

## Respiratory Tract Infections in Diabetes – Lessons From Tuberculosis and Influenza to Guide Understanding of COVID-19 Severity

Amnah Al-Sayyar<sup>1</sup>, Katina D. Hulme<sup>2</sup>, Ronan Thibaut<sup>3</sup>, Jagadeesh Bayry<sup>4</sup>, Frederick J. Sheedy<sup>5</sup>, Kirsty R. Short<sup>2,6</sup> and Fawaz Alzaid<sup>1,3\*</sup>

<sup>1</sup> Dasman Diabetes Institute, Dasman, Kuwait, <sup>2</sup> School of Chemistry and Molecular Biosciences, The University of Queensland, St Lucia, QLD, Australia, <sup>3</sup> Institut Necker Enfants Malades (INEM), Institut National de la Santé et de la Recherche Médicale (INSERM) U1151/CNRS UMRS8253, Immunity and Metabolism of Diabetes (IMMEDIAB), Université de Paris Cité, Paris, France, <sup>4</sup> Department of Biological Sciences & Engineering, Indian Institute of Technology Palakkad, Palakkad, India, <sup>5</sup> School of Biochemistry and Immunology, Trinity College, Dublin, Ireland, <sup>6</sup> Australian Infectious Diseases Research Centre, The University of Queensland, St Lucia, QLD, Australia

#### **OPEN ACCESS**

#### Edited by:

Hamad Ali, Kuwait University, Kuwait

## Reviewed by:

Athanasia Papazafiropoulou, Tzaneio Hospital, Greece William B. Grant, Sunlight Nutrition and Health Research Center, United States

#### \*Correspondence:

Fawaz Alzaid fawaz.alzaid@dasmaninstitute.org; fawaz.alzaid@inserm.fr

#### Specialty section:

This article was submitted to Diabetes: Molecular Mechanisms, a section of the journal Frontiers in Endocrinology

> Received: 13 April 2022 Accepted: 17 June 2022 Published: 26 July 2022

#### Citation:

Al-sayyar A, Hulme KD, Thibaut R, Bayry J, Sheedy FJ, Short KR and Alzaid F (2022) Respiratory Tract Infections in Diabetes – Lessons From Tuberculosis and Influenza to Guide Understanding of COVID-19 Severity. Front. Endocrinol. 13:919223. doi: 10.3389/fendo.2022.919223 Patients with type-2 diabetes (T2D) are more likely to develop severe respiratory tract infections. Such susceptibility has gained increasing attention since the global spread of Coronavirus Disease 2019 (COVID-19) in early 2020. The earliest reports marked T2D as an important risk-factor for severe forms of disease and mortality across all adult age groups. Several mechanisms have been proposed for this increased susceptibility, including pre-existing immune dysfunction, a lack of metabolic flexibility due to insulin resistance, inadequate dietary quality or adverse interactions with antidiabetic treatments or common comorbidities. Some mechanisms that predispose patients with T2D to severe COVID-19 may indeed be shared with other previously characterized respiratory tract infections. Accordingly, in this review, we give an overview of response to Influenza A virus (IAV) and to *Mycobacterium tuberculosis* (Mtb) infections. Similar risk factors and mechanisms are discussed between the two conditions and in the case of COVID-19. Lastly, we address emerging approaches to address research needs in infection and metabolic disease, and perspectives with regards to deployment or repositioning of metabolically active therapeutics.

Keywords: diabetes, infection, influenza, COVID - 19, tuberculosis, inflammation

# 1 INTRODUCTION: TYPE-2 DIABETES AND SEVERITY OF RESPIRATORY TRACT INFECTIONS

Diabetes is a chronic condition characterized by persistently high levels of glucose in blood due to insufficient insulin action or secretion. The international diabetes federation (IDF) estimates that there will be 783 million adults with diabetes in 2045, which is 46% higher compared to the number of cases in 2021 (1). The IDF also reported Type-2 Diabetes (T2D) to be the most prevalent form,

accounting for 90% of all cases worldwide (1). T2D is a condition of insulin resistance, defective insulin secretion and  $\beta$ -cell destruction, these are associated with inflammation and metabolic stress. It results from several risk-factors, such as aging, lifestyle and genetic predisposition (2). Metabolic stress due to loss of glycemic homeostasis results in detrimental microvascular and macrovascular complications and hepatic comorbidities (3). Studies have also shown that T2D increases patient susceptibility to severe infections, with this being attributed to microenvironmental dysmetabolism impairing the immune responses (4, 5). Importantly, this applies to several forms of infection: skin and soft tissue infections, urinary tract infections and respiratory infections, with increased hospitalization and mortality rates (6, 7).

Respiratory infections are considered one of the major severe infections associated with diabetes. Hyperglycemia and increased protein glycosylation were found to be associated with microangiopathic alterations in the lungs of patients with T2D (8). Based on the pneumonia severity index, a higher proportion of diabetic patients suffer severe respiratory infections, compared to non-diabetic patients (52.3% vs. 9.4% of patients) (9). In addition, continuous exposure to high glucose also leads to formation of advanced glycation end-products (AGEs) which are implicated in the development of diabetic vascular complications, induce reactive oxygen species (ROS) and development of pulmonary fibrosis (10). These effects in the lung have been observed in common respiratory diseases that gravely affect patients with diabetes. These include asthma (11), pneumonia (7), tuberculosis (12), influenza (5) and Coronavirus Disease (COVID)-19 (13).

In such conditions, immune responses are often studied for potential impairments and as mechanistic targets contributing to severity (14). In normal physiology, immunity coordinates detection of pathogens, phagocytosis, antigen presentation and production of specific antibodies. In diabetes, the increased susceptibility to severe infection has been associated with defects in these functions at multiple levels (4). Results from a cross-sectional study showed that pulmonary functions were reduced in people with inadequate glucose control and high levels of inflammatory markers like TNFa, IL-6 and C-Reactive Protein (CRP) (15). These inflammatory markers are elevated in diabetes, under non-infectious conditions, and associated with hyperglycemia (16). It was also found that hyperglycemia suppressed IL-2, IL-6 and IL-10 in peripheral blood mononuclear cells (PBMCs) suggesting impaired cellular defense mechanism in patients with diabetes (17). Such markers in circulation reflect localized problems at a given site of infection, such as the lungs. Alveolar macrophages, that populate the lungs, are among the first points of contact with respiratory pathogens, and their dysfunction is a characteristic abnormality in severe cases of COVID-19 (18). These macrophages are specialized tissue resident cells of the innate immune system, generally detecting and disposing of invading foreign debris or pathogens. They also retain the capacity to raise an alarm via signaling to cells in the microenvironment (e.g., endothelial or dendritic cells, T-cells etc.).

Every stage of the immune response also relies on micronutrient balance or availability. Micronutrients have synergistic roles with the molecular machinery that facilitates immune effector functions. Vitamin D has been gaining attention as one of the most important micronutrients contributing to efficiency of immune reponses (19). Vitamin D is most widely known as a regulator of calcium and phosphate levels in the body and is necessary for healthy teeth, bones and muscles. However, Vitamin D also ensures integrity of innate immune responses at mucosal barriers, these are the first line of defense against invading pathogens (19). It also plays a role in cell-mediated processes, decrease pro-inflammatory cytokines expression and has inhibitory actions against pathogens (19). The inhibitory effect of the active form of vitamin D [Calcitriol (1,25 (OH)<sub>2</sub>D<sub>3</sub>)] is modulated by vitamin D receptors (VDRs) that are present in the heart, brain, pancreatic islets, immune cells, muscles and adipose tissue. These are all common target organs in the development of diabetes and its complications (20). Typically, during infection immune cells would trigger VDR signaling, to convert vitamin D to calcitriol, then induce the production of antimicrobial proteins such as cathelicidin (LL-37) via Toll-like receptor (TLR) activation (21).

Several studies have reported an inverse relationship between vitamin D status and T2D incidence (22–24) as its deficiency is associated with  $\beta$ -cell dysfunction and insulin resistance (25). Results from an observational study showed that the prevalence of hypovitaminosis D was higher in T2D compared to control (39% vs 25%) (26). Similarly, Plataki et al. also reported that T2D patients tend to have low levels of vitamin D, as well as of LL-37, providing a mechanism for impaired immune responses and antimicrobial peptide production (27). Such alterations of vitamin D could also increase susceptibility to severe respiratory infections especially in patients with asthma and pulmonary diseases (28). Interestingly, the lung's epithelium was found to "self-generate" the active form of vitamin D that increases LL-37 expression. Dysfunction of this process can also contribute to severity of respiratory infections (29).

Such immune-mediated mechanisms, and others, have been a central focus of COVID-19 research. Indeed, T2D is one of the top 3 risk-factors leading to severe infection and undesirable outcome of COVID-19 across all adult age groups (13). Interactions between infection, immunity, and metabolic disease has been studied to decipher which criteria contribute to risk and by which mechanisms does diabetes impair response to infection. However, prior to the emergence of SARS-CoV-2, decades of research into respiratory infection in T2D have given a wealth of information. This review brings together current knowledge in T2D and respiratory infections, mainly interactions between systemic metabolism and the immune system's response to bacterial and viral infectious pathogens. We discuss tuberculosis (TB) and influenza, that predate COVID-19, as longer-studied conditions well-known to be affected by diabetic status. We discuss risk-factors and mechanisms that contribute to severity in both conditions, then relate this to the current knowledge on COVID-19.

## 2 MYCOBACTERIUM TUBERCULOSIS AND METABOLIC DISEASE

The observation that metabolic factors determined the various outcomes of COVID-19, with severe COVID-19 manifesting primarily in people with metabolic syndrome (30, 31), indicates how host metabolic fitness underlines immune function. A similar spectrum of infection outcomes is observed in TB, caused by infection with the intracellular bacteria Mycobacterium tuberculosis (Mtb). These range from early clearance to latent tuberculosis infections (LTBI) to active TB and severe disease (32). In recent years, T2D emerged as a major risk-factor for developing severe disease, with TB-diabetes recently classified as a syndrome by WHO, particularly prevalent in Southeast Asia (33). Although both pre-diabetes and T2D can enhance risk of contracting TB in the form of LTBI in low burden TB countries (34), in areas where TB is endemic T2D certainly increases risk of active TB and is associated with poorer responses to treatment (12,

35, 36). Interestingly, in these environments, particularly Asia, T2D is associated with worse TB disease (37) irrespective of body mass index (BMI), while obesity in the absence of hyperglycemia is associated with a better prognosis (38). These observations suggest that multiple factors associated with metabolic syndrome including hyperglycemia, insulin resistance and hyperlipidemia can affect TB host immunity differently and ultimately affect disease outcome. Therefore, TB can serve as a case study for how metabolic factors promote severe infection risk. Insights into metabolic factor contribution to TB immunity, from clinical data and limited animal models, are discussed below (**Figures 1A, 2A**).

## 2.1 Factors Affecting Tuberculosis in Diabetes

#### 2.1.1 Hyperglycemia

Hyperglycemia affects immune responses to Mtb infection in mouse models and glycemic control has emerged as critical to disease outcome in patient cohorts (**Figure 1A**). Poorer glycemic





control is associated with more severe TB disease (39), and strategies to treat this reduce the relative risk. However, glycemia centered strategies do not return relative risk of severe disease to non-T2D levels (40). Mechanistically, peritoneal macrophages from diabetic (db/db) mice display impaired phagocytosis and cytokine responses (41, 42). *In vitro* studies suggest that increasing glucose concentrations enhance Mtb-induced pro-inflammatory cytokine responses in bone-marrow derived macrophages (BMDM), but this is only noted at high glucose concentrations which mimic hyperglycemia (43) (**Figure 2A**).

With the realization that immunometabolic reprogramming underlines macrophage responses in Mtb, these processes should be re-examined. In particular, monocyte-derived macrophages recruited to sites of Mtb infection up-regulate expression of the main macrophage glucose transporter GLUT1 and glucose consumption to promote anti-microbial functions and proinflammatory cytokine production (44, 45). Whether this is impacted by the systemic hyperglycemia, characteristic of T2D has not yet been described. However, circulating monocytes derived from pre-diabetic patients display enhanced cytokine production (46), while monocyte-derived macrophages from patients with T2D have impaired anti-microbial responses to Mtb (47). Similar impairments have been observed in alveolar macrophages derived from diabetic mice (48, 49), these cells seem to rely on oxidative metabolism (45) and are described further in the following sections. In vivo Mtb infections in diabetic mouse models display heightened lung inflammation and bacillary burden (50, 51). This correlates with clinical observations of increased lesions, cavities and increased transmissibility of TB in T2D patients (52, 53). Although the antigen-presenting ability of innate cells was not affected by hyperglycemia (54), altered cytokine production can affect the nature of the subsequent T-cell response contributing to poorer bacterial containment (48). Importantly, in vivo studies have also found a profound defect in inherent lymphocyte function in streptozotocin-treated hyperglycemic mice. Martinez et al. found that the accumulation of Receptor for Advanced Glycation End-Products (RAGE)-ligands during chronic hyperglycemia activates a p38 dependent pathway that epigenetically reprograms naïve T-cells, making them hyper-responsive to subsequent activation by Mtb antigens (54) (Figure 2A). Thus, both myeloid and lymphocyte function is inherently impaired under hyperglycemic conditions and can escape the regulatory mechanisms active under homeostasis.

