

The Role of Insulin Receptor Substrate Proteins in Bronchopulmonary Dysplasia and Asthma: New Potential Perspectives

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Abstract: Insulin receptor substrates (IRSs) are proteins that are involved in signaling through the insulin receptor (IR) and insulin-like growth factor (IGFR). They can also interact with other receptors including growth factor receptors. Thus, they represent a critical node for the transduction and regulation of multiple signaling pathways in response to extracellular stimuli. In addition, IRSs play a central role in processes such as inflammation, growth, metabolism, and proliferation. Previous studies have highlighted the role of IRS proteins in lung diseases, in particular asthma. Further, the members of the IRS family are the common proteins of the insulin growth factor signaling cascade involved in lung development and disrupted in bronchopulmonary dysplasia (BPD). However, there is no study focusing on the relationship between IRS proteins and BPD yet. Unfortunately, there is still a significant gap in knowledge in this field. Thus, in this review, we aimed to summarize the current knowledge with the major goal of exploring the possible roles of IRS in BPD and asthma to foster new perspectives for further investigations.

Keywords: insulin receptor substrates; asthma; bronchopulmonary dysplasia; pediatric lung disease

1. Introduction

Insulin receptor (IR) and insulin-like growth factor (IGF) receptor (IGFR) signaling regulate a variety of cellular processes including glucose metabolism, differentiation, and cell growth [1]. Although insulin and IGF are commonly involved in induction processes and overlapping signaling pathways due to shared downstream elements during biological processes, they may also harbor unique effects in cellular physiology [2]. Insulin and IGF signaling pathways are mainly known as metabolic regulators in the cell, but they are also implicated in allergic lung and developmental disorders related to metabolic diseases [3,4]. Further, recent studies have elaborated that the dysregulation of IGF signaling pathways may contribute to the disrupted development of the lung and lung disorders including lung cancer [5]. Common adaptor molecules that transmit extracellular signaling through IGFR and IR are insulin receptor substrate (IRS) proteins [6]. Although they were discovered to be substrates of IR, they can also interact with other receptors such as those for growth hormones, cytokines, integrins, and vascular endothelial growth factors (VEGF) [7]. They generate a critical node for the regulation of multiple signaling pathways depending on



Citation: Gorgisen, G.; Aydin, M.; Mboma, O.; Gökyildirim, M.Y.; Chao, C.-M. The Role of Insulin Receptor Substrate Proteins in Bronchopulmonary Dysplasia and Asthma: New Potential Perspectives. *Int. J. Mol. Sci.* 2022, 23, 10113. https://doi.org/10.3390/ijms 231710113

Academic Editor: Cenk Suphioglu

Received: 14 August 2022 Accepted: 1 September 2022 Published: 4 September 2022

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Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). extracellular stimuli and signal diversion. In addition, they have pivotal roles in processes including inflammation, growth, metabolism, and proliferation [8]. Previous studies differentially emphasized the role of IRS proteins in the development of several diseases, e.g., allergic lung inflammation and diabetes [3,9–11]. However, there is still a significant gap in knowledge on how IRSs are related to the development of pediatric lung diseases such as asthma and bronchopulmonary dysplasia (BPD). This review summarizes the current evidence of IRS possibly being involved in asthma and BPD.

2. The Biological Role of Insulin Receptor Substrate Proteins in Human Tissue

The insulin receptor substrate family consists of six members, IRS1–6 [12]. All family members have evolutionary conserved two domains in their N-terminus. The Plecstrin homology (PH) domain modulates the interaction of IRS proteins with the cell membrane and induces the translocation of IRS1 from the cytosol to the nucleus through the nuclear localization signal located in the PH domain [13–15]. The Phosphotyrosine binding (PTB) domain is another conserved region located in the N-terminal of the IRS protein that plays a central role in the activation of IRS proteins after binding to tyrosine phosphorylated insulin and other receptors [13–15]. Moreover, there are insufficiently conserved regions at the carboxy-terminal ends of the IRS proteins. These contain various tyrosine, serine, and threonine motifs that regulate signal transduction and diversion [12]. This region contributes to the unique specificity owing to the unique regulation of each IRS protein. Importantly, IRS5 and IRS6 have a truncated C-terminal compared to the other members of the family [16–18].

In detail, the IRS members IRS1, IRS2, IRS5, and IRS6, are widely expressed in human tissues, whereas IRS4 presents a tissue-restricted pattern including the brain, thymus, and embryonic tissues. IRS3 acts as a pseudogene and is not expressed in humans [17–21].

The IRS signaling pathway starts with the binding of ligands such as insulin and the insulin-like growth factor (IGF) to their corresponding receptors. This leads to autophosphorylation of the receptor via tyrosine residues (Figure 1) [22]. IRS proteins do not have kinase or other intrinsic enzymatic activity; however, they become phosphorylated by receptor tyrosine kinases following their interactions [23]. The tyrosine phosphorylation of IRS proteins triggers the transmission of signals from receptors to downstream targets in the cytosol. However, the tyrosine phosphorylation of IRS proteins mainly induces two signaling pathways including MAPK and PI3K-AKT [24].

In detail, the PI3K-AKT pathway is a critical node for cell signaling pathways [25]. It regulates many cellular processes such as metabolism, survival, and apoptosis [26]. IRS proteins contain YXXM motifs and phosphorylation of the tyrosine residues of the motifs YXXM leads to the activation of PI3K and the induction of mTOR and AKT activations (Figure 1) [24].

In addition to the PI3K-AKT pathway, upon activation of the IRS proteins, Grb2 binds to the YVNI motifs of the IRS proteins and triggers the activation of RAS-RAF-ERK1/2 signaling cascades (Figure 1) that regulate the expression of the genes responsible for cell proliferation and differentiation [27].

IRS proteins contain more than 50 S/T phosphorylation sites for the several IRS kinases at their COOH terminals [28,29]. In contrast to tyrosine phosphorylations, S/T phosphorylations usually inhibit signal transduction [10]. Under normal conditions, these phosphorylations balance each other, are well-regulated, and occur as a physiological negative feedback mechanism mainly by downstream targets of IRS proteins [29]. In addition to these downstream elements, other signaling mediators can also phosphorylate IRS proteins at S/T motifs such as TNF- α , NF- κ B, and JNK [30–33]. The dysregulation of IRS S/T phosphorylations leads to the development of pathological conditions such as insulin resistance, Type 2-diabetes, cancer, and inflammatory diseases [11,24,33].

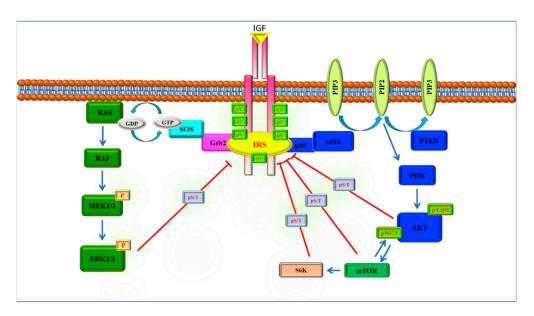


Figure 1. Canonical IRS signaling pathway adapted from [24]. IRS proteins mainly induce the activations of the PI3K-AKT and MAPK pathways. After the binding of receptor tyrosine kinases, IRS proteins are phosphorylated by their tyrosine residues and this activation triggers the IRS-induced signaling pathways. Activations of IRS proteins are regulated through Ser/Thr phosphorylations as a feedback mechanism of their downstream targets.

3. Possible Roles of IRS in Pediatric Lung Diseases

3.1. Pediatric Asthma

As a chronic inflammatory disease, asthma plays a special role among lung disorders. The exact etiology of asthma remains not fully understood, but it has a multifactorial and heterogeneous background where genetic and environmental factors have pivotal roles [34]. In recent decades, asthma has been increasingly recognized as a 'syndrome' with different etiologies and underlying pathophysiologic mechanisms, and it is subclassified into several subgroups or phenotypes [35–40]. Currently, one of the widest and best characterized endotype is allergic (eosinophilic) asthma with a type 2 immune response/inflammation, including type 2 T-helper cell lymphocytes (Th2) and other involved cell subsets [35,41]. However, chronic lower airway inflammation in asthma is caused by the infiltration of inflammatory cells, such as eosinophils, neutrophils, and T-helper cells, as well as the activation of mast cells. In addition, IgE production triggered by B lymphocytes and epithelial cell damage are some aspects occurring during the pathogenesis of asthma [41–43].

