

Homophilic Binding of PTP μ , a Receptor-Type Protein Tyrosine Phosphatase, Can Mediate Cell-Cell Aggregation

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Abstract. The receptor-like protein tyrosine phosphatase, PTP μ , displays structural similarity to cell-cell adhesion molecules of the immunoglobulin superfamily. We have investigated the ability of human PTP μ to function in such a capacity. Expression of PTP μ , with or without the PTPase domains, by recombinant baculovirus infection of Sf9 cells induced their aggregation. However, neither a chimeric form of PTP μ , containing the extracellular and transmembrane segments of the EGF receptor and the intracellular segment of PTP μ , nor the intracellular segment of PTP μ expressed as a soluble protein induced aggregation. PTP μ mediates aggregation via a homophilic mechanism, as judged by lack of incorporation of uninfected Sf9 cells into aggregates of PTP μ -expressing cells.

Homophilic binding has been demonstrated between PTP μ -coated fluorescent beads (Covaspheres) and endogenously expressed PTP μ on MvLu cells. Additionally the PTP μ -coated beads specifically bound to a bacterially expressed glutathione-S-transferase fusion protein containing the extracellular segment of PTP μ (GST/PTP μ) adsorbed to petri dishes. Covaspheres coated with the GST/PTP μ fusion protein aggregated in vitro and also bound to PTP μ expressed endogenously on MvLu cells. These results suggest that the ligand for this transmembrane PTPase is another PTP μ molecule on an adjacent cell. Thus homophilic binding interactions may be an important component of the function of PTP μ in vivo.

THE reversible phosphorylation of tyrosyl residues in proteins is an essential component of the modulation of signal transduction processes involved in cell growth and differentiation. The dynamic balance of cellular phosphotyrosine levels is achieved by the opposing actions of the protein tyrosine kinases (PTKs)¹ and protein tyrosine phosphatases (PTPases). The PTKs are a structurally diverse group that includes growth factor receptors and a substantial number of oncogene products, including members of the *src* family. It is now apparent that PTPases rival the PTKs in structural diversity and complexity. PTPases have the potential to oppose the effects of kinases as illustrated by their ability to suppress the transforming potential of oncogenic PTKs (Brown-Shimer et al., 1992; Woodford-Thomas et al., 1992; Zander et al., 1993). However, the PTPases should not be viewed simply as PTK antagonists. Overexpression of PTP α leads to dephosphorylation and activation of *c-src* and induces cell transformation (Zheng et al., 1992). In addition, CD45 has been shown to play an essential role in signal transduction through the T cell receptor, also potentially exerting its effect through the dephosphorylation and activation of *src*-family PTKs such as *lck* and *fyn* (reviewed

in Trowbridge, 1991). Current evidence clearly indicates an essential role for the PTPases in cellular signaling.

PTPases exist in both soluble and transmembrane, receptor-like forms, (Charbonneau and Tonks, 1992). The structural features of some of the enzymes suggest a role for subcellular localization in the regulation of activity. In the case of the receptor PTPases, similar to the receptor PTKs, there is the potential for regulating activity by the binding of ligands to the extracellular segment of the protein. Several of the transmembrane PTPases are members of the immunoglobulin superfamily and display structural motifs in their extracellular segments that are suggestive of a role in cell-cell adhesion (Streuli et al., 1989; Fischer et al., 1991).

Cell-cell adhesion molecules are grouped into two major families on the basis of homology and conditions for binding: the immunoglobulin superfamily (generally calcium-independent) and the cadherin family (calcium-dependent). N-CAM is prototypical of the immunoglobulin superfamily of adhesion molecules. It contains five immunoglobulin domains and two fibronectin type-III repeats in the extracellular segment (reviewed by Edelman and Crossin, 1991). Cadherins are calcium-dependent cell-cell adhesion molecules that associate with the actin cytoskeleton by interactions of their cytoplasmic domains with proteins termed catenins (Ozawa et al., 1989). Both N-CAM and the cadherins bind by a homophilic mechanism (Edelman et al., 1987; Hall et al., 1990; Rao et al., 1992; Takeichi, 1991).

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1. *Abbreviations used in this paper:* GST, glutathione S-transferase; PTK, protein tyrosine kinase; PTPases, protein tyrosine phosphatases.

The receptor-type PTPase, PTP μ , shows homology to N-CAM in that it bears an extracellular segment with an immunoglobulin domain and four fibronectin type III repeats (see Fig. 1) (Gebbinck et al., 1991). This arrangement of multiple Ig-domains and fibronectin type-III repeats was first observed in LAR (Streuli et al., 1988) and has also been seen in the extracellular segments of several other transmembrane PTPases (reviewed in Charbonneau and Tonks, 1992). In this manuscript we investigate whether PTP μ can participate in homophilic binding interactions.

Introduction of potential adhesion molecules into non-adhesive *Drosophila* S2 insect cells has been used to demonstrate adhesive functions for fasciclin III, connectin and *Dtrk* molecules directly (Snow et al., 1989; Nose et al., 1992; Pulido et al., 1992). In a similar approach, we demonstrate that the full-length form of PTP μ induced aggregation, via a homophilic binding mechanism, when expressed in non-adhesive Sf9 insect cells, which are derived from the Fall armyworm *Spodoptera frugiperda*. Cells expressing mutant forms of the enzyme lacking the natural extracellular segment did not aggregate. In this system, PTPase activity and the aggregation response are not mutually dependent. We have also reconstituted the binding reaction in vitro between baculovirus-expressed PTP μ linked to beads and surfaces coated with bacterially-expressed extracellular segment of PTP μ . Finally we show that endogenously expressed PTP μ in lung cells binds homophilically to baculovirus-expressed PTP μ . These data represent the first demonstration of homophilic binding and a potential adhesive function for a member of the PTPase family of enzymes.

Materials and Methods

Cell Culture and PTPase Activity

Sf9 cells derived from the ovary of the Fall armyworm *Spodoptera frugiperda* (American Type Culture Collection [ATCC] number CRL 1711) were maintained at 27°C in Grace's Insect Medium Supplemented (GIBCO-BRL, Gaithersburg, MD) containing 10% FBS and 10 μ g/ml gentamicin (GIBCO-BRL). MvLu and 3T3 cells (ATCC numbers CCL 64 and CCL 92, respectively) were grown at 37°C, 5% CO₂ in DME containing penicillin and streptomycin plus 10% FBS (GIBCO-BRL).

PTPase assays were performed as described (Flint et al., 1993). Total cell lysates were prepared in detergent containing buffer (20 mM Tris pH 7.6, 5 mM EDTA, 1% Triton X-100, 5 μ g/ml leupeptin, 5 μ g/ml aprotinin, 1 mM benzamide, 1 mM PMSF, and 1 mM DTT). PTPase activity was measured in vitro by dephosphorylation of tyrosine-phosphorylated reduced carboxamidomethylated and maleylated (RCM) lysozyme. 1 U is defined as the amount of enzyme that catalyzes the release of 1 nmol phosphate from the substrate per min.

Plasmid and Baculovirus Construction

The soluble form (80 kD) and the full-length form (200 kD) of human PTP μ were expressed in Sf9 cells infected with the Ac-pS and Ac-pFL baculoviruses, respectively. The soluble form (Ac-pS) contains the DNA sequence coding for the intracellular domain of PTP μ (amino acids 774-1452). pS was generated by digesting bluescript TR-PTP μ (Gebbinck et al., 1991) with BamHI and XbaI and ligating the fragment into pVL1392 (Invitrogen Corp., San Diego, CA). The full-length form of PTP μ was cloned out of bluescript containing the full-length cDNA of PTP μ using the NotI and XbaI sites and ligated into pVL1392 (PTP μ cDNA was provided by M. Gebbinck and W. Moolenaar, Netherlands Cancer Institute). A chimeric form of PTP μ , pEGFR/PTP μ (provided by R. Beijersbergen, Netherlands Cancer Institute) was generated that contained the extracellular and transmembrane domains of the EGF receptor (673 amino acids; base pair 170-2190) and the intracellular domain of PTP μ (678 amino acids; base pair 2320-4865). These three viruses were generated by calcium phosphate

mediated cotransfection of Sf9 cells with plasmid and viral DNA. Recombinant viruses were isolated as described in Summers and Smith (1987). A mutant form of PTP μ was generated that contained the extracellular domain, the transmembrane domain and 54 amino acids of the intracellular domain (Ac-PTP μ -extra). This mutant form lacks both phosphatase domains and was called PTP μ -extra. This form was cloned into the pVL1393 plasmid (Invitrogen Corp.) using a Not I site from the full length cDNA in bluescript and the Bgl II site of the PTP μ intracellular domain. The recombinant virus was made using the BaculoGold Transfection System (Invitrogen Corp.). A 2.2-kb fragment encoding the 79 kD extracellular segment of PTP μ (amino acids 21-740) was generated using oligonucleotides and PCR and ligated into the pGEX-KG vector (provided by K. Guan and J. Dixon, University of Michigan, Ann Arbor, MI) to produce a GST fusion protein. This construct was designated GST/PTP μ . The BIRK baculovirus has been described (Villalba et al., 1989). The tropomyosin baculovirus was provided by M. Pittenger and D. Helfman (Cold Spring Harbor Laboratory).

