

## ORIGINAL RESEARCH

# Curcumin encapsulation and protection based on lysozyme nanoparticles

Jilai Cui<sup>1,2,3</sup>  | Jie Zhou<sup>1,2,3</sup> | Lu Huang<sup>1</sup> | Junxiang Jing<sup>1</sup> | Ningze Wang<sup>1</sup> | Luyuan Wang<sup>1</sup>

<sup>1</sup>College of Life Science, Xinyang Normal University, Xinyang, China

<sup>2</sup>Tea Plant Biology Key Laboratory of Henan Province, Xinyang, China

<sup>3</sup>Institute for Conservation and Utilization of Agro-bioresources in Dabie Mountains, Xinyang, China

## Correspondence

Jilai Cui, College of Life Science, Xinyang Normal University, Xinyang 464000, China.  
Email: jilaicui@xynu.edu.cn

## Funding information

Young Backbone Teachers Program, Grant/Award Number: 2018GGJS-13; Nanhu Scholars Program for Young Scholars

## Abstract

Curcumin possesses antioxidant, anti-inflammatory, and other properties. However, this compound exhibits low bioavailability because of its poor solubility and stability. In this paper, lysozyme nanoparticles were fabricated through solvent evaporation, and then, the solubilization and protection capability of curcumin were investigated. Lysozyme nanoparticles were characterized by dynamic light scattering technique, atomic force microscopy, transmission electron microscopy, and Fourier transform infrared spectroscopy. The load capacity and stability in thermal environment were further explored. Results showed that the lysozyme nanoparticle displayed a spherical structure ( $127.9 \pm 2.12$  nm) with favorable distribution. The solubility of curcumin can increase to 22  $\mu\text{g/mL}$ . After encapsulation by lysozyme nanoparticles, the retentive curcumin can reach up to 67.9% and 30.25% at 25°C and 50°C, respectively, significantly higher than that of free curcumin. Meanwhile, experiments on DPPH free radical scavenging activity indicated the curcumin loaded by lysozyme nanoparticle possessed higher free radical scavenging activity than that of free curcumin with same treatments. The results confirmed that lysozyme nanoparticles exhibit potential applications in solubilizing and protecting the environment-sensitive hydrophobic functional components.

## KEYWORDS

curcumin, lysozyme, nanoparticles, protection, solubilization

## 1 | INTRODUCTION

Curcumin (Cur) is a yellow pigment from traditional medicine turmeric and is commonly used as a spice and food-coloring agent. Cur has drawn much attention because Cur shows multiple properties, such as anti-inflammatory, antioxidant, decreasing hematic fat, resisting sclerosis of arterial congee appearance, and antitumor (Karimian, Pirro, Johnston, Majeed, & Sahebkar, 2017; Saberi-Karimian et al., 2018; Wang et al., 2017). However, as a functional component and effective therapeutic agent, Cur exhibits low water solubility and relatively high environmental sensitivity to

temperature and ultraviolet radiation, which severely limit its application (Nimiya et al., 2016; Zhao, Pan, Nitin, & Tikekar, 2014). Various systems, including emulsion, polysaccharide/protein complex, and nanogel, have been explored and applied to improve the stability and bioavailability of Cur (Mangalathillam et al., 2011; Tapal & Tikku, 2012; Zou et al., 2016).

In recent years, nanoparticles are commonly used in loading, protection, and delivery systems for functional components, such as polyphenols, carotenoids, vitamins, fatty acids, sterols, and other active ingredients (Chen et al., 2013). Nanoparticles improve the solubility and stability of active ingredients and expand the application fields of

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2019 The Authors. *Food Science & Nutrition* published by Wiley Periodicals, Inc.

functional components, especially for the compounds being sensitive to the environment, such as Cur and folic acid (Das, Kasoju, & Bora, 2010; Sneharani, Karakkat, Singh, & Rao, 2010). A carrageenan/lysozyme soluble complex was prepared by one-step self-assembly and was found to be a suitable vehicle for solubilizing and protecting curcumin in heating and ultraviolet radiation environment (Xu et al., 2014).

In this paper, lysozyme nanoparticles (Ly NPs) were prepared by desolvation method. Ly NPs were subsequently characterized through particle size and morphology. Therefore, the encapsulation and protection of Cur were explored, and the antioxidant activity was further verified using DPPH. This endeavor attempted to provide a simple and feasible strategy for solubilization and protection of the hydrophobic functional components.

## 2 | MATERIALS AND METHODS

### 2.1 | Materials

Curcumin (95.0% purity), as well as lysozyme (Ly, 14.3 kDa) from chicken egg white, was purchased from Sinopharm Chemical Reagent Co., Ltd. Other chemicals were of reagent grade and used without purification. All solutions used in the experiments were prepared using ultrapure water through a Millipore (Millipore) Milli-Q water purification system.

