolerated. However, another meta-analysis of remote ischaemic preconditioning in open cardiac surgery showed the cardioprotective effect was most marked in studies without full blinding, emphasising the need for further double-blind randomised studies.⁸¹

Clinical trials are needed to establish whether RIPostC is safe and protective in NE. It will be important to address the safety and reproducibility of inducing intermittent limb ischaemia, with and without concomitant cooling therapy. There remain difficult hurdles such as the dose–response of RIPostC (how many cycles and for how long achieves best protection and

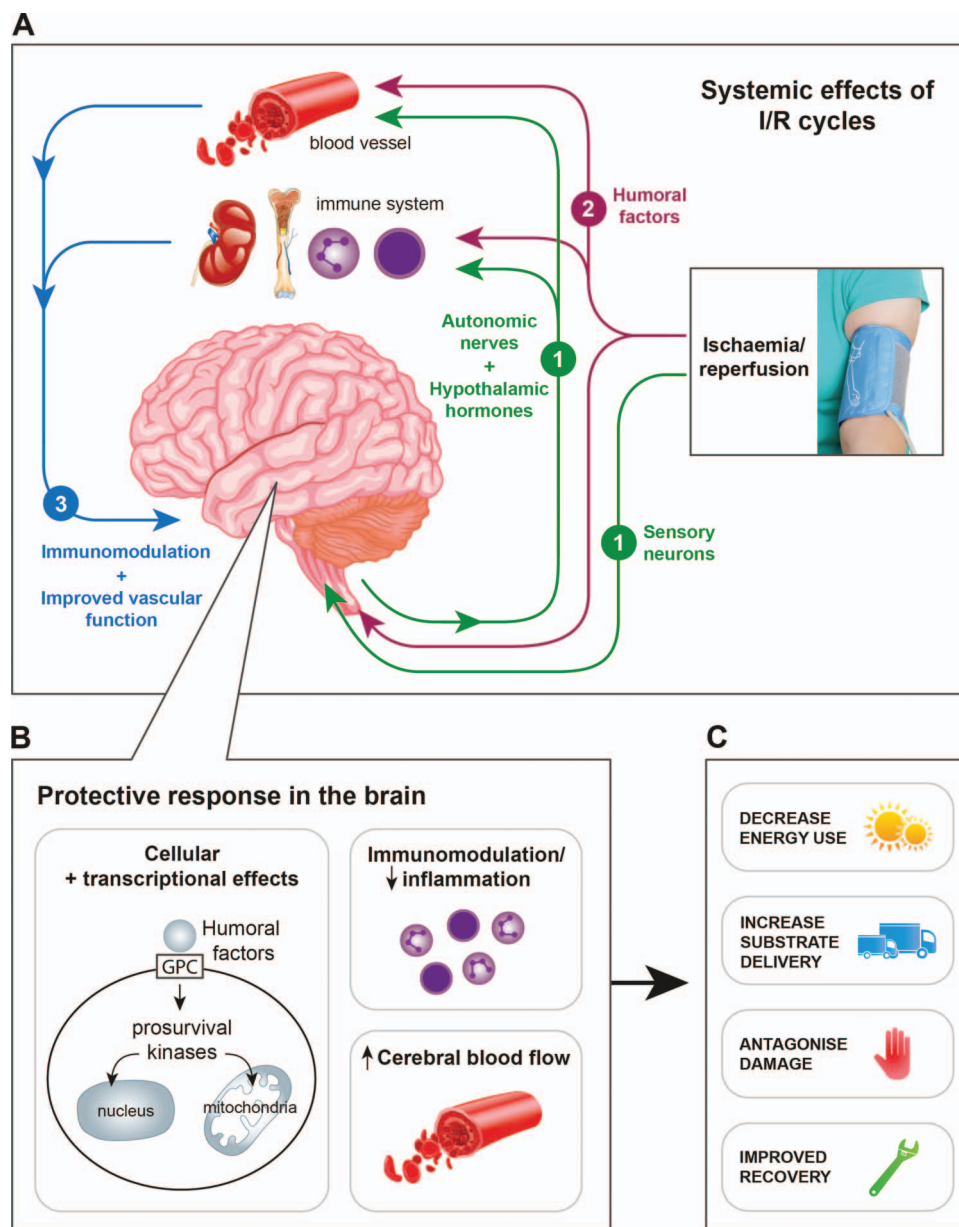


Figure 2 (A) The neuroprotective mechanisms of R1PostC are thought to involve three inter-related pathways induced by remote limb ischaemia. (1) The neuronal pathway involves activation of both local sensory nerves and the autonomic nervous system to mediate protective effects, including the release of humoral factors; (2) the humoral pathway involves endogenous protective factors, including locally acting autocoids and bloodborne humoral factors that travel to the brain and (3) the systemic response includes immune modulation and blood pressure regulation. (B) Within the brain, the three pathways converge to increase cerebral blood flow, ameliorate neuroinflammation and to activate cell survival mechanisms. Direct pro-survival actions within cells are mediated via G-protein-coupled (GPC) receptors and include mitochondrial protection (maintenance of potassium-sensitive ATP channel, prevention of mitochondrial permeability transition pore opening) and transcriptional regulation (both genetic and epigenetic modulation) in the nucleus. (C) Following remote ischaemic stimulus after HI, the effects of these neuroprotective mechanisms are to decrease energy consumption; to increase substrate delivery and offset cerebral secondary energy failure; to protect against cell death and to augment long-term recovery and repair. HI, hypoxia-ischaemia; I/R, ischaemia/reperfusion; R1PostC, remote ischaemic postconditioning.

avoids any detrimental effects), the therapeutic time windows and the precise protective mechanisms.⁸²

MELATONIN Background

Melatonin is a naturally occurring neuroendocrine molecule secreted in response to environmental light–dark cycles. Melatonin is both lipophilic and hydrophilic. It easily crosses biological membranes and acts via receptor-dependent and receptor-independent processes to modulate cell signalling and

gene expression.^{83 84} While melatonin's key and probably best-known role is to regulate the body's multifarious circadian rhythms,²⁶ it influences numerous physiological functions, including growth and development, reproduction and the immune response.

Endogenous melatonin is integral to normal neurodevelopment and protects the developing brain from injury.^{85–90} Maternal melatonin levels are raised in pregnancy^{91 92} and melatonin readily crosses the placenta and blood–brain barrier.^{93 94} Healthy term-born neonates have relatively low

pineal melatonin production, which lacks diurnal variation for the first weeks of life.^{95–96} However, we observed a 6- to 15-fold increase in plasma melatonin following HI in our experimental model of perinatal HI²⁷ and a similar response has also been observed in human stroke and in critically ill children,^{97–98} implying a role for melatonin in an endogenous protective response.

Mechanism

Acting via specific cell membrane and nuclear receptors, melatonin achieves powerful neuroprotective effect via anti-oxidant, anti-apoptotic and anti-inflammatory processes^{85–88} and by promoting neuronal and glial development.^{99–101} Developing brain tissue is highly susceptible to free radical damage^{102–104} and the potent free radical-scavenging properties of melatonin and its metabolites provide a fundamental neuroprotective mechanism.^{105–109} Additional indirect anti-oxidant effects of melatonin include upregulation of anti-oxidant enzymes⁹⁴ and crucially the preservation of mitochondrial integrity.^{106–107–109} Numerous rodent and large animal studies have shown that melatonin reduces oxidative damage to cerebral lipids^{104–110–116} and notably ameliorates secondary cerebral energy failure²⁷ and apoptosis.^{116–121}

Further, melatonin's wide-ranging immune-modulating properties^{122–123} facilitate neuroprotection following HI.^{27–118–121–124} Importantly, melatonin is protective in lipopolysaccharide-sensitised HI.¹²⁵ Given the evidence outlined above indicating therapeutic cooling may lack efficacy following infection-sensitised HI,^{44–45} melatonin may prove an effective immune-modulating neuroprotectant in such cases.

