



Synthesis and Biological Evaluation of Thio-Derivatives of 2-Hydroxy-1, 4-Naphthoquinone (Lawsone) as Novel Antiplatelet Agents

Matías Monroy-Cárdenas^{1†}, Diego Méndez^{2†}, Andrés Trostchansky³, Maximiliano Martínez-Cifuentes⁴, Ramiro Araya-Maturana^{1*} and Eduardo Fuentes^{2*}

OPEN ACCESS

Edited by:

Simone Brogi, University of Pisa, Italy

Reviewed by:

Jean Jacques Vanden Eynde, University of Mons, Belgium Masakazu Atobe, Asahi Kasei, Japan Assem Barakat, King Saud University, Saudi Arabia

*Correspondence:

Ramiro Araya-Maturana raraya@utalca.cl Eduardo Fuentes edfuentes@utalca.cl

[†]These authors have contributed equally to this work

Specialty section:

This article was submitted to Medicinal and Pharmaceutical Chemistry, a section of the journal Frontiers in Chemistry

Received: 21 April 2020 Accepted: 25 May 2020 Published: 04 August 2020

Citation:

Monroy-Cárdenas M, Méndez D, Trostchansky A, Martínez-Cifuentes M, Araya-Maturana R and Fuentes E (2020) Synthesis and Biological Evaluation of Thio-Derivatives of 2-Hydroxy-1,4-Naphthoquinone (Lawsone) as Novel Antiplatelet Agents. Front. Chem. 8:533. doi: 10.3389/fchem.2020.00533 ¹ Instituto de Química de Recursos Naturales, Programa de Investigación Asociativa en Cáncer Gástrico (PIA-CG), Universidad de Talca, Talca, Chile, ² Department of Clinical Biochemistry and Immunohaematology, Thrombosis Research Center, Medical Technology School, Faculty of Health Sciences, Universidad de Talca, Talca, Chile, ³ Departamento de Bioquimica and Center for Free Radical and Biomedical Research, Facultad de Medicina, Universidad de la República, Montevideo, Uruguay, ⁴ Centro Integrativo de Biología y Química Aplicada (CIBQA), Escuela de Tecnología Médica, Facultad de Salud, Universidad Bernardo O'Higgins, Santiago, Chile

We designed and synthesized in water, using conventional heating and microwave irradiation, new thio-derivatives of 2-hydroxy-1,4-naphthoquinone, a naturally occurring pigment known as lawsone or hennotannic acid, thus improving their antiplatelet activity with relevance to their potential future use in thrombus formation treatment. The structure-activity relationship showed that the thiophenyl moiety enhances the antiplatelet activity. Moreover, the position and nature of the substituent at the phenyl ring have a key effect on the observed biological activity. Compound **4** (2-((4-bromophenyl)thio)-3-hydroxynaphthalene-1,4-dione) was the most active derivative, presenting IC₅₀ values for platelet aggregation inhibition of 15.03 \pm 1.52 μ M for TRAP-6, and 5.58 \pm 1.01 μ M for collagen. Importantly, no cytotoxicity was observed. Finally, we discussed the structure-activity relationships of these new lawsone thio-derivatives on inhibition of TRAP-6- and collagen-induced platelet aggregation.

Keywords: lawsone, naphthoquinone, small compounds, platelet, thrombosis

INTRODUCTION

Cardiovascular diseases (CVD), one of the leading causes of deaths from non-communicable disease in the world, all have thrombus formation as a common process, with platelet activation at the dysfunctional vessel wall being one of the main steps during thrombosis (ISTH Steering Committee for World Thrombosis Day, 2014; Diamond, 2016).

Platelets are fragments of megakaryocytes circulating within the bloodstream aiding in maintaining hemostasis with both blood components and endothelial cells (Paes et al., 2019). Although platelets have a normal function in primary hemostasis, increased platelet activation has been widely recognized as a key player in the initiation of thrombotic events (Franco et al., 2015). An increase in reactive oxygen species (ROS), [e.g., superoxide and hydrogen peroxide production at the mitochondrial electron transport chain] has been associated with platelet activation

1

(Geisler et al., 2007; Becatti et al., 2013; Caruso et al., 2015). On the other hand, it has been observed that some guinone and hydroquinone compounds protect mitochondrial damage and platelet activation generated by ROS (Kim et al., 2001; Cocheme et al., 2007); in this way, some structure-activity relationship studies with quinone derivatives have been reported (Chang et al., 1997; Cho et al., 1997; Kim et al., 2001). However, given the variability of the mechanisms of action of guinoneshydroquinones it has been suggested that there is a low selectivity on their antiplatelet activity (Bolibrukh et al., 2015; Chang et al., 2019; Mendez et al., 2020). Recent work has shown conflicting results (Fuentes et al., 2018). In this way, 3'-methoxyflavone quinone inhibited arachidonic acid-induced platelet aggregation at low micromolar concentration with an IC_{50} of $\sim 10\,\mu M$ (Chang et al., 2000). Moreover, the presence of OH groups in addition to their methylation or glycosylation in anthraquinones increases the antiplatelet activity (Yun-Choi et al., 1990; Xu et al., 2014). The available drugs with therapeutic ability to regulate platelet activation are still scarce, thus this research area is the high relevance (Mousa, 1999; Moura et al., 2016).

Lawsone [2- Hydroxy-1,4-naphthoquinone, (1)] or hennotannic acid, is a natural compound from the naphthoquinones family and can be obtained from leaves of henna (Lawsonia spp., family Lythraceae) as a red-orange pigment. The structure of Lawsone (1) has served for the development of new bioactive compounds and its scaffold is present in compounds with several valuable biological activities such as lapachol (2), atovaquone (3), parvaquone (4), and NQ1 (2-OH-3-(2-methyl-trifluorooctyl)-naphthoquinone) (5) (Figure 1) (Jordão et al., 2015). Lawsone (1) and its synthetic derivatives have shown promising biological results with antibacterial, antifungal, antitumor, and antiparasitic effects (Rahmoun et al., 2012; Barani et al., 2018; Al Nasr et al., 2019), their use, or study as antiplatelet agents have not been described, although the antiplatelet activity of other quinones has been reported (Fuentes et al., 2018). The current work aims to evaluate the antiplatelet effects of a series of lawsone thio-derivatives, which were obtained using on-water methodology, which was optimized by microwave irradiation for those cases that yielded poor results with conventional heating.

