

# In this issue of *Gut Microbes*

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It is an amazing time to be a microbiologist. We have progressed from the study of microbes in monoculture to the study of microbes in mixed culture to the study of complex microbial communities in natural environments. Colony purification was an extremely powerful reductionist tool, but after decades of genetic analyses, many genes of model microbes still have no defined function. It is clear that many of these genes may be expressed, or may have phenotypes, only in the presence of other organisms. The early discovery of antibiotics was a step in this direction. The parallel discoveries of quorum sensing and biofilms led to the realization that individual cells of a species can cooperate with one another, in a coordinated multi-cellular fashion, to achieve common goals and to provide three dimensional structures to communities.<sup>1-3</sup> These findings were followed by numerous discoveries of chemical signaling between microbial species and even between kingdoms. Often neglected, the metabolic interactions between organisms are essential to microbial community structure and function.<sup>4</sup> Even the totality of these chemical signaling and metabolic interactions are probably just scratching the surface of the complexity with which microbes interact. The state of the “microbial interactions” field today seems similar to the state of “microbial pathogenesis” in the 1980s. At the time it was thought that pathogens, such as *Salmonella*, were simply “tougher” than organisms such as commensal *E. coli*. *Salmonella* could survive in hostile environments such as host immune cells because it was more resistant to digestive enzymes, acid, and oxidative stress than *E. coli*. It was later found that the mechanisms by which *Salmonella* evades host defenses are vastly more sophisticated, with secretion systems injecting more than 40 different proteins into host cells to manipulate cellular physiology in specific and elegant ways that are still far from understood. Although much less is known regarding microbial interactions, it is likely that they will turn out to be highly sophisticated as well. A striking example is the recent discovery that *P. aeruginosa* uses its Type 6 Secretion System (T6SS) to inhibit other bacteria only in response to attacks from other bacteria that also yield a T6SS, or even in response to mating attempts.<sup>5,6</sup>

In this issue of *Gut Microbes*, we focus on “microbial interactions” specifically within the gut. While this field is in its infancy, its importance is staggering. The development of a healthy microbial community is critical to the development of a healthy person. It seems like every day, more and more human diseases including inflammatory bowel disease (IBD), diabetes, obesity, cardiovascular disease, and allergies are suspected of originating with microbiota imbalances or dysbiosis.<sup>7-12</sup> There is

even increasing evidence for a link between the gut microbiota and brain function, including autism spectrum disorder (ASD).<sup>13</sup> The microbiota is required for the nutrition of the host, the development of intestinal tissues, and the development of the host immune system.<sup>9,11,14,15</sup> The microbiota is also critically important for protecting the host from pathogens (often called colonization resistance [CR]). For instance, disruption of the mouse intestinal microbiota decreases the LD<sub>50</sub> for *Salmonella* from 10<sup>6</sup> cfu to less than 10 cfu.<sup>16-20</sup> Partial restoration of resistance to *Salmonella* is achieved by inoculating the disrupted mice with a fecal suspension from untreated mice.<sup>21</sup> Clearly the host’s normal microbiota plays a very important role in preventing pathogen colonization. Not surprisingly, gut communities are specifically adapted to the species, even the genotype, of the host.<sup>22,23</sup> For instance, mouse microbial communities are better than human communities at preventing *Salmonella* from inflaming the intestine.<sup>15</sup> A germ-free mouse can be inoculated with conventional mouse feces and become resistant to *Salmonella*-mediated inflammation within days. But germ-free mice inoculated with human feces do not become resistant.<sup>15</sup> Elucidating the mechanisms underlying phenomena such as these might one day lead to the rational design of novel probiotics and antibiotics, provide new insights into pathogen host ranges, and contribute to our understanding of the ecology of diseases and epidemics. This isn’t just hype. Antibiotic-induced dysbiosis is the most clinically prevalent dysbiosis in the US healthcare system.<sup>24-28</sup> Broad-spectrum antibiotics can induce long-lasting effects on gut bacterial communities that ultimately result in gastrointestinal pathology.<sup>29,30</sup> Approximately 25% of cases of antibiotic-associated diarrhea are due to *Clostridium difficile*.<sup>26,31</sup> The spectrum of resulting disease can range from a state of asymptomatic carrier to pseudomembranous colitis and death.<sup>27,28</sup> In fact, what one might consider the ultimate probiotic, fecal transplantation, is remarkably successful at curing recurrent *C. difficile* infection.<sup>32</sup> A new industry will spring up if fecal transplantation proves successful in the treatment of obesity or ASD. The FDA held a public workshop in May 2013 to discuss the issues surrounding fecal transplantation, two of which are quality control and patient aversion. Both of these can be solved if combinations of isolated microbial species could be developed into effective probiotics.<sup>33,34</sup> One combination has already shown effectiveness in treating recurrent *C. difficile* infection and comes with the catchy name of “RePOOPulate”.<sup>35</sup> This type of treatment will put more focus on researchers to determine the mechanism of action of probiotics and the mechanisms that