Clinical use and safety

Melatonin is an extremely safe neurotherapeutic. No study of antenatal or postnatal melatonin treatment has shown any serious side effects,¹²⁶ nor were any serious adverse events identified in 3000 children taking melatonin for up to 6 years.¹²⁷ In small neonatal clinical studies, melatonin improved outcomes in sepsis,¹²⁸ prematurity¹²⁹ and perinatal asphyxia.¹³⁰ In our large animal model of perinatal asphyxia, we showed neuroprotective efficacy conferred by melatonin-augmented cooling when compared with cooling alone.²⁷ In our study, melatonin 30 mg/kg (which is 100 times the dose administered for disordered sleep in children) administered to newborn piglets immediately after HI over 6 h did not alter any physiological variable.²⁷ A study of 30 term infants with NE randomised to cooling alone or cooling plus oral melatonin (five daily doses of 10 mg/kg per day made up from melatonin tablets crushed and dissolved in distilled water) suggested improved neurological outcome at 6 months in the melatonin group.¹³¹ However, four patients in the hypothermia group had severe encephalopathy, whereas only two in the hypothermia/melatonin group had severe encephalopathy at birth; this may lend bias to the results in this small study. Further, the blood levels of melatonin on day 5 in the cooling group were 32.1±3.5 pg/mL, while in the melatonin/cooling group were 42.7±5.1 pg/mL; preclinical data suggest that significantly higher pharmacological levels of melatonin are needed for optimal protection and work is currently underway to determine the lowest effective dose of melatonin for neuroprotection. Nevertheless, phase I and II clinical studies of melatonin-augmented hypothermia for NE are keenly awaited. In 2011, melatonin was rated by an international group of leading perinatal neuroscientists as the most promising of 13 neuroprotectants nearing clinical translation.¹³²

CANNABINOIDS

Background

Endocannabinoids are emerging as a potential neurotherapeutic for NE. The endocannabinoid system is a neuromodulatory system that participates in a wide range of physiological processes in mammals.¹³³ This endogenous system consists of target receptors, endogenous ligands and the enzymes responsible for endocannabinoid biosynthesis, transport and degradation.^{134–135} Accumulating evidence indicates that endocannabinoids, like melatonin, are inherently involved in the normal development of the fetal central nervous system and its functions.^{136–141} Moreover, the levels of endocannabinoids, which are normally found at low concentrations in the brain, dramatically increase upon neuronal injury,^{136–142–144} suggesting that endocannabinoids provide an endogenous neuroprotective system.¹⁴⁵

Mechanism

Endocannabinoids modulate the intensity and extension of neurotoxic processes^{146–150} and the inflammatory response^{151–155} and promote cell survival.^{156–161} Synthetic cannabinoid agonists have shown significant grey and white matter protection in animal studies of brain injury.^{162–167} In large animal models of perinatal asphyxia, cannabinoid WIN55212-2 administered immediately after HI protected mitochondrial injury and prevented apoptosis.^{162–163} Cannabidiol given immediately after HI reduced neuronal injury, cerebral haemodynamic impairment, brain oedema and seizures and restored motor and behavioural performance in the 72 h after HI.^{166–167} In rodent models of stroke, prolonged 7-day administration of cannabinoid WIN55212-2 started immediately after injury enhanced long-term neuronal and oligodendrocyte recovery and regeneration.^{164–165} Cannabinoids, however, achieve neuroprotection in part through hypothermia. For example, the cannabinoid agonist HU10 induced hypothermia in an experimental stroke model and was protective, but this benefit was completely abolished by rewarming animals to the temperature of the control group.¹⁶⁸

Clinical use and safety

The main established clinical uses of cannabinoids are for chronic pain, for muscle spasms and as an appetite stimulant.^{169–170} Additionally, the synthetic cannabinoid dexamabinol is in a number of phase I clinical trials for primary and secondary solid tumours (<http://www.clinicaltrials.gov>). A previous phase III randomised controlled trial of 861 adult patients given dexamabinol as a neuroprotectant following traumatic brain injury showed that dexamabinol was safe but not efficacious.¹⁷¹ However, we did not identify any reported clinical studies of cannabinoids for stroke or perinatal brain injury, or studies where cannabinoids were combined with therapeutic hypothermia.

Reported side effects of cannabinoids have all been mild and transient, including sedation, anxiety, dizziness and nausea.^{169–170} Maas *et al*¹⁷¹ found no toxic cardiac, hepatic or renal effects in their study of dexamabinol for traumatic brain injury. However, cannabinoids have been shown to accumulate selectively in the brain and their clearance is relatively slow,¹⁶⁹ thus preclinical pharmacokinetic studies would be imperative prior to clinical trials of cannabinoids for NE.

ERYTHROPOIETIN (EPO)

Background

Epo is a pleiotropic cytokine with multiple roles in addition to that of a haemopoietic growth factor. As with melatonin and cannabinoids, the role of Epo in normal brain development and

neuroprotection is becoming clear (for review see Rangarajan and Juul¹⁷²). Epo receptors (EpoR) are located throughout the central nervous system on neurons,¹⁷³ glia¹⁷⁴ and endothelial cells;¹⁷⁵ they participate in proliferation and differentiation of these cells¹⁷⁶ and are upregulated following brain injury. In a similar way to the exaggerated hypoxic-ischaemic injury observed when the endogenous melatonin response is abolished in pinealectomised animals,¹⁷⁷ the absence of endogenous Epo and EpoR augments ischaemic damage and impairs neuronal survival.¹⁷⁸

Epo is a key component of the body's endogenous 'conditioning' response to injurious paradigms, including ischaemia. Hypoxic preconditioning occurs when Epo is expressed after brief hypoxia, reducing damage following a second insult.¹⁷⁹ This effect can be replicated by treatment with exogenous Epo prior to HI.¹⁸⁰ Preclinical and clinical studies have harnessed the conditioning and regenerative potential of Epo, which is now emerging as a promising neuroprotectant that promotes repair into the tertiary phase of NE.

Mechanism

Hypoxia and pro-inflammatory cytokines activate hypoxia-inducible factor to induce expression of Epo and EpoR. Following brain injury, Epo is anti-apoptotic,¹⁸¹ anti-oxidative¹⁸² and anti-inflammatory.¹⁸³ However, a key role for Epo is repair; Epo binding stimulates neurogenesis, oligodendrogenesis and angiogenesis, all of which are upregulated following brain injury.^{184–185} Additionally, Epo increases neuronal and glial migration around the injured area via the secretion of matrix metalloproteinases.^{186–187} In animal studies of term and preterm perinatal HI, Epo treatment results in reduced brain volume loss and improved cognitive and motor outcomes^{188–190} and augments the neuroprotection conferred by cooling alone.¹⁹¹

Epo in clinical trials

Numerous animal studies of Epo for brain injuries, including stroke and perinatal HI, have shown that high-dose recombinant Epo and Epo-mimetics are safe and cross the blood–brain barrier, resulting in neuroprotection.¹⁹² Epo pharmacokinetics has been studied using doses from 250 U/kg to 2500 U/kg.¹⁹³ Phase I/II studies in human preterm¹⁹³ and term^{194–195} neonates, performed to establish feasibility, safety and appropriate dosing, have not identified any of the common side effects observed in adults (polycythaemia, thrombosis, hypertension). Epo-mimetics have been developed to improve neuroprotection without stimulating erythropoiesis.¹⁹⁶