In detail, during the sensitization phase, antigen-presenting cells (APCs), mainly dendritic cells (DCs), internalize allergens that have passed the epithelial barrier and migrate toward the draining lymph nodes [41,44–46]. DCs present allergen-derived peptides to naive T-helper lymphocytes (CD4⁺) through major histocompatibility complex type II (MHC-II) in conjunction with costimulatory molecules such as CD80, CD86, and OX40L. Together with IL-4, this process triggers the differentiation of CD4⁺ to Th2 and T follicular helper cells (Tfh), which favors the humoral antibody production of IgE by inducing B cell isotypic commutation and plasma cell differentiation [41,45–48]. Th2 produces proinflammatory cytokines, also known as type 2 cytokines, which promote IgE production, activation of eosinophils (IL-5), mast cell development (IL-9), and airway hyperresponsiveness (IL-13) [41,46,47].

In the late phase, locally produced chemokines may cause the recruitment of macrophages, eosinophils, neutrophils, and Th2 lymphocytes [41,42,44]. The latter release specific cytokines such as IL-4, IL-5, IL-9, and IL-13, as well as a granulocyte-macrophage colony-stimulating factor (GM-CSF), which contribute to the maintenance of inflammation and can lead to remodeling, fibrosis, and hyperplasia [41–44,49–51].

Understanding the precise contribution of these signaling pathways to M2 macrophage polarization is a key factor in the search for potential therapeutic strategies to disrupt the proinflammatory signaling pathways in severe asthma and allergic diseases in the future [11,52,53].

3.2. Pediatric Asthma and IRS Signaling: A Forgotten Gap?

IL-4 and IL-13 have multiple important functions including the regulation of allergic responses [54]. As stated above, IRSs have been shown to be involved in allergic inflammation as well. Therefore, it is important to understand the interactions between IRSs, IL-4, and IL-13 proteins to explore the possible roles implicated in asthma. IRS1 was identified as a tyrosine-phosphorylated large molecular weight protein after insulin stimulation while IRS2 was determined in Factor Dependent Continuous-Paterson 2 (FDC-P2) cells after IL-4 treatment [55–57]. In IL-4 and IL-13 signaling, Janus kinase (JAK) proteins are required for the tyrosine phosphorylation of the IRS proteins (Figure 2) [58].

Recently, IRS2 has received increasing attention due to its role in the signaling cascade of type 2 cytokines, such as IL-4 and IL-13 [11,52]. IRS2, similar to its homolog IRS1, is an approximately 170 kDa adaptor protein that mediates the downstream signaling of receptor tyrosine kinases [11,59,60]. It is a member of the insulin receptor substrate family and is involved in signaling through IGF-1, insulin, and EPO, among others, and therefore plays a role in insulin-induced responses [11,59,60]. While its role in promoting insulin resistance and type II diabetes has been explored and its function as a regulator of M2 macrophage polarization has been described, not much is known about its precise contribution to the pathophysiology of asthma/allergy through the IL-4 pathway [11,52,60–62].

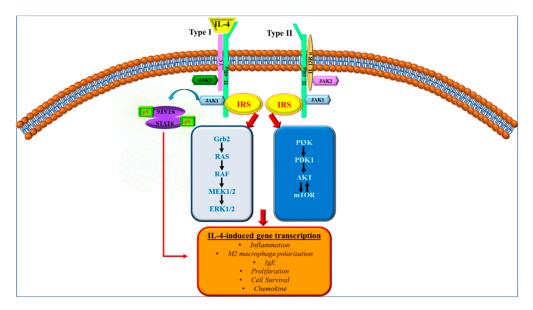


Figure 2. Interleukin-4-induced gene transcriptional effects through IRS and STAT6 activations. Upon binding of IL-4, tyrosine phosphorylated IL receptors generate binding sites for JAK1/2 and IRS proteins. JAK1/2 are mainly responsible for the activations of IRSs and STAT6 that trigger the IL4 -induced cellular effects such as inflammation, cell survival, and proliferation (adapted from [61]).

During IL-4 and IL-13 signal transductions, IL-4 binds to IL4R α and induces the dimerization with the IL-4 γ chain or IL13R α 1, while IL-13 interacts with IL13R α 1 and leads to the 'heterodimerization' of IL4R- α [63]. These complexes trigger the activation of JAK that phosphorylate the specific tyrosine motifs [63]. During these auto- and cross-phosphorylations, JAK1, JAK2, and JAK3 consequently interact with IL4R- α , IL13R- α 1, and - γ -chain, respectively, which are crucial to present docking sites for IRS proteins [64]. The IRS proteins specifically bind to the tyrosine phosphorylated NPXY motif of the cytoplasmic tail of the receptors, which is also defined as the insulin and IL-4 receptor (I4R) motif [65].

The mutation of this motif may block the association between receptor and IRS proteins and inhibits the proliferation effect of IL4R- α signaling [65]. In the IL4R- α signaling pathway, IRS1 and IRS2 are activated by an IL-4 treatment [66]. The activation of IRS1 and IRS2 can trigger distinct signaling cascades including PI3K, AKT, and MAPK, which was already mentioned above. In addition to IRS proteins, STAT6 also binds to the IL4R- α chain upon ligand activation [67]. The activation of STAT6 induces the translocation of STAT6 from the cytosol to the nucleus and mainly regulates gene expressions [68] (Figure 2).

One of the downstream effectors of IRS2 is the $p85\alpha$ subunit of phosphatidylinositol 3-kinase (PI3K), which in turn activates several downstream signaling pathways and regulates epithelial cell migration, suggesting that IL-4 may have a beneficial effect on airway epithelial cell repair via recruitment of the IRS2 pathway [66]. Interestingly, further M2 gene expression, occurs only when activation of the IRS2 signaling pathways occurs via the type 1 IL-4 receptor rather than type II [53,62].

While tyrosine phosphorylation appears to activate IRS2 signaling, serine phosphorylation does the opposite by inhibiting p85 α binding and PI3K activation and promoting IRS2 degradation [59,62]. The precise pathway involves two targets of rapamycin kinase complex 1 (TORC1)-activated proteins, namely GRB10 and p70S6K, the common γ -chain (γ C), and IL-4R α , and the latter initiating the serine phosphorylation of IRS2 [62]. The end result of these two negative regulatory mechanisms is a reduction in the M2 polarization of human macrophages in diseases such as asthma [62]. Another potential process for the downregulation of IRS2 is the Suppressors of Cytokine Signaling Protein Family (SOCS) 1 [53]. This protein family is induced by JAK-STAT activation, nuclear displacement of Signal Transducers and Activators of Transcription (STAT)6, and intracellular signaling through PI3K [53]. SOCS1 inhibits a negative feedback loop by forming an E3 ubiquitin ligase with other proteins, increasing the poly-ubiquitination of phosphorylated IRS2 and promoting the proteasomal degradation of IRS2 [53]. In addition, SOCS1 expression in response to IL-4 appears to be decreased in allergic individuals compared with healthy individuals [53].

3.3. Bronchopulmonary Dysplasia

Bronchopulmonary dysplasia is the most common pulmonary complication in infants born before 30 weeks of gestational age and contributes to long-term morbidity and mortality [69]. According to the severity-based definition of BPD, up to 30–40% of preterm neonates born with a gestational age \leq 28 weeks suffer from BPD [69]. The pathogenesis of this chronic pulmonary disease is multifactorial and most commonly seen in premature infants undergoing mechanical ventilation with oxygen therapy (old BPD). Furthermore, other risk factors such as intrauterine growth restrictions and pre-/postnatal infections may lead to a pulmonary growth arrest resulting in alveolar simplification and pulmonary vasculature disturbance [70–72]. Clinically, BPD is defined by the need for supplemental oxygen or ventilator support at day 28 of life or 36 weeks of gestational age [73]. Despite numerous advances in neonatal care leading to improvement in the survival of preterm infants, options for the prevention and treatment of BPD are still very limited, and curative therapy is still lacking [74]. The development of exogenous surfactant protein application supporting preterm infants to breathe postnatally allowed a less injurious ventilation strategy and has changed the histomorphological phenotype of BPD (new BPD) defined by a decrease in alveologenesis and reduction in small vessel development, alterations in the growth factor signaling, and extracellular matrix changes [72,75].

Hyperoxic injury is one of the main risk factors in BPD pathogenesis, which is thought to disrupt critical signaling pathways that induce lung development, including branching and septation [76]. Several signaling pathways contributing to these processes have been described, such as IGFR [5], FGF10 [77,78], and TGF- β [79] signaling pathways, where the IRS proteins might serve as a common component (Figure 3) [80,81].