Antibodies

For isolation of the soluble (80 kD) form of PTP μ , Sf9 cells were infected with Ac-pS recombinant baculovirus for 4 d. Soluble PTP μ was purified from the cytoplasmic fraction of Sf9 cell lysates by sequential application to fast flow Q Sepharose, Mono Q, and Mono S as described (Brady-Kalnay and Tonks, 1993). The purified protein was used to generate six mAbs. Each of the mAbs was characterized in terms of binding to PTP μ ; none recognized any proteins in extracts from uninfected Sf9 cells (manuscript in preparation). All of the monoclonals recognized the full-length and soluble PTP μ proteins from baculovirus-infected Sf9 cells and COS cells transiently transfected with PTP μ . One of the mAbs (SBK-15) was used for coupling PTP μ to Covaspheres and another antibody (SBK-10) was used in immunoblotting. A rabbit polyclonal antipeptide antibody to the amino terminus of PTP μ (residues 42-57, provided by M. Gebbinck) was used for detection of PTP μ in immunoblots, coating of the MvLu cells and Fab' production. Fab' fragments of this antibody were generated according to Brackenbury et al. (1977). Monoclonal anti-phosphotyrosine antibody (4G10) was purchased from Upstate Biotechnology Inc. (catalog no. 05-321; Lake Placid, NY) and used at 1:1,000 dilution.

Electrophoresis and Immunoblotting

Sf9 cells were infected with Ac-pS, Ac-pFL, Ac-pEGFR/PTP μ , or Ac-PTP μ -extra recombinant baculoviruses to express the various forms of PTP μ . For analysis of expression of the various proteins, cells were harvested 42 h post-infection by centrifugation for 5 min at 3,000 g, and lysed in 1 ml of buffer (20 mM Tris pH 7.6, 5 mM EDTA, 1% Triton X-100, 5 μ g/ml leupeptin, 5 μ g/ml aprotinin, 1 mM benzamide, 1 mM PMSF, 200 μ M phenyl arsine oxide, 1 mM vanadate and 0.1 mM molybdate). After incubation on ice for 30 min, the lysate was centrifuged at 10,000 g for 10 min and the supernatant was used for immunoblotting. The amount of protein was determined by the Bradford method using BSA as a standard (Bradford, 1976). 4 \times sample buffer was added to the lysates and they were separated by electrophoresis on 8% SDS polyacrylamide gels (Laemmli, 1970). Proteins were transferred to nitrocellulose (Towbin, 1979), blocked in 5% nonfat dry milk in TTBS, incubated with mAb (SBK-10) or polyclonal amino-terminal anti-peptide antibody against PTP μ . A secondary antibody conjugated to HRP was added and the bound antibody was visualized by enhanced chemiluminescence (ECL, Amersham Corp., Arlington Heights, IL).

Cell surface expression of the various forms of PTP μ was verified by examining their protease sensitivity on intact cells. Sf9 cells were removed from a T25 flask in 2 mls Grace's medium in the presence or absence of 0.05% trypsin (wt/vol) (Worthington Biochemical Corp., Freehold, NJ) and incubated for 15 min at room temperature. The cells were centrifuged, extracted and immunoblotted as described above except that lima bean trypsin inhibitor (Worthington Biochemical Corp., Freehold, NJ) was added to the lysis buffer (60 μ g/ml).

Phosphotyrosine levels were compared by reactivity to an anti-phosphotyrosine antibody on immunoblots. Co-infections were performed by simultaneously adding both viruses to Sf9 cells and incubating for 42 h. The immunoblots were performed as described above.

Cell Aggregation Assays

Sf9 adhesion assays were similar to those performed in Snow et al. (1989).

The quantitation of the extent of aggregation followed the procedure of Brackenbury et al. (1977). Sf9 cells were harvested 2 or 3 d post-infection. 1×10^6 cells were added to 2 mls Grace's medium containing 150 $\mu\text{g/ml}$ DNase I in a glass scintillation vial (Fisher Scientific, Pittsburgh, PA) and incubated at 25°C at 90–100 rpm in a gyratory shaker. Aliquots were diluted 50-fold and the number of particles was determined using a Coulter Counter. The Coulter Counter settings were: lower threshold–5.0, upper threshold–99.9, current–500 mA, full scale–1, polarity–auto, attenuation–16, and preset gain–1. The percent aggregation was calculated by subtracting the particle number after the 30 min or 1 h time point (N_t) from the initial particle number (N_0) and dividing by the initial number $\{[(N_0 - N_t)/N_0] \times 100\}$. Aggregates were visualized with a phase contrast microscope using the 10 \times objective.

Fluorescent labeling of cells using Di I was performed according to the protocol supplied by Molecular Probes Inc. (Eugene, Oregon). Sf9 cells were incubated with 20 μM Di I for 15 min at room temperature, then washed two times in Grace's medium. Di I labeled, uninfected Sf9 cells were mixed with an equal number of cells expressing full-length PTP μ and aggregation assays were performed as described above. Aliquots were spotted onto microscope slides and Di I labeled cells were observed with a Zeiss Axiophot microscope equipped for epifluorescence using a 20 \times lens.

In a subset of assays, a chelating resin, iminodiacetic acid cross-linked to polystyrene (Sigma Chemical Co., St. Louis, MO), was used to remove divalent cations from the insect cell medium. One ml swollen resin was used to deplete 3 mls media by mixing for 1 h at room temperature. The effect of this procedure was assessed in aggregation assays as described above.

Preparation of Covaspheres

MX Covaspheres (0.9 μm) were purchased from Duke Scientific (Palo Alto, CA). After brief sonication 100 μl of Covaspheres were added to 100 μg of mAb-SBK15. The protein and the Covaspheres were incubated for 75 min at room temperature then centrifuged at 10,000 g for 2 min. Any unbound sites were blocked by incubation for 15 min in 20 mM Tris pH 7.3 containing 1% BSA. The beads were washed with PBS and briefly sonicated before use in immunoprecipitation of full length PTP μ or the EGFR/PTP μ chimera. A triton soluble lysate from full length PTP μ or EGFR/PTP μ chimera-expressing Sf9 cells was added to the SBK15-Covaspheres, incubated at 4°C for 1 h, washed with lysis buffer three times, resuspended in PBS and used in binding assays. Successful immunoprecipitation by the antibody-linked Covaspheres was verified by depletion of the PTP μ reactivity upon immunoblotting. This antibody has been used to purify full-length PTP μ from Sf9 cells (Brady-Kalnay and Tonks, 1993).

The in vitro binding assays also utilized a purified fusion protein, com-

prising glutathione S-transferase linked to the NH $_2$ -terminus of the extracellular segment of PTP μ (GST/PTP μ). The GST/PTP μ fusion protein was expressed in *E. coli* and purified according to Guan and Dixon (1991). The GST protein alone (27 kD) was used as a negative control. These GST proteins were coupled directly to Covaspheres by adding 3.9 μg protein per μl of Covaspheres as described above.

Covasphere Binding Assays

The binding assay was performed as described in Mauro et al. (1992). Briefly, 10 μg of purified GST proteins were adsorbed to 35-mm petri dishes for 30 min, then remaining unbound sites were blocked with 2% BSA in PBS. PTP μ -linked Covaspheres (50 μl) were added to the dishes in a final volume of 1 ml PBS. Bound Covaspheres were visualized with a Zeiss Axiophot microscope equipped for epifluorescence using a 20 \times lens.

The ability of GST/PTP μ -linked Covaspheres to self-aggregate was assessed by adding 30 μl of the Covasphere preparation to 2 ml of PBS followed by rotation for 30 min under low shear conditions. Controls utilizing Covaspheres coated with GST alone were performed in parallel. Aggregates were visualized with a Zeiss Axiophot microscope as above.

Various protein-coated Covaspheres described above were also used in assays to assess binding to the surface of MvLu mink lung cells or 3T3 cells. Covaspheres (50 μl) were added in DME plus 10% FBS to confluent 35-mm plates of MvLu cells, then incubated with the cells for 30 min at room temperature with rocking. The plates were washed 3 times with DME and visualized as described above. All antibody competition experiments were performed using the anti-peptide antibody to the amino-terminus of the extracellular segment of PTP μ . In experiments in which binding was blocked with Fab' fragments the Covaspheres were preincubated for 30 min at room temperature in 0.6 mg pre-immune or immune Fab' per 50 μl Covaspheres in a final volume of 500 μl . The Fab'/Covasphere solution was added to the cells and incubated for 30 min. Alternatively, the MvLu cells were incubated for 30 min with pre-immune or immune serum to PTP μ , diluted 1:500 in culture media, before the addition of 50 μl of full-length PTP μ -linked Covaspheres.

Results

PTP μ Is Expressed at the Cell Surface

Human PTP μ was expressed in Sf9 insect cells by infection with recombinant baculoviruses. Three forms of human PTP μ were expressed: (a) a full-length form (200 kD); (b) a soluble form, comprising only the intracellular segment

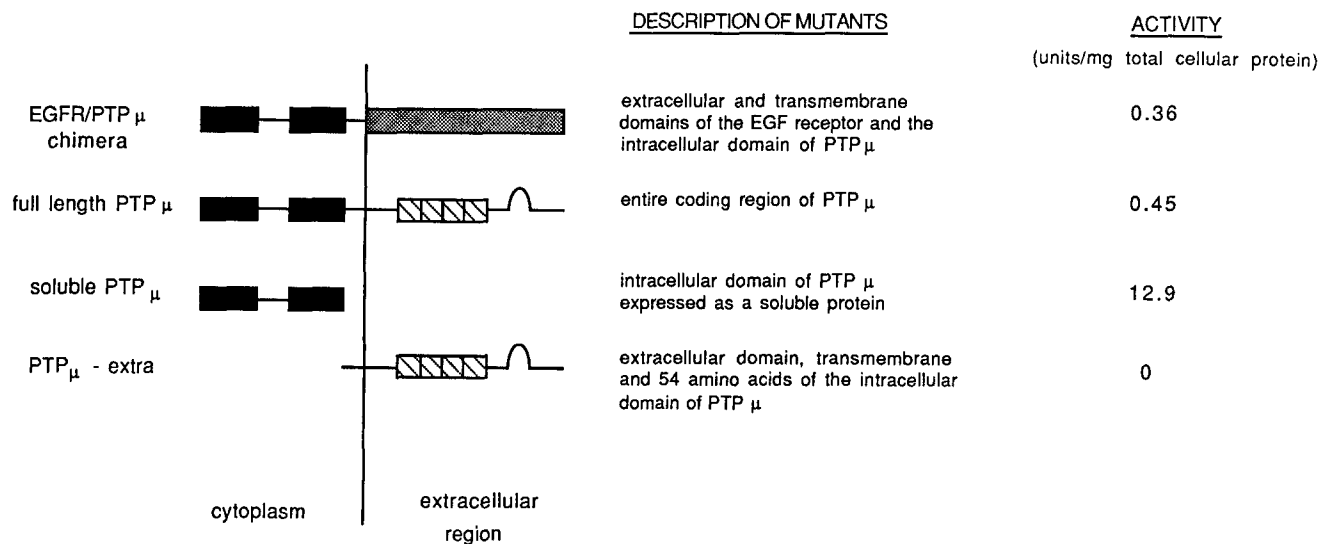
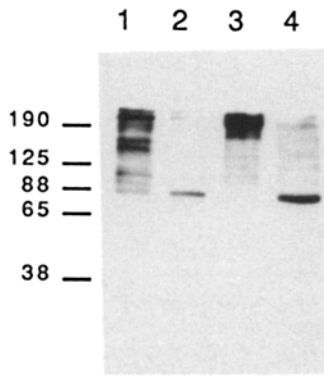


