

# 

**Citation:** Teeuw WJ, Laine ML, Bizzarro S, Loos BG (2015) A Lead *ANRIL* Polymorphism Is Associated with Elevated CRP Levels in Periodontitis: A Pilot Case-Control Study. PLoS ONE 10(9): e0137335. doi:10.1371/journal.pone.0137335

Editor: Kimon Divaris, UNC School of Dentistry, University of North Carolina-Chapel Hill, UNITED STATES

Received: June 24, 2015

Accepted: August 15, 2015

Published: September 8, 2015

**Copyright:** © 2015 Teeuw et al. This is an open access article distributed under the terms of the <u>Creative Commons Attribution License</u>, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

**Data Availability Statement:** All relevant data are within the paper and its Supporting Information files.

Funding: The Department of Periodontology at ACTA, the Netherlands is supported in part by a grant of the University of Amsterdam for its participation in the focal point "Oral Infections and Inflammation". The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript. For the rest of the study, the authors did not receive specific funding. There was no additional external funding received for this study. **RESEARCH ARTICLE** 

# A Lead *ANRIL* Polymorphism Is Associated with Elevated CRP Levels in Periodontitis: A Pilot Case-Control Study

#### Wijnand J. Teeuw\*, Marja L. Laine, Sergio Bizzarro, Bruno G. Loos

Department of Periodontology, Academic Centre for Dentistry Amsterdam (ACTA), University of Amsterdam and VU University Amsterdam, Amsterdam, the Netherlands

\* W.Teeuw@acta.nl

# Abstract

Elevated high sensitive C-reactive protein (hsCRP) is a marker for systemic inflammation and a risk marker for atherosclerotic cardiovascular disease (ACVD), and has also been associated with periodontitis. Inter-individual variation for hsCRP in periodontitis has been shown. ANRIL is the strongest genetic susceptibility locus for both periodontitis and ACVD, and it is speculated that genetic variation in ANRIL may modulate inflammatory processes. Therefore, we explored the possible association between hsCRP plasma levels and a leading ANRIL single nucleotide polymorphism (SNP) in periodontitis patients and controls. 171 healthy subjects with North European descent (115 periodontitis and 56 controls) were included in this case-control study. hsCRP levels were determined and subjects were genotyped for the leading ANRIL SNP rs1333048. In a multivariate analysis, periodontitis, female gender, increasing BMI and homozygosity for the major allele (AA-genotype) of rs1333048 were significantly associated with elevated hsCRP plasma levels (p = 0.012, p = 0.004, p =0.007 and p = 0.003, respectively). Periodontitis patients with rs1333048 AA-genotype showed higher levels of hsCRP than those carrying the minor C allele (median: 4.5 mg/L vs. 1.6 mg/L, p<sub>adiusted</sub> = 0.007). This study is the first to show that, in addition to gender and BMI, also a leading SNP in ANRIL is explanatory for inter-individual variation in hsCRP levels in periodontitis patients of North European descent.

# Introduction

Periodontitis is a complex, chronic inflammatory disease, resulting in loss of connective tissue and alveolar bone support of the teeth [1]. It is the major cause of tooth loss in adults above 40 years and affects human populations worldwide at prevalence rates up to 10–20% for the most severe forms [2,3]. Periodontitis is associated with moderately increased concentration of high sensitive C-reactive protein (hsCRP) in the blood circulation [4]. These elevated levels of hsCRP in periodontitis are 'dose dependent' with higher levels in severe forms of periodontitis, compared to moderate forms and periodontal health and range between 1–4 mg/L [5].



**Competing Interests:** The authors have declared that no competing interests exist.

Periodontal therapy reduces periodontal inflammation and recent meta-analyses are highly suggestive that this reduction results in lower plasma levels of hsCRP (overall: -0.5 mg/L) [6].

Although on average periodontitis patients show elevated hsCRP, variations in levels have been shown both between patients and between populations [4,6]. Also the reduction of hsCRP levels after periodontal treatment varies substantially between study groups [6], suggesting that additionally several other factors may influence the systemic inflammatory response in patients suffering from periodontitis. Interestingly, hsCRP is also widely accepted as key marker of atherosclerosis and is strongly associated with increased risk for atherosclerotic cardiovascular disease (ACVD); levels of 1-3 mg/L and >3 mg/L are considered to give medium and high risk for ACVD, respectively [7,8]. In this context, also increasing BMI and female gender have been suggested as factors associated with chronically elevated hsCRP [9,10].