Follow-up of 22 infants enrolled in a phase I clinical trial of Epo-augmented hypothermia (no comparison group) for treatment of NE found no deaths and only one infant with moderate–severe disability at age 2 years.¹⁹⁷ A number of larger phase II/III studies of Epo safety and efficacy in neonatal populations are underway (reviewed in Rangarajan and Juul¹⁷²). The optimal dose and regimen for human Epo neuroprotection is still not known; however, key points have been learnt from rodent studies such as the requirement for multiple injections and late 1 week dosing for maximal protection¹⁹⁸; a study of 45 term infants comparing single-dose Epo alone on day 0 with 72 h therapeutic hypothermia alone for treatment of NE found superior protection in the hypothermia group.¹⁹⁹ Further studies are needed to fully understand the specific role of Epo in the tertiary phase of brain injury and repair. Thus, the combined safety and efficacy of Epo administered alongside established and novel treatments that ameliorate secondary energy failure (cooling, melatonin) must be determined as key next steps in clinical translation.

CONCLUSION

Perinatal HI leading to NE sets up a cascade of processes that lead to an evolving brain injury, which includes a latent phase, secondary energy failure phase and tertiary brain injury phase. Coexisting infection/inflammation may exacerbate this injury. The brain mounts a potent, though only partially successful, defensive response against many of the deleterious secondary mechanisms of injury. Therapies that augment the endogenous neuroprotective response such as RPostC are attractive but need further study to define optimal protocols. Part of the endogenous neuroprotective response is lowering of the core body temperature as well as increased melatonin, Epo and cannabinoid levels. Augmenting these endogenous responses have shown protection in preclinical studies and therapeutic hypothermia is now a routine therapy for moderate-to-severe NE. Clinical trials are now ongoing for Epo-augmented hypothermia. We anticipate that future newborn brain protection will comprise a tailored combination of therapies; the challenge will be to ensure the timing and dose of each neuroprotectant are appropriate for the phase of injury to ensure optimal and lasting protection.

Funding The RPostC study was funded by the MRC (MR/J00457X/1). DAA is funded by the Basque Government Postdoctoral Program (POS_2013_1_191). DH is funded by the British Heart Foundation (grant number FS/10/039/28270) and the Rosetrees Trust. This work was undertaken at University College London Hospitals/ University College London, which received a proportion of funding from the UK Department of Health's National Institute for Health Research Biomedical Research Centres funding scheme.

Contributors NJR, KJH, DA-A and ME wrote specific sections for the review. JH and NJR put the review together and drew the diagrams. DJH advised on the conditioning section and diagram.

Competing interests None.

Provenance and peer review Commissioned; externally peer reviewed.

Open Access This is an Open Access article distributed in accordance with the terms of the Creative Commons Attribution (CC BY 4.0) license, which permits others to distribute, remix, adapt and build upon this work, for commercial use, provided the original work is properly cited. See: <http://creativecommons.org/licenses/by/4.0/>

REFERENCES

- Liu L, Oza S, Hogan D, *et al*. Global, regional, and national causes of child mortality in 2000–13, with projections to inform post-2015 priorities: an updated systematic analysis. *Lancet* 2015;385:430–40.
- Lee AC, Kozuki N, Blencowe H, *et al*. Intrapartum-related neonatal encephalopathy incidence and impairment at regional and global levels for 2010 with trends from 1990. *Pediatr Res* 2013;74(Suppl 1):50–72.
- Lawn JE, Blencowe H, Oza S, *et al*. Every Newborn: progress, priorities, and potential beyond survival. *Lancet* 2014;384:189–205.
- Edwards AD, Brocklehurst P, Gunn AJ, *et al*. Neurological outcomes at 18 months of age after moderate hypothermia for perinatal hypoxic ischaemic encephalopathy: synthesis and meta-analysis of trial data. *BMJ* 2010;340:c363.
- Roka A, Azzopardi D. Therapeutic hypothermia for neonatal hypoxic ischaemic encephalopathy. *Early Hum Dev* 2010;86:361–7.
- Nelson KB, Leviton A. How much of neonatal encephalopathy is due to birth asphyxia? *Am J Dis Child* 1991;145:1325–31.
- Eklind S, Mallard C, Leverin AL, *et al*. Bacterial endotoxin sensitizes the immature brain to hypoxic–ischaemic injury. *Eur J Neurosci* 2001;13:1101–16.
- Wang X, Stridh L, Li W, *et al*. Lipopolysaccharide sensitizes neonatal hypoxic-ischemic brain injury in a MyD88-dependent manner. *J Immunol* 2009;183:7471–7.
- Nelson KB, Willoughby RE. Infection, inflammation and the risk of cerebral palsy. *Curr Opin Neurol* 2000;13:133–9.
- Szydłowska K, Tymianski M. Calcium, ischemia and excitotoxicity. *Cell Calcium* 2010;47:122–9.
- Jensen EC, Bennet L, Hunter CJ, *et al*. Post-hypoxic hypoperfusion is associated with suppression of cerebral metabolism and increased tissue oxygenation in near-term fetal sheep. *J Physiol* 2006;572(Pt 1):131–9.
- Iwata O, Iwata S, Thornton JS, *et al*. "Therapeutic time window" duration decreases with increasing severity of cerebral hypoxia-ischaemia under normothermia and delayed hypothermia in newborn piglets. *Brain Res* 2007;1154:173–80.

