

VEGF Stimulates RCAN1.4 Expression in Endothelial Cells via a Pathway Requiring Ca^{2+} /Calcineurin and Protein Kinase C- δ

Katherine Holmes¹, Elinor Chapman^{1,2}, Violaine See³, Michael J. Cross^{1*}

1 Department of Pharmacology and Therapeutics, School of Biomedical Sciences, University of Liverpool, Liverpool, United Kingdom, **2** North West Cancer Research Institute, School of Biological Sciences, Bangor University, Bangor, United Kingdom, **3** School of Biological Sciences, University of Liverpool, Liverpool, United Kingdom

Abstract

Background: Vascular endothelial growth factor (VEGF) has previously been shown to upregulate the expression of the endogenous calcineurin inhibitor, regulator of calcineurin 1, variant 4 (RCAN1.4). The aim of this study was to determine the role and regulation of VEGF-mediated RCAN1.4 expression, using human dermal microvascular endothelial cells (HDMECs) as a model system.

Methodology/Principal Findings: We show that VEGF is able to induce RCAN1.4 expression during cellular proliferation and differentiation, and that VEGF-mediated expression of RCAN1.4 was inhibited by the use of inhibitors to protein kinase C (PKC) and calcineurin. Further analysis revealed that siRNA silencing of PKC- δ expression partially inhibited VEGF-stimulated RCAN1.4 expression. Knockdown of RCAN1.4 with siRNA resulted in a decrease in cellular migration and disrupted tubular morphogenesis when HDMECs were either stimulated with VEGF in a collagen gel or in an endothelial/fibroblast co-culture model of angiogenesis. Analysis of intracellular signalling revealed that siRNA mediated silencing of RCAN1.4 resulted in increased expression of specific nuclear factor of activated T-cells (NFAT) regulated genes.

Conclusions/Significance: Our data suggests that RCAN1.4 expression is induced by VEGFR-2 activation in a Ca^{2+} and PKC- δ dependent manner and that RCAN1.4 acts to regulate calcineurin activity and gene expression facilitating endothelial cell migration and tubular morphogenesis.

Citation: Holmes K, Chapman E, See V, Cross MJ (2010) VEGF Stimulates RCAN1.4 Expression in Endothelial Cells via a Pathway Requiring Ca^{2+} /Calcineurin and Protein Kinase C- δ . PLoS ONE 5(7): e11435. doi:10.1371/journal.pone.0011435

Editor: Alma Zernecke, Universität Würzburg, Germany

Received: January 19, 2010; **Accepted:** June 3, 2010; **Published:** July 6, 2010

Copyright: © 2010 Holmes et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: K.H. and M.J.C. are supported by North West Cancer Research Fund (NWCRCF). V.S. is a recipient of a BBSRC David Phillips Fellowship (BB/C520471/1). E.C. is funded by Cancer Research Wales. The funders had no role in the study design, data collection and analysis, decision to publish or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

* E-mail: m.j.cross@liv.ac.uk

Introduction

Angiogenesis is defined as the formation of new blood vessels from pre-existing vessels, and is an essential process in embryonic development and normal physiology. However, an imbalance in angiogenesis may play a role in a number of pathological conditions, including cancer, atherosclerosis, and ischaemia [1]. The vascular endothelial growth factor (VEGF) family of homodimeric glycoproteins have been shown to be critical for angiogenesis. The VEGF family comprises 5 members; VEGF-A, VEGF-B, VEGF-C, VEGF-D and placenta growth factor (PLGF). In addition there are a number of structurally related proteins, including parapoxvirus *Ory* VEGF (VEGF-E). These ligands bind in an overlapping pattern to 3 receptors; VEGF Receptor 1 (VEGFR-1), VEGFR-2 and VEGFR-3 (reviewed in [2]). Vascular endothelial cells express both VEGFR-1 and VEGFR-2, with VEGFR-2 generally accepted as the principal receptor through which VEGF signals are transmitted in the vascular endothelium. Binding of VEGF to VEGFR-2 results in the activation of a number of intracellular signalling pathways including mitogen-

activated protein kinases (MAPKs) and protein kinase B (PKB)/Akt. VEGFR-2 also activates phospholipase C- γ (PLC γ) leading to an increase in intracellular calcium and activation of protein kinase C (PKC; reviewed in [2]).

Proteins within the regulator of calcineurin (RCAN) family are able to bind and regulate the protein phosphatase calcineurin. This family comprises 3 members; RCAN1, 2 and 3. RCAN1 was initially given the name Down syndrome critical region 1 (DSCR1) due to its location on chromosome 21 [3]. Other names include Adapt 78, myocyte-enriched calcineurin interacting protein 1 (MCIP1) and calcipressin 1 [4,5,6].

The human *RCAN1* gene comprises 7 exons, the first 4 of which are alternative first exons, resulting in different isoforms which show different patterns of expression and regulation [7]. Exon one gives rise to the isoform RCAN1.1 [8]. Exon 2 lacks a methionine start site required for translation, and exon 3 encodes just 3 amino acids [7]. Exon 4 gives rise to the isoform RCAN1.4, and is under the control of a calcineurin responsive promoter, comprising multiple consensus binding sites for the transcription factor nuclear factor of activated T-cells (NFAT) [9] and GATA-2/3 sites [10].

More recently, 5 consensus binding sites for activator protein 1 (AP-1) transcription factors have been identified in the region flanking exon 4 [11].

The serine/threonine protein phosphatase 2B (PP2B)/calcineurin is a heterodimer composed of a calcineurin catalytic subunit A (CnA), and a calcineurin regulatory subunit B (CnB). Upon Ca^{2+} induced activation, CnA dephosphorylates members of the NFAT family [12]. This dephosphorylation allows the translocation of NFAT to the nucleus where it binds to the NFAT consensus sequence present in the promoter region of various genes, including RCAN1.4, resulting in an increase in transcription [12]. RCAN1.4 binds to the catalytic domain within CnA and inhibits its activity [5,6,13,14]. Phosphorylation of RCAN1.4 by MAPK and glycogen synthase kinase 3 (GSK-3), allows RCAN1.4 to act as a substrate for CnA [15]. Thus, RCAN1.4 has been suggested to act as a negative feedback inhibitor for CnA signalling.

RCAN1 has previously been shown to be upregulated in a variety of endothelial cell lines in response to VEGF, including human umbilical vein endothelial cells (HUVEC), human aortic endothelial cells (HAEC), human dermal microvascular endothelial cells (HDMEC), and human retinal endothelial cells (HREC) [10,16,17]. In each of these studies RCAN1.4, but not RCAN1.1 was found to be upregulated in response to VEGF treatment. Knockdown of RCAN1 in endothelial cells has also been shown to inhibit VEGF stimulated migration *in vitro*, and VEGF induced angiogenesis, *in vivo* [16]. Most recently, an increase in *Rcan1* expression in mice, by the introduction of an extra transgenic copy, has been shown to suppress tumour growth [18]. In this study, we have analysed the intracellular signalling pathways utilised by VEGFR-2 to stimulate RCAN1.4 expression in primary human dermal microvascular endothelial cells (HDMECs). Our results show that the calcineurin/NFAT pathway and PKC-delta pathway are required for maximal induction of RCAN1.4. Furthermore, RCAN1.4 was required for migration and efficient tubular morphogenesis in these cells.

Materials and Methods

Materials

Recombinant human VEGF-A₁₆₅, epidermal growth factor (EGF), hepatocyte growth factor (HGF) and colony stimulating factor-1 (CSF-1) were obtained from Peprotech. Recombinant human VEGF-B₁₆₇ and recombinant *Osf* virus VEGF-E were obtained from Promokine. Fibroblast growth factor-2 (FGF-2) was obtained from R&D Systems. Phorbol-12-myristate-13-acetate (PMA), A23187, GF109203X (GFX), G66976, Cyclosporin A (CsA) and BAPTA-AM were obtained from Calbiochem. ZM323881 was obtained from Tocris. Oleoyl-L- α -lysophosphatidic acid (LPA) and ethylene glycol tetraacetic acid (EGTA) were obtained from Sigma. All other materials were obtained from Invitrogen unless otherwise stated.

Cell Culture

Human dermal microvascular endothelial cells (HDMEC), human umbilical vein endothelial cells (HUVEC) and human aortic endothelial cells (HAEC) were purchased from Promocell and were cultured in endothelial cell basal media (EBM) MV2 growth media (C-22221; Promocell), supplemented with 5% (v/v) foetal calf serum (FCS) and EGF (5 ng/ml), VEGF (0.5 ng/ml), FGF-2 (10 ng/ml), long R3 insulin growth factor-1 (20 ng/ml), hydrocortisone (0.2 $\mu\text{g}/\text{ml}$) and ascorbic acid (1 $\mu\text{g}/\text{ml}$) (supplement pack C-39221; Promocell). HEK 293 cells were cultured in Dulbecco's Modified Eagles Medium (DMEM) supplemented with 10% FCS. Human dermal fibroblasts were obtained from

Promocell, and cultured in fibroblast growth media (C-23010; Promocell) supplemented with FGF-2 (1 ng/ml) and Insulin (5 $\mu\text{g}/\text{ml}$) (supplement pack; Promocell). Cells were routinely cultured on gelatin coated plates in a humidified incubator under 5% CO_2 at 37°C. For growth factor stimulation, cells were serum starved for at least 16 hours by changing media to MV2 media supplemented with 1% (v/v) FCS without growth factor supplements.

Quantitative Real Time-Polymerase Chain Reaction (RT-qPCR)

Total RNA was extracted from HDMECs using the RNeasy mini kit (Qiagen). DNase treatment was performed using the on-column DNase digestion (Qiagen). One μg total RNA was used for cDNA synthesis with M-MLV reverse transcriptase and oligodT primers. RT-qPCR was performed using Power SYBR Green Mastermix (Applied Biosystems) and the primer sequences as detailed below. Reactions were analysed on an ABI 7000 real-time PCR machine using the following cycle conditions; 2 minutes at 50°C, 10 minutes at 95°C, followed by 45 cycles of 15 seconds at 95°C and 60 seconds at 60°C. Results were normalised against β -actin mRNA expression. Primer sequences were as follows; RCAN1.4 forward – CTCAC-TAGGGGCTTGACTGC, reverse – CAGGCAATCAGGGAGC-TAAA; RCAN1.1 forward – TCATTGACTGCGAGATGGAG, reverse – TGATGTCCTTGTGATACGTCCT; COX-2 forward – GAAGAAAGTTCATCCCTGATCCC, reverse – CTGGGCA-AAGAATGCCAAACA; IL-8 forward – CAGGAATTGAATGGG-TTTGC, reverse – AGCAGACTAGGGTTGCCAG; ICAM-1 forward – CAAGGCCTCAGTCAGTGTGA, reverse – CCTC-TGGCTTCGTCAGAATC.

