Review Article Antiviral Perspectives for Chikungunya Virus

Deepti Parashar and Sarah Cherian

National Institute of Virology, 20-A Dr. Ambedkar Road, Pune 411001, India

Correspondence should be addressed to Deepti Parashar; deeptiparasharster@gmail.com

Received 29 January 2014; Revised 22 April 2014; Accepted 30 April 2014; Published 15 May 2014

Academic Editor: Fumio Imazeki

Copyright © 2014 D. Parashar and S. Cherian. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Chikungunya virus (CHIKV) is a mosquito-borne pathogen that has a major health impact in humans and causes acute febrile illness in humans accompanied by joint pains and, in many cases, persistent arthralgia lasting for weeks to years. CHIKV reemerged in 2005-2006 in several parts of the Indian Ocean islands and India after a gap of 32 years, causing millions of cases. The reemergence of CHIKV has also resulted in numerous outbreaks in several countries in the eastern hemisphere, with a threat to further expand in the near future. However, there is no vaccine against CHIKV infection licensed for human use, and therapy for CHIKV infection is still mainly limited to supportive care as antiviral agents are yet in different stages of testing or development. In this review we explore the different perspectives for chikungunya treatment and the effectiveness of these treatment regimens and discuss the scope for future directions.

1. Introduction

The reemergence of chikungunya virus (CHIKV) in many parts of the world is a significant public health concern. Since the 2005-2006 chikungunya fever epidemic in the Indian Ocean island of La Réunion [1, 2] millions of people in more than 40 countries including India, Malaysia, Indonesia, Thailand, Singapore, the United States, and some European countries have been infected [3–5].

The CHIKV was first isolated from febrile individuals in Tanzania in 1952 [6, 7]. Initially, researchers classified it among viruses such as Sindbis and Semliki Forest virus. Later studies characterized it as an alpha virus that belongs to the family Togaviridae. CHIKV is an enveloped virus with a genome approximately 11.8 kb in size. It consists of a single stranded, positive sense RNA genome with two open reading frames (ORFs) [8]. The one at the 5' end encodes four nonstructural proteins (nsP1–nsP4) and the second ORF at the 3' end encodes the structural proteins, the capsid (C), envelope glycoproteins E1 and E2, and two small cleavage products (E3, 6K).

CHIKV is transmitted to humans by several species of mosquitoes, with *Aedes aegypti* and *A. albopictus* being the two main vectors. The symptoms generally start 4–7 days after the bite. Acute infection lasts for 1–10 days and is

characterized by abrupt onset of fever, headache, fatigue, nausea, vomiting, rash, myalgia, and severe arthralgia. Painful polyarthralgia is the typical symptom and may persist in \sim 10% of cases for several months causing serious economic and social impacts on both the individual and the affected communities in some instances [9].

Thus far, due to the lack of licensed vaccines or effective antivirals against CHIKV, most of the treatment regimens are symptomatic and based on the clinical manifestations. Nonsalicylate analgesics and nonsteroidal anti-inflammatory drugs are most commonly used for symptomatic relief [10]. Immune-based control strategies have also been tested and have shown effectiveness. The elaboration of mouse [11, 12] and nonhuman primate models [13, 14] together with antivirals currently undergoing clinical trials and new approaches involving screening of libraries of small molecules and synthetic compounds with antiviral potential as well as natural products of plants [15] is further providing the stimulus for improving the development of antiviral candidates.

Recently Rashad et al. [16] have presented a detailed review of the current medicinal chemistry aspects supporting the development of chemotherapeutics targeting the CHIKV through drug discovery and design. On the other hand, this review is an attempt to compile and explore the various strategies that have been tried for the management and treatment of CHIKV from a virological perspective and further discuss possible directions for the future.

2. Chemical Compounds

We describe here some of the commonly used nonsteroidal anti-inflammatory drugs (NSAIDs) and nonsalicylate analgesics that have played a major role in symptomatic treatment of the chikungunya disease along with some newer compounds that have been recently tested against the CHIKV infection.

(A) Symptomatic Treatment Strategies

Nonsteroidal Anti-Inflammatory Drugs. Current treatments for viral arthropathies rely mainly on NSAIDs though these often provide only partial relief [17]. A number of patients who experienced chronic rheumatic symptoms following infection with CHIKV during the 2005-2006 La Réunion outbreak were successfully treated with methotrexate (MTX) [18]. MTX, which was originally developed as a chemotherapeutic, forms the basis of most rheumatoid arthritis (RA) treatment regimens due to its anti-inflammatory effects at low doses [19]. Despite this, details of its anti-inflammatory mechanism and its effect on acute viral induced arthritis remain unclear. Some rare chronic patients did benefit from MTX but seem to be in a different diagnostic class "postchikungunya rheumatism" [20]. However, treatment efficacy is hard to evaluate when multiple diseases are present. In a study of post-CHIKV chronic arthritis in Maharashtra, a south-western state of India, antirheumatic drugs including Sulfasalazine and MTX were needed for effective treatment [21]. In a recent study aimed at understanding the antiinflammatory mechanism of MTX and its effect on acute viral induced arthritis, a mouse model of Ross River virus-induced inflammatory disease demonstrated that MTX treatment caused early onset of disease through increased monocyte production [22]. The findings of MTX effectiveness thus seem to be contradictive.

Further, although adverse effects of NSAIDs occur in only a small proportion of users, the widespread use of these drugs has resulted in serious gastrointestinal complications in a substantial overall number of affected persons. Selective COX-II inhibitors such as rofecoxib, celecoxib, and parecoxib have shown consistently comparable efficacy to that of conventional NSAIDs in patients with rheumatoid arthritis, with a significantly reduced propensity to cause gastrointestinal toxicity. The safety benefits of COX-II inhibitors given alone appear similar to combined therapy with conventional NSAIDs and gastroprotective agents, justifying the consideration of such inhibitors as first-line therapy in patients requiring long-term pain control [23].

Nonsalicylate Analgesics (Paracetamol, Acetaminophen, Morphine, and Traditional Herbal Medicine). Most data available regarding the use of nonsalicylate analgesics against the CHIKV is from the follow-up of La Réunion patients. Based on clinical manifestations, the analgesic drug Paracetamol was the most used (in 95.4% of treatments) and often was combined with NSAIDs. The wide use of paracetamol led to the emergence of severe liver afflictions diagnosed during the epidemic, particularly when doses >3 g/day were taken [24]. The hepatotoxicity of paracetamol in combination with interferon and vinblastine has also been reported [25]. Further, studies have shown that acetaminophen (APAP), another analgesic which is commonly used for the relief of fever, and flu-like symptoms, modulates the transcriptional response to recombinant interferon-beta [26]. This also suggests that the use of this class of drugs might need some caution. Morphine was seldom used in the La Réunion although it was reported to be effective during early treatments in a period when the CHIKV was first discovered [27]. Plants are an integral part of the Réunion Island pharmacopeia and therefore were mainly used during the outbreaks to treat fever, pain, and inflammation. Some plant species such as Fernelia spp. are also known for their antiviral properties. However, the efficacy of these plants or association of plants has not been studied, particularly for the hepatotoxic risk that might be present when taken with paracetamol [24].