- 13 Iwata O, Iwata S, Bainbridge A, *et al.* Supra- and sub-baseline phosphocreatine recovery in developing brain after transient hypoxia-ischaemia: relation to baseline energetics, insult severity and outcome. *Brain* 2008;131(Pt 8):2220–6.
- 14 Cady EB, Iwata O, Bainbridge A, *et al.* Phosphorus magnetic resonance spectroscopy 2 h after perinatal cerebral hypoxia-ischemia prognosticates outcome in the newborn piglet. *J Neurochem* 2008;107:1027–35.
- 15 Lorek A, Takei Y, Cady EB, *et al.* Delayed (“secondary”) cerebral energy failure after acute hypoxia-ischemia in the newborn piglet: continuous 48-hour studies by phosphorus magnetic resonance spectroscopy. *Pediatr Res* 1994;36:699–706.
- 16 Azzopardi D, Wyatt JS, Cady EB, *et al.* Prognosis of newborn infants with hypoxic-ischemic brain injury assessed by phosphorus magnetic resonance spectroscopy. *Pediatr Res* 1989;25:445–51.
- 17 Martin E, Buchli R, Ritter S, *et al.* Diagnostic and prognostic value of cerebral 31P magnetic resonance spectroscopy in neonates with perinatal asphyxia. *Pediatr Res* 1996;40:749–58.
- 18 Robertson NJ, Cox IJ, Cowan FM, *et al.* Cerebral intracellular lactic alkalosis persisting months after neonatal encephalopathy measured by magnetic resonance spectroscopy. *Pediatr Res* 1999;46:287–96.
- 19 Robertson NJ, Cowan FM, Cox IJ, *et al.* Brain alkaline intracellular pH after neonatal encephalopathy. *Ann Neurol* 2002;52:732–42.
- 20 Hagberg H, Mallard C, Rousset CI, *et al.* Mitochondria: hub of injury responses in the developing brain. *Lancet Neurol* 2014;13:217–32.
- 21 Northington FJ, Chavez-Valdez R, Martin LJ. Neuronal cell death in neonatal hypoxia-ischemia. *Ann Neurol* 2011;69:743–58.
- 22 Fleiss B, Gressens P. Tertiary mechanisms of brain damage: a new hope for treatment of cerebral palsy? *Lancet Neurol* 2012;11:556–66.
- 23 Gonzalez FF, Ferriero DM. Therapeutics for neonatal brain injury. *Pharmacol Ther* 2008;120:43–53.
- 24 Dirnagl U, Simon RP, Hallenbeck JM. Ischemic tolerance and endogenous neuroprotection. *Trends Neurosci* 2003;26:248–54.
- 25 Burnard ED, Cross KW. Rectal temperature in the newborn after birth asphyxia. *BMJ* 1958;2:1197–9.
- 26 Reiter RJ, Tan D-X, Fuentes-Broto L. Melatonin: a multitasking molecule. *Prog Brain Res* 2010;181:127–51.
- 27 Robertson NJ, Faulkner S, Fleiss B, *et al.* Melatonin augments hypothermic neuroprotection in a perinatal asphyxia model. *Brain* 2013;136(Pt 1):90–105.
- 28 Robertson NJ, Nakakeeto M, Hagmann C, *et al.* Therapeutic hypothermia for birth asphyxia in low-resource settings: a pilot randomised controlled trial. *Lancet* 2008;372:801–3.
- 29 Bona E, Hagberg H, Loberg EM, *et al.* Protective effects of moderate hypothermia after neonatal hypoxia-ischemia: short- and long-term outcome. *Pediatr Res* 1998;43:738–45.
- 30 Thoresen M, Penrice J, Lorek A, *et al.* Mild hypothermia after severe transient hypoxia-ischemia ameliorates delayed cerebral energy failure in the newborn piglet. *Pediatr Res* 1995;37:667–70.
- 31 Jacobs SE, Berg M, Hunt R, *et al.* Cooling for newborns with hypoxic ischaemic encephalopathy. *Cochrane Database Syst Rev* 2013;1:CD003311.
- 32 Wassink G, Gunn ER, Drury PP, *et al.* The mechanisms and treatment of asphyxial encephalopathy. *Front Neurosci* 2014;8:40.
- 33 Edwards AD, Azzopardi DV, Gunn AJ. *Neonatal neural rescue: a clinical guide.* Cambridge University Press, 2013.
- 34 Busto R, Globus MY, Dietrich WD, *et al.* Effect of mild hypothermia on ischemia-induced release of neurotransmitters and free fatty acids in rat brain. *Stroke* 1989;20:904–10.
- 35 Drury PP, Bennet L, Gunn AJ. Mechanisms of hypothermic neuroprotection. *Semin Fetal Neonatal Med* 2010;15:287–92.
- 36 Yenari MA, Han HS. Neuroprotective mechanisms of hypothermia in brain ischaemia. *Nat Rev Neurosci* 2012;13:267–78.
- 37 Azzopardi DV, Strohm B, Edwards AD, *et al.* Moderate hypothermia to treat perinatal asphyxial encephalopathy. *N Engl J Med* 2009;361:1349–58.
- 38 Gluckman PD, Wyatt JS, Azzopardi D, *et al.* Selective head cooling with mild systemic hypothermia after neonatal encephalopathy: multicentre randomised trial. *Lancet* 2005;365:663–70.
- 39 Sarkar S, Donn SM, Bapuraj JR, *et al.* Distribution and severity of hypoxic-ischaemic lesions on brain MRI following therapeutic cooling: selective head versus whole body cooling. *Archives of disease in childhood. Fetal Neonatal Ed* 2012;97:F335–9.
- 40 Tagin MA, Woolcott CG, Vincer MJ, *et al.* Hypothermia for neonatal hypoxic ischemic encephalopathy: an updated systematic review and meta-analysis. *Arch Pediatr Adolesc Med* 2012;166:558–66.
- 41 Shankaran S. Therapeutic hypothermia for neonatal encephalopathy. *Curr Treat Options Neurol* 2012;14:608–19.
- 42 Azzopardi D, Strohm B, Linsell L, *et al.* Implementation and conduct of therapeutic hypothermia for perinatal asphyxial encephalopathy in the UK—analysis of national data. *PLoS ONE* 2012;7:e38504.
- 43 Mourvillier B, Tubach F, van de Beek D, *et al.* Induced hypothermia in severe bacterial meningitis: a randomized clinical trial. *JAMA* 2013;310:2174–83.
- 44 Osredkar D, Thoresen M, Maes E, *et al.* Hypothermia is not neuroprotective after infection-sensitized neonatal hypoxic-ischemic brain injury. *Resuscitation* 2014;85:567–72.
- 45 Wintermark P, Boyd T, Gregas MC, *et al.* Placental pathology in asphyxiated newborns meeting the criteria for therapeutic hypothermia. *Am J Obstet Gynecol* 2010;203:579 e1–9.
- 46 An C, Shi Y, Li P, *et al.* Molecular dialogs between the ischemic brain and the peripheral immune system: dualistic roles in injury and repair. *Prog Neurobiol* 2014;115:6–24.
- 47 Buck BH, Liebeskind DS, Saver JL, *et al.* Early neutrophilia is associated with volume of ischemic tissue in acute stroke. *Stroke* 2008;39:355–60.
- 48 Liesz A, Suri-Payer E, Veltkamp C, *et al.* Regulatory T cells are key cerebroprotective immunomodulators in acute experimental stroke. *Nat Med* 2009;15:192–9.
- 49 Kline AE, Bolinger BD, Kochanek PM, *et al.* Acute systemic administration of interleukin-10 suppresses the beneficial effects of moderate hypothermia following traumatic brain injury in rats. *Brain Res* 2002;937:22–31.
- 50 Klehmet J, Harms H, Richter M, *et al.* Stroke-induced immunodepression and post-stroke infections: lessons from the preventive antibacterial therapy in stroke trial. *Neuroscience* 2009;158:1184–93.
- 51 Kimura A, Sakurada S, Ohkuni H, *et al.* Moderate hypothermia delays proinflammatory cytokine production of human peripheral blood mononuclear cells. *Crit Care Med* 2002;30:1499–502.
- 52 Polderman KH. Hypothermia, immune suppression and SDD: can we have our cake and eat it? *Crit Care* 2011;15:144.
- 53 Jenkins DD, Lee T, Chiuzan C, *et al.* Altered circulating leukocytes and their chemokines in a clinical trial of therapeutic hypothermia for neonatal hypoxic ischemic encephalopathy*. *Pediatr Crit Care Med* 2013;14:786–95.
- 54 Geurts M, Macleod MR, Kollmar R, *et al.* Therapeutic hypothermia and the risk of infection: a systematic review and meta-analysis. *Crit Care Med* 2014;42:231–42.
- 55 Lin HY, Huang CC, Chang KF. Lipopolysaccharide preconditioning reduces neuroinflammation against hypoxic ischemia and provides long-term outcome of neuroprotection in neonatal rat. *Pediatr Res* 2009;66:254–9.
- 56 McAuliffe JJ, Loepeke AW, Miles L, *et al.* Desflurane, isoflurane, and sevoflurane provide limited neuroprotection against neonatal hypoxia-ischemia in a delayed preconditioning paradigm. *Anesthesiology* 2009;111:533–46.
- 57 Wegener S, Gottschalk B, Jovanovic V, *et al.* Transient ischemic attacks before ischemic stroke: preconditioning the human brain? A multicenter magnetic resonance imaging study. *Stroke* 2004;35:616–21.
- 58 Weih M, Kallenberg K, Bergk A, *et al.* Attenuated stroke severity after prodromal TIA: a role for ischemic tolerance in the brain? *Stroke* 1999;30:1851–4.
- 59 Rezkalla SH, Kloner RA. Ischemic preconditioning and preinfarction angina in the clinical arena. *Nat Clin Pract Cardiovasc Med* 2004;1:96–102.
- 60 Vinten-Johansen J, Zhao ZQ, Jiang R, *et al.* Myocardial protection in reperfusion with postconditioning. *Expert Rev Cardiovasc Ther* 2005;3:1035–45.
- 61 Zhao ZQ, Corvera JS, Halkos ME, *et al.* Inhibition of myocardial injury by ischemic postconditioning during reperfusion: comparison with ischemic preconditioning. *Am J Physiol Heart Circ Physiol* 2003;285:H579–88.
- 62 Zhao ZQ, Vinten-Johansen J. Postconditioning: reduction of reperfusion-induced injury. *Cardiovasc Res* 2006;70:200–11.
- 63 Ren C, Gao X, Niu G, *et al.* Delayed postconditioning protects against focal ischemic brain injury in rats. *PLoS ONE* 2008;3:e3851.
- 64 Lim SY, Hausenloy DJ. Remote ischemic conditioning: from bench to bedside. *Front Physiol* 2012;3:27.
- 65 Malhotra S, Naggar I, Stewart M, *et al.* Neurogenic pathway mediated remote preconditioning protects the brain from transient focal ischemic injury. *Brain Res* 2011;1386:184–90.
- 66 Pignataro G, Esposito E, Sirabella R, *et al.* nNOS and p-ERK involvement in the neuroprotection exerted by remote postconditioning in rats subjected to transient middle cerebral artery occlusion. *Neurobiol Dis* 2013;54:105–14.
- 67 Denning GM, Ackermann LW, Barna TJ, *et al.* Proenkephalin expression and enkephalin release are widely observed in non-neuronal tissues. *Peptides* 2008;29:83–92.
- 68 Kanoria S, Jalan R, Seifalian AM, *et al.* Protocols and mechanisms for remote ischemic preconditioning: a novel method for reducing ischemia reperfusion injury. *Transplantation* 2007;84:445–58.
- 69 Shimizu M, Saxena P, Konstantinov IE, *et al.* Remote ischemic preconditioning decreases adhesion and selectively modifies functional responses of human neutrophils. *J Surg Res* 2010;158:155–61.
- 70 Mergenthaler P, Dirnagl U. Protective conditioning of the brain: expressway or roadblock? *J Physiol* 2011;589(Pt 17):4147–55.
- 71 Saxena P, Newman MA, Shehatha JS, *et al.* Remote ischemic conditioning: evolution of the concept, mechanisms, and clinical application. *J Card Surg* 2010;25:127–34.
- 72 Tapuria N, Kumar Y, Habib MM, *et al.* Remote ischemic preconditioning: a novel protective method from ischemia reperfusion injury—a review. *J Surg Res* 2008;150:304–30.

