## REVIEW

**Open Access** 

# Roles of host mitochondria in the development of COVID-19 pathology: Could mitochondria be a potential therapeutic target?

Kavya Srinivasan<sup>1,2</sup>, Ashutosh Kumar Pandey<sup>3</sup>, Ashlena Livingston<sup>4</sup> and Sundararajan Venkatesh<sup>1\*</sup>

## Abstract

The recent emergence of severe acute respiratory syndrome-Corona Virus 2 (SARS-CoV-2) in late 2019 and its spread worldwide caused an acute pandemic of Coronavirus disease 19 (COVID-19). Since then, COVID-19 has been under intense scrutiny as its outbreak led to significant changes in healthcare, social activities, and economic settings worldwide. Although angiotensin-converting enzyme-2 (ACE-2) receptor is shown to be the primary port of SARS-CoV-2 entry in cells, the mechanisms behind the establishment and pathologies of COVID-19 are poorly understood. As recent studies have shown that host mitochondria play an essential role in virus-mediated innate immune response, pathologies, and infection, in this review, we will discuss in detail the entry and progression of SARS-CoV-2 and how mitochondria could play roles in COVID-19 disease. We will also review the potential interactions between SARS-CoV-2 and mitochondria and discuss possible treatments, including whether mitochondria as a potential therapeutic target in COVID-19. Understanding SARS-CoV-2 and mitochondrial interactions mediated virus establishment, inflammation, and other consequences may provide a unique mechanism and conceptual advancement in finding a novel treatment for COVID-19.

Keywords: COVID-19, SARS-CoV-2, Mitochondria, ACE-2 receptor, Cytokine storm, Inflammation

## Introduction

COVID-19 is caused by a new coronavirus, identified as a severe acute respiratory syndrome- coronavirus 2 (SARS-CoV-2), stirring the globe since the start of its outbreak in late 2019. At the time of the final submission of this article on 20<sup>th</sup> October 2021, around ~246 million people had been infected with SARS-CoV-2 and close to 5 million had died, which is approximately a 2% death rate across the globe [1]. Although the zoonotic origin of the virus is still not clear, it is believed that most likely the





© The Author(s) 2021. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/.

<sup>\*</sup>Correspondence: sundarve@njms.rutgers.edu

<sup>&</sup>lt;sup>1</sup> Department of Microbiology, Biochemistry and Molecular Genetics, Rutgers -New Jersey Medical School, The State University of New Jersey, Newark, NJ, USA

Full list of author information is available at the end of the article



also been reported in vaccinated groups as well [5–7]. However, the symptoms in these groups are mostly mild or none [7]. Therefore, understanding SARS-CoV-2's mechanisms of host interaction and disease establishment is important to develop a novel therapeutic tool to combat this highly contagious disease.

Angiotensin-converting enzyme-2 (ACE-2) receptormediated entry of the virus is considered as the primary infection mechanism; however, the consequence after entry and mechanisms of pathologies leading to mortality, cytokine storm, and inflammation are not well known. Furthermore, many newfound studies suggest the potential involvement of host mitochondria in COVID-19 infection is believed to be one of the key mechanisms for COVID-19 pathologies [8–11]. Therefore, in this review, we seek more clarity surrounding mitochondria and their role in establishing COVID-19 infection and explore their possible role as a potential therapeutic target in COVID-19 treatment.

## SARS-CoV-2 Structure

Coronaviruses belong to the family of *coronaviridae* and order *Nidovirales*, contain a large single-stranded RNA, and have a lipid envelope with spike proteins. When observed under the electron microscope, coronavirus appears as a crown-like structure due to their spike glycoproteins [12] (Fig. 1). Like most notable human disease-causing viruses, SARS-CoV-2 also contains a positive-sense single-stranded RNA, belonging to a subgroup of the coronavirus family known as beta-coronavirus. Other notable RNA viruses are those that cause the common cold, dengue, Ebola, hepatitis C, hepatitis E, influenza, measles, MERS, polio, rabies, SARS, and West Nile fever. Among these, SARS-CoV-2 is more contagious than its counterparts, transferred to humans at an exceptionally high infection rate that it has been estimated that the transmissibility rate  $(R_0)$  for SARS-CoV-2 to be in the average of 2.5 compared to 2.4 for SARS-CoV, 2.0 for the 1918 influenza pandemic, 1.7 for the 2009 influenza pandemic and 0.9 for MERS-CoV [13]. From so far identified coronaviruses, HCoV 229E, HKU1, NL63, and OC43 cause mild upper respiratory tract infections and are responsible for most of the common cold [14]. In contrast, SARS-CoV, MERS-CoV, and the recently identified SARS-CoV-2 are highly infectious and cause severe life-threatening diseases [14–19].

The SARS-CoV-2's RNA genome is about 29.9 kb long and encodes four structural proteins (S, E, M, and N) (Fig. 1) [20]. The structural proteins are surface spike glycoprotein (S), the envelope glycoprotein protein (E), membrane glycoprotein (M), and nucleocapsid protein (N) (Fig. 1) [21, 22]. SARS-CoV-2's RNA genome was also shown to encode sixteen non-structural proteins (nsp1–16). In addition, many open reading frames (ORFs) are being identified and reported. Although ORFs are not essential for viral replication but are believed to be necessary for viral pathogenesis. It has been predicted and reported several ORFs (3a, 3b, 6, 7a, 7b, 8, 9b, 9c, 10) in SARS-CoV-2 [23, 24].

Like SARS-CoV, SARS- CoV-2 also infects the host cell using the 150 kDa 'S' spike protein [25, 26]. However, 'N' glycoprotein is the most abundant structural protein, conserved among other coronaviruses, associated with the viral genome, and forms a ribonucleoprotein core of the virus [27]. While the coronavirus 'M' protein is involved in determining the virus's structure and shape, including stabilizing the N protein-RNA complex, the 'E' structural protein, which is the smallest of all structural proteins, helps in the production and maturation of the virus [28, 29]. Although ACE-2 is widely expressed in many organs and tissues, the respiratory tract cells remain the major port of entry. The ACE2 receptor, through which SARS-CoV-2 binds and enters the host cells, is a carboxypeptidase that functions as a counterbalance to the angiotensin-converting enzyme (ACE) of the renin-angiotensin-aldosterone system (RAAS) by converting angiotensin II to angiotensin (1-7). As a result, cells expressing ACE-2 are at a higher risk of establishing infection. SARS-CoV-2 has Open Reading Frames (ORFs) responsible for various steps of its disease progression, where ORF1ab is responsible for most enzymatic proteins, including S, E, M, and N [30].

## SARS-CoV-2 host cell entry mechanisms and its affinity to various tissues

One of the significant mechanisms of coronavirus entry known to date is that the virus uses the 'S' protein and the host's ACE-2 receptor to facilitate its access into the host cell. In contrast, S protein is primed by the host cell serine protease, transmembrane protease, serine 2 (TMPRSS2) [31, 32] (Fig. 2). Importantly, SARS-CoV-2 spike protein 'S' binds human ACE2 receptors through its receptorbinding domain (RBD), which then subsequently proteolytically primed by human proteases to enter the cell [25]. However, furin, a human protease (convertase) predominantly preactivates SARS-CoV-2 entry, reducing the virus dependency on human target cell proteases [25]. Interestingly, SARS-CoV-2 RBD, which has a high affinity to ACE2, also possesses hidden RBD to evade innate immune responses [25].

The ACE-2 receptor is commonly expressed in the lungs, intestine, kidneys, and heart [33–36]. Compared to the MERS-CoV and SARS-CoV viruses, the SARS-CoV-2 is highly infectious and aggressive, as evident from its exponential spread, and it is shown to have a higher

affinity for the ACE-2 receptor than other coronaviruses [18, 25, 37]. While MERS-CoV presents more gastrointestinal manifestations and kidney failure, SARS-CoV mainly manifests in the lower respiratory tract (LRT), possibly due to the host surface receptor affinity and distribution difference [38]. SARS-CoV entry is also through binding ACE-2 receptor, which is predominantly found in the lower respiratory tract cells [37, 39]. On the other hand, MERS-CoV facilitates access through surface enzyme dipeptidyl peptidase 4 (DPP4), also known as CD26, which is widely expressed in the LRT, kidney, and gastrointestinal tract (GIT) [40]. Genomic sequence and structure analysis of ACE2 and its side distributions across different species suggests a potential SARS-CoV-2 transmission from animals to humans [41]. Interestingly, ACE-2 expression is shown to be less in children than adults, which could be one of the reasons that SARS-CoV-2 is not highly contagious in children so far [42, 43]. The SARS-Cov-2 infection has also been reported in other tissues, including the heart, liver, skin, kidney, intestinal tract, and adipose tissue; however, why particular populations are affected with organ-specific infection is not clear [44–49].

In addition to ACE-2 receptor, recent evidence shows that SARS-CoV-2 can also bind to other cell surface markers such as Neuropilin-1, Neuropilin-2, and CD147 (EMMPRIN, a transmembrane glycoprotein of the immunoglobulin superfamily) facilitating virus cell entry, implying there could be more cell surface molecule involved in SARS-CoV-2 entry [50–53].

Mitochondria as an emerging target in SARS-CoV-2

Mitochondria is not only the power generator of the cell but also regulates various other physiological and pathological processes, including calcium and iron homeostasis, steroid synthesis, and apoptosis [54-56]. Although mitochondrial ATP is required to maintain cellular homeostasis, unfortunately, it is also utilized for in-host viral replication [57, 58]. It is known that many single-stranded RNA viruses alter the host mitochondrial dynamics [59-62]. SARS-CoV-2 is closely related to SARS-CoV, which interacts with the host mitochondria in manipulating immune response to evade innate immunity [60]. A recent study shows that coronaviruses SARS-CoV-1, SARS-CoV-2, and MERS-CoV have similar proteins and localization patterns, supporting the hypothesis that these conserved proteins function similarly [63]. In addition, many of the SARS-CoV-2 structural and non-structural proteins encoded by various open reading frames (ORFs) has been shown to have the potential to interact with mitochondria, which will be discussed further below.

**SARS-CoV-2 and mitochondrial interaction** Mitochondria and viral interaction is a longstanding mystery yet to be resolved, but emerging evidence suggests potential interactions between them. Evidence shows that the mitochondrial network is highly susceptible to viral manipulation to evade mitochondria-mediated innate immune response [8, 64]. Since mitochondrial function is essential for various cellular processes, including cellular defense systems, exploiting its dynamics and function by the virus is highly likely. This section discusses the potential interaction between certain SARS-CoV-2 proteins, including ORFs, with mitochondria and outlines how it possibly manipulates them to establish viral infection.

## Structural protein 'M'

The M protein is one of the abundantly found structural proteins in SARS-CoV 2. It is the central organizer of the virus assembly and is responsible for the membrane curvature of the protein that helps interact with other structural proteins [28]. M protein has been shown to adopt two conformations; elongated M protein, which participates in spike clusters, rigidity, and membrane curvature. The compact M protein participates in flexibility and low spike density. The homotypic interaction between the M proteins is the primary foundation for the virion envelope [65]. M proteins interact with other structural proteins such as S and N proteins in regulating the size and variability of the virion [65].

Recently, most of the SARS-CoV-2 proteins have been cloned and expressed in the human cell, and through affinity-purified mass spectrometry, SARS-CoV-2 and human protein interactions have been identified [23]. This includes structural protein M, which interacts with many host mitochondrial proteins such as acyl-CoA dehydrogenase medium chain (ACADM), mitochondrial-processing peptidase subunit beta (PMPCB), pitrilysin metallopeptidase 1 (PITRM1), mitochondrialprocessing peptidase subunit alpha (PMPCA), and



coenzyme Q8B (COQ8B) [23]. These interactions are critical because the virus can induce cellular apoptosis, which is accomplished by major structural proteins like M protein. This is supported by a study in SARS-CoV that showed that M protein itself when overexpressed, induces apoptosis in human pulmonary fibroblast cells [66]. For apoptosis, a mitochondrial event called the mitochondrial outer membrane permeabilization releases caspases, a type of cysteine protease that executes the programmed cell death. However, many characteristic features are still unknown about the M protein and its interactions with mitochondria. Most of the M- interacting mitochondrial proteins belong to electron transport chain (ETC) subunits, metabolic enzymes, and processing peptidase [23]. However, the consequences and significance of these interactions are yet to be determined. Therefore, increasing our understanding of the interaction between structural protein M and mitochondria could help establish the targets for a better approach in treating COVID-19.

### Non Structural proteins Nsp2, 7, 8, 10

In addition to structural protein M, SARS-CoV-2's nonstructural proteins Nsp2, 7, 8, and 10 are also predicted to interact with mitochondria. This is based on the study employed in SARS-CoV in which multidimensional protein identification technology (MudPIT) with C-terminally tagged SARS-CoV Nsp2 identified that Nsp2 interacts with Prohibitin 1 (PHB1) and PHB2, which are inner mitochondrial membrane proteins [67, 68]. However, the exact molecular function of the PHB complex is unknown, but it is believed to possess a chaperoning role in helping to maintain mitochondrial morphology. Additionally, SARS-CoV-2's Nsp7 is also shown to interact with NDUFAF2, an NADH dehydrogenase [ubiquinone] 1 alpha subcomplex assembly factor 2 that is required for optimal assembly of Complex-I at the mitochondrial inner membrane [23]. Whereas, Nsp8 is shown to interact with various mitochondrial ribosomal subunits such as MRPS2, MRPS5, MRPS25, MRPS27, which are part of mitochondrial translation machinery [23]. Furthermore, by employing human embryo lung fibroblast cDNA library using a yeast trap method, it has been found that SARS-CoV nsp10 protein interacts with the NADH 4L subunit and cytochrome oxidase II, resulting in reduced NADHcytochrome activity and depolarization of the inner mitochondrial membrane, suggesting a possible role of Nsp10 in the cytopathic effect of SARS-CoV [69]. Taken together, most of the SARS-CoV-2 non-structural proteins may support viral establishment and replication by modulating the mitochondrial function.