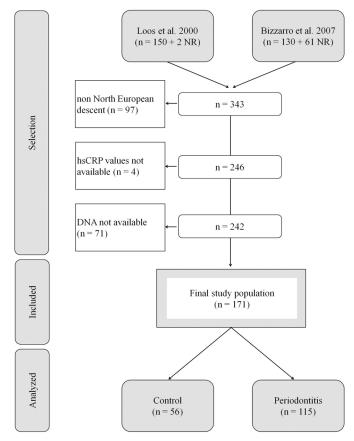
In general, inflammatory responses are modulated by specific polymorphisms in inflammatory genes [11]. We speculate that polymorphisms in the ANRIL locus on chromosome 9p21 may be involved. ANRIL is a genetic risk factor for several conditions with inflammatory components in Caucasians, and is the strongest genetic susceptibility locus for periodontitis [12,13] and several types of ACVD, like coronary artery disease (CAD), myocardial infarction, abdominal aortic aneurysm and intracranial aneurysm [14-16]. It has been shown that the diseaseassociated single nucleotide polymorphisms (SNPs) of chromosome 9p21 have been associated with the expression of ANRIL [17]. In particular, the CAD-associated polymorphisms within the core risk haplotype region have been shown to regulate ANRIL expression in vitro [18] and also in vivo [19]. But so far, the function of ANRIL in relation to periodontitis and ACVD is not fully understood. At least one pathway is known, ANRIL is regulated by STAT1 signaling [18], a pathway that mediates response to inflammation upon stimulation of the pro-inflammatory cytokine interferon- $\gamma$ . Furthermore, ANRIL transcription was up-regulated in gingival tissues by bacterial infection [13]. ANRIL may therefore be an important regulator of inflammatory and immune reactions, such as in the pathophysiology of periodontitis, ACVD, diabetes mellitus and cancer [19]. If so, variation in plasma hsCRP (as endpoint and reporter molecule of a variety of inflammatory pathways) may reflect inter-individual heterogeneity of ANRIL activity based on genetic variation. Therefore, we hypothesize that the observed variation in hsCRP levels in periodontitis patients might be associated with genetic variation in the ANRIL gene in addition to other factors and may explain inter-individual differences in hsCRP levels among periodontitis patients.

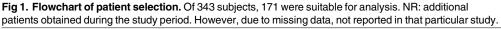
The aim of the present explorative pilot case-control study was to investigate the possible association between levels of hsCRP and the leading ACVD- and periodontitis-associated gene polymorphism (*ANRIL* rs1333048, [12,13]) in Caucasian periodontitis patients and controls.

# **Materials and Methods**

#### Study population

A total of 171 subjects from North European descent, including 115 patients with periodontitis and 56 controls, were suitable to participate in this pilot study. These individuals were retrieved from a total of 343 individuals from two previous studies [20,21] (Fig 1). For all study subjects, age, gender, body mass index (BMI) and current smoking were extracted out of the databases [20,21]. Briefly, referred periodontitis patients were recruited during their first visit at the Department of Periodontology of the Academic Centre Dentistry Amsterdam (ACTA). Clinical measurements were performed at six sites per tooth and a full radiographic status was available to analyze interproximal alveolar bone levels. Patients were classified as suffering from periodontitis using the CDC-AAP case definition for moderate to severe periodontitis [22];





doi:10.1371/journal.pone.0137335.g001

patients showed at least 2 interproximal sites with attachment loss (AL)  $\geq$ 4 mm on different teeth in conjunction with alveolar bone loss on the basis of peri-apical radiographs. All patients were untreated and showed generalized bleeding on probing. Controls were selected among subjects registered for restorative dental procedures or who visited ACTA for regular dental check-ups. Control subjects were included if they were not missing more than one tooth per quadrant (3rd molar excluded) and no probing pocket depth (PPD)  $\geq$ 3 mm. These subjects showed on dental bitewing radiographs  $\leq$ 1-year-old a distance between the cemento-enamel junction and the alveolar bone crest of  $\leq$ 3 mm on all teeth.

Exclusion criteria for the current study subjects were: (i) the presence of a systemic disease, (ii) recent history of any acute or chronic infection, periodontitis not included, (iii) systemic antibiotic treatment within the last 3 months, (iv) the use of any medication (including sporadic NSAID's), (v) pregnancy or lactation, vi) non North European descent, and vii) no DNA available (Fig 1).

The Medical Ethical Committee of the Academic Medical Center, Amsterdam, approved this study and all participants gave their written informed consent to participate.