MATERIALS AND METHODS

Chemicals, Reagents, General Procedures, and Apparatus

¹H and ¹³C NMR spectra were obtained from a spectrometer operating at either 400.13 MHz (¹H) or 100.61 MHz (¹³C). Measurements were carried out at 300°K. Chemical shifts are reported as ppm downfield from TMS for ¹H NMR. ¹³C NMR spectra were recorded with the NMR spectrometer calibrated with CDCl₃. All melting points are uncorrected and were determined using an Electrothermal 9100 apparatus. IR spectra (KBr discs) were recorded on an FT-IR spectrophotometer; wave numbers are reported in cm⁻¹. High-resolution mass spectra were obtained on an orthogonal time-of-flight (TOF) mass spectrometer (QTOF Micro, Micromass UK) or a Q Exactive

Focus (Thermo Scientific, USA). Silica gel 60 (230–400 mesh ASTM) and TLC aluminum sheets silica gel 60 F254 were used for flash-column chromatography and analytical TLC, respectively (Urra et al., 2016; Mendez et al., 2020).

Synthesis of Compounds

General Procedure Using Conventional Heating

A mixture of 2.87 mmol of lawsone and 1.44 mmol of the respective thiol in 15 mL of water was heated at 50°C overnight, and then the mixture was extracted with ethyl acetate ($30 \text{ mL} \times 3$) the organic phase was dried with anhydrous sodium sulfate and then evaporated under a vacuum.

General Procedure With Microwave Irradiation

In a 100 mL ACE pressure tube with a magnetic stir bar, a mixture of 1.44 mmol of lawsone and 0.72 mmol of the respective thiol in water (6 mL) was heated at 50°C for 20 min under microwave irradiation (CEM Discover, Matthews, NC, USA). Then the mixture was extracted with ethyl acetate (30 mL \times 3), and the organic phase dried with anhydrous sodium sulfate and evaporated under a vacuum (Tandon and Maurya, 2009; Tandon et al., 2010). Afterward, lawsone thioderivatives **1–8**, were purified by flash chromatography with ethyl acetate:hexane:methanol 5:2:2 (**Table 1**).

2-Hydroxy-3-(phenylthio)naphthalene-1,4-dione (1) (Tandon and Maurya, 2009)

By reaction of lawsone with benzenethiol compound **1** was obtained. ¹H NMR δ (DMSO-d6): 7.2-7.3 (m, 5H), 7.9 (d, J = 7.3 Hz, 1H), 8.0 (t, J = 7.3 Hz, 2H), 8.1 (t, J = 7.4 Hz, 1H).

2-((2-bromophenyl)thio)-3-hydroxynaphthalene-1,4-dione (2)

By reaction of lawsone with 2-bromobenzenethiol, compound **2** was obtained: red solid; mp 270.8 (d); ¹H NMR δ (DMSO-d6): 6.71 (t, J = 7.4 Hz, 1H), 6.78 (d, J = 7.6 Hz, 1H), 6.86 (t, J = 7.4 Hz, 1H), 7.28 (d, J = 7.9 Hz, 1H), 7.49 (t, J = 7.4 Hz, 1H), 7.65 (t, J = 7.6 Hz, 1H), 7.86 (d, J = 7.6 Hz, 1H), 8.05 (d, J = 7.6 Hz, 1H); ¹³C NMR δ (DMSO-d6): 105.51, 122.32, 124.45, 126.28, 126.41, 126.66, 127.64, 129.87, 131.19, 134.60, 136.05, 143.29, 172.94, 180.09, 186.86; IR (KBr, cm⁻¹): 3426, 2919, 2850, 1712, 1656, 1646, 1583, 1531, 1276, 744; HRMS (ESI) m/z calculated. for C₁₆H₉BrO₃S M⁺: 359.9456, found: 359.9459.

2-((3-bromophenyl)thio)-3-hydroxynaphthalene-1,4-dione (3)

By reaction of lawsone with 3-bromobenzenethiol, compound **3** was obtained: red solid; mp 231.3°C (d); ¹H NMR δ (DMSO-d6): 6.92 (t, J = 7.8 Hz, 1H), 6.99 (ddd, J = 8.1 Hz, $J_2 = J_3 = 1.6$ Hz, 1H), 7.02 (ddd $J_1 = 7.6$ Hz, $J_2 = J_3 = 1.2$ Hz, 1H), 7.10 (dd, $J_1 = J_2 = 1.6$ Hz, 1H), 7.52 (td, $J_1 = 7.4$ Hz $J_2 = 1$ Hz, 1H), 7.65 (td, $J_1 = 7.6$ Hz $J_2 = 1$ Hz, 1H), 7.76 (s,1H), 7.89 (d, J = 7.6 Hz, 1H), 8.04 (dd, $J_1 = 7.6$ Hz, $J_2 = 1$ Hz, 1H); ¹³C NMR δ (DMSO-d6): 107.34, 116.94, 126.41, 126.94, 127.65, 130.57, 131.12, 131.46, 135.12, 135.72, 138.99, 180.75, 188.11; IR (KBr, cm⁻¹): 3428, 2919, 2850, 1664, 1587, 1529, 1276, 738; HRMS (ESI) m/z calculated. for C₁₆H₉BrO₃S M⁺: 359.9456, found: 359.9474.



TABLE 1 | Synthesis of lawsone thio-derivatives.