#### 2.1.2 Hyperinsulinemia

Hyperinsulinemia is the result of insulin resistance, given that pre-diabetes is associated with increased risk of LTBI and that cells from pre-diabetic patients display altered responses to stimulation (40, 46), there is an interest in identifying if hyperinsulinemia in pre-diabetes contributes to altered immune responses (Figure 1A). The recent interest in immunometabolism has demonstrated that Mtb-infected macrophages upregulate glycolysis to promote anti-bacterial and pro-inflammatory cytokine functions (44, 45), processes enhanced by T-cell-derived interferon (IFN)- $\gamma$  and limited by virulent and drug-resistant forms of Mtb (55-57). Although GLUT1 expression is insulin-independent, and insulindependent GLUT4 is not expressed in macrophages (58), insulin receptor signaling can have both pro- and antiinflammatory roles and could impact Mtb immunity. Mtb infection itself can induce transient hyperglycemia through stimulating insulin production which, in guinea pig models, has been linked to more severe disease (59). Identifying these insulin-dependent mechanisms could help explain the complex interdependency between insulin signaling and glucose homeostasis in increased disease severity linked to T2D.

#### 2.1.3 Dyslipidemia

Since glycemic control does not completely reduce risk of severe TB in T2D (40), other metabolic factors have been considered, chiefly, altered lipid homeostasis. Increased BMI contributes to resilience against severe disease (38) and limited human data suggests cholesterol-rich diets may actually protect against TB disease (60). Larger epidemiological studies in Asia have demonstrated that lower total cholesterol levels are linked to more severe TB and poorer treatment outcomes (61). These data suggest a protective role for total cholesterol against TB, which have not been always observed in animal models or in vitro studies. Indeed, hyperlipidemic mice display enhanced TB disease with increased lung inflammation characterized by myeloid nitrous oxide production, increased bronchoalveolar fluid (BALF) T helper type 1 (Th1) cytokines, poorer lung histology and higher bacterial burdens and dissemination to organs (62, 63)(Figure 1A). The increased myeloid inflammation - both macrophage and neutrophil driven, promotes tissue degradation while simultaneously limiting the development of protective T-cell responses. As a caveat, the mouse models used to illustrate this (Apoe-/-, Ldlr-/-; highfat fed genetically-deficient mice) display abnormally high cholesterol levels, only seen in humans with familial hypercholesterolemia and may not accurately reflect TB-diabetes.

Mtb itself is a lipid rich bacillus and must obtain cholesterol for biosynthesis and growth from the host (64). This promotes long-term persistence in the host since mutant strains which lack cholesterol transporter systems (e.g. mce4) succumb to the proinflammatory effects of IFN- $\gamma$  (65). The TB granuloma, contains a lipid-rich core surrounded by various foamy macrophage cell types (66). Although initially thought to represent a nutrient source for the mycobacteria, recent lipidomics and single-cell analysis suggests that heterogeneity exists in both the lipid content and inflammatory phenotype of granuloma macrophages (67, 68). While Mtb infection can alter both cholesterol and fatty acid metabolism to favor mycobacterial growth (69, 70), the process of lipid droplet formation appears to be host encoded and augmented by IFN-y treatment (Figure 2A). This drives initiation of triglyceride biosynthesis and lipid uptake to maintain lipid droplets in infected macrophages. This process is linked to the production of eicosanoids which promote host-defence (67). Much of this work has been performed in BMDM cultures and it is now appreciated that the alveolar macrophages, initial host cells rely basally on fatty acid oxidation and do not employ anti-Mtb glycolysis, which is the opposite to infiltrating macrophages. Dodd et al. illustrated that Mtb-infected alveolar macrophages and monocyte-derived macrophages upregulate the lipid transporter CD36 to drive an anti-inflammatory, promycobacterial phenotype (71, 72). Surfactant lipoproteins in the pulmonary space facilitate this and are internalized during infection (71). The lipid-rich environment of the lung itself undoubtedly shapes alveolar macrophage development and may explain the tropism of Mtb for the pulmonary compartment. Whether these processes are altered in the setting of hyperlipidemia is unclear and may depend on the nature of the dyslipidemia. Hypertriglyceridemia in western populations was found to be particularly pathogenic and linked to poorer treatment outcomes than hypercholesterolemia (73). Another confounding factor is that often it is not simply elevated lipid levels themselves, but modified lipid species generated during disease (74). Oxidized forms of low-density lipoprotein (LDL)-cholesterol and sterols have been shown to impact

macrophage Mtb responses and macrophage recruitment to the lung respectively (75, 76).

#### 2.1.4 Vitamin D Deficiency

Alterations in vitamin D levels in individuals with diabetes and Mtb can disrupt the maintenance of immune homeostasis. Several studies have reported that cases of Mtb infection are associated with low levels of vitamin D, in particular active Mtb (77, 78). A randomized controlled trial confirmed that upon vitamin D supplementation, patients had significantly reduced symptoms by 36.6% in the first month (79). Since TLRs are linked to the antimicrobial function of vitamin D, Liu et al. investigated this mechanism and found that the induction of antimicrobial peptide LL-37 inhibits Mtb proliferation in human monocytes and macrophages (80). This was confirmed further in vitro where the inhibition of LL-37 led to enhanced growth of Mtb, confirming LL-37's suppressive role (81). Very limited number of studies have investigated the status of vitamin D and LL-37 in patients with TB and diabetes. Zhan and Jiang assessed the levels of vitamin D in patients with Mtb and diabetes mellitus and found that the concentration of vitamin D was 35% lower in patients with Mtb and DM compared to the control group (82). In addition, patients with both conditions exhibited high levels of LL-37 that is correlated with a positive acid-fast bacillus smear which indicates the presence of TB (82).

## 2.2 Chronic Inflammation and Immune Training

As a consequence of both hyperlipidemia and hyperglycemia, systemic inflammation is triggered which underlines many of the metabolic sequelae of T2D. This baseline meta-inflammation could also impact host-defense responses and is already implicated in defective anti-tumor immunity (83). Chronic inflammation inhibits both resolution of inflammation and the development of protective lymphocyte responses and can also alter TB granuloma composition (51, 62). Recently, chronic inflammation associated with hyperlipidemia and western diets was shown to reprogram myeloid progenitor cells for heightened activity through the process of innate immune training (84, 85). Following this, Choudhury and colleagues demonstrated that hyperglycemic but not hyperlipidemic mice also displayed heightened monocyte cytokine production due to epigenetic modification and alterations to bone-marrow myelopoiesis, characteristic of innate immune training (84, 86). This process contributes to the development of T2D, but may also impair host-defense in TB-diabetes. With hematopoietic progenitor cells emerging as a key target of inflammation affecting immune cell composition and fate (87), the chronic inflammation in T2D could alter immune cell fate and affect granuloma composition. Recent profiling studies examining active TB granulomas demonstrate a more regulatory phenotype dominated by immunosuppressive myeloid and Regulatory T cells (Treg) (68) and the impact of meta-inflammation on this warrants examination.

# 2.3 (Immuno)-Metabolic Treatments for TB-Diabetes

Although control of glycemia in T2D doesn't completely reduce the risk of increased TB (40), metabolic drugs are being assessed for their ability to prevent disease due to their immunometabolic effects. They can also reveal novel targets in pathogenesis which could form the basis for future novel immuno-therapies or vaccine targets. In a normoglycemic mouse model of TB, metformin was shown to exacerbate late-stage disease by ameliorating inflammation (88). In particular, IL-1 $\beta$  expression was reduced, which can be regulated through metformininduced inhibition of mitochondrial complex 1 activity (89). Similar results were observed in humans receiving metformin, with improved macrophage responses to Mtb infection (90). More pressingly, in the context of T2D, a recent meta-study revealed that diabetes controlled by metformin led to a reduced risk of TB in humans (91). In contrast, a recent mouse study of Mtb infection in diet induced obesity (DIO)-mice suggests that metformin is only beneficial in the hyperglycemic context (92). More recently, cytotoxic CD8<sup>+</sup> lymphocytes emerged as targets for metformin in Mtb-infection models in euglycemic mice. Metformin metabolically reprograms these cells toward oxidative metabolism and promotes anti-microbial function. Traditionally TB vaccines target CD4<sup>+</sup> Th1 cells, so this study is important as it places these CD8<sup>+</sup> T-cells as key in protecting against severe disease, particularly relevant in the metabolically dysregulated context of T2D.

Because of Mtb's tropism for cholesterol, the HMG-CoA reductase targeting drugs, statins, have received attention both as a direct anti-Mtb drug and for use in the context of TB diabetes. By reducing cholesterol biosynthesis at this early ratelimiting step (93), statins lead to compensatory low-denisty lipoprotein receptor (LDLR) expression and reduced serum LDL-cholesterol which should be beneficial in the context of hyperlipidemia and its effect on TB pathogenesis. Clinical studies are examining the feasibility of this (94), however given the controversial role of elevated total cholesterol in TB host defense, the results thus far are unclear. Since targeting HMGR leads to knock-on effects on the production of multiple lipid species (93, 95), statins may affect mycobacterial growth. In vitro studies suggest that statins may directly alter axenic Mtb growth in culture (96), Mtb growth in macrophages is also reduced after statin treatment (97). This may be due to cholesterol depletion but also could be mediated through the immunomodulatory functions of statins (95). Finally, novel lymphocyte subsets which present lipid antigens via CD1 have recently been associated with early immune responses in TB including  $\gamma\delta$  T and invariant natural killer T (iNKT) cells (98). The impact of altered systemic lipid homeostasis as well as statin treatment on their function needs to be considered, as does the nature of vaccine strategies used to boost TB immunity, with current targets focusing solely on MHC-restricted peptide antigens that promote CD4<sup>+</sup> T-cells.

## 3 INFLUENZA A VIRUS AND METABOLIC DISEASE

Influenza A virus (IAV) presents a continuous threat to human health, infecting up to 15% of the population annually (99, 100). Influenza infection severity largely depends on the immune and health status of the individual (101). Although considered to be relatively mild, the 2009 H1N1 pandemic served to emphasize that certain host comorbidities increase the risk of severe disease upon influenza virus infection (102, 103). Prior to 2009, reports already emphasized that diabetes increases the risk of lower respiratory tract infections (LRTI) and pneumonia related hospitalization, and indeed led to more severe influenza outcomes (104-106). Both patients with Type 1 Diabetes (T1D) and T2D were at greater risk of developing LRTI (including influenza), while there was no difference in the risk of upper respiratory tract infections over a 12-month time period (105). Interestingly, in this study T2D patients in particular had an increased risk of developing two or more episodes of a LRTI within the 12-month period. This indicates that hyperglycemia per se may be an important factor in LRTI severity, but not necessarily in frequency of LRTIs. A 7-year retrospective study reported that individuals with diabetes are at a higher risk of being hospitalized due to pneumonia (104). This increased risk was also associated with increasing blood glucose levels (104). Furthermore, during a one year period, individuals with diabetes were more likely to have influenza or pneumonia listed as cause of death compared to people without diabetes, regardless of race, sex, and socioeconomic status (106). Strikingly, it was estimated that during this time, individuals with diabetes made up more than 10% of the recorded influenza and pneumonia associated deaths in the U.S (106). Together, these studies demonstrate the enhanced risk of severe influenza outcomes in individuals with diabetes during seasonal influenza epidemics.

Much of the current literature regarding the susceptibility of individuals with diabetes to severe IAV infection emerged in response to the 2009 H1N1 pandemic (pH1N1) and has been addressed in a previous review (107). Specifically, diabetes was identified as one of the most common host comorbidities in those hospitalized with pH1N1 (108). During the 2009 pandemic, individuals with diabetes had around a 3- to 4-fold increased risk of hospitalization (109–112), intensive care unit (ICU) admission (109, 113, 114), and death (113–115). Indeed, one study reported up to a 9-fold increased risk of hospital mortality in individuals with diabetes (116). While diabetes was identified as an independent risk-factor for severe pH1N1 outcomes, this risk of ICU admission and death becomes higher in patients with additional underlying medical conditions (117).

## 3.1 Factors Influencing Influenza in Diabetes

#### 3.1.1 Hyperglycemia

Experimentally, murine models of both T1D and T2D show that diabetic mice have more severe influenza outcomes when compared to healthy controls (107, 118–120), providing

experimental evidence that hyperglycemia *per se* is a major contributor to disease severity. Studies using an *in vivo* STZ-induced model of T1D, demonstrated that diabetic mice had increased viral load with a more extensive infection, and lower rates of survival (118, 120, 121). This increase in viral load has also been correlated to increasing blood glucose levels observed in T1D mice (119). Similar results have been observed using a leptin receptor-deficient model for obesity and T2D. Specifically, elevated blood glucose levels correlated with increased viral copy number and an increase in the pulmonary pro-inflammatory cytokine response (122) (**Figure 1B**).

Despite the abundance of data that indicates the role of diabetes in influenza severity, the research into mechanisms remains limited. Chronic hyperglycemia can induce oxidative stress *via* the production of ROS and is thought to be one of the key sources of hyperglycemia-induced diabetes complications (123, 124). There are several mechanisms by which hyperglycemia causes increased oxidative stress, such as the production of AGEs, the activation of Protein Kinase-C (PKC), the accumulation of sorbitol, and the hyperactivity of the hexosamine pathway (125). Cumulatively, these lead to the over production of ROS and a decrease in the endogenous antioxidant defense systems (125). Given the causal role of hyperglycemia-induced ROS has in the development of the non-infectious complications of diabetes, it is likely that hyperglycemia-induced ROS production is a driving factor in increased severity of influenza in individuals with diabetes (**Figure 2B**).

There is evidence that hyperglycemia-driven ROS production alters cellular metabolism that can shape the inflammatory response during viral infections (126), however this has yet to be explored in influenza. Hyperglycemia can upregulate glycolysis (127), which can promote a pro-inflammatory immune cell phenotype (128), and increase viral replication (129) (Figure 2B). Despite not coming in direct contact with circulating blood, glucose concentrations in the airway surface liquid increase as a direct result of elevated blood glucose levels (130). High levels of glucose cause a dose-specific increase in IAV infection and replication in Madin-Darby Canine Kidney (MDCK) epithelial cells (129). This was associated with a glycolysis-dependent increase in the assembly of the cellular V-ATPase which is necessary for viral release into the cytoplasm (129). Indeed, increased viral replication has been observed in murine models of both T1D and T2D, and correlated with increased blood glucose levels (119-122). This increase in viral replication as a result of abundance of glucose is likely due to ROS-driven alterations to metabolic function. While this is yet to be directly demonstrated in the context of influenza, high levels of glucose have been shown to increase SARS-CoV-2 viral replication and monocyte cytokine production in a ROS/ glycolysis-dependent manner (131).