### 2.2 | Preparation of Ly NPs

Ly NPs were prepared by desolvation (Jahanban-Esfahlan, Dastmalchi, & Davaran, 2016). Ly was dissolved in purified water with gentle magnetic stirring for 2 hr at room temperature at the concentration of 1.0 mg/mL. Ethanol was added to Ly solutions at the volume ratio of 15:5 with further stirring for 1 hr. After becoming turbid, the solution was incubated at 80°C for 15 min. Subsequently, Ly NPs were prepared by removing ethanol through rotary evaporation at 30°C under vacuum, and the NPs were stored at 4°C for further use.

### 2.3 | Characterization of Ly NPs

The particle size and zeta potential ( $\zeta$ ) of Ly NPs were measured with Nano-ZS90 at 25°C (Malvern Instruments) according to Xu (Xu, Ge, et al., 2018). The Fourier transform infrared (FT-IR) spectra were recorded using a Nicolet Nexus 470 spectrometer with 32 scans and resolution of 4 cm<sup>-1</sup> in the range of 400–4000 cm<sup>-1</sup>. The morphology of the complex was investigated with a JEOL transmission electron microscope (TEM) (H-7650, Hitachi). During the TEM test, a drop of Ly NPs solution was dispensed directly onto a carbon-coated copper grid and allowed to dry spontaneously.

Morphology of the particles was determined using atomic force microscopy (AFM) (Wei et al., 2015). Tapping mode measurements were performed in air using a Dimension Icon with a Nanoscope IV controller (Bruker Corporation) and silicon cantilevers with tetrahedral tips (OMCL-AC160TS—Olympus) with a nominative force constant of

42 Nm<sup>-1</sup> and a resonance frequency of around 300 kHz. For sample preparation, undiluted nanosuspensions were dropped onto a freshly cleaved mica surface (Plano GmbH). After 5 min, the liquid was removed with a tissue and the samples were completely dried before imaging.

### 2.4 | Cur encapsulation based on Ly NPs

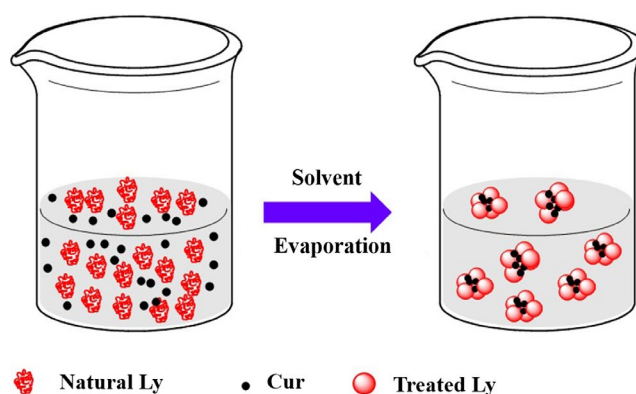
Cur was firstly dissolved in ethanol and then added into the Ly NPs solution with different final concentrations (5, 8, 10, 13 mg/mL) for encapsulation according to previous reports as described in Figure 1 (Naksuriya et al., 2015). Cur concentration was determined using a UV-vis spectrophotometer (UV-1100, MAPADA) at 428 nm (Moussa, Hmadeh, Abiad, Dib, & Patra, 2016; Nazari-Vanani, Moezi, & Heli, 2017). Free Cur (the supernatant solution) solution was obtained after centrifugation at 1888 g for 30 min at 4°C, and the content of Cur was estimated from a standard X-Y plot of Cur (dissolved in ethanol) versus concentration. Encapsulation efficiency (EE) and loading capacity (LC) were defined as the drug content encapsulated into Ly NPs and calculated as follows (Xu et al., 2019):

$$EE(\%) = \frac{\text{Total Cur} - \text{Free Cur}}{\text{Total Cur}} \times 100$$

$$LC (\mu\text{g/g}) = \frac{\text{Total Cur} - \text{Free Cur}}{\text{Weight of Ly NPs}} \times 100$$

### 2.5 | Cur protection by Ly NPs

To evaluate the effect of encapsulation on the stability of Cur against heat treatment, Cur retention rate was comparatively studied. Free Cur solution and Cur-loaded NPs solution at 10 μg/mL (in water) shared 25°C and 50°C treatment for 30 min. Free Cur was prepared equally to the encapsulated Cur in which water was replaced by the Ly NPs solution. For the Cur-encapsulated Ly NPs, ethanol was added equally, stirred for 4 hr, and then evaporated overnight at 40°C under vacuum (Kumar & Ahuja, 2013). The retention rate of Cur was calculated through the ratio of absorbance at 428 nm.