	O O O H	x on water	OH X + OH OH OH		
Compound	x		Yield (%)		
		Conventional heating	Microwave irradiation		
1	Н	64	98		
2	o-Br	26	36		
3	<i>m</i> -Br	34	71		
4	p-Br	8	18		
5	o-CO ₂ Me	8	20		
6	0-F	84	-		
7	<i>m</i> -F	84	-		
8	p-F	86	-		

2-((4-Bromophenyl)thio)-3-hydroxynaphthalene-1,4-dione (4)

By reaction of lawsone with 4-bromobenzenethiol, compound 4 was obtained: red solid; mp 271.2°C (d); ¹H RMN δ (DMSOd6): 6.95 (d, J = 8.2 Hz, 2H), 7.08 (d, J = 8.2 Hz, 2H), 7.50 (t, J = 7.4 Hz, 1H), 7.53 (s, 1H), 7.66 (t, J = 7.5 Hz, 1H), 7.86 (d, J = 7.4 Hz, 1H), 8.04 (d, J = 7.6 Hz, 1H); ¹³C RMN δ (DMSO-d6): 116.95, 126.41, 126.95, 127.66, 130.57, 131.12, 131.46, 135.12, 135.71, 138.93, 180.77, 188.14; IR (KBr, cm⁻¹): 3430, 2919, 2850, 1643, 1585, 1536, 1274, 740; HRMS (ESI) m/z calcd. for C₁₆H₉BrO₃S M⁺: 359.9456, found: 359.9479.

Methyl 2-((3-hydroxy-1,4-dioxo-1,4-dihydronaphthalen-2-yl)thio)benzoate (5)

By reaction of lawsone with methyl thiosalicylate, compound **5** was obtained: red solid; mp 235°C (d); ¹H NMR δ (DMSO-d6): 3.84 (s, 3H, CH₃), 6.95 (t, J = 7.4 Hz, 1H), 7.04 (d, J = 8.1 Hz, 1H), 7.15 (t, J = 7.4, 1H), 7.57 (t, J = 7.5 Hz, 1H), 7.70 (t,

 $J = 7.5 \text{ Hz}, 1\text{H}), 7.80(\text{d}, J = 7.8 \text{ Hz}, 1\text{H}), 7.91(\text{d}, J = 7.6 \text{ Hz}, 1\text{H}), 8.00(\text{d}, J = 7.6 \text{ Hz}, 1\text{H}), 8.07(\text{s}, 1\text{H});^{13}\text{C} \text{ NMR } \delta \text{ (DMSO-d6)}: 51.94, 105.71, 122.66, 126.13, 126.15, 126.17, 126.52, 130.94, 131.20, 131.47, 134.49, 136.27, 144.90, 166.70, 173.26, 179.86, 186.42; IR (KBr, cm⁻¹): 3428, 2919, 2850, 1706, 1662, 1587, 1531, 1276, 1251; HRMS (ESI) m/z calcd. for <math>C_{18}H_{12}O_5S$: 340.0405, found: 340.0392.

2-((2-Fluorophenyl)thio)-3-hydroxynaphthalene-1,4-dione (6)

By reaction of lawsone with 2-fluorobenzenethiol, compound **6** was obtained: red solid; mp 260.8°C (d); ¹H NMR δ (DMSO-d6): 6.61–6.64 (m, 1H), 6.70-6.80 (m, 2H), 6.84 (t, *J* = 7.5 Hz, 1H), 7.43-7.51 (m, 1H), 7.62 (t, *J* = 7.5 Hz, 1H), 7.84 (d, *J* = 7.5 Hz, 1H), 8.03(d, J = 7.5 Hz, 1 H); ¹³C NMR δ (DMSO-d6): 105.00, 114.62 (d, *J* = 21.8 Hz), 124.05 (d, *J* = 0.9 Hz), 125.08 (d, *J* = 7.3 Hz), 126.39, 126.47, 126.70, 127.68, 130.93, 131.21, 134.68, 135.80, 159.22 (d, *J* = 241.2 Hz), 172.96, 180.70, 187.31;

IR (KBr, cm⁻¹): 3428, 2917, 2850, 1673, 1587, 1533, 1469, 1276, 1213; HRMS (ESI) m/z calcd. for $C_{16}H_9BrO_3S$: 300.0256, found: 300.0270.

2-((3-Fluorophenyl)thio)-3-hydroxynaphthalene-1,4dione (7)

By reaction of lawsone with 3-fluorobenzenethiol, compound 7 was obtained: red solid; mp 268.7°C(d); ¹H NMR δ (DMSO-d6): 6.50-6.77(m, 2H), 6.82 (t, J = 7.0 Hz, 1H), 7.34 (s, 1H), 7.46(t, J = 6.9 Hz, 1H), 7.63 (t, J = 7.3 Hz, 1H), 7.84 (d, J = 7.0Hz, 1H), 8.04 (d, J = 7.3 Hz, 1H); ¹³C NMR δ (DMSO-d6): 102.26, 114.87 (d, J = 21.1 Hz), 124.55 (d, J = 2.9 Hz), 124.69 (d, J = 7.3 Hz), 126.21, 126.45, 127.14 (d, J = 2.9 Hz), 127.95 (d, J = 16 Hz), 131.37, 131.61, 134.75, 136.35, 158.92 (d, J = 239.8 Hz), 173.29, 179.13, 185.73; IR (KBr, cm⁻¹): 3430, 2360, 2339, 1664, 1587, 1533, 1469, 1278, 1222; HRMS (ESI) m/z calculated. for C₁₆H₉FO₃S: 300.0256, found: 300.0233.