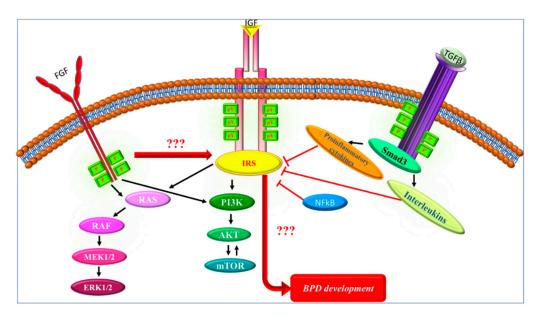


Figure 3. Hypothetical role of IRS proteins in BPD development. IGFR, FGF, and TGF- β signaling pathways have pivotal roles in BPD. IRS proteins are the common proteins of these signaling pathways. In addition to the downstream targets of IRS proteins, IRS-related signaling pathway members such as proinflammatory cytokines, NF- κ B, and interleukins inhibit IRS signaling through Ser/Thr phosphorylations. These inhibitions may have an important role in the development of BPD.

3.4. Aspects of IRS Signaling Possibly Involved in BPD

IRS proteins are the main adaptor molecules of IGF signaling [8,82]. One of the main risk factors of BPD is hyperoxia. Hyperoxia interferes with IGF-1 signaling [5,83,84] and inhibits IGF-1/IGF-1R signal transduction by decreasing the binding affinity in the lungs of preterm infants or the distal epithelial cells in fetal lungs [5]. Decreasing the binding affinity of IGF results in impaired proliferation, formation of secondary ridges, and alveologenesis [85,86]. Although it may seem paradoxical at first, in vivo experiments have shown that mice are protected from 90% oxygen-induced lung injury when IGF-1R expression is reduced and leads to less edema, vascular extravasation, and respiratory failure compared to control mice [87].

IGF signaling is important for normal lung development. Thus, in in vivo experiments, components of IGF signaling were shown to have different expression patterns in different compartments during embryogenesis [88].

Serum IGF-1 levels are significantly reduced in patients with BPD [89]. However, the expression of IGF-1 in epithelial mucosal fluid, epithelial cells, and peribronchial myofibroblasts was increased in BPD [90,91]. This was also demonstrated in the hyperoxia-stimulated ex vivo model of neonatal rat lung. The IGF-1 staining was most pronounced in the airway and alveolar epithelial cells [85]. The cell proliferation was associated with increased IGF-1 mRNA and protein expression and could be inhibited by IGF-1 antibody [85].

It has been shown in previous studies that the downstream targets of IGF signaling play a role in the development of BPD [92]. However, the effect of IRS in relation to IGF in the context of BPD has not been explored. The PI3K-AKT pathway is one downstream target of IGF signaling (Figure 3) [5]. Several studies have shown the relevance of PI3K in parenchymal cells in lung disease [93]. In particular, in adult respiratory distress syndrome (ARDS), the PI3K-dependent activation of PKB in lung endothelial cells was shown to be triggered by overventilation [93]. This overventilation causes a PI3K-dependent shift of NF- κ B which can decrease *Fg10* expression during the development of BPD [94–96].

It is also known that the activation of PI3K/AKT via hydrogen has a protective effect on alveolar epithelial cells type 2 (AEC2) during hyperoxia treatment in animal experiments [97].

The mammalian target of rapamycin (mTOR) is a part of the PI3K/AKT pathway and can be induced by hyperoxia [98–101]. It was shown that inhibition of the mTOR signaling pathway suppresses the proliferation of lung fibroblasts [102]. It is also described that blocking from the mTOR pathway could downregulate the TGF- β expression [103].

ERK1/2 is the other main downstream target of IGF signaling (Figure 3). It has been demonstrated that hyperoxia can activate ERK1/2 in in vivo models [104,105].

Fgf10 is one of the most significant developmental genes expressed in the submesothelial mesenchyme of the developing lung [106,107]. Fgf10 encodes a secreted diffusible protein that acts in a paracrine manner through the epithelial receptor Fgfr2b [108,109]. Most important, FGF10 has been shown to be downregulated in children suffering from fatal severe BPD [110]. There is growing evidence supporting a close interaction between lung vasculature and branching morphogenesis via endothelial-epithelial crosstalk [77,111,112]. It was also demonstrated that Fgf10 is critical for the differentiation of the Fgf10-positive progenitor cells towards the lipofibroblast lineage, a subset of fibroblast believed to play an important role in de novo alveologenesis during regeneration after lung injury [113]. Although previous studies have shown that FGF family members induce the activation of IRS1 and IRS2 proteins through FGFR and IGFR receptors, we do not yet know the exact role of IRS proteins in the development of BPD through the activation of these receptors [114]. Other studies mainly focused on the role of FGF and IRS1 signaling pathways in cancer and different diseases [115,116]. In MCF7 cells, after treatment with FGF, IRS1 expression showed a rapidly increasing profile [117]. In a study regarding the role of insulin resistance in chronic liver disease, Manzano-Nunez and colleagues revealed IRS2 as a positive regulator for the intercellular interaction and the transition from stromal to epithelial repair via Fgf7-Fgfr2b signaling [118].

TGF- β takes a central role in postnatal lung development and alveologenesis. TGF- β signaling is activated by the binding of TGF- β to the type II TGF- β receptor (T β RII) [119]. This complex subsequently binds to one of two variants of the type I receptor (ALK-1 or ALK-5) [119]. The type I receptor transmits signals into the cell by the second-messenger SMAD proteins, namely SMAD1 m and SMAD3 [119,120]. This happens in combination with the co-SMAD, SMAD4, or by SMAD-independent pathway [119,120]. In vivo experiments have revealed that TGF- β also acts as a positive regulator of branching and alveologenesis [121–123]. However, experiments in various in vivo models have shown that the increased expression of TGF- β and activation of the TGF- β signaling pathways are associated with the onset and development of BPD [124–127].

As increased pulmonary TGF-β expression precedes most BPD-related pathophysiological manifestations and infiltration of neutrophils and monocytes into the lungs occurs during disease development [127–129]. These recruited monocytes and macrophages are the main source of TGF- β [130], which leads to an increase in secreted pro-inflammatory cytokines such as many interleukins (IL), e.g., IL-1 β , IL-6, IL-8, and TNF- α [79,131,132]. The development of pulmonary edema is also associated with anti-inflammatory factors, including IL-4, IL-10, IL-12, and IL-13 or the IL-1 receptor antagonist [133–135]. A previous study showed that IRS1 and IRS2 are the key molecules in the T β R-V/LRP-1-induced growth inhibition in mink lung epithelial cells [136]. Another study that focused on the interaction between TGF- β and IRS signaling showed that IRS1 may suppress TGF- β induced epithelial-mesenchymal transition in A549 cells (non-small cell lung cancer cell line) through inhibition of Snail and Slug expressions [137]. In contrast, TGF- β inhibits cell proliferation and increases apoptosis via inhibition of IRS1 expression and activation in colon cancer cells [138]. All these studies focused on cancer development and mainly showed the negative regulation between IRS signaling and TGF- β signaling. This regulation may also have a role in the development of BPD.

Although IRS family members are the common proteins for the IGF signaling cascade, there is no study that focuses on the relationship between IRS proteins and BPD. Previous studies showed that ERK1/2, AKT, and mTOR phosphorylate IRS proteins at Ser/Thr

motifs and negatively regulate the IRS protein's actions (Figure 3). Therefore, IRS proteins may have a pivotal role in the development of BPD, but this needs to be further investigated.

4. Conclusions

It has been extensively shown that IRS proteins are major players involved in many processes such as inflammation, growth, metabolism, and proliferation. However, their roles in lung disorders are insufficiently understood. Here, we have comprehensively summarized the current knowledge on how IRS might be possibly involved in the pathogenesis of asthma and BPD, focusing on the signaling pathways of IGF, IL-4, FGF, and TGF- β receptors. Even though some of the evidence is still weak and hypothetical, it allows new perspectives for further investigations in order to shed more light on the role of IRS in asthma, BPD, and other pediatric lung diseases.

Author Contributions: Project idea, G.G., M.A., and C.-M.C.; conceptualization, G.G., M.A., M.Y.G., and C.-M.C.; supervision and project administration, G.G., M.A., and C.-M.C.; systematic literature search, all authors; methodology, G.G., M.A., O.M., M.Y.G., and C.-M.C.; formal analyses and interpretation of data, G.G., M.A., O.M., M.Y.G., and C.-M.C.; writing—original draft preparation, all authors; writing—review and editing, G.G., M.A., O.M., M.Y.G., and C.-M.C.; writing—revision of the manuscript, G.G., M.A., M.Y.G., and C.-M.C. All authors have read and agreed to the published version of the manuscript.

Funding: M.A. received funding from the Internal Research Grant of the Faculty of Health at Witten/Herdecke University, Germany (project number: IFF 2020-02). C.-M.C. is supported by the DFG grant (CH2361/2-1) and the University Medical Center Rostock.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Conflicts of Interest: The authors declare no conflict of interest. The funders had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript, or in the decision to publish the results.