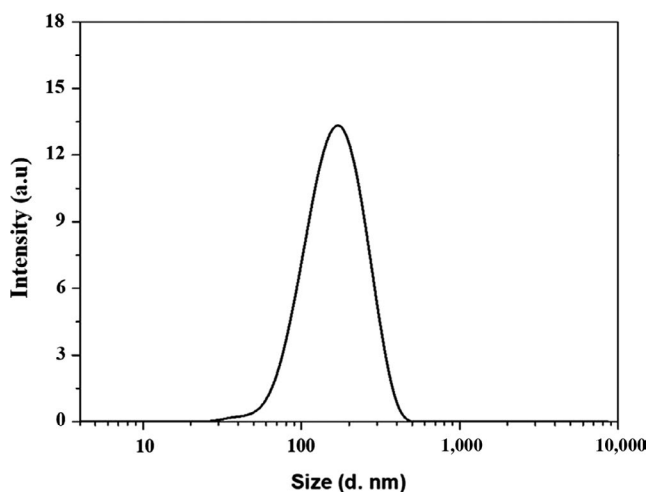
**FIGURE 1** Tentative encapsulation scheme of Cur based on Ly NPs

## 2.6 | Antioxidant activity of free and encapsulated Cur

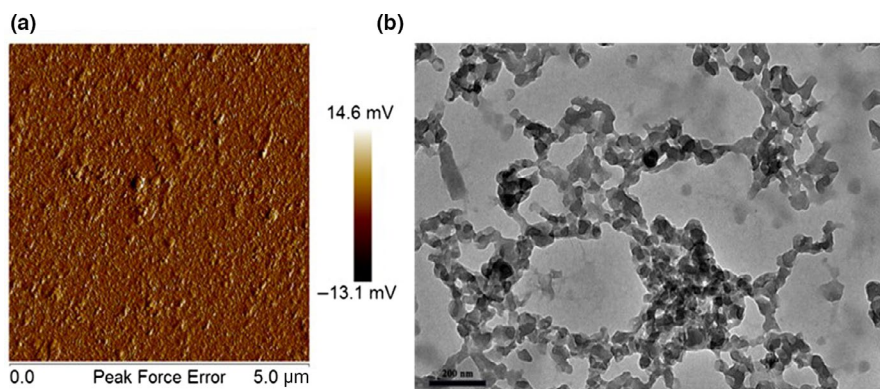
Antioxidant activity was further verified using DPPH method after heat treatment. In brief, free Cur solution and Cur-loaded NPs solution at 10  $\mu\text{g}/\text{mL}$  (in ethanol) were suffered at 25°C and 50°C for 30 min. The scavenging activity assay was carried out by recording the absorbance of DPPH solution (100  $\mu\text{M}$ ) at 517 nm in the presence and absence of the remnant Cur with a UV-vis spectrophotometer (Hamlaoui, Bencheraiet, Bensegueni, & Bencharif, 2017; Tonelli et al., 2019). The free radical scavenging potency of the curcumin was expressed as the percentage of DPPH that was decreased compared with that of the control condition after 30 min of preservation in the dark.

## 3 | RESULTS AND DISCUSSION

Figure 2 shows that the average particle size of the prepared Ly NPs was  $127.9 \pm 2.12$  nm. The size distribution index (PDI) of Ly NPs was  $0.29 \pm 0.03$ . The microscopic morphology of Ly NPs was observed through atomic force microscopy (AFM) and TEM



**FIGURE 2** Size distribution of Ly NPs

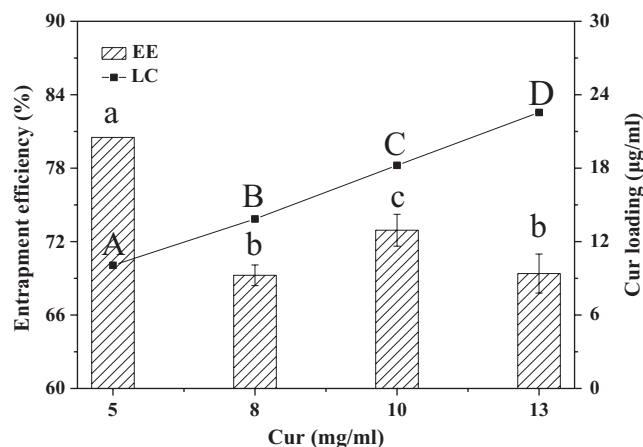


**FIGURE 3** AFM (a) and TEM (b) microstructure of Ly NPs

(Figure 3). AFM and TEM results showed that Ly NPs displayed elliptical appearance with a diameter probably around 90 nm and good dispersivity. The measured particle size was smaller than the hydrate particle size.