#### Analysis of hsCRP plasma levels

Non-fasting, venous blood was collected from each subject in EDTA containing tubes between 8:30 and 11:30am. Plasma levels of hsCRP were determined using a high sensitivity

nephelometric method on the BN ProSpec analyzer (Dade Behring, Marburg, Germany). The detection limit was 0.3 mg/L, linearity was from 0.3 to 230 mg/L, and the coefficient of variation was <3% at a concentration of 2 mg/L. Subjects with hsCRP values <0.3 mg/L were regarded sero-negative [20,21].

# Analysis of gene polymorphisms

For all participants, the lead and validated periodontitis- and ACVD-associated intronic SNP rs1333048 in the core risk region of the ANRIL gene [12,13,23] was investigated. This SNP is also in strong linkage disequilibrium with other representative SNPs that are associated with both periodontitis and ACVD [24]. The polymorphism was assayed at the Institute for Clinical Molecular Biology of the Christian Albrechts University, Kiel, Germany, with TaqMan (Applied Biosystems [ABI], Foster City, CA, USA) on an automated platform. In brief, genomic DNA was arrayed on 96-well or 384-well plates. Samples were amplified with ABI 9700 PCR machines and fluorescence was measured with ABI 7700 fluorometer. The assay included two locus-specific PCR primers that flank the SNP of interest, and two allele-specific oligonucleotide probes (TTCATGCTATTTTGAGGAGATGTTT[ A/C] AATGTCGAATTATTGAAATATTTAT (Context Sequence [VIC/FAM]). For the polymorphism assayed, a PCR-protocol was used as follows: a) activation: 95°C, 10 min, 1 cycle; b) denaturation (95°C, 15s), annealing, elongation, nucleolytic cleavage of hybridized probes (60°C, 1 min), 45 cycles; c) storage: 4°C. For the endpoint measurement the ABIPrism 7900 HT Sequence Detection System was used. The assay had a call rate of 100%. The A allele of rs1333048 has been determined as the major allele, while the C allele is the minor allele [13,15]; subjects were genotyped as AA, AC or CC.

# Data analyses

Data analyses were performed with the SPSS 18.0 package (SPSS Inc., Chicago, IL, USA). Means, standard deviations, medians, interquartile ranges (IQR) and frequency distributions were calculated. Because of the non-normal distribution, hsCRP plasma levels were log transformed for all calculations. The general characteristics of the study population, hsCRP levels, genotype and allelic distribution within the study population were compared with parametric and non-parametric tests (ANOVA and chi-squared test). In addition, boxplots were generated. Where applicable, analyses were corrected for multiple testing (Bonferroni). Deviations from Hardy-Weinberg equilibrium were tested in the control group by using a chi-squared test and a type I error level of 0.05. Odds Ratios (OR) and confidence intervals (CI), adjusted for age, gender, smoking habits and BMI, were calculated. Logistic regression analysis was performed. Significance was assessed by a Wald test and by a likelihood-ratio test. Univariate analysis was performed (ANCOVA), using hsCRP levels as the outcome parameter and periodontitis as a fixed factor. Age, gender, smoking habits, BMI, and carriage of the minor allele (AC or CC) of rs1333048 were included as covariates. For all analyses, the significance level was set to p < 0.05.

# Results

Demographic, clinical and genotypic characteristics of the study population are presented in Table 1. The mean ages for the control and periodontitis group were 43.9, 45.5 years, respectively. Among periodontitis patients, 53.9% were smokers, while this was 30% among controls (p = 0.004). The gender distribution was not significantly different among the control and periodontitis groups (males: 46.4% and 35.7%, respectively). Periodontitis patients had significantly less teeth than controls (p < 0.001), and the periodontitis group presented on average 6.7 teeth with  $\geq$ 50% alveolar bone loss. Controls had no alveolar bone loss.



#### Table 1. General characteristics of the study population.

	Control (n = 56)	Periodontitis (n = 115)	P-value
Background characteristics			
Age	43.9 ± 13.2	45.5 ± 9.9	0.430
Gender (males)	26 (46.4%)	41 (35.7%)	0.175
BMI (kg/m²)	24.9 ± 3.5	24.5 ± 3.6	0.567
# teeth present	27.7 ± 1.9	25.8 ± 3.2	<0.001
# teeth with $\geq$ 50% alveolar bone loss	-	6.7 ± 5.7	NA
# patients with $\geq$ 7 teeth with $\geq$ 50% alveolar bone loss	-	51 (44.3%)	NA
# Smokers	17 (30.4%)	62 (53.9%)	0.004
Systemic inflammatory marker			
hsC-reactive protein (mg/L), median (IQR)	1.3 (0.6–2.5)	1.8 (0.8–4.5)	0.031
ANRIL rs1333048 (A>C)			
Genotypes frequency			
A/A	20 (35.7%)	23 (20.0%)	
A/C	26 (46.4%)	60 (52.2%)	0.065
C/C	10 (17.9%)	32 (27.8%)	
MAF (C-allele)	0.41	0.54 <sup>a</sup>	0.026
C allele carrier (%)	36 (64.3%)	92 (80.0%)	0.026