## Open reading frame 3a (ORF3a)

Open reading frame 3a (ORF3a) encoded protein in SARS-CoV-2 is shown to target mitochondrial ubiquitin-specific peptidase 30 (USP30), a mitochondrial deubiguitinase involved in mitochondrial homeostasis and mitophagy control [70]. SARS-CoV-2 can control mitochondrial function through USP30 to help with host immunosuppression. Like other viruses, SARS-CoV-2 can also induce the neutrophil extracellular trap (NET) osis, an inflammatory response involving mitochondrial biogenesis, mitochondrial fusion, and fission to release mitochondrial DNA (mtDNA) outside of the cell [8, 71]. The release of mtDNA into the cytoplasm triggers the innate immune response and inflammation, a well-known phenomenon that is shown to occur during SARS-CoV infection [72-74]. As mtDNA levels increase, the damage and severity of the illness can progress to multiorgan failure, which is most likely cause mortality [75]. Although a certain level of mtDNA release may be necessary to elicit an immune response, an excess release may lead to severe pathologies in COVID-19.

SARS-CoV-2's ORF3b protein is 154 amino acids, believed to be expressed from the subgenomic RNA3. It has been shown to localize to the nucleus and then translocate to mitochondria through the typical mitochondrial-targeted amphipathic  $\alpha$ -helix [76]. It has also been demonstrated that ORF3b inhibits IFN-1 induction, mediated by retinoic acid-induced gene 1 (RIG-1) and the mitochondrial antiviral signaling protein. Thus, it could be one of the mechanisms for SARS-CoV-2 to evade innate immunity.

## Open reading frame 7a (ORF7a)

Alongside other open reading proteins, ORFs 7a may aid with localization of the viral genome to the mitochondria, interact directly or indirectly and favor disease progression [8, 77, 78]. ORF7a is a transmembrane protein highly conserved between SARS-CoV and 2 [8]. It has also been shown that when ORF7a and Bcl-X are overexpressed in 293T and Vero cells and when ORF7a was immunoprecipitated, they found that ORF7a pulled down Bcl-X<sub>L</sub> and other prosurvival proteins such as Bcl-2, Bcl-w, Mcl-1, and A 1[78]. Further, fractionation experiments demonstrate that ORF7a is localized to mitochondria and additionally learned that the transmembrane domain of ORF7a is critical in interacting with Bcl-X<sub>L</sub> and for inducing apoptosis [78]. Similarly, the C-terminal transmembrane domain of Bcl-X<sub>L</sub> is essential for this interaction and apoptosis induction.

Interestingly, ORF7a does not interact with pro-apoptotic proteins such as Bax, Bak, Bad, and Bid. Although ORF7a is shown to localize majorly to Endoplasmic Reticulum (ER) and Golgi Apparatus (GA), its partial localization to mitochondria is also shown [78, 79]. Thus, the partial interaction with mitochondrial is possibly through the transmembrane domain of 7a interacting with pro-apoptotic proteins essential for apoptosis induction, which could favor SARS-CoV-2 release and subsequent infection of neighboring cells [78].

## Open reading frame 8a (ORF8a)

ORF8a has a 68% similarity in sequence between SARS-CoV and SARS-CoV-2, whose function is unclear [8]. However, it has been shown that SARS-CoV ORF8a, when expressed in Vero cells, ORF8a is localized explicitly to mitochondria [80]. In addition, ORF8a increases mitochondrial transmembrane potential, oxygen consumption, reactive oxygen species (ROS), and apoptosis by increasing caspase-3 activity [80]. Unlike ORF3a and ORF7a, which also induce apoptosis, ORF8a enhances viral replication in cultured cells [80]. Additionally, ORF8 possesses ER import signal and interacts with many ER proteins [63], which could also cause mitochondrial stress as most of the mitochondrial proteins are synthesized in ER and transported into mitochondria. Interaction with ORF8 is also evident from the work that demonstrates ORF8 could impair INF-1 signaling when expressed in cells [81].

### Open reading frame 9b (ORF9b)

Specifically, the open reading frame ORF9b protein of SARS-CoV bind the outer mitochondrial membrane in A549, HEK 293, and THP-1 cellular models and facilitate the degradation of Dynamin-related protein (Drp1) through ubiquitination that ultimately causes mitochondrial elongation [60]. Drp1 is a GTPase required for mitochondrial fission, an essential process by which dysfunctional mitochondria are eliminated through mitophagy [82-86]. However, the exact consequence of this mitochondrial elongation is unclear, but it is believed to alter mitochondrial dynamics, favoring viral replication. In addition, ORF9b also manipulates the mitochondrial antiviral signaling (MAVS) protein through modulation of poly (C)-binding protein 2 (PCBP2) and a E3 Ubiquitin Protein Ligase, AIP4 in a way that degrades MAVS and TNF Receptor Associated Factors (TRAF) 3 and 6, which lead to decrease in IFN mediated antiviral response [60]. Since SARS-CoV-2 also possesses ORF9b, similar interactions may be possible between SARS-CoV-2 and mitochondria. A recent study confirms that SARS-CoV-2 ORF9b localizes to the outer mitochondrial membrane, associates with TOM70 and inhibits IFN-1 response [87]. The same study clearly shows the interaction between SARS-COV-2 ORF9b and TOM70 demonstrated by co-immunoprecipitation and biotin-streptavidin affinity purification mass spectrometry approaches [87]. Further, it has also been shown that the core and C-terminal domains, but not the transmembrane and clamp domains of TOM70 are required for such interaction [87]. Interestingly, overexpression of Tom70 rescued ORF9a inhibited INF signaling. This further supports the notion that Tom70's role in activating MAVS protein that promotes apoptosis during viral infection, which is probably altered by SARS-CoV-2 infection [88, 89].

## Open reading frame 9c (ORF9c)

Another SARS-CoV-2 viral protein, ORF9c interacts with mitochondrial proteins such as NDUFAF1 and NDUFAB1 [8, 23]. It is known that NDUFAF1 is one of the critical players involved in the assembly of Complex I, which could be affected by the interaction of ORF9c. However, NDUFAB1 is one of the subunits of Complex I required for optimal bioenergetics [90]. The interaction of ORF9c with these mitochondrial proteins may affect the initiation of electron flow and ROS production, which is necessary for optimal cellular signaling and immune response [91, 92].

So far, it is evident that SARS-CoV-2 proteins interact with mitochondrial proteins (NDUFAF1, NDUFAF2, NDUFB9, MRPS2, MRPS5, MRPS25, MRPS27) that play crucial roles in the mitochondrial metabolic pathways, required investigation to understand the mechanistic pathway of COVID-19 pathologies [23]. Other viral interactions of significance also include Tom 70, a mitochondrial importer that plays a critical role in transporting proteins into the mitochondria and, more importantly, in modulating antiviral cellular defense pathways [23, 93]. Such interactions provide viral manipulation of the host mitochondria that suppresses immunity and promotes disease progression (Fig. 3). However, a detailed and comprehensive study may require identifying the crucial mitochondrial proteins targeted by SARS-CoV-2. Preventing interactions between SARS-CoV-2 and its potential mitochondrial protein targets during virus establishment is a possible area of research in identifying potential therapeutic strategies. Recent studies have shown that MAVS is activated by the RIG-1, which can sense the presence of viral RNA [94, 95]. By interacting with viperin, an antiviral protein, MAVS can also affect interferon (IFN $\beta$ ) levels, thereby acting as a means of antiviral defense [96]. Although no substantial evidence is available, it's been proposed that SARS-CoV-2 hijack host mitochondria by all the above means to suppress immunity that aids in manipulating mitochondrial function, including the immune pathways involving the MAVS protein.



## SARS-CoV-2 manipulation of mitochondrial machinery and emergence of cytokine storm

Mitochondria are capable of altering signaling and metabolic pathways and the transcription of genes within immune cells. For example, mitochondria can switch the phenotype between pro and anti-inflammatory in immune cells [97, 98]. Furthermore, the virus relies on the mitochondria's energy production for sustenance, which leads to the theory that modulation of mitochondrial metabolism may be effective against the virus. It is also known that replication of this virus relies on the production of doublemembrane vesicles (DMVs) from the endoplasmic reticulum [99–102]. The virus replicates on these DMVs and uses them to escape the host cell's immune defenses. Thus, it is theorized that SARS-CoV-2 manipulates mitochondria by forming double-membrane mitochondrial-derived vesicles (MDVs) [8, 103]. However, there is still no clear evidence proving MDVs promote viral replication.

Although evidence for direct manipulation of SARS-CoV-2 on mitochondria is emerging, the indirect effect of SARS-CoV-2 on mitochondria through modulating cell surface entry proteins is also possible. For example, studies have shown that ACE-2 regulates mitochondrial function [104, 105]. Also, genetic variations in ACE-2 are shown to affect mitochondrial function, as substantial variation in ACE-2 has been reported around the world, but whether such variations alter mitochondrial function and SARS-CoV-2 infection are yet to be established [106].

The cytokine storm, an extremely severe immune response, is a detrimental effect of COVID-19. The viral infection triggers inflammatory signals such as interleukins, interferons, and tumor necrosis factors during this response, leading to an excessive release of cytokines into the bloodstream. This process leads to the development of adverse side effects in the body, including tissue damage, increased cell debris known as damage-associated molecular patterns (DAMPs), and hypoxia. In addition, acute injury can ultimately lead to multiorgan failure in the most severe of cases.

As mitochondria are shown to be one of the critical components in eliciting the innate immune response, specifically in response to viruses, the SARS-CoV-2 mediated immune response involves mitochondria as well [61, 107, 108]. In support of this hypothesis, a master regulator analysis of combined SARS-CoV-2 specific interactome with MERS and SARS- CoV transcriptome using the human lung RNA-sequence data set validates not only the interaction of SARS-CoV-2 with ACE-2 and TMPRSS but also with specific mitochondrial proteins [10]. These mitochondrial proteins include MCL-1, a regulator of apoptosis, and the Complex I subunit NDUFA10 network, which are downregulated by SARS-CoV-2 [10]. However, the proposed mechanism is that the virus strategizes to down-regulate mitochondrial function and uses host cells for its replication. At normal physiological conditions, the MAVS protein present on the mitochondrion's outer membrane interacts with mitofusin-2 (Mfn2), which is essential for mitochondrial fusion. During viral entry, associated mitochondrial membranes at the ER tether mitochondria using Mfn2 and RIG-1. Using other protein recruits, the virus binds to MAVS, which then induces phosphorylation and nuclear translocation of IRF3, resulting in the production of cytokines, including interferon I and III, through activation of NFKba and IRFs (3/7), respectively [109]. As in vitro data show that MAVS is required to induce IFN production by activating NFK $\beta$  and IRF3, recent evidence also shows that mice lacking MAVS failed to induce IFN production in response to viral infection. This suggests the possible involvement of mitochondria in the viral-mediated immune response [108] (Fig. 3).

A clue from a previous study carried out in peripheral blood mononuclear cells (PBMCs) infected with SARS-CoV shows that upregulation of genes encoding mtDNA as well as genes involved in oxidative stress, heat shock, and transcription, coincides with cytokine elevation, compared to PBMCs from control samples [110–112]. The upregulated mtDNA genes include 16S rRNA (mito-chondrial ribosomal subunit), NADH dehydrogenase subunit 1 (ND1), and cytochrome c oxidase subunit I (COX1), whereas the genes involved in oxidative stress are peroxiredoxin 1 (PRDX1) and ferritin heavy polypeptide 1 (FTH1), heat shock response DnaJ (Hsp40), Dnaj homolog, subfamily B, member 1 (DNAJB1), and cytokine IL-1B as well [112]. These SARS-CoV-infected

patients are also shown to have a significantly increased number of mitochondria in their PBMCs compared to PBMCs from control subjects. This may be one of the reasons why more mtDNA gene expression is observed. Interestingly, the electron microscopic structures of PBMCs from SARS-CoV infected patients show increased lysosome-like granules, which is not seen in the control. This could be due to mitochondria's activation in PBMCs, causing an increased immune response and cytokine storm. However, we hypothesize that this could be more cell-specific as it depends on whether the cell is more immunogenic or not. Likewise, a recent study has shown a similar observation of SARS-CoV-2 infection in PBMCs and lymphocytes along with the cytokine storm. However, the involvement of mitochondria, which is likely, is not investigated [113]. This study further shows that cytokines IL-6, IL-10, IL-2 and IFN-γ are significantly increased in severe COVID-19 cases than mild cases, displaying that these levels are associated with the disease severity. Therefore, a possible cytokine storm induced by mitochondria in COVID-19 could be heterogeneous among the infected population and also could be influenced by pre-existing metabolic dysfunction, which further determines the intensity of the cytokine storm and whether it eliminates the virus or causes multiorgan failure.