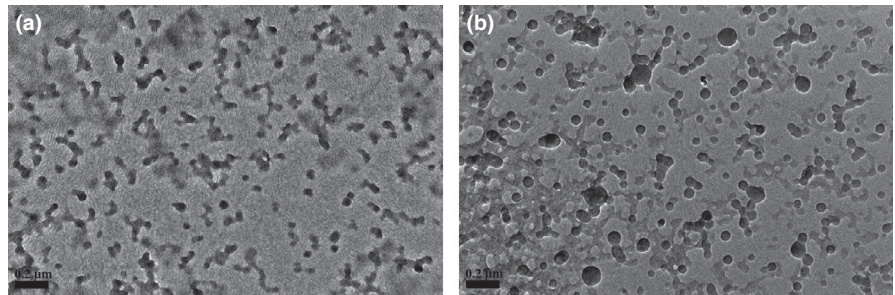
Cur is recognized as a physiological and pharmacological functional nutritional component. Figure 4 illustrates that the hydrophobic micro area of Ly NPs can improve the solubility of Cur. The EEs of Cur were different at different concentrations. At the concentration of 5 mg/mL, EE reached 80.5%. The EE then decreased with increasing Cur concentration, indicating lower loading capacity at lower Cur concentration. By contrast, LC continued to increase with higher concentration of Cur, and LC reached  $22.55 \pm 0.08$   $\mu\text{g}/\text{mg}$  at the Cur concentration of  $13 \pm 0.02$  mg/mL. When the concentration of Cur was 10 mg/mL, EE and LC were  $72.93\% \pm 1.3\%$  and  $18.23 \pm 0.05$   $\mu\text{g}/\text{mg}$ , respectively.

For exploring the influence of Cur loading on the structure of Ly NPs, the microscopic appearance was visualized through TEM after Cur encapsulation (Figure 5). The result showed that the Cur packaged Ly NPs displayed better dispersion. FT-IR spectroscopy was used to explore the change of molecular group during Ly NPs



**FIGURE 4** EE and LC of Cur based on Ly NPs. EE change was shown as bar chart (left axis), and LC change was shown as line chart. Different letters in lowercase mean statistical differences in Entrapment efficiency (EE), and different letters in upper case mean statistical difference in curcumin loading (LC)

**FIGURE 5** TEM of free Ly NPs (a) and Cur-loaded Ly NPs (b). Scale bar = 0.2  $\mu\text{m}$



fabrication and the interaction between Ly and Cur (Figure 6). The FT-IR spectra of Ly NPs showed coincident absorption peaks with those of Ly, including the characteristic absorption peaks of amide I, amide II, and amide II. The significant change observed around the  $3,500\text{ cm}^{-1}$  range, possibly due to N-H or O-H stretches. However, the absorption peak of Ly NPs shifted compared with that of Ly. The phenomenon was resulted from the partly denaturation during NPs preparation. It also found in the other protein during NPs fabrication through heat treatment (Liufeng et al., 2015; Xu, Jin, et al., 2018). After encapsulation of the Cur-loaded Ly NPs did not show new absorption peak compared with Cur and Ly NPs.

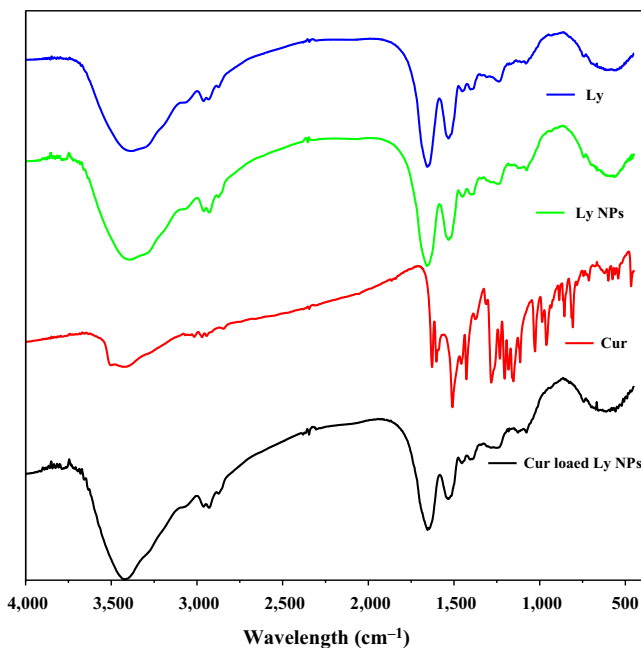
For free Cur, the degradation behavior with different rates occurred at  $25^\circ\text{C}$  and  $50^\circ\text{C}$ . The survival rates were 58.6% and 15.4% after 30 min of treatment. The heat-induced Cur degradation behavior has been confirmed by other reports (Ko, Chang, Wang, Wang, & Hsieh, 2015; Refat, 2013). Figure 7 shows that in all cases the antioxidant activity of Cur decreased after heat treatment, indicating its sensibility in thermal environment (Paramera, Konteles, & Karathanos, 2011). The free Cur exhibited high radical scavenging activity at 56.3% and decreased to 50% after heat treatment at  $50^\circ\text{C}$  for 30 min. The

DPPH radical scavenging activity of packaged Cur increased to 71.9% and 76.5% after treatment at  $25^\circ\text{C}$  and  $50^\circ\text{C}$  for 30 min, respectively.