Abbreviations

AEC2 = alveolar epithelial cell type 2; APCs = antigen-presenting cells; BPD = bronchopulmonary dysplasia; DCs = dendritic cells; FDC-P2 = Factor Dependent Continuous-Paterson 2; GM-CSF = granulocyte-macrophage colony-stimulating factor; I4R = IL-4 receptor; IGFR = insulinlike growth factor receptor; IR = Insulin receptor; IRS= insulin receptor substrates; JAK = Janus kinase; mTOR = Mammalian target of rapamycin; PH = Plecstrin homology; PTB = Phosphotyrosine binding; SOCS = Suppressors of Cytokine Signaling Protein Family; STAT = Signal Transducers and Activators of Transcription; T β RII = TGF- β receptor; Tfh = T follicular helper cells; Th2 = type 2 T-helper cell lymphocyte; TORC1 = target of rapamycin kinase complex 1; TYK = tyrosine kinases; VEGF = Vascular endothelial growth factor

References

- 1. Cao, J.; Yee, D. Disrupting Insulin and IGF Receptor Function in Cancer. Int. J. Mol. Sci. 2021, 22, 555. [CrossRef] [PubMed]
- 2. Okuyama, T.; Kyohara, M.; Terauchi, Y.; Shirakawa, J. The Roles of the IGF Axis in the Regulation of the Metabolism: Interaction and Difference between Insulin Receptor Signaling and IGF-I Receptor Signaling. *Int. J. Mol. Sci.* **2021**, *22*, 6817. [CrossRef]
- 3. Guo, S. Insulin signaling, resistance, and the metabolic syndrome: Insights from mouse models into disease mechanisms. *J. Endocrinol.* **2014**, 220, T1–T23. [CrossRef] [PubMed]
- Han, Y.Y.; Yan, Q.; Chen, W.; Forno, E.; Celedon, J.C. Serum insulin-like growth factor-1, asthma, and lung function among British adults. Ann. Allergy Asthma Immunol. 2021, 126, 284–291.e282. [CrossRef] [PubMed]
- Wang, Z.; Li, W.; Guo, Q.; Wang, Y.; Ma, L.; Zhang, X. Insulin-Like Growth Factor-1 Signaling in Lung Development and Inflammatory Lung Diseases. *Biomed Res. Int.* 2018, 2018, 6057589. [CrossRef]

- 6. Gorgisen, G.; Karatas, U.; Ates, C.; Oksuz, M.; Gulacar, I.M. Association of IRS1 Gly972Arg and IRS2 Gly1057Asp polymorphisms with gastric cancer in Turkish subjects. *Oncol. Lett.* **2020**, *20*, 2016–2020. [CrossRef]
- Machado-Neto, J.A.; Fenerich, B.A.; Rodrigues Alves, A.P.N.; Fernandes, J.C.; Scopim-Ribeiro, R.; Coelho-Silva, J.L.; Traina, F. Insulin Substrate Receptor (IRS) proteins in normal and malignant hematopoiesis. *Clinics* 2018, 73, e566s. [CrossRef]
- Taniguchi, C.M.; Emanuelli, B.; Kahn, C.R. Critical nodes in signaling pathways: Insights into insulin action. *Nat. Rev. Mol. Cell Biol.* 2006, 7, 85–96. [CrossRef]
- 9. Lavin, D.P.; White, M.F.; Brazil, D.P. IRS proteins and diabetic complications. Diabetologia 2016, 59, 2280–2291. [CrossRef]
- Gorgisen, G.; Hapil, F.Z.; Yilmaz, O.; Cetin, Z.; Pehlivanoglu, S.; Ozbudak, I.H.; Erdogan, A.; Ozes, O.N. Identification of novel mutations of Insulin Receptor Substrate 1 (IRS1) in tumor samples of non-small cell lung cancer (NSCLC): Implications for aberrant insulin signaling in development of cancer. *Genet. Mol. Biol.* 2019, 42, 15–25. [CrossRef]
- 11. Dasgupta, P.; Dorsey, N.J.; Li, J.; Qi, X.; Smith, E.P.; Yamaji-Kegan, K.; Keegan, A.D. The adaptor protein insulin receptor substrate 2 inhibits alternative macrophage activation and allergic lung inflammation. *Sci. Signal.* **2016**, *9*, ra63. [CrossRef] [PubMed]
- 12. Hanke, S.; Mann, M. The phosphotyrosine interactome of the insulin receptor family and its substrates IRS-1 and IRS-2. *Mol. Cell. Proteom.* **2009**, *8*, 519–534. [CrossRef] [PubMed]
- Voliovitch, H.; Schindler, D.G.; Hadari, Y.R.; Taylor, S.I.; Accili, D.; Zick, Y. Tyrosine phosphorylation of insulin receptor substrate-1 in vivo depends upon the presence of its pleckstrin homology region. *J. Biol. Chem.* 1995, 270, 18083–18087. [CrossRef] [PubMed]
- 14. Yenush, L.; Makati, K.J.; Smith-Hall, J.; Ishibashi, O.; Myers, M.G., Jr.; White, M.F. The pleckstrin homology domain is the principal link between the insulin receptor and IRS-1. *J. Biol. Chem.* **1996**, *271*, 24300–24306. [CrossRef]
- 15. Burks, D.J.; Pons, S.; Towery, H.; Smith-Hall, J.; Myers, M.G., Jr.; Yenush, L.; White, M.F. Heterologous pleckstrin homology domains do not couple IRS-1 to the insulin receptor. *J. Biol. Chem.* **1997**, 272, 27716–27721. [CrossRef]
- 16. Smith-Hall, J.; Pons, S.; Patti, M.E.; Burks, D.J.; Yenush, L.; Sun, X.J.; Kahn, C.R.; White, M.F. The 60 kDa insulin receptor substrate functions like an IRS protein (pp60IRS3) in adipose cells. *Biochemistry* **1997**, *36*, 8304–8310. [CrossRef]
- Lavan, B.E.; Fantin, V.R.; Chang, E.T.; Lane, W.S.; Keller, S.R.; Lienhard, G.E. A novel 160-kDa phosphotyrosine protein in insulin-treated embryonic kidney cells is a new member of the insulin receptor substrate family. *J. Biol. Chem.* 1997, 272, 21403–21407. [CrossRef]
- 18. Cai, D.; Dhe-Paganon, S.; Melendez, P.A.; Lee, J.; Shoelson, S.E. Two new substrates in insulin signaling, IRS5/DOK4 and IRS6/DOK5. J. Biol. Chem. 2003, 278, 25323–25330. [CrossRef]
- 19. Bjornholm, M.; He, A.R.; Attersand, A.; Lake, S.; Liu, S.C.; Lienhard, G.E.; Taylor, S.; Arner, P.; Zierath, J.R. Absence of functional insulin receptor substrate-3 (IRS-3) gene in humans. *Diabetologia* **2002**, *45*, 1697–1702. [CrossRef]
- 20. Favre, C.; Gerard, A.; Clauzier, E.; Pontarotti, P.; Olive, D.; Nunes, J.A. DOK4 and DOK5: New Dok-related genes expressed in human T cells. *Genes Immun.* 2003, *4*, 40–45. [CrossRef]
- Mardilovich, K.; Pankratz, S.L.; Shaw, L.M. Expression and function of the insulin receptor substrate proteins in cancer. *Cell Commun. Signal.* 2009, 7, 14. [CrossRef] [PubMed]
- 22. Singh, P.; Alex, J.M.; Bast, F. Insulin receptor (IR) and insulin-like growth factor receptor 1 (IGF-1R) signaling systems: Novel treatment strategies for cancer. *Med. Oncol.* 2014, *31*, 805. [CrossRef] [PubMed]
- White, M.F. IRS proteins and the common path to diabetes. Am. J. Physiol. Endocrinol. Metab. 2002, 283, E413–E422. [CrossRef] [PubMed]
- Gorgisen, G.; Gulacar, I.M.; Ozes, O.N. The role of insulin receptor substrate (IRS) proteins in oncogenic transformation. *Cell. Mol. Biol.* 2017, 63, 1–5. [CrossRef]
- 25. Hoxhaj, G.; Manning, B.D. The PI3K-AKT network at the interface of oncogenic signaling and cancer metabolism. *Nat. Rev. Cancer* 2020, 20, 74–88. [CrossRef]
- Deng, S.; Leong, H.C.; Datta, A.; Gopal, V.; Kumar, A.P.; Yap, C.T. PI3K/AKT Signaling Tips the Balance of Cytoskeletal Forces for Cancer Progression. *Cancers* 2022, 14, 1652. [CrossRef]
- 27. Tanaka, S.; Ito, T.; Wands, J.R. Neoplastic transformation induced by insulin receptor substrate-1 overexpression requires an interaction with both Grb2 and Syp signaling molecules. *J. Biol. Chem.* **1996**, 271, 14610–14616. [CrossRef]
- Hancer, N.J.; Qiu, W.; Cherella, C.; Li, Y.; Copps, K.D.; White, M.F. Insulin and metabolic stress stimulate multisite serine/threonine phosphorylation of insulin receptor substrate 1 and inhibit tyrosine phosphorylation. *J. Biol. Chem.* 2014, 289, 12467–12484. [CrossRef]
- 29. White, M.F.; Kahn, C.R. Insulin action at a molecular level—100 years of progress. Mol. Metab. 2021, 52, 101304. [CrossRef]
- 30. Tanti, J.F.; Jager, J. Cellular mechanisms of insulin resistance: Role of stress-regulated serine kinases and insulin receptor substrates (IRS) serine phosphorylation. *Curr. Opin. Pharmacol.* **2009**, *9*, 753–762. [CrossRef]
- Ozes, O.N.; Akca, H.; Mayo, L.D.; Gustin, J.A.; Maehama, T.; Dixon, J.E.; Donner, D.B. A phosphatidylinositol 3kinase/Akt/mTOR pathway mediates and PTEN antagonizes tumor necrosis factor inhibition of insulin signaling through insulin receptor substrate-1. *Proc. Natl. Acad. Sci. USA* 2001, *98*, 4640–4645. [CrossRef] [PubMed]
- 32. Hiratani, K.; Haruta, T.; Tani, A.; Kawahara, J.; Usui, I.; Kobayashi, M. Roles of mTOR and JNK in serine phosphorylation, translocation, and degradation of IRS-1. *Biochem. Biophys. Res. Commun.* **2005**, 335, 836–842. [CrossRef] [PubMed]
- Peng, J.; He, L. IRS posttranslational modifications in regulating insulin signaling. J. Mol. Endocrinol. 2018, 60, R1–R8. [CrossRef] [PubMed]
- 34. Toskala, E.; Kennedy, D.W. Asthma risk factors. Int. Forum. Allergy Rhinol. 2015, 5 (Suppl. 1), S11–S16. [CrossRef]