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allow these organisms to persist, or not, among disparate gut communities.<sup>36,37</sup>

Continuing on these themes, in this issue Vincent Young's group provides a review on the metabolic environment of the intestine and how disruptions of this environment facilitate infection by *Clostridium difficile*.<sup>38</sup> Bruce McClane's group provides a review on *Clostridium perfringens* and how it detects compounds produced by epithelial cells.<sup>39</sup> Zhongtang Yu's lab provides a comprehensive review on the microbiome of poultry,

what factors are known to affect this microbiome and what affects the microbiome has on the host.<sup>40</sup> Jun Zhu's group provides a review on quorum sensing by *Vibrio cholerae* within the intestine,<sup>41</sup> and Dennis Kasper's lab provides a review on the host response to commensals, more specifically, on innate lymphocytes.<sup>42</sup>

#### Disclosure of Potential Conflicts of Interest

No potential conflict of interest was disclosed.

#### References

- Costerton JW, Geesey GG, Cheng KJ. How bacteria stick. *Sci Am* 1978; 238:86-95; PMID:635520; <http://dx.doi.org/10.1038/scientificamerican0178-86>
- Fuqua WC, Winans SC, Greenberg EP. Quorum sensing in bacteria: the LuxR-LuxI family of cell density-responsive transcriptional regulators. *J Bacteriol* 1994; 176:269-75; PMID:8288518
- Hall-Stoodley L, Costerton JW, Stoodley P. Bacterial biofilms: from the natural environment to infectious diseases. *Nat Rev Microbiol* 2004; 2:95-108; PMID:15040259; <http://dx.doi.org/10.1038/nrmicro821>
- Fischbach MA, Sonnenburg JL. Eating for two: how metabolism establishes interspecies interactions in the gut. *Cell Host Microbe* 2011; 10:336-47; PMID:22018234; <http://dx.doi.org/10.1016/j.chom.2011.10.002>
- Ho BT, Basler M, Mekalanos JJ. Type 6 secretion system-mediated immunity to type 4 secretion system-mediated gene transfer. *Science* 2013; 342:250-3; PMID:24115441; <http://dx.doi.org/10.1126/science.1243745>
- Basler M, Ho BT, Mekalanos JJ. Tit-for-tat: type VI secretion system counterattack during bacterial cell-cell interactions. *Cell* 2013; 152:884-94; PMID:23415234; <http://dx.doi.org/10.1016/j.cell.2013.01.042>
- Wen L, Ley RE, Volchkov PY, Stranges PB, Avanesyan L, Stonebraker AC, Hu C, Wong FS, Szot GL, Bluestone JA, et al. Innate immunity and intestinal microbiota in the development of Type 1 diabetes. *Nature* 2008; 455:1109-13; PMID:18806780; <http://dx.doi.org/10.1038/nature07336>
- Dinalo JE, Relman DA. Cross-talk in the gut. *Genome Biol* 2009; 10:203; PMID:19216729; <http://dx.doi.org/10.1186/gb-2009-10-1-203>
- Sekirov I, Russell SL, Antunes LCM, Finlay BB. Gut microbiota in health and disease. *Physiol Rev* 2010; 90:859-904; PMID:20664075; <http://dx.doi.org/10.1152/physrev.00045.2009>
- Huffnagle GB. The microbiota and allergies/asthma. *PLoS Pathog* 2010; 6:e1000549; PMID:20523892; <http://dx.doi.org/10.1371/journal.ppat.1000549>
- Tremaroli V, Bäckhed F. Functional interactions between the gut microbiota and host metabolism. *Nature* 2012; 489:242-9; PMID:22972297; <http://dx.doi.org/10.1038/nature11552>
- Brown EM, Arrieta M-C, Finlay BB. A fresh look at the hygiene hypothesis: how intestinal microbial exposure drives immune effector responses in atopic disease. *Semin Immunol* 2013; 25:378-87; PMID:24209708; <http://dx.doi.org/10.1016/j.smim.2013.09.003>
- Hsiao EY, McBride SW, Hsien S, Sharon G, Hyde ER, McCue T, Codelli JA, Chow J, Reisman SE, Petrosino JF, et al. Microbiota modulate behavioral and physiological abnormalities associated with neurodevelopmental disorders. *Cell* 2013; 155:1451-63; PMID:24315484; <http://dx.doi.org/10.1016/j.cell.2013.11.024>
- Bäckhed F, Ley RE, Sonnenburg JL, Peterson DA, Gordon JI. Host-bacterial mutualism in the human intestine. *Science* 2005; 307:1915-20; PMID:15790844; <http://dx.doi.org/10.1126/science.1104816>
- Chung H, Pamp SJ, Hill JA, Surana NK, Edelman SM, Troy EB, Reading NC, Villablanca EJ, Wang S, Mora JR, et al. Gut immune maturation depends on colonization with a host-specific microbiota. *Cell* 2012; 149:1578-93; PMID:22726443; <http://dx.doi.org/10.1016/j.cell.2012.04.037>
- Nardi RM, Silva ME, Vieira EC, Bambirra EA, Nicoli JR. Intra gastric infection of germfree and conventional mice with *Salmonella typhimurium*. *Braz J Med Biol Res* 1989; 22:1389-92; PMID:2700668
- Que JU, Hentges DJ. Effect of streptomycin administration on colonization resistance to *Salmonella typhimurium* in mice. *Infect Immun* 1985; 48:169-74; PMID:3884509
