



New Viewpoint in Exaggerated Increase of PtiO₂ With Normobaric Hyperoxygenation and Reasons to Limit Oxygen Use in Neurotrauma Patients

Gurgen Harutyunyan¹*, Garnik Harutyunyan² and Gagik Mkhoyan³

¹ Emergency, Hospital 9 de Octubre, Valencia, Spain, ² Faculty of Pharmacy, Universitat De València, Valencia, Spain, ³ Anesthesiology and Intensive Care, Erebouni Medical Center, Yerevan, Armenia

Keywords: normobaric hyperoxia, PtiO₂, metabolism, brain trauma, blood-buffering capacity

OPEN ACCESS

Edited by:

Samir G. Sakka, Universität Witten/Herdecke, Germany

Reviewed by:

Suren Soghomonyan, The Ohio State University, United States

*Correspondence:

Gurgen Harutyunyan varsenik@hotmail.es

Specialty section:

This article was submitted to Intensive Care Medicine and Anesthesiology, a section of the journal Frontiers in Medicine

Received: 13 September 2017 Accepted: 10 April 2018 Published: 22 May 2018

Citation:

Harutyunyan G, Harutyunyan G and Mkhoyan G (2018) New Viewpoint in Exaggerated Increase of PtiO₂ With Normobaric Hyperoxygenation and Reasons to Limit Oxygen Use in Neurotrauma Patients. Front. Med. 5:119. doi: 10.3389/fmed.2018.00119 Multiple studies have shown that some of cerebral metabolic abnormalities occurring after traumatic brain injuries (TBI) are not of ischemic origin (1-4). The described pattern is called "metabolic crisis without brain ischemia" (5), and is associated with poor outcome within 6 months after trauma (6-8). Nevertheless, targeting adequate cerebral blood flow (CBF) and oxygen delivery (DO_2) remains the cornerstone in the clinical management of TBI, especially in the early phase. Surprisingly, DO₂ targeted therapy based on an escalation of dissolved O_{2} , has been shown positive outcome in a few cases only, although it causes a significant increase in O₂ pressure in cerebral tissue (PtiO₂). High inspired O_2 fraction or normobaric hyperoxia (NH) leads to a dramatic elevation of $PtiO_2$ to "arterial" levels of $147 \pm 36 \text{ mmHg}(9)$. This daily and well-known phenomenon so far does not have definitive explanation and differs from the classical notions: oxyhemoglobin saturation (SO₂) must remain nearly 100% over a wide range of O_2 partial pressures (PO₂) greater than 80 mmHg (10). NH induces a negligible increase in the amount of arterial O₂ content and DO₂, whereas it is associated with an important augmentation of PtiO₂, without significant change in cerebral metabolic rate of O₂ consumption (CMRO₂) (11-14). Furthermore, studies using positron emission tomography, magnetic resonance imaging (MRI), and near-infrared spectroscopy showed an active O₂ extraction fraction (OEF) in the non-necrotic tissue during NH (9, 15-18) with negligible increase of regional SO_2 (rSO₂) at 2.8 ± 1.82% (9) or without changes in rSO₂ in tissue with intact autoregulation (16). The data of OEF at 0.56 \pm 0.06 in reversible tissue, which although reduced from viable tissue to infarction (19), discards the possibility of a luxury perfusión in these cases. In general, NH causes an insignificant increase or no in rSO₂ with an exaggerated elevation of PtiO₂, which is equal to or less than end capillary PO_2 (20). Thus, with NH the PO_2 of end capillary venous blood reaches to "arterial" levels. Therefore, the exaggerated increase of $PtiO_2$ with a negligible increase of rSO_2 is incompatible with the classical sigmoidal form of the oxyhemoglobin dissociation curve (ODC). Hereby, the circulated hypotheses of mitochondrial dysfunction as a contributor (21) to high PtiO₂ and the loss of O_2 homeostatic mechanisms in the injured tissue during NH (22) requires an alternative explanation based on the conformational change of hemoglobin (Hb) quaternary structure (Max Perutz—the Nobel prize in chemistry 1962). This change produces a significant decrease in $Hb-O_2$ affinity, considerable Hb buffering capacity augmentation, and convert the sigmoidal form of ODC to hyperbola. In this commentary, we explain the mechanisms underlying the exaggerated rise in PtiO₂ with NH and consider the factors that limiting oxygen use in the damaged cerebral tissue.