Figure 1. Structure and PTPase activity of the various baculovirus-expressed recombinant forms of PTP μ . The solid black boxes represent PTPase domains. The hatched boxes are fibronectin type-III repeats and the loop represents the immunoglobulin-like domain. The stippled box represents the extracellular domain of the EGF receptor. The PTPase activity is expressed as units per mg of total cellular protein above the PTPase activity of lysates from uninfected Sf9 cells which was 0.09 U/mg.

Figure 2. Expression of PTP μ and the EGFR/PTP μ chimera at the cell surface. This figure is an immunoblot of whole cell lysates using mAb SBK-10 generated against the intracellular segment of PTP μ . Lane 1 is from cells expressing full-length PTP μ , Lane 2 is from PTP μ -expressing cells that were subjected to limited trypsin treatment. Extracts from EGFR/PTP μ chimera-infected cells without or with trypsin treatment are shown in lanes 3 and 4, respectively. Each lane contains 5 μ g total protein.



containing the PTPase domains (80 kD); and (c) a chimera containing the extracellular and transmembrane domains of the EGF receptor and the intracellular PTPase domains of PTP μ (195 kD) (see Fig. 1). The activity of these PTPases in lysates of infected cells was measured using tyrosine-phosphorylated RCM lysozyme as a substrate (Fig. 1). The uninfected Sf9 cells displayed low levels of endogenous PTPase activity (0.09 U/mg total protein). The soluble form of PTP μ was expressed at much higher levels (\sim 140-fold over background) than either of the transmembrane forms (\sim fivefold over the endogenous activity) (see Fig. 1). The expression of the various forms of PTP μ was also assessed by immunoblotting with antibodies to its intracellular segment. Fig. 2 shows that the full-length form of PTP μ (lane 1) and the EGFR/PTP μ chimera (lane 3) are expressed at similar levels. Immunoblotting of Sf9 cell lysates expressing the

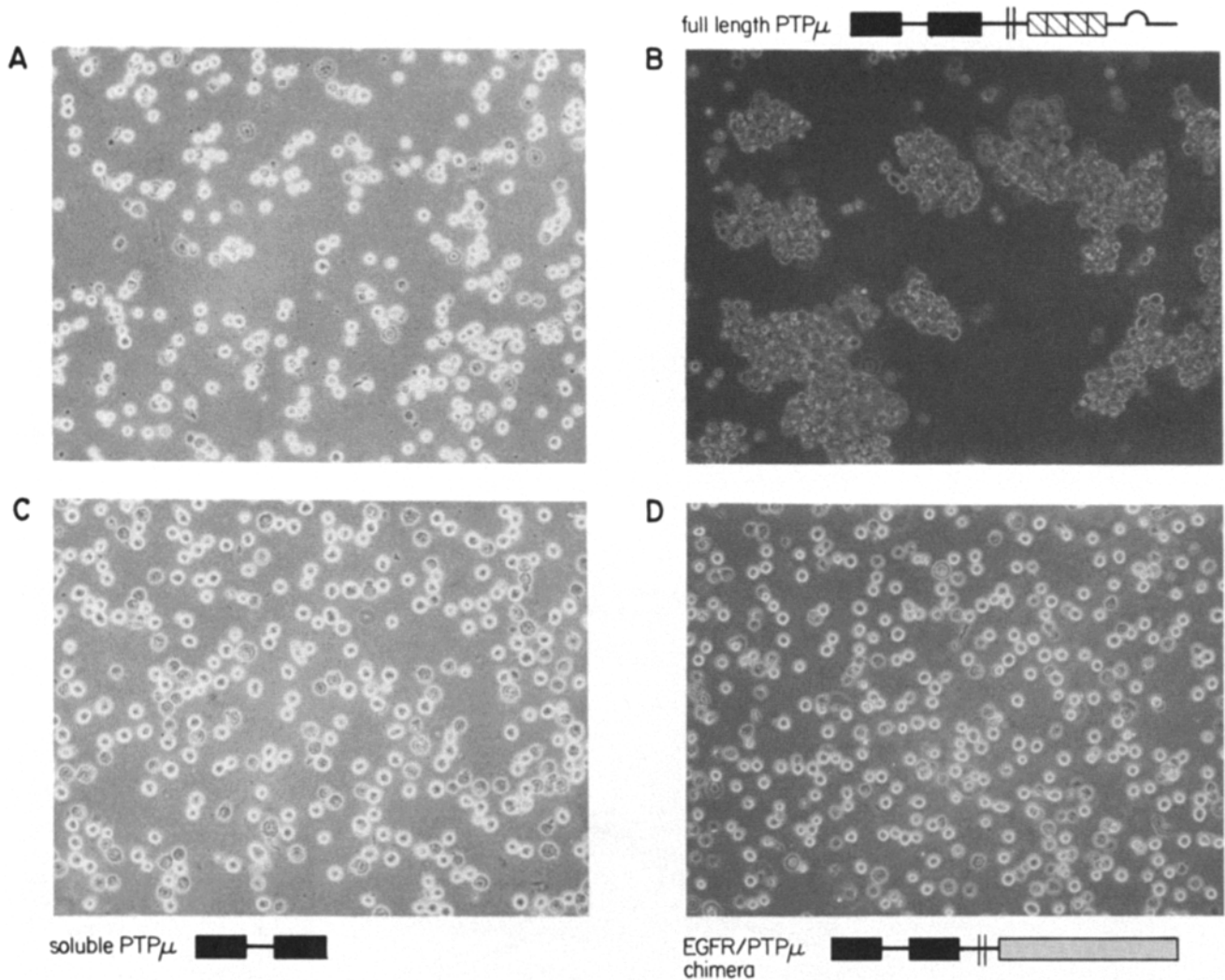


Figure 3. Aggregation of baculovirus-infected Sf9 cells. This figure shows phase contrast micrographs of uninfected or infected Sf9 cells allowed to aggregate under low shear conditions for 1 h. *a* shows uninfected cells which did not aggregate while the full-length PTP μ infected cells (*b*), formed large aggregates. Cells infected with recombinant baculoviruses expressing soluble PTP μ (*c*) or EGFR/PTP μ chimera (*d*) did not aggregate.

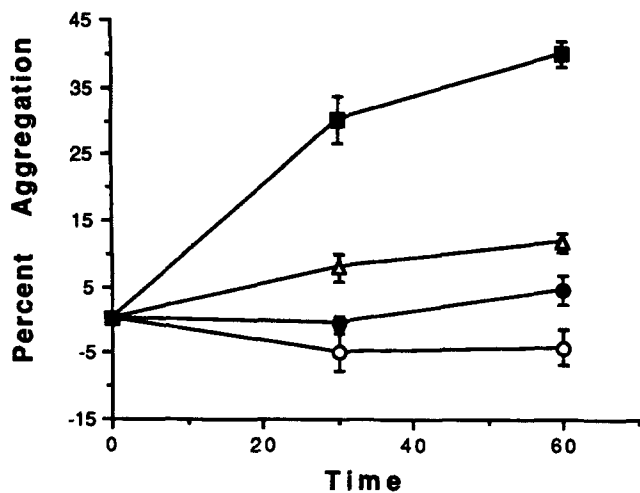


Figure 4. Quantitation of aggregation. A Coulter counter is used to measure a decrease in particle number as single cells are incorporated into aggregates (*percent aggregation*). Full-length PTP μ -infected cells are represented by the solid squares. Soluble PTP μ (solid circles) or EGFR/PTP μ chimera (open circles)-expressing cells showed even lower levels of aggregation than the uninfected cells (open triangles). Error bars indicate SEM from at least five determinations.

soluble form of PTP μ confirmed that it is present at much higher levels than the transmembrane forms.

To verify that PTP μ and the EGFR/PTP μ chimera were expressed at the cell surface, trypsin sensitivity assays were performed (Brady-Kalnay et al., 1993). This assay involves incubation of intact cells with a 0.05% (wt/vol) trypsin solution which allows the protease to act only on the portion of molecules expressed at the cell surface. Full-length forms are converted into smaller fragments still associated with the cell, i.e., possessing their transmembrane and cytoplasmic segments. The viability and integrity of the trypsinized cells was confirmed by trypan blue dye exclusion. Lysates of treated cells were immunoblotted with monoclonal antibody SBK-10 directed against the carboxy-terminal domain of PTP μ , which thus reacts with both full-length PTP μ and the EGFR/PTP μ chimera. Fig. 2 shows ~90% of full-length PTP μ (lane 2) and the EGFR/PTP μ chimera (lane 4) were degraded after trypsinization, thus suggesting that these proteins are expressed predominantly on the cell surface.