Immunoblotting

Protein lysates from HDMECs were prepared in LDS sample buffer containing β -mercaptoethanol. Proteins were resolved by SDS-PAGE on 4–12% NuPage Gels, and transferred onto nitrocellulose membranes (Amersham). Membranes were blocked with 5% (w/v) BSA. Proteins were detected using the following antibodies; RCAN1 (D6694; Sigma), NFAT1 (610702; BD Transductions Labs), phospho-p44/42 MAPK (Thr202/Tyr204; #4377), phospho-PKC (pan; #9371), phospho-PLC γ (Tyr 783; #2821), PKC α (#2056), PKC δ (~2058; all Cell Signalling Technology), PKC ϵ (06-991; Upstate), PKC η (sc-215), colony stimulating factor-1 receptor (CSF-1R; sc-13949) and Actin (sc-1625; all Santa Cruz Biotechnology). Membranes were washed 6 times with TBS-T, and incubated with peroxidase-conjugated secondary antibodies (Amersham/Sigma). Blots were detected using an enhanced chemiluminescence (ECL) detection kit (Amersham).

siRNA transfection

HDMECs were transfected with either two separate small interfering RNA (siRNA) duplexes at a concentration of 10 nM, or a mixture of two siRNA duplexes at a concentration of 10 nM each, using 0.1% (v/v) Lipofectamine RNAiMAX, according to the manufacturer's instructions. All siRNA duplexes were validated by RT-qPCR and western blotting prior to use [19]. Transfection reactions were performed in serum-free OptiMEM. Cell media was changed to serum containing media 4 hours after transfection. The following siRNA duplexes were obtained from Qiagen; RCAN1 (Hs_DSCR1_5 HP, SI03224900; Hs_DSCR1_6 HP, SI03246208), PKC α (Hs_PRKCA_5 HP, SI00301308; Hs_PRKCA_6, HP SI00605927), PKC δ (Hs_PRKCD_8 HP, SI00288337; Hs_PRKCD_11 HP, SI02660539), PKC ϵ (Hs_PRKCE_5 HP,

SI00287784; Hs_PRKCE_6 HP, SI02622088), and PKC η (Hs_PRKCH_5, SI02224075; Hs_PRKCH_6 HP, SI02224082) and All-stars Negative Control siRNA.

Overexpression of Chimeric Receptors

The pcDNA3.1 mammalian expression plasmids encoding the extracellular domain of human CSF-1 receptor (CSF-1R)/c-fms fused to the transmembrane and intracellular domain of murine VEGFR-2 (CKRwt) and murine VEGFR-2 Y1173F point mutation (CKR Y1173F mutant) were kindly provided by Dr. Nader Rahimi (Boston University, Boston, MA). HDMEC were combined with the plasmid DNA and HMVEC-L Nucleofector™ solution (AMAXA) according to the manufacturer's instructions and transfected using program S-05 on the AMAXA Nucleofector™. Cells were then transferred to cell culture dishes and allowed to adhere in 1% FCS containing media overnight.

Intracellular Calcium Measurements

HDMECs plated upon glass based cell culture dishes (Iwaki) were loaded with the Ca²⁺ indicator dyes Fluo-4-AM (5 μ M) and Fura Red-AM (5 μ M) for 20 minutes. Intracellular calcium was monitored using fluorescent microscopy to measure the emission ratio of Fluo-4/Fura Red.

Inositol Phosphate assay

PLC activity was determined by measuring the agonist-stimulated accumulation of [³H]inositol phosphates in the presence of 20 mM LiCl from [³H]myo-inositol labelled cells as previously described [20].

Cellular Proliferation Assay

HDMECs were seeded in 24 well plates in normal growth media for 24 hours. Media was then changed to 1% (v/v) FCS containing media for 24 hours. Cells were stimulated with VEGF (50 ng/ml) for 72 hours, and the number of viable cells determined using a luminescent based Cell-titre glo assay (Promega).

Scratch Wound Migration Assay

HDMECs were seeded in 12 well plates in normal growth media for 48 hours. Media was then changed to 1% (v/v) FCS containing media for 24 hours. A scratch was introduced to the cell monolayer using a sterile 200 μ l pipette tip. Cells were stimulated with VEGF (50 ng/ml) for 18 hours. The cells were then fixed with 2% (w/v) paraformaldehyde and then stained with 0.5% (w/v) crystal violet solution to allow enhanced imaging.

Tube Formation Assay

Collagen gels were prepared by mixing bovine collagen type-I (Vitrogen) with 0.1 M NaOH and 10x Ham's F-12 media (Promocell) in the ratio 8:1:1, and supplemented with 20 mM HEPES, 0.117% (w/v) bicarbonate solution, and 2 mM Glutamax-I. HDMEC were starved in 1% (v/v) FCS containing media for 24 hours prior to being seeded onto pre-prepared collagen gels. Once cells had adhered, a top layer of collagen was added and allowed to set. Cells were then stimulated with growth factors in 1% (v/v) FCS containing media for 20 hours to induce tubular morphogenesis. Collagen gels were fixed with 2% (w/v) paraformaldehyde and cells visualized by phase contrast microscopy.

Co-culture Assay

Human dermal fibroblasts were seeded in 24 well plates, and allowed to become confluent. On day 4 HDMEC were seeded

upon the fibroblast monolayer. Co-cultures were serum starved, and then stimulated with VEGF for 5 days. To assess the effects of gene knockdown siRNA transfections were performed upon days 3, 5 and 8. In order to visualise tube formation by HDMEC co-cultures were fixed with 70% (v/v) ethanol, and then stained with the endothelial specific marker CD31 (Dako) [21].

Statistics

Results are expressed as mean \pm standard deviation (S.D.). Statistical analyses were performed using an unpaired Student's *t*-test. Statistical significance was set at a P-value less than 0.05 (*), or 0.01 (**).

Results

VEGF-A, PMA and A23187 stimulate an increase in RCAN1.4 expression in HDMEC

Human *RCAN1* comprises seven exons, the first four of which are alternatively spliced to give rise to different isoforms (Fig. 1A). Quantitative RT-PCR was used to analyse the expression of *RCAN1.1* and *RCAN1.4* mRNA in response to VEGF-A in HDMECs. Under basal conditions, the amount of *RCAN1.4* mRNA was found to be 100-fold greater than the amount of *RCAN1.1* mRNA (data not shown). Following VEGF-A stimulation, levels of *RCAN1.4* mRNA were increased 8-fold (Fig. 1B); *RCAN1.1* mRNA levels were not increased by VEGF-A stimulation (Fig. 1B). The expression of RCAN1.1 protein, which was practically undetectable on a western blot using an antibody to the C-terminal of RCAN1 which is able to detect all RCAN1 isoforms, did not increase upon VEGF-A stimulation (data not shown).

Vascular endothelial cells express both VEGFR-1, and VEGFR-2, both of which are receptors for VEGF-A. Expression of both receptors in HDMEC was confirmed by PCR (data not shown). In order to establish the role of each receptor in RCAN1.4 upregulation, VEGF-B and VEGF-E, which bind VEGFR-1 and VEGFR-2 respectively, were used to stimulate HDMEC. VEGF-B did not elicit an RCAN1.4 upregulation, while VEGF-E was able to induce a similar increase in RCAN1.4 expression as VEGF-A at both the mRNA and protein level (Fig. 1C and D). For the purposes of this study, all further experiments were conducted using VEGF-A.

In contrast to VEGF-A, stimulation of HDMECs with a range of growth factors (FGF-2, EGF and HGF), and the G-protein agonist LPA did not evoke an increase in *RCAN1.4* mRNA (Fig. 1E) or RCAN1.4 protein (Fig. 1F); active signalling by all growth factors was confirmed by MAPK phosphorylation. VEGF-A was also unique in its ability to stimulate PLC- γ phosphorylation and NFAT activation (Fig. 1F). A23187, a Ca²⁺ ionophore, and PMA, a potent PKC activator, were capable of inducing increased RCAN1.4 mRNA (Fig. 1E) and increased RCAN1.4 protein levels in the absence of increased PLC- γ phosphorylation (Fig. 1F). Interestingly, unlike VEGF-A and A23187, PMA did not appear to stimulate the dephosphorylation of NFAT.

RCAN1.4 is downstream of PLC- γ , PKC and Ca²⁺/ Calcineurin

The ability of VEGF-A to uniquely stimulate PLC activity in HDMECs compared with other growth factors was verified by analysing the generation of inositol phosphates (Fig. 2A). This data correlates with the finding that increases in intracellular Ca²⁺ were only observed following stimulation with VEGF-A and A23187, a Ca²⁺ ionophore (Fig. 2B).