Identifying the critical players of sepsis and inflammatory response resulting from SARS-CoV-2 infection is essential to developing better diagnostic and therapeutic strategies. It is known that during sepsis, the infected bacteria release their DNA, which, along with the other proteins, are recognized as pathogen-associated molecular patterns (PAMPs), which stimulate the inflammatory response in the host cells. In this process, released DNA binds to the toll-like receptor 9 (TLR 9) and the formyl peptides bind to the formyl peptide receptor-1 (FPR1) on the surface of host cells. This further releases cytokines by activating p38 MAP kinase (MAPK) and attracting neutrophils while triggering immune response [114]. On the other hand, a similar pattern is also observed with no infection, but any trauma or damage to the system by external stimuli could result in the release of DAMPs, which could initiate an inflammatory response similar to PAMPs. A breakthrough study has found that mitochondria are evolutionarily conserved bacteria, sharing a similar structural motif with prokaryotes. Mitochondria could release their DNA (mtDNA) and peptides (formyl peptides), which are recognized as DAMPS similar to PAMPs, suggesting that bacteria and mitochondria use a similar tactic while eliciting an immune response. Some of the DAMPs released by the mitochondria are mtDNA, TFAM, formyl peptides and ROS [115-118]. Specifically, formyl peptides are present only in bacteria and mitochondria in nature, suggesting that the injury response caused by DAMPs is analogous to sepsis caused by bacterial infection.

Furthermore, these groups show that intravenous injection of mitochondrial DAMPs causes severe systemic inflammation, including severe lung injury, most commonly observed in COVID-19 [117, 119-121]. Also, ATP required for immune cells comes from mitochondria. In contrast, mitochondria regulate calcium buffering and ROS, which are critical components for antigen-presenting, processing and activation of signaling pathways containing inflammatory proteins [122, 123]. Specifically, in T cells, deficiency of mitochondrial transcription factor TFAM causes energy deficiency, resulting in T cell metabolic failure. This induces the circulation of cytokines, thereby establishing chronic inflammation and senescence phenotype [124]. Additionally, autophagy, a mechanism in which viruses and their proteins are eliminated by being presented to lysosomes, is shown to be impaired during mitochondrial dysfunction in SARS-CoV/CoV-2, resulting in decreased autophagy in T cells, thereby establishing the infection [125]. This is in agreement with the observation of life-threatening casualties in COVID-19 subjects with metabolic compromised pre-existing conditions such as cancer, heart diseases, diabetes, aging, obesity and chronic obstructive pulmonary disease (COPD) [126-136].

## Mitochondria's role in SARS-CoV-2 associated comorbidities

Most of the metabolic syndromes such as diabetes, obesity, and aging are associated with mitochondrial dysfunction, which influences high-risk groups susceptible to SARS-CoV-2 infection, leading to an increase in mortality rate in these groups [131-135, 137-139]. Metabolic dysfunctions in states such as obesity and diabetes are significant players in comorbidities with COVID-19 [140-142]. This makes one speculate the importance of good mitochondrial health in resisting COVID-19 infection. Based on recent observations, COVID-19 can progress to a more severe version of respiratory disease accompanied by hyper inflammation, multiorgan failure, and death in patients with immunocompromising and pre-existing conditions such as diabetes, cardiovascular disease, and renal disease, digestive, cancer, COPD, and immunodeficiency [143]. A study also revealed an association between heart disease and COVID-19, in which congestive heart failure is one of the most common comorbidity with a 42.9% rate in patients diagnosed with COVID-19, which itself can cause heart injury [144, 145].

Interestingly, a recent study found evidence that children possess healthy mitochondria, which could protect them from severe COVID-19 pathology compared to the adult population [146]. For example, Lymphopenia is most frequently observed in the adult population infected with SARS-CoV-2 compared to children, who show milder or no symptoms of lymphopenia [146]. In this condition, T lymphocytes are reduced in adults with worsening conditions of COVID-19 when compared to children. Interestingly, it has been found that mitochondria-dependent apoptosis is elevated in T cells of adults infected with SARS-CoV-2, whereas it is not in children with COVID-19. Based on these in vitro studies, it has also been shown that increased IL-6 and TNF-  $\alpha$ co-stimulated by ROS decreased the removal of dysfunctional mitochondria, resulting in increased mitochondrial-mediated T cell apoptosis [146]. One of the reasons could be that association of dysfunctional mitochondria with aging may cause such differences in the pathologies. Although there is a high possibility for dysfunctional mitochondrial involved in these conditions, detailed studies are required to confirm these observations.

Another theory put forward is impaired mitophagy, causing mitochondrial dysfunction in COVID-19. Mitophagy removes damaged or dysfunctional mitochondria to maintain optimal bioenergetics and cellular metabolism, further maintaining cellular homeostasis. Impaired mitophagy is one of the reasons for mitochondrial dysfunction in metabolic disorders and aging [147, 148]. In addition, viruses are believed to affect mitophagy by interfering with mitochondrial fission, fusion, and membrane potential that further causes mitochondrial stress, ROS generation and decrease ATP generation to suppress overall mitochondrial health to establish its infection [64]. Some of the viruses directly or indirectly induce mitophagy to inhibit apoptosis and prevent innate immune responses from developing their infection [149]. A recent study shows that SARS-CoV-2 interfere and inhibits autophagy by reprogramming cellular metabolism, which could be prevented by treating with autophagy inducing drugs that further prevented SARS-CoV-2 propagation [150]. Therefore, existing drugs that improve mitochondrial health could help prevent or reduce SARS-CoV-2 infection is a potential area of research.

## Potential COVID-19 treatments and role of mitochondria

Until the recent rollout of vaccines, the prevention for COVID-19 had been unpredictable. Despite vaccine administration, people infected with SARS-CoV-2 are at high risk for developing and spreading COVID-19. Hence, treatment strategies to cure or prevent severe COVID-19 pathology or morbidity is an unmet need.



Currently, a variety of compounds and targets have been investigated as potential drug molecules in treating COVID-19 (Fig. 4).

The first-ever USFDA approved drug to treat COVID-19 is remdesivir, an adenine nucleoside analog that targets the RNA-dependent RNA polymerase, which catalyzes the replication of viral RNA. Remedesivir interferes with the action of the RNA polymerization resulting in decreased viral RNA replication, shown to be effective against SARS-CoV and MERS-CoV in early studies [151]. Although remdesivir showed a superior effect against SARS-CoV-2 when compared to placebo in shortening the time to recovery in adults with SARS-CoV 2 mediated lower respiratory tract infection, remedesivir alone did not change the mortality rate, suggesting additional therapecutic approaches needed in efficetnly treating COVID-19. Interestingly, remdesivir also inhibits mtDNA-dependent RNA polymerase but at lower efficiency and inhibits mitochondrial fission mediator Drp1 [152, 153]. Also, remdesivir has been shown to produce severe adverse effects, which are probably associated with mitochondrial toxicity because mitochondrial polymerases lack the selectivity of mammalian counterparts to exclude nucleoside analogs [154, 155]. It has also been identified that remdesivir causes inhibition of mitochondrial respiration through targeting various mitochondrial proteins, including those belonging to respiratory chain complexes [156]. Inhibiting the mitochondrial respiration pathway can lead to the failure of many other significant biochemical pathways in human that could be one of the reasons limiting remdesivir's high dose toxicity, however whether its inhibitory action of viral replication is through mitochondrial modulation not clear.

Ribavirin is another guanine nucleoside analog that prevents viral replication and translation by targeting the RNA-dependent RNA polymerase (RdRp), similar to remdesivir. A study conducted regarding the therapeutic effects of ribavirin with interferon-alpha 2b against MERS-CoV showed some improvement in patients but showed severe adverse impact on SARS-CoV patients [157, 158]. Also, in a recent study in patients with severe COVID-19, ribavirin has not improved the mortality rate [159]. Furthermore, being a nucleoside analog, ribavirin showed various toxicities, including multiorgan dysfunction, lactic acidemia, and mitochondrial toxicity [160]. Another purine nucleotide analog is a favipiravir obtained from the pro drug ribofuranosyl-5'-triphosphate or T-705, which has also been recently investigated for SARS-CoV-2 and showed viral clearance and clinical improvement within 14 days in patients with a range of mild to moderate illness. However, the study also suggested further detailed dose and duration-dependent trials for the definitive conclusion [161]. Umifenovir, which is an antiviral drug that inhibits the fusion of virus and host cell, is believed to target spike protein and host cell ACE-2 interaction. Umifenovir, when compared to lopinavir and ritonavir combined, significantly improved clinical outcomes in a recent study that suggested conducting a larger multicenter trial for umifenovir against COVID-19 [162].

Drugs such as lopinavir, ritonavir, and darunavir alone or in combination are also considered as potential drug candidates for the COVID-19 treatment. These drugs are inhibitors of protease that are crucial in processing the human immunodeficiency virus-1 (HIV-1) replication. Initially, these drug combinations were USFDA approved for HIV-1 treatment as the combination of drugs acts as antiretroviral agents. Lopinavir-ritonavir has shown past clinical studies on SARS-CoV infection, where mild improvements have been demonstrated in patients, but nothing significantly different [163]. In contrast, a clinical study on COVID- 19 patients regarding the drug efficacy showed no therapeutic benefits in hospitalized patients, but further trials in severely ill populations were recommended [164]. Cobicistat is another drug that acts as a cytochrome P450 3A isoform inhibitor, thereby increasing the absorption of co-administered drugs that act on viral replication. A clinical study showed that darunavir has no substantial antiviral activity against the SARS-CoV-2 virus [165]. Recently, a combination of darunavir/ cobicistat for five days has been tested and did not show

any substantial effect againt COVID-19. However, the limitation of small sample size and open-label design has been limited in this study [166].

Other potential drug candidates to treat COVID-19 under controversy are chloroquine diphosphate (CQ) and hydroxychloroquine (HCQ), which are 4-aminoquinoline class drugs, USFDA approved for the treatment of malaria and rheumatoid arthritis. Multiple clinical trials have been initiated to study the effects of these anti-malarial drugs on COVID-19 disease, and it is getting controversial as only one report showed a beneficial effect [167]. There are many proposed theories that CQs inhibit cellular entry of the virus by inhibiting SARS-CoV- 2 uptakes by ACE-2 receptor and endocytosis [168-171]. It has also been known that CQ and HCQ behave like a weak base and act through various intracellular mechanisms such as inhibiting the intracellular enzyme. It inhibits viral entry through pH-dependent endocytosis as the virus majorly relies on pH for its establishment [169, 172, 173]. These drugs are shown to targets explicitly the ACE2 receptor, which prevents the virus from entering the cell membrane by inhibiting interaction with S-protein [168]. Although in vitro and some human clinical studies show promising results, the outcomes of various human clinical trials showed no clear benefit of using these drugs to treat COVID-19 [174-176]. Also, the impact of CQs on mitochondrial function are not so intensively investigated, but it has been shown that CQ inhibits mitochondrial respiration, ATP production, and function and whether this influences the treatment is not clear [177, 178]. Also, whether its combination with other therapeutic strategies would be beneficial against COVID-19 requires further investigation.

Interestingly, toremifene, an FDA-approved anti-cancer drug that belongs to the category of a selective estrogen receptor modulator that selectively targets breast cancer, has also been shown to be a potential SARS-CoV 2 inhibitor [179, 180]. An initial study using human coronavirus host interaction and network proximity analyses of drug targets, and systems pharmacology approach identified that toremifene could be a potential drug againt SARS-CoV-2 [179]. Earlier, toremifene was shown effective against MERS-CoV and SARS-CoV [181]. Furthermore, by employing homology modeling and other bioinformatic tools, toremifene has been shown to inhibit the spike glycoprotein. In addition, a strong interaction has also been observed between toremifene and NSP14 of SARS-CoV-2, suggesting a potential therapeutic drug for COVID-19 that needs detailed investigation [180].