<sup>a</sup> Carriage of the C-allele was significantly associated with periodontitis: adjusted OR = 2.56 (95%CI: 1.20–5.50), p = 0.015

doi:10.1371/journal.pone.0137335.t001

hsCRP was significantly different among the two groups (p = 0.031, <u>Table 1</u>). The median levels of hsCRP levels for controls and periodontitis were 1.3 mg/L and 1.8 mg/L, respectively.

Genotyping results showed no significant deviation from the Hardy-Weinberg equilibrium in the control group. There was no significant difference in the genotype frequencies of *ANRIL* rs1333048 between periodontitis and controls (p = 0.065, <u>Table 1</u>). However, minor allele frequencies and carriage of the C-allele of rs1333048 (genotypes AC and CC combined) was significantly associated with periodontitis (adjusted OR = 2.56 [95% CI: 1.20–5.50], p = 0.015).

#### Univariate analysis of hsCRP levels

Univariate analysis was performed to explore the relation of several covariates with hsCRP levels (<u>Table 2</u>). Including the potential confounders, periodontitis was significantly associated

#### Table 2. Univariate analysis for hsCRP levels.

-			
	<i>F</i> -ratio	<i>p</i> -value	
Intercept	6.83	0.010	
Periodontitis <sup>a</sup>	6.50	0.012	
Age	2.82	0.095	
Gender (female)	8.63	0.004	
Smoking	3.03	0.084	
BMI	7.58	0.007	
ANRIL rs1333048 (AA-genotype)	9.00	0.003	

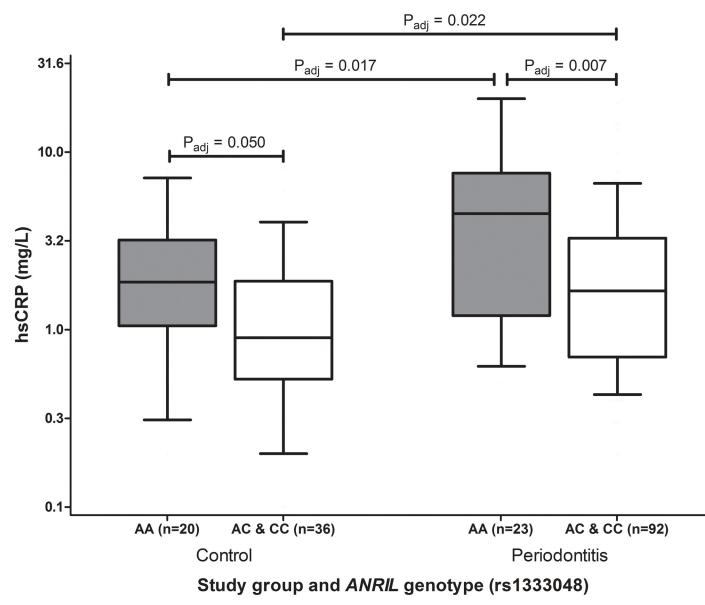
<sup>a</sup> hsCRP means and 95% Confidence Interval (CI) adjusted for all covariates: Control: 1.19 mg/L, CI: 0.90– 1.57; Periodontitis: 1.86 mg/L, CI: 1.11–2.22)

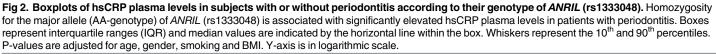
doi:10.1371/journal.pone.0137335.t002

MAF = Minor Allele Frequency; NA = Not applicable

with elevated hsCRP plasma levels (p = 0.012). After adjustment for the included covariates, the adjusted mean hsCRP levels for the control and periodontitis group were 1.19 mg/L (CI: 0.90–1.57), and 1.86 mg/L (CI: 1.11–2.22), p = 0.012, respectively. In addition, the covariates gender, BMI and genotype were significant: females (p = 0.004), individuals with increased BMI (p = 0.007) and subjects homozygous for the major allele (AA-genotype) of rs1333048 showed increased plasma concentrations of hsCRP (p = 0.003).