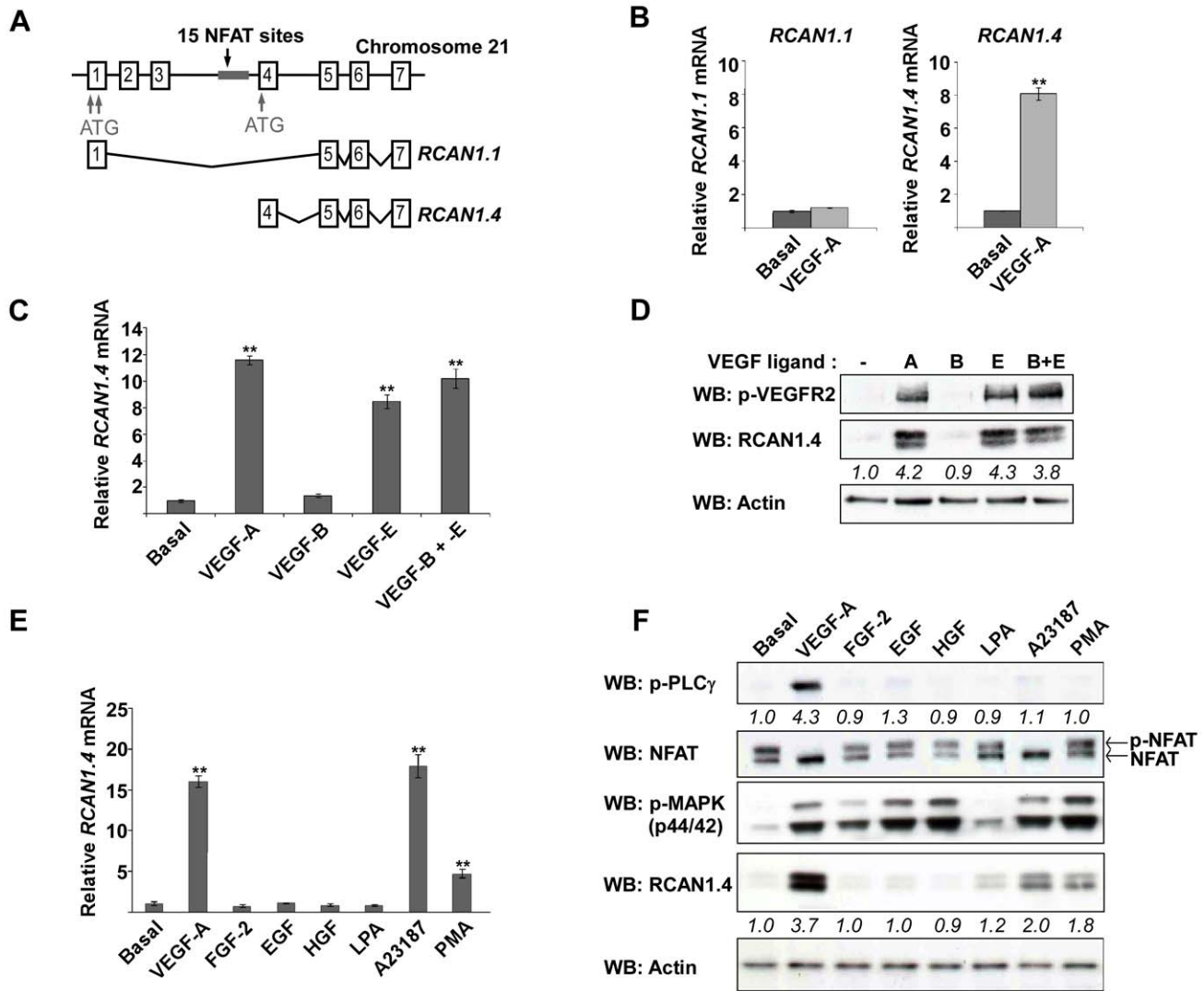


Figure 1. VEGF-A stimulates RCAN1.4 expression, PLC- γ activation and NFAT dephosphorylation in HDMEC. (A) Exon structure of RCAN1, showing the alternative first exons used by the RCAN1.1 and RCAN1.4 isoforms. (B) HDMEC were stimulated with VEGF-A (50 ng/ml) for 1 hour, and the increase in expression of *RCAN1.1* and *RCAN1.4* mRNA, relative to the level of each mRNA in unstimulated (basal) cells, determined by RT-qPCR. Results were normalised to β -actin mRNA expression. Data shows mean \pm SD (n=3). **P<0.01, versus Basal. (C) HDMEC were stimulated with VEGF-A (50 ng/ml), VEGF-B (50 ng/ml), VEGF-E (50 ng/ml) or VEGF-B and -E (50 ng/ml each) for 1 hour and the expression of *RCAN1.4* mRNA determined by RT-qPCR. Results were normalised to β -actin mRNA expression. Data shows mean \pm SD (n=3). **P<0.01, versus Basal. (D) HDMEC were stimulated with agonists as in 1C for 10 mins or 3 hours and the expression levels of RCAN1.4 (3 hour stimulation), and phospho-VEGFR2 (pVEGFR2) proteins determined by SDS-PAGE followed by western blotting. Actin was used as a loading control. Numbers below the blots denote the relative density of each band. Results are representative of 3 experiments. (E) HDMEC were stimulated with VEGF-A (50 ng/ml), FGF-2 (50 ng/ml), EGF (50 ng/ml), HGF (50 ng/ml), LPA (10 μ M), A23187 (5 μ M) and PMA (100 nM) for 1 hour, and the expression of *RCAN1.4* mRNA, relative to unstimulated (basal) cells, determined by RT-qPCR. Results were normalised to β -actin mRNA expression. Data shows mean \pm SD (n=3). **P<0.01, versus Basal. (F) HDMEC were stimulated with agonists as in 1C for 10 mins or 3 hours and the expression levels of RCAN1.4 (3 hour stimulation), phospho-PLC γ (p-PLC γ), NFAT and phospho-p44/42 MAPK (p-MAPK) proteins determined by SDS-PAGE followed by western blotting. Actin was used as a loading control. Numbers below the blots denote the relative density of each band. Results are representative of 3 experiments.
doi:10.1371/journal.pone.0011435.g001

Phosphorylation of the tyrosine 1175 site in VEGFR-2, following VEGF-A stimulation, allows the binding and subsequent phosphorylation of PLC- γ [22]. PLC- γ is then able to catalyze the hydrolysis of the membrane phospholipid phosphatidylinositol (4,5)-bisphosphate (PIP₂) resulting in the generation of diacylglycerol (DAG), and inositol 1,4,5-trisphosphate (IP₃). DAG is a physiological activator of PKC, whilst IP₃ acts upon the endoplasmic reticulum to release calcium stores, resulting in a rise in intracellular calcium ([Ca²⁺]_i). In order to further assess the importance of PLC- γ activation by VEGFR-2 in the increase of

RCAN1.4 expression, HDMECs were transiently transfected using AMAXA nucleofection with plasmids encoding chimeric receptors, comprising the extracellular domain of the human CSF-1R fused with the transmembrane and intracellular domain of the murine VEGFR-2 (Flk-1; Fig. 3A) [23,24]. Two forms of the chimeric receptor were used; the wild-type (CKRwt), and a mutated form in which the tyrosine residue at position 1173 (corresponding to position 1175 in the human receptor) is replaced with a phenylalanine residue (CKR Y1173F). In cells expressing CKRwt, CSF-1 ligand may be used to elicit a VEGF-A-like

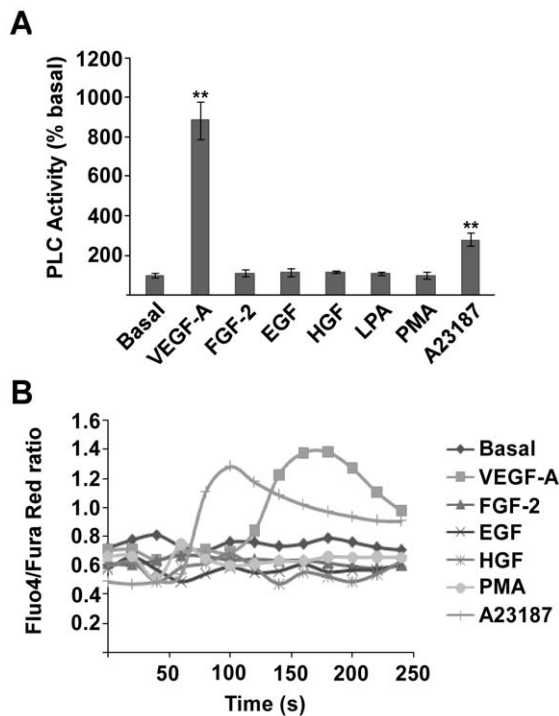


Figure 2. VEGF-A stimulates PLC activity and Ca^{2+} release. (A) HDMEC were stimulated with VEGF-A (50 ng/ml), FGF-2 (50 ng/ml), EGF (50 ng/ml), HGF (50 ng/ml), LPA (10 μ M), A23187 (5 μ M) and PMA (100 nM) for 30 mins, and PLC activity determined by inositol phosphate assay. Data shows mean \pm SD (n=3). **P<0.01, versus Basal. (B) HDMEC were loaded with Fluo4 and Fura Red dyes and stimulated with agonists as in 2A and changes in intracellular calcium monitored by measuring the fluorescent intensity of the two dyes. Results are representative of 3 experiments. doi:10.1371/journal.pone.0011435.g002

response, including phosphorylation of PLC- γ following its binding to tyrosine residue 1173. However, in cells expressing CKR Y1173F, stimulation with CSF-1 can no longer elicit PLC- γ phosphorylation. Expression of the chimeric receptors was confirmed by western blotting with an antibody to the extracellular domain of CSF-1R, and active signalling confirmed by the phosphorylation of PLC- γ and MAPK in response to the ligand CSF-1 (Fig. 3B). Phosphorylated PLC- γ was observed in CKRwt transfected cells in response to CSF-1 stimulation. This phosphorylation was lost in cells transfected with the CKR Y1173F mutant confirming that PLC- γ was not activated by the CKR Y1173F receptor.

The importance of PLC- γ activation in the upregulation of *RCAN1.4* mRNA was assessed by RT-qPCR (Fig. 3C). In cells transfected with CKRwt, levels of *RCAN1.4* mRNA were found to be upregulated when stimulated with CSF-1 to a similar magnitude as that seen when stimulating with VEGF. However, in cells transfected with CKR Y1173F, stimulation with CSF-1 did not induce a rise in the levels of *RCAN1.4* mRNA. Stimulating untransfected cells with CSF-1 did not induce an increase in *RCAN1.4* mRNA, consistent with the lack of expression of endogenous CSF-1R in these cells.

A range of inhibitors were used to further investigate the signalling pathways resulting in the upregulation of RCAN1.4 by VEGF-A (Fig. 4A). Use of ZM323881, an inhibitor to VEGFR-2 [25], inhibited RCAN1.4 upregulation, confirming further that upregulation of RCAN1.4 by VEGF-A was occurring through

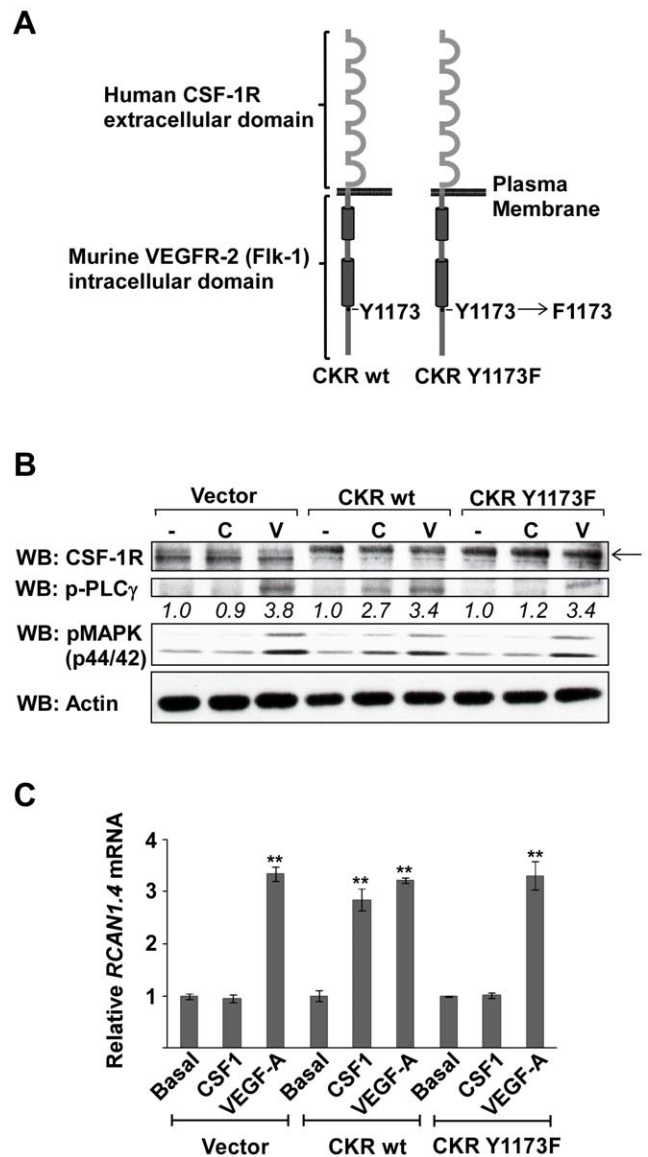


Figure 3. Induction of RCAN1.4 requires phosphorylation of Y1175 on VEGFR-2. (A) Structure of chimeric receptors used to transfected HDMEC. (B) HDMEC were transfected with chimeric receptors and stimulated with CSF-1 (C; 50 ng/ml) or VEGF-A (V; 50 ng/ml) for 10 mins. The levels of CSF-1R, phospho-PLC γ (p-PLC γ) and phospho-p44/42 MAPK (pMAPK) were determined by SDS-PAGE followed by western blotting. Actin was used as a loading control. Numbers below the blots denote the relative density of each band. Results are representative of 3 experiments. (C) HDMEC were transfected with chimeric receptors, stimulated with CSF-1 (50 ng/ml) or VEGF-A (50 ng/ml) for 1 hour, and the expression of *RCAN1.4* mRNA, relative to unstimulated vector transfected cells, determined by RT-qPCR. Results were normalised to β -actin mRNA expression. Data shows mean \pm SD (n=3). **P<0.01, versus unstimulated vector control. doi:10.1371/journal.pone.0011435.g003

VEGFR-2 signalling. BAPTA and EGTA, chelators of intracellular and extracellular calcium respectively [26], were both found to inhibit VEGF-A induced RCAN1.4. GF109203X, a broad spectrum PKC inhibitor [27] was found to inhibit VEGF-A induced RCAN1.4, whilst G δ 6976, a classical PKC inhibitor [28], did not, implicating a novel PKC in the upregulation of RCAN1.4 expression. Finally cyclosporin A (CsA), a potent inhibitor of calcineurin [29], was found to block VEGF-A induced RCAN1.4.