2-((4-Fluorophenyl)thio)-3-hydroxynaphthalene-1,4-dione (8)

By reaction of lawsone with 4-fluorobenzenethiol, compound **8** was obtained: red solid; mp 268.0°C (d); ¹H NMR δ (DMSO-d6): 6.76 (t, *J* = 8.6 Hz, 2H), 7.03 (dd, *J*₁ = 8 Hz, *J*₂ = 5.5 Hz, 1H), 7.50(t, *J* = 7.4 Hz, 1H), 7.63(t, *J* = 7.5 Hz, 1H), 7.79(s, 1H), 7.88(d, *J* = 7.6 Hz, 1H), 8.0(d, *J* = 7.6 Hz, 1H); ¹³C NMR δ (DMSO-d6): 106.91, 115.14, (d, *J* = 21.80 Hz), 126.09, 126.54, 127.40 (d, *J* = 7.3 HZ), 130.98, 131.29 (d, *J* = 13.8 Hz), 134.32, 135.51, 136.18, 159.98 (d, *J* = 241.2 Hz), 172.83, 179.97, 186.60; IR (KBr, cm⁻¹): 3424, 2919, 2850, 2358, 1661, 1587, 1526, 1381, 1273, 1221; HRMS (ESI) m/z calculated. for C₁₆H₉FO₃S: 300.0256, found: 300.0262.

Human Washed Platelets

Platelets were obtained from citrated whole blood of healthy volunteers (two weeks drugs-free) with written informed consent, as previously described (Fuentes et al., 2014a).

Lactate Dehydrogenase (LDH)-Based Cytotoxicity Assay

Washed platelets $(200 \times 10^9 \text{ platelets/L})$ were incubated for 10 min at 37°C with DMSO (vehicle) 0.2% or 100 μ M of compounds. Then, platelets are centrifuged (800 × g for 8 min), and 50 μ L aliquots of the resulting supernatant analyzed by the LDH assay (Cayman Chemical, USA). The reported results correspond to the percentage of total enzyme activity from a control incubation lysed with 0.3% Triton X-100 (Kim et al., 2009; Mendez et al., 2020).

Platelet Aggregation

Platelet aggregation was evaluated using a lumi-aggregometer (Chrono-Log, Haverton, PA, USA) and monitored by light transmission according to Fuentes et al. (Fuentes et al., 2014a; Mendez et al., 2020). Briefly, washed platelets (200×10^9 platelets/L) were pre-incubated for 5 min at 37° C and pH 7.4 with vehicle or different doses of compounds; then platelet aggregation was induced by TRAP-6 (5μ M) or collagen (1μ g/mL) and measured for 5 min at 37° C while stirring.

Flow Cytometry Study

Externalization of phosphatidylserine, glycoprotein (GP) IIb/IIIa activation, P-selectin expression, CD63 release, and ROS levels on platelets were determined by flow cytometry (Ritchie et al., 2000; Fuentes et al., 2014b; Walsh et al., 2014). All results were analyzed in the Accuri C6 flow cytometer (BD, Biosciences, USA).

Statistical Analysis

The data obtained were presented as a mean \pm standard deviation (*SD*) of three independent experiments and analyzed using Prism 8.3 software (GraphPad Inc., San Diego CA, USA). The half-maximal inhibitory concentration (IC₅₀) was calculated from dose-response curves. Differences between samples were analyzed using Tukey or paired t-test (Vaiyapuri et al., 2013). *P*-values < 0.05 were considered as significant (Fuentes et al., 2014b; Munoz-Gutierrez et al., 2017).

RESULTS AND DISCUSSION

Synthesis

Several attempts to reproduce the reported yield in the reaction of unsubstituted benzenethiol with lawsone were unsuccessful, even raising the described time to overnight. However, the reaction with fluoro-substituted benzenethiols gave good yields,

 TABLE 2 | Cytotoxic activity of thio-derivatives of 2-hydroxy-1,4-naphthoquinone (lawsone) on washed platelets.

Compound	Cytotoxic activity Mean (%) \pm <i>SD</i>
Lawsone	32.80 ± 3.13**
1	22.35 ± 2.94
2	14.72 ± 10.92
3	36.51 ± 4.23***
4	22.38 ± 5.78
5	$31.12 \pm 3.06^{**}$
6	21.77 ± 2.37
7	21.50 ± 2.37
8	21.62 ± 4.17
Vehicle (DMSO 0.2%)	12.49 ± 2.69

p < 0.01 and *p < 0.001 analyzed by Tukey test.

TABLE 3 | Inhibition of platelet aggregation by thio-derivatives of 2-hydroxy-1,4-naphthoquinone (lawsone) on TRAP-6- or collagen-stimulated platelets.

Compound	IC_{50} (μ M) ± SD		
	TRAP-6 (5 μM)	Collagen (1 μg/mL)	
1	155.40±55.20	14.58 ± 5.43	
2	111.63 ± 17.53	11.17 ± 3.90	
4	15.03 ± 1.52	5.58 ± 1.01	
6	> 500	10.59 ± 2.25	
7	134.16 ± 17.34	11.13 ± 1.69	
8	107.56 ± 52.98	12.05 ± 1.21	

contrary to Bromo substituted derivatives which gave very low yields. These results show a correlation between yields of the reaction and the solubility of benzenethiols in water. To improve the low yields obtained with the less soluble thiols, microwave irradiation, instead of conventional heating was used, in this way better yields and very reduced reaction times were obtained. The structural assignment of the pure new products relied on the disappearing of the quinone proton of lawsone and the signals reported by the aromatic ring of the thiols, besides high-resolution mass spectrometry. ¹H and ¹³C NMR spectrum of each compound were included as **Supplementary Information**.

Cytotoxicity Activity on Platelets

Before analyzing the potential antiplatelet activity of the synthesized compounds, we evaluated the cytotoxic activity that lawsone and its derivative may have on platelets. After incubating platelets for 10 min at 37°C with each compound, cytotoxicity detection by LDH leakage showed that compounds **1**, **2**, **4**, **6**, 7, and **8** at the higher concentration tested (100 μ M) did not exert cytotoxicity on platelets (**Table 2**). Quinones represent a class of organic compounds that have previously shown that through

quite complex mechanisms can have certain levels of cytotoxicity (Bolton et al., 2000; Bolton and Dunlap, 2017). Quinone-based compounds have been regarded as among the most troublesome of all pan-assay interference compounds (PAINS) chemotypes, limiting their medical potential (Baell and Nissink, 2018; Qin et al., 2018). Alternatively, quinones are highly redox-active molecules which can generate oxidative damage in intracellular macromolecules such as lipids, proteins, and DNA (Bolton et al., 2000). In this study, we showed that lawsone and compounds **3** and **5** had a cytotoxic activity with an increase of LDH released by damaged platelets.

