

RESEARCH ARTICLE

Progesterone Treatment Shows Benefit in Female Rats in a Pediatric Model of Controlled Cortical Impact Injury

Rastafa I. Geddes, Bethany L. Peterson, Donald G. Stein*, Iqbal Sayeed

Department of Emergency Medicine, Emory University, Atlanta, GA 30322 United States of America

* dstei04@emory.edu



Abstract

Purpose

We recently showed that progesterone treatment can reduce lesion size and behavioral deficits after moderate-to-severe bilateral injury to the medial prefrontal cortex in immature male rats. Whether there are important sex differences in response to injury and progesterone treatment in very young subjects has not been given sufficient attention. Here we investigated progesterone's effects in the same model of brain injury but with pre-pubescent females.

Methods

Twenty-eight-day-old female Sprague-Dawley rats received sham (n = 14) or controlled cortical impact (CCI) (n = 21) injury, were given progesterone (8 mg/kg body weight) or vehicle injections on post-injury days (PID) 1–7, and underwent behavioral testing from PID 9–27. Brains were evaluated for lesion size at PID 28.

Results

Lesion size in vehicle-treated female rats with CCI injury was smaller than that previously reported for similarly treated age-matched male rats. Treatment with progesterone reduced the effect of CCI on extent of damage and behavioral deficits.

Conclusion

Pre-pubescent female rats with midline CCI injury to the frontal cortex have reduced morphological and functional deficits following progesterone treatment. While gender differences in susceptibility to this injury were observed, progesterone treatment produced beneficial effects in young rats of both sexes following CCI.

OPEN ACCESS

Citation: Geddes RI, Peterson BL, Stein DG, Sayeed I (2016) Progesterone Treatment Shows Benefit in Female Rats in a Pediatric Model of Controlled Cortical Impact Injury. PLoS ONE 11(1): e0146419. doi:10.1371/journal.pone.0146419

Editor: Cheryl M McCormick, Brock University, CANADA

Received: October 14, 2015

Accepted: December 16, 2015

Published: January 22, 2016

Copyright: © 2016 Geddes et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: All relevant data are within the paper and its Supporting Information files.

Funding: This work was supported by the Emory Center for Injury Control, National Institutes of Health grant no. R49CE001494-04 to DGS, the Emory University Summer Undergraduate Research Experience (SURE) program, and by unrestricted gifts to the Brain Research Laboratory from BHR Pharma and Allen and Company. There were no employer/employee relationships with either of the companies although DGS had a consulting agreement with BHR to provide pre-clinical information on the use of progesterone for the

treatment of TBI. None of the authors currently participate in any licensing agreement or receive any royalties from BHR Pharma. Allen and Company is an investment firm with no commercial interests in pharmacology or any other matter relating to the authors' research. None of the funders had any role in study design, data collection and analysis, decision to publish, or preparation of this or any other manuscript.

Competing Interests: These unrestricted gifts made through the Emory Development Office do not alter the authors' adherence to PLOS ONE policies on sharing data and materials. DGS and IS hold the following patents: Methods for the treatment of a traumatic central nervous system injury, No. EP2030622B1, date approved: 2/23/2011, Method for the treatment of a traumatic central nervous system injury via a tapered administration of progesterone, No. 1871382, date approved: 8/17/2011. They do not have any licensing agreements pertaining to the patents and having the patents does not alter the authors' adherence to PLOS ONE policies on sharing data and materials.

Introduction

Traumatic Brain Injury (TBI) has a world-wide incidence rate of 106 per 100,000 population [1], and no FDA-approved therapy currently exists [2,3]. Globally, adolescents have the highest TBI rates of any age group [4–7] and males are nearly three times as likely as females to die from a TBI [1]. Current statistics suggest that gender may play a role, with females lower in TBI susceptibility, extent of injury and prognosis.

Although progesterone (PROG) has been shown to be beneficial in pre-clinical laboratory research in multiple models of central nervous system (CNS) injuries including TBI [8–15], several dozen trials over the last 15–20 years attempting to treat adult TBI have all produced negative outcomes [16], and two recent Phase III clinical trials, SyNAPSe (ClinicalTrials.gov Identifier: NCT01143064) and ProTECT III (NCT01143064), reported no significant beneficial effects of acute PROG treatment on moderate to severe closed-head TBI in adult males and females [17,18]. Unfortunately, these trials did not conduct dosing and duration of treatment optimization studies prior to testing the hormone in patients and had other design problems [19,20]. In addition, although the trials did not directly study sex differences, sex differences in variability of injury severity, outcomes, dose-optimization [19], post-acute rehabilitation, and co-morbidities could have been a factor in the results (see [20,21] for more details).

Whether PROG's neuroprotective effects after brain injury vary in males and females across the developmental spectrum is still an open question. For example, following neonatal hypoxic-ischemic injury in both male and female rats, PROG-treated males surprisingly showed much more substantial tissue sparing and less reactive gliosis than females and there were significant sex differences in behavioral outcomes when the animals were tested later in life [22]. However, in c57BALB mice, sex differences in response to a cortical contusion injury were seen in only a few measures of activity—in cognitive and motor tasks, the deficits were the same [23]. Recent individual animal meta-analyses of a number of published preclinical studies of PROG in females with stroke [24] showed an increase in the incidence of stroke-related death in adult females, highlighting the need for investigations to evaluate how the female subject may differentially respond to brain injury.

PROG is an important sex steroid as well as a developmental hormone, so young females with brain injury just entering puberty/estrus may be more susceptible to rapid changes in hormonal levels of PROG that could result in different morphological and functional outcomes compared to male conspecifics or older subjects with similar damage. Robertson et al. [25] reported that tissue loss was reduced in PROG-treated female rats at 7d after unilateral contusion injury to the exposed brain and suggested a need for future studies looking at functional outcome measures. Recently, we showed that PROG treatment reduced lesion size and behavioral deficits after moderate-to-severe bilateral injury to the medial prefrontal cortex (mPFC) in post-natal day (PND) 28 male Sprague-Dawley (SD) rats [26]. Here we report on the response of pre-pubescent female rats with a similar TBI to post-injury PROG treatment. We think it is important to analyze responses to brain injury by gender as well as by stage of development, especially when critical sex hormones may affect functional and morphological outcomes. We tested the hypothesis that PND 28 female rats with a controlled cortical impact (CCI) injury would show the same benefits of neurosteroid treatment as their age-matched male conspecifics. Rats do not become sexually mature until about 6–8 weeks [27], so the present study reports a model of pediatric brain injury for evaluating PROG treatment following TBI in animals at an age range equivalent to that of a 9–10-year-old human child. This study could provide important information in designing future clinical trials of PROG treatment in children and adolescents with TBI.

Materials and Methods

Subjects

Forty-eight SD rats (Harlan) were acquired at PND 21 and acclimated to the environment over 2 days. Rats were weighed on PND 23–25 and daily thereafter and housed, fed and maintained on a 12-hour reverse light/dark cycle. The Institutional Animal Care and Use Committee of Emory University approved the procedures used in this study and the research was conducted in an AAALAC-approved facility (Protocol # 2001801).

Surgery

Surgeries were performed on PND 28. For initial anesthesia, the rats were placed in an air-tight induction chamber with oxygen, nitrous oxide, and isoflurane gas (4% induction, 1.5% maintenance, 700 mmHg/min N₂O, and 500 mmHg/min O₂). They were then mounted in a Kopf stereotaxic device (model 900) equipped with a Mouse and Neonatal Rat Adaptor (model BJK-030). The animal's head was held in place by non-traumatic ear bars and a bite bar. Anesthesia was delivered just prior to the surgery by nose cone and the rats' heads shaved and sterilized with 70% ethanol and Betadine™ antiseptic solution. Anesthesia levels were monitored closely throughout surgery and were frequently adjusted between 400–700 mmHg/min, based on heart rate and oxygen saturation. In our previous study [26] we found that a 4.0-mm diameter stainless steel impactor resulted in a survivable, severe CCI injury in 28-day-old male rats so we used the same CCI parameters for the age-matched female rats.

A SurgiVet™ pulse oximeter (model V3304) was attached to the animal's rear paw to monitor and maintain blood SpO₂ at or above 90%. A heart rate monitor with its sensor attached to the other hind paw was used to maintain a rate \geq 300 beats per minute. A homeothermic blanket control unit (Harvard Apparatus, Holliston, MA) was used to monitor and maintain core body temperature (\sim 37°C) and prevent hypothermia throughout surgery. Under aseptic conditions, the cranium and its bony landmarks including bregma (β) and lambda (λ) were exposed by making a midline incision along the scalp into the skin and fascia covering the skull. The craniectomy was centered on the midline at 2.0 mm anterior to β . The cortical impact was made over the midline medial frontal cortex with an Impact One™ Stereotaxic Impactor for CCI (Leica #39463920). The sham-injured groups received the same surgical procedures up to and including craniectomy but no CCI injury. After surgery, the rats were placed on a heating pad, monitored closely and upon awakening were returned to their home cages. Of the 48 females at the beginning of the experiment, 3 died under surgery.