Other potential drugs belong to corticosteroids (e.g., dexamethasone, hydrocortisone, prednisolone), which decrease inflammatory response against influenza infections, SARS-CoV and MERS-CoV, whereas they have also

## Table 1 SARS-CoV-2 interaction with mitochondrial function

SARS-CoV-2 Target	Interaction	Function [Reference]
Acyl-CoA Dehydrogenase Medium Chain (ACADM)	Structural protein "M'	Mitochondrial fatty acid β-oxidation (FAO) [23]
Mitochondrial-processing peptidase subunit beta (PMPCB)	Structural protein "M'	Mitochondrial precursor processing [23]
Pitrilysin metallopeptidase 1 (PITRM1)	Structural protein "M'	Mitochondrial precursor processing and degradation [23]
Mitochondrial-processing peptidase subunit alpha (PMPCA)	Structural protein "M'	Cleavage of the leader peptides of precursor proteins [23]
Coenzyme Q8B (COQ8B)	Structural protein "M'	transporting electrons along the respiratory chain of the mitochondrial inner membrane [23]
<sup>a</sup> Prohibitin 1(PHB1)	NSP2	Chaperone for ETC proteins [68]
<sup>a</sup> Prohibitin2 (PHB2)	NSP2	Chaperone for ETC proteins [68]
NADH dehydrogenase [ubiquinone] 1 alpha subcomplex assembly factor 2 (NDUFAF2)	NSP7	Chaperone for mitochondrial complex I assembly [23]
Mitochondrial ribosomal protein Subunits (MRPS2, S5, S25 and S27)	NSP8	Mitochondrial translation [23]
<sup>a</sup> NADH-ubiquinone oxidoreductase chain 4L (NADH4L)	NSP10	Complex   function [69]
<sup>a</sup> Cytochrome C oxidase subunit II (COX II)	NSP10	Complex IV function [69]
Mitochondrial ubiquitin-specific peptidase 30 (USP30)	ORF3a	Deubiquitinase [22]
<sup>a</sup> B-cell lymphoma-extra-large (Bcl-X <sub>L)</sub>	ORF7a	Apoptosis [78]
<sup>a</sup> Mitochondrial antiviral-signaling protein (MAVS)	ORF9b	Antiviral signaling [60]
<sup>a</sup> TRAF3 (TNF receptor associated factor 3),	ORF9b	Antiviral signaling [60]
<sup>a</sup> TRAF 6 (TNF receptor associated factor 6).	ORF9b	Antiviral signaling [60]
<sup>a</sup> Dynamin-related protein 1 (Drp1)	ORF9b	Mitochondrial fission [60]
Translocases of outer membrane 70, mitochondrial (Tom70)	ORF9b	Mitochondrial import [23, 87]
NADH dehydrogenase [ubiquinone] 1 alpha subcomplex assembly factor 1 (NDUFAF1)	ORF9c	Complex-I assembly factor [8, 23]
NADH: Ubiquinone Oxidoreductase Subunit AB1(NDUFAB1)	ORF9c	Complex I function [8, 23]

<sup>a</sup> Predicted interactions based on conserved structure and functions from SARS-CoV

been investigated against SARS-CoV-2 [182, 183]. Dexamethasone is effective in SARS-CoV-2 infected groups receiving mechanical ventilation and receiving oxygen that lowered the 28-day mortality rate, but not in groups with no kinds of support [183]. In addition, administration of methylprednisolone followed by prednisolone was also effective and showed better outcomes than the dexamethasone group against SARS-CoV-2 [184]. Out of many human clinical trials, dexamethasone is shown to be the first effective treatment that saved lives against COVID-19 [183, 185]. The primary mechanism of dexamethasone is its anti-inflammatory action through glucocorticoid receptors, which also translocate to mitochondria; however, the mitochondrial contribution to such effect needs to be explored [186].

Recently, melatonin has emerged as one of the potential enhancers/adjuvants in COVID-19 treatment [187]. Melatonin is a multifunctional molecule synthesized in mitochondria and possesses antioxidant, anti-inflammatory, and immunomodulatory functions that are shown to have a protective effect against COVID-19 [187–189]. As melatonin improves mitochondrial function by increasing oxidative phosphorylation, ATP production, upregulation of antioxidant enzymes, scavenging ROS and reactive nitrogen species (RNS), it has been proposed to test for its protective action against COVID-19 [188–194]. Although melatonin is not a standalone treatment against COVID-19, it has been suggested as a prophylactic molecule in combination with other drugs that could be useful in managing COVID-19 treatment [187].

Drugs such as camostat and nafamostate, which are TMPRS22 protease inhibitors, are also shown to be effective against SARS-CoV-2 [31, 195–197]. As TMPRS22 is responsible for SARS-CoV-2 cell entry through S protein activation, inhibiting TMPRS22 is an effective method to contain virus entry; clinical trials are underway worldwide. On the other hand, tocilizumab and sarilumab are monoclonal antibodies against IL-6 and believed to have a potential role against COVID-19 by inhibiting IL-6 mediated inflammation. However, a recent clinical trial found that these antibodies have not shown any better clinical outcomes or lower mortality rates alone or combined with remdesivir [198–201].

Altogether, based on the shreds of evidence that these drugs also influence mitochondrial function, the perspective of mitochondria as a potential target in treating COVID-19 is eminent, which is also supported by the evidence of mitochondrial manipulation by SARS-CoV-2 (Table 1). Therefore, targeting crucial interaction between SARS-CoV-2 components and mitochondrial proteins could be a potential strategy against COVID-19 (Table 1). But, many studies need to be conducted to establish mitochondria as a target in treating COVID-19. Strategies like improving mitochondrial function could by any means be detrimental to virus establishment. Therefore, treatment options can include compounds that modulate mitochondrial bioenergetic processes because the virus relies on mitochondria energy. Also, virus-induced mitochondrial dysfunction could be alleviated by increasing mitochondrial function, thereby preventing mitochondrial hijacking by the virus.

## **Conclusions and perspectives**

The sudden outbreak and rapid spread of SARS-CoV-2 posed a severe challenge to the global welfare framework. Developing treatments and immunizations requires a thorough and profound comprehension of the pathogenesis of these arising viruses. Although we initially know little about the SARS-CoV-2, past knowledge and work on SARS-CoV and MERS-CoV sped up our understanding of this novel COVID-19 in a brief timeframe. We have put forward various hypotheses about SARS-CoV-2 pathogenesis with different investigations, related explicitly to mitochondrial interactions as SARS-CoV-2 is proposed to hijack mitochondria leading to virus establishment in the host cells. Studies show that SARS-CoV-2 uses mitochondria to escape from immunity by altering metabolic pathways and suppressing the innate immune response by interacting with various components of host mitochondria. Furthermore, the other factors related to the mitochondrial function that influence SARS-CoV-2 establishment are age, immunosuppression, and metabolic disorders (age, diabetes, obesity). There is clear evidence that the elderly population is affected with severe infection from the virus, leading to a higher mortality rate. In conclusion, understanding the intricate relationship between SARS-CoV-2 and mitochondria is critical for developing treatment options and fighting against COVID-19 and other novel RNA viruses.

#### Abbreviations

ACADM: Acyl-CoA Dehydrogenase Medium Chain; ACE-2: Angiotensinconverting enzyme-2; Bcl-X<sub>L</sub>: B-cell lymphoma-extra-large; COPD: Chronic obstructive pulmonary disease; COQ8B: Coenzyme Q8B; PHB1: Prohibitin 1; COVID-19: Coronavirus disease 19; COX II: Cytochrome c oxidase subunit II; COX I: Cytochrome c oxidase subunit I; DMVs: Double-membrane vesicles; DNJB1: DNAJ homolog, subfamily B, member 1; DPP4: Dipeptidyl peptidase 4; Drp1: Dynamin-related protein 1; ERR0: Estrogen-related receptor alpha; FPR1: Formyl peptide receptor-1; FTH1: Ferritin heavy polypeptide 1; GTT : Gastrointestinal tract; HCQ: Hydroxychloroquine; MAPK: Mitogen-activated protein kinase; MAVS: Mitochondrial antiviral signaling; MDVs: Mitochondrialderived vesicles; MERS-CoV: Middle East respiratory syndrome-CoV; ND1: NADH dehydrogenase subunit 1; NDUFAF1: NADH dehydrogenase [ubiquinone] 1 alpha subcomplex assembly factor 1; NADH4L: NADH-Ubiquinone oxidoreductase chain 4L; NET: Neutrophil extracellular trap; PAMPs: Pathogenassociated molecular patterns; PCBP2: Poly (C)-binding protein 2; PHB2: Prohibitin2; PITRM1: Pitrilysin metallopeptidase 1; PMPCA: Mitochondrial-processing peptidase subunit alpha; PMPCB: Mitochondrial-processing peptidase subunit beta; PRDX1: Peroxiredoxin 1; SARS-CoV-2: Severe Acute Respiratory Syndrome-Corona Virus 2; TLR: Toll-like receptor 9; Tom70: Translocases of outer membrane 70, mitochondrial; TRAF: TNF Receptor Associated Factor; TRAF3: TNF receptor associated factor 3; TRAF6: TNF receptor associated factor 6; USP30: Ubiquitin specific peptidase 30.

## Acknowledgments

Not applicable.

#### Code availability

Not applicable.

## Authors' contributions

Kavya Srinivasan and Sundararajan Venkatesh drafted the manuscript. Kavya Srinivasan, Ashlena Livingston and Ashutosh Kumar Pandey conducted the literature search. Ashutosh Kumar Pandey prepared all the figures. Ashlena Livingston prepared the table. Sundararajan Venkatesh reviewed, proof read and edited the manuscript. The authors have reviewed and approved the manuscript before submission.

#### Funding

Venkatesh Sundararajan's laboratory is partially supported by the American Heart Association grants (20CDA35260096 and 20TPA3542000)

#### Availability of data and materials

Not applicable

### Declarations

**Ethics approval and consent to participate** Not applicable.

#### **Consent for publication**

Not applicable.

#### **Competing interests**

The authors declare that they have no conflict of interest.

#### Author details

<sup>1</sup>Department of Microbiology, Biochemistry and Molecular Genetics, Rutgers -New Jersey Medical School, The State University of New Jersey, Newark, NJ, USA. <sup>2</sup>New York Institute of Technology, Old Westbury, NY, USA. <sup>3</sup>Department of Pharmacology, Physiology and Neuroscience, Rutgers -New Jersey Medical School, The State University of New Jersey, Newark, NJ, USA. <sup>4</sup>Dallas Baptist University, Dallas, TX, United States.

### Received: 25 April 2021 Accepted: 4 November 2021 Published online: 23 November 2021

## References

- Who. Coronavirus (covid-19) dashboard. Available at: https://covid19. who.int/. Accessed Nov 3, 2021.
- Chakraborty C, Sharma AR, Bhattacharya M, Sharma G, SS L. The 2019 novel coronavirus disease (covid-19) pandemic: a zoonotic prospective. Asian Pac J Trop Med. 2020;(13):242–6.
- 3. Evans SJW, Jewell NP. Vaccine effectiveness studies in the field. N Engl J Med. 2021;385(7):650–1. https://doi.org/10.1056/NEJMe2110605.
- Chakraborty C, Sharma AR, Sharma G, Bhattacharya M, Lee SS. Sarscov-2 causing pneumonia-associated respiratory disorder (covid-19): Diagnostic and proposed therapeutic options. Eur Rev Med Pharmacol Sci. 2020;24(7):4016–26. https://doi.org/10.26355/eurrev\_202004\_ 20871.
- Thompson MG, Burgess JL, Naleway AL, Tyner H, Yoon SK, Meece J, et al. Prevention and attenuation of covid-19 with the bnt162b2 and

mrna-1273 vaccines. N Engl J Med. 2021;385(4):320–9. https://doi.org/ 10.1056/NEJMoa2107058.

- Lopez Bernal J, Andrews N, Gower C, Gallagher E, Simmons R, Thelwall S, et al. Effectiveness of covid-19 vaccines against the b.1.617.2 (delta) variant. N Engl J Med. 2021;385(7):585–94. https://doi.org/10.1056/ NEJMoa2108891.
- Bergwerk M, Gonen T, Lustig Y, Amit S, Lipsitch M, Cohen C, et al. Covid-19 breakthrough infections in vaccinated health care workers. N Engl J Med. 2021. https://doi.org/10.1056/NEJMoa2109072.
- Singh KK, Chaubey G, Chen JY, Suravajhala P. Decoding sars-cov-2 hijacking of host mitochondria in covid-19 pathogenesis. Am J Physiol Cell Physiol. 2020;319(2):C258–67. https://doi.org/10.1152/ajpcell. 00224.2020.
- Edeas M, Saleh J, Peyssonnaux C. Iron: Innocent bystander or vicious culprit in covid-19 pathogenesis? Int J Infect Dis. 2020;97:303–5. https:// doi.org/10.1016/j.ijid.2020.05.110.
- Guzzi PH, Mercatelli D, Ceraolo C, Giorgi FM. Master regulator analysis of the sars-cov-2/human interactome. J Clin Med. 2020;9(4). https://doi. org/10.3390/jcm9040982.
- Kloc M, Ghobrial RM, Kubiak JZ. The role of genetic sex and mitochondria in response to covid-19 infection. Int Arch Allergy Immunol. 2020;181(8):629–34. https://doi.org/10.1159/000508560.
- 12. Tyrrell DAJ, Fielder M. Cold wars : The fight against the common cold. Oxford: Oxford University Press; 2002.
- Petersen E, Koopmans M, Go U, Hamer DH, Petrosillo N, Castelli F, et al. Comparing sars-cov-2 with sars-cov and influenza pandemics. Lancet Infect Dis. 2020;20(9):e238–44. https://doi.org/10.1016/S1473-3099(20) 30484-9.
- 14. Liu DX, Liang JQ, Fung TS. Human coronavirus-229e, -oc43, -nl63, and -hku1. Reference module in life sciences. 2020:B978-970-912-809633-809638.821501-X.DOI:10.1016/B978-0-12-809633-8.21501-X.
- Memish ZA, Zumla AI, Al-Hakeem RF, Al-Rabeeah AA, Stephens GM. Family cluster of middle east respiratory syndrome coronavirus infections. N Engl J Med. 2013;368(26):2487–94. https://doi.org/10.1056/ NEJMoa1303729.
- Tiwari N, Upadhyay J, Nazam Ansari M, Joshi R. Novel beta-coronavirus (sars-cov-2): current and future aspects of pharmacological treatments. Saudi Pharm J. 2020. https://doi.org/10.1016/j.jsps.2020.08.015.
- Zhu N, Zhang D, Wang W, Li X, Yang B, Song J, et al. A novel coronavirus from patients with pneumonia in china, 2019. N Engl J Med. 2020;382(8):727–33. https://doi.org/10.1056/NEJMoa2001017.
- Sanche S, Lin YT, Xu C, Romero-Severson E, Hengartner N, Ke R. High contagiousness and rapid spread of severe acute respiratory syndrome coronavirus 2. Emerg Infect Dis. 2020;26(7):1470–7. https://doi.org/10. 3201/eid2607.200282.
- Peiris JS, Yuen KY, Osterhaus AD, Stohr K. The severe acute respiratory syndrome. N Engl J Med. 2003;349(25):2431–41. https://doi.org/10. 1056/NEJMra032498.
- Yadav R, Chaudhary JK, Jain N, Chaudhary PK, Khanra S, Dhamija P, et al. Role of structural and non-structural proteins and therapeutic targets of sarscov-2 for covid-19. Cells. 2021;10(4). https://doi.org/10.3390/cells10040821.
- Huang Y, Yang C, Xu XF, Xu W, Liu SW. Structural and functional properties of sars-cov-2 spike protein: Potential antivirus drug development for covid-19. Acta Pharmacol Sin. 2020;41(9):1141–9. https://doi.org/10. 1038/s41401-020-0485-4.
- Chan JF, Kok KH, Zhu Z, Chu H, To KK, Yuan S, et al. Genomic characterization of the 2019 novel human-pathogenic coronavirus isolated from a patient with atypical pneumonia after visiting wuhan. Emerg Microbes Infect. 2020;9(1):221–36. https://doi.org/10.1080/22221751. 2020.1719902.
- Gordon DE, Jang GM, Bouhaddou M, Xu J, Obernier K, White KM, et al. A sars-cov-2 protein interaction map reveals targets for drug repurposing. Nature. 2020;583(7816):459–68. https://doi.org/10.1038/ s41586-020-2286-9.
- Kim D, Lee JY, Yang JS, Kim JW, Kim VN, Chang H. The architecture of sars-cov-2 transcriptome. Cell. 2020;181(4):914–921 e910. https://doi. org/10.1016/j.cell.2020.04.011.
- Shang J, Wan Y, Luo C, Ye G, Geng Q, Auerbach A, et al. Cell entry mechanisms of sars-cov-2. Proc Natl Acad Sci U S A. 2020;117(21):11727–34. https://doi.org/10.1073/pnas.2003138117.

