

## Anti-Aging Effects of the *Hanwoo* Leg Bone, Foot and Tail Infusions (HLI, HFI and HTI) on Skin Fibroblast

Ja young Seol, Ji Young Yoon<sup>1</sup>, Hee Sun Jeong<sup>2</sup>, Nami Joo<sup>3</sup>, and Soon Young Choi\*

Division of Biological Science, Sookmyung Women's University, Seoul 04310, Korea

<sup>1</sup>Department of Le Cordon Blue Hospitality Management, Sookmyung Women's University, Seoul 04310, Korea

<sup>2</sup>Traditional Dietary Life Food, Graduate School of Traditional Culture and Arts,

Sookmyung Women's University, Seoul 04310, Korea

<sup>3</sup>Department of Food & Nutrition, Sookmyung Women's University, Seoul 04310, Korea

### Abstract

Many researchers revealed that collagen contribute to maintaining the skin's elasticity and inhibit wrinkling of skin. Korean native cattle (*Hanwoo*) bone (leg bone, foot and tail) infusion contains the various inorganic materials, collagen and chondroitin sulfate. All of this, a large quantity of collagen is included in *Hanwoo* infusion. Therefore, this study emphasized on the effects of collagen in the *Hanwoo* bone infusion. For the first time, *Hanwoo* bone infusions were directly added to the media of Human Dermal Fibroblast (NHDF-c) to test anti-aging effects. First, it was identified that growth rate of skin fibroblast was increased. Furthermore, the *Hanwoo* bone infusion increased a 50% of fibroblast collagen synthesis. Also, suppression of skin fibroblast aging was confirmed by treatment *Hanwoo* bone infusion. In conclusion, this study demonstrates the effects of infusion made from *Hanwoo* leg bone, foot and tail on anti-aging, wrinkle inhibiting and skin fibroblast elasticity maintaining. Therefore, this study identified that traditional infusion has effects that are good for skin elasticity.

**Keywords:** Korean native cattle (*Hanwoo*), leg bone infusion, foot infusion, tail infusion, skin fibroblast (NHDF-c), anti-aging

Received September 6, 2015; Revised February 22, 2016; Accepted February 22, 2016

### Introduction

*Hanwoo* bones were boiled and resulting infusions are commonly ingested as soup in Korea. In general, the *Hanwoo* bones used for soup were the leg bone, foot, and tail. The foot refers to the bone below the kneecap and the leg bone refers to the two leg bones above the foot. The tail consists of several joints including the hip bone. The main components of *Hanwoo* bone infusions are collagen, chondroitin sulfate and inorganic materials, like calcium, sodium and magnesium (Park, 1986; Park and Lee, 1982; Seol and Jang, 1990). Among them, collagen and chondroitin sulfate are known to have effects on the bone and skin (Kim, 2003; Saito and Marumo, 2010; Salaszyk *et al.*, 2004; Shizuka *et al.*, 2013). Collagen has functions such as maintaining skin elasticity, strengthening joints and retaining moisture. However, due to the

large size of the collagen molecule, ingested collagen is unable to absorb into the body. However, previous studies show that collagen was found in blood following the oral ingestion (Ichikawa *et al.*, 2010; Koji *et al.*, 2005; Sugihara *et al.*, 2012), and the effects of the collagen on skin health have been published (Vivian *et al.*, 2011; Zague, 2008). In another paper, the BioCellcollagen that is the gelatin made from the collagen of the chicken heart cartilage bone, improved blood circulation and reduced the aging of the skin (Schwartz and Park, 2012). In addition, Matsuda *et al.* reported the collagen peptides taken by pigs through oral administration increased the density of fibroblast and the diameter and density of collagen fibrils (Matsuda *et al.*, 2006). Also, it was reported that mouse skin cell growth was promoted when treated with the collagen peptides (Yasutaka *et al.*, 2009).