- Bohnhoff M, Drake BL, Miller CP. Effect of streptomycin on susceptibility of intestinal tract to experimental *Salmonella* infection. *Proc Soc Exp Biol Med* 1954; 86:132-7; PMID:13177610; <http://dx.doi.org/10.3181/00379727-86-21030>
- Hapfelmeier S, Hardt WD. A mouse model for *S. typhimurium*-induced enterocolitis. *Trends Microbiol* 2005; 13:497-503; PMID:16140013; <http://dx.doi.org/10.1016/j.tim.2005.08.008>
- Spees AM, Lopez CA, Kingsbury DD, Winter SE, Bäuml AJ. Colonization resistance: battle of the bugs or Ménéage à Trois with the host? *PLoS Pathog* 2013; 9:e1003730; PMID:24278012; <http://dx.doi.org/10.1371/journal.ppat.1003730>
- Miller CP, Bohnhoff M. Changes in the Mouse's Enteric Microflora Associated with Enhanced Susceptibility to *Salmonella* Infection Following Streptomycin Treatment. *J Infect Dis* 1963; 113:59-66; PMID:14044094; <http://dx.doi.org/10.1093/infdis/113.1.59>
- Rawls JF, Mahowald MA, Ley RE, Gordon JI. Reciprocal gut microbiota transplants from zebrafish and mice to germ-free recipients reveal host habitat selection. *Cell* 2006; 127:423-33; PMID:17055441; <http://dx.doi.org/10.1016/j.cell.2006.08.043>
- Turnbaugh PJ, Hamady M, Yatsunenko T, Cantarel BL, Duncan A, Ley RE, Sogin ML, Jones WJ, Roe BA, Affourtit JP, et al. A core gut microbiome in obese and lean twins. *Nature* 2009; 457:480-4; PMID:19043404; <http://dx.doi.org/10.1038/nature07540>
- Lemon KP, Armitage GC, Relman DA, Fischbach MA. Microbiota-targeted therapies: an ecological perspective. *Sci Transl Med* 2012; 4:rv5; PMID:22674555; <http://dx.doi.org/10.1126/scitranslmed.3004183>
- Dethlefsen L, Relman DA. Incomplete recovery and individualized responses of the human distal gut microbiota to repeated antibiotic perturbation. *Proc Natl Acad Sci U S A* 2011; 108(Suppl 1):4554-61; PMID:20847294; <http://dx.doi.org/10.1073/pnas.1000087107>
- Pacheco SM, Johnson S. Important clinical advances in the understanding of *Clostridium difficile* infection. *Curr Opin Gastroenterol* 2013; 29:42-8; PMID:23207596; <http://dx.doi.org/10.1097/MOG.0b013e32835a68d4>
- Bouza E. Consequences of *Clostridium difficile* infection: understanding the healthcare burden. *Clin Microbiol Infect* 2012; 18(Suppl 6):5-12; PMID:23121549; <http://dx.doi.org/10.1111/1469-0691.12064>
- Cecil JA. *Clostridium difficile*: Changing Epidemiology, Treatment and Infection Prevention Measures. *Curr Infect Dis Rep* 2012; 14:612-9; PMID:23054932; <http://dx.doi.org/10.1007/s11908-012-0298-9>
- Relman DA. The human microbiome: ecosystem resilience and health. *Nutr Rev* 2012; 70(Suppl 1):S2-9; PMID:22861804; <http://dx.doi.org/10.1111/j.1753-4887.2012.00489.x>
- Cotter PD, Stanton C, Ross RP, Hill C. The impact of antibiotics on the gut microbiota as revealed by high throughput DNA sequencing. *Discov Med* 2012; 13:193-9; PMID:22463795
- Dubberke ER, Olsen M. Burden of *Clostridium difficile* on the healthcare system. *Clin Infect Dis* 2012; 55(Suppl 2):S88-92; PMID:22752870; <http://dx.doi.org/10.1093/cid/cis335>
- van Nood E, Vrieze A, Nieuwdorp M, Fuentes S, Zoetendal EG, de Vos WM, Visser CE, Kuijper EJ, Bartsman JFWM, Tijssen JGP, et al. Duodenal infusion of donor feces for recurrent *Clostridium difficile*. *N Engl J Med* 2013; 368:407-15; PMID:23323867; <http://dx.doi.org/10.1056/NEJMoal205037>
- Goodman AL, Kallstrom G, Faith JJ, Reyes A, Moore A, Dantas G, Gordon JI. Extensive personal human gut microbiota culture collections characterized and manipulated in gnotobiotic mice. *Proc Natl Acad Sci U S A* 2011; 108:6252-7; PMID:21436049; <http://dx.doi.org/10.1073/pnas.1102938108>
- Faith JJ, Rey FE, O'Donnell D, Karlsson M, McNulty NP, Kallstrom G, Goodman AL, Gordon JI. Creating and characterizing communities of human gut microbes in gnotobiotic mice. *ISME J* 2010; 4:1094-8; PMID:20664551; <http://dx.doi.org/10.1038/ismej.2010.110>
- Petrof EO, Gloor GB, Vanner SJ, Weese SJ, Carter D, Daigneault MC, Brown EM, Schroeter K, Allen-Vercoe E. Stool substitute transplant therapy for the eradication of *Clostridium difficile* infection: "RePOOPulating" the gut. *Microbiome* 2013; 1:3; <http://dx.doi.org/10.1186/2049-2618-1-3>
- Goodman AL, McNulty NP, Zhao Y, Leip D, Mitra RD, Luzopune CA, Knight R, Gordon JI. Identifying genetic determinants needed to establish a human gut symbiont in its habitat. *Cell Host Microbe* 2009; 6:279-89; PMID:19748469; <http://dx.doi.org/10.1016/j.chom.2009.08.003>