Steroids. Corticoids were prescribed to treat arthralgia (27.7%) of cases), particularly invalidating forms during the chikungunya disease in the Reunion Island hospital staff [24]. A study carried out in South India during the chikungunya epidemic in 2007 with the objective of evolving a uniform treatment protocol demonstrated that addition of prednisolone to aceclofenac reduced pain and improved the quality of life in patients with acute chikungunya arthritis, compared to aceclofenac given alone in the management of early chikungunya fever. Their studies thus recommended coadministration of low-dose systemic corticosteroids with NSAIDs as the best regimen in treating acute chikungunya cases with arthralgia [28]. In another report, regarding a case of post chikungunya reversible demyelinating encephalitis that was presented with vertigo, dysarthria, and ataxia, there was complete clinical as well as radiological improvement with steroids [29]. However, corticosteroid use during acute viral arthritis is considered to be contraindicated as a result of the risk of immunosuppression causing enhanced infection and disease exacerbation [30].

(B) Specific Chemical Compounds Tested against CHIKV

2.1. Chloroquine. Chloroquine was first reported to inhibit Sindbis virus (SINV) and Semliki Forest Virus (SFV) infectivity in vitro more than 35 years ago [31-35], but studies in mice suggested that the drug might enhance viral replication and aggravate the disease [36]. However, chloroquine was found to be effective in treating chronic CHIKV-induced arthralgia with the possible mechanism attributed most likely to its antiinflammatory properties that are adapted for the treatment of some autoimmune illnesses [37]. Recent research on the efficacy of chloroquine has focused on the dosage used to treat acute CHIKV infections [38, 39]. To advance the studies with chloroquine and CHIKV, a double blind placebo-controlled randomized trial [38, 40] was conducted in the Réunion Island. No statistical difference was observed between the chloroquine and placebo groups, either in mean duration of febrile arthralgia or rate of decrease of viraemia. However, the number of patients included in the study was too small to draw definitive conclusions regarding the efficacy of the chloroquine treatment. In another in vitro study [41] since chloroquine exerted its antiviral effects in all the three modes of treatment (pretreatment, concurrent, and posttreatment), it was suggested that it has prophylactic and therapeutic potentials. However, in a recent controlled evaluation of chloroquine in treating patients with early persistent musculoskeletal pain and arthritis following CHIKV infection, selected cytokine assays that were carried out to study the inflammatory nature and the response of the symptoms to it, do not support a meaningful therapeutic role of oral chloroquine in the management of the symptoms studied [42]. Further, the studies carried out by Padmakumar et al. [28] also suggested that coadministration of hydroxychloroquine with NSAIDs in acute stages of the disease does not offer any additional benefits.

2.2. Ribavirin and 6-Azauridine. Ribavirin has shown wide in vitro inhibitory activity against RNA viruses with different modes of action depending on the virus [40]. Some of the suggested mechanisms of action are through the inhibition of IMP dehydrogenase, error catastrophe mechanism, by causing mutations and by interaction with viral polymerase [43]. Ravichandran and Manian [44] determined whether antiviral treatment would make any difference in those patients who continue to have arthritis even after two weeks after the febrile episode. Preliminary observations showed that ribavirin may have a direct antiviral activity against chikungunya. Compared to ribavirin, 6-azauridine, a broadspectrum antimetabolite, was more effective against CHIKV and showed a similar antiviral activity against SFV [45, 46]. Further, interferon-alpha and ribavirin in combination also showed a synergistic effect on the in vitro inhibition of CHIKV [46]. Human infection with CHIKV appears to induce immunological dysfunction. Though interferon usually boosts the immune response, caution in its usage is recommended until more extensive nonhuman primate models have been studied. Polyethylene glycol-conjugated (pegylated, PEG) alpha interferon appears to be an effective treatment against infection with Venezuelan equine encephalitis virus (VEEV) and has profound effects on the host immune response to infection [47]. Treatment of VEEVinfected BALB/c mice with PEG IFN- α resulted in a greatly enhanced survival from either a subcutaneous or an aerosol infection. Treatment, results in a number of changes to the immune response characteristics normally associated with VEEV infection including increased macrophage activation, absence of splenic CD4, CD8, and B cells by day 2 post infection normally associated with VEEV infection, and reduction in the high tumor necrosis factor alpha production by macrophages.

In an animal model study for CHIKV, Couderc et al. [11] have demonstrated that adult mice with a partially or totally abrogated type-I IFN pathway develop a mild or severe infection; thus, it might be justified to test pegylated alpha interferon on CHIKV and other pathogenic alphaviruses. 2.3. Arbidol. The antiviral drug arbidol (ARB) was originally developed at the Russian Research Chemical and Pharmaceutical Institute about 20 years ago. This drug has been used in Russia for prophylaxis and treatment of acute respiratory infection including influenza from 1990. It has been shown that ARB exhibits a wide range of activities against a number of RNA and DNA enveloped/nonenveloped viruses. In cellulo, antiviral ARB activity against CHIKV has been investigated [48]. This compound was found to present potent inhibitory activity against the virus propagated onto immortalized primary human fibroblast and vero cells. Their study also demonstrated the growth of an ARB resistant CHIKV mutant possessing a single amino acid substitution (G407R) localized in the E2 envelope protein. Though this study helps explain the mechanism of action of arbidol, it also brings out the point that ARB usage may be impaired as in vivo there could be selection of a resistant virus that is able to be disseminated.

2.4. Harringtonine. In a recent study, harringtonine, a cephalotaxine alkaloid, displayed potent inhibition of CHIKV infection with minimal cytotoxicity. Treatment of harringtonine against SINV, a related alphavirus, suggested that harringtonine could inhibit other alphaviruses as well. [49]. It was indicated that harringtonine inhibited an early stage of the CHIKV replication cycle which occurred after viral entry into cells. Interestingly, harringtonine displayed greater potency with a CHIKV strain carrying the E1 A226V mutation as compared with a strain carrying the wild type E1.

Table 1 presents a summary of the assays used, the hypothesized target, and the pros and cons of the compounds described above.

3. Inhibitors Based on RNA-Mediated Interference

RNA interference (RNAi) is a posttranscriptional process triggered by the introduction of double-stranded RNA (dsRNA) which leads to gene silencing in a sequence-specific manner. The method is based on targeting specific viral proteins which consequently leads to the shutdown of the protein expression process, thereby stopping viral replication. RNAi mediated inhibition of viral replication has emerged as a promising antiviral strategy. The small interfering RNA (siRNA) and small hairpin RNA (shRNA) molecules are central to RNA interference.

Small interfering RNAs (siRNAs) are formed by cleaving the dsRNAs by the RNase III family member, Dicer, into 19– 23 nucleotide (nt) fragments with 5' phosphorylated ends and 2-nt unpaired and unphosphorylated 3' ends. The processing of the target RNA is achieved by endonuclease argonaute 2 (Ago 2) which is further incorporated into the RNAi specificity complex (RISC). On the other hand, plasmidexpressed short hairpin RNA (shRNA) requires the activity of endogenous exportin 5 and Ago2 (Argonaute 2) that forms a dimer with Dicer, which then receives the shRNA. The shRNA is cleaved in one step by Dicer generating a 19–23 nt duplex siRNA with 2 nt 3' overhangs. The siRNAs guide RISC to the target mRNA. RISC delivers the mRNA to