Full Length PTP μ Expressed in Sf9 Cells Mediates Aggregation

We have used the nonadhesive Sf9 insect cells and the baculovirus expression system to demonstrate that the protein tyrosine phosphatase PTP μ functions in cell-cell aggregation. The infected cells were tested in aggregation assays which involve the rotation of cells under low shear conditions (Brackenbury et al., 1977). Samples are counted at various time points to measure the incorporation of single cells into aggregates.

Sf9 cells were infected with recombinant baculoviruses, harvested and assayed for aggregation. The results of a visual inspection of the cells is illustrated in the phase contrast micrographs in Fig. 3. While uninfected cells do not aggregate (*a*) expression of the full-length form of PTP μ , shown in *b*, induces the formation of large aggregates. Cells expressing the soluble PTP μ (*c*) or the EGFR/PTP μ chimera (*d*) also did not aggregate suggesting that increased phosphatase activity alone was not sufficient for this response. Neither overexpression of a nonspecific transmembrane protein nor the localization of PTPase activity at the membrane was sufficient to induce aggregation.

Quantitative analysis of the aggregation of Sf9 cells is displayed in Fig. 4. The quantitation, measured using a Coulter Counter, is expressed as percent aggregation which is a function of decrease in particle number as single cells are incorporated into multicellular aggregates. Uninfected cells display low levels of aggregation (mean = 11.7%) whereas full-length PTP μ -expressing cells showed high levels of aggregation (mean = 39.8%). Cells expressing soluble PTP μ (mean = 4.6%) or the EGFR/PTP μ chimera (mean = -4.1%) showed low levels of aggregation. The negative number indicates that the final cell count was higher than the initial cell count due to the dissociation of cells during the aggregation assay. Control infected cells showed lower levels of aggregation than uninfected Sf9 cells.

Adhesion molecules are classified by their dependence on divalent cations, primarily calcium (Takeichi, 1977). The aggregation assay described above was routinely performed in Grace's insect cell medium which contains 7 mM calcium chloride. To test whether divalent cations were important for aggregation, we used a chelating resin to remove them from the medium prior to performing the assay. The full-length PTP μ -expressing cells aggregated equally well with or without divalent cations (data not shown) suggesting that, like

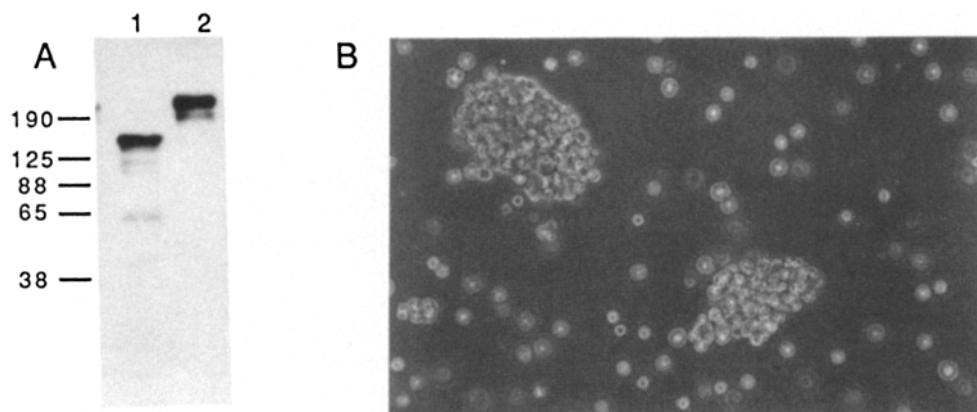


Figure 5. Aggregation of PTP μ -extra expressing cells. In *A*, the immunoblot shows that PTP μ -extra (lane 1) and full length PTP μ (lane 2) are expressed at similar levels. The anti-peptide antibody used in this immunoblot recognizes a sequence (residues 42-57) at the amino-terminus of PTP μ . Each lane contains 5 μ g protein. *B* is a phase contrast micrograph of PTP μ -extra expressing cells after a 1 h aggregation assay.

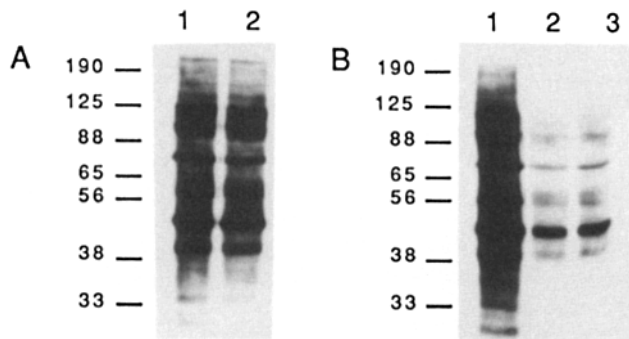


Figure 6. Analysis of phosphotyrosine levels of infected Sf9 cells. The BIRK (tyrosine kinase)-infected (*a*, lane 1, and *b*, lane 1) or BIRK-tropomyosin co-infected (*a*, lane 2) cells showed high levels of anti-phosphotyrosine immunoreactivity. Co-infection of BIRK and full length PTP μ produced a reduction in signal that was unchanged before (*b*, lane 2) and after aggregation of the cells (*b*, lane 3). The same amount of recombinant PTP μ baculovirus was added in single or co-infection experiments. Each lane contains 30 μ g protein.

other members of the Ig-superfamily of adhesion molecules, it functions in a divalent cation-independent manner.

Phosphatase Activity Is not Required for Aggregation

To test whether phosphatase activity is required for the adhesive function, we generated a mutant form of PTP μ containing the entire extracellular domain, transmembrane domain and 54 amino acids of the intracellular domain, which we termed PTP μ -extra. This protein lacks both phosphatase domains. As expected, in lysates of cells expressing PTP μ -extra, no PTPase activity above that found in uninfected cells was detected. An immunoblot using an antipeptide antibody directed against the NH₂-terminal extracellular segment of PTP μ (Fig. 5 *a*) illustrates that PTP μ -extra (lane 1) is expressed at similar levels to the full-length protein (lane 2). The ability of PTP μ -extra expressing cells to aggregate is shown in Fig. 5 *b*. Quantitative analysis of the assays indicated a percent aggregation of the PTP μ -extra infected cells of 30.3% \pm 4.3 (mean \pm SEM, *n* = 8), compared to 38.2% \pm 1.9 (mean \pm SEM, *n* = 6) for full-length PTP μ -infected cells. Thus, the adhesive function of PTP μ and its capacity to dephosphorylate proteins are not mutually dependent.

A common feature of receptor-like molecules is that the enzymatic activity of their intracellular domains is regulated by binding of ligands to their extracellular segments. We thus sought to examine whether aggregation might affect the activity of PTP μ . To this end, the levels of total cellular phosphotyrosine were analyzed. Sf9 cells contain very low levels of phosphotyrosine-containing proteins. To increase the amount of cellular phosphotyrosine and the variety of potential substrates, we infected the cells with a recombinant baculovirus expressing BIRK, a soluble protein comprising the catalytic domain of the mammalian insulin receptor tyrosine kinase (Villalba et al., 1989). There are many examples of direct protein:protein and enzyme:substrate interactions that have been investigated by co-infection of Sf9 cells with distinct recombinant baculoviruses (Kaplan et al., 1990; Parker et al., 1991; Kato et al., 1993). In light of these reports, we co-infected two batches of Sf9 cells. One batch

was co-infected with baculoviruses expressing BIRK and a nonphosphatase control (tropomyosin) and the other batch with BIRK and full-length PTP μ . Cell lysates were then analyzed for changes in phosphotyrosine levels. Fig. 6 is an immunoblot, using anti-phosphotyrosine antibodies, of extracts from cells infected with BIRK (Fig. 6 *a*, lane 1) or co-infected with BIRK and tropomyosin (*a*, lane 2). The co-infection with the non-phosphatase control did not alter the BIRK-induced increase in phosphotyrosine levels. The presence of tropomyosin protein was confirmed by immunoblotting (data not shown). Fig. 6 *b* shows that while high levels of phosphotyrosine containing proteins were seen in BIRK-infected cells (lane 1) co-infection with BIRK and full-length PTP μ (lane 2) resulted in a dramatic decrease in phosphotyrosine levels, presumably due to high basal activity of the PTPase. Immunoblotting verified that full-length PTP μ was expressed at similar levels in the single or co-infected Sf9 cell lysates. The overall levels and pattern of anti-phosphotyrosine immunoreactive proteins were unchanged by aggregation. BIRK/full-length PTP μ expressing cells before (lane 2) and after (lane 3) aggregation are shown in Fig. 6 *b*. Co-infection of BIRK and the soluble PTP μ , which is expressed at \sim 25-fold higher levels than the transmembrane forms, as determined by activity assays, contained even lower levels of anti-phosphotyrosine reactivity, suggesting that the immunoreactive bands were indeed tyrosine phosphorylated proteins. In addition to this *in vivo* assay, total cell lysates of full-length PTP μ -infected cells were assayed for PTPase activity with RCM lysozyme as a substrate, before and after aggregation. The activity was essentially unchanged after aggregation.

PTP μ Mediates Homophilic Aggregation

We have tested whether the aggregation of Sf9 cells expressing PTP μ is mediated via a homophilic mechanism in which molecules of PTP μ on different cells interact with one another or by a heterophilic mechanism in which the insect cells express a molecule that binds to human PTP μ . Uninfected nonadhesive Sf9 cells were labeled with the fluorescent lipophilic dye, Di I (Snow et al., 1989) and mixed with unlabeled full-length PTP μ -infected cells. After aggregation the cells were visualized using an epifluorescence microscope. The Di I labeled cells did not aggregate and did not contribute to the aggregates formed by the full-length PTP μ -infected cells. Fig. 7 is a representative example of 32 aggregates examined in three independent experiments; no Di I-positive cells were seen in any of the aggregates. These results strongly suggest that PTP μ binds in a homophilic fashion.