In addition to altering cellular metabolic function, a history of exposure to hyperglycemia can lead to ROS-driven endothelial dysfunction in diabetes (132). While endothelial cells are not normally infected by influenza virus in humans, they play a crucial role in influenza pathogenesis given their close proximity to the pulmonary alveolar epithelium. Specifically, during IAV infection, it is the pulmonary endothelial cells that are believed to

be a major source of cytokine production in the lungs (133, 134). We have previously demonstrated using both an *in vitro* and *in vivo* model, that exposure to high glucose conditions prior to IAV infection increased virus-induced pulmonary barrier damage (122). This was associated with an increased pro-inflammatory response in endothelial cells and the subsequent damage of the epithelial junctional complex. This is likely to be further exacerbated in conditions of glycemic variability (**Figure 2B**).

#### 3.1.2 Glycemic Variability

Typically, individuals without diabetes have blood glucose levels ranging between 70-100 mg/dl (135). These glucose levels fluctuate during the day, particularly post-meal up to almost 140 mg/dl (135). These glucose fluctuations are referred to as glycemic instability or glycemic variability and can be more extreme and prolonged in individuals with uncontrolled diabetes.

There is mounting evidence that glycemic variability is associated with greater ROS production than steady hyperglycemia (125), and could further impair the immune response (107, 136). However, glycemic variability as a factor in severe influenza infections remains a relatively new topic, with an extremely limited amount of research into its role, and most studies are outside the scope of influenza. Nevertheless, there has been one key study that has investigated the role of glycemic variability in the context of influenza. Using an in vitro coculture model mimicking the respiratory epithelial-endothelial barrier we have shown that compared to hyperglycemia, glycemic variability increased viral replication, cell death, and inflammation of both the epithelial and endothelial cells (136). This was correlated with an increase in a marker of oxidative stress. These results were confirmed using an in vivo model, where mice that experience glycemic variability in the weeks leading up to infection suffered more severe influenza (136). Specifically, mice with glycemic variability had increased weightloss, decreased lung function, and increased apoptotic cell death. This was again associated with increased pulmonary inflammation and oxidative stress. Together, these data suggest a key role of dysregulated glucose levels, both elevated and variable, driving severe influenza in the context of a primary infection (Figure 1B).

## 3.2 Adaptive Immune Responses

In addition to its effect on the innate immune system and viral replication, diabetes may also have direct effects on the cellular adaptive immune response to viral infection. There is evidence that hyperglycemia induces hyperresponsiveness, enhanced activation, and proliferation of T cells (54, 137) (**Figure 2B**). However, there is perhaps a larger body of evidence that hyperglycemia impairs responses, increases the frequency of senescent cells, and impairs the proliferation of T cells (138–140). *In vitro* evidence suggest that high glucose concentrations reduce the production of IFN- $\gamma$  by CD8<sup>+</sup> T cells (141), and reduces their viability (142). Consistent with this, it has been shown that when compared to healthy controls, CD8<sup>+</sup> T cells (143), and using genome-wide expression analysis of PBMCs from donors with diabetes showed a reduction in activity of cytotoxic

genes compared to controls (144). Much like what has been observed in innate immunity, there is an emerging role of glycemic variability in negatively shaping the adaptive immune response to influenza. Compared to hyperglycemia, we have shown that glycemic variability increases pulmonary inflammation, oxidative stress and influenza severity following a secondary influenza infection in a murine model (136). It was speculated that this increase in severity could be driven by glycemic variability-induced oxidative stress reducing the CD8<sup>+</sup> T cell function. Currently, there is very little known about the effect of both hyperglycemia and glycemic variability on the adaptive immune response to influenza, and further indepth investigation is warranted.

Taken together these studies suggested that individuals with diabetes suffer more severe influenza as a result of i) metabolic dysfunction, ii) increased viral replication, iii) endothelial dysfunction, and iv) dysregulation of the immune response.

## **3.3 Response to Vaccination in Diabetes**

As individuals with diabetes are at a higher risk of developing serious influenza complication, influenza vaccination is currently recommended by the Centers for Disease Control and Prevention (CDC) for patients with diabetes (145). During recent influenza seasons, almost 30% of adults hospitalized with influenza had diabetes (145), highlighting the need for vaccination in this vulnerable group. During influenza epidemics of the early 1990s, vaccination of individuals with diabetes (both T1D and T2D) reduced hospital admission by almost 80% (146). A more recent meta-analysis determined that when individuals with diabetes (both T1D and T2D) are vaccinated, there is a reduced risk of hospitalization for pneumonia, and a lower mortality rate (147). Furthermore, while individuals with diabetes are at risk of developing cardiovascular complications following influenza virus infection (148, 149), influenza vaccination is associated with a reduced risk of cardiovascular mortality in adults with diabetes (150). While there are rare case study reports of adverse reactions to influenza vaccination in individuals with diabetes, overall, reactogenicity is similar in individuals with diabetes and healthy adults (151, 152). Although influenza vaccines are proven to be safe and reduced the development of severe complications, there have been some questions over vaccine efficacy in people with diabetes. Specifically, there is evidence that whilst patients with T2D were more likely to have received the influenza vaccination in the last 12 months they still experienced a greater number of respiratory infections than their non-diabetic counterparts (153). This is likely due to their reduced T cell and antibody response to influenza vaccines. In a small cohort, patients with T1D had decreased T cell response to influenza A-H1N1 subunit vaccine compared to controls, and this was associated with hyperglycemia (143). A larger study that encompassed both T1D and T2D reported that in the T1D group there was a significant increase in antibody non-responders to two of the three vaccine components (143). While this may be cause for concern, it has been demonstrated in vivo that vaccination via a higher dose, or a second low dose, increases antigen levels in diabetic mice to the point that they are able to protect against an

Respiratory Tract Infections in Diabetes

otherwise lethal challenge (120, 121). Together, this emphasizes the importance of maintaining up-to-date vaccination in individuals with diabetes. Unfortunately, vaccination rates for influenza are often lagging amongst those with diabetes. The current target vaccination rate for patients with diabetes is 75% (99). However, in developed countries coverage values are around 50-70% (154, 155), with some as low as 10% (156). Recently, in China, the reported vaccination rate of individuals with diabetes was below 8%, despite more than 46% of participants reporting an original intention to receive the vaccine (157). On top of lagging vaccine coverage rates in individuals with diabetes, the overall efficacy of the vaccine is known to change from season to season, as the circulating strains change. For example, the CDC has reported the current influenza vaccination for the 2021-2022 flu season in the U.S. did not reduce the risk of outpatient illness caused by influenza A (H3N2) (158). This suggests that not all influenza associated complications and death in diabetic individuals can be prevented with vaccination alone. Lagging vaccination rates, combined with sub-optimal vaccine efficacy in some influenza seasons means that severe influenza infections remain an issue for those living with diabetes.

## 4 COVID-19 SEVERITY AND TYPE-2 DIABETES

The above examples of respiratory infections in patients with diabetes clearly indicate a strong interaction between metabolism and successful detection and clearance of invading pathogens from the lung. Whether this be at the level of systemic metabolism or cellular immunometabolic defects that alter immune responses, a wealth of knowledge from cases of influenza and of TB can be used to draw comparisons with COVID-19.

Global events since March 2020 have made COVID-19 a near singular preoccupation of medical and research professionals due to the unprecedented strain on healthcare services. Early in the outbreak, T2D and associated metabolic syndrome were identified as risk factors for severity and death from COVID-19 (159). Diabetes is underpinned by inflammation and systemic dysmetabolism putting patients at-risk of other comorbidities. Although, initial reports focused on the immunology of SARS-CoV-2 infection and clinical trials applied antiviral or antiinflammatory therapies (160); higher mortality in patients with pre-existing dysmetabolism indicates that metabolic mechanisms are also attractive, to establish risk, or redeploy therapeutics to mitigate severity. Indeed, the increased severity of other respiratory infections (e.g., TB, influenza) in diabetes and the interactions with specific metabolic traits support those common features of metabolic decline that are more important to focus on to mitigate risk.

The earliest report dedicated to COVID-19 patients with T2D revealed that glycemic instability/variability increases risk of severe disease (13). Further meta-analyses found that of the components of metabolic syndrome, diabetes is the biggest contributor to adverse

outcome compared to hyperlipidemia, obesity and hypertension (161). Multiple reviews have addressed the interactions between COVID-19 and T2D (162, 163). Here we will cover common points with risk factors for TB and influenza, as well more recent questions regarding metabolically active therapeutics and responses to vaccination in the population with TD2 and COVID-19.

## 4.1 Factors Influencing COVID-19 in Diabetes

Identified factors that increase risk of COVID-19 include increased inflammation, increased ROS, insulin resistance, hyperglycemia and vascular endothelial damage. All of these factors are pre-existent or accentuated in patients with T2D prior to infection with SARS-CoV-2 (163). These risk factors are generally shared with IAV and Mtb infection (Figures 1C, 2C). The particularity of SARS-CoV-2 is its activation of the renin-angiotensin-aldosterone system (RAAS) via the virus's main entry point, the Angiotensin converting enzyme (ACE)-2 (164). The RAAS is defective in T2D and has long been known to contribute to the development of complications in diabetes (nephropathy, macroangiopathy) (165, 166). However, dysfunction of this system cannot solely explain the increased susceptibility of patients with diabetes to a plethora of infectious respiratory conditions. Therefore, other metabolic factors are valuable candidates to look into to better understand risk.

#### 4.1.1 Hyperglycemia

Hyperglycemia increases SARS-CoV-2 replication in circulating monocytes (131). Interestingly, on the immunometabolic front, mitochondrial adaptation also occurs in these cells to produce more ROS, contributing to severity. Our own work demonstrated that patients most severely affected exhibit morphological and functional changes in their monocyte pool (167). These changes are related to hyperinflammation and interferon signaling, associated with severity and are more pronounced in T2D. Thus, potentially *via* an interaction with monocytes, hyperglycemia may be an important mechanism-based risk-factor for severe COVID-19. Of note, glycemic control tends to deteriorate with infection in patients with T2D (**Figure 2C**). Those on insulin will require increasing doses to lower glycemia, and this requirement is associated with increased levels of inflammatory cytokines (168, 169).

Local and systemic inflammation are characteristic of COVID-19. Autopsy studies have reported inflammatory infiltrate, in the form of macrophages and lymphocytes, in a number of tissues, including the lungs, myocardium, liver and the brain (170, 171). Systemically, the cytokine storm syndrome and hyperinflammation are common and potentially life threatening in severe cases (172). The major immune signals that are impaired in COVID-19 at the transcriptional and translational levels are pro-inflammatory IL-6 signaling and the type-1 interferon system (173, 174). Mechanisms shared with other coronavirus infections have been proposed to link immune responses to disturbed metabolic homeostasis and worsening disease course. Notably, the large burden of

inflammatory infiltrate affects key insulin-responsive tissues, including the liver and skeletal muscle (175).

#### 4.1.2 Dyslipidemia

The role of dyslipidemia in COVID-19 severity on a background of T2D is unclear. Studies have found either increased risk of severe COVID-19 with dyslipidemia or no effect (176). The lack of clarity comes from confounding factors (such as multiple treatments, age and comorbid conditions) and from heterogeneity of studied populations.

#### 4.1.3 Vitamin D Deficiency

Hypovitaminosis D is associated with severe COVID-19, increased hospitalization and increased mortality (177). Several studies reported the inverse relationship between vitamin D supplementation and COVID-19 and its role as a preventive measure (178, 179). Generally, the protective effects of vitamin D against COVID-19 are linked to respiratory epithelial cell production of LL-37 that contributes to host-defense mechanisms through the disruption of viral membranes and replication (180). This was confirmed by Roth et al. as they demonstrated the role of LL-37 in preventing viral cell entry by binding to the ACE2 receptor of SARS-CoV-2 (181). It was also noted that vitamin D deficiency leads to a reduction in modulatory potential towards the cytokine storm during viral infection. This phenomenon is implicated in braking the lung epithelium, resulting in alveolar edema (182).

To date, very limited data are available on the link between hypovitaminoses in diabetic individuals and COVID-19. Results from a clinical study found that 76% of patients with vitamin D deficiency and hyperglycemia had severe COVID-19 with increased hospitalization and mortality rates compared to patients with normal vitamin D and glucose levels (183). They also confirmed that hypovitaminosis D in hyperglycemia resulted in worse respiratory parameters and increased levels of IL-1b, IL-6 and IFN- $\gamma$  (183). Similar results were also found in obese and vitamin D deficient individuals with 72% increase in infection severity compared to the control group (183). These studies suggest that optimal nutrition and supplementation with vitamin D is a promising candidate as a preventive measure or potentially an adjunct treatment for COVID-19 (184).

#### 4.1.4 Obesity

Obesity is considered as a major risk factor for COVID-19 infection due to its significant role in increasing systematic inflammation through the dysregulation of adipose tissue. Compared to nonobese individuals, obese individuals had greater COVID-19 severity, hypoxemic respiratory failure and higher baseline initial serum levels of CRP and IL-6 (associated with low-grade-chronic inflammation) (185). This suggests that obesity leads to worse COVID-19 outcomes that are associated increased inflammation and metabolic dysregulation.

## 4.2 Response to Vaccination and Metabolically Active Therapeutics

Efficiency of the COVID-19 vaccine response has been evaluated in patients with T2D. Vaccination has been reported to be

efficient in patients with good glycemic control compared to patients with uncontrolled T2D. Therefore, the adaptive immune compartment does not seem to be compromised with regards to antibody production in T2D. However, hyperglycemia in uncontrolled diabetes results in weak immunity, indicating that poor glycemic control can indeed impair the antibody producing branch of adaptive immunity. Two independent studies reported similar results (186, 187).