Products	Assay type	Hypothesized target	Pros	Cons	References
Chloroquine	<i>In vitro</i> (vero cells)	Disrupted endosome-mediated CHIKV internalization, possibly through the prevention of endosomal acidification.	<i>In vitro</i> study proved that it blocks the production of proinflammatory cytokines and the proliferation of monocytes, macrophages, and lymphocytes.	<i>In vivo</i> study required.	Delogu and de Lamballerie, 2011 [50] Khan et al., 2010 [41]
Ribavirin	Human	Can interact with the intracellular viral RNA production.	Faster resolution of joint and soft tissue manifestations.	Involvement of a small number of patients and lack of planning as randomly distributed patients were not compared with a placebo group.	Ravichandran and Manian, 2008 [44]
6-Azauridine	In vitro (vero cells)	Inhibition of orotidine monophosphate decarboxylase, an enzyme involved in the de novo biosynthesis of pyrimidine, cytidine, and thymidine.	Showed a significant inhibition of CHIKV at a low concentration.	The antiviral activity has been difficult to replicate <i>in vivo.</i>	Briolant et al., 2004 [46]
Arbidol	<i>In vitro</i> (vero and primary human fibroblast cells)	Inhibition of virus mediated fusion and blocking of the viral entry into the target cells through inhibition of glycoprotein conformational changes that are essential for the fusion process.	Well-tolerated with minimal side effects.	Not tested in <i>in vivo</i> system.	Delogu et al., 2011 [48]
Harringtonine	<i>In vitro</i> (BHK21 cells)	Affects CHIKV RNA production inside the infected cell as well as viral protein expression such as the nsP3 and the E2 proteins.	Minimal cytotoxicity.	Not tested in <i>in vivo</i> system.	Kaur et al., 2013 [49]

TABLE 1: Major tested anti-CHIKV chemical compounds.

cytoplasmic foci named processing bodies (P-bodies or GWbodies), wherein mRNA decay factors are concentrated. The target mRNA is cleaved by Ago2 and degraded.

3.1. Small Interfering RNA (siRNA) Molecules. The therapeutic application of siRNA for the inhibition of CHIKV replication has been examined in vero cells [51]. siRNAs against the conserved regions of nsP3 and E1 genes of CHIKV have been designed and siRNA activity was assessed by detecting both the infectious virus and its genome. The titers of the virus yield in the supernatant were determined at 24 and 48 hours postinfection. Results showed reduction of virus titre by ~1.5 log10- ~2.5 log10 in comparison to the control. The study thus showed limited success and further needs the investigation of these siRNA sequences in an in vivo model for their ultimate therapeutic application. Studies have also been conducted at National Institute of Virology, Pune, India, by designing eight siRNAs, either targeting the E2 or the ns1 genes of CHIKV. The efficiency of these siRNAs to inhibit the CHIKV production was assessed in vero cells. Two of the siRNAs were able to reduce CHIKV E3 transcripts by ~5log10 and ~2.5log10 when cells were

transfected after 1 hour post infection. 100 pmol of siRNA was found optimum in perturbing CHIKV production. The combination of these siRNAs was found to be very effective in inhibiting the production of CHIKV. Importantly, these siRNAs could inhibit the CHIKV replication in Swiss albino mice when administered 72 hours after viral infection. A siRNA dose of 1 mg/kg body weight (1700 pmol) was found optimal to inhibit the CHIKV replication in Swiss albino mice. Our findings suggested that RNAi is capable of silencing sequence-specific genes of CHIKV and might constitute a new therapeutic strategy for CHIKV infection [52].

3.2. Small Hairpin RNA (shRNA) Molecules. The efficacy of plasmid-based short hairpin RNA (shRNA) against CHIKV replication in three CHIKV-permissive cell lines, namely, HeLa, RD, and BHK cells has been investigated. Cell clones expressing shRNAs against CHIKV E1 and nsP1 genes displayed significant inhibition of infectious CHIKV production, while shRNA capsid demonstrated a modest inhibitory effect as compared to scrambled shRNA cell clones and nontransfected cell controls. Analysis of CHIKV E2 protein expression and visualization of shRNA (E1 and nsP1) cell

clones collectively demonstrated similar inhibitory trends against CHIKV replication. shRNA E1 showed non cell-type specific anti-CHIKV effects and broad-spectrum silencing against different geographical strains of CHIKV. Furthermore, shRNA E1 clones did not exert any inhibition against dengue virus and SINV replication, thus indicating the high specificity of shRNA against CHIKV replication. Moreover, no shRNA-resistant CHIKV mutant was generated after 50 passages of CHIKV in stable cell clones. More importantly, strong and sustained anti-CHIKV protection was conferred in suckling mice pretreated with different concentrations (3.1, 9.5 and 19 pmol) of shRNA (E1). Results have thus suggested the promising efficacy of anti-CHIKV shRNAs, in particular, plasmid-shRNA E1, as a novel antiviral strategy against CHIKV infection [53].

4. Cellular Factors

Another approach to inhibiting the CHIKV infection is by targeting cellular factors such as proteases that are involved in the processing of the CHIKV particles and cellular receptors that may have a role in viral replication, or in another way by the induction of immune-based cellular enzymes that possess antiviral activity.

4.1. Furin Inhibitors. Molecules that inhibit alphavirus entry into susceptible cells by blocking the intracellular cleavage of viral envelope glycoproteins have been investigated. CHIKV infection of cultured human cells was shown to be inhibited by impairing the maturation of the CHIKV E2 surface glycoprotein using the furin inhibitor peptide, decanoyl-RVKR-chloromethyl ketone [54]. Another study reported the possible inhibition of the replication of VEEV using polyclonal antibodies to laminin-binding protein [55].

4.2. 2',5'-Oligoadenylate Synthetase (OAS3). CHIKV is highly sensitive to the antiviral activity of Type-I interferons (IFN- α/β). Bréhin et al. [56] investigated the role of IFNinduced 2',5'-oligoadenylate synthetase (OAS) family in innate immunity to CHIKV. They have established inducible human epithelial HeLa cell lines expressing either the large form of human OAS and OAS3 or the genetic variant OAS3-R844X which is predicted to lack about 20% of the OAS3 protein from the carboxy terminus. HeLa cells respond to ectopic OAS3 expression by efficiently inhibiting CHIKV growth. The characteristic of the antiviral effect was a blockade in early stages of virus replication. Thus, OAS3 pathway may represent a novel antialphaviral mechanism by which IFN- α/β controls CHIKV growth. HeLa cells expressing the truncated form of OAS3 were less resistant to CHIKV infection, raising the question on the involvement of OAS3 genetic polymorphism in human susceptibility to alphavirus infection [56]. In another study [57] a CHIKV variant exhibiting remarkable resistance towards the antiviral activity of OAS3, through an enhancement of viral RNA replication, was identified. It was shown that a single amino acid change in the E2 glycoprotein allows the rescue of viral growth in OAS3 expressing HeLa cells by acting on the early

stages of viral life cycle. These observations also provide a new insight into the role of E2 into the pathogenicity of CHIKV in human cells [57].

4.3. Cellular IMPDH Enzyme. Mycophenolic acid (MPA) is a weak organic acid and a well-known immunosuppressive agent. It has been investigated as an effective agent against growth and multiplication of several microbial pathogens including viruses [58]. MPA inhibits the replication of several viruses *in vitro* to varying degrees [59]. MPA is a noncompetitive inhibitor of inosine monophosphate dehydrogenase (IMPDH). MPA is used clinically in the prevention of rejection of transplanted organs [60]. Various studies have reported that MPA blocked the cytopathic effect and replications of several viruses [61].

Khan et al. [62] have assessed the antiviral potential of MPA against CHIKV via inhibition of IMPDH enzyme in vero cells. Inhibition of virus induced apoptosis was observed as measured by caspase-3, PARP, and Bcl-2. Total genome infectivity was determined by analyzing the ratio of total infectious viral particles to the genome copy number. Nontoxic concentration of MPA (10 μ M) reduced \geq 99.9% CHIKV titre in vero cells. MPA via depletion of substrate for polymerase (GTP) inhibited CHIKV-induced apoptosis.