- 26. Ord M, Faustova I, Loog M. The sequence at spike s1/s2 site enables cleavage by furin and phospho-regulation in sars-cov2 but not in sars-cov1 or mers-cov. Sci Rep. 2020;10(1):16944. https://doi.org/10.1038/s41598-020-74101-0.
- Cubuk J, Alston JJ, Incicco JJ, Singh S, Stuchell-Brereton MD, Ward MD, et al. The sars-cov-2 nucleocapsid protein is dynamic, disordered, and phase separates with rna. Nat Commun. 2021;12(1):1936. https://doi. org/10.1038/s41467-021-21953-3.
- Neuman BW, Kiss G, Kunding AH, Bhella D, Baksh MF, Connelly S, et al. A structural analysis of m protein in coronavirus assembly and morphology. J Struct Biol. 2011;174(1):11–22. https://doi.org/10.1016/j.jsb.2010. 11.021.
- 29. Sarkar M, Saha S. Structural insight into the role of novel sars-cov-2 e protein: A potential target for vaccine development and other therapeutic strategies. PLoS One. 2020;15(8):e0237300. https://doi.org/10. 1371/journal.pone.0237300.
- Khailany RA, Safdar M, Ozaslan M. Genomic characterization of a novel sars-cov-2. Gene Rep. 2020;19:100682. https://doi.org/10.1016/j.genrep. 2020.100682.
- Hoffmann M, Kleine-Weber H, Schroeder S, Kruger N, Herrler T, Erichsen S, et al. Sars-cov-2 cell entry depends on ace2 and tmprss2 and is blocked by a clinically proven protease inhibitor. Cell. 2020;181(2):271– 280 e278. https://doi.org/10.1016/j.cell.2020.02.052.
- Walls AC, Park YJ, Tortorici MA, Wall A, McGuire AT, Veesler D. Structure, function, and antigenicity of the sars-cov-2 spike glycoprotein. Cell. 2020;181(2):281–292 e286. https://doi.org/10.1016/j.cell.2020.02.058.
- Hamming I, Timens W, Bulthuis ML, Lely AT, Navis G, van Goor H. Tissue distribution of ace2 protein, the functional receptor for sars coronavirus. A first step in understanding sars pathogenesis. J Pathol. 2004;203(2):631–7. https://doi.org/10.1002/path.1570.
- Crackower MA, Sarao R, Oudit GY, Yagil C, Kozieradzki I, Scanga SE, et al. Angiotensin-converting enzyme 2 is an essential regulator of heart function. Nature. 2002;417(6891):822–8. https://doi.org/10.1038/natur e00786.
- Donoghue M, Hsieh F, Baronas E, Godbout K, Gosselin M, Stagliano N, et al. A novel angiotensin-converting enzyme-related carboxypeptidase (ace2) converts angiotensin i to angiotensin 1-9. Circ Res. 2000;87(5):E1–9. https://doi.org/10.1161/01.res.87.5.e1.
- Tikellis C, Johnston CI, Forbes JM, Burns WC, Burrell LM, Risvanis J, et al. Characterization of renal angiotensin-converting enzyme 2 in diabetic nephropathy. Hypertension. 2003;41(3):392–7. https://doi.org/10.1161/ 01.HYP.0000060689.38912.CB.
- Song W, Gui M, Wang X, Xiang Y. Cryo-em structure of the sars coronavirus spike glycoprotein in complex with its host cell receptor ace2. PLoS Pathog. 2018;14(8):e1007236. https://doi.org/10.1371/journal.ppat. 1007236.
- Paules CI, Marston HD, Fauci AS. Coronavirus infections-more than just the common cold. JAMA. 2020. https://doi.org/10.1001/jama.2020.0757.
- Li F, Li W, Farzan M, Harrison SC. Structure of SARS coronavirus spike receptor-binding domain complexed with receptor. Science. 2005;309(5742):1864–8. https://doi.org/10.1126/science.1116480.
- Park YJ, Walls AC, Wang Z, Sauer MM, Li W, Tortorici MA, et al. Structures of mers-cov spike glycoprotein in complex with sialoside attachment receptors. Nat Struct Mol Biol. 2019;26(12):1151–7. https://doi.org/10. 1038/s41594-019-0334-7.
- Li R, Qiao S, Zhang G. Analysis of angiotensin-converting enzyme 2 (ace2) from different species sheds some light on cross-species receptor usage of a novel coronavirus 2019-ncov. J Infect. 2020;80(4):469–96. https://doi.org/10.1016/j.jinf.2020.02.013.
- 42. Sungnak W, Huang N, Becavin C, Berg M, Network HCALB. Sars-cov-2 entry genes are most highly expressed in nasal goblet and ciliated cells within human airways. ArXiv. 2020
- Saheb Sharif-Askari N, Saheb Sharif-Askari F, Alabed M, Temsah MH, Al Heialy S, Hamid Q, et al. Airways expression of sars-cov-2 receptor, ace2, and tmprss2 is lower in children than adults and increases with smoking and copd. Mol Ther Methods Clin Dev. 2020;18:1–6. https://doi.org/ 10.1016/j.omtm.2020.05.013.
- Al-Benna S. Association of high level gene expression of ace2 in adipose tissue with mortality of covid-19 infection in obese patients. Obes Med. 2020;19:100283. https://doi.org/10.1016/j.obmed.2020.100283.

- Edler C, Schroder AS, Aepfelbacher M, Fitzek A, Heinemann A, Heinrich F, et al. Dying with sars-cov-2 infection-an autopsy study of the first consecutive 80 cases in hamburg, germany. Int J Legal Med. 2020;134(4):1275–84. https://doi.org/10.1007/s00414-020-02317-w.
- Sarkesh A, Daei Sorkhabi A, Sheykhsaran E, Alinezhad F, Mohammadzadeh N, Hemmat N, et al. Extrapulmonary clinical manifestations in covid-19 patients. Am J Trop Med Hyg. 2020;103(5):1783–96. https:// doi.org/10.4269/ajtmh.20-0986.
- Dahiya DS, Kichloo A, Albosta M, Pagad S, Wani F. Gastrointestinal implications in covid-19. J Investig Med. 2020;68(8):1397–401. https:// doi.org/10.1136/jim-2020-001559.
- Gavriatopoulou M, Korompoki E, Fotiou D, Ntanasis-Stathopoulos I, Psaltopoulou T, Kastritis E, et al. Organ-specific manifestations of covid-19 infection. Clin Exp Med. 2020;20(4):493–506. https://doi.org/10.1007/ s10238-020-00648-x.
- Gupta A, Madhavan MV, Sehgal K, Nair N, Mahajan S, Sehrawat TS, et al. Extrapulmonary manifestations of covid-19. Nat Med. 2020;26(7):1017– 32. https://doi.org/10.1038/s41591-020-0968-3.
- Daly JL, Simonetti B, Klein K, Chen KE, Williamson MK, Anton-Plagaro C, et al. Neuropilin-1 is a host factor for sars-cov-2 infection. Science. 2020;370(6518):861–5. https://doi.org/10.1126/science.abd3072.
- Cantuti-Castelvetri L, Ojha R, Pedro LD, Djannatian M, Franz J, Kuivanen S, et al. Neuropilin-1 facilitates sars-cov-2 cell entry and infectivity. Science. 2020;370(6518):856–60. https://doi.org/10.1126/science. abd2985.
- 52. Wang K, Chen W, Zhang Z, Deng Y, Lian JQ, Du P, et al. Cd147-spike protein is a novel route for sars-cov-2 infection to host cells. Signal Transduct Target Ther. 2020;5(1):283. https://doi.org/10.1038/s41392-020-00426-x.
- Perlman S, Netland J. Coronaviruses post-sars: update on replication and pathogenesis. Nat Rev Microbiol. 2009;7(6):439–50. https://doi.org/ 10.1038/nrmicro2147.
- Pandey AK, Pain J, Dancis A, Pain D. Mitochondria export iron-sulfur and sulfur intermediates to the cytoplasm for iron-sulfur cluster assembly and trna thiolation in yeast. J Biol Chem. 2019;294(24):9489–502. https://doi.org/10.1074/jbc.RA119.008600.
- Spinelli JB, Haigis MC. The multifaceted contributions of mitochondria to cellular metabolism. Nat Cell Biol. 2018;20(7):745–54. https://doi.org/ 10.1038/s41556-018-0124-1.
- Duchen MR. Mitochondria and calcium: from cell signalling to cell death. J Physiol. 2000;529(Pt 1):57–68. https://doi.org/10.1111/j.1469-7793.2000.00057.x.
- Silva da Costa L, Pereira da Silva AP, Da Poian AT, El-Bacha T. Mitochondrial bioenergetic alterations in mouse neuroblastoma cells infected with sindbis virus: Implications to viral replication and neuronal death. PLoS One. 2012;7(4):e33871. https://doi.org/10.1371/journal.pone.0033871.
- Jang KJ, Jeong S, Kang DY, Sp N, Yang YM, Kim DE. A high atp concentration enhances the cooperative translocation of the sars coronavirus helicase nsp13 in the unwinding of duplex rna. Sci Rep. 2020;10(1):4481. https://doi.org/10.1038/s41598-020-61432-1.
- Kim SJ, Syed GH, Khan M, Chiu WW, Sohail MA, Gish RG, et al. Hepatitis c virus triggers mitochondrial fission and attenuates apoptosis to promote viral persistence. Proc Natl Acad Sci U S A. 2014;111(17):6413–8. https://doi.org/10.1073/pnas.1321114111.
- Shi CS, Qi HY, Boularan C, Huang NN, Abu-Asab M, Shelhamer JH, et al. Sars-coronavirus open reading frame-9b suppresses innate immunity by targeting mitochondria and the mavs/traf3/traf6 signalosome. J Immunol. 2014;193(6):3080–9. https://doi.org/10.4049/jimmunol. 1303196.
- Chatel-Chaix L, Cortese M, Romero-Brey I, Bender S, Neufeldt CJ, Fischl W, et al. Dengue virus perturbs mitochondrial morphodynamics to dampen innate immune responses. Cell Host Microbe. 2016;20(3):342– 56. https://doi.org/10.1016/j.chom.2016.07.008.
- Barbier V, Lang D, Valois S, Rothman AL, Medin CL. Dengue virus induces mitochondrial elongation through impairment of drp1-triggered mitochondrial fission. Virology. 2017;500:149–60. https://doi.org/ 10.1016/j.virol.2016.10.022.
- Gordon DE, Hiatt J, Bouhaddou M, Rezelj VV, Ulferts S, Braberg H, et al. Comparative host-coronavirus protein interaction networks reveal panviral disease mechanisms. Science. 2020;370(6521). https://doi.org/10. 1126/science.abe9403.