PROG solution and injection schedule

The remaining 45 rats were randomly assigned to one of 4 groups: CCI+PROG (8 mg/kg; n = 12); CCI+vehicle (n = 11); sham-vehicle (n = 11); and sham+PROG. CCI+PROG received injections of 8 mg/kg of PROG (4-pregnene-3, 20-dione, Sigma Aldrich, St. Louis, MO) dissolved in 22.5% 2-hydroxypropyl- β -cyclodextrin (HBC) (Sigma). The incoming animals were allocated to each of the groups by non-systematic selection and assignment soon after they were delivered to the animal housing quarters. Once assigned to a group they were numbered and then the group was coded for blinding purposes until the completion of the experiment. The solutions containing PROG and vehicle for CCI and sham groups were independently coded prior to administration so that investigators were blinded to the agents being injected.

The dose used here was the same as that found to be most beneficial in our previous experiment in similarly aged males [26]. PROG was administered at a volume of 0.02 ml/100 gm body weight. However, the sham+PROG group was given 16 mg/kg doses of the drug (1) to

permit comparisons with the highest dose previously tested in the sham-injured juvenile male rats [26], and (2) to demonstrate the safety of a “higher-than-effective” dose of PROG in our normal or non-injured juvenile female subjects. A total of 9 injections were administered to each animal in all experimental conditions at the following post-injury times: 1, 3, and 24 h, and 2, 3, 4, 5, 6, and 7 days. All injections were given subcutaneously except the first, which was administered intraperitoneally to ensure more rapid absorption following injury [28,29]. To avoid withdrawal effects, the 8th and 9th injections of PROG were tapered (one-half and one-quarter of the original dose, respectively) [30,31]. PROG was prepared just prior to surgery and again on the 4th post-injury night.

Behavioral testing

We used the same regimen of behavioral testing for the females as for the males [26]. To obtain baseline data, rotarod performance and behavior in the elevated plus maze (EPM) were assessed in all rats prior to surgery. All post-surgery testing was delayed for one week, during which the rats received daily vehicle or PROG injections as described above. Animal IDs were coded to keep experimenters blind to group identity throughout behavioral testing and histological analysis.

Anxiety-like behavior in the EPM. In the EPM, anxiety is computed by determining the amount of time spent in the open vs. the closed arms. This is because in novel situations, rats tend to exhibit thigmotactic behavior—huddling next to walls or enclosed spaces, which provide mechanical stimulation. Because lab rats are naturally exploratory, reduced thigmotaxis (inferred from time spent exploring the open arm) is taken as an indication of lowered stress or anxiety. Thus the total number of open vs. closed arm entries was used as the operational dependent measure. If the animal fell off the open arm it was returned to the start position in the center square and the fall recorded. Individual trial data for each rat was used to determine group averages.

Testing was conducted under red light in a quiet environment. Baseline EPM data was obtained between PND 25 and 27 and rats were tested twice post-surgery (on post-injury (PID) 9 and 17). Each trial lasted 5 min, and the total number of open arm entries (or crosses over the center square with both front and hind paws) was reported as percent of visits to the open and closed arms.

Rotarod testing. An accelerating Exconomex™ rotarod (Columbus Instruments, Columbus, OH) was used under red light to assess balance and motor coordination. Rats were given initial habituation training and then baseline testing prior to surgery. For habituation training, rats were placed on a stationary rod for at least 3 min, then slowly habituated to the rotating rod from a starting speed of 1 rpm, which was accelerated to 30 rpm over 5 min. A baseline score was obtained on PND 27 and rats were again tested on PID 9, 15, 21, 25 and 27. Rats were scored on latency to fall off the rod (maximum score was 300 s), and each testing day consisted of three trials separated by a 10-min break. Scores from the three trials for each day were averaged.

Spatial navigation performance and memory in the MWM. MWM tests were conducted under dim white light in a white plastic pool measuring 135 cm in diameter [26]. The rat's position in the maze, swim distance, and latency to reach the platform was recorded with an overhead camera and computer-assisted tracking system (CleverSys, Reston, VA).

Beginning on PID 10, all the rats were tested for acquisition in the MWM. Each animal received two trials per day, separated by a 5-min interval. A trial consisted of placing the subject in the pool facing the wall and allowing it to swim until it reached the platform or until 90 sec had elapsed. When a rat was unable to locate the MWM platform within the allotted time,

the experimenter led the rat to the platform. Rats were allowed to remain on the platform for 20 sec and then removed from the pool. After 5 min, subjects were again released into the tank, but this time from the opposite position from that in the previous trial and allowed to swim to the platform. Animals were placed in holding cages in front of an air heater between trials and before being returned to their home cages.

Histology

Like the previously tested males, at 1 month post-injury, the females were fatally anesthetized and their brains perfused with saline, then with 10% formalin, and then extracted for histological analysis. The brains were cut into 20- μ m sections on a cryostat and stored at -80°C on 1% gelatin coated (subbed) slides. Slides were stained in 0.1% cresyl violet solution (0.1 g cresyl violet acetate and 0.3 ml of glacial acetic acid dissolved in 100 ml dH_2O) for 10 min at 45°C and then rinsed in distilled water. The percent of damaged issue was identified in six 20-micron Nissl-stained sections and analyzed from between 4.5 and -0.5 mm from β for lesion size using the Image J[™] System (Media Cybernetics, Silver Spring, MD).

Stained slides were scanned with Silverfast Pathscan software (PathScan Enabler IV, Meyer Instruments, Houston TX) and the scanned images were analyzed using ImageJ[™]. The percentage of injured tissue from a single section was calculated by tracing the perimeter of the injury and determining its surface area, dividing this by an estimate of the total surface area of the section (taken by tracing both the remaining tissue and the estimated perimeter of the necrotic cavity), and multiplying by 100.

Qualitative observations on females surviving the TBI

Of the 22 sham rats, the data for 8 were removed from final analysis. The reasons for removal included: failure to meet the criterion of 180 ms on the rotarod during baseline testing ($n = 2$); evidence of unreliable data during post-surgery testing ($n = 5$) due to attempts to escape the apparatus or hugging the rod (thus missing the sensor); and an unintentional injury to brain from the drill bit during craniectomy ($n = 1$). The data for 14 out of 22 sham-injured females (8 in the sham+vehicle and 6 in the sham+PROG groups) are presented in Figs 1–3. Of the 23 surviving rats with CCI injury, one failed to meet rotarod baseline criteria and another repeatedly attempted to escape the apparatus rather than run on the wheel. Thus, the data for 21 out of 23 rats with CCI injury are also presented in Figs 1–3, with $n = 10$ rats in the CCI+vehicle and $n = 11$ rats in the CCI+PROG group.

Statistical Analysis

A mixed factorial analysis of variance (ANOVA) for repeated measures was performed on the behavioral results and lesion reconstruction data, which were expressed as the mean \pm SEM. Mean comparisons were used for *post-hoc* analyses of repeated ANOVAs. Independent paired *t*-tests were also used to compare the differences between baseline (pre-injury) and post-injury data when data were normally distributed. Statistical significance was established at $p \leq 0.05$. Based on a delta-value of 1.5 (using latency to find the hidden platform on the MWM task), we calculated the sample sizes and power needed to reject the null hypothesis (of no differences between the CCI+PROG and CCI+vehicle-treated rats) to achieve $>80\%$ power to detect a 30% difference (medium effect). The number of rats per group at these criteria was determined to be 9.

Effect of CCI on lesion size

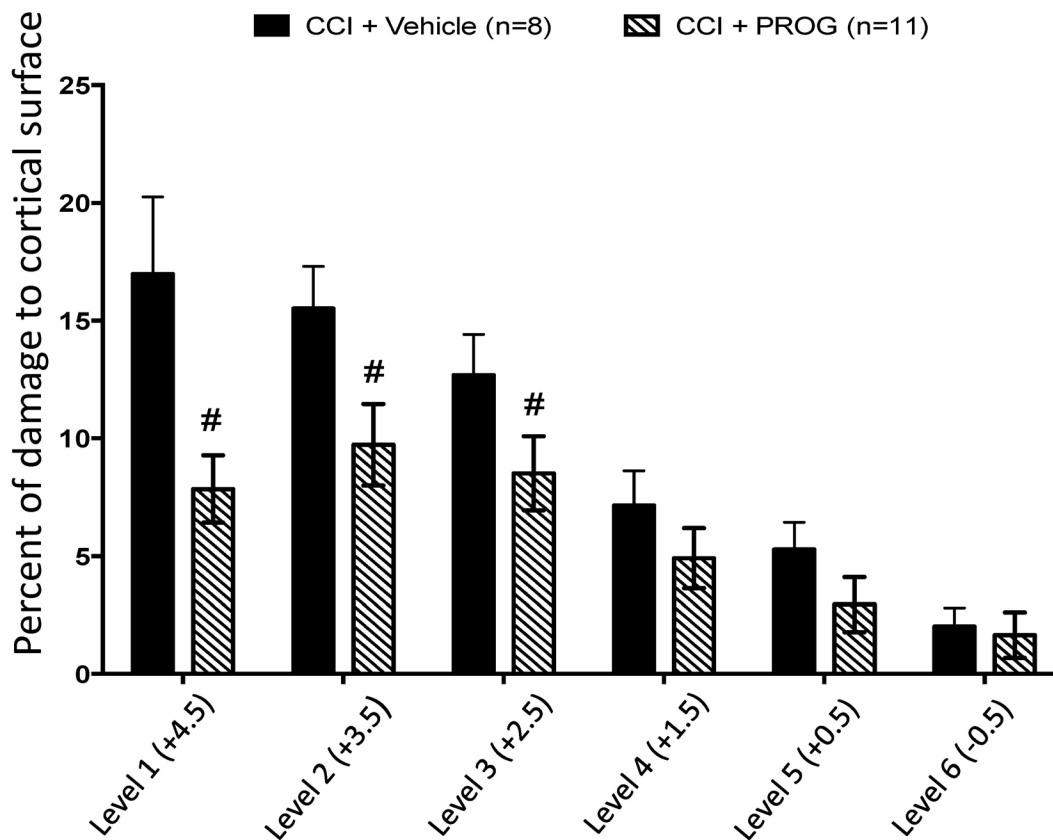


Fig 1. Lesion reconstruction analysis of juvenile female rats with controlled cortical impact (CCI) treated with progesterone (PROG) vs. vehicle. Mean percent (\pm SEM) of volumetric tissue loss at 6 anterior-to-posterior (A/P) levels at 4 weeks post-injury. # = difference between the CCI+vehicle-treated group and CCI rats given 8 mg/kg PROG. Values are mean \pm SEM (n = 6–11 / group).