Reconstitution of PTP μ -mediated Homophilic Binding *In Vitro*

We have examined the ability of baculovirus-expressed PTP μ to bind to surfaces coated with bacterially-expressed extracellular segment of PTP μ *in vitro*. A mAb to the intracellular domain of PTP μ was covalently linked to Covaspheres (fluorescent beads). These antibody-linked Covaspheres were used to immunoprecipitate either the full length PTP μ or the EGFR/PTP μ chimera from lysates of infected cells. The entire extracellular domain of PTP μ was expressed as a glutathione S-transferase (GST) fusion protein

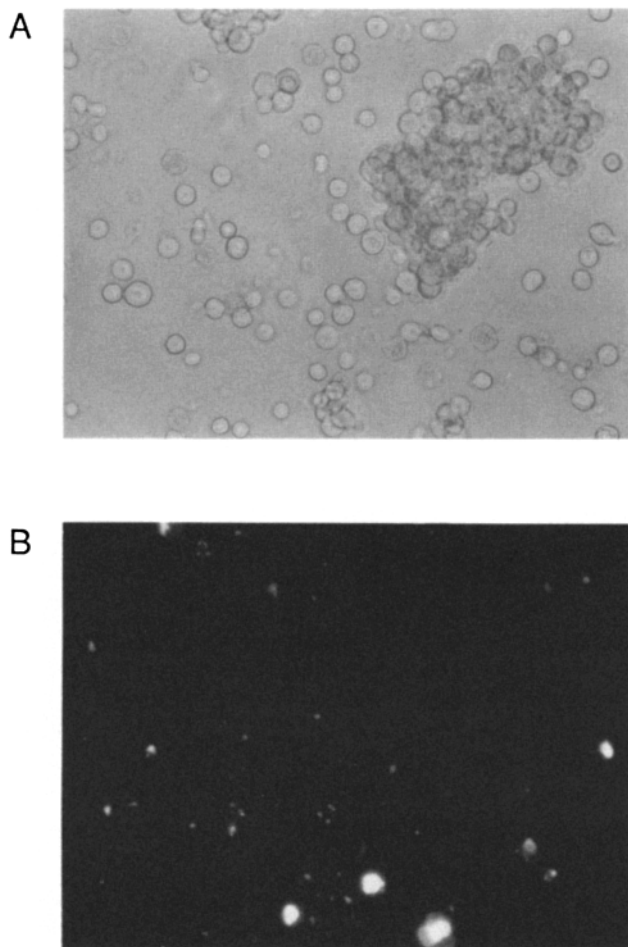


Figure 7. Aggregation of Sf9 cells expressing PTP μ occurs through homophilic binding. *a* is a representative phase contrast micrograph of an aggregation experiment in which Di I labeled uninfected Sf9 cells were mixed with full-length PTP μ -expressing cells before aggregation. *b* shows the same field as *a* photographed using an epifluorescence microscope to visualize the Di I. The Di I labeled uninfected cells did not contribute to the large aggregate. This is a representative picture of 32 such aggregates examined in three independent experiments.

and purified from *E. coli* using a glutathione Sepharose affinity column. This purified protein was adsorbed onto petri dishes and the binding of PTP μ or EGFR/PTP μ chimera-coated Covaspheres was visualized using an epifluorescence microscope after washing the dishes to remove nonadherent Covaspheres. There was no significant binding of the full-length PTP μ Covaspheres to GST alone [29 ± 0.6 Covaspheres per field (mean \pm SEM; $n = 6$) shown in Fig. 8 *a*]. Similarly, Fig. 8 *b* shows that there was no significant binding of EGFR/PTP μ chimera-coated Covaspheres to the GST/PTP μ extracellular segment (28 ± 3.9 Covaspheres per field; $n = 8$). However, Fig. 8 *c* demonstrates that the full-length PTP μ -coated Covaspheres bound specifically to the GST/PTP μ extracellular segment fusion protein (764 ± 41 Covaspheres per field; $n = 5$). These data establish that the homophilic binding in vitro, which was performed in PBS, was independent of any added divalent cations. Also because the GST/PTP μ protein was isolated from bacteria, glycosy-

lation of the extracellular domain is unlikely to contribute to the binding reaction. Similar observations were made for Nr-CAM (Mauro et al., 1992).

Baculovirus Expressed PTP μ Binds by a Homophilic Mechanism to PTP μ that Is Normally Expressed in MvLu Cells

We have examined the ability of PTP μ to bind homophilically when expressed at physiological levels. At this time, the only cell line we have found that expresses PTP μ is MvLu mink lung cells. This cell line expressed low but detectable levels of the enzyme as determined by immunoblotting with intracellular segment-directed monoclonal anti-PTP μ antibodies (data not shown). The ability of PTP μ -coated Covaspheres to bind to these cells was tested. As shown in Fig. 9, *a* and *b*, only low levels of the EGFR/PTP μ chimera-coated Covaspheres bound nonspecifically to the cells while high levels of binding of PTP μ -coated Covaspheres were observed (*c* and *d*). This binding was inhibited by pre-incubating the MvLu cells with antibodies to the extracellular domain of PTP μ (*e* and *f*). Pre-immune IgG did not affect the binding of PTP μ -coated Covaspheres (Table I). This "sided" experiment selectively blocks PTP μ on the MvLu cells, thus illustrating that binding occurs through a homophilic interaction. In a separate experiment, Fab' fragments of antibodies to PTP μ were pre-incubated with the PTP μ -coated Covaspheres, which also prevented binding. Incubation with Fab' fragments derived from pre-immune serum did not affect the binding. A quantitative summary of the results of Covasphere binding are presented in Table I. These data demonstrate that purified PTP μ was capable of binding homophilically to PTP μ expressed on the surface of MvLu cells.

GST/PTP μ -coated Covaspheres Aggregate In Vitro and Bind to MvLu Cells

In light of the preceding data, one might anticipate that Covaspheres coated with PTP μ would self-aggregate. To test this possibility the GST/PTP μ fusion protein was covalently coupled directly to Covaspheres to ensure a high protein:bead ratio. The Covaspheres were then rotated under low shear conditions to allow any aggregation to occur. As shown in Fig. 10, the Covaspheres coated with GST alone did not aggregate (*a*) whereas the GST/PTP μ -coated Covaspheres formed aggregates (*b*). These Covaspheres were also examined for their ability to bind to the surface of MvLu cells (Table II). While there was no significant binding of control GST Covaspheres, GST/PTP μ Covaspheres bound at high levels to MvLu cells. Furthermore, the GST/PTP μ Covaspheres did not bind to 3T3 cells which do not express detectable amounts of immunoreactive PTP μ .

Discussion

The cloning of receptor-type PTPases has led to the convergence of the fields of cell adhesion and protein tyrosine dephosphorylation. The structural similarity of some of the receptor-type PTPases to the immunoglobulin superfamily of cell adhesion molecules suggests that these phosphatases may participate in homophilic binding interactions, i.e., the "ligand" for one of these PTPases may be a molecule of the same enzyme expressed on an adjacent cell.