Normal epidermal cell differentiation continues as skin ages. However, the dermal matrix or connective tissue is reduced due to the reduction of keratinocyte division, causing skin become thin, dry and wrinkled (Kim *et al.*, 2010). The connective tissue of the skin consists mostly of collagen and elastin. Collagen and elastin provide elas-

\*Corresponding author: Soon Young Choi, Division of Biological Science, Sookmyung Women's University, Seoul 04310, Korea. Tel:+82-2-710-9510, Fax:+82-2-2077-7322, E-mail: sychoi@sookmyung.ac.kr

ticity and strength to the skin. When elasticity and strength are weakened due to aging, the skin becomes prone to damages and aging (Makrantonaki and Zouboulis, 2007). Skin wrinkling is caused by an imbalance in collagen synthesis and decomposition. In young skin, the matrix metalloproteinase (MMP)-1 and 9 collagenases help to balance collagen synthesis and decomposition. As skin ages, the synthesis of the collagen decreases and the activity of MMP-1, 9 collagenase increases (Talwar *et al.*, 1995). The increased MMPs within skin fibroblast due to repeated exposure to ultraviolet (UV) make skin collagen decomposition and causes skin-wrinkling (Imokawa, 2008). Skin aging can be identified by the growth rate of the skin fibroblast, suppression of collagenase and the amount of collagen synthesis. In order to determine the extent of skin-aging, activity level of senescence-associated  $\beta$ -galactosidase can be used for assessing senescence in mammalian cells. Through this study, we verified the effects of the *Hanwoo bone infusions*, which contain large amounts of collagen, on skin elasticity and senescence suppression.

## Materials and Methods

### Cell culture

The skin fibroblast used in the experiments was NHDF-c (adult human dermal fibroblast) obtained from Promo-Cell (Germany). Skin fibroblast was cultured in DMEM media (Welgene Inc., Korea) supplemented with 10% fetal bovine serum (Gibco BRL, USA) and 1% penicillin/streptomycin (Welgene Inc., Korea) at 37°C in a 5% CO<sub>2</sub> incubator (ASTEC, Japan). The used cell passage numbers is between three and ten. Skin fibroblast was cultured for 24 h until they reached 70% confluency, and then further cultured in the FBS-free media for 24 h. Each infusion of *Hanwoo's* leg bone, foot and tail was added at the DMEM media without FBS, cells were cultured in 5% CO<sub>2</sub> incubator (ASTEC, Japan) for 48 h. The control group was cultured in the media without the infusions.

### Reagents

The Bovine Collagen, that is the pure collagen was purchased from Biocolor Life Science (U.K.) and used for the control experiment.

### Preparation of the *Hanwoo* leg bone, foot and tail infusions

The sample infusion that used in these experiments was prepared according to the optimization recipe presented by the Yoon *et al.* (2015). Four kg of each *Hanwoo* leg

bone, foot, and tail was boiled in 20 L water for 12 h. Final volume is about 15 L. The *Hanwoo* leg bone infusion used the four *Hanwoo* leg bones was marked *leg bone infusion* or HLI. The *Hanwoo* foot infusion used the part of the *Hanwoo's* foot under the kneecap was marked *foot infusion* or HFI. The *Hanwoo* tail infusion used the *Hanwoo's* coccyx was marked *tail infusion* or HTI.

### Cell proliferation assay

The cell proliferation rate was measured by using the Cell Proliferation Reagent WST-1 (Roche, Germany). The skin fibroblast was cultured in 96-well plates ( $4 \times 10^3$ /well) in DMEM media with 10% FBS for 24 h and cultured additionally in the media without FBS for 24 h. Next, each infusion was added to the DMEM media without FBS and skin fibroblast was cultured in 5% CO<sub>2</sub> incubator for 72 h. Skin fibroblast was cultured at 37°C in a 5% CO<sub>2</sub> incubator (ASTEC, Japan) for 4 h after adding 10  $\mu$ L of Cell Proliferation Reagent WST-1 to each well. Absorbance was measured at 450 nm by using of ELISA Reader (Molecular Devices, UK).