- 35. Guibas, G.V.; Mathioudakis, A.G.; Tsoumani, M.; Tsabouri, S. Relationship of Allergy with Asthma: There Are More Than the Allergy "Eggs" in the Asthma "Basket". *Front. Pediatr.* 2017, *5*, 92. [CrossRef]
- 36. Quirt, J.; Hildebrand, K.J.; Mazza, J.; Noya, F.; Kim, H. Asthma. Allergy Asthma Clin. Immunol. 2018, 14, 50. [CrossRef]
- 37. Sandrock, C.E.; Norris, A. Infection in severe asthma exacerbations and critical asthma syndrome. *Clin. Rev. Allergy Immunol.* **2015**, *48*, 104–113. [CrossRef]
- Schivo, M.; Phan, C.; Louie, S.; Harper, R.W. Critical asthma syndrome in the ICU. *Clin. Rev. Allergy Immunol.* 2015, 48, 31–44. [CrossRef] [PubMed]
- 39. Wenzel, S.E. Asthma phenotypes: The evolution from clinical to molecular approaches. Nat. Med. 2012, 18, 716–725. [CrossRef]
- 40. Wenzel, S. Severe asthma: From characteristics to phenotypes to endotypes. Clin. Exp. Allergy 2012, 42, 650–658. [CrossRef]
- 41. Komlosi, Z.I.; van de Veen, W.; Kovacs, N.; Szucs, G.; Sokolowska, M.; O'Mahony, L.; Akdis, M.; Akdis, C.A. Cellular and molecular mechanisms of allergic asthma. *Mol. Asp. Med.* **2022**, *85*, 100995. [CrossRef] [PubMed]
- 42. Locksley, R.M. Asthma and allergic inflammation. *Cell* 2010, 140, 777–783. [CrossRef] [PubMed]
- 43. Mims, J.W. Asthma: Definitions and pathophysiology. Int. Forum. Allergy Rhinol. 2015, 5 (Suppl. 1), S2–S6. [CrossRef] [PubMed]
- 44. Agrawal, D.K.; Shao, Z. Pathogenesis of allergic airway inflammation. *Curr. Allergy Asthma Rep.* 2010, 10, 39–48. [CrossRef]
- 45. Finn, P.W.; Bigby, T.D. Innate immunity and asthma. *Proc. Am. Thorac. Soc.* **2009**, *6*, 260–265. [CrossRef]
- 46. Gauvreau, G.M.; El-Gammal, A.I.; O'Byrne, P.M. Allergen-induced airway responses. Eur. Respir. J. 2015, 46, 819–831. [CrossRef]
- Kim, H.Y.; DeKruyff, R.H.; Umetsu, D.T. The many paths to asthma: Phenotype shaped by innate and adaptive immunity. *Nat. Immunol.* 2010, 11, 577–584. [CrossRef]
- 48. Madore, A.M.; Laprise, C. Immunological and genetic aspects of asthma and allergy. J. Asthma Allergy 2010, 3, 107–121. [CrossRef]
- 49. Hammad, H.; Lambrecht, B.N. The basic immunology of asthma. *Cell* **2021**, *184*, 1469–1485. [CrossRef]
- 50. Lambrecht, B.N.; Hammad, H. The immunology of asthma. Nat. Immunol. 2015, 16, 45–56. [CrossRef]
- 51. Lambrecht, B.N.; Hammad, H.; Fahy, J.V. The Cytokines of Asthma. Immunity 2019, 50, 975–991. [CrossRef] [PubMed]
- 52. Heller, N.M.; Qi, X.; Junttila, I.S.; Shirey, K.A.; Vogel, S.N.; Paul, W.E.; Keegan, A.D. Type I IL-4Rs selectively activate IRS-2 to induce target gene expression in macrophages. *Sci. Signal.* 2008, *1*, ra17. [CrossRef] [PubMed]
- McCormick, S.M.; Gowda, N.; Fang, J.X.; Heller, N.M. Suppressor of Cytokine Signaling (SOCS)1 Regulates Interleukin-4 (IL-4)-activated Insulin Receptor Substrate (IRS)-2 Tyrosine Phosphorylation in Monocytes and Macrophages via the Proteasome. J. Biol. Chem. 2016, 291, 20574–20587. [CrossRef]
- 54. Gour, N.; Wills-Karp, M. IL-4 and IL-13 signaling in allergic airway disease. Cytokine 2015, 75, 68–78. [CrossRef]
- Sun, X.J.; Rothenberg, P.; Kahn, C.R.; Backer, J.M.; Araki, E.; Wilden, P.A.; Cahill, D.A.; Goldstein, B.J.; White, M.F. Structure of the insulin receptor substrate IRS-1 defines a unique signal transduction protein. *Nature* 1991, 352, 73–77. [CrossRef]
- 56. Sun, X.J.; Wang, L.M.; Zhang, Y.; Yenush, L.; Myers, M.G., Jr.; Glasheen, E.; Lane, W.S.; Pierce, J.H.; White, M.F. Role of IRS-2 in insulin and cytokine signaling. *Nature* **1995**, *377*, 173–177. [CrossRef] [PubMed]
- Rabega, C.; Alexandrescu, R.; Rabega, M. Study of purified aldolase from Saccharomyces cerevisiae, using irradiated fructose-1,6-diphosphate. *Rev. Ig Bacteriol. Virusol. Parazitol. Epidemiol. Pneumoftiziol. Bacteriol. Virusol. Parazitol. Epidemiol.* 1976, 21, 37–41.
- Brisson-Lougarre, A.; Blum, C.J. Specific receptors for triiodothyronine in nuclei isolated from normal human polynuclear neutrophils. C. R. Acad. Sci. III 1985, 300, 287–292.
- Manohar, S.; Yu, Q.; Gygi, S.P.; King, R.W. The Insulin Receptor Adaptor IRS2 is an APC/C Substrate That Promotes Cell Cycle Protein Expression and a Robust Spindle Assembly Checkpoint. *Mol. Cell. Proteom.* 2020, 19, 1450–1467. [CrossRef]
- Nakahara, M.; Ito, H.; Skinner, J.T.; Lin, Q.; Tamosiuniene, R.; Nicolls, M.R.; Keegan, A.D.; Johns, R.A.; Yamaji-Kegan, K. The inflammatory role of dysregulated IRS2 in pulmonary vascular remodeling under hypoxic conditions. *Am. J. Physiol. Lung Cell. Mol. Physiol.* 2021, 321, L416–L428. [CrossRef]
- Keegan, A.D.; Zamorano, J.; Keselman, A.; Heller, N.M. IL-4 and IL-13 Receptor Signaling From 4PS to Insulin Receptor Substrate
 There and Back Again, a Historical View. *Front. Immunol.* 2018, *9*, 1037. [CrossRef] [PubMed]
- Warren, K.J.; Fang, X.; Gowda, N.M.; Thompson, J.J.; Heller, N.M. The TORC1-activated Proteins, p70S6K and GRB10, Regulate IL-4 Signaling and M2 Macrophage Polarization by Modulating Phosphorylation of Insulin Receptor Substrate-2. *J. Biol. Chem.* 2016, 291, 24922–24930. [CrossRef]
- Karo-Atar, D.; Bitton, A.; Benhar, I.; Munitz, A. Therapeutic Targeting of the Interleukin-4/Interleukin-13 Signaling Pathway: In Allergy and Beyond. *BioDrugs* 2018, 32, 201–220. [CrossRef] [PubMed]
- 64. Wu, W.J.; Wang, S.H.; Wu, C.C.; Su, Y.A.; Chiang, C.Y.; Lai, C.H.; Wang, T.H.; Cheng, T.L.; Kuo, J.Y.; Hsu, T.C.; et al. IL-4 and IL-13 Promote Proliferation of Mammary Epithelial Cells through STAT6 and IRS-1. *Int. J. Mol. Sci.* **2021**, *22*, 12008. [CrossRef]
- Keegan, A.D.; Nelms, K.; White, M.; Wang, L.M.; Pierce, J.H.; Paul, W.E. An IL-4 receptor region containing an insulin receptor motif is important for IL-4-mediated IRS-1 phosphorylation and cell growth. *Cell* 1994, 76, 811–820. [CrossRef]
- White, S.R.; Martin, L.D.; Abe, M.K.; Marroquin, B.A.; Stern, R.; Fu, X. Insulin receptor substrate-1/2 mediates IL-4-induced migration of human airway epithelial cells. *Am. J. Physiol. Lung Cell. Mol. Physiol.* 2009, 297, L164–L173. [CrossRef] [PubMed]
- 67. Takeda, K.; Tanaka, T.; Shi, W.; Matsumoto, M.; Minami, M.; Kashiwamura, S.; Nakanishi, K.; Yoshida, N.; Kishimoto, T.; Akira, S. Essential role of Stat6 in IL-4 signaling. *Nature* **1996**, *380*, 627–630. [CrossRef] [PubMed]
- Fu, C.; Jiang, L.; Hao, S.; Liu, Z.; Ding, S.; Zhang, W.; Yang, X.; Li, S. Activation of the IL-4/STAT6 Signaling Pathway Promotes Lung Cancer Progression by Increasing M2 Myeloid Cells. *Front. Immunol.* 2019, 10, 2638. [CrossRef]