4.4. Viperin. The antiviral role of viperin has been demonstrated in CHIKV infection by studying the longitudinal transcriptional profiles of the innate immune response in PBMCs from a cohort of 24 CHIKV-infected patients. Results showed that type I IFNs controlled CHIKV infection via RSAD2 (which encodes viperin), an enigmatic multifunctional IFNstimulated gene (ISG). Viperin was highly induced in monocytes, the major target cell of CHIKV in blood. *In vivo* study demonstrated the direct role of viperin in controlling CHIKV replication. Mice lacking Rsad2 had higher viremia and severe joint inflammation compared with wild-type mice [63].

4.5. Human Antibodies. Immunotherapy, in the form of human polyclonal antibody, has been used for treatment of human viral infections, and in alphavirus-infected animal models, passive immunization with convalescent sera from animals was protective. Sera from patients and monkeys who have recovered from acute alphavirus-infections have been shown to contain neutralizing antibodies in suckling mouse models. Human polyclonal antibodies (CHIKV Ig) have been purified from plasma of convalescent donors and used in mouse models of CHIKV infection [64]. Results are promising with CHIKV Ig showing both prophylactic and therapeutic potential. In both IFN- α/β R-/- and immunecompetent mouse neonates, a single prophylactic dose of CHIKV Ig was found to be protective against lethality associated with CHIKV, with undetectable viral levels in the serum and no dissemination to the central nervous system. The degree of protection correlated to the dose of antibodies administered. CHIKV Ig also has a therapeutic effect as it is protective against lethality when given up to 8 hours postinfection. The results also supported the hypothesis that viremia precedes CNS dissemination and controlling the viremia prevents neurologic complications [64].

From another point of view, the therapeutic utility of monoclonal/polyclonal antibodies might be limited as patients with a serodiagnosis of an alphaviral disease clearly already have antibodies, and administration of antibodies before endogenous antibody production is likely to be impractical. Nevertheless, such antibodies might find utility in preventing mother-to-child infections. Conceivably, neutralizing antibodies could be used prophylactically during an epidemic. However, parenterally administered antibodies (such as tocilizumab and infliximab) have serum half-lives of only 8–14 days, so repeated dosing would be envisaged in an extended epidemic [65].

A summary of the assays used, the hypothesized target, and the pros and cons of the cellular inhibitors described herein is presented in Table 2.

5. High Throughput Screening and Structure Based Drug Discovery Approaches

Recently, cell-based high throughput assays have been developed to identify potential CHIKV inhibitors. One study reported a focus screen of 356 natural compounds and clinically approved drugs using a CHIKV replicon and a concomitant screen with SFV surrogate infection model [66], while another study screened 3,040 small molecules for inhibitors of CHIKV nsP2 using a novel target-based phenotypic assay approach [67]. Cruz et al. [68] used a cell-based high throughput screening assay using resazurin against a kinase inhibitor library combined with the imagebased high content assay approach to identify compounds against CHIKV having novel antiviral activity.

In addition, the availability of the crystal structures of several proteins of the CHIKV RNA genome and other related alphaviruses is encouraging the drug discovery process through target structure-based pharmacophore modeling, virtual library screening, and drug docking approaches. When used prior to experimental screening, it can be considered as a powerful computational filter for reducing the size of a chemical library that can be further experimentally tested [69]. Upon the identification of compound hits through such screening, structural models are further used to develop a structure activity relationship to optimize the compound's activity [70]. Rashad and Keller [71] used in silico techniques of virtual screening and docking studies for structure based design towards the identification of novel binding sites and inhibitors for the chikungunya virus envelope proteins. Based on multiple enzymatic activities and functional roles of the nsP2 [72, 73], recent studies have considered it as a potential target for CHIKV inhibitors [48, 56]. Bassetto et al. [74] used a bioinformatics approach considering a homology-based model of CHIKV virus based on the crystal structure of nsP2 of the alphavirus VEE as a template. The study identified a few compounds that selectively inhibited CHIKV in a virus-cell based CPE reduction assay, though further experimental studies are ongoing to prove that the compounds are indeed nsP2 inhibitors. Lucas-Hourani et al. [67], on the other hand, used a phenotypic assay to identify

one natural compound that partially blocks nsP2 activity and inhibits CHIKV replication *in vitro*. A recent work [75] showed that the nsP4 protein of CHIKV is involved in the mechanism of action of T-705 (favipiravir) that was seen to inhibit CHIKV replication *in vitro* and *in vivo*. Thus it is imperative to target other viral proteins also for structurebased drug design.

6. Antiviral Vaccine Approaches

An alternative approach to virus disease control involves the use of vaccines. There is still no effective vaccine to prevent the disease. In 1967, formalin-inactivated CHIKV vaccines were prepared in chick embryo and suckling mouse brain in 1967 and 1972, and in green monkey kidney cell tissue culture [76, 77]. In 1970, CHIKV vaccine was prepared by tween ether extraction [78]. In 1973, Nakao and Hotta [79] indicated that the UV-inactivated virus was superior to the formalinized virus in regard to its immunogenicity in monkeys. In 2000, Edelman et al. [80] published a phase II assay with an attenuated vaccine (strain obtained from an infected patient during the 1962 outbreak in Thailand).

A number of preclinical CHIKV vaccines have been described, including inactivated virus formulations [81–83], live-attenuated virus vaccines [84–86], chimeric virus vaccines [87], DNA vaccines [88, 89], T cell based peptide vaccine [90], a recombinant adenovirus vaccine [91], subunit protein vaccines [92–94], and a virus-like particle (VLP) formulation [95, 96].

The immunogenic potential of recombinant CHIKV envelope proteins has been evaluated in mice. Recombinant protein elicited a strong humoral response and a balanced Th1/Th2 response. Recombinant CHIKV antigens can be proposed as potent subunit vaccine candidates [97].

Live CHIKV vaccine (CHIKV/IRES) that is highly attenuated yet immunogenic in mouse models and is incapable of replicating in mosquito cells has been developed [98]. The development and evaluation of CHIKV vaccine candidates that were attenuated by deleting a large part of the gene encoding nsP3 or the entire gene encoding 6K have been described. This stable and efficient attenuated CHIKV vaccine candidate can be administered either as viral particles or as infectious genomes launched by DNA [99]. Recently two live-attenuated vaccine candidates based on the insertion of a picornavirus internal ribosome entry site (IRES) sequence into the genome of CHIKV have been generated. These CHIKV/IRES vaccine candidates appear to be safe and efficacious, supporting their strong potential as a human vaccine to protect against CHIKV infection and reduce transmission and further spread [100]. The immunogenicity profile and the efficacy of a novel CHIKV vaccine candidate based on the highly attenuated poxvirus vector modified vaccinia virus Ankara (MVA) expressing the CHIKV structural genes C, E3, E2, 6K, and E1 (termed MVA-CHIKV) have been generated and characterized [101]. MVA-CHIKV was found to be an effective vaccine against CHIKV infection, and it induced strong, broad, highly polyfunctional and longlasting CHIKV-specific CD8⁺ T cell responses, together with

Cellular factors	Assays	Target/effectors	Pros	Cons	References
Furin inhibitors	<i>In vitro</i> (myoblast cells).	Intracellular furin-mediated cleavage of viral envelope glycoproteins: the E2E3 or p62 precursor.	Able to induce a stronger inhibition of viral infection.	Not tested in <i>invivo</i> system.	Ozden et al., 2008 [54]
2′,5′-Oligoadenylate synthetase (OAS3)	Human epithelial HeLa cell lines.	Affects CHIKV replication through a RNase L-dependent pathway.	Ability of OAS3 to inhibit alphavirus growth may be important for the development of antiviral molecules against CHIKV.	Cannot rule out the possibility that OAS3-mediated inhibition of CHIKV was also due to a block early in virus life cycle, for example, viral entry and uncoating of virus particles.	Bréhin et al., 2009 [56]
Cellular IMPDH enzyme	<i>In vitro</i> (vero cells).	Depletion of intracellular guanosine pool.	CHIKV utilizes IMPDH activity for its growth and multiplication which is a potential and effective target to prevent its infection.	It would be useful to explore similar findings by targeting IMPDH in case of other alphaviruses which are more lethal than chikungunya like Sindbis virus, Semliki forest virus, and so forth.	Khan et al., 2011 [62]
Viperin	In vivo (monocytes).	Endoplasmic reticulum.	Critical antiviral host protein that controls CHIKV infection and provides a preclinical basis for the design of effective control strategies against CHIKV.	Large gaps in our understanding of the precise mechanisms at play for viperin to exert such a wide variety of roles within the cell.	Teng et al., 2012 [63]

TABLE 2: Some of the major cellular inhibitors against chikungunya virus.

neutralizing antibodies against CHIKV. A recent paper [102] has reviewed several vaccine approaches for CHIKV.