### Collagen quantitative analysis

The amount of the collagen synthesized by skin fibroblast was measured using the Sircol Collagen Assay Kit (Biocolor Life Science, U.K.), according to the manufacturer's instructions. Skin fibroblast was cultured in 6-well plates ( $1 \times 10^5$ /well) in DMEM media with 10% FBS for 24 h. Next, each HLI, HFI and HTI was added to the separate DMEM medium with 1% FBS then incubated in a 5% CO<sub>2</sub> incubator for 72 h. After collecting the culture media in microcentrifuge tube, the collagen in the media was extracted and concentrated for 18 h with collagen isolation and concentration reagent which contains polyethylene glycol in a TRIS-HCL buffer, pH7.6 (Biocolor Life Science). Then, microcentrifuge tube was centrifuged at 12,000 rpm for 10 min, without delay. The pellet of hydrated transparent collagen is invisible. The Sircol dye that selectively combines with collagen was added to the concentrated culture media, and ice-cold acid-salt wash reagent was added gently to the collagen-dye pellet to remove the unbound dye. The bound dye was released and dissolved by adding 0.5 M sodium hydroxide. The released Sirius Red Dye was measured calorimetrically at 550 nm by ELISA Reader (Molecular Devices).

### Senescent cell assay

Senescence-associated  $\beta$ -galactosidase (SA- $\beta$ -Gal) activity, which is increased through aging, was measured by

the Senescent Cells Histochemical Staining Kit (Sigma, USA) (Joseph *et al.*, 2000; Ronald and Susan 2005). Skin fibroblast was cultured in 35-mm culture dishes ( $1 \times 10^5$  / well) with DMEM media with each HLI, HFI and HTI for 72 h. After fixing for 7 min with 0.2% glutaraldehyde and 2% formaldehyde, the staining solution (5mM potassium ferricyanide, 5mM potassium ferrocyanide, X-gal solution) was added to the media and incubated at 37°C without CO<sub>2</sub> for 24 h. The senescent skin fibroblast, dyed in blue, was examined under a microscope ( $\times 200$ ) (Olympus, Japan)

### Western blot analysis

Total protein was extracted from cells using lysis buffer. After SDS-polyacrylamide gel electrophoresis, proteins were transferred onto PVDF membranes using a semi-dry transfer unit-TE 70 (Amersham Biosciences, USA). The membranes were blocked with 4% skim milk and probed with the primary antibody at 4°C for 12 h. Blots were washed and incubated with the second antibody for 1 h. Using the LAS-3000 (Fujifilm Co., Japan), the changes in protein expression were identified. MMP1, MMP9 (Santa Cruz, USA) and actin (Sigma-Aldrich Co., USA) were used as primary antibodies.

### Statistical analysis

All results are expressed as means  $\pm$  standard deviation from the three independent experiments. Statistical analysis was performed using SAS9.1 (SAS Institute, USA).  $p < 0.05$  was considered statistically significant.

## Results and Discussion

### The HLI, HFI and HTI increase the proliferation of skin fibroblast

In order to confirm the effects of the HLI, HFI and HTI on the activation of the skin fibroblast metabolism, incre-

ments of the skin fibroblast division and cell growth rate were measured by treating skin fibroblast with the HLI, HFI and HTI. When treating skin fibroblast with the HLI, HFI and HTI, the protein concentration in each infusion was measured. When treating skin fibroblast with each infusion, skin fibroblast proliferation increased in comparison to the untreated control fibroblast. When comparing infusion concentrations, the concentration of infusion that displayed the highest growth rates was as follows: the 20 ug of *Hanwoo* leg bone protein showing 54% increased in growth rate, 10 ug *Hanwoo* foot protein at 60% and 10 ug *Hanwoo* tail protein at 65% (Fig. 1). Statistical analysis was carried out to configure the statistical significance of protein concentration differences ( $p < 0.05$ ). Consequently, the HLI, HFI and HTI increased the proliferation of skin fibroblast. Increase of the growth rate of skin fibroblast means increment in dermal tissue and connective tissue. Therefore, it was expected that the increase in the skin fibroblast growth rate may be effective in suppressing skin wrinkling.