Of the different classes of metabolically active drugs applied in metabolic syndrome and T2D, some have been studied for their contribution to risk or better outcome of infection with SARS-CoV-2. Given the risk factors described above, one would hypothesize that rigorous glycemic control and use of insulin would be beneficial, whilst use of lipid lowering drugs may not affect outcome. Metanalyses have found that user of metformin and sulfonylureas had lower mortality risk (188). Metformin acts by lowering hepatic glucose output, with a number of proposed molecular mechanisms, and Sulfonylureas are insulin secretagogues. Interestingly, some studies report that insulin use was associated with worse outcome (188). This is at odds with the proposed action on stimulating CD8<sup>+</sup> T-cell function, however in an already diabetic context, the use of insulin may be a sign of progressed diabetes and thus an overall less healthy metabolic status of the patient. Use of dipeptidyl peptidase-4 inhibitors did not have an effect on disease outcome. So, these studies indicate that use of antidiabetic agents either improve or do not affect outcome in COVID-19 in patients with T2D; unless patients with T2D have progressed to the stage of use of exogenous insulin. With regards to controlling dyslipidemia, the most commonly used class of drugs is Statins and studies have been ambivalent. Some meta-analyses conclude better outcome from retrospective studies (189) whilst others report improvement, worsening or no effect (190).

## 5 PERSPECTIVES: METABOLOMIC SHIFTS AND METABOLIC DEPRESSION IN COVID-19

Beyond pre-existing T2D, and clinical and immunological factors mentioned above, another way of addressing interactions between metabolism and response to infection is the application of metabolome studies. Such approaches are most valuable to basic research when applied translationally, studies are often data-driven in nature and when combined with clinical and immunometabolic observations, metabolomic approaches may lead to further hypothesis generation to direct future mechanistic work. Two reports by Shen et al (191) and by Wu et al (192) actually characterized the systemic metabolomic, lipidomic and proteomic responses to SARS-CoV-2 infection. Concertedly, these studies show that COVID-19 is characterized by a generalized systemic metabolic depression, although they were carried out in non-diabetic patients, metabolomic and lipidomic changes may indicate which specific pathways are dysregulated in COVID-19, and potentially subject to worsening in patients with metabolic disease.

The study by Shen et al. investigated proteomic and metabolomic profiles of sera from patients with severe and non-severe COVID-19 and non-COVID-19 patients with similar clinical presentation (other respiratory infections). In this study, the COVID-19-dependent proteome and metabolome represented three major pathways: complement system, macrophage function and platelet degranulation (191). Lipoproteins, sphingolipids, glycerolipids, steroid hormones and their intermediates related to macrophage function were downregulated in COVID-19 and correlated to severity. These molecules play important roles in immune function, including signaling, regulating membrane properties, apoptosis, migration and importantly in the resolution of inflammation. This lipid repressive profile is specific to SARS-CoV-2, as studies with other viruses (HCoV-229E, MERS-CoV) found increases of several of these lipid mediators relative to healthy subjects (193). As for liver-derived molecules, increased bilirubin degradation products and bile acid derivatives indicate impaired hepatic detoxification and urea cycle activity is also altered, which is a typical consequence of the interferon response associated with viral infection. Interestingly, the proteomic profile also showed decreased serotonin correlating to disease severity; which would influence sickness behaviors (e.g., lack of appetite, lethargy) also altering systemic metabolism.

The publication by Wu et al (192) carried out kinetic targeted lipidomic and metabolomic profiling of sera from healthy subjects and from COVID-19 patients ranging from mild to severe, and to fatal disease. When compared to healthy subjects, 87 out of 431 metabolites tested were altered in patients with fatal COVID-19 at inclusion, which increased to 162 at last sampling; with fewer metabolites altered in severe and mild cases. Pathway analyses over all severities reflected enrichment of pyrimidine, fructose and mannose and carbon metabolism as well as taste transduction pathways; whereas fatal case features reflected thyroid hormone synthesis and signaling, and purine metabolism pathways. The last time points sampled in fatal cases were marked by an acute reduction of metabolites, with malate and aspartate being the most affected, indicating breakdown of mitochondrial respiration. Targeted lipidomic analyses found most lipids to be upregulated in COVID-19 compared to healthy subjects, with magnitude increasing with severity. Lipids dysregulated over all severities enriched pathways in phosphatidylinositol signaling, inositol phosphate metabolism and long-term depression. Dysregulated lipids in fatal COVID-19 enrich endocannabinoid signaling, bacterial and viral infection and glycerophospholipid metabolism pathways. Interestingly, the serum metabolome did not normalize after recovery. This could be due to residual effects of hyperinflammation, lasting damage to metabolic tissues, a particularity of COVID-19; or a combination of the above. In any case, this indicates that clinical recovery is not dependent on re-establishing metabolic homeostasis.

Taken together the above studies report generalized metabolic depression in COVID-19. Although metabolic diseases were not discussed, it is reasonable to assume that pre-existing dysmetabolism, sub-optimal liver function, impaired glucose homeostasis, dyslipidemia, or any other disequilibrium associated with metabolic disease will adversely affect the metabolic response to COVID-19. The above studies indicate that higher risk of severity in metabolic disease may be due to a lack of metabolic flexibility, as required to efficiently respond to SARS-CoV-2 infection. Indeed, the 80% of COVID-19 cases that are mild or asymptomatic, may be so because of their adaptable metabolism, capable of ramping up or down their substrate production or utilization to cope with pathogen burden.

Several overarching themes can be drawn in the context of these studies. COVID-19 is associated with: i) compromised liver function; ii) dyslipidemia; iii) depression or a depression-like molecular profile; and 4) Altered cellular metabolism. The liver response in releasing alarm molecules is common in severe infection and sepsis, compromised function may arise from high viral loads, congestion due to outstripped filtering capacity, or pre-existing intrahepatic triglycerides that impair liver function. Dyslipidemia may be due to increased lipolysis from a highly inflammatory background, which would be aggravated in T2D where fatty tissues lose sensitivity to insulin's anti-lipolytic effects. Importantly, dyslipidemia and increased lipolysis may also arise due to sickness behaviors that modify energy intake and expenditure, notably a loss of appetite and drop in blood glucose levels causing ketogenesis and reliance on lipid metabolism. Finally, effects were reported on cellular pathways of nucleic acid synthesis, lipid synthesis and oxidative metabolism. The loss of substrates for nucleic acid and lipid synthesis, may be explained by virus-host cell dynamics where viral particles usurp nucleotides for their own replication, and make use of cellular lipids to construct their envelopes. SARS-CoV-2 seems particularly efficient in diverting the use of cellular substrates for its own needs, the authors describe this as a 'hijacking' of host cell metabolic machinery (191). A recent study by Zhang et al. supports such dynamics by demonstrating that intercellular glucose and folate are depleted in SARS-CoV-2 infected cells. This is the result of SARS-CoV-2 infection orienting glucose and folate metabolism towards needs supporting viral replication, destabilizing host mRNA abundance and protein translation processes by its use for viral biosynthesis (194).

To put findings into context, over recent years immuno metabolism research has characterized systemic immunity in metabolic diseases as well as cellular metabolism associated with immune functions. Yet the metabolic demands of viral infection remain relatively understudied. A hallmark publication by Wang et al. in 2017 (130) highlights the systemic metabolism associated with bacterial and viral infection. Whilst both cause similar symptoms or sickness behaviors: loss of appetite, weight loss, fever, lethargy; the energy imbalance and systemic metabolic shift is favourable only in bacterial infection. Of note, fever and marked weight loss are common to SARS-CoV-2, where decreased energy intake is aggravated due to the loss of taste and smell and depression of central reward systems with decreased serotonin reported in the above studies (191, 192, 195). The resulting undernutrition shifts systemic metabolism from glucose-dependence to reliance on ketone bodies and fatty acids, reminiscent of fasting metabolism. This is maladaptive in viral infection, where nutritional supplementation, particularly with glucose, is protective, independently of inflammatory status

or pathogen burden. The systemic metabolic shift in infection is also favorable for viral replication, where free fatty acid substrates are used to form and close virion membranes (196). Whilst all may also apply to COVID-19, the added respiratory depression will also starve the host of oxygen and thus the capacity to maintain efficient oxidative metabolism, largely dependent on lipid-substrates, the mediators of which also sustain lipogenesis.

To date, few specific therapies have proven to be effective against COVID-19 (197, 198). Generally patients receive standard supportive care and antiviral therapies, trials have been ambivalent with regards to alternative therapies including chloroquine, steroids or their related compounds (199). These studies and many others have applied these approaches to classify patient risk, importantly these findings can also be used to redirect trials towards metabolically active therapeutics already applied in metabolic diseases. Analyses from medical databases based around prescription history will also give signs whether lipid lowering, glycogenolytic or anti-hyperglycemic agents may mitigate or aggravate COVID-19, as well as other respiratory infections with a high burden of disease.

## **AUTHOR CONTRIBUTIONS**

AA, FA, RT, FS, KH, KS, JB wrote the manuscript. All authors contributed to the article and approved the submitted version.

#### FUNDING

This work was supported by the Kuwait Foundation for the Advancement of Sciences (KFAS) grant (RA HM-2019-009). FA was supported by the French National Research Agency (Agence Nationale de la Recherche; ANR) ANR-JCJC grant for the MitoFLAME Project ANR-19-CE14-0005. RT was supported by a grant from the European Foundation for the Study of Diabetes (EFSD). FS was supported by Science Foundation Ireland Award 19/FFP/6625. KS is funded by NHMRC investigator grant 2007919.

#### REFERENCES

- 1. International Diabetes Federation. *IDF Diabetes Atlas 2021*, *10th edition*, Belgium: International Diabetes Federation (2021).
- American Diabetes Association. Classification and Diagnosis of Diabetes. Diabetes Care (2017) 40:S11–24. doi: 10.2337/dc17-S005
- Ferlita S, Yegiazaryan A, Noori N, Lal G, Nguyen T, To K, et al. Type 2 Diabetes Mellitus and Altered Immune System Leading to Susceptibility to Pathogens, Especially Mycobacterium Tuberculosis. J Clin Med (2019) 8:2219. doi: 10.3390/jcm8122219
- Klekotka RB, Mizgała E, Król W. The Etiology of Lower Respiratory Tract Infections in People With Diabetes. *Pneumonol Alergol Pol* (2015) 83:401–8. doi: 10.5603/PiAP.2015.0065
- Casqueiro J, Casqueiro J, Alves C. Infections in Patients With Diabetes Mellitus: A Review of Pathogenesis. *Indian J Endocr Metab* (2012) 16:27. doi: 10.4103/2230-8210.94253
- Hine JL, de Lusignan S, Burleigh D, Pathirannehelage S, McGovern A, Gatenby P, et al. Association Between Glycaemic Control and Common Infections in People With Type 2 Diabetes: A Cohort Study. *Diabet Med* (2017) 34:551–7. doi: 10.1111/dme.13205
- Kornum JB, Thomsen RW, Riis A, Lervang HH, Schønheyder HC, Sørensen HT. Type 2 Diabetes and Pneumonia Outcomes. *Diabetes Care* (2007) 30:2251–7. doi: 10.2337/dc06-2417
- Hsia CCW, Raskin P. The Diabetic Lung: Relevance of Alveolar Microangiopathy for the Use of Inhaled Insulin. Am J Med (2005) 118:205–11. doi: 10.1016/j.amjmed.2004.09.019
- Vishwakarma P, Usman K, Garg R, Bajpai J, Sethi R, Pradhan A. Clinical and Radiological Presentations of Various Pulmonary Infections in Hospitalized Diabetes Mellitus Patients: A Prospective, Hospital-Based, Comparative, Case Series Study. *Pulm Med* (2021) 2021:1–8. doi: 10.1155/ 2021/8878746
- Zheng H, Wu J, Jin Z, Yan L-J. Potential Biochemical Mechanisms of Lung Injury in Diabetes. *Aging Dis* (2017) 8:7. doi: 10.14336/AD.2016.0627
- Cardet JC, Ash S, Kusa T, Camargo CA, Israel E. Insulin Resistance Modifies the Association Between Obesity and Current Asthma in Adults. *Eur Respir J* (2016) 48:403–10. doi: 10.1183/13993003.00246-2016
- Al-Rifai RH, Pearson F, Critchley JA, Abu-Raddad LJ. Association Between Diabetes Mellitus and Active Tuberculosis: A Systematic Review and Meta-Analysis. *PloS One* (2017) 12:e0187967. doi: 10.1371/journal.pone.0187967