Antiplatelet Activity of New Thio-Derivatives of 2-Hydroxy-1, 4-Naphthoquinone

Antiplatelet therapy is called the treatment for the prevention of platelet activation-associated thrombosis (Harrington et al., 2003). Although aspirin and clopidogrel have been the cornerstone in the management of platelet regulation, a high number of patients continued suffering recurrent thrombotic events (Fox et al., 2004; Price et al., 2011). Since the available



Frontiers in Chemistry | www.frontiersin.org

therapeutic drugs to control non-desired platelet activation remains limited, the search for new antiplatelet agents is necessary (Mousa, 1999; Moura et al., 2016). In this regard, some structure-activity relationship studies with quinone derivatives have shown that the variability of inhibition of platelet activation by these type of compounds is governed by the nature of the substituent (Kim et al., 2001; Fuentes et al., 2018), as explained below: (i) The oxidation of the B-ring of hydroxyl-methoxyisoflavone to methoxyflavone quinone, and the conversion of flavone and isoflavones into their corresponding flavanguinone or isoflavanguinones, result in an increase and potent antiplatelet activity (Chang et al., 2000). (ii) The inhibition of platelet aggregation by 2-alkoxy-1,4-naphthoquinone increases with 2-alkoxy chain length (Lien et al., 2002). (iii) The inclusion of a free amino group in the thiosulfonate moiety of quinone (4-aminobenzenesulfonothioic acid S-[(9,10-dihydro-1,4dimethoxy-9,10-dioxo2-anthracenyl)methyl] ester) increased its antiplatelet activity (Bolibrukh et al., 2015), (iv) The inhibition of platelet activation by alpha-tocopherol quinone is greater than that of vitamin E (Rao et al., 1981), and (v) *gem*-diethyl/methyl substitutions and the addition/modifications of the third ring of *ortho*-carbonyl hydroquinone scaffold influence on the selective index (IC₅₀ TRAP-6/IC₅₀ Collagen) and the inhibitory capacity of platelet aggregation (Mendez et al., 2020).

Lawsone is a natural naphthoquinone that is easily obtained from natural sources and is currently commercially available in bulk. It has been used as the starting material for the synthesis of a variety of biologically active compounds and materials with interesting properties in fungi, bacteria, and viruses (Jordão et al., 2015). The synthesis reaction from lawsone described in this article has allowed the development of a variety of lawsone thioderivatives that could be potentially useful for the treatment of platelets aggregation-associated thrombosis diseases. In this way, we observed that the platelet antiaggregant activity depends on the nature and position of the substituent at the phenyl moiety (**Table 3**). All tested compounds showed higher effectiveness (lower IC_{50} values) against platelet aggregation induced by collagen than TRAP-6 (**Table 3**). It has described that given the variability of quinone in biological activities it is likely that there



FIGURE 3 | Antiplatelet activity of compound **4**. (**A**) Externalization of phosphatidylserine was evaluated by annexin-V binding. (**B**) Levels of CD63 on the surface of platelets after activation by TRAP-6 (5μ M) or collagen (1μ g/mL). (**C**) P-selectin expression was measured by flow cytometry in platelets stimulated with TRAP-6 (5μ M) or collagen (1μ g/mL). (**C**) P-selectin expression was measured by flow cytometry in platelets stimulated with TRAP-6 (5μ M) or collagen (1μ g/mL). (**D**) GPIIb/IIIa activation stimulated by TRAP-6 (5μ M) or collagen (1μ g/mL) was determined by flow cytometry using the PAC-1 antibody. The graph depicts the mean \pm *SD* of *n* = 3 experiments. The black graph is the control of resting platelets. **p* < 0.05, ***p* < 0.01, and ****p* < 0.001 vs. 0 (vehicle DMSO 0.2%).

is a low selectivity in its antiplatelet activity (Ma and Long, 2014). Our work, as shown in **Table 3**, shows that compound **6** presents a specific inhibition of collagen-induced platelet aggregation. Atherothrombosis is initiated by collagen exposure from endothelial damage to circulating platelets. These platelets, via the GPVI receptor, bind directly to collagen triggering platelet activation (Nieswandt and Watson, 2003). In this context, our results suggest that pretreatment of platelets with compound **6** down-regulates the GPVI-mediated signaling pathway (Jung et al., 2017).

Small structural changes led to the synthesis of non-toxic lawsone thio-derivatives with potent antiplatelet activity (Tables 1, 3). Our experimental design led to the synthesis of antiplatelets compounds, being compound 4 the most potent among the non-toxic compounds. Compound 4 presents the lower IC_{50} values, being 15.03 \pm 1.52 μM for TRAP-6, and $5.58 \pm 1.01 \mu$ M for collagen respectively (Figure 2). Thus, we decided to deeply analyze the effects of compound 4 in platelets. In addition to its non-toxic activity (Table 2), we demonstrated that compound 4 was unable to affect phosphatidylserine exposure (Figure 3A), a modification associated with platelet mitochondrial apoptotic-like events (Augereau et al., 2004). In non-activated platelets, CD63 is present on the dense granules and exposed in membranes after platelet activation (Israels and McMillan-Ward, 2005). CD63 is essential for P-selectin function, which is a thrombo-inflammatory molecule that participates in platelet activation and aggregation (Théorêt et al., 2011). As shown in Figure 3B, compound 4 decreased the expression of CD63 on platelets' membrane after activation with either TRAP-6 or collagen. As expected, platelets' P-selectin expression on activated platelets by TRAP-6 or collagen was inhibited by compound 4 at 10 and $40\,\mu\text{M}$ (Figure 3C). In activated platelets, GPIIb/IIIa is very important at the final step of platelet activation leading to aggregation (Chen et al., 2015). We analyzed the activated form of GPIIb/IIIa by studying the binding of FITC-labeled PAC-1 antibody by flow cytometry (Fuentes et al., 2014a). Figure 3D demonstrated that compound 4 inhibited TRAP-6- and collagen-induced PAC-1 binding to platelets at compound levels similar to those that inhibited platelet aggregation (Table 3).