### The HLI, HFI and HTI increase skin fibroblast collagen synthesis

The amount of skin fibroblast collagen synthesis was examined as the next indicator regarding effects of the HLI, HFI and HTI on skin fibroblast metabolism. The collagen content in each infusion was measured to analyze the correlation between the amount of skin fibroblast collagen synthesis and the collagen content in each infusion (Fig. 2A). When comparing the collagen content in 1 mL of each infusion, the collagen content was highest in the HFI followed by the HLI and the HTI. This finding coincided with the results from the preceding studies by Yoon *et al.* (2015). Next, the amount of skin fibroblast collagen synthesis was measured after adding each infusion to the media of the skin fibroblast. Skin fibroblast treated with each infusion synthesized more collagen than untreated

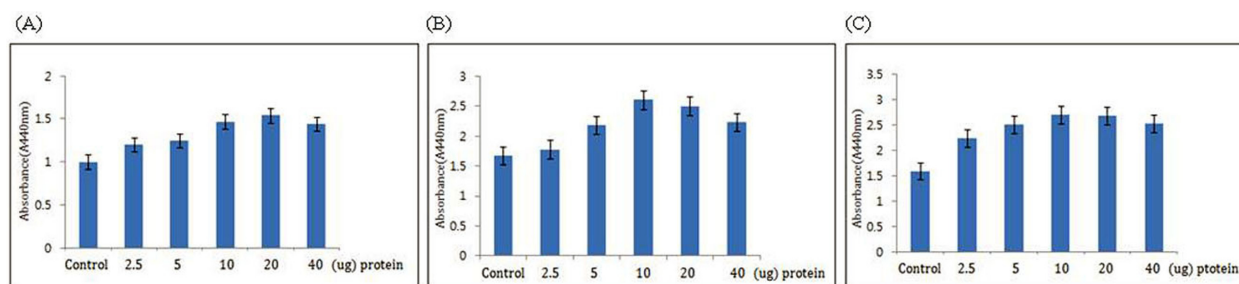
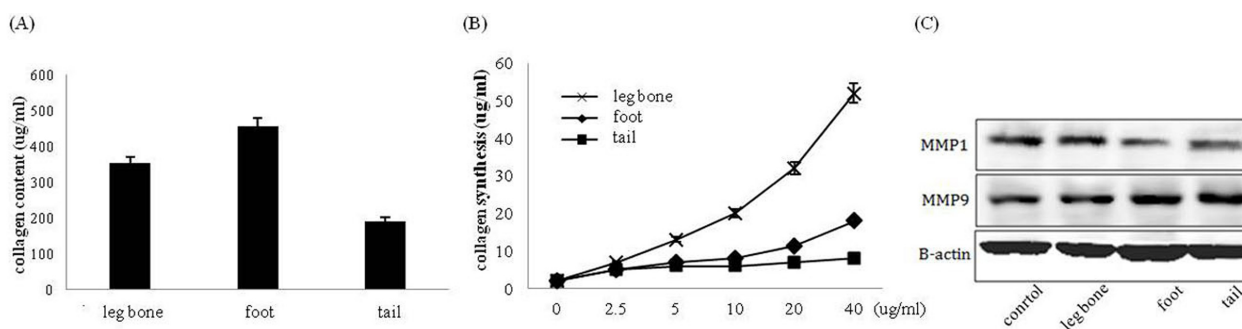


Fig. 1. The effect of the *Hanwoo* bone (leg bone, foot and tail) infusions on the cell proliferation of skin fibroblast by WST-1. (A) *Hanwoo* leg bone infusion. (B) *Hanwoo* foot infusion. (C) *Hanwoo* tail infusion.