BASIC BIOCHEMISTRY

Hemoglobin plays an important role in the transport of CO_2 by the blood. In the brain tissue with a respiratory quotient (RQ) of 1, metabolism of 1 mol of O_2 results in 1 mol of CO_2 . Also, 1 mol of

 CO_2 in the erythrocyte generates 1 mol of protons (H^+) by two mechanisms:

$$CO_2 + H_2O \leftrightarrow HCO_3^- + H^+ *$$
$$CO_2 + Hb-NH_2 \leftrightarrow Hb-NH-COO^- + H^+$$

(*This reaction is catalyzed by carbonic anhydrase ranging between 10^4 and 10^6 reactions per second (23); therefore, the participation of other buffers of blood in CO₂ buffering is not discussed).

The H⁺ uptake by Hb facilitates CO_2 transport by stimulating bicarbonate formation: erythrocyte membranes do have a rapid anion exchange protein (capnoporin), which exchanges HCO_3^- for Cl⁻ at a ratio of 1:1. The H⁺ released by carbamate formation is taken up by the Hb as well. Thus, Hb remains the main buffer for protons (i.e., for CO_2), but its capacity is biochemically limited (24):

$$Hb(O_{2})_{k} - O_{2} \leftrightarrow Hb(O_{2})_{k-1} + 0.6H^{+}$$
(1)

where k = 1, 2, 3 or 4; 0.6 is the Haldane coefficient (the amount of the oxygen-linked H⁺ binding of Hb).

Thus, the release of 1 mol of O_2 will allow the Hb to bind a maximum 0.6 mol of H⁺ or the Hb buffering capacity is lost in the brain tissue when OEF exceeded 60% (i.e., full saturation of Hb with H⁺) and CO₂ begins to accumulate intensively in the brain tissue, provoking important acidosis. The imbalance of Hb capacities as a H⁺ buffer and as an O₂ transporter in this setting (i.e., Eq. 1) makes the buffering capacity of Hb more important for brain cell survival to maintain a stable pH than its O₂ carrier function. With the constant CMRO₂, unchanged capillary permeability, temperature, pH, and DO₂, the PtiO₂ generated depends only on the Hb–O₂ affinity, which decreases when Hb is saturated with negative allosteric effectors [2,3-diphosphoglycerate (2,3 DPG), CO₂, H⁺, Cl⁻] (24).

The Hill coefficient (*n*) and the ODCs provide a way to assess these effects:

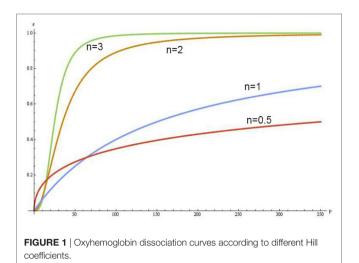
$$SO_2 = \frac{(PO2)^n}{(P^{50}O2)^n + (PO2)^n}$$
 (2)

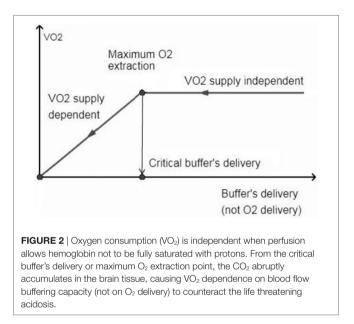
where P⁵⁰O₂ is O₂ half-saturation pressure of Hb.

The increase of concentrations of the negative allosteric effectors in erythrocytes shifts the sigmoidal shape of ODC to the right and when Hb saturates with them, the ODC becomes hyperbolic in form (**Figure 1**). In this condition, Hb converts from the quaternary conformational state R (relax) to a state T (tense), which has a lower Hb–O₂ affinity, hyperbolic form of ODC and increases up to six times the buffer capacity of Hb (24).