doi:10.1371/journal.pone.0146419.g001

Results

Lesion (tissue) reconstruction analysis

Briefly, the 6 sections from each rat brain were photographed with an Epson scanner (and ImagePro™ software). Group means of percent of damaged tissue averaged across 6 anterior-to-posterior (A/P) sections were used to determine overall lesion size. One of the sham rats had visible drill-induced injury ($\geq 1\%$) that was unintentional and thus its data was excluded from behavioral statistical analysis. Tissue sections from the remaining sham rats had no observable damage. Results from the lesion reconstruction analysis of CCI-injured animals treated with vehicle or PROG are shown in Fig 1. Compared to intact sham-operated animals, there was a significant lesion effect ($F(3,29) = 27.54, p < 0.01$). *Post-hoc* comparisons further revealed significant differences at A/P brain levels 1 to 3 between PROG and vehicle-treated CCI rats ($\# = p < 0.05$).

Physiological and functional effects of PND 28 CCI-induced injury

Body Weight. One-way ANOVA with repeated measures on PND or PID conducted on the mean pre- and post-injury body weights found no main effect on mean body weight (F

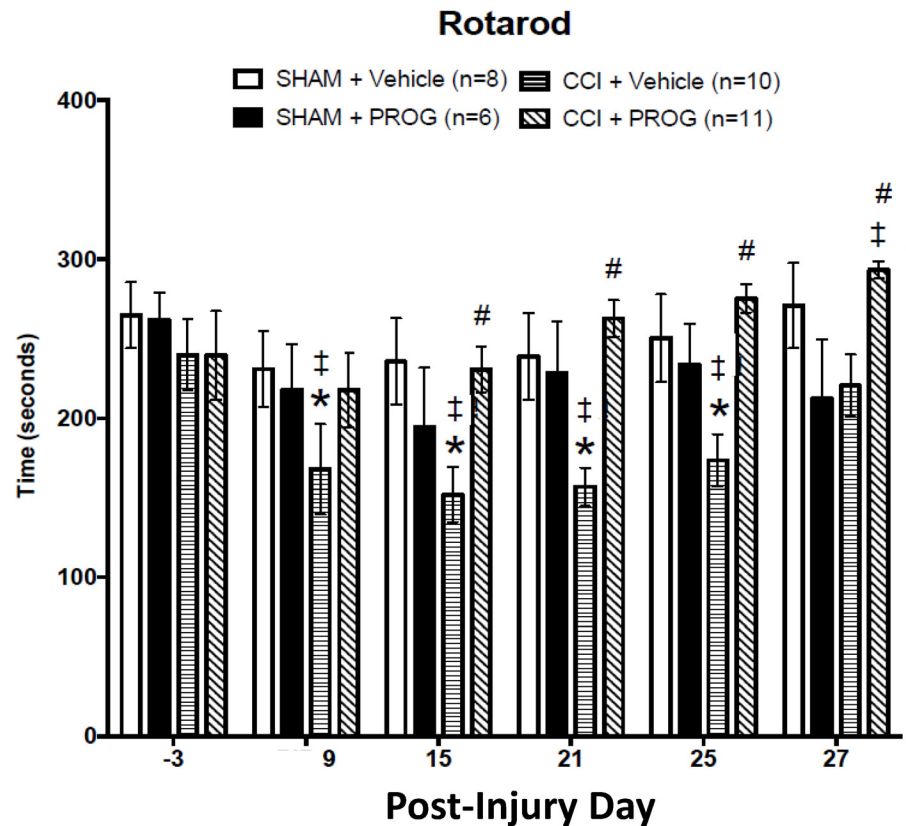


Fig 2. Effect of progesterone (PROG) treatment on vestibulomotor function in female CCI rats treated with PROG vs. vehicle. PROG (8 mg/kg) -treated CCI rats showed improvement in balancing and walking on the rotarod tasks. The Sham+PROG group was given 16-mg/kg doses of PROG. * = different from sham + vehicle; # = different from CCI + vehicle (p 's < 0.05). ‡ = different from baseline within each group. Values are mean \pm SEM ($n = 6-11$).

doi:10.1371/journal.pone.0146419.g002

(3,31) = 0.1351, $p > 0.05$) (S1 Fig). As might be expected, there was, however, a main effect of PID on mean body weight ($F(30,930) = 2265.0$, $p < .001$). *Post-hoc* pair-wise comparisons indicated that the group mean body weights were significantly increased over time ($p < 0.01$), except for the first two days post-surgery ($p > 0.05$).

Elevated Plus Maze. Repeated measures ANOVA were performed on EPM behaviors. The independent measure was Lesion/Treatment and the dependent measures were the percent of entries in the open vs. closed arms. The ANOVA revealed no significant main effect of Lesion/Treatment on the percent of entries in each arm or total crossings (F 's < 1, $p > 0.05$; see S2 Fig). One-way ANOVA revealed that on all 3 test days, all groups preferred the closed arms of the maze (9 to 1).

Rotarod. Activity on the rotarod was measured as latency to fall off the rotating/accelerating rod over a 300-sec period. A repeated 4 x 6 (Lesion/Treatment x Test Days) measures ANOVA, with repeated measure on PID, revealed a main effect of Lesion/Treatment on rotarod performance ($F(5,45) = 3.87$, $p < 0.05$). *Post-hoc* analysis further revealed that rotarod performance during baseline testing was not different among the groups ($p > 0.05$). As indicated in Fig 2, CCI injury and administration of 8 mg/kg of PROG had a significant effect on rotarod performance in the developing female rats. The rats given a CCI performed significantly worse on PID 9, 15, 21 and 25 ($^{\ddagger} = p < 0.05$) compared to their baseline performance.

A MWM Latency (Run 1)

B MWM Latency (Run 2)

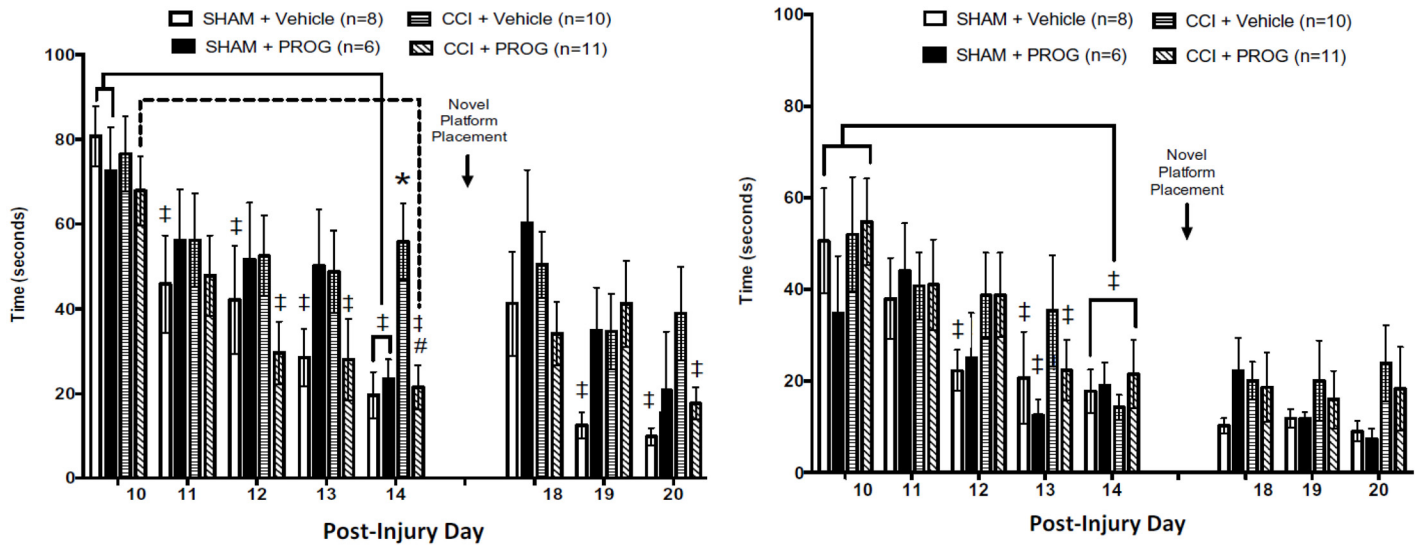


Fig 3. Effects of progesterone (PROG) on learning and memory task as assessed by latency to locate the Morris water maze (MWM) platform. During Run 1 (a) on the acquisition phase there was a significant effect of controlled cortical impact injury. Eight mg/kg PROG proved beneficial by decreasing the mean latency to find a hidden platform in the MWM compared to the CCI+vehicle-treated group. There was no clear effect of PROG treatment on Run 2 (b) or during the reversal phase ($p > 0.05$). The Sham+PROG group was given 16 mg/kg doses of PROG. * = different from Sham+Vehicle; # = different from CCI+Vehicle (p 's < 0.05). ‡ = different from baseline within each group; $n = 6-11$.

doi:10.1371/journal.pone.0146419.g003

Rotarod performance in the CCI+PROG group, in contrast, did not differ from baseline on these days and was significantly improved by PID 27 ($p < 0.05$). As also indicated in Fig 2, rotarod performance in the CCI+vehicle group was worse than in the sham+vehicle-treated rats on PID 9, 15, 21 and 25 ($p < 0.05$). The CCI+PROG group was not different from sham+vehicle and had significantly better rotarod scores than CCI+vehicle on PID 15, 21, 25 and 27 ($p < 0.05$). Finally, while the 8 mg/kg dose of PROG reduced performance deficits in the CCI-injured rats, the 16 mg/kg dose noticeably increased the variability in performance in the sham group.