- Singh SP, Amar S, Gehlot P, Patra SK, Kanwar N, Kanwal A. Mitochondrial modulations, autophagy pathways shifts in viral infections: consequences of covid-19. Int J Mol Sci. 2021;22(15). https://doi.org/10.3390/ ijms22158180.
- de Haan CA, Vennema H, Rottier PJ. Assembly of the coronavirus envelope: Homotypic interactions between the m proteins. J Virol. 2000;74(11):4967–78. https://doi.org/10.1128/jvi.74.11.4967-4978.2000.
- Zhao G, Shi SQ, Yang Y, Peng JP. M and n proteins of sars coronavirus induce apoptosis in hpf cells. Cell Biol Toxicol. 2006;22(5):313–22. https://doi.org/10.1007/s10565-006-0077-1.
- Cornillez-Ty CT, Liao L, Yates JR 3rd, Kuhn P, Buchmeier MJ. Severe acute respiratory syndrome coronavirus nonstructural protein 2 interacts with a host protein complex involved in mitochondrial biogenesis and intracellular signaling. J Virol. 2009;83(19):10314–8. https://doi.org/10.1128/JVI.00842-09.
- Tatsuta T, Model K, Langer T. Formation of membrane-bound ring complexes by prohibitins in mitochondria. Mol Biol Cell. 2005;16(1):248–59. https://doi.org/10.1091/mbc.e04-09-0807.
- Li Q, Wang L, Dong C, Che Y, Jiang L, Liu L, et al. The interaction of the sars coronavirus non-structural protein 10 with the cellular oxidoreductase system causes an extensive cytopathic effect. J Clin Virol. 2005;34(2):133–9. https://doi.org/10.1016/j.jcv.2004.12.019.
- Pasquier C, Robichon A. Computational search of hybrid human/sarscov-2 dsrna reveals unique viral sequences that diverge from those of other coronavirus strains. Heliyon. 2021;7(6):e07284. https://doi.org/10. 1016/j.heliyon.2021.e07284.
- 71. Schonrich G, Raftery MJ. Neutrophil extracellular traps go viral. Front Immunol. 2016;7:366. https://doi.org/10.3389/fimmu.2016.00366.
- White MJ, McArthur K, Metcalf D, Lane RM, Cambier JC, Herold MJ, et al. Apoptotic caspases suppress mtdna-induced sting-mediated type i ifn production. Cell. 2014;159(7):1549–62. https://doi.org/10.1016/j.cell. 2014.11.036.
- Rongvaux A, Jackson R, Harman CC, Li T, West AP, de Zoete MR, et al. Apoptotic caspases prevent the induction of type i interferons by mitochondrial DNA. Cell. 2014;159(7):1563–77. https://doi.org/10.1016/j.cell. 2014.11.037.
- West AP, Khoury-Hanold W, Staron M, Tal MC, Pineda CM, Lang SM, et al. Mitochondrial DNA stress primes the antiviral innate immune response. Nature. 2015;520(7548):553–7. https://doi.org/10.1038/nature14156.
- Aswani A, Manson J, Itagaki K, Chiazza F, Collino M, Wupeng WL, et al. Scavenging circulating mitochondrial DNA as a potential therapeutic option for multiple organ dysfunction in trauma hemorrhage. Front Immunol. 2018;9:891. https://doi.org/10.3389/fimmu.2018.00891.
- Freundt EC, Yu L, Park E, Lenardo MJ, Xu XN. Molecular determinants for subcellular localization of the severe acute respiratory syndrome coronavirus open reading frame 3b protein. J Virol. 2009;83(13):6631–40. https://doi.org/10.1128/JVI.00367-09.
- Schaecher SR, Touchette E, Schriewer J, Buller RM, Pekosz A. Severe acute respiratory syndrome coronavirus gene 7 products contribute to virus-induced apoptosis. J Virol. 2007;81(20):11054–68. https://doi.org/ 10.1128/JVI.01266-07.
- Tan YX, Tan TH, Lee MJ, Tham PY, Gunalan V, Druce J, et al. Induction of apoptosis by the severe acute respiratory syndrome coronavirus 7a protein is dependent on its interaction with the bcl-xl protein. J Virol. 2007;81(12):6346–55. https://doi.org/10.1128/JVI.00090-07.
- Nelson CA, Pekosz A, Lee CA, Diamond MS, Fremont DH. Structure and intracellular targeting of the sars-coronavirus orf7a accessory protein. Structure. 2005;13(1):75–85. https://doi.org/10.1016/j.str.2004.10.010.
- Chen CY, Ping YH, Lee HC, Chen KH, Lee YM, Chan YJ, et al. Open reading frame 8a of the human severe acute respiratory syndrome coronavirus not only promotes viral replication but also induces apoptosis. J Infect Dis. 2007;196(3):405–15. https://doi.org/10.1086/519166.
- Li JY, Liao CH, Wang Q, Tan YJ, Luo R, Qiu Y, et al. The orf6, orf8 and nucleocapsid proteins of sars-cov-2 inhibit type i interferon signaling pathway. Virus Res. 2020;286:198074. https://doi.org/10.1016/j.virusres. 2020.198074.
- Rana A, Oliveira MP, Khamoui AV, Aparicio R, Rera M, Rossiter HB, et al. Promoting drp1-mediated mitochondrial fission in midlife prolongs healthy lifespan of drosophila melanogaster. Nat Commun. 2017;8(1):448. https://doi.org/10.1038/s41467-017-00525-4.
- 83. Lin XH, Qiu BQ, Ma M, Zhang R, Hsu SJ, Liu HH, et al. Suppressing drp1-mediated mitochondrial fission and mitophagy increases

mitochondrial apoptosis of hepatocellular carcinoma cells in the setting of hypoxia. Oncogenesis. 2020;9(7):67. https://doi.org/10.1038/ s41389-020-00251-5.

- Vantaggiato C, Castelli M, Giovarelli M, Orso G, Bassi MT, Clementi E, et al. The fine tuning of drp1-dependent mitochondrial remodeling and autophagy controls neuronal differentiation. Front Cell Neurosci. 2019;13:120. https://doi.org/10.3389/fncel.2019.00120.
- Ikeda Y, Shirakabe A, Brady C, Zablocki D, Ohishi M, Sadoshima J. Molecular mechanisms mediating mitochondrial dynamics and mitophagy and their functional roles in the cardiovascular system. J Mol Cell Cardiol. 2015;78:116–22. https://doi.org/10.1016/j.yjmcc.2014. 09.019.
- Shirakabe A, Zhai P, Ikeda Y, Saito T, Maejima Y, Hsu CP, et al. Drp1dependent mitochondrial autophagy plays a protective role against pressure overload-induced mitochondrial dysfunction and heart failure. Circulation. 2016;133(13):1249–63. https://doi.org/10.1161/CIRCULATIO NAHA.115.020502.
- Jiang HW, Zhang HN, Meng QF, Xie J, Li Y, Chen H, et al. Sars-cov-2 orf9b suppresses type i interferon responses by targeting tom70. Cell Mol Immunol. 2020;17(9):998–1000. https://doi.org/10.1038/ s41423-020-0514-8.
- Lin R, Paz S, Hiscott J. Tom70 imports antiviral immunity to the mitochondria. Cell Res. 2010;20(9):971–3. https://doi.org/10.1038/cr.2010. 113.
- Wei B, Cui Y, Huang Y, Liu H, Li L, Li M, et al. Tom70 mediates sendai virus-induced apoptosis on mitochondria. J Virol. 2015;89(7):3804–18. https://doi.org/10.1128/JVI.02959-14.
- Hou T, Zhang R, Jian C, Ding W, Wang Y, Ling S, et al. Ndufab1 confers cardio-protection by enhancing mitochondrial bioenergetics through coordination of respiratory complex and supercomplex assembly. Cell Res. 2019;29(9):754–66. https://doi.org/10.1038/s41422-019-0208-x.
- 91. Jin Z, Wei W, Yang M, Du Y, Wan Y. Mitochondrial complex i activity suppresses inflammation and enhances bone resorption by shifting macrophage-osteoclast polarization. Cell Metab. 2014;20(3):483–98. https://doi.org/10.1016/j.cmet.2014.07.011.
- 92. West AP, Brodsky IE, Rahner C, Woo DK, Erdjument-Bromage H, Tempst P, et al. TIr signalling augments macrophage bactericidal activity through mitochondrial ros. Nature. 2011;472(7344):476–80. https://doi.org/10.1038/nature09973.
- Liu XY, Wei B, Shi HX, Shan YF, Wang C. Tom70 mediates activation of interferon regulatory factor 3 on mitochondria. Cell Res. 2010;20(9):994–1011. https://doi.org/10.1038/cr.2010.103.
- Furr SR, Moerdyk-Schauwecker M, Grdzelishvili VZ, Marriott I. Rig-i mediates nonsegmented negative-sense rna virus-induced inflammatory immune responses of primary human astrocytes. Glia. 2010;58(13):1620–9. https://doi.org/10.1002/glia.21034.
- Kowalinski E, Lunardi T, McCarthy AA, Louber J, Brunel J, Grigorov B, et al. Structural basis for the activation of innate immune patternrecognition receptor rig-i by viral rna. Cell. 2011;147(2):423–35. https:// doi.org/10.1016/j.cell.2011.09.039.
- Hee JS, Cresswell P. Viperin interaction with mitochondrial antiviral signaling protein (mavs) limits viperin-mediated inhibition of the interferon response in macrophages. PLoS One. 2017;12(2):e0172236. https://doi. org/10.1371/journal.pone.0172236.
- Mills EL, Kelly B, Logan A, Costa ASH, Varma M, Bryant CE, et al. Succinate dehydrogenase supports metabolic repurposing of mitochondria to drive inflammatory macrophages. Cell. 2016;167(2):457–470 e413. https://doi.org/10.1016/j.cell.2016.08.064.
- Mills EL, O'Neill LA. Reprogramming mitochondrial metabolism in macrophages as an anti-inflammatory signal. Eur J Immunol. 2016;46(1):13– 21. https://doi.org/10.1002/eji.201445427.
- Cortese M, Goellner S, Acosta EG, Neufeldt CJ, Oleksiuk O, Lampe M, et al. Ultrastructural characterization of zika virus replication factories. Cell Rep. 2017;18(9):2113–23. https://doi.org/10.1016/j.celrep.2017.02. 014.
- Knoops K, Kikkert M, Worm SH, Zevenhoven-Dobbe JC, van der Meer Y, Koster AJ, et al. Sars-coronavirus replication is supported by a reticulovesicular network of modified endoplasmic reticulum. PLoS Biol. 2008;6(9):e226. https://doi.org/10.1371/journal.pbio.0060226.
- 101. Maier HJ, Hawes PC, Cottam EM, Mantell J, Verkade P, Monaghan P, et al. Infectious bronchitis virus generates spherules from zippered

endoplasmic reticulum membranes. mBio. 2013;4(5):e00801–13. https://doi.org/10.1128/mBio.00801-13.