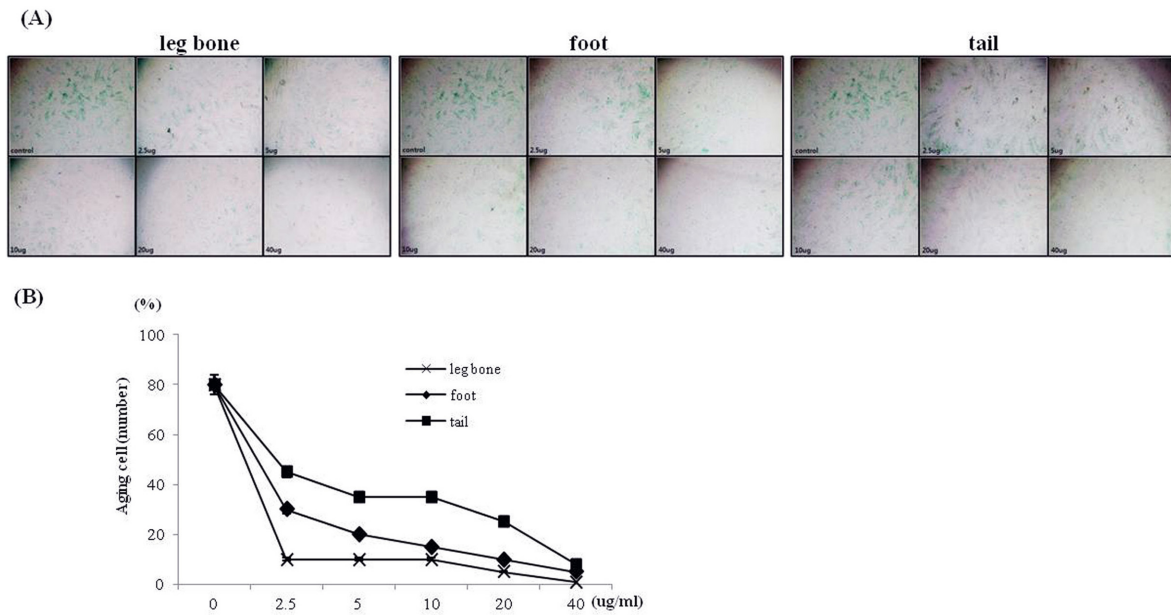
**Fig. 2.** The effect of the *Hanwoo bone* (leg bone, foot and tail) infusions on the collagen synthesis assay of skin fibroblast by Sircol™. (A) Collagen content contained in the *Hanwoo bone* (leg bone, foot and tail) infusions. (B) Collagen synthesis of skin fibroblast. (C) Expression of MMP1, MMP-9 protein in skin fibroblast treated with the *Hanwoo bone* (leg bone, foot and tail) infusions by western-blot.

skin fibroblast. When cells were treated with 40 ug of protein from each infusion, the amount of the collagen synthesis was increased by approximately 50% with HLI, 20% with HFI, and 10% with HTI (Fig. 2B). The HLI, HFI and HTI increased the amount of skin fibroblast collagen synthesis, which showed its effectiveness in maintaining the skin elasticity. To this effect, the HLI showed the highest amount of the collagen synthesis, followed by the HFI and the HTI, respectively. In order to determine the exact amount of the collagen synthesis caused by infusion treatments, the amount of collagen content in each infusion was subtracted from the total amount of the collagen synthesis in media. In contrast to our expectations, direct correlation between the collagen content in infusion and the amount of the collagen synthesis of skin fibroblast treated with the HLI, HFI and HTI was not found. Thus, it was confirmed that the activation of the skin fibroblast metabolism was not directly related to the collagen content in the infusion. The results suggested that there is a synergistic effect from ingredients other than collagen. In order to confirm this expectation, the bovine collagen was only treated at skin fibroblast with different concentrations to examine the amount of the skin fibroblast collagen synthesis (Fig. 4A). The amount of collagen synthesis did not increase even when treated with the highest collagen concentration (1,000 ug/mL). Thus, the effect of each infusion on the synthesis of collagen may be the result of the synergistic effects of the collagen and the other ingredients in bone infusion. Next, it was necessary to confirm whether the increase in the amount of collagen synthesis was a result of the reduction of collagenase MMP-1 and MMP-9 amount or the result of increase in the amount of the collagen synthesis. By the western blot assay, the protein amount of MMP-1 and MMP-9 was analyzed following