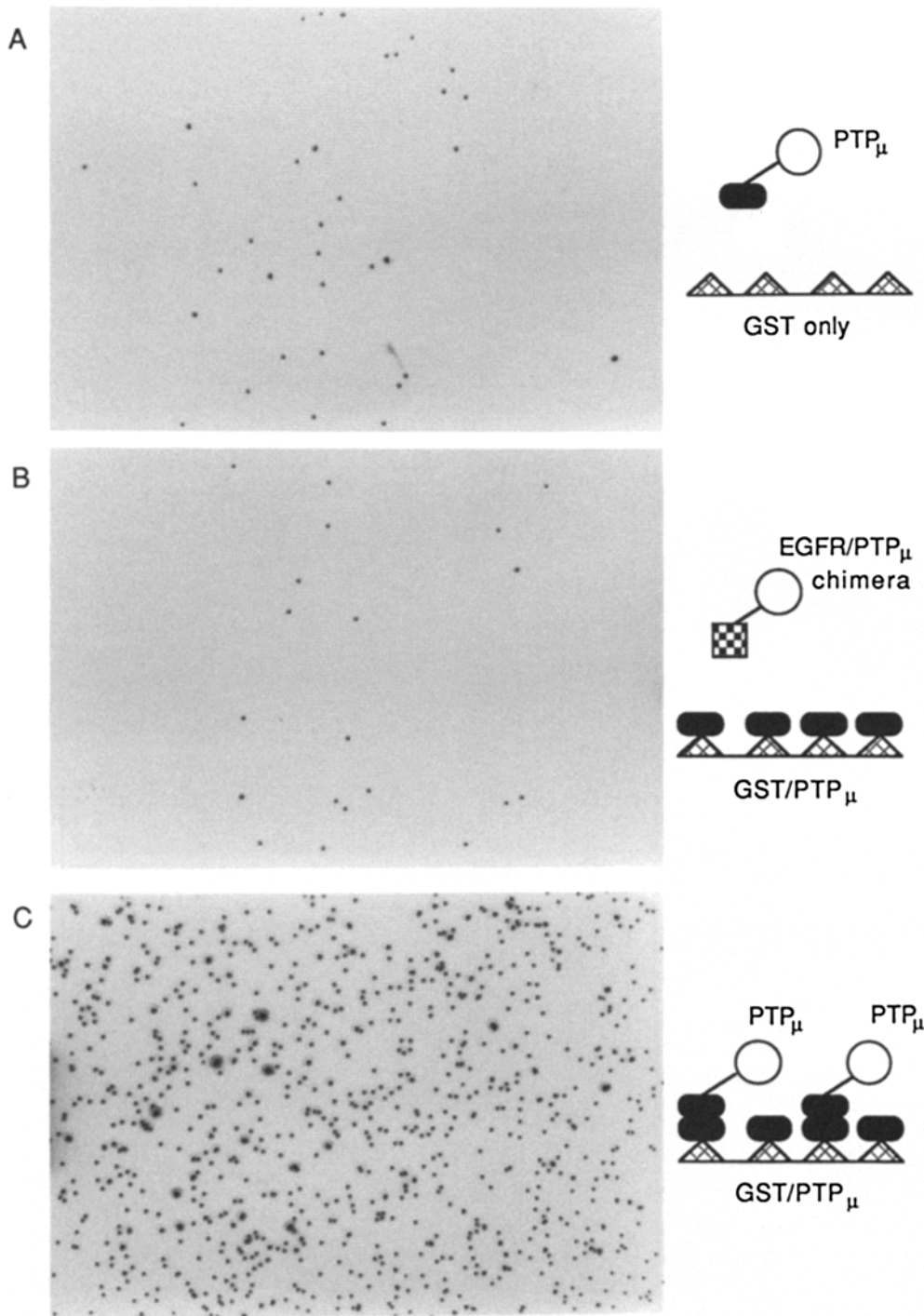


Figure 8. Binding of baculovirus-expressed PTP μ -linked Covaspheres to surfaces coated with purified fusion protein containing the extracellular segment of PTP μ (GST/PTP μ). *a* shows that there were only low levels of nonspecific binding of full-length PTP μ -coated Covaspheres to petri dishes coated with GST protein alone. Similarly, only low levels of nonspecific binding were seen when the EGFR/PTP μ chimera-coated Covaspheres were added to GST/PTP μ fusion protein-coated dishes (*b*). However, numerous full length PTP μ -coated Covaspheres bound to the GST/PTP μ fusion protein-coated dishes (*c*).

In this manuscript we report experiments which demonstrate homophilic binding between molecules of PTP μ by several criteria: (*a*) Expression of full-length PTP μ , or mutant forms with an intact extracellular segment, in the presence or absence of the PTPase domains, caused Sf9 cells to aggregate. Neither a chimeric phosphatase bearing the extracellular segment of the EGF-receptor and the intracellular segment of PTP μ nor the PTPase domains of PTP μ expressed as a soluble protein induced aggregation. When uninfected, fluorescently labeled Sf9 cells were mixed with

unlabeled PTP μ -expressing cells, the labeled cells were excluded from the aggregates. Furthermore, no immunoreactive species were detected after immunoblot analysis of lysates of uninfected Sf9 cells using a variety of mAbs to PTP μ . These data indicate that the counter-receptor for PTP μ was only found on the surfaces of other PTP μ -expressing cells. (*b*) The binding reaction has been reconstituted in vitro. First, fluorescent Covaspheres linked to baculovirus expressed PTP μ will bind to petri dishes coated with a GST fusion protein (GST/PTP μ) containing the ex-

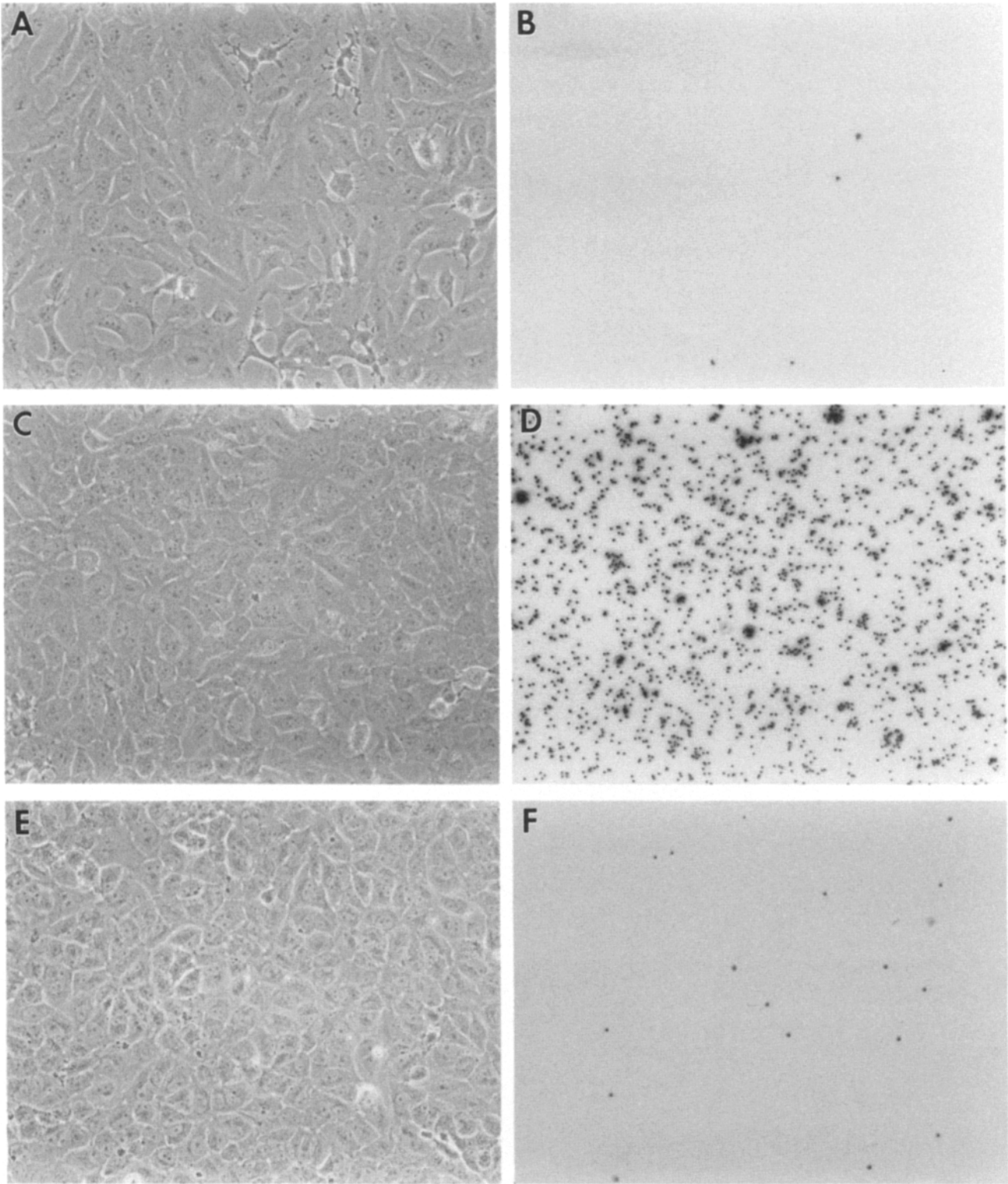


Figure 9. Homophilic binding of baculovirus-expressed PTP μ -coated Covaspheres to endogenous PTP μ in MvLu cells. Phase contrast micrographs of MvLu cells (*a*, *c*, and *e*) and micrographs of fluorescent Covasphere binding (*b*, *d*, and *f*) are shown. *a* and *b* show the low level of nonspecific binding when EGFR/PTP μ chimera-coated Covaspheres are applied to the cells. *c* and *d*, show full-length PTP μ -coated Covaspheres bound to the cells. In *e* and *f*, the MvLu cells were preincubated with an anti-peptide antibody to the amino-terminus of PTP μ before addition of the PTP μ -bound Covaspheres.

Table I. Quantitation of the Binding of PTP μ -coated Covaspheres to MvLu Cells

	Number bound/field*
PTP μ -Covaspheres	539 (\pm 94)
EGFR/PTP μ chimera-Covaspheres	9 (\pm 2)
PTP μ -Covaspheres pretreated with preimmune Fab'	475 (\pm 39)
PTP μ -Covaspheres pretreated with anti-PTP μ Fab'	15 (\pm 3)
PTP μ -Covaspheres added to MvLu cells pretreated with pre-immune antibody	401 (\pm 10)
PTP μ -Covaspheres added to MvLu cells pretreated with anti-PTP μ antibody	19 (\pm 3)

* The number of Covaspheres bound is represented as the mean of at least three fields. The field is 0.09 mm². The number in parentheses indicates SEM. Some of these results are illustrated in Fig. 9.

tracellular segment of PTP μ purified from *E. coli*, but will not bind to GST alone. Furthermore, Covaspheres linked to the EGFR/PTP μ chimera bind neither GST nor GST/PTP μ . Secondly, Covaspheres coated with GST/PTP μ fusion protein self-aggregate, whereas those coated with GST alone do not. (c) We show that PTP μ , as it is expressed endogenously on MvLu cells, retains the capacity to interact with baculovirus-expressed PTP μ linked to Covaspheres. While PTP μ -coated Covaspheres bind to MvLu cells, EGFR/PTP μ Covaspheres do not. Binding of PTP μ Covaspheres is blocked by pretreating the cells with antibody to the extracellular segment of PTP μ but not by a control pre-immune antibody.

Table II. Quantitation of Covasphere Binding to Cell Surfaces

	Number bound/field*
GST-Covaspheres added to MvLu cells	12 (\pm 2)
GST/PTP μ -Covaspheres added to MvLu cells	1,006 (\pm 78)
GST/PTP μ -Covaspheres added to 3T3 cells	9 (\pm 2)

* The number of Covaspheres bound is represented as the mean of at least six fields. The field is 0.09 mm² and contained approximately equal numbers of MvLu and 3T3 cells. The number in parentheses indicates SEM.

Also if the PTP μ Covaspheres are pretreated with Fab' fragments from the anti-PTP μ antibody, binding to MvLu cells is blocked whereas control Fab's are without effect. Collectively, these data establish homophilic binding between PTP μ molecules in a heterologous expression system, in vitro binding assays and in cells in which the molecule is normally expressed.