Spatial navigation and cognition in the MWM. Mean latency to reach the MWM platform within 90 sec during acquisition and novel platform placement learning served as the dependent measures. Group latency scores from Runs 1 and 2 are shown in Fig 3a and 3b and were analyzed using a 4 x 2 x 9 (Lesion/Treatment x Run x Test Day) mixed factorial ANOVA, with repeated measures on PID.

As discussed below, on the first MWM trial the female rat pups were slow compared to their male counterparts. Among the females, there was a significant main effect of Lesion/Treatment ($F(3,31) = 5.94, p < 0.01$) and Test Day ($F(7,217) = 11.82, p < 0.01$) in finding the submerged MWM platform. As shown in Fig 3a, during Run 1 on the second and third days of acquisition training, the vehicle-treated and CCI+PROG-treated groups did significantly better than on the first trial. On the last day of acquisition training (PID 14), before the platform was moved to a novel location, all rats except those in the CCI+vehicle-treated group performed better than they did on PID 10 ($^{\ddagger} = p < 0.05$). On PID 14 the latency to reach the platform during Run 1 was significantly delayed in the CCI+vehicle group compared to the CCI+PROG ($\# = p < 0.05$) or sham groups ($* = p < 0.05$).

Post-hoc Tukey's multiple comparison tests revealed (1) a significant difference between CCI and sham rats treated with vehicle ($p < 0.05$) and (2) a significant difference between PROG- and vehicle-treated CCI rats ($p < 0.05$), but (3) the differences between the CCI +PROG and the sham groups were not significant (p 's > 0.05). Finally, as shown in Fig 3a, placing the MWM platform in a novel quadrant was transiently disruptive to the performance pattern observed in sham-injured rats at the end of acquisition (PID 14).

As shown in Fig 3b, neither sham-injured nor CCI female rats appeared to use the Run 1 experience to find the MWM platform more efficiently in Run 2 (which was conducted 5 min after Run 1). Overall, CCI had no effect on acquisition learning and performance on the three novel platform placement trials. Taken together, our data show that the CCI injury was sufficient to produce observable, histopathological, specific behavioral and cognitive deficits in female PND 28 rat pups.

Discussion

Sex differences in 1-month-old rat pups with CCI injury

Despite the physiological and cognitive differences between healthy young males and females, there is no consensus on whether human females with a TBI have better functional outcomes than males [32–36]. It is also not clear whether the levels of female hormones like estrogen (or estradiol) and PROG, which increases exponentially during pregnancy, play a role in sex-specific neuroprotection or neurodegeneration following a TBI [37–45]. Interestingly, Wagner et al. [46] found no sex differences in the acute elevation of serum sex hormone levels in adult patients with a severe TBI but noted that elevated levels of testosterone in women and estradiol in men were associated with a poor prognosis and increased mortality at 6 and 12 months after injury. Less is known about whether such outcomes would be observed in younger subjects, so gathering similar data in juveniles could be important in planning for clinical investigation of neurosteroid treatment for male and female juvenile TBI patients [47]. This is not a trivial issue. Two independent studies collectively analyzing 56,994 juvenile patients with moderate-to-severe brain trauma found that adolescent, but not prepubescent, brain-injured girls had a lower incidence of mortality than age-matched brain-injured boys [48,49]. These data were interpreted to suggest that an “active” (and developing) reproductive system affords juvenile females a sex-specific advantage during a TBI event and recovery.

A growing number of preclinical studies show that the effectiveness of post-injury PROG treatment for brain trauma in the developing rat may be affected by age, gender and injury model [50–55]. Mannix et al. [56] recently found that PROG treatment improved grip strength in CCI-injured 8-week-old male mice but worsened performance in females, while the MWM spatial learning deficit was unaffected by PROG treatment in either sex. As noted in our introduction, two recently reported failures in clinical trials of PROG for the treatment of TBI in adults found no benefits of PROG treatment on very blunt and simplified quality of life outcome measures, but potential sex differences in biomarkers and other neuropsychological or lesion parameters have not yet been evaluated [17,18], and there were issues of dose optimization and duration of treatment that limit more sophisticated interpretation of the poor results [19,20].

The disappointing trial results make it more important to examine the varying efficacy of PROG treatment by sex, age, dosing and duration of treatment as well as to identify the most appropriate dosing and comparable outcome measures [19,20]. Here we compared pre-estrous, prepubescent PND28 female SD rats from the current investigation and PND28 male SD rats from our previous publication [26]. Fig 4 summarizes the comparative analysis of the male and

	Post-Injury Day (PID)	Male Sham + vehicle (n=8)	Female Sham + vehicle (n=8)	Male Sham + PROG-16 (n=8)	Female Sham + PROG-16 (n=6)	Male CCI + vehicle (n=9)	Female CCI + vehicle (n=10)	Male CCI + PROG-8 (n=8)	Female CCI + PROG-8 (n=11)
BODY WEIGHT (grams)	0	80.13 ± 2.00	77.13 ± 1.59	82.63 ± 1.19	78.67 ± 2.36	83.33 ± 2.36	78.5 ± 1.78	80.33 ± 2.30	77.64 ± 0.79
	28	258.70 ± 3.71	207.38 ± 5.62	263.63 ± 4.88	210.83 ± 4.36	261.67 ± 4.79	207.2 ± 4.44	255.22 ± 4.53	203.36 ± 4.22
LESION VOLUME (% of whole hemisphere)	28	-	-	-	-	22.99 ± 3.11	9.94 ± 2.83	15.41 ± 3.68*	5.94 ± 3.64
ROTAROD (seconds)	-3	240.38 ± 12.42	264.88 ± 20.94	232.63 ± 12.87	261.83 ± 17.27	243.11 ± 15.08	240.05 ± 22.50	245.11 ± 11.49	239.77 ± 27.71
	9	205.50 ± 23.52	231.06 ± 23.70	204.00 ± 29.74	218.00 ± 28.85	113.00 ± 24.28	168.15 ± 28.40	202.67 ± 26.63*	218.05 ± 23.56
	15	264.88 ± 22.25	235.88 ± 27.18	236.00 ± 23.62	194.33 ± 37.71	151.56 ± 19.84	152.10 ± 17.21	211.56 ± 29.81*	230.86 ± 14.24*
	21	253.38 ± 20.09	238.88 ± 27.29	231.88 ± 25.10	228.67 ± 32.06	141.67 ± 15.16	156.95 ± 12.06	224.22 ± 22.98*	262.82 ± 11.55*
	25	260.50 ± 19.93	250.50 ± 27.52	263.13 ± 25.51	234.00 ± 25.41	154.11 ± 21.08	173.60 ± 16.48	222.78 ± 22.94*	275.64 ± 9.08*
	27	251.88 ± 22.33	271.00 ± 26.79	229.50 ± 28.06	212.67 ± 36.72	153.89 ± 14.18	220.65 ± 19.67	219.44 ± 16.76*	293.36 ± 5.31*
MWM Acquisition LATENCY 2 PLATFORM (seconds)	10 (Run 1)	60.38 ± 10.32	80.82 ± 7.12	45.38 ± 13.60	72.51 ± 10.40	71.22 ± 8.92	76.68 ± 8.95	71.22 ± 9.86	67.93 ± 8.22
	10 (Run 2)	52.32 ± 14.37	50.56 ± 11.47	33.25 ± 9.01	34.76 ± 12.58	66.11 ± 11.27	52.01 ± 12.47	73.80 ± 8.91	54.73 ± 9.45
	14 (Run 1)	15.50 ± 2.26	19.66 ± 5.50	14.00 ± 3.41	23.43 ± 4.68	45.33 ± 14.17	55.92 ± 8.99	19.00 ± 5.71*	21.50 ± 5.23*
	14 (Run 2)	14.24 ± 6.14	17.78 ± 4.75	13.91 ± 3.87	18.97 ± 5.13	21.36 ± 4.36	14.25 ± 2.73	22.05 ± 9.40	21.53 ± 7.40
EPM (% visits)	-2 (open)	15.38 ± 4.52	12.68 ± 3.29	21.56 ± 3.59	10.52 ± 3.06	19.33 ± 3.63	7.99 ± 1.79	25.11 ± 4.56	9.76 ± 3.12
	-2 (closed)	84.63 ± 4.52	87.32 ± 3.29	78.44 ± 3.59	89.48 ± 3.06	80.67 ± 3.63	92.01 ± 1.79	70.89 ± 4.56	90.24 ± 3.12
	9 (open)	14.44 ± 1.50	3.95 ± 2.25	10.00 ± 2.23	4.96 ± 4.08	12.44 ± 1.56	10.43 ± 3.93	15.72 ± 3.12	7.77 ± 2.77
	9 (closed)	85.56 ± 1.50	96.05 ± 2.25	90.00 ± 2.27	95.04 ± 4.08	87.56 ± 1.56	89.57 ± 3.93	84.28 ± 3.12	92.24 ± 2.77
	27 (open)	20.44 ± 2.17	16.22 ± 4.43	22.00 ± 4.62	10.24 ± 1.57	9.44 ± 3.72	13.27 ± 4.44	14.94 ± 3.06	12.23 ± 1.68
	27 (closed)	79.56 ± 2.17	84.24 ± 4.43	78.00 ± 4.62	89.76 ± 1.57	90.56 ± 3.72	86.73 ± 4.44	85.06 ± 3.06	87.77 ± 1.68
OPEN FIELD Distance travelled (centimeters)	-1	1729.75 ± 104.71	1548.00 ± 116.83	1472.75 ± 78.78	1505.50 ± 154.69	1618.45 ± 82.91	1726.11 ± 108.08	1697.22 ± 91.12	1799.64 ± 78.40
	9	1978.50 ± 217.93	1891.62 ± 116.72	1688.75 ± 161.35	2249.33 ± 168.34	2238.67 ± 238.83	2556.22 ± 165.87	2341.00 ± 109.83	2427.55 ± 133.57
	15	1784.38 ± 100.50	2181.38 ± 123.83	1732.25 ± 103.37	2406.00 ± 303.66	1923.89 ± 134.18	2197.22 ± 122.82	1980.00 ± 94.19	2280.46 ± 181.94
	21	1907.13 ± 126.59	1950.25 ± 180.16	1705.88 ± 124.79	2115.33 ± 136.53	1781.56 ± 156.19	2126.78 ± 203.61	1958.33 ± 93.71	2153.18 ± 91.45
	25	1868.25 ± 89.24	1819.50 ± 112.94	1519.88 ± 157.40	2176.67 ± 256.84	1815.11 ± 104.98	2236.33 ± 307.31	2019.67 ± 134.96	2215.82 ± 144.32
	27	1815.25 ± 61.05	1595.75 ± 96.46	1589.38 ± 85.36	1794.83 ± 146.67	1884.56 ± 156.40	2236.33 ± 293.00	1986.67 ± 107.32	2215.82 ± 138.55