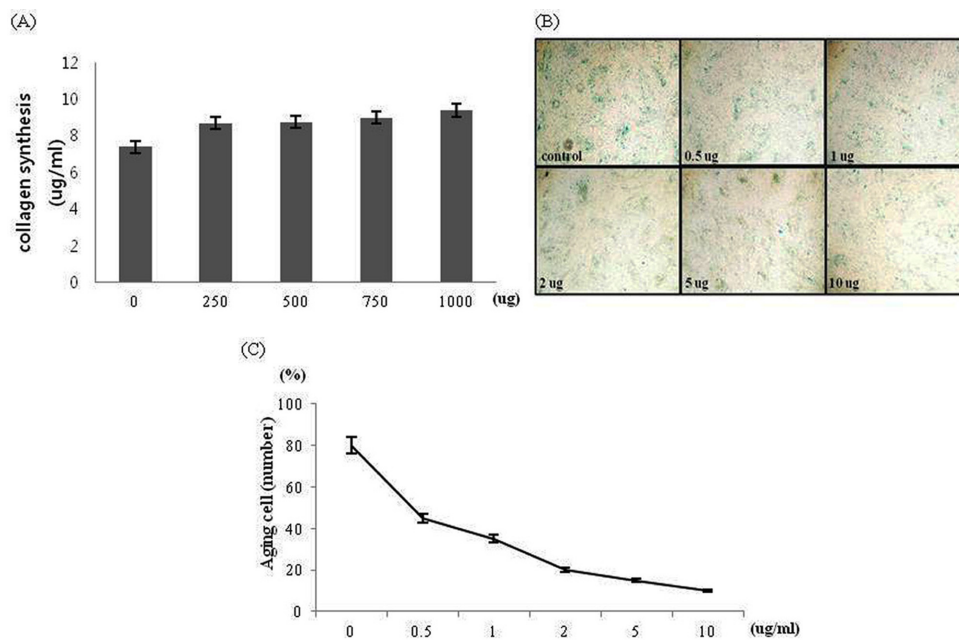
the treatment of skin fibroblast with the *Hanwoo bone* infusions. In case of the HLI, there was no significant changes in the protein amount of MMP-1 and MMP-9; small decrease in MMP-1 and MMP-9 protein amount was observed in the case of the HFI and the HTI (Fig. 2C). Thus, the increased amount of skin fibroblast collagen synthesis may be the effect of the increase in the collagen synthesis rather than the suppression of the collagen decomposition.

#### The HLI, HFI and HTI inhibit senescence of skin fibroblast

Since the treatment of each infusion increased the amount of skin fibroblast collagen synthesis, the infusions may affect skin elasticity. It was verified whether skin fibroblast senescence is effectively suppressed by infusion treatment, senescence-associated  $\beta$ -galactosidase was measured. Using the senescent cell staining kit, the aged cells were dyed so as to distinguish them from healthy cells (Fig. 3A). The number of aged cells in the fixed sector was counted (Fig. 3B). The results displayed high anti-aging efficacy of the *Hanwoo bone* infusions even in low concentration of each infusion. In the case of the 40 ug protein concentrate, only few senescent cells were observed. Statistical analysis was performed in order to identify whether there was a difference according to the concentrations. Analysis showed significant differences according to protein concentrations ( $p < 0.001$ ). The number of aged cells decreased remarkably in comparison to the fibroblast control group without the infusion treatment. The anti-aging effects of collagen was confirmed by culturing skin fibroblast with the bovine collagen in order to prove the effects of the collagen itself on the anti-aging of skin fibroblast (Fig. 4B, C). When skin fibroblast was treated



**Fig. 3.** The effect of the *Hanwoo* bone (leg bone, foot and tail) infusions on the senescence of skin fibroblast by senescent cells histochemical staining kit. (A) The senescent skin fibroblast was observed under a fluorescent microscope after cells were stained with senescent cells histochemical staining kit ( $\times 200$ ). (B) Aged cells were numbered.



**Fig. 4.** The effect of the bovine collagen on skin fibroblast metabolism. (A) Collagen synthesis. (B) The senescent skin fibroblast were observed under a fluorescent microscope after cells were stained with senescent cells histochemical staining kit ( $\times 200$ ). (C) Aged cells were numbered.

with the highest concentration of bovine collagen, 10 ug/mL, the aging of the fibroblast was inhibited. However, the result was not as effective as when skin fibroblast was cultured with the *Hanwoo* bone infusions. From these

results, it could be concluded that the anti-aging effects of each infusion on skin fibroblast was caused by collagen and other ingredients. This assumption would coincide with the result that bovine collagen was unable to increase

the collagen synthesis in comparison to the *Hanwoo* bone infusions (Fig. 4A).