- Zhu L, She ZG, Cheng X, Qin JJ, Zhang XJ, Cai J, et al. Association of Blood Glucose Control and Outcomes in Patients With COVID-19 and Pre-Existing Type 2 Diabetes. *Cell Metab* (2020) 31:1068–1077.e3. doi: 10.1016/j.cmet.2020.04.021
- Meyer KC. The Role of Immunity in Susceptibility to Respiratory Infection in the Aging Lung. *Respir Physiol* (2001) 128:23–31. doi: 10.1016/S0034-5687(01)00261-4
- Dennis RJ, Maldonado D, Rojas MX, Aschner P, Rondón M, Charry L, et al. Inadequate Glucose Control in Type 2 Diabetes is Associated With Impaired Lung Function and Systemic Inflammation: A Cross-Sectional Study. *BMC Pulm Med* (2010) 10:38. doi: 10.1186/1471-2466-10-38
- Phosat C, Panprathip P, Chumpathat N, Prangthip P, Chantratita N, Soonthornworasiri N, et al. Elevated C-Reactive Protein, Interleukin 6, Tumor Necrosis Factor Alpha and Glycemic Load Associated With Type 2 Diabetes Mellitus in Rural Thais: A Cross-Sectional Study. *BMC Endocr Disord* (2017) 17:44. doi: 10.1186/s12902-017-0189-z
- Reinhold' D, Ansorge' S. Elevated Glucose Levels Stimulate Transforming Growth Factor-131 (TGF-131), Suppress Interleukin 11-2, 11-6 and 11-10 Production and DNA Synthesis in Peripheral Blood Mononuclear Cells. *Horm Metab Res* (1996) 28:267–70.
- Wang C, Xie J, Zhao L, Fei X, Zhang H, Tan Y, et al. Alveolar Macrophage Dysfunction and Cytokine Storm in the Pathogenesis of Two Severe COVID-19 Patients. *EBioMedicine* (2020) 57:102833. doi: 10.1016/ j.ebiom.2020.102833
- Gombart AF, Pierre A, Maggini S. A Review of Micronutrients and the Immune System–Working in Harmony to Reduce the Risk of Infection. *Nutrients* (2020) 12:236. doi: 10.3390/nu12010236
- Baeke F, Takiishi T, Korf H, Gysemans C, Mathieu C. Vitamin D: Modulator of the Immune System. *Curr Opin Pharmacol* (2010) 10:482–96. doi: 10.1016/ j.coph.2010.04.001
- Chung C, Silwal P, Kim I, Modlin RL, Jo E-K. Vitamin D-Cathelicidin Axis: At the Crossroads Between Protective Immunity and Pathological Inflammation During Infection. *Immune Netw* (2020) 20:e12. doi: 10.4110/in.2020.20.e12
- 22. Tahrani AA, Ball A, Shepherd L, Rahim A, Jones AF, Bates A. The Prevalence of Vitamin D Abnormalities in South Asians With Type 2 Diabetes Mellitus in the UK. *Int J Clin Pract* (2010) 64:351–5. doi: 10.1111/j.1742-1241.2009.02221.x
- 23. Dawson-Hughes B, Staten MA, Knowler WC, Nelson J, Vickery EM, LeBlanc ES, et al. Intratrial Exposure to Vitamin D and New-Onset

Diabetes Among Adults With Prediabetes: A Secondary Analysis From the Vitamin D and Type 2 Diabetes (D2d) Study. *Diabetes Care* (2020) 43:2916–22. doi: 10.2337/dc20-1765

- Song Y, Wang L, Pittas AG, Del Gobbo LC, Zhang C, Manson JE, et al. Blood 25-Hydroxy Vitamin D Levels and Incident Type 2 Diabetes. *Diabetes Care* (2013) 36:1422–8. doi: 10.2337/dc12-0962
- 25. Chiu KC, Chu A, Go VLW, Saad MF. Hypovitaminosis D Is Associated With Insulin Resistance and  $\beta$  Cell Dysfunction. *Am J Clin Nutr* (2004) 79:820–5. doi: 10.1093/ajcn/79.5.820
- Isaia G, Giorgino R, Adami S. High Prevalence of Hypovitaminosis D in Female Type 2 Diabetic Population. *Diabetes Care* (2001) 24:1496–6. doi: 10.2337/diacare.24.8.1496
- Plataki MN, Vamvoukaki R, Samonis G, Bikis C, Gorgomiti M, Papadakis JA, et al. Vitamin D and Cathelicidin (LL-37) Status in Patients With Type 2 Diabetes and *Staphylococcus aureus* Nasal Carriage. *Rev Diabetic Stud* (2021) 17:30–7. doi: 10.1900/RDS.2021.17.30
- Ginde AA, Mansbach JM, Camargo CA. Association Between Serum 25-Hydroxyvitamin D Level and Upper Respiratory Tract Infection in the Third National Health and Nutrition Examination Survey. *Arch Intern Med* (2009) 169:384. doi: 10.1001/archinternmed.2008.560
- Hansdottir S, Monick MM, Hinde SL, Lovan N, Look DC, Hunninghake GW. Respiratory Epithelial Cells Convert Inactive Vitamin D to Its Active Form: Potential Effects on Host Defense. *J Immunol* (2008) 181:7090–9. doi: 10.4049/jimmunol.181.10.7090
- Lighter J, Phillips M, Hochman S, Sterling S, Johnson D, Francois F, et al. Obesity in Patients Younger Than 60 Years Is a Risk Factor for COVID-19 Hospital Admission. *Clin Infect Dis* (2020) 71:896–7. doi: 10.1093/cid/ ciaa415
- Richardson S, Hirsch JS, Narasimhan M, Crawford JM, McGinn T, Davidson KW, et al. Presenting Characteristics, Comorbidities, and Outcomes Among 5700 Patients Hospitalized With COVID-19 in the New York City Area. *JAMA* (2020) 323:2052. doi: 10.1001/jama.2020.6775
- Divangahi M, Khan N, Kaufmann E. Beyond Killing Mycobacterium Tuberculosis: Disease Tolerance. Front Immunol (2018) 9:2976. doi: 10.3389/fimmu.2018.02976
- 33. Global Tuberculosis Reports . Available at: https://www.who.int/teams/ control-of-neglected-tropical-diseases/lymphatic-filariasis/morbiditymanagement-and-disability-prevention/global-tuberculosis-programme.
- 34. Barron MM, Shaw KM, Bullard KM, Ali MK, Magee MJ. Diabetes is Associated With Increased Prevalence of Latent Tuberculosis Infection: Findings From the National Health and Nutrition Examination Survey, 2011-2012. *Diabetes Res Clin Pract* (2018) 139:366–79. doi: 10.1016/ j.diabres.2018.03.022
- Baker MA, Harries AD, Jeon CY, Hart JE, Kapur A, Lönnroth K, et al. The Impact of Diabetes on Tuberculosis Treatment Outcomes: A Systematic Review. *BMC Med* (2011) 9:81. doi: 10.1186/1741-7015-9-81
- Workneh MH, Bjune GA, Yimer SA. Prevalence and Associated Factors of Tuberculosis and Diabetes Mellitus Comorbidity: A Systematic Review. *PloS* One (2017) 12:e0175925. doi: 10.1371/journal.pone.0175925
- 37. Kubiak RW, Sarkar S, Horsburgh CR, Roy G, Kratz M, Reshma A, et al. Interaction of Nutritional Status and Diabetes on Active and Latent Tuberculosis: A Cross-Sectional Analysis. *BMC Infect Dis* (2019) 19:627. doi: 10.1186/s12879-019-4244-4
- Aibana O, Acharya X, Huang CC, Becerra MC, Galea JT, Chiang SS, et al. Nutritional Status and Tuberculosis Risk in Adult and Pediatric Household Contacts. *PloS One* (2016) 11:e0166333. doi: 10.1371/journal.pone.0166333
- Chiang CY, Bai KJ, Lin HH, Chien ST, Lee JJ, Enarson DA, et al. The Influence of Diabetes, Glycemic Control, and Diabetes-Related Comorbidities on Pulmonary Tuberculosis. *PloS One* (2015) 10:e0121698. doi: 10.1371/journal.pone.0121698
- Critchley JA, Carey IM, Harris T, DeWilde S, Hosking FJ, Cook DG. Glycemic Control and Risk of Infections Among People With Type 1 or Type 2 Diabetes in a Large Primary Care Cohort Study. *Diabetes Care* (2018) 41:2127–35. doi: 10.2337/dc18-0287
- Zykova SN, Jenssen TG, Berdal M, Olsen R, Myklebust R, Seljelid R. Altered Cytokine and Nitric Oxide Secretion *In Vitro* by Macrophages From Diabetic Type II-Like Db/Db Mice. *Diabetes* (2000) 49:1451–8. doi: 10.2337/diabetes.49.9.1451

- 42. Liu HF, Zhang HJ, Hu QX, Liu XY, Wang ZQ, Fan JY, et al. Altered Polarization, Morphology, and Impaired Innate Immunity Germane to Resident Peritoneal Macrophages in Mice With Long-Term Type 2 Diabetes. J BioMed Biotechnol (2012) 2012:867023. doi: 10.1155/2012/ 867023
- Lachmandas E, Vrieling F, Wilson LG, Joosten SA, Netea MG, Ottenhoff TH, et al. The Effect of Hyperglycaemia on *In Vitro* Cytokine Production and Macrophage Infection With Mycobacterium Tuberculosis. *PloS One* (2015) 10:e0117941. doi: 10.1371/journal.pone.0117941
- 44. Gleeson LE, Sheedy FJ, Palsson-McDermott EM, Triglia D, O'Leary SM, O'Sullivan MP, et al. Cutting Edge: Mycobacterium Tuberculosis Induces Aerobic Glycolysis in Human Alveolar Macrophages That Is Required for Control of Intracellular Bacillary Replication. *J Immunol* (2016) 196:2444–9. doi: 10.4049/jimmunol.1501612
- Huang L, Nazarova EV, Tan S, Liu Y, Russell DG. Growth of Mycobacterium Tuberculosis *In Vivo* Segregates With Host Macrophage Metabolism and Ontogeny. *J Exp Med* (2018) 215:1135–52. doi: 10.1084/jem.20172020
- 46. Kumar NP, Banurekha VV, Nair D, Sridhar R, Kornfeld H, Nutman TB, et al. Coincident Pre-Diabetes is Associated With Dysregulated Cytokine Responses in Pulmonary Tuberculosis. *PloS One* (2014) 9:e112108. doi: 10.1371/journal.pone.0112108
- 47. Restrepo BI, Khan A, Singh VK, de-Leon E, Aguillón-Durán GP, Ledezma-Campos E, et al. Human Monocyte-Derived Macrophage Responses to M. Tuberculosis Differ by the Host's Tuberculosis, Diabetes or Obesity Status, and are Enhanced by Rapamycin. *Tuberculosis (Edinb)* (2021) 126:102047. doi: 10.1016/j.tube.2020.102047
- Martinez N, Ketheesan N, West K, Vallerskog T, Kornfeld H. Impaired Recognition of Mycobacterium Tuberculosis by Alveolar Macrophages From Diabetic Mice. J Infect Dis (2016) 214:1629–37. doi: 10.1093/infdis/ jiw436
- Yamasawa H, Nakayama M, Bando M, Sugiyama Y. Impaired Inflammatory Responses to Multiple Toll-Like Receptor Ligands in Alveolar Macrophages of Streptozotocin-Induced Diabetic Mice. *Inflamm Res* (2012) 61:417–26. doi: 10.1007/s00011-011-0426-2
- Alim MA, Kupz A, Sikder S, Rush C, Govan B, Ketheesan N. Increased Susceptibility to Mycobacterium Tuberculosis Infection in a Diet-Induced Murine Model of Type 2 Diabetes. *Microbes Infect* (2020) 22:303–11. doi: 10.1016/j.micinf.2020.03.004
- Martens GW, Arikan MC, Lee J, Ren F, Greiner D, Kornfeld H. Tuberculosis Susceptibility of Diabetic Mice. *Am J Respir Cell Mol Biol* (2007) 37:518–24. doi: 10.1165/rcmb.2006-0478OC
- Arriaga MB, Rocha MS, Nogueira BMF, Nascimento V, Araújo-Pereira M, Souza AB, et al. The Effect of Diabetes and Prediabetes on Mycobacterium Tuberculosis Transmission to Close Contacts. J Infect Dis (2021) 224:2064– 72. doi: 10.1093/infdis/jiab264
- 53. Magee MJ, Kempker RR, Kipiani M, Gandhi NR, Darchia L, Tukvadze N, et al. Diabetes Mellitus is Associated With Cavities, Smear Grade, and Multidrug-Resistant Tuberculosis in Georgia. *Int J Tuberc Lung Dis* (2015) 19:685–92. doi: 10.5588/ijtld.14.0811
- Martinez N, Vallerskog T, West K, Nunes-Alves C, Lee J, Martens GW, et al. Chromatin Decondensation and T Cell Hyperresponsiveness in Diabetes-Associated Hyperglycemia. *J Immunol* (2014) 193:4457–68. doi: 10.4049/ jimmunol.1401125
- Braverman J, Sogi KM, Benjamin D, Nomura DK, Stanley SA. HIF-1α Is an Essential Mediator of IFN-γ-Dependent Immunity to Mycobacterium Tuberculosis. J Immunol (2016) 197:1287–97. doi: 10.4049/jimmunol.1600266
- Hackett EE, Charles-Messance H, O'Leary SM, Gleeson LE, Muñoz-Wolf N, Case S, et al. Mycobacterium Tuberculosis Limits Host Glycolysis and IL-1β by Restriction of PFK-M *via* MicroRNA-21. *Cell Rep* (2020) 30:124–136.e4. doi: 10.1016/j.celrep.2019.12.015
- 57. Howard NC, Marin ND, Ahmed M, Rosa BA, Martin J, Bambouskova M, et al. Mycobacterium Tuberculosis Carrying a Rifampicin Drug Resistance Mutation Reprograms Macrophage Metabolism Through Cell Wall Lipid Changes. *Nat Microbiol* (2018) 3:1099–108. doi: 10.1038/s41564-018-0245-0
- Freemerman AJ, Johnson AR, Sacks GN, Milner JJ, Kirk EL, Troester MA, et al. Metabolic Reprogramming of Macrophages: Glucose Transporter 1 (GLUT1)-Mediated Glucose Metabolism Drives a Proinflammatory Phenotype. J Biol Chem (2014) 289:7884–96. doi: 10.1074/jbc.M113.522037