- 73 Zhou Y, Fathali N, Lekic T, *et al.* Remote limb ischemic postconditioning protects against neonatal hypoxic-ischemic brain injury in rat pups by the opioid receptor/Akt pathway. *Stroke* 2011;42:439–44.
- 74 Liu X, Zhao S, Liu F, *et al.* Remote ischemic postconditioning alleviates cerebral ischemic injury by attenuating endoplasmic reticulum stress-mediated apoptosis. *Transl Stroke Res* 2014;5:692–700.
- 75 Drunalini Perera PN, Hu Q, Tang J, *et al.* Delayed remote ischemic postconditioning improves long term sensory motor deficits in a neonatal hypoxic ischemic rat model. *PLoS ONE* 2014;9:e90258.
- 76 Ezzati M, Bainbridge A, Broad KD, *et al.* Limb remote ischemic post-conditioning protects cerebral white matter in a piglet model of perinatal asphyxia. *PAS* 2014;4:118.305.
- 77 Thayyil S, Chandrasekaran M, Taylor A, *et al.* Cerebral magnetic resonance biomarkers in neonatal encephalopathy: a meta-analysis. *Pediatrics* 2010;125:e382–95.
- 78 Brevoord D, Kranke P, Kuijpers M, *et al.* Remote ischemic conditioning to protect against ischemia-reperfusion injury: a systematic review and meta-analysis. *PLoS ONE* 2012;7:e42179.
- 79 Zhong H, Gao Z, Chen M, *et al.* Cardioprotective effect of remote ischemic postconditioning on children undergoing cardiac surgery: a randomized controlled trial. *Paediatr Anaesth* 2013;23:726–33.
- 80 Hougaard KD, Hjort N, Zeidler D, *et al.* Remote ischemic preconditioning as an adjunct therapy to thrombolysis in patients with acute ischemic stroke: a randomized trial. *Stroke* 2014;45:159–67.
- 81 Pilcher JM, Young P, Weatherall M, *et al.* A systematic review and meta-analysis of the cardioprotective effects of remote ischaemic preconditioning in open cardiac surgery. *J R Soc Med* 2012;105:436–45.
- 82 Zhao H. Hurdles to clear before clinical translation of ischemic postconditioning against stroke. *Transl Stroke Res* 2013;4:63–70.
- 83 Reiter RJ, Tan DX, Manchester LC, *et al.* Medical implications of melatonin: receptor-mediated and receptor-independent actions. *Adv Med Sci* 2007;52:11–28.
- 84 Slominski RM, Reiter RJ, Schlabritz-Loutsevitch N, *et al.* Melatonin membrane receptors in peripheral tissues: distribution and functions. *Mol Cell Endocrinol* 2012;351:152–66.
- 85 Acuna-Castroviejio D, Martin M, Macias M, *et al.* Melatonin, mitochondria, and cellular bioenergetics. *J Pineal Res* 2001;30:65–74.
- 86 Cardinali DP, Pagano ES, Scacchi Bernasconi PA, *et al.* Melatonin and mitochondrial dysfunction in the central nervous system. *Horm Behav* 2013;63:322–30.
- 87 Jou MJ, Peng TI, Yu PZ, *et al.* Melatonin protects against common deletion of mitochondrial DNA-augmented mitochondrial oxidative stress and apoptosis. *J Pineal Res* 2007;43:389–403.
- 88 Pandi-Perumal SR, BaHammam AS, Brown GM, *et al.* Melatonin antioxidative defense: therapeutical implications for aging and neurodegenerative processes. *Neurotox Res* 2013;23:267–300.
- 89 Sharma R, Ottenhof T, Rzeczowska PA, *et al.* Epigenetic targets for melatonin: induction of histone H3 hyperacetylation and gene expression in C17.2 neural stem cells. *J Pineal Res* 2008;45:277–84.
- 90 Thomas L, Purvis CC, Drew JE, *et al.* Melatonin receptors in human fetal brain: 2-[(125)I]iodomelatonin binding and MT1 gene expression. *J Pineal Res* 2002;33:218–24.
- 91 Tamura H, Nakamura Y, Korkmaz A, *et al.* Melatonin and the ovary: physiological and pathophysiological implications. *Fertil Steril* 2009;92:328–43.
- 92 Tamura H, Nakamura Y, Terron MP, *et al.* Melatonin and pregnancy in the human. *Reprod Toxicol* 2008;25:291–303.
- 93 Okatani Y, Okamoto K, Hayashi K, *et al.* Maternal-fetal transfer of melatonin in pregnant women near term. *J Pineal Res* 1998;25:129–34.
- 94 Okatani Y, Wakatsuki A, Kaneda C. Melatonin increases activities of glutathione peroxidase and superoxide dismutase in fetal rat brain. *J Pineal Res* 2000;28:89–96.
- 95 Ardura J, Gutierrez R, Andres J, *et al.* Emergence and evolution of the circadian rhythm of melatonin in children. *Horm Res* 2003;59:66–72.
- 96 Kennaway DJ, Stamp GE, Goble FC. Development of melatonin production in infants and the impact of prematurity. *J Clin Endocrinol Metab* 1992;75:367–9.
- 97 Marseglia L, Aversa S, Barberi I, *et al.* High endogenous melatonin levels in critically ill children: a Pilot study. *J Pediatr* 2013;162:357–60.
- 98 Seifman MA, Adamides AA, Nguyen PN, *et al.* Endogenous melatonin increases in cerebrospinal fluid of patients after severe traumatic brain injury and correlates with oxidative stress and metabolic disarray. *Cereb Blood Flow Metab* 2008;28:684–96.
- 99 Fu J, Zhao SD, Liu HJ, *et al.* Melatonin promotes proliferation and differentiation of neural stem cells subjected to hypoxia in vitro. *J Pineal Res* 2011;51:104–12.
- 100 Husson I, Mesples B, Bac P, *et al.* Melatonergic neuroprotection of the murine periventricular white matter against neonatal excitotoxic challenge. *Ann Neurol* 2002;51:82–92.
- 101 Villapol S, Fau S, Renolleau S, *et al.* Melatonin promotes myelination by decreasing white matter inflammation after neonatal stroke. *Pediatr Res* 2011;69:51–5.
- 102 Pearce W. Hypoxic regulation of the fetal cerebral circulation. *J Appl Physiol* 2006;100:731–8.
- 103 Vento M, Escobar J, Cernada M, *et al.* The use and misuse of oxygen during the neonatal period. *Clin Perinatol* 2012;39:165–76.
- 104 Wakatsuki A, Okatani Y, Izumiya C, *et al.* Melatonin protects against ischemia and reperfusion-induced oxidative lipid and DNA damage in fetal rat brain. *J Pineal Res* 1999;26:147–52.
- 105 Galano A, Tan DX, Reiter RJ. On the free radical scavenging activities of melatonin's metabolites, AFMK and AMK. *J Pineal Res* 2013;54:245–57.
- 106 Gilad E, Cuzzocrea S, Zingarelli B, *et al.* Melatonin is a scavenger of peroxynitrite. *Life Sci* 1997;60:PL169–74.
- 107 Hardeland R, Tan DX, Reiter RJ. Kynuramines, metabolites of melatonin and other indoles: the resurrection of an almost forgotten class of biogenic amines. *J Pineal Res* 2009;47:109–26.
- 108 Ressmeyer AR, Mayo JC, Zelosko V, *et al.* Antioxidant properties of the melatonin metabolite N1-acetyl-5-methoxykynuramine (AMK): scavenging of free radicals and prevention of protein destruction. *Redox Rep* 2003;8:205–13.
- 109 Tan DX, Manchester LC, Terron MP, *et al.* One molecule, many derivatives: a never-ending interaction of melatonin with reactive oxygen and nitrogen species? *J Pineal Res* 2007;42:28–42.
- 110 Miller SL, Yan EB, Castillo-Melendez M, *et al.* Melatonin provides neuroprotection in the late-gestation fetal sheep brain in response to umbilical cord occlusion. *Dev Neurosci* 2005;27:200–10.
- 111 Wakatsuki A, Izumiya C, Okatani Y, *et al.* Oxidative damage in fetal rat brain induced by ischemia and subsequent reperfusion. Relation to arachidonic acid peroxidation. *Biol Neonate* 1999;76:84–91.
- 112 Wakatsuki A, Okatani Y, Shinohara K, *et al.* Melatonin protects against ischemia/reperfusion-induced oxidative damage to mitochondria in fetal rat brain. *J Pineal Res* 2001;31:167–72.
- 113 Wakatsuki A, Okatani Y, Shinohara K, *et al.* Melatonin protects fetal rat brain against oxidative mitochondrial damage. *J Pineal Res* 2001;30:22–8.
- 114 Watanabe K, Hamada F, Wakatsuki A, *et al.* Prophylactic administration of melatonin to the mother throughout pregnancy can protect against oxidative cerebral damage in neonatal rats. *J Matern Fetal Neonatal Med* 2012;25:1254–9.
- 115 Watanabe K, Wakatsuki A, Shinohara K, *et al.* Maternally administered melatonin protects against ischemia and reperfusion-induced oxidative mitochondrial damage in premature fetal rat brain. *J Pineal Res* 2004;37:276–80.
- 116 Yawno T, Castillo-Melendez M, Jenkin G, *et al.* Mechanisms of melatonin-induced protection in the brain of late gestation fetal sheep in response to hypoxia. *Dev Neurosci* 2012;34:543–51.
- 117 Cetinkaya M, Alkan T, Ozyener F, *et al.* Possible neuroprotective effects of magnesium sulfate and melatonin as both pre- and post-treatment in a neonatal hypoxic-ischemic rat model. *Neonatology* 2011;99:302–10.
- 118 Hutton LC, Abbas M, Dickinson H, *et al.* Neuroprotective properties of melatonin in a model of birth asphyxia in the spiny mouse (*Acomys cahirinus*). *Dev Neurosci* 2009;31:437–51.
- 119 Kaur C, Sivakumar V, Ling EA. Melatonin protects periventricular white matter from damage due to hypoxia. *J Pineal Res* 2010;48:185–93.
- 120 Ozyener F, Cetinkaya M, Alkan T, *et al.* Neuroprotective effects of melatonin administered alone or in combination with topiramate in neonatal hypoxic-ischemic rat model. *Restor Neurol Neurosci* 2012;30:435–44.
- 121 Welin AK, Svedin P, Lapatto R, *et al.* Melatonin reduces inflammation and cell death in white matter in the mid-gestation fetal sheep following umbilical cord occlusion. *Pediatr Res* 2007;61:153–8.
- 122 Carrillo-Vico A, Lardone PJ, Fernandez-Santos JM, *et al.* Human lymphocyte-synthesized melatonin is involved in the regulation of the interleukin-2/interleukin-2 receptor system. *J Clin Endocrinol Metab* 2005;90:992–1000.
- 123 Srinivasan V, Pandi-Perumal SR, Spence DW, *et al.* Melatonin in septic shock: some recent concepts. *J Crit Care* 2010;25:656 e1–6.
- 124 Balduini W, Carloni S, Perrone S, *et al.* The use of melatonin in hypoxic-ischemic brain damage: an experimental study. *J Matern Fetal Neonatal Med* 2012;25 (Suppl 1):119–24.
- 125 Wang X, Svedin P, Nie C, *et al.* N-acetylcysteine reduces lipopolysaccharide-sensitized hypoxic-ischemic brain injury. *Ann Neurol* 2007;61:263–71.
- 126 Jahnke G, Marr M, Myers C, *et al.* Maternal and developmental toxicity evaluation of melatonin administered orally to pregnant Sprague-Dawley rats. *Toxicol Sci* 1999;50:271–9.
- 127 Buscemi N, Vandermeer B, Hooton N, *et al.* Efficacy and safety of exogenous melatonin for secondary sleep disorders and sleep disorders accompanying sleep restriction: meta-analysis. *BMJ* 2006;332:385–93.
- 128 Gitto E, Karbownik M, Reiter RJ, *et al.* Effects of melatonin treatment in septic newborns. *Pediatr Res* 2001;50:756–60.
- 129 Gitto E, Reiter RJ, Cordaro SP, *et al.* Oxidative and inflammatory parameters in respiratory distress syndrome of preterm newborns: beneficial effects of melatonin. *Am J Perinatol* 2004;21:209–16.
- 130 Fullia F, Gitto E, Cuzzocrea S, *et al.* Increased levels of malondialdehyde and nitrite/nitrate in the blood of asphyxiated newborns: reduction by melatonin. *J Pineal Res* 2001;31:343–9.