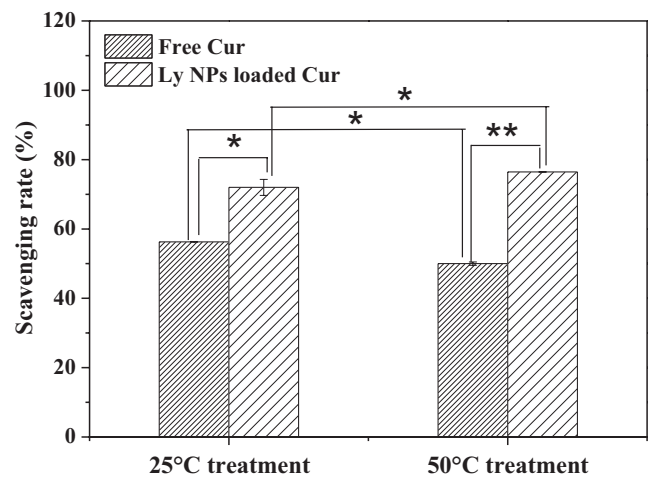
## 4 | DISCUSSION

Ly NPs displayed a single favorable peak distribution with the PDI less than 0.3 and good dispersivity of the prepared Ly NPs (Zhu et al., 2013). The size distribution of Ly NPs was resulted from swelling behavior in the water environment and dehydration during drying. The size of Ly NPs became smaller as measured by TEM. This phenomenon was also widely observed in other similar studies (Lin et al., 2015).

Cur offers potential applications in the fields of functional food and medicine, and has attracted considerable attention and research in recent years. However, its low water solubility and stability result in low bioavailability, which notably limits its wide application. Entrapment of Cur based on Ly NPs indicated that  $72\%$  ( $7.3 \pm 0.13\ \mu\text{g}/\text{mL}$ ,  $5.07 \times 10^{-5}\ \text{mol}/\text{L}$ ) of Cur was loaded in Ly NPs. Solubility improved by approximately 664 times compared with that in the water ( $11\ \text{ng}/\text{mL}$ ) (Kaminaga et al., 2003). The improvement in Cur water solubility was an order of magnitude higher than those in previous reports based on beta-casein micelles and hydrophobic modified starch ( $7.5 \times 10^{-5}\ \text{mol}/\text{L}$ ,  $1 \times 10^{-5}\ \text{mol}/\text{L}$ ) (Letchford & Burt, 2007; Yu & Huang, 2010). This phenomenon may be induced by the exposure of the hydrophobic group in the hydrophobic cavity of



**FIGURE 6** FT-IR spectra of Ly, Cur, Ly NPs, and Cur-loaded Ly NPs, respectively, from top to bottom

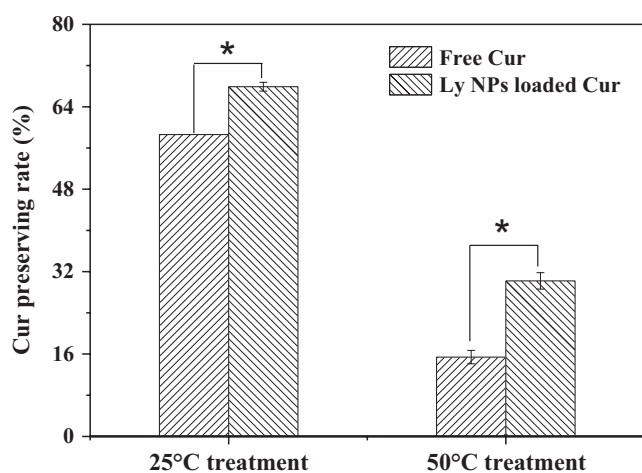


**FIGURE 7** Radical scavenging activity improvements by Cur entrapment. One asterisk means statistically  $p < 0.05$ , and two asterisks mean very significant difference with  $p < 0.01$

Ly during self-assembly. This result illustrated that Ly NPs exhibit its efficient encapsulation ability of Cur and potential for loading other hydrophobic health-beneficial compounds. After entrapment, the Cur loading endowed high steric hindrance effect between Ly NPs. Ly NPs were difficult to aggregate and exhibited improved elliptic morphology. FT-IR further illustrated that the structure of Ly was partially damaged. These results indicated that Cur was loaded driven by thermodynamics and self-assembled by physical interaction. This phenomenon was also confirmed by Cur-encapsulated zein nanoparticle, which indicates that hydrophobic interaction was the main force between the Cur and Ly NPs (Liang et al., 2015).