The association of the *ANRIL* genotype with hsCRP levels is a new finding. Therefore, we explored the hsCRP levels in relation to periodontitis, subgrouped by carriership of the minor allele. Fig 2 shows that subjects with periodontitis and homozygous for the major allele





doi:10.1371/journal.pone.0137335.g002

(AA-genotype) of rs1333048 have significantly elevated hsCRP levels (4.5 mg/L [median]; IQR: 1.2–7.6 mg/L) compared to subjects with periodontitis and carriage of the minor allele (AC and CC genotypes) (1.6 mg/L [median; IQR: 0.7–3.3 mg/L; p = 0.007] and healthy controls homozygous for the major allele (1.9 mg/L [median]; IQR: 1.1–3.0 mg/L; p = 0.017). A trend for intragroup differences in hsCRP levels between subjects of the control group homozygous for the major allele and subjects carrying the minor allele was also observed (p = 0.050). Notably, carriage of the minor allele of rs1333048 is associated with relative low hsCRP levels, nevertheless the periodontitis patients with genotypes AC or CC showed higher hsCRP levels than the corresponding individuals in the control group (periodontitis: 1.6 mg/L [median]; IQR: 0.7–3.2 mg/L; control: 0.9 mg/L [median]; IQR: 0.6–1.7 mg/L; p = 0.022).

#### Discussion

The present pilot study aimed to investigate the possible association between hsCRP plasma levels and potential confounding factors, including the leading polymorphism in *ANRIL*, in periodontitis patients and controls. *ANRIL* is an important genetic locus as it is associated with several chronic systemic conditions, including ACVD and periodontitis. The main finding of this study is that not only periodontitis, BMI and female gender are related with elevated CRP: also the AA-genotype of *ANRIL* (rs1333048) is associated with significantly elevated hsCRP plasma levels in patients with periodontitis. While periodontitis, female gender and elevated BMI have been associated with increased hsCRP in the literature [4,9,10], this is the first study that shows that a SNP in *ANRIL* is also associated with hsCRP levels in periodontitis patients. Interestingly, a trend was also seen for controls, but this did not reach statistical significance (p = 0.050). The observation suggests that among other and currently unknown factors, the *ANRIL* locus may be involved in 'generic' inflammatory processes of immune-mediated diseases with hsCRP as final biomarker at one or several pathways. The challenge is to turn the increasing knowledge of pleiotropic pathways to clinical relevance [25].

We recognize that periodontitis is a complex disease with multiple causal factors playing a role simultaneously [26]. These can be grouped in genetic, bacteria and lifestyle related factors, including diet, systemic diseases, notably diabetes, and other factors as of yet unknown. Currently, more genetic loci, in addition to *ANRIL*, shared between ACVD and periodontitis have been identified: *PLASMINOGEN* and a conserved noncoding element within *CAMTA1* upstream of *VAMP3* [27,28]. Recent experimental and epidemiological studies suggested that *ANRIL*, *VAMP3* and *PLASMINOGEN* are involved in several regulatory networks that relate to glucose and fatty acid metabolism, host-microbiome interactions and TGF- $\beta$  signaling, providing evidence for a mechanistic link between ACVD, periodontitis, obesity and inflammation [27–30]. Any disturbances in these pathways by genetic variants may be a common pathogenic trait of ACVD and periodontitis.

Previous studies showed that carriership of the minor allele of rs1333048 is associated with ACVD and periodontitis. In the present study we corroborate this; the C-allele was more frequent in periodontitis patients than controls. However, the clinical effects of this confirmed association have never been reported, although functionality of the SNP is suggested *in vitro/ex vivo* experiments [27]. We show here that, in the inflammatory disease periodontitis, C-carriage is associated with lower plasma levels of hsCRP compared with those that had the AA-genotype. The role of CRP in periodontitis is not yet established. CRP is regarded as a sensitive, non-specific marker of inflammation. However, although chronic elevated hsCRP levels are considered as a prognostic risk marker for ACVD [7,8], in acute infection or inflammation the role of CRP might be protective [31]. In that sense, genetic variations causing low CRP levels might play an etiologic role. It is known that CRP can inhibit the alternative complement

pathway by binding factor H [<u>32–35</u>]. This results in restricted complement activation favoring opsonization and not in the formation of the membrane attack complex, subsequently leading to a less strong inflammatory response and therefore less inflammation derived tissue damage [<u>31</u>]. Nevertheless, we have to realize that for certain cases, even with high levels of CRP, other factors might play a more dominant role in the onset and/or progression of periodontal disease.