Limitation and Perspectives

The set of experiments presented were designed to demonstrate that thio-derivatives of lawsone are useful novel scaffolds to synthesize antiplatelet compounds. The small number of compounds comprising the series analyzed was limited in most part due to the difficulty in accessing human platelets from a significant number of donors. Besides this fact, we were able

REFERENCES

- Al Nasr, I., Jentzsch, J., Winter, I., Schobert, R., Ersfeld, K., Koko, W. S., et al. (2019). Antiparasitic activities of new lawsone Mannich bases. *Archiv. Pharmazie* 352:1900128. doi: 10.1002/ardp.201900128
- Augereau, O., Rossignol, R., DeGiorgi, F., Mazat, J. P., Letellier, T., and Dachary-Prigent, J. (2004). Apoptotic-like mitochondrial events associated to

to determine that the modification inserted for compound **4** synthesis improved the activity on platelet aggregation. Future studies will be directed toward increasing the series of lawsone-derivatives and comparing their effects in different biological models. Although it was not in the scope of the current work, the lack of testing of cytotoxicity, bio-disponibility, and mechanisms of action on *in vivo* systems decrease the impact of compound **4** for pharmacological purposes. However, these will be solved in the future with studies in cells and animal models at different pathophysiological conditions.

In conclusion, our study revealed that thio-derivatives of lawsone are useful novel scaffolds to synthesize antiplatelet compounds, with compound 4 being the most potent.

DATA AVAILABILITY STATEMENT

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

ETHICS STATEMENT

The studies involving human participants were reviewed and approved by Scientific ethics committee, Universidad de Talca. The patients/participants provided their written informed consent to participate in this study.

AUTHOR CONTRIBUTIONS

EF, RA-M, and MM-Ci: conceptualization. MM-Cá and DM: formal analysis. EF, and RA-M: writing—original draft preparation and funding acquisition. AT: writing—review and editing. All authors contributed to the article and approved the submitted version.

FUNDING

This work was supported by ANID/CONICYT, FONDECYT grants #1180427 (EF) and #1180069 (RA-M), and ANID/REDES #170003 (EF). FONDECYT 11170142 (MM-Ci). AT was funded by Comision Sectorial de Investigación Científica (CSIC)-Uruguay, CSIC-Grupos N°536.

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fchem. 2020.00533/full#supplementary-material

phosphatidylserine exposure in blood platelets induced by local anaesthetics. *Thromb. Haemost* 92, 104–113. doi: 10.1160/TH03-10-0631

- Baell, J. B., and Nissink, J. W. M. (2018). Seven year itch: Pan-Assay Interference Compounds (PAINS) in 2017-utility and limitations. ACS Chem. Biol. 13, 36–44. doi: 10.1021/acschembio.7b00903
- Barani, M., Mirzaei, M., Torkzadeh-Mahani, M., and Nematollahi, M. H. (2018). Lawsone-loaded Niosome and its antitumor activity in MCF-7 breast

Cancer cell line: a Nano-herbal treatment for Cancer. Daru 26, 11–17. doi: 10.1007/s40199-018-0207-3