- Ulasli M, Verheije MH, de Haan CA, Reggiori F. Qualitative and quantitative ultrastructural analysis of the membrane rearrangements induced by coronavirus. Cell Microbiol. 2010;12(6):844–61. https://doi.org/10. 1111/j.1462-5822.2010.01437.x.
- Wu KE, Fazal FM, Parker KR, Zou J, Chang HY. Rna-gps predicts sarscov-2 rna residency to host mitochondria and nucleolus. Cell Syst. 2020;11(1):102–108 e103. https://doi.org/10.1016/j.cels.2020.06.008.
- 104. Cao X, Lu XM, Tuo X, Liu JY, Zhang YC, Song LN, et al. Angiotensin-converting enzyme 2 regulates endoplasmic reticulum stress and mitochondrial function to preserve skeletal muscle lipid metabolism. Lipids Health Dis. 2019;18(1):207. https://doi.org/10.1186/s12944-019-1145-x.
- Shi TT, Yang FY, Liu C, Cao X, Lu J, Zhang XL, et al. Angiotensin-converting enzyme 2 regulates mitochondrial function in pancreatic beta-cells. Biochem Biophys Res Commun. 2018;495(1):860–6. https://doi.org/10. 1016/j.bbrc.2017.11.055.
- Cao Y, Li L, Feng Z, Wan S, Huang P, Sun X, et al. Comparative genetic analysis of the novel coronavirus (2019-ncov/sars-cov-2) receptor ace2 in different populations. Cell Discov. 2020;6:11. https://doi.org/10.1038/ s41421-020-0147-1.
- Burtscher J, Cappellano G, Omori A, Koshiba T, Millet GP. Mitochondria: In the cross fire of sars-cov-2 and immunity. iScience. 2020;23(10):101631. https://doi.org/10.1016/j.isci.2020.101631.
- Sun Q, Sun L, Liu HH, Chen X, Seth RB, Forman J, et al. The specific and essential role of mavs in antiviral innate immune responses. Immunity. 2006;24(5):633–42. https://doi.org/10.1016/j.immuni.2006.04.004.
- Lin R, Heylbroeck C, Pitha PM, Hiscott J. Virus-dependent phosphorylation of the irf-3 transcription factor regulates nuclear translocation, transactivation potential, and proteasome-mediated degradation. Mol Cell Biol. 1998;18(5):2986–96. https://doi.org/10.1128/mcb.18.5.2986.
- Li L, Wo J, Shao J, Zhu H, Wu N, Li M, et al. Sars-coronavirus replicates in mononuclear cells of peripheral blood (pbmcs) from sars patients. J Clin Virol. 2003;28(3):239–44. https://doi.org/10.1016/s1386-6532(03)00195-1.
- 111. Li T, Qiu Z, Zhang L, Han Y, He W, Liu Z, et al. Significant changes of peripheral t lymphocyte subsets in patients with severe acute respiratory syndrome. J Infect Dis. 2004;189(4):648–51. https://doi.org/10. 1086/381535.
- Shao H, Lan D, Duan Z, Liu Z, Min J, Zhang L, et al. Upregulation of mitochondrial gene expression in pbmc from convalescent sars patients. J Clin Immunol. 2006;26(6):546–54. https://doi.org/10.1007/ s10875-006-9046-y.
- 113. Liu J, Li S, Liu J, Liang B, Wang X, Wang H, et al. Longitudinal characteristics of lymphocyte responses and cytokine profiles in the peripheral blood of sars-cov-2 infected patients. EBioMedicine. 2020;55:102763. https://doi.org/10.1016/j.ebiom.2020.102763.
- 114. Dorward DA, Lucas CD, Chapman GB, Haslett C, Dhaliwal K, Rossi AG. The role of formylated peptides and formyl peptide receptor 1 in governing neutrophil function during acute inflammation. Am J Pathol. 2015;185(5):1172–84. https://doi.org/10.1016/j.ajpath.2015.01.020.
- 115. Wenceslau CF, McCarthy CG, Goulopoulou S, Szasz T, NeSmith EG, Webb RC. Mitochondrial-derived n-formyl peptides: novel links between trauma, vascular collapse and sepsis. Med Hypotheses. 2013;81(4):532–5. https://doi.org/10.1016/j.mehy.2013.06.026.
- Wenceslau CF, McCarthy CG, Szasz T, Goulopoulou S, Webb RC. Mitochondrial n-formyl peptides induce cardiovascular collapse and sepsislike syndrome. Am J Physiol Heart Circ Physiol. 2015;308(7):H768–77. https://doi.org/10.1152/ajpheart.00779.2014.
- Zhang Q, Raoof M, Chen Y, Sumi Y, Sursal T, Junger W, et al. Circulating mitochondrial damps cause inflammatory responses to injury. Nature. 2010;464(7285):104–7. https://doi.org/10.1038/nature08780.
- Goulopoulou S, Matsumoto T, Bomfim GF, Webb RC. Toll-like receptor 9 activation: A novel mechanism linking placenta-derived mitochondrial DNA and vascular dysfunction in pre-eclampsia. Clin Sci. 2012;123(7):429–35. https://doi.org/10.1042/CS20120130.
- Chen LD, Zhang ZY, Wei XJ, Cai YQ, Yao WZ, Wang MH, et al. Association between cytokine profiles and lung injury in covid-19 pneumonia. Respir Res. 2020;21(1):201. https://doi.org/10.1186/s12931-020-01465-2.
- Li L, Huang Q, Wang DC, Ingbar DH, Wang X. Acute lung injury in patients with covid-19 infection. Clin Transl Med. 2020;10(1):20–7. https://doi.org/10.1002/ctm2.16.

- 121. Zhang L, Wang DC, Huang Q, Wang X. Significance of clinical phenomes of patients with covid-19 infection: a learning from 3795 patients in 80 reports. Clin Transl Med. 2020;10(1):28–35. https://doi.org/10.1002/ctm2.17.
- 122. Carr EL, Kelman A, Wu GS, Gopaul R, Senkevitch E, Aghvanyan A, et al. Glutamine uptake and metabolism are coordinately regulated by erk/ mapk during t lymphocyte activation. J Immunol. 2010;185(2):1037–44. https://doi.org/10.4049/jimmunol.0903586.
- Le A, Lane AN, Hamaker M, Bose S, Gouw A, Barbi J, et al. Glucoseindependent glutamine metabolism via tca cycling for proliferation and survival in b cells. Cell Metab. 2012;15(1):110–21. https://doi.org/10. 1016/j.cmet.2011.12.009.
- Desdin-Mico G, Soto-Heredero G, Aranda JF, Oller J, Carrasco E, Gabande-Rodriguez E, et al. T cells with dysfunctional mitochondria induce multimorbidity and premature senescence. Science. 2020;368(6497):1371–6. https://doi.org/10.1126/science.aax0860.
- Baixauli F, Acin-Perez R, Villarroya-Beltri C, Mazzeo C, Nunez-Andrade N, Gabande-Rodriguez E, et al. Mitochondrial respiration controls lysosomal function during inflammatory t cell responses. Cell Metab. 2015;22(3):485–98. https://doi.org/10.1016/j.cmet.2015.07.020.
- Robilotti EV, Babady NE, Mead PA, Rolling T, Perez-Johnston R, Bernardes M, et al. Determinants of covid-19 disease severity in patients with cancer. Nat Med. 2020;26(8):1218–23. https://doi.org/10.1038/ s41591-020-0979-0.
- Liang W, Guan W, Chen R, Wang W, Li J, Xu K, et al. Cancer patients in sars-cov-2 infection: A nationwide analysis in China. Lancet Oncol. 2020;21(3):335–7. https://doi.org/10.1016/S1470-2045(20)30096-6.
- 128. Montone RA, lannaccone G, Meucci MC, Gurgoglione F, Niccoli G. Myocardial and microvascular injury due to coronavirus disease 2019. Eur Cardiol. 2020;15:e52. https://doi.org/10.15420/ecr.2020.22.
- 129. Bohm M, Frey N, Giannitsis E, Sliwa K, Zeiher AM. Coronavirus disease 2019 (covid-19) and its implications for cardiovascular care: expert document from the german cardiac society and the world heart federation. Clin Res Cardiol. 2020. https://doi.org/10.1007/ s00392-020-01656-3.
- Guo T, Fan Y, Chen M, Wu X, Zhang L, He T, et al. Cardiovascular implications of fatal outcomes of patients with coronavirus disease 2019 (covid-19). JAMA Cardiol. 2020;5(7):811–8. https://doi.org/10.1001/ jamacardio.2020.1017.
- Zhu L, She ZG, Cheng X, Qin JJ, Zhang XJ, Cai J, et al. Association of blood glucose control and outcomes in patients with covid-19 and pre-existing type 2 diabetes. Cell Metab. 2020;31(6):1068–1077.e1063. https://doi.org/10.1016/j.cmet.2020.04.021.
- 132. Palaiodimos L, Kokkinidis DG, Li W, Karamanis D, Ognibene J, Arora S, et al. Severe obesity, increasing age and male sex are independently associated with worse in-hospital outcomes, and higher in-hospital mortality, in a cohort of patients with covid-19 in the bronx, new york. Metabolism. 2020;108:154262. https://doi.org/10.1016/j.metabol.2020. 154262.
- Anderson MR, Geleris J, Anderson DR, Zucker J, Nobel YR, Freedberg D, et al. Body mass index and risk for intubation or death in sars-cov-2 infection: a retrospective cohort study. Ann Intern Med. 2020. https:// doi.org/10.7326/M20-3214.
- 134. Klang E, Kassim G, Soffer S, Freeman R, Levin MA, Reich DL. Severe obesity as an independent risk factor for covid-19 mortality in hospitalized patients younger than 50. Obesity. 2020;28(9):1595–9. https://doi.org/ 10.1002/oby.22913.
- Lighter J, Phillips M, Hochman S, Sterling S, Johnson D, Francois F, et al. Obesity in patients younger than 60 years is a risk factor for covid-19 hospital admission. Clin Infect Dis. 2020;71(15):896–7. https://doi.org/ 10.1093/cid/ciaa415.
- Lippi G, Henry BM. Chronic obstructive pulmonary disease is associated with severe coronavirus disease 2019 (covid-19). Respir Med. 2020;167:105941. https://doi.org/10.1016/j.rmed.2020.105941.
- Pinti MV, Fink GK, Hathaway QA, Durr AJ, Kunovac A, Hollander JM. Mitochondrial dysfunction in type 2 diabetes mellitus: an organ-based analysis. Am J Physiol Endocrinol Metab. 2019;316(2):E268–85. https:// doi.org/10.1152/ajpendo.00314.2018.
- Kelley DE, He J, Menshikova EV, Ritov VB. Dysfunction of mitochondria in human skeletal muscle in type 2 diabetes. Diabetes. 2002;51(10):2944–50. https://doi.org/10.2337/diabetes.51.10.2944.

- 139. Jang JY, Blum A, Liu J, Finkel T. The role of mitochondria in aging. J Clin Invest. 2018;128(9):3662–70. https://doi.org/10.1172/JCl120842.
- Ritter A, Kreis NN, Louwen F, Yuan J. Obesity and covid-19: Molecular mechanisms linking both pandemics. Int J Mol Sci. 2020;21(16). https:// doi.org/10.3390/ijms21165793.
- 141. Costa FF, Rosario WR, Ribeiro Farias AC, de Souza RG, Duarte Gondim RS, Barroso WA. Metabolic syndrome and covid-19: an update on the associated comorbidities and proposed therapies. Diabetes Metab Syndr. 2020;14(5):809–14. https://doi.org/10.1016/j.dsx.2020.06.016.
- 142. Pugliese G, Vitale M, Resi V, Orsi E. Is diabetes mellitus a risk factor for coronavirus disease 19 (covid-19)? Acta Diabetol. 2020;57(11):1275–85. https://doi.org/10.1007/s00592-020-01586-6.
- 143. Sanyaolu A, Okorie C, Marinkovic A, Patidar R, Younis K, Desai P, et al. Comorbidity and its impact on patients with covid-19. SN Compr. Clin Med. 2020:1–8. https://doi.org/10.1007/s42399-020-00363-4.
- 144. Arentz M, Yim E, Klaff L, Lokhandwala S, Riedo FX, Chong M, et al. Characteristics and outcomes of 21 critically ill patients with covid-19 in washington state. JAMA. 2020;323(16):1612–4. https://doi.org/10.1001/ jama.2020.4326.
- 145. Lippi G, Salvagno GL, Pegoraro M, Militello V, Caloi C, Peretti A, et al. Assessment of immune response to sars-cov-2 with fully automated maglumi 2019-ncov igg and igm chemiluminescence immunoassays. Clin Chem Lab Med. 2020;58(7):1156–9. https://doi.org/10.1515/ cclm-2020-0473.
- 146. Yang Y, Liangjian K, Li L, Yongjian W, Zhong B, Huang X. Distinct mitochondria-mediated t cells apoptosis responses in children and adults with covid-19. J Infect Dis. 2021. https://doi.org/10.1093/infdis/jiab400.
- Liu L, Liao X, Wu H, Li Y, Zhu Y, Chen Q. Mitophagy and its contribution to metabolic and aging-associated disorders. Antioxid Redox Signal. 2020;32(12):906–27. https://doi.org/10.1089/ars.2019.8013.
- 148. Tong M, Saito T, Zhai P, Oka SI, Mizushima W, Nakamura M, et al. Mitophagy is essential for maintaining cardiac function during high fat diet-induced diabetic cardiomyopathy. Circ Res. 2019;124(9):1360–71. https://doi.org/10.1161/CIRCRESAHA.118.314607.
- 149. Zhang L, Qin Y, Chen M. Viral strategies for triggering and manipulating mitophagy. Autophagy. 2018;14(10):1665–73. https://doi.org/10.1080/15548627.2018.1466014.
- Gassen NC, Papies J, Bajaj T, Emanuel J, Dethloff F, Chua RL, et al. Sarscov-2-mediated dysregulation of metabolism and autophagy uncovers host-targeting antivirals. Nat Commun. 2021;12(1):3818. https://doi.org/ 10.1038/s41467-021-24007-w.
- 151. de Wit E, Feldmann F, Cronin J, Jordan R, Okumura A, Thomas T, et al. Prophylactic and therapeutic remdesivir (gs-5734) treatment in the rhesus macaque model of mers-cov infection. Proc Natl Acad Sci U S A. 2020;117(12):6771–6. https://doi.org/10.1073/pnas.1922083117.
- Tchesnokov EP, Feng JY, Porter DP, Gotte M. Mechanism of inhibition of ebola virus rna-dependent rna polymerase by remdesivir. Viruses. 2019;11(4). https://doi.org/10.3390/v11040326.
- Kwok M, Lee C, Li HS, Deng R, Tsoi C, Ding Q, et al. Remdesivir induces persistent mitochondrial and structural damage in human induced pluripotent stem cell derived cardiomyocytes. Cardiovasc Res. 2021. https://doi.org/10.1093/cvr/cvab311.
- Beigel JH, Tomashek KM, Dodd LE, Mehta AK, Zingman BS, Kalil AC, et al. Remdesivir for the treatment of covid-19 - preliminary report. N Engl J Med. 2020. https://doi.org/10.1056/NEJMoa2007764.
- 155. Xu Y, Barauskas O, Kim C, Babusis D, Murakami E, Kornyeyev D, et al. Off-target in vitro profiling demonstrates that remdesivir is a highly selective antiviral agent. Antimicrob Agents Chemother. 2021;65(2). https://doi.org/10.1128/AAC.02237-20.
- Akinci E, Cha M, Lin L, Yeo G, Hamilton MC, Donahue CJ, et al. Elucidation of remdesivir cytotoxicity pathways through genome-wide crisprcas9 screening and transcriptomics. bioRxiv. 2020. https://doi.org/10. 1101/2020.08.27.270819.
- 157. Falzarano D, de Wit E, Rasmussen AL, Feldmann F, Okumura A, Scott DP, et al. Treatment with interferon-alpha2b and ribavirin improves outcome in mers-cov-infected rhesus macaques. Nat Med. 2013;19(10):1313–7. https://doi.org/10.1038/nm.3362.
- Chiou HE, Liu CL, Buttrey MJ, Kuo HP, Liu HW, Kuo HT, et al. Adverse effects of ribavirin and outcome in severe acute respiratory syndrome: experience in two medical centers. Chest. 2005;128(1):263–72. https:// doi.org/10.1378/chest.128.1.263.