- 69. Northway, W.H., Jr.; Rosan, R.C.; Porter, D.Y. Pulmonary disease following respirator therapy of hyaline-membrane disease. Bronchopulmonary dysplasia. *N. Engl. J. Med.* **1967**, *276*, 357–368. [CrossRef]
- Tracy, M.K.; Berkelhamer, S.K. Bronchopulmonary Dysplasia and Pulmonary Outcomes of Prematurity. *Pediatr. Ann.* 2019, 48, e148–e153. [CrossRef]
- Bhandari, A.; Bhandari, V. Pitfalls, problems, and progress in bronchopulmonary dysplasia. *Pediatrics* 2009, 123, 1562–1573. [CrossRef] [PubMed]
- 72. Jobe, A.J. The new BPD: An arrest of lung development. Pediatr. Res. 1999, 46, 641–643. [CrossRef] [PubMed]
- Jobe, A.H.; Bancalari, E. Bronchopulmonary dysplasia. *Am. J. Respir. Crit. Care Med.* 2001, 163, 1723–1729. [CrossRef] [PubMed]
 Davidson, L.M.; Berkelhamer, S.K. Bronchopulmonary Dysplasia: Chronic Lung Disease of Infancy and Long-Term Pulmonary Outcomes. *J. Clin. Med.* 2017, 6, 4. [CrossRef]
- 75. Thebaud, B.; Goss, K.N.; Laughon, M.; Whitsett, J.A.; Abman, S.H.; Steinhorn, R.H.; Aschner, J.L.; Davis, P.G.; McGrath-Morrow, S.A.; Soll, R.F.; et al. Bronchopulmonary dysplasia. *Nat. Rev. Dis. Primers* **2019**, *5*, 78. [CrossRef] [PubMed]
- 76. Warner, B.B.; Stuart, L.A.; Papes, R.A.; Wispe, J.R. Functional and pathological effects of prolonged hyperoxia in neonatal mice. *Am. J. Physiol.* **1998**, 275, L110–L117. [CrossRef] [PubMed]
- Jones, M.R.; Chong, L.; Bellusci, S. Fgf10/Fgfr2b Signaling Orchestrates the Symphony of Molecular, Cellular, and Physical Processes Required for Harmonious Airway Branching Morphogenesis. Front. Cell Dev. Biol. 2020, 8, 620667. [CrossRef] [PubMed]
- 78. Taghizadeh, S.; Chao, C.M.; Guenther, S.; Glaser, L.; Gersmann, L.; Michel, G.; Kraut, S.; Goth, K.; Koepke, J.; Heiner, M.; et al. FGF10 triggers de novo alveologenesis in a BPD model: Impact on the resident mesenchymal niche cells. *Stem. Cells* 2022, 40, 605–617. [CrossRef]
- 79. Oak, P.; Hilgendorff, A. The BPD trio? Interaction of dysregulated PDGF, VEGF, and TGF signaling in neonatal chronic lung disease. *Mol. Cell. Pediatr.* 2017, 4, 11. [CrossRef]
- 80. Huang, S.S.; Leal, S.M.; Chen, C.L.; Liu, I.H.; Huang, J.S. Cellular growth inhibition by TGF-beta1 involves IRS proteins. *FEBS Lett.* 2004, 565, 117–121. [CrossRef]
- Yoneyama, Y.; Lanzerstorfer, P.; Niwa, H.; Umehara, T.; Shibano, T.; Yokoyama, S.; Chida, K.; Weghuber, J.; Hakuno, F.; Takahashi, S.I. IRS-1 acts as an endocytic regulator of IGF-I receptor to facilitate sustained IGF signaling. *Elife* 2018, 7, e32893. [CrossRef] [PubMed]
- 82. Rabiee, A.; Kruger, M.; Ardenkjaer-Larsen, J.; Kahn, C.R.; Emanuelli, B. Distinct signaling properties of insulin receptor substrate (IRS)-1 and IRS-2 in mediating insulin/IGF-1 action. *Cell. Signal.* **2018**, 47, 1–15. [CrossRef] [PubMed]
- 83. Day, C.L.; Ryan, R.M. Bronchopulmonary dysplasia: New becomes old again! Pediatr. Res. 2017, 81, 210–213. [CrossRef]
- 84. Capoluongo, E.; Ameglio, F.; Zuppi, C. Insulin-like growth factor-I and complications of prematurity: A focus on bronchopulmonary dysplasia. *Clin. Chem. Lab. Med.* 2008, *46*, 1061–1066. [CrossRef]
- Chetty, A.; Nielsen, H.C. Regulation of cell proliferation by insulin-like growth factor 1 in hyperoxia-exposed neonatal rat lung. Mol. Genet. Metab. 2002, 75, 265–275. [CrossRef] [PubMed]
- Belcastro, R.; Lopez, L.; Li, J.; Masood, A.; Tanswell, A.K. Chronic lung injury in the neonatal rat: Up-regulation of TGFbeta1 and nitration of IGF-R1 by peroxynitrite as likely contributors to impaired alveologenesis. *Free Radic. Biol. Med.* 2015, *80*, 1–11. [CrossRef] [PubMed]
- 87. Ahamed, K.; Epaud, R.; Holzenberger, M.; Bonora, M.; Flejou, J.F.; Puard, J.; Clement, A.; Henrion-Caude, A. Deficiency in type 1 insulin-like growth factor receptor in mice protects against oxygen-induced lung injury. *Respir. Res.* 2005, *6*, 31. [CrossRef]
- Kheirollahi, V.; Khadim, A.; Kiliaris, G.; Korfei, M.; Barroso, M.M.; Alexopoulos, I.; Vazquez-Armendariz, A.I.; Wygrecka, M.; Ruppert, C.; Guenther, A.; et al. Transcriptional Profiling of Insulin-like Growth Factor Signaling Components in Embryonic Lung Development and Idiopathic Pulmonary Fibrosis. *Cells* 2022, *11*, 1973. [CrossRef]
- 89. Lofqvist, C.; Hellgren, G.; Niklasson, A.; Engstrom, E.; Ley, D.; Hansen-Pupp, I.; Consortium, W. Low postnatal serum IGF-I levels are associated with bronchopulmonary dysplasia (BPD). *Acta Paediatr.* **2012**, *101*, 1211–1216. [CrossRef] [PubMed]
- Chetty, A.; Andersson, S.; Lassus, P.; Nielsen, H.C. Insulin-like growth factor-1 (IGF-1) and IGF-1 receptor (IGF-1R) expression in human lung in RDS and BPD. *Pediatr. Pulmonol.* 2004, 37, 128–136. [CrossRef]
- Capoluongo, E.; Vento, G.; Ameglio, F.; Lulli, P.; Matassa, P.G.; Carrozza, C.; Santini, S.A.; Antenucci, M.; Castagnola, M.; Giardina, B.; et al. Increased levels of IGF-1 and beta2-microglobulin in epithelial lining fluid of preterm newborns developing chronic lung disease. effects of rhG-CSF. Int. J. Immunopathol. Pharmacol. 2006, 19, 57–66. [CrossRef] [PubMed]
- Zhang, S.; Luan, X.; Li, H.; Jin, Z. Insulin-like growth factor-1: A potential target for bronchopulmonary dysplasia treatment (Review). *Exp. Ther. Med.* 2022, 23, 191. [CrossRef] [PubMed]
- Kuebler, W.M.; Uhlig, U.; Goldmann, T.; Schael, G.; Kerem, A.; Exner, K.; Martin, C.; Vollmer, E.; Uhlig, S. Stretch activates nitric oxide production in pulmonary vascular endothelial cells in situ. *Am. J. Respir. Crit. Care Med.* 2003, 168, 1391–1398. [CrossRef] [PubMed]
- 94. Uhlig, U.; Fehrenbach, H.; Lachmann, R.A.; Goldmann, T.; Lachmann, B.; Vollmer, E.; Uhlig, S. Phosphoinositide 3-OH kinase inhibition prevents ventilation-induced lung cell activation. *Am. J. Respir. Crit. Care Med.* **2004**, *169*, 201–208. [CrossRef]
- Benjamin, J.T.; Carver, B.J.; PLoSa, E.J.; Yamamoto, Y.; Miller, J.D.; Liu, J.H.; van der Meer, R.; Blackwell, T.S.; Prince, L.S. NF-kappaB activation limits airway branching through inhibition of Sp1-mediated fibroblast growth factor-10 expression. *J. Immunol.* 2010, 185, 4896–4903. [CrossRef]