37. Deriu E, Liu JZ, Pezeshki M, Edwards RA, Ochoa RJ, Contreras H, Libby SJ, Fang FC, Raffatellu M. Probiotic bacteria reduce salmonella typhimurium intestinal colonization by competing for iron. [Internet]. *Cell Host Microbe* 2013; 14:26-37; <http://linkinghub.elsevier.com/retrieve/pii/S1931312813002230>; PMID:23870311; <http://dx.doi.org/10.1016/j.chom.2013.06.007>.
38. Theriot CM, Young VB. Microbial and metabolic interactions between the gastrointestinal tract and *Clostridium difficile* infection. *Gut Microbes* 2013; 5; PMID:24335555; <http://dx.doi.org/10.4161/gmic.27131>
39. Chen J, Ma M, Uzal FA, McClane BA. Host cell-induced signaling causes *Clostridium perfringens* to upregulate production of toxins important for intestinal infections. *Gut Microbes* 2013; 5; PMID:24061146; <http://dx.doi.org/10.4161/gmic.26419>
40. Pan D, Yu Z. Intestinal microbiome of poultry and its interaction with host and diet. *Gut Microbes* 2013; 5:5; PMID:24256702; <http://dx.doi.org/10.4161/gmic.26945>
41. Rothenbacher FP, Zhu J. Efficient responses to host and bacterial signals during *Vibrio cholerae*-colonization. *Gut Microbes* 2014;5; <http://dx.doi.org/10.4161/gmic.26944>
42. Chen VL, Kasper DL. Interactions between the intestinal microbiota and innate lymphoid cells. *Gut Microbes* 2013; 5; PMID:24418741; <http://dx.doi.org/10.4161/gmic.27289>