These data support roles for Ca^{2+} , calcineurin and PKC in the pathway linking VEGFR-2 to RCAN1.4 expression.

The same inhibitors were used in combination with PMA (Fig. 4B) and A23187 (Fig. 4C). As expected GF109203X inhibited PMA stimulation of RCAN1.4. However, as with VEGF-A, Gö6976 did not affect PMA stimulated RCAN1.4 expression. The use of CsA supported our previous data (Fig. 1D), showing that PMA is not acting through the calcineurin pathway to stimulate expression of RCAN1.4. A23187 induction of RCAN1.4 was inhibited by the Ca^{2+} chelators, and by CsA, but not by PKC inhibitors. Use of the MEK1 inhibitor U0126 [30] did not affect RCAN1.4 induction by VEGF, PMA or A23187, suggesting that RCAN1.4 is not downstream of the MEK-p44/p42 MAPK pathway.

PKC δ is required for induction of RCAN1.4

The phosphorylation of different PKC isoforms following stimulation with VEGF-A, FGF-2, PMA or A23187 was investigated using a pan-phospho PKC antibody (Fig. 5). The antibody used detects serine residues within the hydrophobic regions of PKC α , δ , ϵ and η , the phosphorylation of which is required for PKC activation [31]. PKC α , δ , ϵ and η were all rapidly phosphorylated in response to VEGF-A. However, only the phosphorylation of PKC δ was found to be sustained above basal levels over a 60 minute period, while the phosphorylation of PKC α , ϵ and η were found to return to basal levels within 30 minutes (Fig. 5). In response to FGF-2 stimulation, small transient increases in the phosphorylation of PKC α , PKC ϵ and PKC η were observed (Fig. 5). As with VEGF-A, the phosphorylation of these isoforms returned to basal levels within 30 minutes of stimulation. Similar data to FGF-2 was obtained when stimulating HDMECs with EGF (data not shown). In response to PMA, the phosphorylation of all PKC isoforms is sustained over 60 minutes. In contrast, in response to A23187 the PKCs were seen to undergo a weak transient phosphorylation.

Use of the PKC activator, PMA, and PKC inhibitors, GF109203X and Gö6976, implicated the involvement of a novel PKC in the upregulation of RCAN1.4. PKC is a multi-protein family, comprising several isoforms [32]. The classical isoforms α , β , and γ have a requirement for Ca^{2+} and DAG for activity, whereas the novel isoforms δ , ϵ , η and θ require DAG but not Ca^{2+} . A third class of atypical isoforms require neither Ca^{2+} nor DAG. The PKC expression profile in HDMECs was characterised using end-point PCR, and visualised on an agarose gel (Fig. 6A). The PKC profile of 293 cells was also characterised as a positive control for the primer sets used. HDMECs were found to express the classical PKC isoform α , the novel isoforms δ , ϵ , and η , and the atypical isoform ι .

HDMEC were transfected with siRNAs to the expressed classical (α) and novel isoforms (δ , ϵ and η) using non-silencing (N.S.) siRNA as a control. The knockdown of each isoform was confirmed by western blotting (Fig. 6B). Knockdown of each isoform was quantified by RT-qPCR, and found to be between 80 and 90% (data not shown). Knockdown of PKC δ was found to reduce VEGF-A upregulation of RCAN1.4 mRNA by approximately 50% (Fig. 6C). Knockdown of other PKC isoforms present in HDMECs did not affect VEGF-A upregulation of RCAN1.4. This was confirmed at the protein level by western blotting, with a 40% reduction in RCAN1.4 protein observed following PKC δ knockdown (Fig. 6D). Knockdown of PKC δ was not found to decrease the ability of PMA to stimulate RCAN1.4. However, knockdown of PKC ϵ reduced PMA stimulated RCAN1.4 mRNA (Fig. 6C), and reduced protein levels by 40% (Fig. 6D).

In order to confirm that the role of PKC δ in VEGF-A stimulated upregulation of RCAN1.4 is not confined to HDMEC, PKC δ knockdown was also performed in HUVEC and HAEC

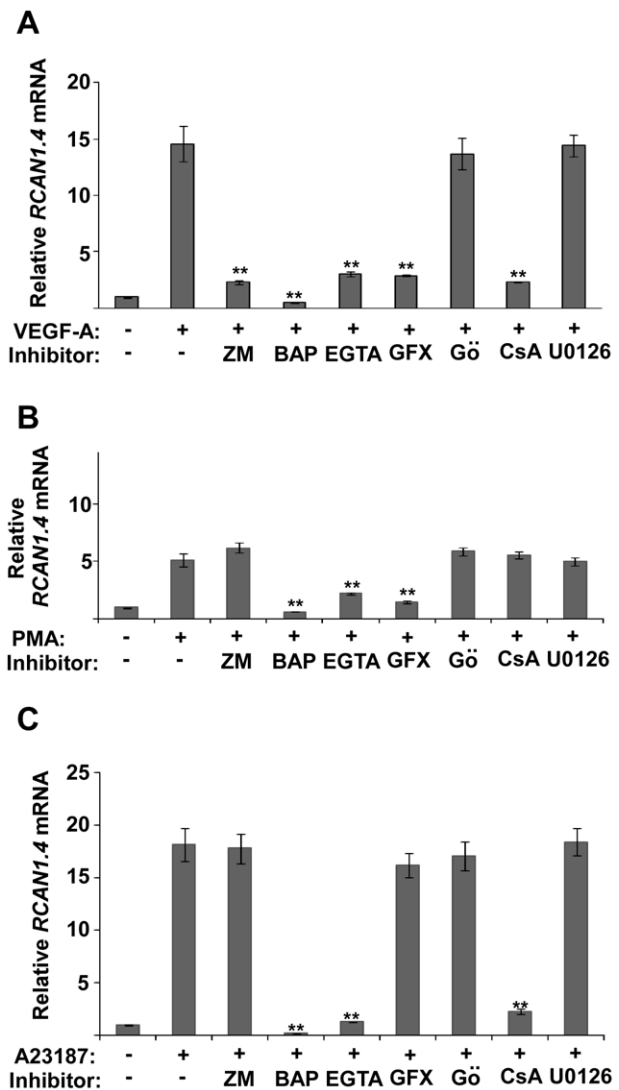


Figure 4. RCAN1.4 induction requires Ca^{2+} , calcineurin and PKC activity. HDMEC were incubated with the inhibitors ZM (3 μM), BAP (20 nM), EGTA (2.5 mM), GFX (3 μM), Go (3 μM), CsA (100 nM) or U0126 (5 μM) for 30 minutes, and then stimulated with: (A) VEGF-A (50 ng/ml), (B) PMA (100 nM), or (C) A23187 (5 μM) for 1 hour. Expression of RCAN1.4 mRNA, relative to unstimulated (basal) cells, was determined by RT-qPCR. Results were normalised to β -actin expression. Data shows mean \pm SD (n = 3). **P < 0.01, versus VEGF/PMA/A23187 treated. BAP – BAPTA/AM, CsA – cyclosporin A, GFX – GF109203X, Go – Gö6976, ZM – ZM323881.

doi:10.1371/journal.pone.0011435.g004

using two separate PKC δ specific siRNA duplexes (Fig. 6E). In all 3 endothelial cell lines knockdown of PKC δ resulted in an approximate 50% reduction in RCAN1.4 expression in response to VEGF-A. In addition, CsA was successfully used to inhibit RCAN1.4 upregulation in all 3 cell lines (Fig. 6E).

Knockdown of RCAN1.4 affects expression of NFAT regulated genes

RCAN1.4 has previously been shown to regulate calcineurin signalling via a negative feedback loop [5,6,9]. In order to investigate the role of RCAN1.4 in the regulation of VEGFR-2 signalling we utilised siRNA against RCAN1 and analysed signalling and gene expression in response to VEGF-A. Due to

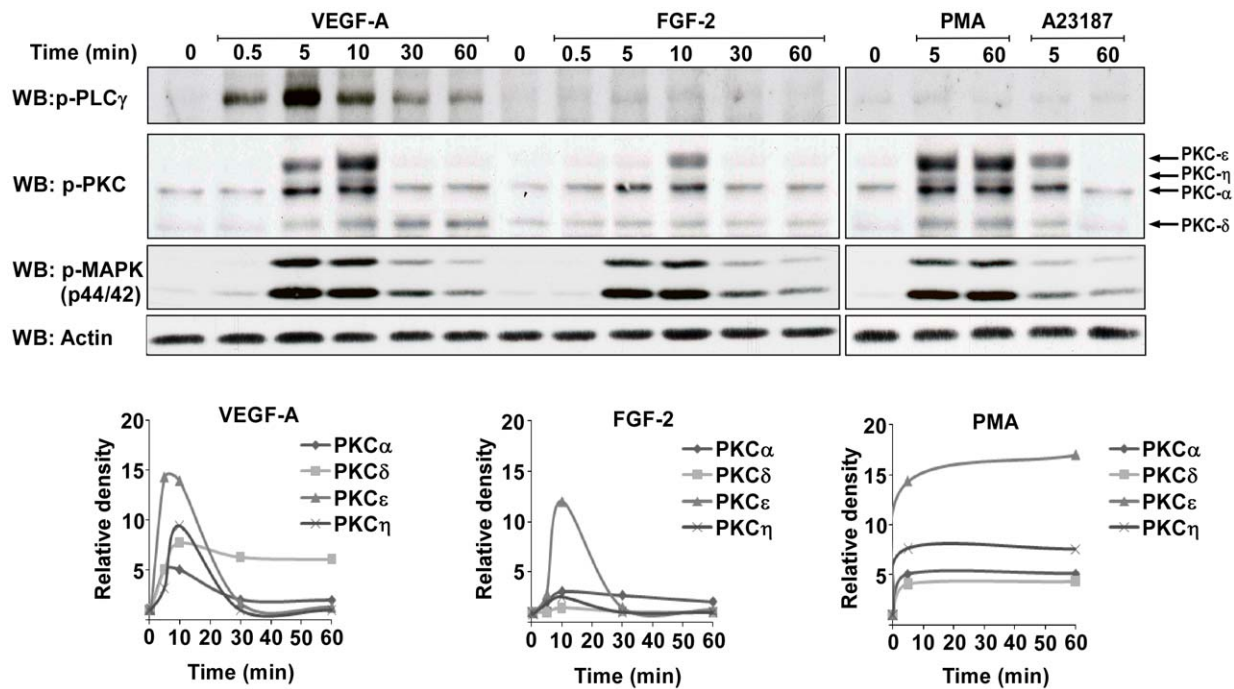


Figure 5. PKC δ phosphorylation is sustained in response to VEGF-A. HDMEC were stimulated with VEGF-A (50 ng/ml), FGF-2 (50 ng/ml) PMA (100 nM) or A23187 (5 μ M) for the time points indicated. The expression levels of phospho-PLC γ (p-PLC γ), phospho-PKC (p-PKC) and phospho-p44/42 MAPK (p-MAPK) were determined by SDS-PAGE followed by western blotting. Actin was used as a loading control. Graphs show the relative expression of each phosphorylated PKC isoform in response to either VEGF-A, FGF-2 or PMA, as determined from the relative density of the bands compared to the unstimulated (basal) condition. Results are representative of 3 experiments. doi:10.1371/journal.pone.0011435.g005

the lack of availability of isoform specific RCAN1.4 siRNAs, the two duplexes used in this study are capable of knocking down the expression of both RCAN1 isoforms. Since RCAN1.4 is the only detectable isoform and is VEGF-inducible, we attribute the effects of the siRNA on VEGF signalling to changes in RCAN1.4 expression. Efficient knockdown of RCAN1.4 was confirmed at the protein level by western blotting (Fig. 7A) and at the mRNA level by qRT-PCR with a 90% reduction in mRNA compared with the N.S. siRNA control (data not shown). No effect of RCAN1.4 knockdown was observed on the phosphorylation of PLC- γ and MAPK in response to VEGF (Fig. 7A). In order to analyse the effect of RCAN1.4 silencing on gene expression we chose a panel of NFAT-regulated genes: *COX-2* [33], *IL- β* [34] and *ICAM-1* [35]. RCAN1.4 knockdown increased the expression of *COX-2*, *IL- β* and *ICAM-1* mRNA in response to VEGF-A (Fig. 7B), confirming that RCAN1.4 acts as a negative regulator of NFAT-mediated gene expression.