Temperature and other factors affect the structure and activity of Cur. However, heating is a conventional technique in food processing, especially for sterilization. Figure 8 shows that the Ly NPs package strategy is a simple and effective method to alleviate the degradation of Cur against thermal treatment. Obviously, compared with free Cur, the stability of Cur was significantly improved through encapsulation into Ly NPs. Interestingly, high processing temperature brought high Cur rate. Comparatively, the Cur retention rate increased to 67.9% and 30.25% after Ly NPs package with the same treatment. Therefore, the strategy of Ly NPs package not only solubilized Cur but also alleviated the degradation behavior in the adverse environments. The attempt may expand the application of Cur in food field, especially in the application of high temperature rapid processing products. The antioxidant capacity improved by 20% for the packaged Cur at 50°C. The DPPH experiment verified that encapsulating Cur in Ly NPs alleviated its breakdown during food heating process and protected its antioxidant activity.

In conclusion, Ly NPs with average size of  $127.9 \pm 2.12$  nm were prepared through desolvation method. The prepared Ly NPs exhibit regular elliptical appearance and good size distribution. During fabrication, the structure of Ly was partially damaged. Cur was loaded as driven by thermodynamics and self-assembled by physical interaction, which resulted in improved appearance and dispersion. At the Cur concentration of 10 mg/mL, EE and LC values were  $72.93 \pm 1.3\%$



**FIGURE 8** Cur protections by Ly NPs in the heat treatment. Cur preserving rates are significant different ( $p < 0.05$ ) for free and encapsulated Cur in both 25°C and 50°C treatments

and  $18.23 \pm 0.05$   $\mu\text{g}/\text{mg}$ , respectively. Water solubility of encapsulated Cur increased by approximately 660 times. Compared with free Cur, packaged Cur displayed significantly improved survival rate after heat treatment. Increased processing temperature translates to enhanced Cur survival rate. DPPH test also showed that packaged Cur exhibited high antioxidant capability after 50°C treatment. Thus, encapsulation of Cur in Ly NPs is a practical method for the encapsulation and protection of Cur, possibly providing a workable reference for other environment-sensitive hydrophobic functional components.

## ACKNOWLEDGEMENT

This work was financially supported by the Young Backbone Teachers Program (Grant No. 2018GGJS-13) and Nanhu Scholars Program for Young Scholars of XYNU.

## CONFLICT OF INTEREST

The authors declare that they do not have any conflict of interest.

## ETHICAL APPROVAL

This study does not involve any human or animal testing.

## ORCID

Jilai Cui  <https://orcid.org/0000-0002-6949-2424>

## REFERENCES

- Chen, W., Zheng, M., Meng, F., Cheng, R., Deng, C., Feijen, J., & Zhong, Z. (2013). In situ forming reduction-sensitive degradable nanogels for facile loading and triggered intracellular release of proteins. *Biomacromolecules*, 14(4), 1214–1222. <https://doi.org/10.1021/bm400206m>
- Das, R. K., Kasoju, N., & Bora, U. (2010). Encapsulation of curcumin in alginate-chitosan-pluronic composite nanoparticles for delivery to cancer cells. *Nanomedicine*, 6(1), 153–160. <https://doi.org/10.1016/j.nano.2009.05.009>
- Hamlouli, I., Bencheraiet, R., Bensegueni, R., & Bencharif, M. (2017). Experimental and theoretical study on DPPH radical scavenging mechanism of some chalcone quinoline derivatives. *Journal of Molecular Structure*, 1156, 385–389. <https://doi.org/10.1016/j.molstruc.2017.11.118>
- Jahanban-Esfahlan, A., Dastmalchi, S., & Davaran, S. (2016). A simple improved desolvation method for the rapid preparation of albumin nanoparticles. *International Journal of Biological Macromolecules*, 91, 703–709. <https://doi.org/10.1016/j.ijbiomac.2016.05.032>
- Kaminaga, Y., Nagatsu, A., Akiyama, T., Sugimoto, N., Yamazaki, T., Maitani, T., & Mizukami, H. (2003). Production of unnatural glucosides of curcumin with drastically enhanced water solubility by cell suspension cultures of *Catharanthus roseus*. *FEBS Letters*, 555(2), 311–316. [https://doi.org/10.1016/S0014-5793\(03\)01265-1](https://doi.org/10.1016/S0014-5793(03)01265-1)
- Karimian, M. S., Pirro, M., Johnston, T. P., Majeed, M., & Sahebkar, A. (2017). Curcumin and endothelial function: Evidence and mechanisms of protective effects. *Current Pharmaceutical Design*, 23(17), 2462–2473. <https://doi.org/10.2174/1381612823666170222122822>