Fig 4. Tabulated comparative deficits between male and female controlled cortical impact injury. Bold numbers with asterisk (*) indicate differences between treatment groups within the same gender, $p < .05$. Darker-shaded cells indicate where male rats either found the hidden MWM platform faster or made fewer visits to the closed arm of the EPM than similarly treated female rat pups (between-gender analysis), $p < .05$. Lighter-shaded cells indicate where female rat pups either weighed less, had smaller lesions, spent less time

in the open arm of the EPM, tended to travel longer distances (Open Field), remained on the rotarod longer, or found the MWM platform faster than similarly treated male counterparts (b/w gender analysis), $p < .05$; PID 0 = day of injury but prior to surgery; PID -1 = day(s) before surgery; MWM: Morris Water Maze; CCI: controlled cortical impact; Mean \pm SEM; $n = 6-11$. Male data (columns 3, 5, 7, and 9) were previously published [26].

doi:10.1371/journal.pone.0146419.g004

female CCI rat pup data where the specific location of impact and impactor size, velocity, and impactor contact times were identical.

Sex differences and body weight: Effects of CCI injury and PROG treatment

It has long been known that in peri-adolescent humans, around age 14, a gender-based divergence in mean body weight arises [57]. This difference intensifies and becomes significant during puberty and into young adulthood. In the present study, by PID 28, the sham-injured/vehicle-treated females weighed approximately 50 grams less (Fig 4, row 2, lighter-shaded cells) than male counterparts. These data are not surprising, but it is important to note that whatever sexually dimorphic developmental factors are leading to differences in mean body weight in the maturing human, these differences are preserved in our growing, vehicle-treated control rat pups despite sham surgery. These weight and body size sex differences could affect how a similar dose of a lipophilic hormone like PROG could be metabolized and transported to the brain. These considerations could be especially important in planning for clinical dose optimization with any therapeutic agent and would certainly be important in considering the use of a key sex hormone in developing females. Body weight and mass could very likely affect drug metabolism and other pharmacokinetic factors affecting the response to treatment [19].

Neither CCI injury nor PROG treatment produced any noticeable changes in mean body weight in male or female rat pups, although rats in the Sham+PROG groups weighed slightly more, and CCI+PROG rats weighed slightly less, than their vehicle-treated counterparts (Fig 4, row 1, column 1). Further studies are warranted to determine whether there are long-term differences in mean body weight between sexes as a result of treatment with PROG, or any other neuroprotective drug, for that matter.

Lesion extent/severity: Effect of PROG treatment

An early study on sex differences and brain injury reported lower post-injury cerebral edema in pseudopregnant female SD rats compared to normally cycling females and males [58–61]. In the present study, lesion reconstruction data indicate significant differences in TBI neuropathology between sexes in the developing rodent. As shown in Fig 4, row 3, compared to males, CCI surgery clearly produced less physical damage to brain tissue (% total hemispheric tissue loss) of juvenile female rats. Previously, in males we observed a significant effect of PROG on mean percent of volumetric tissue loss at all six A/P levels examined, but in females we observed (Fig 1) a significant effect of PROG only at the three anterior levels (+4.5, +3.5, +2.5). Robertson and Saraswathihere [54] were recently surprised to find that their vehicle-treated juvenile SD female CCI rats tended to have smaller lesions compared to male counterparts, but, interestingly, Peterson et al. reported the opposite effect, with males exhibiting far more neuronal sparing after neonatal hypoxic injury [22]. These contradictory data may be the result of the effects of critical periods in development on injury outcome. The inherent differences in response to acquired brain trauma may have a profound effect on treatment effectiveness and delayed functional and morphological sparing [62]. For instance, the PROG treatment that reduced lesion volume by ~33% in SD male rat pups (from 22.99% \pm 3.11 to 15.41% \pm 3.68)

reduced lesion volume by ~40% (from $9.94\% \pm 2.83$ to $5.94\% \pm 3.64$) in age-matched female rats. In this case, owing to the over two-fold difference in the lesion volume compared to age-matched male rats, the injury was too small to see any beneficial treatment effect of PROG on overall lesion volume in the female rats, so the differences between the treated and untreated females was not statistically significant. Given these factors, the potential sex-specific difference in lesion susceptibility (higher in males) and treatment efficacy (lower in females) imply a need for caution when interpreting relative lesion severity and designing injury models, and especially when comparing treatment efficacy across sexes in developing humans.

Rotarod: Effects of CCI injury and PROG treatment

The rotarod tests motor coordination and vestibular balance [63]. Acute CCI injuries to the unilateral parietal [64] and bilateral mPFC [26] have been independently shown to disrupt rotarod performance in PND17 and PND28 male rats, respectively. In the present study, rotarod performance of sham-injured rat groups did not differ by sex or treatment when tested before 1 month of age (Fig 4, rows 4–9, columns 2–5). In contrast to our findings, Wagner et al. [65] reported that vehicle-treated SD female rats with diffuse axonal injury performed better on the rotarod than vehicle-treated sham-injured males. Here, in contrast to its effect on the sham-injured rats, CCI surgery produced a slightly less sustained deficit on rotarod performance in juvenile females on PID 9 and 27 and PROG treatment was clearly more beneficial in female rats on PID 21, 25, 27 compared to their male counterparts evaluated in our previous study [26] (Fig 4, rows 4–9, columns 6–9). It is possible that exogenous and endogenous PROG levels during puberty synergistically interact to combat a TBI sustained prior to adolescence. Only longer-term tests and gender comparisons from pre- to post-puberty will begin to address this issue.

Activity level: Effects of CCI injury and PROG treatment

Open Field and EPM tests are used to study hyperactivity and exploratory and anxiety-like behavior rodents [66,67]. Parker and Morinan [68] demonstrated that rodent behavior in the EPM is primarily driven by anxiety-related behaviors in juvenile and adult rats of both sexes. We have previously demonstrated that both EPM and open field results are affected by CCI injury and damage to the mPFC [26,28,31]. In contrast to adult rats, immature rats with moderate brain injury displayed relatively minor changes in activity level compared to controls. We found that female rat pups sporadically spent more time in the open arm of the EPM than males, and PROG-treated sham-injured females tended to be more active than sham-injured males given PROG. Future studies will confirm or discount the strength of these trends.

Spatial learning: Effects of CCI injury and PROG treatment

The rodent MWM was designed to use spatial learning in rodents as a measure of cognitive, spatial and memory performance [69]. More recently, gender differences have been reported using virtual MWM tasks in young adults (undergraduate students) and prepubescent children [70,71]. While both studies found that males were better at visual tasks, higher levels of anxiety during spatial tasks have been reported in young adult human females [72], and shown to selectively affect hippocampus-dependent learning in children [73]. This spatial anxiety may retard spatial navigation as a function of sex or age, suggesting an increased vulnerability in the developing females.