PHYSIOLOGY AND PATHOPHYSIOLOGY

In brain tissue (RQ at 1.0), use of 1 mol of O_2 will result in the production of more CO_2 than in the other tissues (RQ at 0.8). That is, the Hb buffering capacity is depleted with higher SO_2 in the brain tissue (i.e., $SO_2 \approx 40\%$, Eq. 1) than in other tissues. From this critical point, the generated CO_2 is transported only





in dissolved states and accumulated in the brain tissue causing clinically important acidosis with "normal" values of $PtiO_2$. High levels of CO_2 and intracellular acidosis directly or indirectly suppress the mitochondrial respiration: the CMRO₂ is inhibited proportionally to decrease the perfusion by compensatory mechanisms [i.e., OEF is unable to raise (25)] trying to counteract the life-threatening intracellular acidosis. This is a blood-buffering capacity (BBC)-dependent mitochondrial suppression while independent of tissue O_2 availability for cell aerobic metabolism. The O_2 consumption (VO₂)/buffer's delivery inter-independency is shown in **Figure 2**.

Therefore, the accepted term " O_2 delivery" does not reflect the pathophysiology of metabolism and has to be replaced by "buffer's delivery," which depends on the amount of Hb, the CBF, the total content of CO_2 in arterial blood, and the Hb quaternary conformational state at the end of capillaries. The exaggerated increase in PtiO₂ by NH in the VO₂ supply-dependent phase has no metabolic effect due to the depletion of BBC. Moreover, augmentation in BBC in this phase through the increases of Hb concentration, augmentation of CBF or controlled hyperventilation without a change of Hb conformational state at the end of capillaries, is able to unlock the mitochondrial suppression and raise the CMRO₂. Importantly, the high concentrations of 2,3 DPG as a significant negative allosteric effector can reduce the Hill coefficient to one without depleting the buffering capacity of Hb and can convert the ODC to hyperbola. In this case, the NH also causes an exaggerated increase in PtiO₂ in relatively healthy tissue by physiological reduction on rCBF through vasoconstriction.

CLINICAL INTERPRETATION

Many drugs and techniques used commonly during therapy of severe TBI, including mannitol, thiopental, ketorolac, nimodipine, hypothermia, deep sedation, transfusion of red blood cells (RBC), etc., can reduce the $PtiO_2$ in the damaged tissue (26–30). This is because it improves the relationship within BBC and CMRO₂ causing a decrease of Hb loading by negative effectors (i.e., increase in Hb–O₂ affinity). The effects of medically induced augmentation of cerebral perfusion pressure on cerebral oxygenation are difficult to predict (31, 32). Indeed, the effects depend on which phase (VO₂ buffer supply dependent or independent) the rCBF (i.e., BBC) change is made. Coles et al. showed that with hyperventilation, many regions of brain demonstrate an increase in CMRO₂ despite the reduction in CBF (33). In fact, the reduction of Hb loading by negative allosteric effectors in the VO₂ buffer supply-dependent phase, the hyperventilation rehabilitates the suppressed mitochondrial function by augmentation of BBC. Often uncontrolled hyperventilation causes an important decrease in PtiO₂ due to an increase in Hb-O₂ affinity. The worst is, when hyperventilation causes end capillary T to R conformational change of Hb with a decrease in BBC, which is dangerous for damaged tissue. Unlike hyperventilation, hypoventilation improves tissue oxygenation (34). Despite this effect, hypoventilation has no clinical use in patients with TBI. The increase in PtiO₂ is related not so much to the increase of CBF by vasodilatation as to the increase in Hb loading by the negative allosteric effectors and the decrease in Hb-O₂ affinity. Depletion of BBC will cause the decrease of CMRO₂. Several studies have shown an association between anemia and poor outcome after TBI (35-37). As expected, the higher Hb levels are associated with improved outcome after subarachnoid hemorrhage (38) and post-cardiac arrest patients (39-41) (i.e., higher BBC maintains the metabolism in the VO₂ supply independent phase). On the other hand, the transfusion of RBC is an independent risk factor for poor neurological outcome after 3 and 6 months in TBI patients (42). The stored blood with low values of erythrocyte 2,3-DPG can be used without hesitation when correcting a chronic anemia for instance (43). However, in acute situations with important tissue acidosis, the organism needs to rapidly augment the buffering capacity of Hb by R to T states shift, but re-synthesis of 2,3-DPG is obviously insufficient in stored blood. In such situations, fresh blood or blood with a near normal 2,3-DPG content should be used (43). According to proposed hypothesis, the augmentation of BBC by increasing the Hb concentration is the main role of RBC transfusion in TBI patients. Inconsistent results and contradictory findings have been reported for the effects of NH in brain energy metabolism (8, 10, 19, 44–46). The lactate MR spectroscopy demonstrates that in ischemic core and contralateral striatum, the lactate levels are not affected by NH, whereas, in the region of mismatch, lactate levels changed with changes in O₂ delivery (47). The T2*-weighted MRI studies showed positive CBF changes to O₂ challenge in the mismatch region, in contrast to negative CBF changes in normal tissue due to O₂-induced vasoconstriction (16, 48). Indeed, the increase in BBC through augmentation of rCBF in the VO₂ buffer supply dependent phase (i.e., in mismatch region) improves local acidosis, unlock the mitochondrial suppression, and decrease the lactate levels.