- Ko, W. C., Chang, C. K., Wang, H. J., Wang, S. J., & Hsieh, C. W. (2015). Process optimization of microencapsulation of curcumin in  $\gamma$ -polyglutamic acid using response surface methodology. *Food Chemistry*, 172, 497–503. <https://doi.org/10.1016/j.foodchem.2014.09.100>
- Kumar, A., & Ahuja, M. (2013). Carboxymethyl gum kondagogu-chitosan polyelectrolyte complex nanoparticles: Preparation and characterization. *International Journal of Biological Macromolecules*, 62(1), 80–84. <https://doi.org/10.1016/j.ijbiomac.2013.08.035>
- Letchford, K., & Burt, H. (2007). A review of the formation and classification of amphiphilic block copolymer nanoparticulate structures: Micelles, nanospheres, nanocapsules and polymersomes. *European Journal of Pharmaceutics & Biopharmaceutics*, 65(3), 259–269. <https://doi.org/10.1016/j.ejpb.2006.11.009>
- Liang, H., Huang, Q., Zhou, B., He, L., Lin, L., An, Y., ... Li, B. (2015). Self-assembled zein-sodium carboxymethyl cellulose nanoparticles as an effective drug carrier and transporter. *Journal of Materials Chemistry B*, 3(16), 3242–3253. <https://doi.org/10.1039/C4TB01920B>
- Lin, L., Xu, W., Liang, H., He, L., Liu, S., Li, Y., ... Chen, Y. (2015). Construction of pH-sensitive lysozyme/pectin nanogel for tumor methotrexate delivery. *Colloids & Surfaces B Biointerfaces*, 126, 459–466. <https://doi.org/10.1016/j.colsurfb.2014.12.051>
- Lin, L., Xu, W., Liang, H., He, L., Liu, S., Li, Y., ... Chen, Y. (2015). Construction of pH-sensitive lysozyme/pectin nanogel for tumor methotrexate delivery. *Colloids and Surfaces B: Biointerfaces*, 126, 459–466. <https://doi.org/10.1016/j.colsurfb.2014.12.051>
- Mangalathillam, S., Rejinold, N. S., Nair, A., Lakshmanan, V. K., Nair, S. V., & Jayakumar, R. (2011). Curcumin loaded chitin nanogels for skin cancer treatment via the transdermal route. *Nanoscale*, 4(1), 239–250. <https://doi.org/10.1039/c1nr11271f>
- Moussa, Z., Hmadeh, M., Abiad, M. G., Dib, O. H., & Patra, D. (2016). Encapsulation of curcumin in cyclodextrin-metal organic frameworks: Dissociation of loaded CD-MOFs enhances stability of curcumin. *Food Chemistry*, 212, 485–494. <https://doi.org/10.1016/j.foodchem.2016.06.013>
- Naksuriya, O., Shi, Y., van Nostrum, C. F., Anuchapreeda, S., Hennink, W. E., & Okonogi, S. (2015). HPMA-based polymeric micelles for curcumin solubilization and inhibition of cancer cell growth. *European Journal of Pharmaceutics & Biopharmaceutics*, 94(141), 501–512. <https://doi.org/10.1016/j.ejpb.2015.06.010>
- Nazari-Vanani, R., Moezi, L., & Heli, H. (2017). In vivo evaluation of a self-nanoemulsifying drug delivery system for curcumin. *Biomedicine & Pharmacotherapy*, 88, 715–720. <https://doi.org/10.1016/j.biopha.2017.01.102>
- Nimiya, Y., Wang, W., Du, Z., Sukamtoh, E., Zhu, J., Decker, E., & Zhang, G. (2016). Redox modulation of curcumin stability: Redox active antioxidants increase chemical stability of curcumin. *Molecular Nutrition & Food Research*, 60(3), 487–494. <https://doi.org/10.1002/mnfr.201500681>
- Paramera, E. I., Konteles, S. J., & Karathanos, V. T. (2011). Stability and release properties of curcumin encapsulated in *Saccharomyces cerevisiae*,  $\beta$ -cyclodextrin and modified starch. *Food Chemistry*, 125(3), 913–922. <https://doi.org/10.1016/j.foodchem.2010.09.071>
- Refat, M. S. (2013). Synthesis and characterization of ligational behavior of curcumin drug towards some transition metal ions: Chelation effect on their thermal stability and biological activity. *Spectrochimica Acta Part A Molecular & Biomolecular Spectroscopy*, 105, 326–337. <https://doi.org/10.1016/j.saa.2012.12.041>
- Saberi-Karimian, M., Parizadeh, S. M. R., Ghayour-Mobarhan, M., Salahshoh, M. M., Dizaji, B. F., Safarian, H., ... Ahmadinejad, M. (2018). Evaluation of the effects of curcumin in patients with metabolic syndrome. *Comparative Clinical Pathology*, 27(3), 555–563. <https://doi.org/10.1007/s00580-017-2624-y>