- Becatti, M., Fiorillo, C., Gori, A. M., Marcucci, R., Paniccia, R., Giusti, B., et al. (2013). Platelet and leukocyte ROS production and lipoperoxidation are associated with high platelet reactivity in Non-ST elevation myocardial infarction (NSTEMI) patients on dual antiplatelet treatment. *Atherosclerosis* 231, 392–400. doi: 10.1016/j.atherosclerosis.2013.09.030
- Bolibrukh, K., Polovkovych, S., Khoumeri, O., Halenova, T., Nikolaeva, I., Savchuk, O., et al. (2015). Synthesis and anti-platelet activity of thiosulfonate derivatives containing a quinone moiety. *Sci. Pharm.* 83, 221–231. doi: 10.3797/scipharm.1411-14
- Bolton, J. L., and Dunlap, T. (2017). Formation and biological targets of quinones: cytotoxic versus cytoprotective effects. *Chem. Res. Toxicol.* 30, 13–37. doi: 10.1021/acs.chemrestox.6b00256
- Bolton, J. L., Trush, M. A., Penning, T. M., Dryhurst, G., and Monks, T. J. (2000). Role of quinones in toxicology. *Chem. Res. Toxicol.* 13, 135–160. doi: 10.1021/tx9902082
- Caruso, R., Rocchiccioli, S., Gori, A. M., Cecchettini, A., Giusti, B., Parodi, G., et al. (2015). Inflammatory and antioxidant pattern unbalance in "clopidogrelresistant" patients during acute coronary syndrome. *Mediators Inflamm*. 2015:710123. doi: 10.1155/2015/710123
- Chang, C. Y., Huang, L. J., Wang, J. P., Teng, C. M., Chen, S. C., and Kuo, S. C. (2000). Synthesis and anti-platelet, anti-inflammatory and anti-allergic activities of methoxyisoflavanquinone and related compounds. *Chem. Pharm. Bull.* 48, 964–73. doi: 10.1248/cpb.48.964
- Chang, M.-C., Chang, B.-E., Pan, Y.-H., Lin, B.-R., Lian, Y.-C., Lee, M.-S., et al. (2019). Antiplatelet, antioxidative, and anti-inflammatory effects of hydroquinone. *J. Cell. Physiol.* 234, 18123–18130. doi: 10.1002/jcp. 28444
- Chang, T. S., Kim, H. M., Lee, K. S., Khil, L. Y., Mar, W. C., Ryu, C. K., et al. (1997). Thromboxane A2 synthase inhibition and thromboxane A2 receptor blockade by 2-[(4-cyanophenyl)amino]-3-chloro-1,4naphthalenedione (NQ-Y15) in rat platelets. *Biochem. Pharmacol.* 54, 259–268. doi: 10.1016/S0006-2952(97)00179-2
- Chen, Z., Mondal, N. K., Ding, J., Koenig, S. C., Slaughter, M. S., Griffith, B. P., et al. (2015). Activation and shedding of platelet glycoprotein IIb/IIIa under non-physiological shear stress. *Mol. Cell Biochem.* 409, 93–101. doi: 10.1007/s11010-015-2515-y
- Cho, Y. S., Kim, M. J., Lee, J. Y., Chung, J. H. (1997). The role of thiols in protecting against simultaneous toxicity of menadione to platelet plasma and intracellular membranes. J. Pharmacol. Exp. Ther. 280, 1335–1340.
- Cocheme, H. M., Kelso, G. F., James, A. M., Ross, M. F., Trnka, J., Mahendiran, T., et al. (2007). Mitochondrial targeting of quinones: therapeutic implications. *Mitochondrion* 7, S94–S102. doi: 10.1016/j.mito.2007.02.007
- Diamond, S. L. (2016). Systems analysis of thrombus formation. Circ. Res. 118, 1348–1362. doi: 10.1161/CIRCRESAHA.115.306824
- Fox, K. A., Mehta, S. R., Peters, R., Zhao, F., Lakkis, N., Gersh, B. J., et al. (2004). Benefits and risks of the combination of clopidogrel and aspirin in patients undergoing surgical revascularization for non-ST-elevation acute coronary syndrome: the Clopidogrel in Unstable angina to prevent Recurrent ischemic Events (CURE) Trial. *Circulation* 110, 1202–1208. doi: 10.1161/01.CIR.0000140675.85342.1B
- Franco, A. T., Corken, A., and Ware, J. (2015). Platelets at the interface of thrombosis, inflammation, and cancer. *Blood* 126, 582–588. doi: 10.1182/blood-2014-08-531582
- Fuentes, E., Badimon, L., Caballero, J., Padró, T., Vilahur, G., Alarcón, M., et al. (2014a). Protective mechanisms of adenosine 5'-monophosphate in platelet activation and thrombus formation. *Thromb. Haemost* 111, 491–507. doi: 10.1160/TH13-05-0386
- Fuentes, E., Caballero, J., Alarcon, M., Rojas, A., and Palomo, I. (2014b). Chlorogenic acid inhibits human platelet activation and thrombus formation. *PLoS ONE* 9:e90699. doi: 10.1371/journal.pone.0090699
- Fuentes, M., Araya-Maturana, R., Palomo, I., and Fuentes, E. (2018). Platelet mitochondrial dysfunction and mitochondria-targeted quinone-and hydroquinone-derivatives: review on new strategy of antiplatelet activity. *Biochem. Pharmacol.* 156, 215–222. doi: 10.1016/j.bcp.2018.08.035
- Geisler, T., Anders, N., Paterok, M., Langer, H., Stellos, K., Lindemann, S., et al. (2007). Platelet response to clopidogrel is attenuated in diabetic

patients undergoing coronary stent implantation. *Diabetes Care* 30, 372–374. doi: 10.2337/dc06-1625