- Podell BK, Ackart DF, Kirk NM, Eck SP, Bell C, Basaraba RJ. Non-Diabetic Hyperglycemia Exacerbates Disease Severity in Mycobacterium Tuberculosis Infected Guinea Pigs. *PloS One* (2012) 7:e46824. doi: 10.1371/journal.pone.0046824
- Pérez-Guzmán C, Vargas MH, Quiñonez F, Bazavilvazo N, Aguilar A. A Cholesterol-Rich Diet Accelerates Bacteriologic Sterilization in Pulmonary Tuberculosis. *Chest* (2005) 127:643–51. doi: 10.1378/chest.127.2.643
- 61. Jo YS, Han K, Kim D, Yoo JE, Kim Y, Yang B, et al. Relationship Between Total Cholesterol Level and Tuberculosis Risk in a Nationwide Longitudinal Cohort. *Sci Rep* (2021) 11:16254. doi: 10.1038/s41598-021-95704-1
- Martens GW, Arikan MC, Lee J, Ren F, Vallerskog T, Kornfeld H. Hypercholesterolemia Impairs Immunity to Tuberculosis. *Infect Immun* (2008) 76:3464–72. doi: 10.1128/IAI.00037-08
- Martens GW, Vallerskog T, Kornfeld H. Hypercholesterolemic LDL Receptor-Deficient Mice Mount a Neutrophilic Response to Tuberculosis Despite the Timely Expression of Protective Immunity. *J Leukoc Biol* (2012) 91:849–57. doi: 10.1189/jlb.0311164
- 64. Wipperman MF, Sampson NS, Thomas ST. Pathogen Roid Rage: Cholesterol Utilization by Mycobacterium Tuberculosis. Crit Rev Biochem Mol Biol (2014) 49:269–93. doi: 10.3109/10409238.2014.895700
- Pandey AK, Sassetti CM. Mycobacterial Persistence Requires the Utilization of Host Cholesterol. *Proc Natl Acad Sci U S A* (2008) 105:4376–80. doi: 10.1073/pnas.0711159105
- Russell DG, Cardona P-J, Kim M-J, Allain S, Altare F. Foamy Macrophages and the Progression of the Human Tuberculosis Granuloma. *Nat Immunol* (2009) 10:943–8. doi: 10.1038/ni.1781
- Knight M, Braverman J, Asfaha K, Gronert K, Stanley S. Lipid Droplet Formation in Mycobacterium Tuberculosis Infected Macrophages Requires IFN-γ/HIF-1α Signaling and Supports Host Defense. *PloS Pathog* (2018) 14: e1006874.
- McCaffrey EF, Donato M, Keren L, Chen Z, Delmastro A, Fitzpatrick MB, et al. The Immunoregulatory Landscape of Human Tuberculosis Granulomas. *Nat Immunol* (2022) 23:318–29. doi: 10.1038/s41590-021-01121-x
- Laval T. *Et al. De Novo* Synthesized Polyunsaturated Fatty Acids Operate as Both Host Immunomodulators and Nutrients for Mycobacterium Tuberculosis. *eLife* (2021) 10:e71946.
- Ouimet M, Koster S, Sakowski E, Ramkhelawon B, van Solingen C, Oldebeken S, et al. Mycobacterium Tuberculosis Induces the miR-33 Locus to Reprogram Autophagy and Host Lipid Metabolism. Nat Immunol (2016) 17:677–86. doi: 10.1038/ni.3434
- Dodd CE, Pyle CJ, Glowinski R, Rajaram MVS, Schlesinger LS. CD36-Mediated Uptake of Surfactant Lipids by Human Macrophages Promotes Intracellular Growth of *Mycobacterium Tuberculosis. J. Immunol.* (2016) 197:4727–35.
- Henning LN, Azad AK, Parsa KVL, Crowther JE, Schlesinger LS. Pulmonary Surfactant Protein-A Regulates Toll-Like Receptor Expression and Activity in Human Macrophages. J Immunol. (2009) 180(12):1–28.
- Shivakoti R, Newman JW, Hanna LE, Queiroz ATL, Borkowski K, Gupte AN, et al. Host Lipidome and Tuberculosis Treatment Failure. *Eur Respir J* (2022) 59:2004532. doi: 10.1183/13993003.04532-2020
- Williams KJ, Tabas I. The Response-to-Retention Hypothesis of Early Atherogenesis. Arterioscler Thromb Vasc Biol (1995) 15:551–61. doi: 10.1161/01.ATV.15.5.551
- 75. Ngo MD, Bartlett S, Bielefeldt-Ohmann H, Foo CX, Sinha R, Arachchige BJ, et al. A Blunted GPR183/oxysterol Axis During Dysglycemia Results in Delayed Recruitment of Macrophages to the Lung During M. Tuberculosis Infection. J Infect Dis (2022) 225(12):2219–28. doi: 10.1093/infdis/jiac102
- 76. Vrieling F, Wilson L, Rensen PCN, Walzl G, Ottenhoff THM, Joosten SA. Oxidized Low-Density Lipoprotein (oxLDL) Supports Mycobacterium Tuberculosis Survival in Macrophages by Inducing Lysosomal Dysfunction. *PloS Pathog* (2019) 15:e1007724. doi: 10.1371/journal.ppat.1007724
- 77. Jaimni V, Shasty BA, Madhyastha SP, Shetty GV, Acharya RV, Bekur R, et al. Association of Vitamin D Deficiency and Newly Diagnosed Pulmonary Tuberculosis. *Pulm Med* (2021) 2021:1–6. doi: 10.1155/2021/5285841
- Buonsenso D, Sali M, Pata D, Masiello E, Salerno G, Ceccarelli M, et al. Vitamin D Levels in Active TB, Latent TB, Non-TB Pneumonia and Healthy Children: A Prospective Observational Study. (2018) 37(5):337–47.

- 79. Wang J, Xiong K, Wang Q, Zhao S, Liu Y, Ma A. Adjunctive Vitamin A and D During Pulmonary Tuberculosis Treatment: A Randomized Controlled Trial With a 2 × 2 Factorial Design. *Food Funct* (2020) 11:4672–81. doi: 10.1039/C9FO02751C
- Liu PT, Stenger S, Li H, Wenzel L, Tan BH, Krutzik S, et al. Toll-Like Receptor Triggering of a Vitamin D-Mediated Human Antimicrobial Response. *Science* (2006) 311:5. doi: 10.1126/science.1123933
- Liu PT, Stenger S, Tang DH, Modlin L. Cutting Edge: Vitamin D-Mediated Human Antimicrobial Activity Against Mycobacterium Tuberculosis Is Dependent on the Induction of Cathelicidin. J Immunol (2007) 179 (4):2060–3.
- Zhan Y, Jiang L. Status of Vitamin D, Antimicrobial Peptide Cathelicidin and T Helper-Associated Cytokines in Patients With Diabetes Mellitus and Pulmonary Tuberculosis. *Exp Ther Med* (2015) 9:11–6. doi: 10.3892/ etm.2014.2042
- Christ A, Lauterbach M, Latz E. Western Diet and the Immune System: An Inflammatory Connection. *Immunity* (2019) 51:794–811. doi: 10.1016/ j.immuni.2019.09.020
- 84. Choudhury RP, Edgar L, Rydén M, Fisher EA. Diabetes and Metabolic Drivers of Trained Immunity: New Therapeutic Targets Beyond Glucose. *Arterioscler Thromb Vasc Biol* (2021) 41:1284–90. doi: 10.1161/ ATVBAHA.120.314211
- Christ A, Günther P, Lauterbach MAR, Duewell P, Biswas D, Pelka K, et al. Western Diet Triggers NLRP3-Dependent Innate Immune Reprogramming. *Cell* (2018) 172:162–175.e14. doi: 10.1016/j.cell.2017.12.013
- Edgar L, Akbar N, Braithwaite AT, Krausgruber T, Gallart-Ayala H, Bailey J, et al. Hyperglycemia Induces Trained Immunity in Macrophages and Their Precursors and Promotes Atherosclerosis. *Circulation* (2021) 144:961–82. doi: 10.1161/CIRCULATIONAHA.120.046464
- Chavakis T, Mitroulis I, Hajishengallis G. Hematopoietic Progenitor Cells as Integrative Hubs for Adaptation to and Fine-Tuning of Inflammation. *Nat Immunol* (2019) 20:802–11. doi: 10.1038/s41590-019-0402-5
- Singhal A, Jie L, Kumar P, Hong GS, Leow MKS, Paleja B, et al. Metformin as Adjunct Antituberculosis Therapy. *Sci Transl Med* (2014) 6:263ra159. doi: 10.1126/scitranslmed.3009885
- Kelly B, Tannahill GM, Murphy MP, O'Neill LAJ. Metformin Inhibits the Production of Reactive Oxygen Species From NADH:Ubiquinone Oxidoreductase to Limit Induction of Interleukin-1β (IL-1β) and Boosts Interleukin-10 (IL-10) in Lipopolysaccharide (LPS)-Activated Macrophages. *J Biol Chem* (2015) 290:20348–59. doi: 10.1074/jbc.M115.662114
- Lachmandas E, Eckold C, Böhme J, Koeken VACM, Marzuki MB, Blok B, et al. Metformin Alters Human Host Responses to Mycobacterium Tuberculosis in Healthy Subjects. J Infect Dis (2019) 220:139–50. doi: 10.1093/infdis/jiz064
- Heo E, Kim E, Jang EJ, Lee C-H. The Cumulative Dose-Dependent Effects of Metformin on the Development of Tuberculosis in Patients Newly Diagnosed With Type 2 Diabetes Mellitus. *BMC Pulm Med* (2021) 21:303. doi: 10.1186/s12890-021-01667-4
- 92. Sathkumara HD, Hansen K, Miranda-Hernandez S, Govan B, Rush CM, Henning L, et al. Disparate Effects of Metformin on Mycobacterium Tuberculosis Infection in Diabetic and Nondiabetic Mice. *Antimicrob Agents Chemother* (2020) 65:e01422–20. doi: 10.1128/AAC.01422-20
- Endo A, Kuroda M, Tanzawa K. Competitive Inhibition of 3-Hydroxy-3-Methylglutaryl Coenzyme A Reductase by ML-236A and ML-236B Fungal Metabolites, Having Hypocholesterolemic Activity. *FEBS Lett* (1976) 72:323–6. doi: 10.1016/0014-5793(76)80996-9
- 94. Lai CC, Lee tse M G, Lee SH, Hsu WT, Chang SS, Chen SC, et al. Statin Treatment is Associated With a Decreased Risk of Active Tuberculosis: An Analysis of a Nationally Representative Cohort. *Thorax* (2016) 71:646–51. doi: 10.1136/thoraxjnl-2015-207052
- 95. Davaro F, Forde SD, Garfield M, Jiang Z, Halmen K, Tamburro ND, et al. 3-Hydroxyl-3-Methylglutaryl Coenzyme A (HMG-CoA) Reductase Inhibitor (Statin)-Induced 28-kDa Interleukin-1β Interferes With Mature IL-1β Signaling. J Biol Chem (2014) 289:16214–22. doi: 10.1074/jbc.M114.571505
- Jerwood S, Cohen J. Unexpected Antimicrobial Effect of Statins. J Antimicrob Chemother (2008) 61:362–4. doi: 10.1093/jac/dkm496
- 97. Parihar SP, Guler R, Khutlang R, Lang DM, Hurdayal R, Mhlanga MM, et al. Statin Therapy Reduces the Mycobacterium Tuberculosis Burden in Human

Macrophages and in Mice by Enhancing Autophagy and Phagosome Maturation. *J Infect Dis* (2014) 209:754–63. doi: 10.1093/infdis/jit550

- De Libero G, Mori L. The T-Cell Response to Lipid Antigens of Mycobacterium Tuberculosis. Front Immunol (2014) 5:219. doi: 10.3389/ fimmu.2014.00219
- Goeijenbier M, van Sloten TT, Slobbe L, Mathieu C, van Genderen P, Beyer WEP, et al. Benefits of Flu Vaccination for Persons With Diabetes Mellitus: A Review. Vaccine (2017) 35:5095–101. doi: 10.1016/j.vaccine.2017.07.095
- 100. Stöhr K. Influenza–WHO Cares. Lancet Infect Dis (2002) 2:517. doi: 10.1016/ S1473-3099(02)00366-3
- Nicholson KG, Wood JM, Zambon M. Influenza. Lancet (2003) 362:1733– 45. doi: 10.1016/S0140-6736(03)14854-4
- 102. Morris SK, Parkin P, Science M, Subbarao P, Yau Y, O'Riordan S, et al. A Retrospective Cross-Sectional Study of Risk Factors and Clinical Spectrum of Children Admitted to Hospital With Pandemic H1N1 Influenza as Compared to Influenza A. *BMJ Open* (2012) 2:e000310. doi: 10.1136/ bmjopen-2011-000310
- 103. Nguyen-Van-Tam JS, Openshaw PJM, Hashim A, Gadd EM, Lim WS, Semple MG, et al. Risk Factors for Hospitalisation and Poor Outcome With Pandemic A/H1N1 Influenza: United Kingdom First Wave (May-September 2009). *Thorax* (2010) 65:645–51. doi: 10.1136/thx.2010.135210
- 104. Benfield T, Jensen JS, Nordestgaard BG. Influence of Diabetes and Hyperglycaemia on Infectious Disease Hospitalisation and Outcome. *Diabetologia* (2007) 50:549–54. doi: 10.1007/s00125-006-0570-3
- 105. Muller LMAJ, Gorter KJ, Hak E, Goudzwaard WL, Schellevis FG, Hoepelman AIM, et al. Increased Risk of Common Infections in Patients With Type 1 and Type 2 Diabetes Mellitus. *Clin Infect Dis* (2005) 41:281–8. doi: 10.1086/431587
- 106. Valdez R, Narayan KM, Geiss LS, Engelgau MM. Impact of Diabetes Mellitus on Mortality Associated With Pneumonia and Influenza Among non-Hispanic Black and White US Adults. *Am J Public Health* (1999) 89:1715– 21. doi: 10.2105/AJPH.89.11.1715
- Hulme KD, Gallo LA, Short KR. Influenza Virus and Glycemic Variability in Diabetes: A Killer Combination? *Front Microbiol* (2017) 8:861. doi: 10.3389/ fmicb.2017.00861
- Venkata C, Sampathkumar P, Afessa B. Hospitalized Patients With 2009 H1N1 Influenza Infection: The Mayo Clinic Experience. *Mayo Clin Proc* (2010) 85:798–805. doi: 10.4065/mcp.2010.0166
- Allard R, Leclerc P, Tremblay C, Tannenbaum T-N. Diabetes and the Severity of Pandemic Influenza A (H1N1) Infection. *Diabetes Care* (2010) 33:1491–3. doi: 10.2337/dc09-2215
- Campbell A, Rodin R, Kropp R, Mao Y, Hong Z, Vachon J, et al. Risk of Severe Outcomes Among Patients Admitted to Hospital With Pandemic (H1N1) Influenza. *CMAJ* (2010) 182:349–55. doi: 10.1503/cmaj.091823
- 111. Gilca R, De Serres G, Boulianne N, Ouhoummane N, Papenburg J, Douville-Fradet M, et al. Risk Factors for Hospitalization and Severe Outcomes of 2009 Pandemic H1N1 Influenza in Quebec, Canada. *Influenza Other Respir Viruses* (2011) 5:247–55. doi: 10.1111/j.1750-2659.2011.00204.x
- 112. Lenzi L, Mello ÄMd, Silva LRd, Grochocki MHC, Pontarolo R. Pandemic Influenza A (H1N1) 2009: Risk Factors for Hospitalization. J Bras Pneumol (2012) 38:57–65. doi: 10.1590/S1806-37132012000100009
- 113. Hanslik T, Boelle P-Y, Flahault A. Preliminary Estimation of Risk Factors for Admission to Intensive Care Units and for Death in Patients Infected With A (H1N1)2009 Influenza Virus, France, 2009-2010. *PloS Curr* (2010) 2: RRN1150. doi: 10.1371/currents.RRN1150
- 114. Barrau M, Larrieu S, Cassadou S, Chappert JL, Dussart P, Najioullah F, et al. Hospitalized Cases of Influenza A(H1N1)pdm09 in the French Territories of the Americas, July 2009-March 2010. *Rev Panam Salud Publica* (2012) 32:124–30. doi: 10.1590/S1020-49892012000800006
- 115. Murhekar M, Swamy K, Kumar M, Manickam P, Pandian P, Balaganesakumar S. Risk Factors Associated With Death Among Influenza A (H1N1) Patients, Tamil Nadu, India, 2010. J Postgrad Med (2013) 59:9–14. doi: 10.4103/0022-3859.109481
- 116. Xi X, Xu Y, Jiang L, Li A, Duan J, Du B, et al. Hospitalized Adult Patients With 2009 Influenza A(H1N1) in Beijing, China: Risk Factors for Hospital Mortality. *BMC Infect Dis* (2010) 10:256. doi: 10.1186/1471-2334-10-256
- 117. Cortes Garcia M, Sierra Moros MJ, Santa-Olalla Peralta P, Hernandez-Barrera V, Jimenez-Garcia R, Pachon I, et al. Clinical Characteristics and