7. Conclusions

The explosive chikungunya fever epidemic of 2005 in La Réunion and its rapid dissemination to several countries in the Indian Ocean stressed the need for appropriate CHIKV antivirals. In the sections above, we have tried to explore the different perspectives for chikungunya management and treatment and the effectiveness of these regimens. Though advances in molecular and bioinformatics tools have expedited in identifying novel drug and vaccine candidates, the major thrust that is now required is to explore the efficacy of these for CHIKV treatment. As shown in Tables 1 and 2, evaluation studies in *in vivo* systems and further clinical testing of lead compounds are essential for establishing the effectiveness of the potential compounds.

In the management of CHIKV arthralgia, the differential diagnosis of CHIKV, against presence of other conditions such as chronic rheumatic diseases, would be important to treat patients suffering from arthropathy. Among the chemical compounds that have been attempted for CHIKV, some of the well-known broad-spectrum antivirals like chloroquine, ribavirin, and interferon, which have displayed some efficacy in humans, may prove to be promising. Though there is no evidence supporting the clinical efficacy of ribavirin on CHIKV, the combination with interferon could be subjected to clinical trials for the treatment of CHIKV infections. Further, the in vitro use of siRNA or plasmid based shRNAs has shown promising results in the cell based infection model of CHIKV. In spite of this, further studies are warranted to rationalize the therapeutic significance of these siRNAs in primates. The data obtained from the studies related to cellular factors could form the basis for future development of novel immune-based control strategies. The chemical structure of the furin inhibiting peptide could be used as a starting point for development of more novel and active peptidomimetics that target the intracellular furin cleavage step. Compounds that have emerged from virtual library screening and structure-based drug discovery would be expected to display lower toxicity and side effects owing to their selectivity to specific viral targets. Some of the issues related to vaccine development are the requirement of further in vivo evaluation, the need for clinical trials, and cost and scalability issues. Though the MVA-CHIKV may be considered as a potential vaccine candidate against CHIKV, considering the essential role for B cells and antibodies in controlling CHIKV infection, eliciting strong humoral response through the use of suitable adjuvants would be vital for improvement of the vaccine efficacy.

Nevertheless, fighting CHIKV is not only a matter of drugs and vaccines. Preventive measures should also be applied to avoid mosquito bites responsible for disease transmission. These simple measures include the use of insect repellents, the minimization of skin surface exposed to mosquito bites, the elimination of standing water where mosquitoes can lay eggs, and the installation of window and door screens. Precise measures need to be implemented by the government in the form of surveillance and awareness programs among people and medical practitioners for better management of the disease and for the prevention of new outbreaks.

Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

References

- I. Schuffenecker, I. Iteman, A. Michault et al., "Genome microevolution of Chikungunya viruses causing the Indian Ocean outbreak," *PLoS Medicine*, vol. 3, no. 7, pp. 1058–1070, 2006.
- [2] M. Sourisseau, C. Schilte, N. Casartelli et al., "Characterization of reemerging Chikungunya virus," *PLoS Pathogens*, vol. 3, article e89, 2007.
- [3] H. C. Hapuarachchi, K. B. A. T. Bandara, S. D. M. Sumanadasa et al., "Re-emergence of Chikungunya virus in South-east Asia: virological evidence from Sri Lanka and Singapore," *Journal of General Virology*, vol. 91, no. 4, pp. 1067–1076, 2010.
- [4] V. A. Arankalle, S. Shrivastava, S. Cherian et al., "Genetic divergence of Chikungunya viruses in India (1963–2006) with special reference to the 2005-2006 explosive epidemic," *Journal* of General Virology, vol. 88, no. 7, pp. 1967–1976, 2007.
- [5] S. S. Cherian, A. M. Walimbe, S. M. Jadhav et al., "Evolutionary rates and timescale comparison of Chikungunya viruses inferred from the whole genome/E1 gene with special reference to the 2005–07 outbreak in the Indian subcontinent," *Infection, Genetics and Evolution*, vol. 9, no. 1, pp. 16–23, 2009.
- [6] E. G. S. J. Strauss, Ed., Structure and Replication of the Alphavirus Genome, Plenum Press, New York, NY, USA, 1986.
- [7] C. Peters and J. Dalrymple, "Alphaviruses," in *Fields Virology*, B. N. Fields and D. M. Knipe, Eds., pp. 713–761, Raven Press, New York, NY, USA, 2nd edition, 1990.
- [8] S. K. Singh and S. K. Unni, "Chikungunya virus: host pathogen interaction," *Reviews in Medical Virology*, vol. 21, no. 2, pp. 78– 88, 2011.
- [9] P. G. Jupp and B. M. McIntosh, "Chikungunya virus disease," in *The Arboviruses: Epidemiology and Ecology*, T. P. Monath, Ed., pp. 137–157, CRC Press, Boca Raton, Fla, USA, 1988.
- [10] G. Pialoux, B. A. Gaüzère, S. Jauréguiberry, and M. Strobel, "Chikungunya, an epidemic arbovirosis," *The Lancet Infectious Disease*, vol. 7, no. 5, pp. 319–327, 2007.
- [11] T. Couderc, F. Chrétien, C. Schilte et al., "A mouse model for Chikungunya: young age and inefficient type-I interferon signaling are risk factors for severe disease," *PLoS Pathogens*, vol. 4, no. 2, 2008.
- [12] S. A. Ziegler, L. Lu, A. P. A. Travassos Da Rosa, S.-Y. Xiao, and R. B. Tesh, "An animal model for studying the pathogenesis of Chikungunya virus infection," *The American Journal of Tropical Medicine and Hygiene*, vol. 79, no. 1, pp. 133–139, 2008.