- Carver, B.J.; PLoSa, E.J.; Stinnett, A.M.; Blackwell, T.S.; Prince, L.S. Interactions between NF-kappaB and SP3 connect inflammatory signaling with reduced FGF-10 expression. J. Biol. Chem. 2013, 288, 15318–15325. [CrossRef]
- Wu, D.; Liang, M.; Dang, H.; Fang, F.; Xu, F.; Liu, C. Hydrogen protects against hyperoxia-induced apoptosis in type II alveolar epithelial cells via activation of PI3K/Akt/Foxo3a signaling pathway. *Biochem. Biophys. Res. Commun.* 2018, 495, 1620–1627. [CrossRef]
- Mendoza, M.C.; Er, E.E.; Blenis, J. The Ras-ERK and PI3K-mTOR pathways: Cross-talk and compensation. *Trends Biochem. Sci.* 2011, 36, 320–328. [CrossRef]
- 99. Hay, N.; Sonenberg, N. Upstream and downstream of mTOR. Genes Dev. 2004, 18, 1926–1945. [CrossRef]
- Thomas, G.V.; Tran, C.; Mellinghoff, I.K.; Welsbie, D.S.; Chan, E.; Fueger, B.; Czernin, J.; Sawyers, C.L. Hypoxia-inducible factor determines sensitivity to inhibitors of mTOR in kidney cancer. *Nat. Med.* 2006, 12, 122–127. [CrossRef]
- 101. Wang, S.H.; Li, L.H.; Zou, D.M.; Zheng, X.M.; Deng, J. Roles of the mammalian target of rapamycin (mTOR) signaling pathway in the repair of hyperoxia-induced acute lung injury. *Adv. Clin. Exp. Med.* **2020**, *29*, 13–23. [CrossRef] [PubMed]
- Chung, E.J.; Sowers, A.; Thetford, A.; McKay-Corkum, G.; Chung, S.I.; Mitchell, J.B.; Citrin, D.E. Mammalian Target of Rapamycin Inhibition With Rapamycin Mitigates Radiation-Induced Pulmonary Fibrosis in a Murine Model. *Int. J. Radiat. Oncol. Biol. Phys.* 2016, 96, 857–866. [CrossRef] [PubMed]
- 103. Wang, Y.; Ma, Q.; Ma, X.; Zhang, Z.; Liu, N.; Wang, M. Role of mammalian target of rapamycin signaling in autophagy and the neurodegenerative process using a senescence accelerated mouse-prone 8 model. *Exp. Ther. Med.* 2017, 14, 1051–1057. [CrossRef] [PubMed]
- 104. Porzionato, A.; Sfriso, M.M.; Mazzatenta, A.; Macchi, V.; De Caro, R.; Di Giulio, C. Effects of hyperoxic exposure on signal transduction pathways in the lung. *Respir. Physiol. Neurobiol.* **2015**, *209*, 106–114. [CrossRef]
- 105. Rubinfeld, H.; Seger, R. The ERK cascade: A prototype of MAPK signaling. Mol. Biotechnol. 2005, 31, 151–174. [CrossRef]
- 106. El Agha, E.; Bellusci, S. Walking along the Fibroblast Growth Factor 10 Route: A Key Pathway to Understand the Control and Regulation of Epithelial and Mesenchymal Cell-Lineage Formation during Lung Development and Repair after Injury. *Scientifica* 2014, 2014, 538379. [CrossRef]
- 107. Bellusci, S.; Grindley, J.; Emoto, H.; Itoh, N.; Hogan, B.L. Fibroblast growth factor 10 (FGF10) and branching morphogenesis in the embryonic mouse lung. *Development* **1997**, 124, 4867–4878. [CrossRef]
- 108. Gupte, V.V.; Ramasamy, S.K.; Reddy, R.; Lee, J.; Weinreb, P.H.; Violette, S.M.; Guenther, A.; Warburton, D.; Driscoll, B.; Minoo, P.; et al. Overexpression of fibroblast growth factor-10 during both inflammatory and fibrotic phases attenuates bleomycin-induced pulmonary fibrosis in mice. *Am. J. Respir. Crit. Care Med.* **2009**, *180*, 424–436. [CrossRef]
- 109. Volckaert, T.; Dill, E.; Campbell, A.; Tiozzo, C.; Majka, S.; Bellusci, S.; De Langhe, S.P. Parabronchial smooth muscle constitutes an airway epithelial stem cell niche in the mouse lung after injury. *J. Clin. Investig.* **2011**, 121, 4409–4419. [CrossRef]
- Alejandre-Alcazar, M.A.; Michiels-Corsten, M.; Vicencio, A.G.; Reiss, I.; Ryu, J.; de Krijger, R.R.; Haddad, G.G.; Tibboel, D.; Seeger, W.; Eickelberg, O.; et al. TGF-beta signaling is dynamically regulated during the alveolarization of rodent and human lungs. *Dev. Dyn.* 2008, 237, 259–269. [CrossRef]
- 111. Chao, C.M.; Chong, L.; Chu, X.; Shrestha, A.; Behnke, J.; Ehrhardt, H.; Zhang, J.; Chen, C.; Bellusci, S. Targeting Bronchopulmonary Dysplasia-Associated Pulmonary Hypertension (BPD-PH): Potential Role of the FGF Signaling Pathway in the Development of the Pulmonary Vascular System. *Cells* 2020, *9*, 1875. [CrossRef] [PubMed]
- 112. Scott, C.L.; Walker, D.J.; Cwiklinski, E.; Tait, C.; Tee, A.R.; Land, S.C. Control of HIF-1α and vascular signaling in fetal lung involves cross talk between mTORC1 and the FGF-10/FGFR2b/Spry2 airway branching periodicity clock. *Am. J. Physiol. Lung Cell. Mol. Physiol.* **2010**, 299, L455–L471. [CrossRef] [PubMed]
- 113. Al Alam, D.; El Agha, E.; Sakurai, R.; Kheirollahi, V.; Moiseenko, A.; Danopoulos, S.; Shrestha, A.; Schmoldt, C.; Quantius, J.; Herold, S.; et al. Evidence for the involvement of fibroblast growth factor 10 in lipofibroblast formation during embryonic lung development. *Development* 2015, 142, 4139–4150. [CrossRef] [PubMed]
- 114. Nolan, M.K.; Jankowska, L.; Prisco, M.; Xu, S.; Guvakova, M.A.; Surmacz, E. Differential roles of IRS-1 and SHC signaling pathways in breast cancer cells. *Int. J. Cancer* **1997**, *72*, 828–834. [CrossRef]
- 115. Shi, Y.; Ma, Z.; Cheng, Q.; Wu, Y.; Parris, A.B.; Kong, L.; Yang, X. FGFR1 overexpression renders breast cancer cells resistant to metformin through activation of IRS1/ERK signaling. *Biochim. Biophys. Acta Mol. Cell Res.* 2021, 1868, 118877. [CrossRef] [PubMed]
- 116. Dailey, L.; Laplantine, E.; Priore, R.; Basilico, C. A network of transcriptional and signaling events is activated by FGF to induce chondrocyte growth arrest and differentiation. *J. Cell Biol.* **2003**, *161*, 1053–1066. [CrossRef]
- Lassarre, C.; Ricort, J.M. Growth factor-specific regulation of insulin receptor substrate-1 expression in MCF-7 breast carcinoma cells: Effects on the insulin-like growth factor signaling pathway. *Endocrinology* 2003, 144, 4811–4819. [CrossRef]
- Manzano-Nunez, F.; Arambul-Anthony, M.J.; Galan Albinana, A.; Leal Tassias, A.; Acosta Umanzor, C.; Borreda Gasco, I.; Herrera, A.; Forteza Vila, J.; Burks, D.J.; Noon, L.A. Insulin resistance disrupts epithelial repair and niche-progenitor Fgf signaling during chronic liver injury. *PLoS Biol.* 2019, *17*, e2006972. [CrossRef]
- Alejandre-Alcazar, M.A.; Kwapiszewska, G.; Reiss, I.; Amarie, O.V.; Marsh, L.M.; Sevilla-Perez, J.; Wygrecka, M.; Eul, B.; Kobrich, S.; Hesse, M.; et al. Hyperoxia modulates TGF-beta/BMP signaling in a mouse model of bronchopulmonary dysplasia. *Am. J. Physiol. Lung Cell. Mol. Physiol.* 2007, 292, L537–L549. [CrossRef]
- 120. Massague, J. TGF-beta signal transduction. Annu. Rev. Biochem. 1998, 67, 753–791. [CrossRef]