Knockdown of RCAN1.4 affects cellular migration and tubular morphogenesis, but not cellular proliferation

It has previously been shown that plating telomerase-immortalised HDMEC onto either a collagen or fibronectin matrix causes the cells to respond differently [36]. Upon a collagen matrix the cells undergo tubular morphogenesis in response to VEGF-A stimulation, whereas on a fibronectin matrix the cells proliferate in response to VEGF-A. In our studies, we have used gelatin, rather than fibronectin, as a similar proliferative behaviour was exhibited upon each matrix. The expression of *RCAN1.4* mRNA in response to VEGF-A upon collagen and gelatin matrices over a 24 hour period was determined using RT-qPCR (Fig. 8A). *RCAN1.4* mRNA expression was increased upon both matrices. However, the increase in expression was greater on the collagen matrix, as

opposed to the gelatin matrix. In addition, the expression of *RCAN1.4* mRNA was sustained for 24 hours on collagen, whereas on the gelatin matrix expression levels had returned to close to basal levels within 24 hours. A similar pattern of RCAN1.4 expression was observed at the protein level on both matrices (Fig. 8B).

To investigate the role of RCAN1.4 in cellular proliferation in response to VEGF-A stimulation RCAN1.4 expression was knocked down using siRNA and the extent of cellular proliferation over 72 hours was determined. In untransfected cells there was an 80% increase in cell number following VEGF-A stimulation, compared to those cells left in basal conditions. Knockdown of RCAN1.4 in HDMECs did not impact upon cellular proliferation in response to VEGF-A (Fig. 9A).

The role of RCAN1.4 in cellular migration was determined using a scratch wound migration assay, in which RCAN1.4 expression was knocked down, and the ability of the cells to migrate into a scratched wound in response to VEGF-A stimulation was quantified (Fig. 9B). Following knockdown of RCAN1.4 the ability of HDMEC to migrate into the wound was reduced by 80%, indicating a role for RCAN1.4 in endothelial cell migration.

The importance of RCAN1.4 expression in tubular morphogenesis was assessed using two *in vitro* models of angiogenesis. Firstly, the cells were plated within a collagen gel, and stimulated with VEGF-A (Fig. 9C). The ability of cells to form tubes was determined by measuring the total cellular junctions using AngioQuant software [37]. In response to VEGF-A, the number of cellular junctions increased by 85%, when compared with unstimulated cells. However, when cells were transfected with siRNA to RCAN1.4 their ability to undergo tubular morphogenesis was impaired. Under basal conditions the number of cellular junctions was reduced by 70%. Though VEGF-A was able to induce an increase in tubular morphogenesis in cells transfected

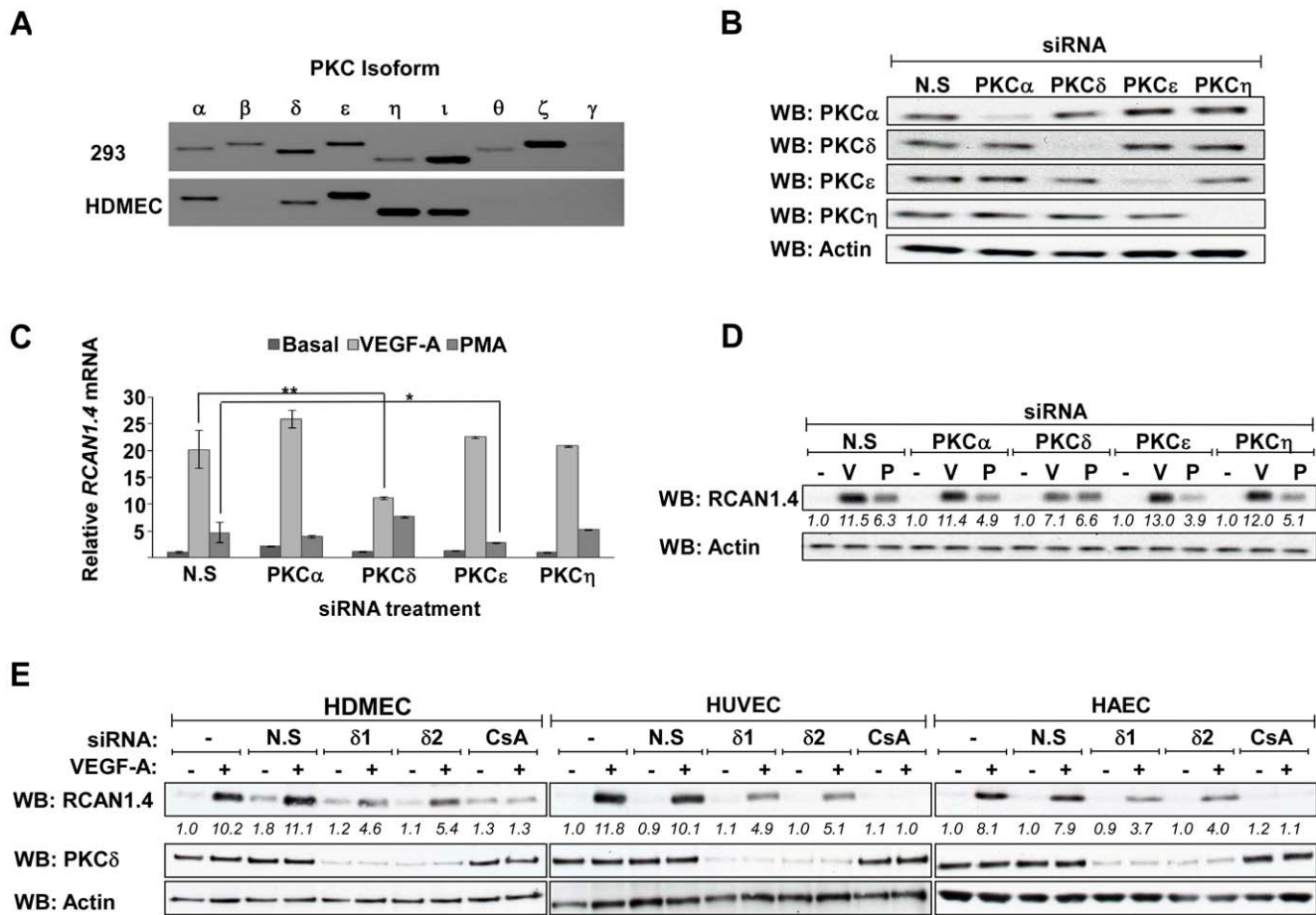


Figure 6. VEGF-A mediated RCAN1.4 expression is regulated by PKC δ . (A) PKC isoforms present in HDMEC and 293 cells were quantified by endpoint PCR run on an agarose gel. Results are representative of 3 experiments (B) HDMEC were transfected with a mixture of two siRNAs to individual PKC isoforms or with non-silencing (N.S.) siRNA, and the knockdown confirmed by SDS-PAGE followed by western blotting. Actin was used as a loading control. Results are representative of 3 experiments (C) HDMEC were transfected with a mixture of two siRNAs to individual PKC isoforms and stimulated 48 hours post-transfection with VEGF-A (50 ng/ml) or PMA (100 nM) for 1 hour. Expression of *RCAN1.4* mRNA, relative to unstimulated (basal) N.S. transfected cells, was determined by RT-qPCR. Results were normalised to β -actin expression. Data shows mean \pm SD, (n=3). *P<0.05, **P<0.01, versus N.S. treated cells. (D) HDMEC were transfected with a mixture of two siRNAs to individual PKC isoforms and stimulated 48 hours post-transfection with VEGF (V; 50 ng/ml) or PMA (P; 100 nM) for 3 hours. Expression levels of RCAN1.4 protein were determined by SDS-PAGE followed by western blotting. Actin was used as a loading control. Numbers below the blot denote the relative density of each band. Results are representative of 3 experiments. (E) HDMEC, HUVEC and HAEC were transfected with two separate siRNAs to PKC δ or N.S. siRNA as indicated, and stimulated 48 hours post-transfection with VEGF (50 ng/ml) for 3 hours. Expression levels of RCAN1.4 protein were determined by SDS-PAGE followed by western blotting. Actin was used as a loading control. Numbers below the blots denote the relative density of each band. Results are representative of 3 experiments. doi:10.1371/journal.pone.0011435.g006

with RCAN1.4 siRNA, compared to unstimulated cells, it did not exceed the level seen in untransfected basal cells.

To further investigate the importance of RCAN1.4 in tubular morphogenesis a second *in vitro* assay was utilised, in which HDMEC were cultured in combination with human dermal fibroblast cells [21]. As with the collagen assay, the ability of HDMECs to undergo tubular morphogenesis in response to VEGF-A was impaired following knockdown of RCAN1.4 (Fig. 9D). Taken together, the data from the collagen gel assay and co-culture assay suggest that RCAN1.4 is required for efficient tubular morphogenesis.

Discussion

VEGF-A has previously been shown to upregulate RCAN1.4 expression in endothelial cells, triggering a negative feedback loop which regulates the activity of calcineurin, and results in decreased angiogenesis [10,17,38]. It has been previously assumed that

VEGF-A is able to regulate RCAN1.4 expression by activation of the classical Ca²⁺/calcineurin pathway leading to activation of the NFAT transcription factor; the potential involvement of other signalling pathways has not been investigated. We now show that in addition to the Ca²⁺/calcineurin pathway, VEGF-A also utilises PKC δ in the regulation of RCAN1.4 expression. In addition, we show that RCAN1.4 is required for efficient tubular morphogenesis in two *in vitro* assays of angiogenesis, including a co-culture model.