- 131 Aly H, Elmahdy H, El-Dib M, *et al.* Melatonin use for neuroprotection in perinatal asphyxia: a randomized controlled pilot study. *J Perinatol* 2014 doi: 10.1038/jp.2014.186. [Epub ahead of print].
- 132 Robertson NJ, Tan S, Groenendaal F, *et al.* Which neuroprotective agents are ready for bench to bedside translation in the newborn infant? *J Pediatr* 2012;160:544–52 e4.
- 133 Pacher P, Batkai S, Kunos G. The endocannabinoid system as an emerging target of pharmacotherapy. *Pharmacol Rev* 2006;58:389–462.
- 134 Ahn K, McKinney MK, Cravatt BF. Enzymatic pathways that regulate endocannabinoid signaling in the nervous system. *Chem Rev* 2008;108:1687–707.
- 135 Di Marzo V, Bisogno T, De Petrocellis L. Endocannabinoids and related compounds: walking back and forth between plant natural products and animal physiology. *Chem Biol* 2007;14:741–56.
- 136 Berger C, Schmid PC, Schabitz WR, *et al.* Massive accumulation of N-acyl ethanolamines after stroke. Cell signalling in acute cerebral ischemia? *J Neurochem* 2004;88:1159–67.
- 137 Fernandez-Ruiz J, Berrendero F, Hernandez ML, *et al.* The endogenous cannabinoid system and brain development. *Trends Neurosci* 2000;23:14–20.
- 138 Gaffuri AL, Ladarre D, Lenkei Z. Type-1 cannabinoid receptor signaling in neuronal development. *Pharmacology* 2012;90:19–39.
- 139 Harkany T, Keimpema E, Barabas K, *et al.* Endocannabinoid functions controlling neuronal specification during brain development. *Mol Cell Endocrinol* 2008;286(1–2 Suppl 1):S84–90.
- 140 Mato S, Del Olmo E, Pazos A. Ontogenetic development of cannabinoid receptor expression and signal transduction functionality in the human brain. *Eur J Neurosci* 2003;17:1747–54.
- 141 Paria BC, Dey SK. Ligand-receptor signaling with endocannabinoids in preimplantation embryo development and implantation. *Chem Phys Lipids* 2000;108:211–20.
- 142 Hansen HH, Ikonomidou C, Bittigau P, *et al.* Accumulation of the anandamide precursor and other N-acyl ethanolamine phospholipids in infant rat models of in vivo necrotic and apoptotic neuronal death. *J Neurochem* 2001;76:39–46.
- 143 Panikashvili D, Simeonidou C, Ben-Shabat S, *et al.* An endogenous cannabinoid (2-AG) is neuroprotective after brain injury. *Nature* 2001;413:527–31.
- 144 Sugiura T, Yoshinaga N, Kondo S, *et al.* Generation of 2-arachidonoylglycerol, an endogenous cannabinoid receptor ligand, in picrotoxinin-administered rat brain. *Biochem Biophys Res Commun* 2000;271:654–8.
- 145 van der Stelt M, Di Marzo V. Cannabinoid receptors and their role in neuroprotection. *Neuromolecular Med* 2005;7:37–50.
- 146 Freund TF, Katona I, Piomelli D. Role of endogenous cannabinoids in synaptic signaling. *Physiol Rev* 2003;83:1017–66.
- 147 Kim SH, Won SJ, Mao XO, *et al.* Molecular mechanisms of cannabinoid protection from neuronal excitotoxicity. *Mol Pharmacol* 2006;69:691–6.
- 148 Marsicano G, Goodenough S, Monory K, *et al.* CB1 cannabinoid receptors and on-demand defense against excitotoxicity. *Science* 2003;302:84–8.
- 149 van der Stelt M, Veldhuis WB, Maccarrone M, *et al.* Acute neuronal injury, excitotoxicity, and the endocannabinoid system. *Mol Neurobiol* 2002;26:317–46.
- 150 Waksman Y, Olson JM, Carlisle SJ, *et al.* The central cannabinoid receptor (CB1) mediates inhibition of nitric oxide production by rat microglial cells. *J Pharmacol Exp Ther* 1999;288:1357–66.
- 151 Klein TW. Cannabinoid-based drugs as anti-inflammatory therapeutics. *Nature Rev Immunol* 2005;5:400–11.
- 152 Murकिनати S, Juttler E, Keinert T, *et al.* Activation of cannabinoid 2 receptors protects against cerebral ischemia by inhibiting neutrophil recruitment. *FASEB J* 2010;24:788–98.
- 153 Stella N. Endocannabinoid signaling in microglial cells. *Neuropharmacology* 2009;56(Suppl 1):244–53.
- 154 Tanasescu R, Constantinescu CS. Cannabinoids and the immune system: an overview. *Immunobiology* 2010;215:588–97.
- 155 Walter L, Stella N. Cannabinoids and neuroinflammation. *Br J Pharmacol* 2004;141:775–85.
- 156 Guzman M, Sanchez C, Galve-Roperh I. Control of the cell survival/death decision by cannabinoids. *J Mol Med* 2001;78:613–25.
- 157 Guzman M, Sanchez C, Galve-Roperh I. Cannabinoids and cell fate. *Pharmacol Ther* 2002;95:175–84.
- 158 Molina-Holgado E, Vela JM, Arevalo-Martin A, *et al.* Cannabinoids promote oligodendrocyte progenitor survival: involvement of cannabinoid receptors and phosphatidylinositol-3 kinase/Akt signaling. *J Neurosci* 2002;22:9742–53.
- 159 Ozaita A, Puighearnan E, Maldonado R. Regulation of PI3K/Akt/GSK-3 pathway by cannabinoids in the brain. *J Neurochem* 2007;102:1105–14.
- 160 Ramirez SH, Hasko J, Skuba A, *et al.* Activation of cannabinoid receptor 2 attenuates leukocyte-endothelial cell interactions and blood-brain barrier dysfunction under inflammatory conditions. *J Neurosci* 2012;32:4004–16.
- 161 Visconti MT, Oddi S, Latini L, *et al.* Selective CB2 receptor agonism protects central neurons from remote axotomy-induced apoptosis through the PI3K/Akt pathway. *J Neurosci* 2009;29:4564–70.
- 162 Alonso-Alconada D, Alvarez A, Alvarez FJ, *et al.* The cannabinoid WIN 55212-2 mitigates apoptosis and mitochondrial dysfunction after hypoxia ischemia. *Neurochem Res* 2012;37:161–70.