Outcomes of Diabetic Patients Who Were Hospitalised With 2009 Pandemic Influenza A H1N1 Infection. *J Infect* (2012) 64:218–24. doi: 10.1016/j.jinf.2011.11.022

- 118. Huo C, Zhang S, Zhang S, Wang M, Qi P, Xiao J, et al. Mice With Type 1 Diabetes Exhibit Increased Susceptibility to Influenza A Virus. *Microb Pathog* (2017) 113:233–41. doi: 10.1016/j.micpath.2017.10.026
- 119. Reading PC, Allison J, Crouch EC, Anders EM. Increased Susceptibility of Diabetic Mice to Influenza Virus Infection: Compromise of Collectin-Mediated Host Defense of the Lung by Glucose? J Virol (1998) 72:6884–7. doi: 10.1128/JVI.72.8.6884-6887.1998
- 120. Wu J, Zhang F, Fang F, Chang H, Wang F, Yang Z, et al. Efficacy of Inactivated Vaccine Against H5N1 Influenza Virus Infection in Mice With Type 1 Diabetes. *Vaccine* (2010) 28:2775-81. doi: 10.1016/ j.vaccine.2010.01.037
- 121. Zhu Q, Chang H, Chen Y, Fang F, Xue C, Zhang F, et al. Protection of Inactivated Influenza Virus Vaccine Against Lethal Influenza Virus Infection in Diabetic Mice. *Biochem Biophys Res Commun* (2005) 329:87–94. doi: 10.1016/j.bbrc.2005.01.109
- 122. Hulme KD, Yan L, Marshall RJ, Bloxham CJ, Upton KR, Hasnain SZ, et al. High Glucose Levels Increase Influenza-Associated Damage to the Pulmonary Epithelial-Endothelial Barrier. *Elife* (2020) 9:e56907. doi: 10.7554/eLife.56907.sa2
- Brownlee M. Biochemistry and Molecular Cell Biology of Diabetic Complications. Nature (2001) 414:813–20. doi: 10.1038/414813a
- 124. Brownlee M, Cerami A. The Biochemistry of the Complications of Diabetes Mellitus. Annu Rev Biochem (1981) 50:385–432. doi: 10.1146/ annurev.bi.50.070181.002125
- 125. Papachristoforou E, Lambadiari V, Maratou E, Makrilakis K. Association of Glycemic Indices (Hyperglycemia, Glucose Variability, and Hypoglycemia) With Oxidative Stress and Diabetic Complications. J Diabetes Res (2020) 2020:7489795. doi: 10.1155/2020/7489795
- 126. Damen MSMA, Alarcon PC, Shah AS, Divanovic S. Greasing the Inflammatory Pathogenesis of Viral Pneumonias in Diabetes. *Obes Rev* (2022) 23(5):e13415. doi: 10.1111/obr.13415
- 127. O'Neill LAJ, Kishton RJ, Rathmell J. A Guide to Immunometabolism for Immunologists. Nat Rev Immunol (2016) 16:553-65. doi: 10.1038/ nri.2016.70
- Makowski L, Chaib M, Rathmell JC. Immunometabolism: From Basic Mechanisms to Translation. *Immunol Rev* (2020) 295:5–14. doi: 10.1111/ imr.12858
- Kohio HP, Adamson AL. Glycolytic Control of Vacuolar-Type ATPase Activity: A Mechanism to Regulate Influenza Viral Infection. Virology (2013) 444:301–9. doi: 10.1016/j.virol.2013.06.026
- 130. Philips BJ, Meguer J-X, Redman J, Baker EH. Factors Determining the Appearance of Glucose in Upper and Lower Respiratory Tract Secretions. *Intensive Care Med* (2003) 29:2204–10. doi: 10.1007/s00134-003-1961-2
- 131. Codo AC, Davanzo GG, Monteiro LB, Souza G, Muraro S, Carregari V, et al. Elevated Glucose Levels Favor SARS-CoV-2 Infection and Monocyte Response Through a HIF-1 $\alpha$ /Glycolysis-Dependent Axis. *Cell Metab* (2020) 32:437-446.e5. doi: 10.2139/ssrn.3606770
- 132. Meza CA, La Favor JD, Kim D-H, Hickner RC. Endothelial Dysfunction: Is There a Hyperglycemia-Induced Imbalance of NOX and NOS? *Int J Mol Sci* (2019) 20:E3775. doi: 10.3390/ijms20153775
- 133. Short KR, Veldhuis Kroeze EJB, Reperant LA, Richard M, Kuiken T. Influenza Virus and Endothelial Cells: A Species Specific Relationship. *Front Microbiol* (2014) 5:653. doi: 10.3389/fmicb.2014.00653
- 134. Teijaro JR, Walsh KB, Cahalan S, Fremgen DM, Roberts E, Scott F, et al. Endothelial Cells are Central Orchestrators of Cytokine Amplification During Influenza Virus Infection. *Cell* (2011) 146:980–91. doi: 10.1016/ j.cell.2011.08.015
- 135. Dhatariya K, Corsino L, Umpierrez GE. Management of Diabetes and Hyperglycemia in Hospitalized Patients. In: KR Feingold, et al, editors. *Endotext* (South Dartmouth (MA)) (2000).
- 136. Marshall RJ, Armart P, Hulme KD, Chew KY, Brown AC, Hansbro PM, et al. Glycemic Variability in Diabetes Increases the Severity of Influenza. *mBio* (2020) 11:e02841–19. doi: 10.1128/mBio.02841-19
- 137. Nyambuya TM, Dludla PV, Mxinwa V, Nkambule BB. T-Cell Activation and Cardiovascular Risk in Adults With Type 2 Diabetes Mellitus: A Systematic

Review and Meta-Analysis. *Clin Immunol* (2020) 210:108313. doi: 10.1016/j.clim.2019.108313

- Giese IM, Schilloks MC, Degroote RL, Weigand M, Renner S, Wolf E, et al. Chronic Hyperglycemia Drives Functional Impairment of Lymphocytes in Diabetic INSC94Y Transgenic Pigs. *Front Immunol* (2021) 11. doi: 10.3389/ fmmu.2020.607473
- 139. Lau EYM, Carroll EC, Callender LA, Hood GA, Berryman V, Pattrick M, et al. Type 2 Diabetes is Associated With the Accumulation of Senescent T Cells. *Clin Exp Immunol* (2019) 197:205–13. doi: 10.1111/cei.13344
- 140. Lee Yho, Kim SR, Han DH, Yu HT, Han YD, Kim JH, et al. Senescent T Cells Predict the Development of Hyperglycemia in Humans. *Diabetes* (2019) 68:156–62. doi: 10.2337/db17-1218
- 141. Recino A, Barkan K, Wong FS, Ladds G, Cooke A, Wallberg M. Hyperglycaemia Does Not Affect Antigen-Specific Activation and Cytolytic Killing by CD8+ T Cells In Vivo. Biosci Rep (2017) 37: BSR20171079. doi: 10.1042/BSR20171079
- 142. Giles A. Metb-08. Increasing Glucose Concentrations is Deleterious to Th1 and Cytotoxic T Cells. *Neuro Oncol* (2016) 18:vi101.
- 143. Diepersloot RJ, Bouter KP, Beyer WE, Hoekstra JB, Masurel N. Humoral Immune Response and Delayed Type Hypersensitivity to Influenza Vaccine in Patients With Diabetes Mellitus. *Diabetologia* (1987) 30:397–401. doi: 10.1007/BF00292541
- 144. van der Pouw Kraan TCTM, Chen WJ, Bunck MCM, van Raalte DH, van der Zijl NJ, van Genugten RE, et al. Metabolic Changes in Type 2 Diabetes are Reflected in Peripheral Blood Cells, Revealing Aberrant Cytotoxicity, a Viral Signature, and Hypoxia Inducible Factor Activity. *BMC Med Genomics* (2015) 8:20. doi: 10.1186/s12920-015-0096-y
- Flu & People With Diabetes | CDC (2021). Available at: https://www.cdc.gov/ flu/highrisk/diabetes.htm.
- 146. Colquhoun AJ, Nicholson KG, Botha JL, Raymond NT. Effectiveness of Influenza Vaccine in Reducing Hospital Admissions in People With Diabetes. *Epidemiol Infect* (1997) 119:335–41. doi: 10.1017/S095026889700825X
- 147. Bechini A, Ninci A, Del Riccio M, Biondi I, Bianchi J, Bonanni P, et al. Impact of Influenza Vaccination on All-Cause Mortality and Hospitalization for Pneumonia in Adults and the Elderly With Diabetes: A Meta-Analysis of Observational Studies. *Vaccines (Basel)* (2020) 8:263. doi: 10.3390/ vaccines8020263
- 148. Kwong JC, Schwartz KL, Campitelli MA, Chung H, Crowcroft NS, Karnauchow T, et al. Acute Myocardial Infarction After Laboratory-Confirmed Influenza Infection. N Engl J Med (2018) 378:345–53. doi: 10.1056/NEJMoa1702090
- 149. Warren-Gash C, Blackburn R, Whitaker H, McMenamin J, Hayward AC. Laboratory-Confirmed Respiratory Infections as Triggers for Acute Myocardial Infarction and Stroke: A Self-Controlled Case Series Analysis of National Linked Datasets From Scotland. *Eur Respir J* (2018) 51:1701794. doi: 10.1183/13993003.01794-2017
- 150. Modin D, Claggett B, Køber L, Schou M, Jensen JUS, Solomon SD, et al. Influenza Vaccination Is Associated With Reduced Cardiovascular Mortality in Adults With Diabetes: A Nationwide Cohort Study. *Diabetes Care* (2020) 43:2226–33. doi: 10.2337/dc20-0229
- 151. Dos Santos G, Tahrat H, Bekkat-Berkani R. Immunogenicity, Safety, and Effectiveness of Seasonal Influenza Vaccination in Patients With Diabetes Mellitus: A Systematic Review. *Hum Vaccin Immunother* (2018) 14:1853–66. doi: 10.1080/21645515.2018.1446719
- 152. Remschmidt C, Wichmann O, Harder T. Vaccines for the Prevention of Seasonal Influenza in Patients With Diabetes: Systematic Review and Meta-Analysis. *BMC Med* (2015) 13:53. doi: 10.1186/s12916-015-0295-6
- 153. Davis TME, Weerarathne T, Foong Y, Mason C, Davis WA. Community-Acquired Infections in Type 2 Diabetic Patients and Their Nondiabetic Partners. The Fremantle Diabetes Study. J Diabetes Complications (2005) 19:259–63. doi: 10.1016/j.jdiacomp.2005.03.003
- 154. Jiménez-Garcia R, Lopez-de-Andres A, Hernandez-Barrera V, Gómez-Campelo P, San Andrés-Rebollo FJ, de Burgos-Lunar C, et al. Influenza Vaccination in People With Type 2 Diabetes, Coverage, Predictors of Uptake, and Perceptions. Result of the MADIABETES Cohort a 7years Follow Up Study. *Vaccine* (2017) 35:101–8. doi: 10.1016/j.vaccine.2016.11.039