- Tong S, Su Y, Yu Y, Wu C, Chen J, Wang S, et al. Ribavirin therapy for severe covid-19: A retrospective cohort study. Int J Antimicrob Agents. 2020;56(3):106114. https://doi.org/10.1016/j.ijantimicag.2020.106114.
- Lafeuillade A, Hittinger G, Chadapaud S. Increased mitochondrial toxicity with ribavirin in hiv/hcv coinfection. Lancet. 2001;357(9252):280–1. https://doi.org/10.1016/S0140-6736(00)03618-7.
- Manabe T, Kambayashi D, Akatsu H, Kudo K. Favipiravir for the treatment of patients with covid-19: a systematic review and metaanalysis. BMC Infect Dis. 2021;21(1):489. https://doi.org/10.1186/ s12879-021-06164-x.
- 162. Nojomi M, Yassin Z, Keyvani H, Makiani MJ, Roham M, Laali A, et al. Effect of arbidol (umifenovir) on covid-19: A randomized controlled trial. BMC Infect Dis. 2020;20(1):954. https://doi.org/10.1186/ s12879-020-05698-w.
- Chu CM, Cheng VC, Hung IF, Wong MM, Chan KH, Chan KS, et al. Role of lopinavir/ritonavir in the treatment of sars: initial virological and clinical findings. Thorax. 2004;59(3):252–6. https://doi.org/10.1136/thorax.2003. 012658.
- Cao B, Wang Y, Wen D, Liu W, Wang J, Fan G, et al. A trial of lopinavirritonavir in adults hospitalized with severe covid-19. N Engl J Med. 2020;382(19):1787–99. https://doi.org/10.1056/NEJMoa2001282.
- 165. De Meyer S, Bojkova D, Cinatl J, Van Damme E, Buyck C, Van Loock M, et al. Lack of antiviral activity of darunavir against sars-cov-2. Int J Infect Dis. 2020;97:7–10. https://doi.org/10.1016/j.ijid.2020.05.085.
- 166. Chen J, Xia L, Liu L, Xu Q, Ling Y, Huang D, et al. Antiviral activity and safety of darunavir/cobicistat for the treatment of covid-19. Open Forum Infect Dis. 2020;7(7):ofaa241. https://doi.org/10.1093/ofid/ ofaa241.
- 167. Gao J, Tian Z, Yang X. Breakthrough: chloroquine phosphate has shown apparent efficacy in treatment of covid-19 associated pneumonia in clinical studies. Biosci Trends. 2020;14(1):72–3. https://doi.org/10.5582/ bst.2020.01047.
- Vincent MJ, Bergeron E, Benjannet S, Erickson BR, Rollin PE, Ksiazek TG, et al. Chloroquine is a potent inhibitor of sars coronavirus infection and spread. Virol J. 2005;2:69. https://doi.org/10.1186/1743-422X-2-69.
- 169. Liu J, Cao R, Xu M, Wang X, Zhang H, Hu H, et al. Hydroxychloroquine, a less toxic derivative of chloroquine, is effective in inhibiting sars-cov-2 infection in vitro. Cell Discov. 2020;6(1):16. https://doi.org/10.1038/ s41421-020-0156-0.
- 170. Hashem AM, Alghamdi BS, Algaissi AA, Alshehri FS, Bukhari A, Alfaleh MA, et al. Therapeutic use of chloroquine and hydroxychloroquine in covid-19 and other viral infections: a narrative review. Travel Med Infect Dis. 2020;35:101735. https://doi.org/10.1016/j.tmaid.2020.101735.
- Mauthe M, Orhon I, Rocchi C, Zhou X, Luhr M, Hijlkema KJ, et al. Chloroquine inhibits autophagic flux by decreasing autophagosomelysosome fusion. Autophagy. 2018;14(8):1435–55. https://doi.org/10. 1080/15548627.2018.1474314.
- Savarino A, Boelaert JR, Cassone A, Majori G, Cauda R. Effects of chloroquine on viral infections: an old drug against today's diseases? Lancet Infect Dis. 2003;3(11):722–7. https://doi.org/10.1016/s1473-3099(03) 00806-5.
- Rolain JM, Colson P, Raoult D. Recycling of chloroquine and its hydroxyl analogue to face bacterial, fungal and viral infections in the 21st century. Int J Antimicrob Agents. 2007;30(4):297–308. https://doi.org/10. 1016/j.ijantimicag.2007.05.015.
- Oscanoa TJ, Romero-Ortuno R, Carvajal A, Savarino A. A pharmacological perspective of chloroquine in sars-cov-2 infection: an old drug for the fight against a new coronavirus? Int J Antimicrob Agents. 2020;56(3):106078. https://doi.org/10.1016/j.ijantimicag.2020.106078.
- Sturrock BR, Chevassut TJ. Chloroquine and covid-19 a potential game changer? Clin Med (Lond). 2020. https://doi.org/10.7861/clinmed. 2020-0129.
- 176. Axfors C, Schmitt AM, Janiaud P, Van't Hooft J, Abd-Elsalam S, Abdo EF, et al. Author correction: Mortality outcomes with hydroxychloroquine and chloroquine in covid-19 from an international collaborative metaanalysis of randomized trials. Nat Commun. 2021;12(1):3001. https://doi. org/10.1038/s41467-021-23559-1.
- 177. Deepalakshmi PD, Parasakthy K, Shanthi S, Devaraj NS. Effect of chloroquine on rat liver mitochondria. Indian J Exp Biol. 1994;32(11):797–9.
- 178. Redmann M, Benavides GA, Berryhill TF, Wani WY, Ouyang X, Johnson MS, et al. Inhibition of autophagy with bafilomycin and chloroquine

decreases mitochondrial quality and bioenergetic function in primary neurons. Redox Biol. 2017;11:73–81. https://doi.org/10.1016/j.redox. 2016.11.004.

- Zhou Y, Hou Y, Shen J, Huang Y, Martin W, Cheng F. Network-based drug repurposing for novel coronavirus 2019-ncov/sars-cov-2. Cell Discov. 2020;6:14. https://doi.org/10.1038/s41421-020-0153-3.
- Martin WR, Cheng F. Repurposing of fda-approved toremifene to treat covid-19 by blocking the spike glycoprotein and nsp14 of sars-cov-2. J Proteome Res. 2020;19(11):4670–7. https://doi.org/10.1021/acs.jprot eome.0c00397.
- Dyall J, Coleman CM, Hart BJ, Venkataraman T, Holbrook MR, Kindrachuk J, et al. Repurposing of clinically developed drugs for treatment of middle east respiratory syndrome coronavirus infection. Antimicrob Agents Chemother. 2014;58(8):4885–93. https://doi.org/10.1128/AAC.03036-14.
- 182. Kandimalla R, John A, Abburi C, Vallamkondu J, Reddy PH. Current status of multiple drug molecules, and vaccines: an update in sars-cov-2 therapeutics. Mol Neurobiol. 2020;57(10):4106–16. https://doi.org/10. 1007/s12035-020-02022-0.
- Group RC, Horby P, Lim WS, Emberson JR, Mafham M, Bell JL, et al. Dexamethasone in hospitalized patients with covid-19. N Engl J Med. 2021;384(8):693–704. https://doi.org/10.1056/NEJMoa2021436.
- Pinzon MA, Ortiz S, Holguin H, Betancur JF, Cardona Arango D, Laniado H, et al. Dexamethasone vs methylprednisolone high dose for covid-19 pneumonia. PLoS One. 2021;16(5):e0252057. https://doi.org/10.1371/ journal.pone.0252057.
- 185. Tomazini BM, Maia IS, Cavalcanti AB, Berwanger O, Rosa RG, Veiga VC, et al. Effect of dexamethasone on days alive and ventilator-free in patients with moderate or severe acute respiratory distress syndrome and covid-19: The codex randomized clinical trial. JAMA. 2020;324(13):1307–16. https://doi.org/10.1001/jama.2020.17021.
- Sionov RV, Kfir S, Zafrir E, Cohen O, Zilberman Y, Yefenof E. Glucocorticoid-induced apoptosis revisited: A novel role for glucocorticoid receptor translocation to the mitochondria. Cell Cycle. 2006;5(10):1017–26. https://doi.org/10.4161/cc.5.10.2738.
- 187. Farnoosh G, Akbariqomi M, Badri T, Bagheri M, Izadi M, Saeedi-Boroujeni A, et al. Efficacy of a low dose of melatonin as an adjunctive therapy in hospitalized patients with covid-19: a randomized, doubleblind clinical trial. Arch Med Res. 2021. https://doi.org/10.1016/j. arcmed.2021.06.006.
- Zhang R, Wang X, Ni L, Di X, Ma B, Niu S, et al. Covid-19: melatonin as a potential adjuvant treatment. Life Sci. 2020;250:117583. https://doi.org/ 10.1016/j.lfs.2020.117583.
- Tan DX, Manchester LC, Qin L, Reiter RJ. Melatonin: a mitochondrial targeting molecule involving mitochondrial protection and dynamics. Int J Mol Sci. 2016;17(12). https://doi.org/10.3390/ijms17122124.
- Bromme HJ, Morke W, Peschke D, Ebelt H, Peschke D. Scavenging effect of melatonin on hydroxyl radicals generated by alloxan. J Pineal Res. 2000;29(4):201–8. https://doi.org/10.1034/j.1600-0633.2002.290402.x.
- 191. Absi E, Ayala A, Machado A, Parrado J. Protective effect of melatonin against the 1-methyl-4-phenylpyridinium-induced inhibition of complex i of the mitochondrial respiratory chain. J Pineal Res. 2000;29(1):40– 7. https://doi.org/10.1034/j.1600-079x.2000.290106.x.
- Jou MJ, Peng TI, Yu PZ, Jou SB, Reiter RJ, Chen JY, et al. Melatonin protects against common deletion of mitochondrial DNA-augmented mitochondrial oxidative stress and apoptosis. J Pineal Res. 2007;43(4):389–403. https://doi.org/10.1111/j.1600-079X.2007.00490.x.
- 193. Ding K, Wang H, Xu J, Li T, Zhang L, Ding Y, et al. Melatonin stimulates antioxidant enzymes and reduces oxidative stress in experimental traumatic brain injury: The nrf2-are signaling pathway as a potential mechanism. Free Radic Biol Med. 2014;73:1–11. https://doi.org/10. 1016/j.freeradbiomed.2014.04.031.
- Kilanczyk E, Bryszewska M. The effect of melatonin on antioxidant enzymes in human diabetic skin fibroblasts. Cell Mol Biol Lett. 2003;8(2):333–6.
- 195. Hofmann-Winkler H, Moerer O, Alt-Epping S, Brauer A, Buttner B, Muller M, et al. Camostat mesylate may reduce severity of coronavirus disease 2019 sepsis: a first observation. Crit Care Explor. 2020;2(11):e0284. https://doi.org/10.1097/CCE.00000000000284.
- 196. Hoffmann M, Hofmann-Winkler H, Smith JC, Kruger N, Arora P, Sorensen LK, et al. Camostat mesylate inhibits sars-cov-2 activation by tmprss2related proteases and its metabolite gbpa exerts antiviral activity.

EBioMedicine. 2021;65:103255. https://doi.org/10.1016/j.ebiom.2021. 103255.

- 197. Li K, Meyerholz DK, Bartlett JA, McCray PB Jr. The tmprss2 inhibitor nafamostat reduces sars-cov-2 pulmonary infection in mouse models of covid-19. mBio. 2021;12(4):e0097021. https://doi.org/10.1128/mBio. 00970-21.
- Rosas IO, Brau N, Waters M, Go RC, Hunter BD, Bhagani S, et al. Tocilizumab in hospitalized patients with severe covid-19 pneumonia. N Engl J Med. 2021;384(16):1503–16. https://doi.org/10.1056/NEJMo a2028700.
- 199. Rosas IO, Diaz G, Gottlieb RL, Lobo SM, Robinson P, Hunter BD, et al. Tocilizumab and remdesivir in hospitalized patients with severe covid-19 pneumonia: a randomized clinical trial. Intensive Care Med. 2021;47(11):1258–70. https://doi.org/10.1007/s00134-021-06507-x.
- Veiga VC, Prats J, Farias DLC, Rosa RG, Dourado LK, Zampieri FG, et al. Effect of tocilizumab on clinical outcomes at 15 days in patients with

severe or critical coronavirus disease 2019: randomised controlled trial. BMJ. 2021;372:n84. https://doi.org/10.1136/bmj.n84.

 Lescure FX, Honda H, Fowler RA, Lazar JS, Shi G, Wung P, et al. Sarilumab in patients admitted to hospital with severe or critical covid-19: a randomised, double-blind, placebo-controlled, phase 3 trial. Lancet Respir Med. 2021;9(5):522–32. https://doi.org/10.1016/S2213-2600(21) 00099-0.

## **Publisher's Note**

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.