- [13] P. Roques, C. Joubert, and B. Malleret, "Chikungunya et autres arboviroses émergentes enmilieu tropical," Reunion Island, http://www.invs.sante.fr/agenda/colloquechikungunya/, 2007.
- [14] K. Labadie, T. Larcher, C. Joubert et al., "Chikungunya disease in nonhuman primates involves long-term viral persistence in macrophages," *Journal of Clinical Investigation*, vol. 120, no. 3, pp. 894–906, 2010.
- [15] Y. Li, L. Wang, S. Li et al., "Seco-pregnane steroids target the subgenomic RNA of alphavirus-like RNA viruses," *Proceedings* of the National Academy of Sciences of the United States of America, vol. 104, no. 19, pp. 8083–8088, 2007.
- [16] A. A. Rashad, S. Mahalingam, and P. A. Keller, "Chikungunya virus: Emerging targets and new opportunities for medical chemistry," *Journal of Medicinal Chemistry*, vol. 57, no. 4, pp. 1147–1166, 2014.
- [17] A. Suhrbier and M. L. Linn, "Clinical and pathologic aspects of arthritis due to Ross River virus and other alphaviruses," *Current Opinion in Rheumatology*, vol. 16, no. 4, pp. 374–379, 2004.
- [18] J.-J. Hoarau, M.-C. J. Bandjee, P. K. Trotot et al., "Persistent chronic inflammation and infection by Chikungunya arthritogenic alphavirus in spite of a robust host immune response," *Journal of Immunology*, vol. 184, no. 10, pp. 5914–5927, 2010.
- [19] M. E. Weinblatt, B. N. Weissman, D. E. Holdsworth et al., "Longterm prospective study of methotrexate in the treatment of rheumatoid arthritis: 84-Month update," *Arthritis and Rheumatism*, vol. 35, no. 2, pp. 129–137, 1992.
- [20] F. Simon and E. Javelle, "Administrative issues linked to health insurance coverage of chronic post-Chikungunya rheumatism," *Medecine Tropicale*, vol. 72, pp. 99–102, 2012.
- [21] M. A. Ganu and A. S. Ganu, "Post-Chikungunya chronic arthritis—our experience with dmards over two year follow up," *Journal of Association of Physicians of India*, vol. 59, no. 2, pp. 83–86, 2011.
- [22] A. Taylor, K. C. Sheng, and L. J. Herrero, "Methotrexate treatment causes early onset of disease in a mouse model of ross river virus-induced inflammatory disease through increased monocyte production," *PLoS ONE*, vol. 8, no. 8, article e71146, Article ID 007114, 2013.
- [23] M. Lazzaroni and G. Bianchi Porro, "Gastrointestinal sideeffects of traditional non-steroidal anti-inflammatory drugs and new formulations," *Alimentary Pharmacology and Therapeutics*, vol. 20, no. 2, pp. 48–58, 2004.
- [24] F. Staikowsky, K. Le Roux, I. Schuffenecker et al., "Retrospective survey of Chikungunya disease in Réunion Island hospital staff," *Epidemiology and Infection*, vol. 136, no. 2, pp. 196–206, 2008.
- [25] P. Kellokumpu-Lehtinen, E. Iisalo, and E. Nordman, "Hepatotoxicity of paracetamol in combination with interferon and vinblastine," *The Lancet*, vol. 1, no. 8647, p. 1143, 1989.
- [26] A. Farnsworth, A. S. Flaman, S. S. Prasad et al., "Acetaminophen modulates the transcriptional response to recombinant interferon-beta," *PLoS ONE*, vol. 5, no. 6, Article ID e11031, 2010.
- [27] M. C. Robinson, "An epidemic of virus disease in Southern Province, Tanganyika territory, in 1952-1953," *Transactions of the Royal Society of Tropical Medicine and Hygiene*, vol. 49, no. 1, pp. 28–32, 1955.
- [28] B. Padmakumar, J. B. Jayan, R. M. R. Menon, B. Krishnankutty, R. Payippallil, and R. S. Nisha, "Comparative evaluation of four therapeutic regimes in Chikungunya arthritis: a prospective randomized parallel-group study," *Indian Journal of Rheumatology*, vol. 4, no. 3, pp. 94–101, 2009.

- [29] L. A. Gauri, B. L. Ranwa, K. Nagar, A. Vyas, and Q. Fatima, "Post Chikungunyabrain stem encephalitis," *Journal of Association of Physicians of India*, vol. 60, no. 4, pp. 68–69, 2012.
- [30] A. D. Mylonas, D. Harley, D. M. Purdie et al., "Corticosteroid therapy in an alphaviral arthritis," *Journal of Clinical Rheumatology*, vol. 10, no. 6, pp. 326–330, 2004.
- [31] S. Cassell, J. Edwards, and D. T. Brown, "Effects of lysosomotropic weak bases on infection of BHK-21 cells by Sindbis virus," *Journal of Virology*, vol. 52, no. 3, pp. 857–864, 1984.
- [32] K. Coombs, E. Mann, J. Edwards, and D. T. Brown, "Effects of chloroquine and cytochalasin B on the infection of cells by Sindbis virus and vesicular stomatitis virus," *Journal of Virology*, vol. 37, no. 3, pp. 1060–1065, 1981.
- [33] A. Helenius, M. Marsh, and J. White, "Inhibition of Semliki Forest virus penetration by lysosomotropic weak bases," *Journal* of General Virology, vol. 58, no. 1, pp. 47–61, 1982.
- [34] A. D. Inglot, "Comparison of the antiviral activity in vitro of some non-steroidal anti-inflammatory drugs," *Journal of General Virology*, vol. 4, no. 2, pp. 203–214, 1969.
- [35] Y. Shimizu, S. Yamamoto, M. Homma, and N. Ishida, "Effect of chloroquine on the growth of animal viruses," *Archiv für die* gesamte Virusforschung, vol. 36, no. 1-2, pp. 93–104, 1972.
- [36] R. K. Maheshwari, V. Srikantan, and D. Bhartiya, "Chloroquine enhances replication of Semliki Forest virus and encephalomyocarditis virus in mice," *Journal of Virology*, vol. 65, no. 2, pp. 992–995, 1991.
- [37] S. W. Brighton, "Chloroquine phosphate treatment of chronic Chikungunya arthritis. An open pilot study," *South African Medical Journal*, vol. 66, no. 6, pp. 217–218, 1984.
- [38] X. De Lamballerie, V. Boisson, J.-C. Reynier et al., "On Chikungunya acute infection and chloroquine treatment," *Vector-Borne* and Zoonotic Diseases, vol. 8, no. 6, pp. 837–839, 2008.
- [39] A. Savarino, R. Cauda, and A. Cassone, "On the use of chloroquine for Chikungunya," *The Lancet Infectious Diseases*, vol. 7, no. 10, p. 633, 2007.
- [40] P. Leyssen, E. De Clercq, and J. Neyts, "The anti-yellow fever virus activity of ribavirin is independent of error-prone replication," *Molecular Pharmacology*, vol. 69, no. 4, pp. 1461–1467, 2006.
- [41] M. Khan, S. R. Santhosh, M. Tiwari, P. V. Lakshmana Rao, and M. Parida, "Assessment of *in vitro* prophylactic and therapeutic efficacy of chloroquine against Chikungunya virus in Vero cells," *Journal of Medical Virology*, vol. 82, no. 5, pp. 817–824, 2010.
- [42] A. Chopra, M. Saluja, and A. Venugopalan, "Effectiveness of chloroquine and inflammatory cytokine response in patients with early persistent musculoskeletal pain and arthritis following Chikungunya virus infection," *Arthritis & Rheumatology*, vol. 66, no. 2, pp. 319–326, 2014.
- [43] X. de Lamballerie, L. Ninove, and R. N. Charrel, "Antiviral treatment of Chikungunya virus infection," *Infectious Disorders— Drug Targets*, vol. 9, no. 2, pp. 101–104, 2009.
- [44] R. Ravichandran and M. Manian, "Ribavirin therapy for Chikungunya arthritis," *Journal of Infection in Developing Countries*, vol. 2, no. 2, pp. 140–142, 2008.
- [45] B. Rada and M. Dragun, "Antiviral action and selectivity of 6 azauridine," Annals of the New York Academy of Sciences, vol. 284, pp. 410–417, 1977.
- [46] S. Briolant, D. Garin, N. Scaramozzino, A. Jouan, and J. M. Crance, "*in vitro* inhibition of Chikungunya and Semliki Forest viruses replication by antiviral compounds: synergistic effect of

interferon-α and ribavirin combination," *Antiviral Research*, vol. 61, no. 2, pp. 111–117, 2004.