- 163 Alonso-Alconada D, Alvarez FJ, Alvarez A, *et al.* The cannabinoid receptor agonist WIN 55212-2 reduces the initial cerebral damage after hypoxic-ischemic injury in fetal lambs. *Brain Res* 2010;1362:150–9.
- 164 Fernandez-Lopez D, Pazos MR, Tolon RM, *et al.* The cannabinoid agonist WIN55212 reduces brain damage in an in vivo model of hypoxic-ischemic encephalopathy in newborn rats. *Pediatr Res* 2007;62:255–60.
- 165 Fernandez-Lopez D, Pradillo JM, Garcia-Yebenes I, *et al.* The cannabinoid WIN55212-2 promotes neural repair after neonatal hypoxia-ischemia. *Stroke* 2010;41:2956–64.
- 166 Lafuente H, Alvarez FJ, Pazos MR, *et al.* Cannabidiol reduces brain damage and improves functional recovery after acute hypoxia-ischemia in newborn pigs. *Pediatr Res* 2011;70:272–7.
- 167 Pazos MR, Mohammed N, Lafuente H, *et al.* Mechanisms of cannabidiol neuroprotection in hypoxic-ischemic newborn pigs: role of 5HT(1A) and CB2 receptors. *Neuropharmacology* 2013;71:282–91.
- 168 Leker RR, Gai N, Mechoulam R, *et al.* Drug-induced hypothermia reduces ischemic damage: effects of the cannabinoid HU-210. *Stroke* 2003;34:2000–6.
- 169 Borgelt LM, Franson KL, Nussbaum AM, *et al.* The pharmacologic and clinical effects of medical cannabis. *Pharmacotherapy* 2013;33:195–209.
- 170 Croxford JL. Therapeutic potential of cannabinoids in CNS disease. *CNS Drugs* 2003;17:179–202.
- 171 Maas AI, Murray G, Henney H III, *et al.* Efficacy and safety of dexanabinol in severe traumatic brain injury: results of a phase III randomised, placebo-controlled, clinical trial. *Lancet Neurol* 2006;5:38–45.
- 172 Rangarajan V, Juul SE. Erythropoietin: emerging role of erythropoietin in neonatal neuroprotection. *Pediatr Neurol* 2014;51:481–8.
- 173 Morishita E, Masuda S, Nagao M, *et al.* Erythropoietin receptor is expressed in rat hippocampal and cerebral cortical neurons, and erythropoietin prevents in vitro glutamate-induced neuronal death. *Neuroscience* 1997;76:105–16.
- 174 Nagai A, Nakagawa E, Choi HB, *et al.* Erythropoietin and erythropoietin receptors in human CNS neurons, astrocytes, microglia, and oligodendrocytes grown in culture. *J Neuropathol Exp Neurol* 2001;60:386–92.
- 175 Yamaji R, Okada T, Moriya M, *et al.* Brain capillary endothelial cells express two forms of erythropoietin receptor mRNA. *Eur J Biochem* 1996;239:494–500.
- 176 Marti HH, Bernaudin M, Petit E, *et al.* Neuroprotection and angiogenesis: dual role of erythropoietin in brain ischemia. *News Physiol Sci* 2000;15:225–9.
- 177 Kilic E, Ozdemir YG, Bolay H, *et al.* Pinealectomy aggravates and melatonin administration attenuates brain damage in focal ischemia. *J Cereb Blood Flow Metab* 1999;19:511–6.
- 178 Chen ZY, Asavaritkrai P, Prchal JT, *et al.* Endogenous erythropoietin signaling is required for normal neural progenitor cell proliferation. *J Biol Chem* 2007;282:25875–83.
- 179 Prass K, Scharff A, Ruscher K, *et al.* Hypoxia-induced stroke tolerance in the mouse is mediated by erythropoietin. *Stroke* 2003;34:1981–6.
- 180 Ferriero DM. Protecting neurons. *Epilepsia* 2005;46(Suppl 7):45–51.
- 181 Juul SE, Beyer RP, Bammler TK, *et al.* Microarray analysis of high-dose recombinant erythropoietin treatment of unilateral brain injury in neonatal mouse hippocampus. *Pediatr Res* 2009;65:485–92.
- 182 Maiese K, Chong ZZ, Hou J, *et al.* Erythropoietin and oxidative stress. *Curr Neurovasc Res* 2008;5:125–42.
- 183 Villa P, van Beek J, Larsen AK, *et al.* Reduced functional deficits, neuroinflammation, and secondary tissue damage after treatment of stroke by nonerythropoietic erythropoietin derivatives. *Cereb Blood Flow Metab* 2007;27:552–63.
- 184 Gonzalez FF, Larphaveesarp A, McQuillen P, *et al.* Erythropoietin increases neurogenesis and oligodendroglial precursor cells after neonatal stroke. *Stroke* 2013;44:753–8.
- 185 Xiong Y, Mahmood A, Meng Y, *et al.* Delayed administration of erythropoietin reducing hippocampal cell loss, enhancing angiogenesis and neurogenesis, and improving functional outcome following traumatic brain injury in rats: comparison of treatment with single and triple dose. *J Neurosurg* 2010;113:598–608.
- 186 Kaneko N, Kako E, Sawamoto K. Enhancement of ventricular-subventricular zone-derived neurogenesis and oligodendrogenesis by erythropoietin and its derivatives. *Front Cell Neurosci* 2013;7:235.
- 187 Wang L, Zhang ZG, Zhang RL, *et al.* Matrix metalloproteinase 2 (MMP2) and MMP9 secreted by erythropoietin-activated endothelial cells promote neural progenitor cell migration. *J Neurosci* 2006;26:5996–6003.
- 188 Fan X, Heijnen CJ, van der KM, *et al.* Beneficial effect of erythropoietin on sensorimotor function and white matter after hypoxia-ischemia in neonatal mice. *Pediatr Res* 2011;69:56–61.
- 189 Gonzalez FF, Abel R, Almlil CR, *et al.* Erythropoietin sustains cognitive function and brain volume after neonatal stroke. *Dev Neurosci* 2009;31:403–11.
- 190 van de Looij Y, Chatagner A, Quairiaux C, *et al.* Multi-modal assessment of long-term erythropoietin treatment after neonatal hypoxic-ischemic injury in rat brain. *PLoS ONE* 2014;9:e95643.
- 191 Traudt CM, McPherson RJ, Bauer LA, *et al.* Concurrent erythropoietin and hypothermia treatment improve outcomes in a term nonhuman primate model of perinatal asphyxia. *Dev Neurosci* 2013;35:491–503.