For immunoglobulin superfamily-type adhesion molecules, such as N-CAM, aggregation is highly dependent upon the concentration of the molecule at the cell surface (Hoffman and Edelman, 1983). Therefore, to assess the aggregation potential of PTP μ a model system had to be used that could generate high level expression of the protein at the cell surface. Introduction of potential adhesion molecules into nonadhesive cells has been used to demonstrate directly their adhesive functions. Transfection for determination of an adhesive function was first described by Takeichi and colleagues for E-cadherin (Nagafuchi et al., 1987). Goodman

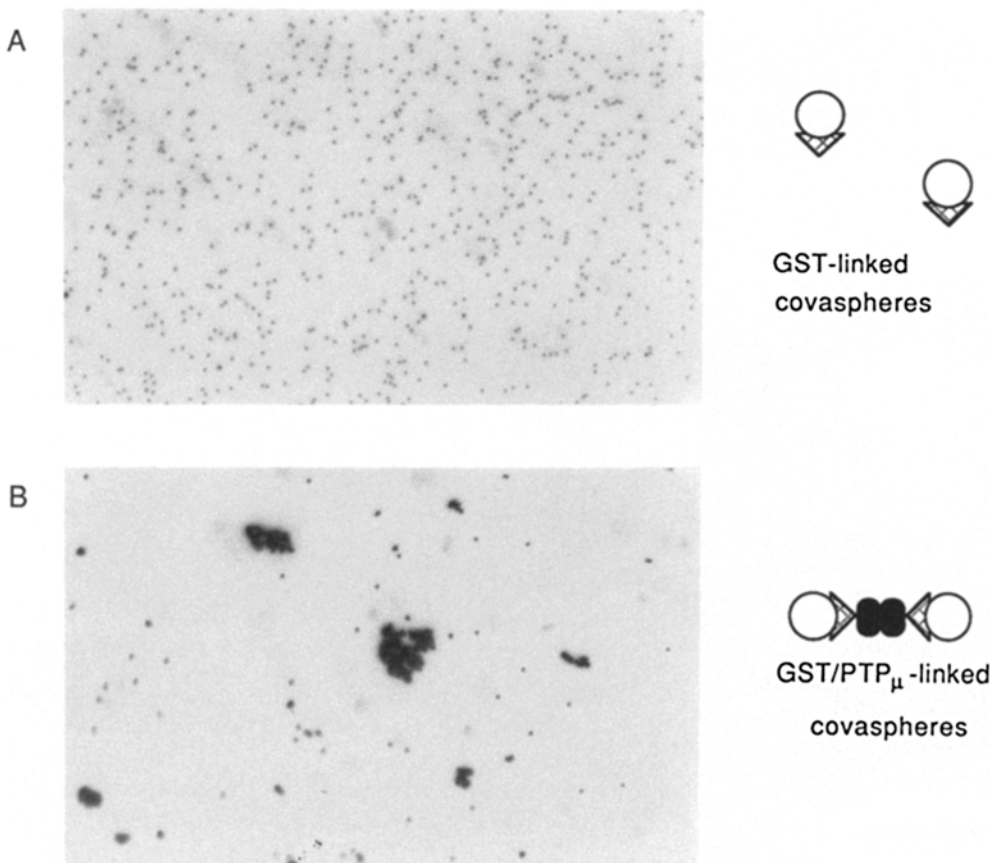


Figure 10. GST/PTP μ -coated Covaspheres aggregate in vitro. Covaspheres coated with GST alone or GST/PTP μ were used in an in vitro aggregation assay and visualized by an epifluorescence microscope. GST Covaspheres did not aggregate (a) whereas GST/PTP μ Covaspheres formed aggregates (b).

and co-workers introduced fasciclin III and connectin molecules into nonadhesive *Drosophila* S2 cells to demonstrate that these molecules mediated homophilic adhesion (Snow et al., 1989; Nose et al., 1992). Similarly, Pulido et al. (1992) have used *Drosophila* S2 cells to demonstrate that the receptor tyrosine kinase, *Dtrk*, may function as an adhesion molecule. We chose to use Sf9 insect cells and the baculovirus expression system (Matsuura et al., 1987; Summers and Smith, 1987) to express PTP μ . This system has been used previously to express high levels of glycosylated, biologically active haemagglutinin (Kuroda et al., 1986) and tissue plasminogen activator (Jarvis and Summers, 1989). An additional requirement for use in adhesion studies is a baculovirus-susceptible cell line with low levels of endogenous aggregation. Sf9 cells fulfill this condition in that they do not aggregate well in an in vitro assay.

The Sf9 cells infected with full length PTP μ showed high levels of aggregation (39%) compared to uninfected cells (11.9%), and the even lower levels of aggregation of cells infected with soluble PTP μ (4.6%) or the EGFR/PTP μ chimera (-4.1%). The level of aggregation induced by PTP μ was similar to that induced by other known adhesion molecules in various model systems. For example, transfection of mouse L-cells with N-CAM resulted in a difference in aggregation of 30% (Edelman et al., 1987) whereas N-CAM expression increased aggregation of an RSV-transformed neuronal cell line from 20 to 40% (Brady-Kalnay et al., 1993). Expression of Nr-CAM in L-cells increased aggregation to 14% (Mauro et al., 1992) while Nose et al. (1989) detected a 50% difference in aggregation of fasciclin III-transfected versus untransfected S2 cells. Therefore, PTP μ promotes similar levels of aggregation to those detected in response to well established cell-cell adhesion molecules. These data suggest that the Sf9 cell/baculovirus expression system may represent a useful tool for revealing potential adhesive functions for other molecules.

Whether or not PTP μ has the ability to perform this adhesive function in its normal environment remains to be determined. Nevertheless, we have shown that PTP μ expressed endogenously on the surface of MvLu cells retains the capacity for homophilic binding. However, MvLu cells do not aggregate in an aggregation assay. The low levels of the PTPase expressed in these cells may preclude the successful application of this assay. We have observed PTP μ protein to be expressed abundantly in lung, however at present we have not found a cell line that reflects that high level of expression. Thus, the ultimate resolution of the issue of whether, as suggested by the data presented in this manuscript, PTP μ can function as an adhesion molecule in vivo may require the use of primary cell systems.

High local concentrations in a specific area of the membrane could allow PTP μ to bind homophilically and function in an adhesive role even though it may be expressed at low overall levels in a particular cell. One novel feature of PTP μ that may be interesting in this regard is that its intracellular juxtamembrane segment is 70 amino acids longer than the equivalent segment in other receptor PTPases. Using the ALIGN program, we have shown that this portion of PTP μ displays sequence similarity to the intracellular segment of members of the cadherin family (Tonks et al., 1992). This homology is interesting because no other receptor type PTPases have such similarity to the cadherins, nor do any

other members of the immunoglobulin superfamily. The intracellular domain of the cadherins is the most highly conserved segment (~90% identity) among members of this family (reviewed by Takeichi, 1991) and interacts indirectly with the actin cytoskeleton (Ozawa et al., 1989). The cadherins are found at areas of cell contact, for example adherens-type cell junctions (Tsukita et al., 1991). Such cell junctions are areas of rapid phosphotyrosine turnover and locations at which tyrosine kinases including *src* and pp125^{src} are concentrated in normal and transformed cells (Volberg et al., 1991; Guan and Shalloway, 1992). In fact, changes in tyrosine phosphorylation may be involved in controlling the structural integrity of these junctions (Volberg et al., 1992). In light of this homology to the cadherins it is possible that PTP μ may also associate with the cytoskeleton through an interaction with accessory molecules at points of cytoskeletal-membrane association such as intercellular junctions.

PTP μ displays phosphatase activity after expression in Sf9 cells. To address the possibility that phosphatase activity was a requirement for cell-cell aggregation, we made a mutant form of PTP μ , PTP μ -extra, that lacked both phosphatase domains. This form of PTP μ induced aggregation with similar efficiency to the full-length form indicating that phosphatase activity and the aggregation response were not mutually dependent. Furthermore, the fact that the bacterially-expressed fusion protein GST/PTP μ containing only the extracellular segment of the PTPase was able to induce binding in vitro also suggested that aggregation was mediated solely by the extracellular domain. Aggregation did not result in dramatic changes in cellular phosphotyrosine levels as determined by anti-phosphotyrosine immunoblots. These data raise the possibility that, unlike the growth factor receptor PTKs where binding of the ligand to the extracellular segment triggers the kinase activity of the intracellular domain, PTP μ activity may not be strictly dependent upon ligand binding. However, this should be considered in light of the caveat that it is unlikely that Sf9 insect cells contain the normal substrates for human PTP μ . Changes in activity or affinity for a particular substrate may be seen when two molecules of PTP μ contact one another in their normal environment. Nevertheless, one could also propose a tethering role for homophilic binding interactions among PTP μ extracellular segments. Thus, the spatial distribution of the enzyme on the cell surface may be restricted, for example to intercellular junctions, with concomitant restriction of the spectrum of substrates or regulatory proteins with which it can interact.

PTP μ offers a unique, potentially direct link between cell adhesion phenomena and the triggering of signal transduction pathways. It remains possible that PTP μ itself may not directly promote cell-cell adhesion in vivo. However, in the context of adhesion driven by PTP μ -independent binding mechanisms, homophilic interactions between apposing PTP μ molecules may alter their phosphatase activity. Thus the homology of PTP μ to the cadherins, N-CAM-like molecules and PTPases predicts an interesting role in cell-cell communication and continuing studies of PTP μ should yield important new insights into the control of signal transduction processes.