For example smoking is known to be a strong etiologic factor for periodontitis. Interestingly, it was in the current study not related to elevated hsCRP plasma levels in periodontitis patients and controls. A possible explanation could be that the effect of smoking depends on the number of cigarettes consumed per day (heavy or light smoking) [36]. Furthermore, the effect of smoking on systemic inflammation is not straight forward and the results of several studies are contradictory [37,38]. It is suggested, that cigarette smoking may affect inflammation via other pathways than elevated hsCRP levels; an association between smoking status and the presence of advanced atherosclerotic lesions but not with hsCRP plasma levels has been shown [39].

In this pilot study we show for the first time that, in addition to gender and BMI, a leading SNP in *ANRIL* is associated with variations in hsCRP plasma levels in periodontitis patients from North European descent. Replication studies in other Caucasian and non-Caucasian periodontitis patients are needed as well as biochemical studies are necessary to explain the role of *ANRIL* as pleiotrophic gene affecting inflammatory pathways of different immune-mediated diseases [25], like periodontitis and ACVD.

### Supporting Information

**S1 Table. STROBE Checklist for Case-Control study.** (DOC)

## Acknowledgments

The authors thank Dr. A.S. Schaefer from the Institute for Clinical Molecular Biology of the Christian Albrechts University, Kiel, Germany, for the performance of the genotyping.

## **Author Contributions**

Conceived and designed the experiments: WT ML SB BL. Performed the experiments: WT ML SB. Analyzed the data: WT ML. Contributed reagents/materials/analysis tools: WT ML SB. Wrote the paper: WT ML SB BL.

#### References

- 1. Pihlstrom BL, Michalowicz BS, Johnson NW. Periodontal diseases. Lancet 2005; 366: 1809–1820. PMID: <u>16298220</u>
- Eke PI, Dye BA, Wei L, Thornton-Evans GO, Genco RJ, Cdc Periodontal Disease Surveillance workgroup: James Beck GDRP. Prevalence of periodontitis in adults in the United States: 2009 and 2010. J Dent Res 2012; 91: 914–920. PMID: <u>22935673</u>
- Hugoson A, Sjodin B, Norderyd O. Trends over 30 years, 1973–2003, in the prevalence and severity of periodontal disease. J Clin Periodontol 2008; 35: 405–414. doi: <u>10.1111/j.1600-051X.2008.01225.x</u> PMID: <u>18433384</u>
- Paraskevas S, Huizinga JD, Loos BG. A systematic review and meta-analyses on C-reactive protein in relation to periodontitis. J Clin Periodontol 2008; 35: 277–290. doi: <u>10.1111/j.1600-051X.2007.01173.x</u> PMID: <u>18294231</u>
- Loos BG. Systemic markers of inflammation in periodontitis. J Periodontol 2005; 76: 2106–2115. PMID: 16277583