- [47] R. A. Lukaszewski and T. J. G. Brooks, "Pegylated alpha interferon is an effective treatment for virulent venezuelan equine encephalitis virus and has profound effects on the host immune response to infection," *Journal of Virology*, vol. 74, no. 11, pp. 5006–5015, 2000.
- [48] I. Delogu, B. Pastorino, C. Baronti, A. Nougairède, E. Bonnet, and X. de Lamballerie, "*in vitro* antiviral activity of arbidol against Chikungunya virus and characteristics of a selected resistant mutant," *Antiviral Research*, vol. 90, no. 3, pp. 99–107, 2011.
- [49] P. Kaur, M. Thiruchelvan, R. C. H. Lee et al., "Inhibition of Chikungunya virus replication by Harringtonine, a novel antiviral that suppresses viral protein expression," *Antimicrobial Agents and Chemotherapy*, vol. 57, no. 1, p. 155, 2013.
- [50] I. Delogu and X. de Lamballerie, "Chikungunya disease and chloroquine treatment," *Journal of Medical Virology*, vol. 83, no. 6, pp. 1058–1059, 2011.
- [51] P. K. Dash, M. Tiwari, S. R. Santhosh, M. Parida, and P. V. Lakshmana Rao, "RNA interference mediated inhibition of Chikungunya virus replication in mammalian cells," *Biochemical and Biophysical Research Communications*, vol. 376, no. 4, pp. 718–722, 2008.
- [52] D. Parashar, M. S. Paingankar, S. Kumar et al., "Administration of E2 and NS1 siRNAs inhibit Chikungunya virus replication *in vitro* and protects mice infected with the virus," *PLoS Neglected Tropical Diseases*, vol. 7, no. 9, article e2405, 2013.
- [53] S. Lam, K. C. Chen, M. M. Ng et al., "Expression of plasmidbased shRNA against the E1 and nsP1 genes effectively silenced Chikungunya virus replication," *PLoS ONE*, vol. 7, no. 10, Article ID e46396, 2012.
- [54] S. Ozden, M. Lucas-Hourani, P.-E. Ceccaldi et al., "Inhibition of Chikungunya virus infection in cultured human muscle cells by furin inhibitors: impairment of the maturation of the E2 surface glycoprotein," *Journal of Biological Chemistry*, vol. 283, no. 32, pp. 21899–21908, 2008.
- [55] E. I. Bondarenko, E. V. Protopopova, I. V. Surovtsev, A. N. Shvalov, and V. B. Loktev, "Inhibition of replication of Venezuelan equine encephalomyelitis with polyclonal antibodies to laminin-binding protein," *Voprosy Virusologii*, vol. 49, no. 5, pp. 32–37, 2004.
- [56] A.-C. Bréhin, I. Casadémont, M.-P. Frenkiel, C. Julier, A. Sakuntabhai, and P. Desprès, "The large form of human 21,51-Oligoadenylate Synthetase (OAS3) exerts antiviral effect against Chikungunya virus," *Virology*, vol. 384, no. 1, pp. 216–222, 2009.
- [57] H. Gad, S. Paulous, E. Belarbi et al., "The E2-E166K substitution restores Chikungunya virus growth in OAS3 expressing cells by acting on viral entry," *Virology*, vol. 434, no. 1, pp. 27–37, 2012.
- [58] J. E. Kitchin, M. K. Pomeranz, G. Pak, K. Washenik, and J. L. Shupack, "Rediscovering mycophenolic acid: a review of its mechanism, side effects, and potential uses," *Journal of the American Academy of Dermatology*, vol. 37, no. 3, pp. 445–449, 1997.
- [59] Z. J. Gong, S. de Meyer, C. Clarysse et al., "Mycophenolic acid, an immunosuppressive agent, inhibits HBV replication *in vitro*," *Journal of Viral Hepatitis*, vol. 6, no. 3, pp. 229–236, 1999.
- [60] J. J. Lipsky, "Mycophenolate mofetil," *The Lancet*, vol. 348, no. 9038, pp. 1357–1359, 1996.
- [61] R. E. Lanford, D. Chavez, B. Guerra et al., "Ribavirin induces error-prone replication of GB virus B in primary tamarin

hepatocytes," *Journal of Virology*, vol. 75, no. 17, pp. 8074–8081, 2001.

- [62] M. Khan, R. Dhanwani, I. K. Patro, P. V. L. Rao, and M. M. Parida, "Cellular IMPDH enzyme activity is a potential target for the inhibition of Chikungunya virus replication and virus induced apoptosis in cultured mammalian cells," *Antiviral Research*, vol. 89, no. 1, pp. 1–8, 2011.
- [63] T. S. Teng, S. S. Foo, D. Simamarta et al., "Viperin restricts Chikungunya virus replication and pathology," *The Journal of Clinical Investigation*, vol. 122, no. 12, pp. 4447–4460, 2012.
- [64] T. Couderc, N. Khandoudi, M. Grandadam et al., "Prophylaxis and therapy for Chikungunya virus infection," *Journal of Infectious Diseases*, vol. 200, no. 4, pp. 516–523, 2009.
- [65] A. Suhrbier, M. C. Jaffar-Bandjee, and P. Gasque, "Arthritogenic alphaviruses—an overview," *Nature Reviews. Rheumatology*, vol. 8, no. 7, pp. 420–429, 2012.
- [66] L. Pohjala, A. Utt, M. Varjak et al., "Inhibitors of alphavirus entry and replication identified with a stable Chikungunya replicon cell line and virus-based assays," *PLoS ONE*, vol. 6, no. 12, Article ID e28923, 2011.
- [67] M. Lucas-Hourani, A. Lupan, P. Desprès et al., "A phenotypic assay to identify Chikungunya virus inhibitors targeting the nonstructural protein nsP2," *Journal of Biomolecular Screening*, vol. 18, no. 2, pp. 172–179, 2013.
- [68] D. J. M. Cruz, R. M. Bonotto, R. G. B. Gomes et al., "Identification of novel compounds inhibiting Chikungunya virusinduced cell death by high throughput screening of a kinase inhibitor library," *PLoS Neglected Tropical Diseases*, vol. 7, no. 10, Article ID e2471, 2013.
- [69] C. Bissantz, G. Folkers, and D. Rognan, "Protein-based virtual screening of chemical databases. 1. Evaluation of different docking/scoring combinations," *Journal of Medicinal Chemistry*, vol. 43, no. 25, pp. 4759–4767, 2000.
- [70] E. Reichert, A. Clase, A. Bacetty, and J. Larsen, "Alphavirus antiviral drug development: scientific gap analysis and prospective research areas," *Biosecurity and Bioterrorism*, vol. 7, no. 4, pp. 413–427, 2009.
- [71] A. A. Rashad and P. P. Keller, "Structure based design towards the identification of novel binding sites and inhibitors for the Chikungunya virus envelope proteins," *Journal of Molecular Graphics & Modelling*, vol. 44, pp. 241–252, 2013.
- [72] J. J. Fros, W. J. Liu, N. A. Prow et al., "Chikungunya virus nonstructural protein 2 inhibits type I/II interferon-stimulated JAK-STAT signaling," *Journal of Virology*, vol. 84, no. 20, pp. 10877–10887, 2010.
- [73] L. K. White, T. Sali, D. Alvarado et al., "Chikungunya virus induces IPS-1-dependent innate immune activation and protein kinase R-independent translational shutoff," *Journal of Virology*, vol. 85, no. 1, pp. 606–620, 2011.
- [74] M. Bassetto, T. de Burghgraeve, L. Delang et al., "Computeraided identification, design and synthesis of a novel series of compounds with selective antiviral activity against Chikungunya virus," *Antiviral Research*, vol. 98, no. 1, pp. 12–18, 2013.
- [75] D. Jochmans, N. Segura Guerrero, and L. Delang, "Inhibition of Chikungunya virus replication by T-705 (favipiravir) and identification of resistance associated mutations in the RNA-dep endent RNA polymerase," http://rega.kuleuven.be/cmt/jn/poster/2013/2013_dj/2013CHIKVT705_Poster_KeystoneMeeting_ V8.pdf.
- [76] V. R. Harrison, L. N. Binn, and R. Randall, "Comparative immunogenicities of Chikungunya vaccines prepared in avian

and mammalian tissues," *The American Journal of Tropical Medicine and Hygiene*, vol. 16, no. 6, pp. 786–791, 1967.