There were no observed differences among the male and female rats in MWM performance on the first (acquisition) trial (Fig 4 row 10). Similar to the rotarod results, sham surgery and PROG treatment in the intact animals showed no evidence of sex differences in acquiring the

task. Others have reported that CCI injury did not affect reversal learning in male or female SD rat pups after excitotoxic lesions of the mPFC [74]. In our study, CCI or PROG treatment had minimal effects on short-term memory, but PROG treatment improved performance on long-term memory tasks by the fifth exposure (on PID 14) equally in female and male CCI rat pups (Fig 4, row 10). This is important because, while the majority of juvenile brain injury studies using hormonal treatment have had encouraging results, longer-term studies have indicated that sustained elevation in neonatal PROG significantly affects sexual behaviors in adulthood in males but not in females [75]. Looking forward, perhaps particularly in females, determining the time when hormonal treatment is appropriate to administer (and during what stage of estrus) following a pediatric TBI could be very important.

Conclusions

Our data demonstrate that the same injury parameters used to produce a moderate TBI in pre-pubescent (PND 28) male SD rat pups [26] were sufficient to produce histopathological and functional deficits in female conspecifics. Sex differences were observed in the severity of CCI injury as well as in the effectiveness of post-injury PROG treatment on some, but not all, functional performance outcomes. For neuroprotective strategy/studies we think that it will be important to take into account the TBI severity, sex and hormonal status of the patient at the time treatment is delivered. Preclinical juvenile rodent brain injury data to date suggest that the effects of PROG treatment post-TBI may vary not only by (a) age at time of injury and (b) drug dose/duration, but also by (c) injury type (e.g., diffused, focal, or penetrating brain injury, blast trauma, hypoxia-ischemia, reperfusion injury) and (d) gender [52,56,76–79]. In particular, a given brain injury could have far more deleterious effects in the early stages of life than if the same damage occurs later in development [80,81]. We suggest that if clinical research is to advance, future studies will have to document TBI progression and the effects of hormonal treatment throughout puberty across genders not only on TBI-induced deficits, but also on normal milestones in maturation like puberty onset and gender-specific sexual-social behavior.

Supporting Information

S1 Fig. Dose-response effect of progesterone on weight. Mean body weight (gm) showing weight changes in each group. No significant differences in body weight between groups ($p > 0.05$) were observed. Values are mean \pm SEM ($n = 6-11$ / group). PROG = progesterone. (TIFF)

S2 Fig. Effect of progesterone on elevated plus maze (EPM) activity. Percent of visits to pair of open vs. closed arms. There were no significant differences between groups in the % of visits to the open and closed arms of the EPM maze either pre- or post-surgery (p 's > 0.05). * = differences in the arm visited (open vs. closed). Values are mean \pm SEM ($n = 6-11$ / group). PROG = progesterone. (TIFF)

Acknowledgments

This work was supported by unrestricted gifts in support of research from BHR Pharma, Allen and Company, the Emory Center for Injury Control, National Institutes of Health grant no. R49CE001494-04 to DGS, and the Emory University Summer Undergraduate Research Experience (SURE) program. The authors would like to acknowledge the contributions of our undergraduate research trainees (William Godwin, Maydda Qureshi, Leea Richardson, and Noah Rosen) towards the completion of this work.

Author Contributions

Conceived and designed the experiments: RIG IS DGS. Performed the experiments: RIG BLP. Analyzed the data: RIG IS DGS. Wrote the paper: RIG DGS IS BLP.

References

1. Coronado VG, McGuire LC, Sarmiento K, Bell J, Lionbarger MR, Jones CD, et al. Trends in Traumatic Brain Injury in the U.S. and the public health response: 1995–2009. *J Safety Res.* 2012; 43: 299–307. doi: [10.1016/j.jsr.2012.08.011](https://doi.org/10.1016/j.jsr.2012.08.011) PMID: [23127680](https://pubmed.ncbi.nlm.nih.gov/23127680/)
2. Centers for Disease Control. Vital signs: avoidable deaths from heart disease, stroke, and hypertensive disease—United States, 2001–2010. *MMWR Morbidity and mortality weekly report.* 2013; 62: 721–727. PMID: [24005227](https://pubmed.ncbi.nlm.nih.gov/24005227/)
3. Loane DJ, Faden AI. Neuroprotection for traumatic brain injury: translational challenges and emerging therapeutic strategies. *Trends Pharmacol Sci.* 2010; 31: 596–604. doi: [10.1016/j.tips.2010.09.005](https://doi.org/10.1016/j.tips.2010.09.005) PMID: [21035878](https://pubmed.ncbi.nlm.nih.gov/21035878/)
4. Babikian T, Asarnow R. Neurocognitive outcomes and recovery after pediatric TBI: meta-analytic review of the literature. *Neuropsychology.* 2009; 23: 283–296. doi: [10.1037/a0015268](https://doi.org/10.1037/a0015268) PMID: [19413443](https://pubmed.ncbi.nlm.nih.gov/19413443/)
5. Farrer TJ, Frost RB, Hedges DW. Prevalence of traumatic brain injury in juvenile offenders: a meta-analysis. *Child Neuropsychol.* 2013; 19: 225–234. doi: [10.1080/09297049.2011.647901](https://doi.org/10.1080/09297049.2011.647901) PMID: [22372420](https://pubmed.ncbi.nlm.nih.gov/22372420/)
6. Perron BE, Howard MO. Prevalence and correlates of traumatic brain injury among delinquent youths. *Crim Behav Ment Health.* 2008; 18: 243–255. doi: [10.1002/cbm.702](https://doi.org/10.1002/cbm.702) PMID: [18803295](https://pubmed.ncbi.nlm.nih.gov/18803295/)
7. Williams WH, Cordan G, Mewse AJ, Tonks J, Burgess CN. Self-reported traumatic brain injury in male young offenders: a risk factor for re-offending, poor mental health and violence? *Neuropsychol Rehabil.* 2010; 20: 801–812. doi: [10.1080/09602011.2010.519613](https://doi.org/10.1080/09602011.2010.519613) PMID: [21069616](https://pubmed.ncbi.nlm.nih.gov/21069616/)
8. Kaore SN, Langade DK, Yadav VK, Sharma P, Thawani VR, Sharma R. Novel actions of progesterone: what we know today and what will be the scenario in the future? *J Pharm Pharmacol.* 2012; 64: 1040–1062. doi: [10.1111/j.2042-7158.2012.01464.x](https://doi.org/10.1111/j.2042-7158.2012.01464.x) PMID: [22775208](https://pubmed.ncbi.nlm.nih.gov/22775208/)
9. Luoma JI, Stern CM, Mermelstein PG. Progesterone inhibition of neuronal calcium signaling underlies aspects of progesterone-mediated neuroprotection. *J Steroid Biochem Mol Biol.* 2012; 131: 30–36. doi: [10.1016/j.jsbmb.2011.11.002](https://doi.org/10.1016/j.jsbmb.2011.11.002) PMID: [22101209](https://pubmed.ncbi.nlm.nih.gov/22101209/)
10. Stein DG. Is progesterone a worthy candidate as a novel therapy for traumatic brain injury? *Dialogues Clin Neurosci.* 2011; 13: 352–359. PMID: [22033509](https://pubmed.ncbi.nlm.nih.gov/22033509/)
11. Coughlan T, Gibson C, Murphy S. Progesterone, BDNF and neuroprotection in the injured CNS. *Int J Neurosci.* 2009; 119: 1718–1740. PMID: [19922383](https://pubmed.ncbi.nlm.nih.gov/19922383/)
12. Cai W, Zhu Y, Furuya K, Li Z, Sokabe M, Chen L. Two different molecular mechanisms underlying progesterone neuroprotection against ischemic brain damage. *Neuropharmacology.* 2008; 55: 127–138. doi: [10.1016/j.neuropharm.2008.04.023](https://doi.org/10.1016/j.neuropharm.2008.04.023) PMID: [18572204](https://pubmed.ncbi.nlm.nih.gov/18572204/)
13. Singh M. Mechanisms of progesterone-induced neuroprotection. *Ann N Y Acad Sci.* 2005; 1052: 145–151. PMID: [16024757](https://pubmed.ncbi.nlm.nih.gov/16024757/)
14. Gonzalez Deniselle MC, Lopez Costa JJ, Gonzalez SL, Labombarda F, Garay L, Guennoun R, et al. Basis of progesterone protection in spinal cord neurodegeneration. *J Steroid Biochem Mol Biol.* 2002; 83: 199–209. PMID: [12650717](https://pubmed.ncbi.nlm.nih.gov/12650717/)
15. Stein DG, Wright DW. Progesterone in the clinical treatment of acute traumatic brain injury. *Expert Opin Investig Drugs.* 2010; 19: 847–857. doi: [10.1517/13543784.2010.489549](https://doi.org/10.1517/13543784.2010.489549) PMID: [20486864](https://pubmed.ncbi.nlm.nih.gov/20486864/)
16. Li LM, Menon DK, Janowitz T. Cross-sectional analysis of data from the U.S. clinical trials database reveals poor translational clinical trial effort for traumatic brain injury, compared with stroke. *PLoS One.* 2014; 9: e84336. doi: [10.1371/journal.pone.0084336](https://doi.org/10.1371/journal.pone.0084336) PMID: [24416218](https://pubmed.ncbi.nlm.nih.gov/24416218/)
17. Wright DW, Yeatts SD, Silbergleit R, Palesch YY, Hertzberg VS, Frankel M, et al. Very early administration of progesterone for acute traumatic brain injury. *N Engl J Med.* 2014; 371: 2457–2466. doi: [10.1056/NEJMoa1404304](https://doi.org/10.1056/NEJMoa1404304) PMID: [25493974](https://pubmed.ncbi.nlm.nih.gov/25493974/)
18. Skolnick BE, Maas AI, Narayan RK, van der Hoop RG, MacAllister T, Ward JD, et al. A Clinical Trial of Progesterone for Severe Traumatic Brain Injury. *N Engl J Med.* 2014; 371: 2467–2476. doi: [10.1056/NEJMoa1411090](https://doi.org/10.1056/NEJMoa1411090) PMID: [25493978](https://pubmed.ncbi.nlm.nih.gov/25493978/)
19. Howard RB, Sayeed I, Stein D. Suboptimal dosing parameters as possible factors in the negative Phase III clinical trials of progesterone in TBI. *J Neurotrauma.* 2015. Sep 15. [Epub ahead of print]doi: [10.1089/neu.2015.4179](https://doi.org/10.1089/neu.2015.4179)