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References

- Brackenbury, R., J. Thiery, U. Rutishauser, and G. M. Edelman. 1977. Adhesion among neural cells of the chick embryo. *J. Biol. Chem.* 252:6835-6840.
- Bradford, M. 1976. A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Anal. Biochem.* 72:248-254.
- Brady-Kalnay, S., E. Boghaert, S. Zimmer, and R. Brackenbury. 1993. Increased N-CAM-mediated cell-cell adhesion does not reduce WC5 rat cerebellar cell invasion. *Clin. Exp. Metastasis.* 11:313-324.
- Brady-Kalnay, S., and N. K. Tonks. 1993. Purification and characterization of the human protein tyrosine phosphatase, PTP μ , from a baculovirus expression system. *Mol. Cell. Biochem.* In Press.
- Brown-Shimer, S., K. Johnson, D. Hill, and A. Bruskin. 1992. Effect of protein tyrosine phosphatase 1B expression on transformation by the human *neu* oncogene. *Cancer Res.* 52:478-482.
- Charbonneau, H., and N. K. Tonks. 1992. 1002 protein phosphatases? *Annu. Rev. Cell Biol.* 8:463-493.
- Edelman, G. M., B. Murray, R. Mege, B. Cunningham, and W. Gallin. 1987. Cellular expression of liver and neural cell adhesion molecules after transfection with their cDNAs results in specific cell-cell binding. *Proc. Natl. Acad. Sci. USA.* 84:8502-8506.
- Edelman, G. M., and K. L. Crossin. 1991. Cell adhesion molecules: implications for a molecular histology. *Annu. Rev. Biochem.* 60:155-190.
- Fischer, E. H., H. Charbonneau, and N. K. Tonks. 1991. Protein tyrosine phosphatases: a diverse family of intracellular and transmembrane enzymes. *Science (Wash. DC).* 253:401-406.
- Flint, A., M. Gebbink, B. Franza, D. Hill, and N. K. Tonks. 1993. Multi-site phosphorylation of the protein tyrosine phosphatase, PTP1B: identification of cell cycle regulated and phorbol ester stimulated sites of phosphorylation. *EMBO (Eur. Mol. Biol. Organ.) J.* 12:1937-1946.
- Gebbink, M., I. van Etten, G. Hateboer, R. Suijkerbuijk, R. Beijersbergen, A. van Kessel, and W. Moolenaar. 1991. Cloning, expression and chromosomal localization of a new putative receptor-like protein tyrosine phosphatase. *FEBS (Fed. Eur. Biochem. Soc.) Lett.* 290:123-130.
- Guan, J., and D. Shalloway. 1992. Regulation of focal adhesion-associated protein tyrosine kinase by both cellular adhesion and oncogenic transformation. *Nature (Lond.)* 358:690-692.
- Guán, K., and J. Dixon. 1991. Eukaryotic proteins expressed in *Escherichia coli*: an improved thrombin cleavage and purification procedure of fusion proteins with glutathione S-transferase. *Anal. Biochem.* 192:262-267.
- Hall, A., R. Nelson, and U. Rutishauser. 1990. Binding properties of detergent-solubilized NCAM. *J. Cell Biol.* 110:817-824.
- Hoffman, S., and G. M. Edelman. 1983. Kinetics of homophilic binding by embryonic and adult forms of the neural cell adhesion molecule. *Proc. Natl. Acad. Sci. USA.* 80:5762-5766.
- Laemmli, U. K. 1970. Cleavage of structural proteins during the assembly of the head of bacteriophage T4. *Nature (Lond.)* 227:680-685.
- Jarvis, D., and M. Summers. 1989. Glycosylation and secretion of human tissue plasminogen activator in recombinant baculovirus-infected insect cells. *Mol. Cell. Biol.* 9:214-223.
- Kaplan, D., D. Morrison, G. Wong, F. McCormick, and L. T. Williams. 1990. PDGF β -receptor stimulates tyrosine phosphorylation of GAP and association of GAP with a signalling complex. *Cell.* 61:125-133.
- Kato, J., H. Matsushima, S. Hiebert, M. Ewen, and C. Sherr. 1993. Direct binding of cyclin D to the retinoblastoma gene product (pRb) and pRb phosphorylation by the cyclin D-dependent kinase CDK4. *Genes Dev.* 7:331-342.
- Kuroda, K., C. Hauser, R. Rott, H. Klenk, and W. Doerfler. 1986. Expression of the influenza virus haemagglutinin in insect cells by a baculovirus vector. *EMBO (Eur. Mol. Biol. Organ.) J.* 5:1359-1365.
- Matsuura, Y., R. Possee, H. Overton, and D. Bishop. 1987. Baculovirus expression vectors: the requirements for high level expression of proteins, including glycoproteins. *J. Gen. Virol.* 68:1233-1250.
- Mauro, V., L. Krushel, B. Cunningham, and G. M. Edelman. 1992. Homophilic and heterophilic binding activities of Nr-CAM, a nervous system cell adhesion molecule. *J. Cell Biol.* 119:191-202.
- Nagafuchi, A., Y. Shirayoshi, K. Okazaki, K. Yasuda, and M. Takeichi. 1987. Transformation of cell adhesion properties by exogenously introduced E-cadherin cDNA. *Nature (Lond.)* 329:341-343.
- Nose, A., V. Mahajan, and C. Goodman. 1992. Connectin: a homophilic cell adhesion molecule expressed on a subset of muscles and the motoneurons that innervate them in *Drosophila*. *Cell.* 70:553-567.
- Ozawa, M., H. Baribault, and R. Kemler. 1989. The cytoplasmic domain of the cell adhesion molecule uvomorulin associates with three independent proteins structurally related in different species. *EMBO (Eur. Mol. Biol. Organ.) J.* 8:1711-1717.
- Parker, L., S. Atherton-Fessler, M. Lee, S. Ogg, J. Falk, K. Swenson, and H. Piwnicka-Worms. 1991. Cyclin promotes the tyrosine phosphorylation of p34^{cdc2} in a *wee1*⁺ dependent manner. *EMBO (Eur. Mol. Biol. Organ.) J.* 10:1255-1263.
- Pulido, D., S. Campuzano, T. Koda, J. Modolell, and M. Barbacid. 1992. *Dtrk*, a *Drosophila* gene related to the *trk* family of neurotrophin receptors, encodes a novel class of neural cell adhesion molecule. *EMBO (Eur. Mol. Biol. Organ.) J.* 11:391-404.
- Rao, Y., X. Wu, J. Garipey, U. Rutishauser, and C. Siu. 1992. Identification of a peptide sequence involved in homophilic binding in the neural cell adhesion molecule NCAM. *J. Cell Biol.* 118:937-949.
- Snow, P., A. Bieber, and C. Goodman. 1989. Fasciclin III: a novel homophilic adhesion molecule in *Drosophila*. *Cell.* 59:313-323.
- Streuli, M., N. Krueger, L. Hall, S. Schlossman, and H. Saito. 1988. A new member of the immunoglobulin superfamily that has a cytoplasmic region homologous to the leukocyte common antigen. *J. Exp. Med.* 168:1523-1530.
- Streuli, M., N. Krueger, A. Tsai, and H. Saito. 1989. A family of receptor-linked protein tyrosine phosphatases in humans and *Drosophila*. *Proc. Natl. Acad. Sci. USA.* 86:8698-8702.
- Summers, M., and G. Smith. 1987. *A Manual of Methods for Baculovirus Vectors and Insect Cell Culture Procedures*. Texas Agriculture Experimental Station, College Station, TX.
- Takeichi, M. 1977. Functional correlation between cell adhesive properties and some cell surface proteins. *J. Cell Biol.* 75:464-474.
- Takeichi, M. 1991. Cadherin cell adhesion receptors as a morphogenetic regulator. *Science (Wash. DC).* 251:1451-1455.
- Tonks, N. K., Q. Yang, A. Flint, M. Gebbink, B. Franza, D. Hill, H. Sun, and S. Brady-Kalnay. 1992. Protein tyrosine phosphatases: the problems of a growing family. *Cold Spring Harbor Symp. Quant. Biol.* 57:87-94.
- Towbin, H., T. Staehelin, and J. Gordon. 1979. Electrophoretic transfer of proteins from polyacrylamide gels to nitrocellulose sheets: procedure and some applications. *Proc. Natl. Acad. Sci. USA* 76:4350-4354.
- Trowbridge, I. S. 1991. CD45: a prototype for transmembrane protein tyrosine phosphatases. *J. Biol. Chem.* 266:23517-23520.
- Tsukita, S., K. Oishi, T. Akiyama, Y. Yamanashi, T. Yamamoto, and S. Tsukita. 1991. Specific proto-oncogene tyrosine kinases of *src* family are enriched in cell-to-cell adherens junctions where the level of tyrosine phosphorylation is elevated. *J. Cell Biol.* 113:867-879.
- Villalba, M., S. Wente, D. Russell, J. Ahn, C. Reichelderfer, and R. Rosen. 1989. Another version of the human insulin receptor kinase domain: Expression, purification, and characterization. *Proc. Natl. Acad. Sci. USA.* 86:7848-7852.
- Volberg, T., B. Geiger, R. Dror, and Y. Zick. 1991. Modulation of intercellular adherens-type junctions and tyrosine phosphorylation of their components in RSV-transformed cultured chicken cells. *Cell Reg.* 2:105-120.
- Volberg, T., Y. Zick, R. Dror, I. Sabanay, C. Gilon, A. Levitzki, and B. Geiger. 1992. The effect of tyrosine-specific protein phosphorylation on the assembly of adherens-type junctions. *EMBO (Eur. Mol. Biol. Organ.) J.* 11:1733-1742.
- Woodford-Thomas, T., J. Rhodes, and J. Dixon. 1992. Expression of a protein tyrosine phosphatase in normal and *v-src*-transformed mouse 3T3 fibroblasts. *J. Cell Biol.* 117:401-414.
- Zander, N., D. Cool, C. Diltz, L. Rohrschneider, E. Krebs, and E. Fischer. 1993. Suppression of *v-fms*-induced transformation by overexpression of a truncated T-cell protein tyrosine phosphatase. *Oncogene.* 8:1175-1182.
- Zheng, X., Y. Wang, and C. Pallen. 1992. Cell transformation and activation of pp60 *c-src* by overexpression of a protein tyrosine phosphatase. *Nature (Lond.)* 359:336-339.