- 155. O'Halloran AC, Lu P-J, Williams WW, Bridges CB, Singleton JA. Influenza Vaccination Coverage Among People With High-Risk Conditions in the U.S. *Am J Prev Med* (2016) 50:e15–26. doi: 10.1016/j.amepre.2015.06.008
- 156. Kroneman M, van Essen GA, John Paget W. Influenza Vaccination Coverage and Reasons to Refrain Among High-Risk Persons in Four European Countries. Vaccine (2006) 24:622–8. doi: 10.1016/j.vaccine.2005.08.040
- 157. Ye L, Fang T, Cui J, Zhu G, Ma R, Sun Y, et al. The Intentions to Get Vaccinated Against Influenza and Actual Vaccine Uptake Among Diabetic Patients in Ningbo, China: Identifying Motivators and Barriers. *Hum Vaccin Immunother* (2021) 17:106–18. doi: 10.1080/21645515.2020.1761201
- 158. Chung JR, Kim SS, Kondor RJ, Smith C, Budd AP, Tartof SY, et al. Interim Estimates of 2021-22 Seasonal Influenza Vaccine Effectiveness - United States, February 2022. MMWR Morb Mortal Wkly Rep (2022) 71:365–70. doi: 10.15585/mmwr.mm7110a1
- Dong E, Du H, Gardner L. An Interactive Web-Based Dashboard to Track COVID-19 in Real Time. *Lancet Infect Dis* (2020) 20:533–4. doi: 10.1016/ S1473-3099(20)30120-1
- 160. Wang Y, Zhang D, Du G, Du R, Zhao J, Jin Y, et al. Remdesivir in Adults With Severe COVID-19: A Randomised, Double-Blind, Placebo-Controlled, Multicentre Trial. *Lancet* (2020) 395:1569–78. doi: 10.1016/S0140-6736(20) 31022-9
- 161. Rico-Martín S, Calderón-García JF, Basilio-Fernández B, Clavijo-Chamorro MZ, Sánchez Muñoz-Torrero JF. Metabolic Syndrome and Its Components in Patients With COVID-19: Severe Acute Respiratory Syndrome (SARS) and Mortality. A Systematic Review and Meta-Analysis. J Cardiovasc Dev Dis (2021) 8:162. doi: 10.3390/jcdd8120162
- Lima-Martínez MM, Carrera Boada C, Madera-Silva MD, Marín W, Contreras M. COVID-19 Y Diabetes Mellitus: Una Relación Bidireccional. *Clín Investig Arterioscler* (2021) 33:151–7. doi: 10.1016/j.arteri.2020.10.001
- 163. Lim S, Bae JH, Kwon H-S, Nauck MA. COVID-19 and Diabetes Mellitus: From Pathophysiology to Clinical Management. Nat Rev Endocrinol (2021) 17:11–30. doi: 10.1038/s41574-020-00435-4
- 164. Wan Y, Shang J, Graham R, Baric RS, Li F. Receptor Recognition by the Novel Coronavirus From Wuhan: An Analysis Based on Decade-Long Structural Studies of SARS Coronavirus. J Virol (2020) 94:e00127–20. doi: 10.1128/JVI.00127-20
- 165. Lim HS, MacFadyen RJ, Lip GYH. Diabetes Mellitus, the Renin-Angiotensin-Aldosterone System, and the Heart. Arch Internal Med (2004) 164:1737-48. doi: 10.1001/archinte.164.16.1737
- 166. Ruggenenti P, Cravedi P, Remuzzi G. The RAAS in the Pathogenesis and Treatment of Diabetic Nephropathy. *Nat Rev Nephrol* (2010) 6:319–30. doi: 10.1038/nrneph.2010.58
- 167. Alzaid F, Julla J, Diedisheim M, Potier C, Potier L, Velho G, et al. Monocytopenia, Monocyte Morphological Anomalies and Hyperinflammation Characterise Severe COVID-19 in Type 2 Diabetes. *EMBO Mol Med* (2020) 12:e13038. doi: 10.15252/emmm.202013038
- Gianchandani R, Esfandiari NH, Ang L, Iyengar J, Knotts S, Choksi P, et al. Managing Hyperglycemia in the COVID-19 Inflammatory Storm. *Diabetes* (2020) 69:2048–53. doi: 10.2337/dbi20-0022
- 169. Wu L, Girgis CM, Cheung NW. COVID-19 and Diabetes: Insulin Requirements Parallel Illness Severity in Critically Unwell Patients. *Clin Endocrinol (Oxf)* (2020) 93:390–3. doi: 10.1111/cen.14288
- Eketunde AO, Mellacheruvu SP, Oreoluwa P. A Review of Postmortem Findings in Patients With COVID-19. *Cureus* (2020) 12:e9438. doi: 10.7759/ cureus.9438
- Peiris S, Mesa H, Aysola A, Manivel J, Toledo J, Borges-Sa M, et al. Pathological Findings in Organs and Tissues of Patients With COVID-19: A Systematic Review. *PloS One* (2021) 16:e0250708. doi: 10.1371/ journal.pone.0250708
- 172. Ragab D, Salah Eldin H, Taeimah M, Khattab R, Salem R. The COVID-19 Cytokine Storm; What We Know So Far. *Front Immunol* (2020) 11:1446. doi: 10.3389/fimmu.2020.01446
- 173. Zeng Z, Yu H, Chen H, Qi W, Chen L, Chen G, et al. Longitudinal Changes of Inflammatory Parameters and Their Correlation With Disease Severity and Outcomes in Patients With COVID-19 From Wuhan, China. *Crit Care* (2020) 24:525. doi: 10.1186/s13054-020-03255-0

- 174. Zhang Q, Bastard P, Liu Z, Le Pen J, Moncada-Velez M, Chen J, et al. Inborn Errors of Type I IFN Immunity in Patients With Life-Threatening COVID-19. Science (2020) 370:eabd4570. doi: 10.1126/science.abd4570
- 175. Groop LC, Bonadonna RC, DelPrato S, Ratheiser K, Zyck K, Ferrannini E, et al. Glucose and Free Fatty Acid Metabolism in non-Insulin-Dependent Diabetes Mellitus. Evidence for Multiple Sites of Insulin Resistance. J Clin Invest (1989) 84:205–13. doi: 10.1172/JCI114142
- 176. Atmosudigdo IS, Lim MA, Radi B, Henrina J, Yonas E, Vania R, et al. Dyslipidemia Increases the Risk of Severe COVID-19: A Systematic Review, Meta-Analysis, and Meta-Regression. *Clin Med Insights Endocrinol Diabetes* (2021) 14:1179551421990675. doi: 10.1177/1179551421990675
- 177. Seal KH, Bertenthal D, Carey E, Grunfeld C, Bikle DD, Lu CM. Association of Vitamin D Status and COVID-19-Related Hospitalization and Mortality. *J Gen Intern Med* (2022) 37:853–61. doi: 10.1007/s11606-021-07170-0
- 178. Villasis-Keever MA, López-Alarcón MG, Miranda-Novales G, Zurita-Cruz JN, Barrada-Vázquez AS, González-Ibarra J, et al. Efficacy and Safety of Vitamin D Supplementation to Prevent COVID-19 in Frontline Healthcare Workers. A Randomized Clinical Trial. Arch Med Res (2022) 53:423–30. doi: 10.1016/j.arcmed.2022.04.003
- Oristrell J, Oliva JC, Casado E, Subirana I, Domínguez D, Toloba A, et al. Vitamin D Supplementation and COVID-19 Risk: A Population-Based, Cohort Study. J Endocrinol Invest (2022) 45:167–79. doi: 10.1007/s40618-021-01639-9
- Honardoost M, Ghavideldarestani M, Khamseh ME. Role of Vitamin D in Pathogenesis and Severity of COVID-19 Infection. Arch Physiol Biochem (2020) 1–7. doi: 10.1080/13813455.2020.1792505
- 181. Roth A, Lütke S, Meinberger D, Hermes G, Sengle G, Koch M, et al. LL-37 Fights SARS-CoV-2: The Vitamin D-Inducible Peptide LL-37 Inhibits Binding of SARS-CoV-2 Spike Protein to its Cellular Receptor Angiotensin Converting Enzyme 2 In Vitro. bioRxiv (2020). doi: 10.1101/ 2020.12.02.408153
- Bilezikian JP, Bikle D, Hewison M, Lazaretti-Castro M, Formenti AM, Gupta A, et al. MECHANISMS IN ENDOCRINOLOGY: Vitamin D and COVID-19. Eur J Endocrinol (2020) 183:R133–47. doi: 10.1530/EJE-20-0665
- 183. di Filippo L, Allora A, Doga M, Formenti AM, Locatelli M, Rovere Querini P, et al. Vitamin D Levels Are Associated With Blood Glucose and BMI in COVID-19 Patients, Predicting Disease Severity. J Clin Endocrinol Metab (2022) 107:e348–60. doi: 10.1210/clinem/dgab599
- 184. de Faria Coelho-Ravagnani C, Corgosinho FC, Sanches FLFZ, Prado CMM, Laviano A, Mota JF. Dietary Recommendations During the COVID-19 Pandemic. Nutr Rev (2021) 79:382–93. doi: 10.1093/nutrit/nuaa067
- 185. McNeill JN, Lau ES, Paniagua SM, Liu EE, Wang JK, Bassett IV, et al. The Role of Obesity in Inflammatory Markers in COVID-19 Patients. *Obes Res Clin Pract* (2021) 15:96–9. doi: 10.1016/j.orcp.2020.12.004
- 186. Marfella R, D'Onofrio N, Sardu C, Scisciola L, Maggi P, Coppola N, et al. Does Poor Glycaemic Control Affect the Immunogenicity of the COVID-19 Vaccination in Patients With Type 2 Diabetes: The CAVEAT Study. *Diabetes Obes Metab* (2022) 24:160–5. doi: 10.1111/dom.14547
- 187. Papadokostaki E, Tentolouris A, Anastasiou IA, Psichogiou M, Iliaki E, Eleftheriadou I, et al. Immunogenicity of SARS-CoV-2 BNT162b2 Vaccine in People With Diabetes: A Prospective Observational Study. Vaccines (Basel) (2022) 10:382. doi: 10.3390/vaccines10030382
- 188. Kan C, Zhang Y, Han F, Xu Q, Ye T, Hou N, et al. Mortality Risk of Antidiabetic Agents for Type 2 Diabetes With COVID-19: A Systematic Review and Meta-Analysis. Front Endocrinol (2021) 12. doi: 10.3389/fendo.2021.708494
- 189. Kow CS, Hasan SS. The Association Between the Use of Statins and Clinical Outcomes in Patients With COVID-19: A Systematic Review and Meta-

Analysis. Am J Cardiovasc Drugs (2021) 22:167–181. doi: 10.1007/s40256-021-00490-w

- 190. Makar AB, McMartin KE, Palese M, Tephly TR. Association Between Statin Use and Poor Outcomes in COVID-19 Patients With Diabetes Mellitus: A Systematic Review. *Curr Diabetes Rev* (2022) 13(2):117–26. doi: 10.2174/ 1573399818666220209095056
- 191. Shen B, Yi X, Sun Y, Bi X, Du J, Zhang C, et al. Proteomic and Metabolomic Characterization of COVID-19 Patient Sera. *Cell* (2020) 182:59–72.e15.
- 192. Wu D, Shu T, Yang X, Song JX, Zhang M, Yao C, et al. Plasma Metabolomic and Lipidomic Alterations Associated With COVID-19. *Natl Sci Rev* (2020) 7:1157–68. doi: 10.1093/nsr/nwaa086
- 193. Yan B, Chu H, Yang D, Sze KH, Lai PM, Yuan S, et al. Characterization of the Lipidomic Profile of Human Coronavirus-Infected Cells: Implications for Lipid Metabolism Remodeling Upon Coronavirus Replication. *Viruses* (2019) 11:E73. doi: 10.3390/v11010073
- 194. Zhang Y, Guo R, Kim SH, Shah H, Zhang S, Liang JH, et al. SARS-CoV-2 Hijacks Folate and One-Carbon Metabolism for Viral Replication. *Nat Commun* (2021) 12:1676. doi: 10.1038/s41467-021-21903-z
- 195. Spinato G, Fabbris C, Polesel J, Cazzador D, Borsetto D, Hopkins C, et al. Alterations in Smell or Taste in Mildly Symptomatic Outpatients With SARS-CoV-2 Infection. JAMA (2020) 323:2089–90. doi: 10.1001/ jama.2020.6771
- 196. Greseth MD, Traktman P. *De Novo* Fatty Acid Biosynthesis Contributes Significantly to Establishment of a Bioenergetically Favorable Environment for Vaccinia Virus Infection. *PloS Pathog* (2014) 10:e1004021. doi: 10.1371/ journal.ppat.1004021
- 197. Wen W, Chen C, Tang J, Wang C, Zhou M, Cheng Y, et al. Efficacy and Safety of Three New Oral Antiviral Treatment (Molnupiravir, Fluvoxamine and Paxlovid) for COVID-19: A Meta-Analysis. Ann Med (2022) 54:516–23. doi: 10.1080/07853890.2022.2034936
- Drożdżal S, Rosik J, Lechowicz K, Machaj F, Szostak B, Przybyciński J, et al. An Update on Drugs With Therapeutic Potential for SARS-CoV-2 (COVID-19) Treatment. Drug Resist Updat (2021) 59:100794. doi: 10.1016/j.drup.2021.100794
- 199. Mehra MR, Desai SS, Kuy S, Henry TD, Patel AN. Retraction: Cardiovascular Disease, Drug Therapy, and Mortality in Covid-19. N Engl J Med (2020) 382:2582. doi: 10.1056/NEJMoa2007621

Conflict of Interest: KS is a consultant for Sanofi, Roche and NovoNordisk.

The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

**Publisher's Note:** All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2022 Al-Sayyar, Hulme, Thibaut, Bayry, Sheedy, Short and Alzaid. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.