- Sneharani, A. H., Karakkat, J. V., Singh, S. A., & Rao, A. G. (2010). Interaction of curcumin with  $\beta$ -lactoglobulin-stability, spectroscopic analysis, and molecular modeling of the complex. *Journal of Agricultural & Food Chemistry*, 58(20), 11130–11139. <https://doi.org/10.1021/jf102826q>
- Tapal, A., & Tiku, P. K. (2012). Complexation of curcumin with soy protein isolate and its implications on solubility and stability of curcumin. *Food Chemistry*, 130(4), 960–965. <https://doi.org/10.1016/j.foodchem.2011.08.025>
- Tonelli, A., Candiani, A., Sozzi, M., Zucchelli, A., Foresti, R., Dall'Asta, C., ... Cucinotta, A. (2019). The geek and the chemist: Antioxidant capacity measurements by DPPH assay in beverages using open source tools, consumer electronics and 3D printing. *Sensors and Actuators B: Chemical*, 282, 559–566. <https://doi.org/10.1016/j.snb.2018.11.019>
- Wang, X., Deng, J., Yuan, J., Tang, X., Wang, Y., Chen, H., ... Zhou, L. (2017). Curcumin exerts its tumor suppressive function via inhibition of NEDD4 oncoprotein in glioma cancer cells. *International Journal of Oncology*, 51(2), 467–477. <https://doi.org/10.3892/ijo.2017.4037>
- Xu, W., Ge, P., Huang, L., Du, Z., Liu, X., Li, J., & Yang, N. (2018). Solubilization and protection of curcumin based on lysozyme/albumin nano-complex. *AIP Advances*, 8(11), 115221. <https://doi.org/10.1063/1.5046534>
- Xu, W., Huang, L., Jin, W., Ge, P., Shah, B. R., Zhu, D., & Jing, J. (2019). Encapsulation and release behavior of curcumin based on nanoemulsions-filled alginate hydrogel beads. *International Journal of Biological Macromolecules*, 134, 210–215. <https://doi.org/10.1016/j.ijbiomac.2019.04.200>
- Xu, W., Jin, W., Huang, K., Huang, L. U., Lou, Y., Li, J., ... Li, B. (2018). Interfacial and emulsion stabilized behavior of lysozyme/xanthan gum nanoparticles. *International Journal of Biological Macromolecules*, 117, 280–286. <https://doi.org/10.1016/j.ijbiomac.2018.05.187>
- Xu, W., Jin, W., Li, Z., Liang, H., Wang, Y., Shah, B. R., ... Li, B. (2015). Synthesis and characterization of nanoparticles based on negatively charged xanthan gum and lysozyme. *Food Research International*, 71, 83–90. <https://doi.org/10.1016/j.foodres.2015.02.007>
- Xu, W., Jin, W., Zhang, C., Li, Z., Lin, L., Huang, Q., ... Li, B. (2014). Curcumin loaded and protective system based on complex of  $\kappa$ -carrageenan and lysozyme. *Food Research International*, 59, 61–66. <https://doi.org/10.1016/j.foodres.2014.01.059>
- Yu, H., & Huang, Q. (2010). Enhanced in vitro anti-cancer activity of curcumin encapsulated in hydrophobically modified starch. *Food Chemistry*, 119(2), 669–674. <https://doi.org/10.1016/j.foodchem.2009.07.018>
- Zhao, Y., Pan, Y., Nitin, N., & Tikekar, R. V. (2014). Enhanced stability of curcumin in colloidosomes stabilized by silica aggregates. *LWT - Food Science and Technology*, 58(2), 667–671. <https://doi.org/10.1016/j.lwt.2014.03.017>
- Zhu, K., Ye, T., Liu, J., Peng, Z., Xu, S., Lei, J., ... Li, B. (2013). Nanogels fabricated by lysozyme and sodium carboxymethyl cellulose for 5-fluorouracil controlled release. *International Journal of Pharmaceutics*, 441(1), 721–727. <https://doi.org/10.1016/j.ijpharm.2012.10.022>
- Zou, L., Zheng, B., Zhang, R., Zhang, Z., Liu, W., Liu, C., ... McClements, D. J. (2016). Influence of lipid phase composition of excipient emulsions on curcumin solubility, stability, and bioaccessibility. *Food Biophysics*, 11(3), 213–225. <https://doi.org/10.1007/s11483-016-9432-9>

**How to cite this article:** Cui J, Zhou J, Huang L, Jing J, Wang N, Wang L. Curcumin encapsulation and protection based on lysozyme nanoparticles. *Food Sci Nutr*. 2019;7:2702–2707. <https://doi.org/10.1002/fsn3.1129>