- Teeuw WJ, Slot DE, Susanto H, Gerdes VE, Abbas F, D'Aiuto F, et al. Treatment of periodontitis improves the atherosclerotic profile: a systematic review and meta-analysis. J Clin Periodontol 2014; 41: 70–79. doi: <u>10.1111/jcpe.12171</u> PMID: <u>24111886</u>
- Geluk CA, Post WJ, Hillege HL, Tio RA, Tijssen JG, van Dijk RB, et al. C-reactive protein and angiographic characteristics of stable and unstable coronary artery disease: data from the prospective PRE-VEND cohort. Atherosclerosis 2008; 196: 372–382. PMID: 17157301
- Ridker PM, MacFadyen J, Libby P, Glynn RJ. Relation of baseline high-sensitivity C-reactive protein level to cardiovascular outcomes with rosuvastatin in the Justification for Use of statins in Prevention: an Intervention Trial Evaluating Rosuvastatin (JUPITER). Am J Cardiol 2010; 106: 204–209. doi: <u>10.</u> <u>1016/j.amjcard.2010.03.018</u> PMID: <u>20599004</u>
- Pradeep AR, Priyanka N, Prasad MV, Kalra N, Kumari M. Association of progranulin and high sensitivity CRP concentrations in gingival crevicular fluid and serum in chronic periodontitis subjects with and without obesity. Dis Markers 2012; 33: 207–213. doi: 10.3233/DMA-2012-0926 PMID: 22960346
- Woloshin S, Schwartz LM. Distribution of C-reactive protein values in the United States. N Engl J Med 2005; 352: 1611–1613.
- Incalcaterra E, Accardi G, Balistreri CR, Caimi G, Candore G, Caruso M, et al. Pro-inflammatory genetic markers of atherosclerosis. Curr Atheroscler Rep 2013; 15: 329. doi: <u>10.1007/s11883-013-0329-5</u> PMID: 23591672
- Schaefer AS, Bochenek G, Manke T, Nothnagel M, Graetz C, Thien A, et al. Validation of reported genetic risk factors for periodontitis in a large-scale replication study. J Clin Periodontol 2013; 40: 563– 572. doi: 10.1111/jcpe.12092 PMID: 23587006
- Schaefer AS, Richter GM, Dommisch H, Reinartz M, Nothnagel M, Noack B, et al. CDKN2BAS is associated with periodontitis in different European populations and is activated by bacterial infection. J Med Genet 2011; 48: 38–47. doi: 10.1136/jmg.2010.078998 PMID: 20978019
- Deloukas P, Kanoni S, Willenborg C, Farrall M, Assimes TL, Thompson JR, et al. Large-scale association analysis identifies new risk loci for coronary artery disease. Nat Genet 2013; 45: 25–33. doi: <u>10.</u> 1038/ng.2480 PMID: 23202125
- Schunkert H, Gotz A, Braund P, McGinnis R, Tregouet DA, Mangino M, et al. Repeated replication and a prospective meta-analysis of the association between chromosome 9p21.3 and coronary artery disease. Circulation 2008; 117: 1675–1684. doi: <u>10.1161/CIRCULATIONAHA.107.730614</u> PMID: 18362232
- Helgadottir A, Thorleifsson G, Magnusson KP, Gretarsdottir S, Steinthorsdottir V, Manolescu A, et al. The same sequence variant on 9p21 associates with myocardial infarction, abdominal aortic aneurysm and intracranial aneurysm. Nat Genet 2008; 40: 217–224. doi: 10.1038/ng.72 PMID: 18176561
- 17. Congrains A, Kamide K, Oguro R, Yasuda O, Miyata K, Yamamoto E, et al. Genetic variants at the 9p21 locus contribute to atherosclerosis through modulation of ANRIL and CDKN2A/B. Atherosclerosis 2012; 220: 449–455. doi: 10.1016/j.atherosclerosis.2011.11.017 PMID: 22178423
- Harismendy O, Notani D, Song X, Rahim NG, Tanasa B, Heintzman N, et al. 9p21 DNA variants associated with coronary artery disease impair interferon-gamma signalling response. Nature 2011; 470: 264–268. doi: 10.1038/nature09753 PMID: 21307941
- Cunnington MS, Santibanez Koref M, Mayosi BM, Burn J, Keavney B. Chromosome 9p21 SNPs Associated with Multiple Disease Phenotypes Correlate with ANRIL Expression. PLoS Genet 2010; 6: e1000899. doi: 10.1371/journal.pgen.1000899 PMID: 20386740
- Bizzarro S, van der Velden U, ten Heggeler JM, Leivadaros E, Hoek FJ, Gerdes VE, et al. Periodontitis is characterized by elevated PAI-1 activity. J Clin Periodontol 2007; 34: 574–580. PMID: <u>17535288</u>
- Loos BG, Craandijk J, Hoek FJ, Wertheim-van Dillen PM, van der Velden U. Elevation of systemic markers related to cardiovascular diseases in the peripheral blood of periodontitis patients. J Periodontol 2000; 71: 1528–1534. PMID: <u>11063384</u>
- 22. Eke PI, Page RC, Wei L, Thornton-Evans G, Genco RJ. Update of the case definitions for populationbased surveillance of periodontitis. J Periodontol 2012; 83: 1449–1454. doi: <u>10.1902/jop.2012.110664</u> PMID: <u>22420873</u>
- Ernst FD, Uhr K, Teumer A, Fanghanel J, Schulz S, Noack B, et al. Replication of the association of chromosomal region 9p21.3 with generalized aggressive periodontitis (gAgP) using an independent case-control cohort. BMC Med Genet 2010; 11: 119. doi: 10.1186/1471-2350-11-119 PMID: 20696043
- Schaefer AS, Richter GM, Groessner-Schreiber B, Noack B, Nothnagel M, El Mokhtari NE, et al. Identification of a shared genetic susceptibility locus for coronary heart disease and periodontitis. PLoS Genet 2009; 5: e1000378. doi: <u>10.1371/journal.pgen.1000378</u> PMID: <u>19214202</u>