- Harrington, R. A., Hodgson, P. K., and Larsen, R. L. (2003). Antiplatelet therapy. *Circulation* 108, e45–e47. doi: 10.1161/01.CIR.0000083142.13956.D6
- Israels, S. J., and McMillan-Ward, E. M. (2005). CD63 modulates spreading and tyrosine phosphorylation of platelets on immobilized fibrinogen. *Thromb Haemost* 93, 311–318. doi: 10.1160/TH.04-08-0503
- ISTH Steering Committee for World Thrombosis Day (2014). Thrombosis: a major contributor to the global disease burden. *J. Thromb. Haemost.* 12, 1580–1590. doi: 10.1111/jth.12698
- Jordão, A. K., Vargas, M. D., Pinto, A. C., da Silva, F. d. C., and Ferreira, V. F. (2015). Lawsone in organic synthesis. *RSC Adv.* 5, 67909–67943. doi: 10.1039/C5RA12785H
- Jung, S. H., Han, J. H., Park, H. S., Lee, J. J., Yang, S. Y., Kim, Y. H., et al. (2017). Inhibition of collagen-induced platelet aggregation by the secobutanolide Secolincomolide A from lindera obtusiloba blume. *Front. Pharmacol.* 8:560. doi: 10.3389/fphar.2017.00560
- Kim, E. J., Lim, K. M., Kim, K. Y., Bae, O. N., Noh, J. Y., Chung, S. M., et al. (2009). Doxorubicin-induced platelet cytotoxicity: a new contributory factor for doxorubicin-mediated thrombocytopenia. J. Thromb. Haemost 7, 1172–1183. doi: 10.1111/j.1538-7836.2009.03477.x
- Kim, S. R., Lee, J. Y., Lee, M. Y., Chung, S. M., Bae, O. N., and Chung, J. H. (2001). Association of quinone-induced platelet anti-aggregation with cytotoxicity. *Toxicol. Sci.* 62, 176–182. doi: 10.1093/toxsci/62.1.176
- Lien, J. C., Huang, L. J., Teng, C. M., Wang, J. P., and Kuo, S. C. (2002). Synthesis of 2-alkoxy 1,4-naphthoquinone derivatives as antiplatelet, antiinflammatory, and antiallergic agents. *Chem. Pharm. Bull.* 50, 672–4. doi: 10.1248/cpb.50.672
- Ma, W., and Long, Y. T. (2014). Quinone/hydroquinone-functionalized biointerfaces for biological applications from the macro- to nano-scale. *Chem. Soc. Rev.* 43, 30–41. doi: 10.1039/C3CS60174A
- Mendez, D., Urra, F. A., Millas-Vargas, J. P., Alarcon, M., Rodriguez-Lavado, J., Palomo, I., et al. (2020). Synthesis of antiplatelet ortho-carbonyl hydroquinones with differential action on platelet aggregation stimulated by collagen or TRAP-6. *Eur. J. Med. Chem.* 192:112187. doi: 10.1016/j.ejmech.2020.112187
- Moura, L. A., de Almeida, A. C., da Silva, A. V., de Souza, V. R., Ferreira, V. F., Menezes, M. V., et al. (2016). Synthesis, anticlotting and antiplatelet effects of 1,2,3-triazoles derivatives. *Med. Chem.* 12, 733–741. doi: 10.2174/1573406412666160502153417
- Mousa, S. A. (1999). Antiplatelet therapies: from aspirin to GPIIb/IIIareceptor antagonists and beyond. *Drug Discov. Today* 4, 552–561. doi: 10.1016/S1359-6446(99)01394-X
- Munoz-Gutierrez, C., Sepulveda, C., Caballero, J., Palomo, I., and Fuentes, E. (2017). Study of the interactions between Edaglitazone and Ciglitazone with PPARgamma and their antiplatelet profile. *Life Sci.* 186, 59–65. doi: 10.1016/j.lfs.2017.07.031
- Nieswandt, B., and Watson, S. P. (2003). Platelet-collagen interaction: is GPVI the central receptor? *Blood* 102, 449–461. doi: 10.1182/blood-2002-12-3882
- Paes, M. A. A., Gaspar, R. S., Fuentes, E., Wehinger, S., Palomo, I., and Trostchansky, A. (2019). Lipid metabolism and signaling in platelet function. *Adv. Exp. Med. Biol.* 1127, 97–115. doi: 10.1007/978-3-030-11488-6_7
- Price, M. J., Angiolillo, D. J., Teirstein, P. S., Lillie, E., Manoukian, S. V., Berger, P. B., et al. (2011). Platelet reactivity and cardiovascular outcomes after percutaneous coronary intervention: a time-dependent analysis of the Gauging Responsiveness with a VerifyNow P2Y12 assay: Impact on Thrombosis and Safety (GRAVITAS) trial. *Circulation* 124, 1132–1137. doi: 10.1161/CIRCULATIONAHA.111.029165
- Qin, X., Peng, Y., and Zheng, J. (2018). *In vitro* and *in vivo* studies of the electrophilicity of physcion and its oxidative metabolites. *Chem. Res. Toxicol.* 31, 340–349. doi: 10.1021/acs.chemrestox.8b00026
- Rahmoun, N. M., Boucherit-Otmani, Z., Boucherit, K., Benabdallah, M., Villemin, D., and Choukchou-Braham, N. (2012). Antibacterial and antifungal activity of lawsone and novel naphthoquinone derivatives. *Médecine et Maladies Infectieuses* 42, 270–275. doi: 10.1016/j.medmal.2012.05.002
- Rao, G. H., Cox, C. A., Gerrard, J. M., and White, J. G. (1981). Alpha tocopherol quinone (alpha TQ): a potent inhibitor of platelet function. *Prog. Lipid Res.* 20, 549–552. doi: 10.1016/0163-7827(81)90097-7
- Ritchie, J. L., Alexander, H. D., and Rea, I. M. (2000). Flow cytometry analysis of platelet P-selectin expression in whole blood-methodological considerations.

Clin. Lab. Haematol. 22, 359–363. doi: 10.1046/j.1365-2257.2000. 00339.x

- Tandon, V. K., and Maurya, H. K. (2009). "On water:" unprecedented nucleophilic substitution and addition reactions with 1,4-quinones in aqueous suspension. *Tetrahedron Lett.* 50, 5896–5902. doi: 10.1016/j.tetlet.2009.07.149
- Tandon, V. K., Maurya, H. K., Verma, M. K., Kumar, R., and Shukla, P. K. (2010).
 "On water" assisted synthesis and biological evaluation of nitrogen and sulfur containing hetero-1,4-naphthoquinones as potent antifungal and antibacterial agents. *Eur. J. Med. Chem.* 45, 2418–2426. doi: 10.1016/j.ejmech.2010. 02.023
- Théorêt, J.-F., Yacoub, D., Hachem, A., and Gillis, M.-A., Merhi, Y. (2011). P-selectin ligation induces platelet activation and enhances microaggregate and thrombus formation. *Thrombosis Res.* 128, 243–250. doi: 10.1016/j.thromres.2011.04.018
- Urra, F. A., Cordova-Delgado, M., Lapier, M., Orellana-Manzano, A., Acevedo-Arevalo, L., Pessoa-Mahana, H., et al. (2016). Small structural changes on a hydroquinone scaffold determine the complex I inhibition or uncoupling of tumoral oxidative phosphorylation. *Toxicol. Appl. Pharmacol.* 291, 46–57. doi: 10.1016/j.taap.2015.12.005
- Vaiyapuri, S., Moraes, L. A., Sage, T., Ali, M. S., Lewis, K. R., Mahaut-Smith, M. P., et al. (2013). Connexin40 regulates platelet function. *Nat. Commun.* 4:2564. doi: 10.1038/ncomms3564

- Walsh, T. G., Berndt, M. C., Carrim, N., Cowman, J., Kenny, D., and Metharom, P. (2014). The role of Nox1 and Nox2 in GPVI-dependent platelet activation and thrombus formation. *Redox Biol.* 2, 178–186. doi: 10.1016/j.redox.2013.12.023
- Xu, K., Wang, P., Wang, L., Liu, C., Xu, S., Cheng, Y., et al. (2014). Quinone derivatives from the genus Rubia and their bioactivities. *Chem. Biodivers* 11, 341–363. doi: 10.1002/cbdv.201200173
- Yun-Choi, H. S., Kim, J. H., and Takido, M. (1990). Potential inhibitors of platelet aggregation from plant sources, V. Anthraquinones from seeds of Cassia obtusifolia and related compounds. J. Nat. Prod. 53, 630–633. doi: 10.1021/np50069a014

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2020 Monroy-Cárdenas, Méndez, Trostchansky, Martínez-Cifuentes, Araya-Maturana and Fuentes. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.