In this research, the infusions made from the *Hanwoo* parts such as the leg bone, foot and tail were added to skin fibroblast culture media in order to measure the effects of the *Hanwoo* bone infusions regarding the improvement of the elasticity of skin fibroblast and the inhibition of the skin aging. It was found that the aged cell number decreased as the proliferation of skin fibroblast and collagen synthesis increased due to culture treatment with the *Hanwoo* bone infusion in comparison to the control group. The skin fibroblast proliferation and the amount of the collagen synthesis increases depended on infusion concentrations. The highest levels of skin fibroblast collagen synthesis and anti-aging efficacy of skin fibroblast observed in the HLI, followed by the HFI and then the HTI. In accordance with the assumption that the main cause of this effect was the collagen from each infusion, skin fibroblast was treated only with bovine collagen and the skin fibroblast metabolism activation experiment was carried out. Unlike the assumption, treatment of the bovine collagen did not show skin fibroblast metabolic activation efficacy. This explains the result in which the collagen content of each infusion and activation of skin fibroblast metabolism did not correspond directly. When comparing collagen content per 1 mL of each infusion, the order of collagen content from the highest to lowest in each infusion was as follows: foot, leg bone, and tail (Fig. 2A). Research conducted by Zague (2008) confirmed that collagen is a major component in the anti-aging of the skin or skin elasticity. However, in the test confirming the collagen content of the *Hanwoo* bone infusion and each of their effects on skin fibroblast metabolic activation, it was observed that the collagen content of each infusion and their effects on the anti-aging of the skin and the skin elasticity does not correspond directly. The HFI contains a greater amount of collagen than the HLI, yet the effects on the activation of skin fibroblast metabolism are the greatest in the case of the HLI. Although the effect of the HFI was less than the effect of the HLI, there was no significant difference between the two. From such data, It may be speculated that collagen peptide type has an effect on the collagen function as well as the total collagen amount.

It is important to note that the effects of the *Hanwoo* bone infusion on skin fibroblast metabolism, rather than the effects of collagen itself, were observed. The other components of the *Hanwoo* bone infusion may have anti-aging effects on the skin and the skin elasticity in addition to collagen. In this study, the *Hanwoo* bone infusions pre-

pared through the traditional method were added to skin fibroblast culture media and the activation of the skin fibroblast metabolism was confirmed through scientific experimental methods. The significance of this research is that it confirmed the effects of the traditionally prepared infusions on skin anti-aging through scientific methods. This research suggests that *Hanwoo* bone infusions may be used in place of other collagen peptides that are used to activate skin fibroblast metabolism. Furthermore, this conclusion highlights the *Hanwoo* bone infusion as natural food that activates skin fibroblast metabolism through oral ingestion.

### Acknowledgements

This research was supported by the *Hanwoo Board* in year 2013.

### References

1. Ichikawa, S., Morifuji, M., Ohara, H., Matsumoto, H., Takeuchi, Y., and Sato, K. (2010) Hydroxyproline-containing dipeptides and tripeptides quantified at high concentration in human blood after oral administration of gelatin hydrolysate. *Int. J. Food Sci. Nutr.* **61**, 52-60.
2. Imokawa, G. (2008) Recent advances in characterizing biological mechanisms underlying UV-induced wrinkles: A pivotal role of fibroblast-derived elastase. *Arch. Dermatol. Res.* **300**, 7-20.
3. Joseph, S., Allen, R. G., Samuel, B., Arthur, B., and Vincent, J. C. (2000) Is  $\beta$ -galactosidase staining a marker of senescence *in vitro* and *in vivo*? *Exp. Cell Res.* **257**, 162-171.
4. Kim, E. J., Kim, M. K., Jin, X. J., Oh, J. H., Kim, J. E., and Chung, J. H. (2010) Skin aging and photoaging alter fatty acids composition, including 11,14,17-eicosatrienoic acid, in the epidermis of human skin. *J. Korean Med. Sci.* **25**, 980-983.
5. Kim, K. S. (2003) Review of health functional ingredient in food. *Pusan Women's Univ. J.* **25**, 79-94.
6. Koji, I., Takanori, H., Yasuki, T., Fumiki, M., Kenji, S., Yasushi, N., Akane, H., Yasuhiro, K., Yukihiko, N., and Kozo, O. (2005) Identification of food-derived collagen peptides in human blood after oral ingestion of gelatin hydrolysates. *J. Agric. Food Chem.* **53**, 6531-6536.
7. Makrantonaki, E. and Zouboulis, C. C. (2007) Molecular mechanisms of skin aging state of the art. *Ann. N Y Acad. Sci.* **1119**, 40-50.
8. Matsuda, N., Koyama, Y. I., Hosaka, Y., Ueda, H., Watanabe, T., Araya, T., Irie, S., and Takehana, K. (2006) Effects of ingestion of collagen peptide on collagen fibrils and glycosaminoglycans in the dermis. *J. Nutr. Sci. Vitaminol.* **52**, 211-215.
9. Park, D. Y. (1986) Total nitrogen and free amino acid contents in shank bone stock according to boiling time. *J. Korean Soc. Food Nutr.* **15**, 243-248.