RCAN1.4 was found to be upregulated at both the mRNA and the protein level in response to VEGF treatment (Fig. 1B). Although RCAN1.1 has also been reported to be upregulated in HUVECs in response to VEGF [39], we did not see upregulation of RCAN1.1 in HDMECs. Furthermore, basal levels of *RCAN1.1* mRNA were 100-fold lower than basal levels of *RCAN1.4* mRNA, and RCAN1.1 expression was practically undetectable on a western blot. VEGF-A and VEGF-E, but not VEGF-B induced

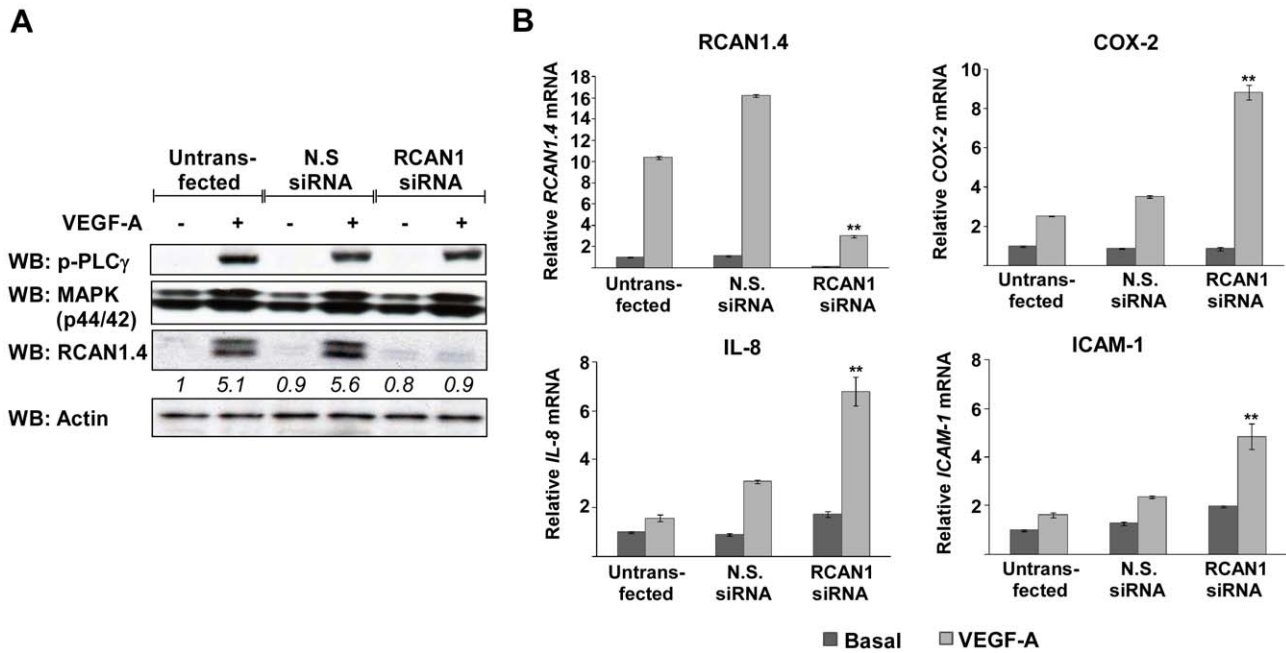


Figure 7. RCAN1.4 regulates NFAT induced gene expression. (A) HDMECs were transfected with a mixture of two siRNAs to RCAN1 or non-silencing (N.S.) siRNA and the knockdown of RCAN1.4 and effects on phospho-PLC γ (p-PLC γ) and phospho-p44/42 MAPK (pMAPK) were determined by SDS-PAGE followed by western blotting. Actin was used as a loading control. Numbers below the blots denote the relative density of each band. Results are representative of 3 experiments. (B) HDMECs were transfected with a mixture of two RCAN1 siRNAs and stimulated with VEGF-A (50 ng/ml) for 1 hour. Expression of *COX-2*, *IL-8* and *ICAM-1* mRNA, relative to unstimulated (basal) untransfected cells, were determined by RT-qPCR. Results were normalised to β -actin mRNA expression. Data shows mean \pm SD (n=3). **P<0.01, versus untransfected. doi:10.1371/journal.pone.0011435.g007

RCAN1.4 expression in HDMECs, indicating that RCAN1.4 upregulation occurs via VEGFR-2 signalling (Fig. 1C). No other growth factor was found to induce RCAN1.4 expression (Fig. 1E and 1F). Furthermore, VEGF-A was the only growth factor found to phosphorylate PLC- γ , and raise intracellular Ca²⁺ (Fig. 1D/2B). Although there is evidence that other growth factors, including FGF-2 and EGF, are able to stimulate PLC- γ activity in other cell

lines [40,41,42], it would appear that in primary HDMECs the ability of VEGF to stimulate PLC- γ activity is unique amongst growth factors.

In addition to VEGF-A, PMA and A23187 were also found to upregulate the expression of RCAN1.4. Treatment of HUVECs with a combination of VEGF-A and PMA has previously been shown to enhance the upregulation of RCAN1.4 [17]. The ability

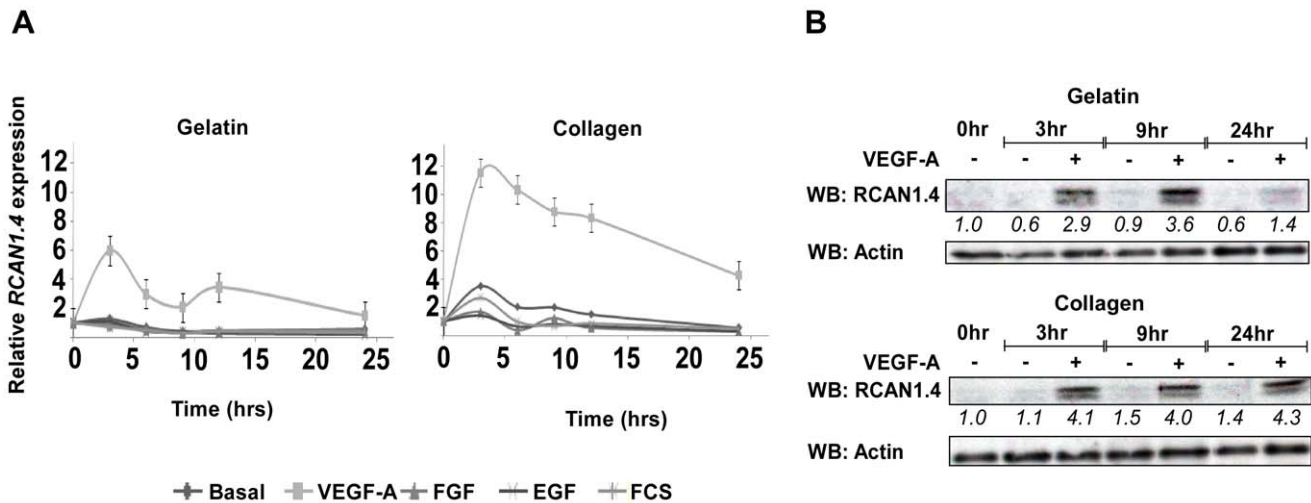


Figure 8. RCAN1.4 is expressed during VEGF-mediated proliferation and tubular morphogenesis. (A) HDMECs were plated onto either a collagen or gelatin matrix and stimulated with VEGF-A (50 ng/ml), FGF-2 (50 ng/ml), EGF (50 ng/ml) or FCS (10%), for the time points indicated. Expression of *RCAN1.4* mRNA, relative to unstimulated (basal) cells at 0 hr, was determined by RT-qPCR. Results were normalised to β -actin mRNA expression. Data shows mean \pm SD, (n=3). (B) HDMECs were treated as in 8(A), and the expression of RCAN1.4 protein determined by SDS-PAGE followed by western blotting. Numbers below the blots denote the relative density of each band. Results are representative of 3 experiments. doi:10.1371/journal.pone.0011435.g008

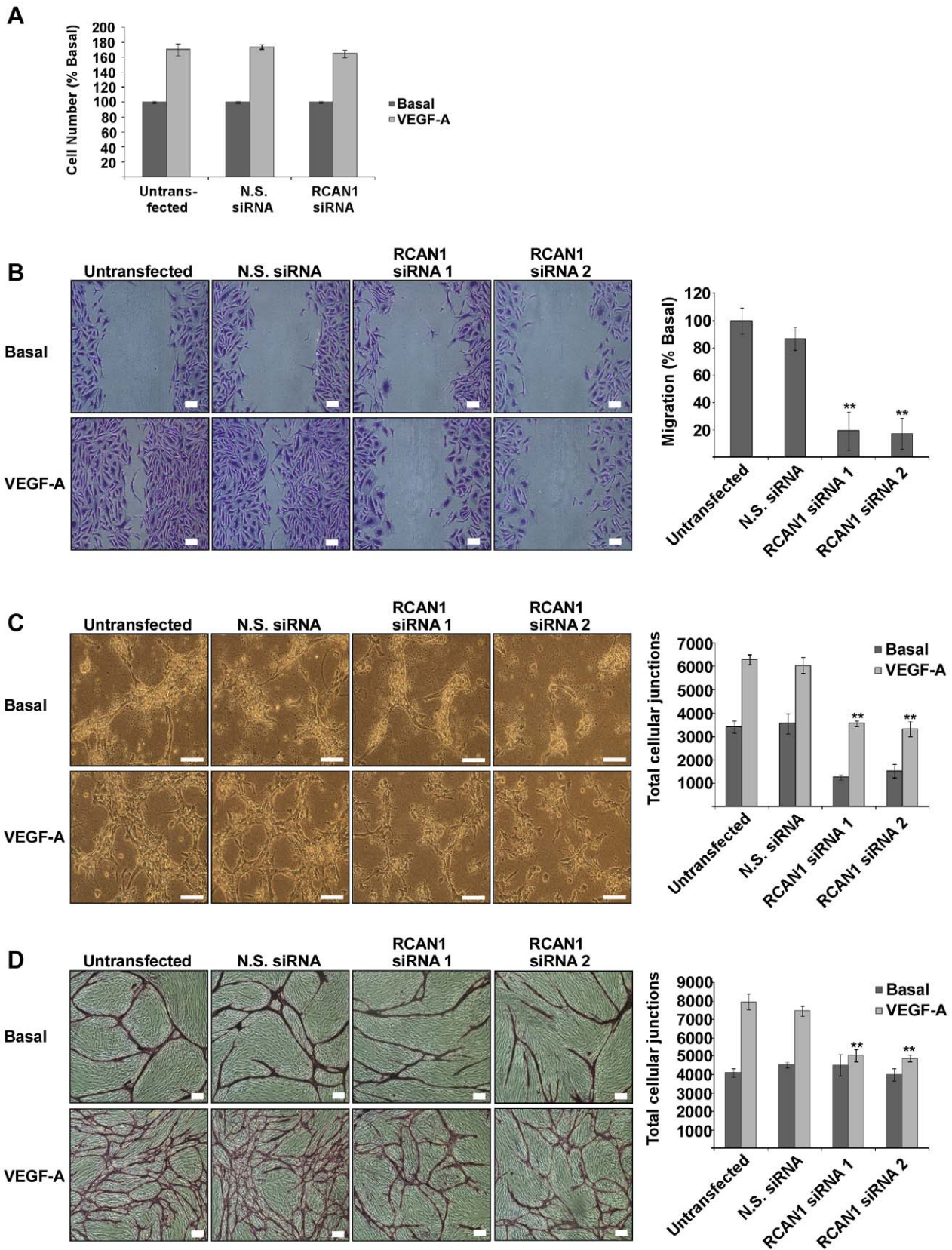


Figure 9. RCAN1.4 is required for migration and tubular morphogenesis, but not proliferation in HDMEC. (A) HDMEC were transfected with a mixture of two siRNAs to RCAN1 or non-silencing (N.S.) siRNA and stimulated with VEGF-A (50 ng/ml) for 72 hours. Cellular proliferation was determined by measuring viable cells. Data shows mean \pm SD, (n = 3). (B) HDMEC were transfected with two individual siRNAs to RCAN1 or N.S. siRNA as indicated. A scratch was introduced into the cell monolayer and the cells then stimulated with VEGF-A (50 ng/ml). Cells were fixed at 18 hours and