- 121. Gauldie, J.; Galt, T.; Bonniaud, P.; Robbins, C.; Kelly, M.; Warburton, D. Transfer of the active form of transforming growth factor-beta 1 gene to newborn rat lung induces changes consistent with bronchopulmonary dysplasia. *Am. J. Pathol.* **2003**, *163*, 2575–2584. [CrossRef]
- 122. Sureshbabu, A.; Syed, M.A.; Boddupalli, C.S.; Dhodapkar, M.V.; Homer, R.J.; Minoo, P.; Bhandari, V. Conditional overexpression of TGFbeta1 promotes pulmonary inflammation, apoptosis and mortality via TGFbetaR2 in the developing mouse lung. *Respir. Res.* **2015**, *16*, 4. [CrossRef]
- 123. Warburton, D.; Bellusci, S.; De Langhe, S.; Del Moral, P.M.; Fleury, V.; Mailleux, A.; Tefft, D.; Unbekandt, M.; Wang, K.; Shi, W. Molecular mechanisms of early lung specification and branching morphogenesis. *Pediatr. Res.* 2005, 57, 26R–37R. [CrossRef] [PubMed]
- 124. Hilgendorff, A.; Parai, K.; Ertsey, R.; Juliana Rey-Parra, G.; Thebaud, B.; Tamosiuniene, R.; Jain, N.; Navarro, E.F.; Starcher, B.C.; Nicolls, M.R.; et al. Neonatal mice genetically modified to express the elastase inhibitor elafin are protected against the adverse effects of mechanical ventilation on lung growth. *Am. J. Physiol. Lung Cell. Mol. Physiol.* 2012, 303, L215–L227. [CrossRef] [PubMed]
- 125. Mokres, L.M.; Parai, K.; Hilgendorff, A.; Ertsey, R.; Alvira, C.M.; Rabinovitch, M.; Bland, R.D. Prolonged mechanical ventilation with air induces apoptosis and causes failure of alveolar septation and angiogenesis in lungs of newborn mice. *Am. J. Physiol. Lung Cell. Mol. Physiol.* **2010**, *298*, L23–L35. [CrossRef] [PubMed]
- 126. Kompass, K.S.; Deslee, G.; Moore, C.; McCurnin, D.; Pierce, R.A. Highly conserved transcriptional responses to mechanical ventilation of the lung. *Physiol. Genom.* **2010**, *42*, 384–396. [CrossRef]
- 127. Speer, C.P. Pulmonary inflammation and bronchopulmonary dysplasia. J. Perinatol. 2006, 26 (Suppl. 1), S57–S62. [CrossRef]
- 128. Ballabh, P.; Simm, M.; Kumari, J.; Krauss, A.N.; Jain, A.; Califano, C.; Lesser, M.L.; Cunningham-Rundles, S. Neutrophil and monocyte adhesion molecules in bronchopulmonary dysplasia, and effects of corticosteroids. *Arch. Dis. Child. Fetal Neonatal Ed.* 2004, *89*, F76–F83. [CrossRef]
- 129. Ogden, B.E.; Murphy, S.; Saunders, G.C.; Johnson, J.D. Lung lavage of newborns with respiratory distress syndrome. Prolonged neutrophil influx is associated with bronchopulmonary dysplasia. *Chest* **1983**, *83*, 31S–33S. [CrossRef]
- 130. Grotendorst, G.R.; Smale, G.; Pencev, D. Production of transforming growth factor beta by human peripheral blood monocytes and neutrophils. *J. Cell. Physiol.* **1989**, *140*, 396–402. [CrossRef]
- 131. Schultz, C.; Tautz, J.; Reiss, I.; Moller, J.C. Prolonged mechanical ventilation induces pulmonary inflammation in preterm infants. *Biol. Neonate* **2003**, *84*, 64–66. [CrossRef] [PubMed]
- 132. Merritt, T.A.; Deming, D.D.; Boynton, B.R. The 'new' bronchopulmonary dysplasia: Challenges and commentary. *Semin. Fetal Neonatal Med.* **2009**, *14*, 345–357. [CrossRef]
- Carlton, D.P.; Albertine, K.H.; Cho, S.C.; Lont, M.; Bland, R.D. Role of neutrophils in lung vascular injury and edema after premature birth in lambs. J. Appl. Physiol. 1997, 83, 1307–1317. [CrossRef] [PubMed]
- Jaarsma, A.S.; Braaksma, M.A.; Geven, W.B.; van Oeveren, W.; Bambang Oetomo, S. Activation of the inflammatory reaction within minutes after birth in ventilated preterm lambs with neonatal respiratory distress syndrome. *Biol. Neonate* 2004, *86*, 1–5. [CrossRef]
- 135. Kotecha, S.; Mildner, R.J.; Prince, L.R.; Vyas, J.R.; Currie, A.E.; Lawson, R.A.; Whyte, M.K. The role of neutrophil apoptosis in the resolution of acute lung injury in newborn infants. *Thorax* 2003, *58*, 961–967. [CrossRef] [PubMed]
- Huang, S.S.; Leal, S.M.; Chen, C.L.; Liu, I.H.; Huang, J.S. Identification of insulin receptor substrate proteins as key molecules for the TbetaR-V/LRP-1-mediated growth inhibitory signaling cascade in epithelial and myeloid cells. *FASEB J.* 2004, 18, 1719–1721. [CrossRef] [PubMed]
- 137. Shi, J.; Wang, D.M.; Wang, C.M.; Hu, Y.; Liu, A.H.; Zhang, Y.L.; Sun, B.; Song, J.G. Insulin receptor substrate-1 suppresses transforming growth factor-beta1-mediated epithelial-mesenchymal transition. *Cancer Res.* 2009, 69, 7180–7187. [CrossRef] [PubMed]
- 138. Bailey, K.L.; Agarwal, E.; Chowdhury, S.; Luo, J.; Brattain, M.G.; Black, J.D.; Wang, J. TGFbeta/Smad3 regulates proliferation and apoptosis through IRS-1 inhibition in colon cancer cells. *PLoS ONE* **2017**, *12*, e0176096. [CrossRef]