10. Park, D. Y. and Lee, Y. S. (1982) An experiment in extracting efficient nutrients from sagol bone stock. *J. Korean Nutr. Food* **11**, 47-52.
11. Ronald, K. G. and Susan, M. K. (2005) Quantitative assay of senescence-associated-galactosidase activity in mammalian cell extracts. *Analyt. Biochem.* **343**, 329-334.
12. Saito, M. and Marumo, K. (2010) Collagen cross-links as a determinant of bone quality: a possible explanation for bone fragility in aging, osteoporosis, and diabetes mellitus. *Osteoporos Int.* **21**, 195-214.
13. Salasznyk, R. M., Williams, W. A., Boskey, A., Batorsky, A., and Plopper, G. E. (2004) Adhesion to vitronectin and collagen I promotes osteogenic differentiation of human mesenchymal stem cells. *J. Biomed. Biotechnol.* **1**, 24-34.
14. Schwartz, S. R. and Park, J. S. (2012) Ingestion of BioCell Collagen®, a novel hydrolyzed chicken sternal cartilage extract; enhanced blood microcirculation and reduced facial aging signs. *Clin. Interv. Aging* **7**, 267-273.
15. Seol, M. Y. and Jang, M. S. (1990) A study on mineral contents in sagol bone stock. *Korean J. Food Cookery Sci.* **6**, 21-26.
16. Shizuka, Y., Yoshizawa, Y., Kawakubo, A., Ikeda, T., Yanagiguchi, K., and Hayashi, Y. (2013) Early gene and protein expression associated with osteoblast differentiation in response to fish collagen peptides powder. *Dental Mater. J.* **32**, 233-240.
17. Sugihara, F., Inoue, N., Kuwamori, M., and Taniguchi, M. (2012) Quantification of hydroxyprolyl-glycine(Hyp-Gly) in human blood after ingestion of collagen hydrolysate. *J. Bio-sci. Bioeng.* **113**, 202-203.
18. Talwar, J. S., Griffiths, C. E. M., Fisher, G. J., Hamilton, T. A., and Vorhees, J. J. (1995) Reduced type I and type III pro-collagens in photodamaged adult human skin. *J. Invest. Dermatol.* **105**, 285-290.
19. Vivian, Z., Vanessa, F., Marina, C. R., Geórgia Álvares, C., Ruy, G. J., and Gláucia, M. M. S. (2011) Collagen hydrolysate intake increases skin collagen expression and suppresses matrix metalloproteinase 2 activity. *J. Medicinal Food* **14**, 618-624.
20. Yasutaka, S., Koji, I., Fumiki, M., Takaaki, I., Toshio, M., Chikako, O., Toshio, T., Park, E. Y., Nakamura, Y., and Sato, K. (2009) Effect of prolyl-hydroxyproline (Pro-Hyp), a food-derived collagen peptide in human blood, on growth of fibroblast from mouse skin. *J. Agric. Chem.* **57**, 444-449.
21. Yoon, J. Y., Choi, S. Y., Jeong, H. S., Park, Y. I., Kim, D. S., and Joo, N. M. (2015) A comparative study on quality and physicochemical characteristics of segmental bone Korean beef infusion. *Korean J. Food Nutr.* **28**, 470-477.
22. Zague, V. (2008) A new view concerning the effects of collagen hydrolysate intake on skin properties. *Arch. Dermatol. Res.* **300**, 479-483.