stained with crystal violet. Bars = 100 μ m. The graph shows the extent of migration in response to VEGF-A relative to the untransfected condition. Data shows mean \pm SD, (n = 3). **P < 0.01, versus untransfected. (C) Cells were transfected with two individual siRNAs to RCAN1 or N.S. siRNA and plated onto a collagen matrix, and stimulated with VEGF-A (50 ng/ml). Collagen gels were fixed at 20 hours to allow visualisation of tubes. Bars = 100 μ m. The graph shows quantification of tubular morphogenesis as measured by the number of cellular junctions. Data shows mean \pm SD, (n = 3). **P < 0.01, versus untransfected, VEGF treated. (D) Cells were cultured upon fibroblast lawns, and transfected with two individual siRNAs to RCAN1 or N.S. siRNA. Cells were stimulated with VEGF-A (50 ng/ml) for 5 days. Cells were fixed and stained for the endothelial specific marker CD31 to allow visualisation of tubes. Bars = 100 μ m. The graph shows quantification of tubular morphogenesis as measured by the number of cellular junctions. Data shows mean \pm SD (n = 3). **P < 0.01, versus untransfected, VEGF treated.
doi:10.1371/journal.pone.0011435.g009

of PMA to cause RCAN1.4 upregulation has been previously noted in the literature [17,38], although to date the exact role of PKC in the regulation of RCAN1.4 expression has not been investigated.

Here we show that unlike VEGF-A, stimulation of HDMECs with PMA does not result in the dephosphorylation of NFAT (Fig. 1D). Furthermore, use of the calcineurin inhibitor CsA did not inhibit PMA induced upregulation of RCAN1.4 (Fig. 4A). Finally, we have shown that stimulation with PMA does not induce a rise in intracellular Ca^{2+} , which is required for calcineurin activity (Fig. 2B). Taken together, these data point

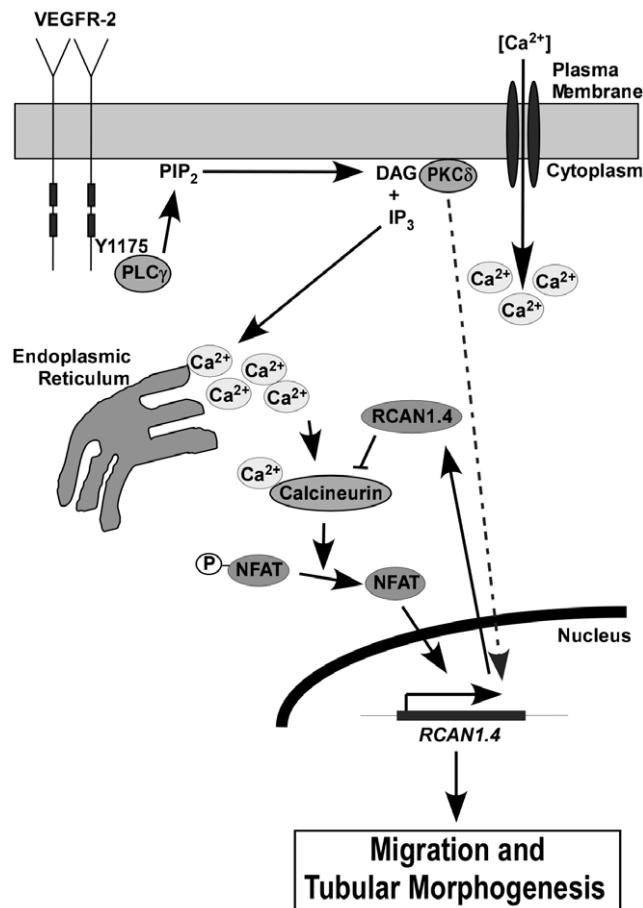


Figure 10. Summary of RCAN1.4 upregulation by VEGF-A in HDMEC. VEGFR-2 activation leads to PLC- γ activation and the subsequent increase in intracellular Ca^{2+} leading to activation of calcineurin and NFAT, which gives rise to increased RCAN1.4 expression. In addition, VEGFR-2 activation results in the sustained phosphorylation of PKC δ , which contributes to the upregulation of RCAN1.4 via an unknown pathway. RCAN1.4 acts to regulate VEGFR-2 mediated cellular migration and tubular morphogenesis.
doi:10.1371/journal.pone.0011435.g010

towards a secondary pathway, involving PKC, and independent from the calcineurin pathway, which regulates RCAN1.4.

Profiling of the PKC isoforms in HDMECs detected the expression of just 5 isoforms; the classical α , the novel δ , ϵ , and η and the atypical ι (Fig. 6A). VEGF-A was found to induce the rapid phosphorylation of PKC α , δ , ϵ and η , with only PKC δ phosphorylation being sustained, whereas PMA induced sustained activation of these PKC isoforms (Fig. 5). FGF-2 was found to only weakly phosphorylate PKC ϵ and η . This is the first data profiling the temporal phosphorylation of multiple PKC isoforms in response to VEGF-A and FGF-2 in human endothelial cells. Activation of PKC δ has been previously observed following stimulation of endothelial cells with VEGF-A, and has been implicated in Akt activation, and secretion of von Willebrand factor (vWF) from endothelial cells, suggesting that PKC δ may regulate a number of pathways required for endothelial cell function. [43,44]. Why the phosphorylation of PKC δ is selectively sustained in response to VEGF-A is unknown, but may be related to the activation and interaction of other VEGF-A responsive proteins with PKC δ .

Targeted knockdown of these PKC isoforms using specific siRNAs was performed to further identify which PKCs had a role in RCAN1.4 upregulation (Fig. 6C–D). Following PKC δ knockdown VEGF-A induced RCAN1.4 was reduced at both the mRNA and the protein level by approximately 50%, suggesting that PKC δ is able to regulate RCAN1.4 expression in HDMEC. In contrast, PMA appears to preferentially use PKC ϵ , rather than PKC δ , to regulate RCAN1.4 expression. This is likely to be due to differences in kinetic profiles of phosphorylated PKCs following stimulation by either VEGF-A or PMA in HDMECs. In contrast to VEGF-A, PMA stimulates a sustained phosphorylation of PKC ϵ and η , and it is possible that in this instance PKC ϵ is able to regulate RCAN1.4 expression independently of calcineurin mediated activation of NFAT. PKC δ was also found to be required for VEGF-A induced RCAN1.4 expression in both HUVEC and HAEC confirming that the importance of PKC δ in VEGF-A signalling is not confined to HDMEC (Fig. 6E).

The induction of RCAN1.4 by VEGF-A, but not PMA, was inhibited by the use of the calcineurin inhibitor, CsA. Calcineurin signalling is therefore necessary for RCAN1.4 induction by VEGF-A. However, since knockdown of PKC δ does not result in the complete inhibition of RCAN1.4 induction by VEGF-A, it would appear that signalling via PKC δ may cooperate with the calcineurin pathway to induce RCAN1.4 expression, probably by regulating the activity of a transcription factor distinct from NFAT. A recent study demonstrated that RCAN1.4 could be upregulated by c-Jun, a component of the AP-1 complex [11]. However, it is unlikely that this is the pathway through which PKC is acting as a known inhibitor of AP-1 activation, MEK1 inhibitor U0126 [30], was found not to affect VEGF or PMA stimulated *RCAN1.4* mRNA expression (Fig. 3).

Proliferation is an important process contributing towards angiogenesis. The role of RCAN1.4 in proliferation is still somewhat ambiguous. Overexpression of RCAN1 has been

shown to reduce endothelial cell proliferation [10], while knockdown of RCAN1.4 has been shown to increase endothelial cell proliferation [39]. RCAN1.4 is thought to exert its effects upon cellular proliferation via the regulation of calcineurin and NFAT induced gene transcription. In our study we were not able to detect an effect of RCAN1.4 knockdown upon proliferation (Fig. 9A). A similar result has also been reported in HUVECs [16]. A recently published study has shown that knockdown of RCAN1.4 decreases MAPK activation and cellular proliferation in human U87MG glioblastoma cells, via a calcineurin independent mechanism [45]. We do not observe any alterations in MAPK signalling following RCAN1.4 knockdown (Fig. 7A), which is consistent with the lack of effect of RCAN1.4 on cell proliferation.

Directed endothelial cell migration in response to VEGF stimulation represents one of the earliest steps in sprouting angiogenesis [46]. Our data shows that RCAN1.4 is able to regulate migration in HDMEC. RCAN1.4 has previously been shown to have a role in angiogenesis *in vitro*, and *in vivo* [10,16]. Here we provide data to show that RCAN1.4 expression is required for efficient tubular morphogenesis in a co-culture model (Fig. 9D). Upon stimulation with VEGF-A, HDMEC undergo tubular morphogenesis, which may be quantified using image analysis software. Following knockdown of RCAN1.4 the ability of HDMECs to undergo tubular morphogenesis was impaired. Though the formation of tubular networks can still occur it would appear that the cells are no longer equipped to fully respond to

VEGF-A stimulation. Similarly, HDMECs plated upon a collagen gel were less able to form tubular networks in response to VEGF-A following RCAN1.4 knockdown (Fig. 9C). Visual analysis of the tubes upon a collagen gel shows that the tubes that are formed appear to be less organised.

RCAN1.4 is known to have a role in the regulation of tumour angiogenesis. Angiogenesis is a multi-step process requiring the proteolytic digestion of the basement membrane, and co-ordinated migration, proliferation, and finally differentiation of endothelial cells into a lumen-containing vessel supported by pericytes [47]. Here we provide evidence that RCAN1.4 is required for the coordination of specific components of angiogenesis.

It has been previously thought that VEGF induces RCAN1.4 via the activation of the Ca^{2+} /calcineurin/NFAT pathway. We now provide evidence for a role of PKC δ in the upregulation of RCAN1.4 expression via an independent pathway (Fig. 10). In addition, we show that VEGF appears unique amongst growth factors in its ability to induce an increase in intracellular Ca^{2+} levels, as well as sustained PKC δ phosphorylation, both of which contribute to the upregulation of RCAN1.4 expression in endothelial cells.

Author Contributions

Conceived and designed the experiments: KH MJC. Performed the experiments: KH EC VS. Analyzed the data: KH EC VS. Contributed reagents/materials/analysis tools: VS. Wrote the paper: KH MJC.