- [77] A. White, S. Berman, and J. P. Lowenthal, "Comparative immunogenicities of Chikungunya vaccines propagated in monkey kidney monolayers and chick embryo suspension cultures," *Applied Microbiology*, vol. 23, no. 5, pp. 951–952, 1972.
- [78] K. H. Eckels, V. R. Harrison, and F. M. Hetrick, "Chikungunya virus vaccine prepared by Tween-ether extraction," *Applied Microbiology*, vol. 19, no. 2, pp. 321–325, 1970.
- [79] E. Nakao and S. Hotta, "Immunogenicity of purified, inactivated Chikungunya virus in monkeys," *Bulletin of the World Health Organization*, vol. 48, no. 5, pp. 559–562, 1973.
- [80] R. Edelman, C. O. Tacket, S. S. Wasserman, S. A. Bodison, J. G. Perry, and J. A. Mangiafico, "Phase II safety and immunogenicity study of live Chikungunya virus vaccine TSI-GSD-218," *American Journal of Tropical Medicine and Hygiene*, vol. 62, no. 6, pp. 681–685, 2000.
- [81] V. R. Harrison, K. H. Eckels, P. J. Bartelloni, and C. Hampton, "Production and evaluation of a formalin-killed Chikungunya vaccine," *Journal of Immunology*, vol. 107, no. 3, pp. 643–647, 1971.
- [82] M. Tiwari, M. Parida, S. R. Santhosh, M. Khan, P. K. Dash, and P. V. L. Rao, "Assessment of immunogenic potential of Vero adapted formalin inactivated vaccine derived from novel ECSA genotype of Chikungunya virus," *Vaccine*, vol. 27, no. 18, pp. 2513–2522, 2009.
- [83] J. Gardner, I. Anraku, T. T. Le et al., "Chikungunya virus arthritis in adult wild-type mice," *Journal of Virology*, vol. 84, no. 16, pp. 8021–8032, 2010.
- [84] N. H. Levitt, H. H. Ramsburg, and S. E. Hasty, "Development of an attenuated strain of Chikungunya virus for use in vaccine production," *Vaccine*, vol. 4, no. 3, pp. 157–162, 1986.
- [85] C. D. Partidos, J. Paykel, J. Weger et al., "Cross-protective immunity against o'nyong-nyong virus afforded by a novel recombinant Chikungunya vaccine," *Vaccine*, vol. 30, no. 31, pp. 4638–4643, 2012.
- [86] K. Plante, E. Wang, C. D. Partidos et al., "Novel Chikungunya vaccine candidate with an ires-based attenuation and host range alteration mechanism," *PLoS Pathogens*, vol. 7, no. 7, Article ID e1002142, 2011.
- [87] E. Wang, E. Volkova, A. P. Adams et al., "Chimeric alphavirus vaccine candidates for Chikungunya," *Vaccine*, vol. 26, no. 39, pp. 5030–5039, 2008.
- [88] K. Muthumani, K. M. Lankaraman, D. J. Laddy et al., "Immunogenicity of novel consensus-based DNA vaccines against Chikungunya virus," *Vaccine*, vol. 26, no. 40, pp. 5128–5134, 2008.
- [89] K. Mallilankaraman, D. J. Shedlock, H. Bao et al., "A DNA vaccine against Chikungunya virus is protective in mice and induces neutralizing antibodies in mice and nonhuman primates," *PLoS Neglected Tropical Diseases*, vol. 5, no. 1, article e928, 2011.
- [90] T. G. Szabó, R. Palotai, P. Antal et al., "Critical role of glycosylation in determining the length and structure of T cell epitopes," *Immunome Research*, vol. 5, no. 1, article 4, 2009.
- [91] D. Wang, A. Suhrbier, A. Penn-Nicholson et al., "A complex adenovirus vaccine against Chikungunya virus provides complete protection against viraemia and arthritis," *Vaccine*, vol. 29, no. 15, pp. 2803–2809, 2011.
- [92] M. Kumar, A. B. Sudeep, and V. A. Arankalle, "Evaluation of recombinant E2 protein-based and whole-virus inactivated

candidate vaccines against Chikungunya virus," *Vaccine*, vol. 30, no. 43, pp. 6142–6149, 2012.

- [93] S. W. Metz and G. P. Pijlman, "Arbovirus vaccines; opportunities for the baculovirus-insect cell expression system," *Journal of Invertebrate Pathology*, vol. 107, pp. s16–s30, 2011.
- [94] S. W. Metz, C. Geertsema, B. E. Martina et al., "Functional processing and secretion of Chikungunya virus E1 and E2 glycoproteins in insect cells," *Virology Journal*, vol. 8, article 353, 2011.
- [95] W. Akahata, Z.-Y. Yang, H. Andersen et al., "A virus-like particle vaccine for epidemic Chikungunya virus protects nonhuman primates against infection," *Nature Medicine*, vol. 16, no. 3, pp. 334–338, 2010.
- [96] R. M. Kramer, Y. Zeng, N. Sahni et al., "Development of a stable virus-like particle vaccine formulation against Chikungunya virus and investigation of the effects of polyanions," *Journal of Pharmaceutical Sciences*, vol. 102, no. 12, pp. 4305–4314, 2013.
- [97] M. Khan, R. Dhanwani, P. V. Rao et al., "Subunit vaccine formulations based on recombinant envelope proteins of Chikungunya virus elicit balanced Th1/Th2 response and virusneutralizing antibodies in mice," *Virus Research*, vol. 167, no. 2, pp. 236–246, 2012.
- [98] H. Chu, S. C. Das, J. F. Fuchs et al., "Deciphering the protective role of adaptive immunity to CHIKV/IRES a novel candidate vaccine against Chikungunya in the A129 mouse model," *Vaccine*, vol. 31, no. 33, pp. 3353–3360, 2013.
- [99] D. Hallengärd, M. Kakoulidou, A. Lulla et al., "Novel attenuated Chikungunya vaccine candidates elicit protective immunity in C57BL/6 mice," *Journal of Virology*, vol. 88, no. 5, pp. 2858–2866, 2013.
- [100] C. J. Roy, A. P. Adams, E. Wang et al., "Chikungunya vaccine candidate is highly attenuated and protects nonhuman primates against telemetrically-monitored disease following a single dose," *Journal of Infectious Diseases*. In press.
- [101] J. García-Arriaza, V. Cepeda, D. Hallengärd et al., "A novel poxvirus-based vaccine (MVA-CHIKV) is highly immunogenic and protects mice against Chikungunya infection," *Journal of Virology*, vol. 88, no. 6, pp. 3527–3547, 2014.
- [102] J. Bettadapura, L. J. Herrero, A. Taylor et al., "Approaches to the treatment of disease induced by Chikungunya virus," *Indian Journal of Medical Research*, vol. 138, pp. 762–765, 2013.