- 192 McPherson RJ, Demers EJ, Juul SE. Safety of high-dose recombinant erythropoietin in a neonatal rat model. *Neonatology* 2007;91:36–43.
- 193 Juul SE, McPherson RJ, Bauer LA, *et al.* A phase I/II trial of high-dose erythropoietin in extremely low birth weight infants: pharmacokinetics and safety. *Pediatrics* 2008;122:383–91.
- 194 Benders MJ, van der Aa NE, Roks M, *et al.* Feasibility and safety of erythropoietin for neuroprotection after perinatal arterial ischemic stroke. *J Pediatr* 2014;164:481–6 e1–2.
- 195 Wu YW, Bauer LA, Ballard RA, *et al.* Erythropoietin for neuroprotection in neonatal encephalopathy: safety and pharmacokinetics. *Pediatrics* 2012;130:683–91.
- 196 Sanchis-Gomar F, Perez-Quilis C, Lippi G. Erythropoietin receptor (EpoR) agonism is used to treat a wide range of disease. *Mol Med* 2013;19:62–4.
- 197 Rogers EE, Bonifacio SL, Glass HC, *et al.* Erythropoietin and hypothermia for hypoxic-ischemic encephalopathy. *Pediatr Neurol* 2014;51:657–62.
- 198 Kellert BA, McPherson RJ, Juul SE. A comparison of high-dose recombinant erythropoietin treatment regimens in brain-injured neonatal rats. *Pediatr Res* 2007;61:451–5.
- 199 El Shimi MS, Awad HA, Hassanein SM, *et al.* Single dose recombinant erythropoietin versus moderate hypothermia for neonatal hypoxic ischemic encephalopathy in low resource settings. *J Matern Fetal Neonatal Med* 2014;27:1295–300.