- Parkes M, Cortes A, van Heel DA, Brown MA. Genetic insights into common pathways and complex relationships among immune-mediated diseases. Nat Rev Genet 2013; 14: 661–673. doi: <u>10.1038/</u> <u>nrg3502</u> PMID: <u>23917628</u>
- Lopez R, Hujoel P, Belibasakis GN. On putative periodontal pathogens: an epidemiological perspective. Virulence 2015; 6: 249–257. doi: 10.1080/21505594.2015.1014266 PMID: 25874553
- Bochenek G, Hasler R, El Mokhtari NE, Konig IR, Loos BG, Jepsen S, et al. The large non-coding RNA ANRIL, which is associated with atherosclerosis, periodontitis and several forms of cancer, regulates ADIPOR1, VAMP3 and C11ORF10. Hum Mol Genet 2013; 22: 4516–4527. doi: <u>10.1093/hmg/ddt299</u> PMID: <u>23813974</u>
- Schaefer AS, Bochenek G, Jochens A, Ellinghaus D, Dommisch H, Guzeldemir-Akcakanat E, et al. Genetic evidence for PLASMINOGEN as a shared genetic risk factor of coronary artery disease and periodontitis. Circ Cardiovasc Genet 2015; 8: 159–167. doi: <u>10.1161/CIRCGENETICS.114.000554</u> PMID: 25466412
- Divaris K, Monda KL, North KE, Olshan AF, Lange EM, Moss K, et al. Genome-wide association study of periodontal pathogen colonization. J Dent Res 2012; 91: 21S–28S. PMID: 22699663
- Schwenk RW, Angin Y, Steinbusch LK, Dirkx E, Hoebers N, Coumans WA, et al. Overexpression of vesicle-associated membrane protein (VAMP) 3, but not VAMP2, protects glucose transporter (GLUT) 4 protein translocation in an in vitro model of cardiac insulin resistance. J Biol Chem 2012; 287: 37530– 37539. doi: 10.1074/jbc.M112.363630 PMID: 22936810
- Du Clos TW. Pentraxins: structure, function, and role in inflammation. ISRN Inflamm 2013; 2013: 379040. doi: <u>10.1155/2013/379040</u> PMID: <u>24167754</u>
- **32.** Holers VM. The spectrum of complement alternative pathway-mediated diseases. Immunol Rev 2008; 223: 300–316. doi: <u>10.1111/j.1600-065X.2008.00641.x</u> PMID: <u>18613844</u>
- Mihlan M, Blom AM, Kupreishvili K, Lauer N, Stelzner K, Bergstrom F, et al. Monomeric C-reactive protein modulates classic complement activation on necrotic cells. FASEB J 2011; 25: 4198–4210. doi: <u>10.</u> <u>1096/fj.11-186460</u> PMID: <u>21856781</u>
- Mold C, Kingzette M, Gewurz H. C-reactive protein inhibits pneumococcal activation of the alternative pathway by increasing the interaction between factor H and C3b. J Immunol 1984; 133: 882–885.
  PMID: 6234363
- Suankratay C, Mold C, Zhang Y, Potempa LA, Lint TF, Gewurz H. Complement regulation in innate immunity and the acute-phase response: inhibition of mannan-binding lectin-initiated complement cytolysis by C-reactive protein (CRP). Clin Exp Immunol 1998; 113: 353–359. PMID: <u>9737662</u>
- 36. van Greevenbroek MM, Jacobs M, van der Kallen CJ, Blaak EE, Jansen EH, Schalkwijk CG, et al. Human plasma complement C3 is independently associated with coronary heart disease, but only in heavy smokers (the CODAM study). Int J Cardiol 2012; 154: 158–162. doi: <u>10.1016/j.ijcard.2010.09.</u> 017 PMID: 20926148
- Helmersson J, Larsson A, Vessby B, Basu S. Active smoking and a history of smoking are associated with enhanced prostaglandin F(2alpha), interleukin-6 and F2-isoprostane formation in elderly men. Atherosclerosis 2005; 181: 201–207. PMID: <u>15939073</u>
- Wannamethee SG, Lowe GD, Shaper AG, Rumley A, Lennon L, Whincup PH. Associations between cigarette smoking, pipe/cigar smoking, and smoking cessation, and haemostatic and inflammatory markers for cardiovascular disease. Eur Heart J 2005; 26: 1765–1773. PMID: 15817606
- Zieske AW, Tracy RP, McMahan CA, Herderick EE, Homma S, Malcom GT, et al. Elevated serum Creactive protein levels and advanced atherosclerosis in youth. Arterioscler Thromb Vasc Biol 2005; 25: 1237–1243. PMID: <u>15802624</u>