20. Stein DG. Embracing failure: What the Phase III progesterone studies can teach about TBI clinical trials. *Brain Injury*. 2015; 29: 1259–1272. PMID: [26274493](#)
21. Schumacher M, Denier C, Oudinet J-P, Adams DH, Guennoun R. Progesterone neuroprotection: The background of clinical trial failure. *J Steroid Biochem Mol Biol*. 2015 Nov 17. pii: S0960-0760(15)30135-7. doi: [10.1016/j.jsbmb.2015.11.010](#) [Epub ahead of print]
22. Peterson BL, Won S, Geddes RI, Sayeed I, Stein DG. Sex-related differences in effects of progesterone following neonatal hypoxic brain injury. *Behav Brain Res*. 2015; 286: 152–165. doi: [10.1016/j.bbr.2015.03.005](#) PMID: [25746450](#)
23. Tucker LB, Fu AH, McCabe JT. Performance of male and female C57BL/6J Mice on motor and cognitive tasks commonly used in pre-clinical Traumatic Brain Injury research. *J Neurotrauma*. 2015 Aug 12. [Epub ahead of print] doi: [10.1089/neu.2015.3977](#)
24. Wong R, Renton C, Gibson CL, Murphy SJ, Kendall DA, Bath PM, et al. Progesterone treatment for experimental stroke: an individual animal meta-analysis. *J Cereb Blood Flow Metab*. 2013; 33: 1362–1372. doi: [10.1038/jcbfm.2013.120](#) PMID: [23838830](#)
25. Robertson CL, Puskar A, Hoffman GE, Murphy AZ, Saraswati M, Fiskum G. Physiologic progesterone reduces mitochondrial dysfunction and hippocampal cell loss after traumatic brain injury in female rats. *Exp Neurol*. 2006; 197: 235–243. PMID: [16259981](#)
26. Geddes RI, Sribnick EA, Sayeed I, Stein DG. Progesterone treatment shows benefit in a pediatric model of moderate to severe bilateral brain injury. *PLoS One*. 2014; 9: e87252. doi: [10.1371/journal.pone.0087252](#) PMID: [24489882](#)
27. Sengupta P. The laboratory rat: relating its age with human's. *Int J Prev Med*. 2013; 4: 624–630. PMID: [23930179](#)
28. Goss CW, Hoffman SW, Stein DG. Behavioral effects and anatomic correlates after brain injury: a progesterone dose-response study. *Pharmacol Biochem Behav*. 2003; 76: 231–242. PMID: [14592674](#)
29. Russell KL, Kutcho KM, Fowler SC, Berman NE, Levant B. Sensorimotor behavioral tests for use in a juvenile rat model of traumatic brain injury: assessment of sex differences. *J Neurosci Methods*. 2011; 199: 214–222. doi: [10.1016/j.jneumeth.2011.05.008](#) PMID: [21600923](#)
30. Cutler SM, Pettus EH, Hoffman SW, Stein DG. Tapered progesterone withdrawal enhances behavioral and molecular recovery after traumatic brain injury. *Exp Neurol*. 2005; 195: 423–429. PMID: [16039652](#)
31. Cutler SM, Vanlandingham JW, Stein DG. Tapered progesterone withdrawal promotes long-term recovery following brain trauma. *Exp Neurol*. 2006; 200: 378–385. PMID: [16797538](#)
32. Farace E, Alves WM. Do women fare worse: a metaanalysis of gender differences in traumatic brain injury outcome. *J Neurosurg*. 2000; 93: 539–545. PMID: [11014529](#)
33. Morrison WE, Arbelaez JJ, Fackler JC, De Maio A, Paidas CN. Gender and age effects on outcome after pediatric traumatic brain injury. *Pediatr Crit Care Med*. 2004; 5: 145–151. PMID: [14987344](#)
34. Wright DW, Kellermann AL, Hertzberg VS, Clark PL, Frankel M, Goldstein FC, et al. ProTECT: A randomized clinical trial of progesterone for acute Traumatic Brain Injury. *Ann Emerg Med*. 2007; 49 391–402 PMID: [17011666](#)
35. Ratcliff JJ, Greenspan AI, Goldstein FC, Stringer AY, Bushnik T, Hammond FM, et al. Gender and traumatic brain injury: do the sexes fare differently? *Brain Inj*. 2007; 21: 1023–1030. PMID: [17891564](#)
36. Gibson CL, Gray LJ, Bath PM, Murphy SP. Progesterone for the treatment of experimental brain injury; a systematic review. *Brain*. 2008; 131: 318–328. PMID: [17715141](#)
37. Davis DP, Douglas DJ, Smith W, Sise MJ, Vilke GM, Holbrook TL, et al. Traumatic brain injury outcomes in pre- and post-menopausal females versus age-matched males. *J Neurotrauma*. 2006; 23: 140–148. PMID: [16503798](#)
38. Stein DG, Hoffman SW. Estrogen and progesterone as neuroprotective agents in the treatment of acute brain injuries. *Pediatr Rehabil*. 2003; 6: 13–22. PMID: [12745891](#)
39. Roof RL, Hall ED. Gender differences in acute CNS trauma and stroke: neuroprotective effects of estrogen and progesterone. *J Neurotrauma*. 2000; 17: 367–388. PMID: [10833057](#)
40. Roof RL, Hall ED. Estrogen-related gender difference in survival rate and cortical blood flow after impact-acceleration head injury in rats. *J Neurotrauma*. 2000; 17: 1155–1169. PMID: [11186229](#)
41. Bazarian JJ, Blyth B, Mookerjee S, He H, McDermott MP. Sex differences in outcome after mild traumatic brain injury. *J Neurotrauma*. 2010; 27: 527–539. doi: [10.1089/neu.2009.1068](#) PMID: [19938945](#)
42. Yeung JH, Mikocka-Walus AA, Cameron PA, Poon WS, Ho HF, Chang A, et al. Protection from traumatic brain injury in hormonally active women vs men of a similar age: a retrospective international study. *Arch Surg*. 2011; 146: 436–442. doi: [10.1001/archsurg.2011.46](#) PMID: [21502452](#)
43. Ma J, Huang S, Qin S, You C. Progesterone for acute traumatic brain injury. *Cochrane Database Syst Rev*. 2012; 10: CD008409. doi: [10.1002/14651858.CD008409.pub3](#) PMID: [23076947](#)