References

- Carmeliet P (2005) Angiogenesis in life, disease and medicine. *Nature* 438: 932–936.
- Holmes K, Roberts OL, Thomas AM, Cross MJ (2007) Vascular endothelial growth factor receptor-2: structure, function, intracellular signalling and therapeutic inhibition. *Cell Signal* 19: 2003–2012.
- Fuentes JJ, Pritchard MA, Planas AM, Bosch A, Ferrer I, et al. (1995) A new human gene from the Down syndrome critical region encodes a proline-rich protein highly expressed in fetal brain and heart. *Hum Mol Genet* 4: 1935–1944.
- Crawford DR, Leahy KP, Abramova N, Lan L, Wang Y, et al. (1997) Hamster adapt78 mRNA is a Down syndrome critical region homologue that is inducible by oxidative stress. *Archives of biochemistry and biophysics* 342: 6–12.
- Rothermel B, Vega RB, Yang J, Wu H, Bassel-Duby R, et al. (2000) A protein encoded within the Down syndrome critical region is enriched in striated muscles and inhibits calcineurin signaling. *J Biol Chem* 275: 8719–8725.
- Fuentes JJ, Genesca L, Kingsbury TJ, Cunningham KW, Perez-Riba M, et al. (2000) DSCR1, overexpressed in Down syndrome, is an inhibitor of calcineurin-mediated signaling pathways. *Hum Mol Genet* 9: 1681–1690.
- Fuentes JJ, Pritchard MA, Estivill X (1997) Genomic organization, alternative splicing, and expression patterns of the DSCR1 (Down syndrome candidate region 1) gene. *Genomics* 44: 358–361.
- Genesca L, Aubareda A, Fuentes JJ, Estivill X, De La Luna S, et al. (2003) Phosphorylation of calcipressin 1 increases its ability to inhibit calcineurin and decreases calcipressin half-life. *Biochem J* 374: 567–575.
- Yang J, Rothermel B, Vega RB, Frey N, McKinsey TA, et al. (2000) Independent signals control expression of the calcineurin inhibitory proteins MCIP1 and MCIP2 in striated muscles. *Circ Res* 87: E61–68.
- Minami T, Horiuchi K, Miura M, Abid MR, Takabe W, et al. (2004) Vascular endothelial growth factor- and thrombin-induced termination factor, Down syndrome critical region-1, attenuates endothelial cell proliferation and angiogenesis. *J Biol Chem* 279: 50537–50554.
- Zhao P, Xiao X, Kim AS, Leite MF, Xu J, et al. (2008) c-Jun inhibits thapsigargin-induced ER stress through up-regulation of DSCR1/Adapt78. *Experimental biology and medicine (Maywood, N.J)* 233: 1289–1300.
- Hogan PG, Chen L, Nardone J, Rao A (2003) Transcriptional regulation by calcium, calcineurin, and NFAT. *Genes Dev* 17: 2205–2232.
- Kingsbury TJ, Cunningham KW (2000) A conserved family of calcineurin regulators. *Genes & development* 14: 1595–1604.
- Chan B, Greenan G, McKeon F, Ellenberger T (2005) Identification of a peptide fragment of DSCR1 that competitively inhibits calcineurin activity *in vitro* and *in vivo*. *Proc Natl Acad Sci U S A*.
- Vega RB, Yang J, Rothermel BA, Bassel-Duby R, Williams RS (2002) Multiple domains of MCIP1 contribute to inhibition of calcineurin activity. *J Biol Chem* 277: 30401–30407.
- Iizuka M, Abe M, Shiiba K, Sasaki I, Sato Y (2004) Down syndrome candidate region 1, a downstream target of VEGF, participates in endothelial cell migration and angiogenesis. *J Vasc Res* 41: 334–344.
- Yao YG, Duh EJ (2004) VEGF selectively induces Down syndrome critical region 1 gene expression in endothelial cells: a mechanism for feedback regulation of angiogenesis? *Biochem Biophys Res Commun* 321: 648–656.
- Back K-H, Zaslavsky A, Lynch RC, Britt C, Okada Y, et al. (2009) Down's syndrome suppression of tumour growth and the role of the calcineurin inhibitor DSCR1. *Nature* 459: 1126–1130.
- Holmes K, Williams CM, Chapman EA, Cross MJ (2010) Detection of siRNA induced mRNA silencing by RT-qPCR: considerations for experimental design. *BMC Res Notes* 3: 53.
- Plevin R, Palmer S, Gardner SD, Wakelam MJ (1990) Regulation of bombesin-stimulated inositol 1,4,5-trisphosphate generation in Swiss 3T3 fibroblasts by a guanine-nucleotide-binding protein. *Biochem J* 268: 605–610.
- Sorrell JM, Baber MA, Caplan AI (2007) A self-assembled fibroblast-endothelial cell co-culture system that supports *in vitro* vasculogenesis by both human umbilical vein endothelial cells and human dermal microvascular endothelial cells. *Cells, tissues, organs* 186: 157–168.
- Takahashi T, Yamaguchi S, Chida K, Shibuya M (2001) A single autophosphorylation site on KDR/Flk-1 is essential for VEGF-A-dependent activation of PLC-gamma and DNA synthesis in vascular endothelial cells. *The EMBO journal* 20: 2768–2778.
- Rahimi N, Dayanir V, Lashkari K (2000) Receptor chimeras indicate that the vascular endothelial growth factor receptor-1 (VEGFR-1) modulates mitogenic activity of VEGFR-2 in endothelial cells. *J Biol Chem* 275: 16986–16992.
- Dayanir V, Meyer RD, Lashkari K, Rahimi N (2001) Identification of tyrosine residues in vascular endothelial growth factor receptor-2/FLK-1 involved in activation of phosphatidylinositol 3-kinase and cell proliferation. *J Biol Chem* 276: 17686–17692.
- Whittles CE, Pocock TM, Wedge SR, Kendrew J, Hennequin LF, et al. (2002) ZM323881, a novel inhibitor of vascular endothelial growth factor-receptor-2 tyrosine kinase activity. *Microcirculation* 9: 513–522.
- Tsien RY (1980) New calcium indicators and buffers with high selectivity against magnesium and protons: design, synthesis, and properties of prototype structures. *Biochemistry* 19: 2396–2404.
- Toullec D, Pianetti P, Coste H, Bellevergue P, Grand-Perret T, et al. (1991) The bisindolylmaleimide GF 109203X is a potent and selective inhibitor of protein kinase C. *J Biol Chem* 266: 15771–15781.
- Qatsha KA, Rudolph C, Marme D, Schachtele C, May WS (1993) Go 6976, a selective inhibitor of protein kinase C, is a potent antagonist of human immunodeficiency virus 1 induction from latent/low-level-producing reservoir cells *in vitro*. *Proc Natl Acad Sci U S A* 90: 4674–4678.

29. Fruman DA, Klee CB, Bierer BE, Burakoff SJ (1992) Calcineurin phosphatase activity in T lymphocytes is inhibited by FK 506 and cyclosporin A. *Proc Natl Acad Sci U S A* 89: 3686–3690.
30. Favata MF, Horiuchi KY, Manos EJ, Daulerio AJ, Stradley DA, et al. (1998) Identification of a novel inhibitor of mitogen-activated protein kinase kinase. *J Biol Chem* 273: 18623–18632.
31. Keranen LM, Dutil EM, Newton AC (1995) Protein kinase C is regulated in vivo by three functionally distinct phosphorylations. *Curr Biol* 5: 1394–1403.
32. Dempsey EC, Newton AC, Mochly-Rosen D, Fields AP, Reyland ME, et al. (2000) Protein kinase C isozymes and the regulation of diverse cell responses. *Am J Physiol Lung Cell Mol Physiol* 279: L429–438.
33. Hernandez GL, Volpert OV, Iniguez MA, Lorenzo E, Martinez-Martinez S, et al. (2001) Selective inhibition of vascular endothelial growth factor-mediated angiogenesis by cyclosporin A: roles of the nuclear factor of activated T cells and cyclooxygenase 2. *J Exp Med* 193: 607–620.
34. Rahman I, MacNee W (1998) Role of transcription factors in inflammatory lung diseases. *Thorax* 53: 601–612.
35. Lindsey KQ, Caughman SW, Olerud JE, Bunnett NW, Armstrong CA, et al. (2000) Neural regulation of endothelial cell-mediated inflammation. *J Invest Dermatol Symp Proc* 5: 74–78.
36. Mellberg S, Dimberg A, Bahram F, Hayashi M, Rennel E, et al. (2009) Transcriptional profiling reveals a critical role for tyrosine phosphatase VE-PTP in regulation of VEGFR2 activity and endothelial cell morphogenesis. *Faseb J* 23: 1490–1502.
37. Niemisto A, Dumire V, Yli-Harja O, Zhang W, Shmulevich I (2005) Robust quantification of in vitro angiogenesis through image analysis. *Ieee Transactions on Medical Imaging* 24: 549–553.
38. Hesser BA, Liang XH, Camenisch G, Yang S, Lewin DA, et al. (2004) Down syndrome critical region protein 1 (DSCR1), a novel VEGF target gene that regulates expression of inflammatory markers on activated endothelial cells. *Blood* 104: 149–158.
39. Qin L, Zhao D, Liu X, Nagy JA, Hoang MV, et al. (2006) Down syndrome candidate region 1 isoform 1 mediates angiogenesis through the calcineurin-NFAT pathway. *Mol Cancer Res* 4: 811–820.
40. Mohammadi M, Honegger AM, Rotin D, Fischer R, Bellot F, et al. (1991) A tyrosine-phosphorylated carboxy-terminal peptide of the fibroblast growth factor receptor (Flg) is a binding site for the SH2 domain of phospholipase C-gamma 1. *Mol Cell Biol* 11: 5068–5078.
41. Nishibe S, Wahl MI, Hernandez-Sotomayor SM, Tonks NK, Rhee SG, et al. (1990) Increase of the catalytic activity of phospholipase C-gamma 1 by tyrosine phosphorylation. *Science* 250: 1253–1256.
42. Okano Y, Mizuno K, Osada S, Nakamura T, Nozawa Y (1993) Tyrosine phosphorylation of phospholipase C gamma in c-met/HGF receptor-stimulated hepatocytes: comparison with HepG2 hepatocarcinoma cells. *Biochem Biophys Res Commun* 190: 842–848.
43. Gliko G, Wheeler-Jones C, Zachary I (2002) Vascular endothelial growth factor induces protein kinase C (PKC)-dependent Akt/PKB activation and phosphatidylinositol 3'-kinase-mediated PKC delta phosphorylation: role of PKC in angiogenesis. *Cell Biol Int* 26: 751–759.
44. Lorenzi O, Frieden M, Villemin P, Fournier M, Foti M, et al. (2008) Protein kinase C-delta mediates von Willebrand factor secretion from endothelial cells in response to vascular endothelial growth factor (VEGF) but not histamine. *Journal of thrombosis and haemostasis: JTH* 6: 1962–1969.
45. Lee HJ, Kim YS, Sato Y, Cho YJ (2009) RCAN1-4 knockdown attenuates cell growth through the inhibition of Ras signaling. *FEBS Lett* 583: 2557–2564.
46. Lamalice L, Le Bocuf F, Huot J (2007) Endothelial cell migration during angiogenesis. *Circ Res* 100: 782–794.
47. Folkman J (1986) How is blood vessel growth regulated in normal and neoplastic tissue? G.H.A. Clowes memorial Award lecture. *Cancer research* 46: 467–473.