44. Wright DW, Stein DG, Sayeed I, Hua F, Atif F, Yousuf S, et al. Response to: Do pregnant women have improved outcomes after traumatic brain injury? *Am J Surg.* 2012; 204: 803–804. doi: [10.1016/j.amjsurg.2011.05.002](https://doi.org/10.1016/j.amjsurg.2011.05.002) PMID: [21880296](https://pubmed.ncbi.nlm.nih.gov/21880296/)
45. Berry C, Mirocha J, Salim A. Response to: Do pregnant women have improved outcomes after traumatic brain injury. *Am J Surg.* 2012; 204: 558–560.
46. Wagner AK, McCullough EH, Niyonkuru C, Ozawa H, Loucks TL, Dobos JA, et al. Acute serum hormone levels: characterization and prognosis after severe traumatic brain injury. *J Neurotrauma.* 2011; 28: 871–888. doi: [10.1089/neu.2010.1586](https://doi.org/10.1089/neu.2010.1586) PMID: [21488721](https://pubmed.ncbi.nlm.nih.gov/21488721/)
47. Hembree WC, Cohen-Kettenis P, Delemarre-van de Waal HA, Gooren LJ, Meyer WJ 3rd, Spack NP, et al. Endocrine treatment of transsexual persons: an Endocrine Society clinical practice guideline. *J Clin Endocrinol Metab.* 2009; 94: 3132–3154. doi: [10.1210/jc.2009-0345](https://doi.org/10.1210/jc.2009-0345) PMID: [19509099](https://pubmed.ncbi.nlm.nih.gov/19509099/)
48. Haider AH, Efron DT, Haut ER, Chang DC, Paidas CN, Cornwell EE 3rd. Mortality in adolescent girls vs boys following traumatic shock: an analysis of the National Pediatric Trauma Registry. *Arch Surg.* 2007; 142: 875–880; discussion 879–880. PMID: [17875843](https://pubmed.ncbi.nlm.nih.gov/17875843/)
49. Ley EJ, Short SS, Liou DZ, Singer MB, Mirocha J, Melo N, et al. Gender impacts mortality after traumatic brain injury in teenagers. *J Trauma Acute Care Surg.* 2013; 75: 682–686. doi: [10.1097/TA.0b013e31829d024f](https://doi.org/10.1097/TA.0b013e31829d024f) PMID: [24064883](https://pubmed.ncbi.nlm.nih.gov/24064883/)
50. Holmes GL, Weber DA. The effect of progesterone on kindling: a developmental study. *Brain Res.* 1984; 318: 45–53. PMID: [6488053](https://pubmed.ncbi.nlm.nih.gov/6488053/)
51. Frye CA. Progesterone reduces depressive behavior of young ovariectomized, aged progesterone receptor knockout, and aged wild type mice in the tail suspension test. *J Psychopharmacol.* 2011; 25: 421–428. doi: [10.1177/0269881109349836](https://doi.org/10.1177/0269881109349836) PMID: [19965943](https://pubmed.ncbi.nlm.nih.gov/19965943/)
52. Tsuji M, Taguchi A, Ohshima M, Kasahara Y, Ikeda T. Progesterone and allopregnanolone exacerbate hypoxic-ischemic brain injury in immature rats. *Exp Neurol.* 2012; 233: 214–220. doi: [10.1016/j.expneurol.2011.10.004](https://doi.org/10.1016/j.expneurol.2011.10.004) PMID: [22020180](https://pubmed.ncbi.nlm.nih.gov/22020180/)
53. Uysal N, Baykara B, Kiray M, Cetin F, Aksu I, Dayi A, et al. Combined treatment with progesterone and magnesium sulfate positively affects traumatic brain injury in immature rats. *Turk Neurosurg.* 2013; 23: 129–137. doi: [10.5137/1019-5149.JTN.5582-11.1](https://doi.org/10.5137/1019-5149.JTN.5582-11.1) PMID: [23546895](https://pubmed.ncbi.nlm.nih.gov/23546895/)
54. Robertson CL, Saraswati M. Progesterone protects mitochondrial function in a rat model of pediatric traumatic brain injury. *J Bioenerg Biomembr.* 2014.
55. Hill CA, Fitch RH. Sex differences in mechanisms and outcome of neonatal hypoxia-ischemia in rodent models: implications for sex-specific neuroprotection in clinical neonatal practice. *Neurol Res Int.* 2012; 2012: 867531. doi: [10.1155/2012/867531](https://doi.org/10.1155/2012/867531) PMID: [22474588](https://pubmed.ncbi.nlm.nih.gov/22474588/)
56. Mannix R, Berglass J, Berkner J, Moleus P, Qiu J, Jantzie LL, et al. Sex differences in the effect of progesterone after controlled cortical impact in adolescent mice: a preliminary study. *J Neurosurg.* 2014; 121: 1337–1341. doi: [10.3171/2014.8.JNS14715](https://doi.org/10.3171/2014.8.JNS14715) PMID: [25280093](https://pubmed.ncbi.nlm.nih.gov/25280093/)
57. Rogol AD, Roemmich JN, Clark PA. Growth at puberty. *J Adolesc Health.* 2002; 31: 192–200. PMID: [12470915](https://pubmed.ncbi.nlm.nih.gov/12470915/)
58. Attella MJ, Nattinville A, Stein DG. Hormonal state affects recovery from frontal cortex lesions in adult female rats. *Behav Neural Biol.* 1987; 48: 352–367. PMID: [3689284](https://pubmed.ncbi.nlm.nih.gov/3689284/)
59. Roof RL, Zhang Q, Glasier MM, Stein DG. Gender-specific impairment on Morris water maze task after entorhinal cortex lesion. *Behav Brain Res.* 1993; 57: 47–51. PMID: [8292254](https://pubmed.ncbi.nlm.nih.gov/8292254/)
60. Roof RL, Duvdevani R, Stein DG. Gender influences outcome of brain injury: progesterone plays a protective role. *Brain Res.* 1993; 607: 333–336. PMID: [8481809](https://pubmed.ncbi.nlm.nih.gov/8481809/)
61. Bramlett HM, Dietrich WD. Pathophysiology of cerebral ischemia and brain trauma: similarities and differences. *J Cereb Blood Flow Metab.* 2004; 24: 133–150. PMID: [14747740](https://pubmed.ncbi.nlm.nih.gov/14747740/)
62. Kolb B, Cioe J. Recovery from early cortical damage in rats, VIII. Earlier may be worse: behavioural dysfunction and abnormal cerebral morphogenesis following perinatal frontal cortical lesions in the rat. *Neuropharmacology.* 2000; 39: 756–764. PMID: [10699442](https://pubmed.ncbi.nlm.nih.gov/10699442/)
63. Hamm RJ, Pike BR, O'Dell DM, Lyeth BG, Jenkins LW. The rotarod test: an evaluation of its effectiveness in assessing motor deficits following traumatic brain injury. *J Neurotrauma.* 1994; 11: 187–196. PMID: [7932797](https://pubmed.ncbi.nlm.nih.gov/7932797/)
64. Adelson PD, Skinner JC, Davis DS, Tran MP, Dixon CE, Kochanek PM, et al. A model of focal TBI in the neonatal rat. *J Neurotrauma.* 2000; 17: 929.
65. Wagner AK, Willard LA, Kline AE, Wenger MK, Bolinger BD, Ren D, et al. Evaluation of estrous cycle stage and gender on behavioral outcome after experimental traumatic brain injury. *Brain Res.* 2004; 998: 113–121. PMID: [14725974](https://pubmed.ncbi.nlm.nih.gov/14725974/)

66. Pellow S, Chopin P, File SE, Briley M. Validation of open:closed arm entries in an elevated plus-maze as a measure of anxiety in the rat. *J Neurosci Methods*. 1985; 14: 149–167. PMID: [2864480](#)
67. Prut L, Belzung C. The open field as a paradigm to measure the effects of drugs on anxiety-like behaviors: a review. *Eur J Pharmacol*. 2003; 463: 3–33. PMID: [12600700](#)
68. Parker V, Morinan A. The socially isolated rat as a model for anxiety. *Neuropharmacology*. 1986; 25: 663–664.
69. Morris R. Developments of a water-maze procedure for studying spatial learning in the rat. *J Neurosci Methods*. 1984; 11: 47–60. PMID: [6471907](#)
70. Newhouse P, Newhouse C, Astur RS. Sex differences in visual-spatial learning using a virtual water maze in pre-pubertal children. *Behav Brain Res*. 2007; 183: 1–7. PMID: [17629971](#)
71. Kallai J, Makany T, Karadi K, Jacobs WJ. Spatial orientation strategies in Morris-type virtual water task for humans. *Behav Brain Res*. 2005; 159: 187–196. PMID: [15817182](#)
72. Astur RS, Tropp J, Sava S, Constable RT, Markus EJ. Sex differences and correlations in a virtual Morris water task, a virtual radial arm maze, and mental rotation. *Behav Brain Res*. 2004; 151: 103–115. PMID: [15084426](#)
73. Mueller SC, Temple V, Cornwell B, Grillon C, Pine DS, Ernst M. Impaired spatial navigation in pediatric anxiety. *J Child Psychol Psychiatry*. 2009; 50: 1227–1234. doi: [10.1111/j.1469-7610.2009.02112.x](#) PMID: [19594834](#)
74. Lacroix L, White I, Feldon J. Effect of excitotoxic lesions of rat medial prefrontal cortex on spatial memory. *Behav Brain Res*. 2002; 133: 69–81. PMID: [12048175](#)
75. Hull EM. Effects of neonatal exposure to progesterone in sexual behavior of male and female rats. *Physiol Behav*. 1981; 26: 401–405. PMID: [7243957](#)
76. Reddy DS, Gangisetty O, Briyal S. Disease-modifying activity of progesterone in the hippocampus kindling model of epileptogenesis. *Neuropharmacology*. 2010; 59: 573–581. doi: [10.1016/j.neuropharm.2010.08.017](#) PMID: [20804775](#)
77. Dableh LJ, Henry JL. Progesterone prevents development of neuropathic pain in a rat model: Timing and duration of treatment are critical. *J Pain Res*. 2011; 4: 91–101. doi: [10.2147/JPR.S17009](#) PMID: [21559355](#)
78. Adelson PD, Fellows-Mayle W, Kochanek PM, Dixon CE. Morris water maze function and histologic characterization of two age-at-injury experimental models of controlled cortical impact in the immature rat. *Childs Nerv Syst*. 2013; 29: 43–53. doi: [10.1007/s00381-012-1932-4](#) PMID: [23089934](#)
79. Kolb B, Mychasiuk R, Muhammad A, Gibb R. Brain plasticity in the developing brain. *Prog Brain Res*. 2013; 207: 35–64. doi: [10.1016/B978-0-444-63327-9.00005-9](#) PMID: [24309250](#)
80. Halliwell C, Comeau W, Gibb R, Frost DO, Kolb B. Factors influencing frontal cortex development and recovery from early frontal injury. *Dev Neurorehabil*. 2009; 12: 269–278. doi: [10.3109/17518420903087715](#) PMID: [20477557](#)
81. Kolb B, Mychasiuk R, Williams P, Gibb R. Brain plasticity and recovery from early cortical injury. *Dev Med Child Neurol*. 2011; 53 Suppl 4: 4–8. doi: [10.1111/j.1469-8749.2011.04054.x](#) PMID: [21950386](#)