DISCUSSION

Martini and colleagues determined that severe TBI management guided by PtiO₂ monitoring was associated with a poor neurological outcome and was an inefficient use of hospital resources (49). Of course, recording of "normal" or high PtiO₂ values with acceptable high inspired O₂ fraction or positive end-expiratory pressures without sufficient BBC may create a false impression of safety and negatively impact clinical decision making. The use of absolute levels of PtiO₂ as a marker of ischemia is confusing because a more important BBC deficit happens much earlier. The transition from the concept of "increase in O₂ supply" to the concept of "increase of buffer supply" has important clinical implications in the strategy of treatment in neurotrauma patient. In addition, the characteristics of T Hb (i.e., hyperbolic ODC, high buffering capacity) allow us to understand and correctly assess the effects of current treatment and evolution of damaged brain tissue through PtiO2 changes with NH. The explanation of known paradoxes in neurotrauma such as an exaggerated rise in PtiO₂ by NH, inexplicable decrease in PtiO₂ after many drugs and techniques commonly used, and metabolic crisis without "ischemia" is explained for the first time by the Hb conformational changes and BBC-dependent mitochondrial suppression. As we have seen the metabolic crisis always coincides with ischemia but in the sense of lack of BBC. The hypothesis is structural and revolutionary. Despite the enormous amount of data available in literature, targeted practical observations are needed to confirm (or deny) this hypothesis.

CONCLUSION

- 1. The exaggerated increase of $PtiO_2$ by NH occurs in the "T" state of Hb when ODC as a hyperbole and does not always coincide with the mitochondrial dysfunction.
- 2. The BBC is a limiting factor of O_2 consumption by damaged brain tissues.

AUTHOR CONTRIBUTIONS

GH, as the first author, he developed the hypothesis and prepared the manuscript. All the co-authors helped with their important input to finalize the hypothesis discussed and to write the manuscript.

REFERENCES

- Chieregato A, Tanfani A, Compagnone C, Turrini C, Sarpieri F, Ravaldini M, et al. Global cerebral blood flow and CPP after severe head injury: a xenon-CT study. *Intensive Care Med* (2007) 33(5):856–62. doi:10.1007/s00134-007-0604-4
- Soustiel JF, Sviri GE. Monitoring of cerebral metabolism: non-ischemic impairment of oxidative metabolism following severe traumatic brain injury. *Neurol Res* (2007) 29(7):654–60. doi:10.1179/016164107X240017
- Glenn TC, Kelly DF, Boscardin WJ, McArthur DL, Vespa P, Oertel M, et al. Energy dysfunction as a predictor of outcome after moderate or severe head injury: indices of oxygen, glucose, and lactate metabolism. *J Cereb Blood Flow Metab* (2003) 23(10):1239–50. doi:10.1097/01.WCB.0000089833.23606.7F
- Soustiel JF, Glenn TC, Vespa P, Rinsky B, Hanuscin C, Martin NA. Assessment of cerebral blood flow by means of blood-flow-volume measurement in the internal carotid artery: comparative study with a 133xenon clearance technique. *Stroke* (2003) 34(8):1876–80. doi:10.1161/01.STR.0000080942.32331.39
- Vespa P, Bergneider M, Hattori N, Wu HM, Huang SC, Martin NA, et al. Metabolic crisis without brain ischemia is common after traumatic brain injury: a combined microdialysis and positron emission tomography study. *J Cereb Blood Flow Metab* (2005) 25:763–74. doi:10.1038/sj.jcbfm.9600073
- Stein NR, McArthur DL, Etchepare M, Vespa PM. Early cerebral metabolic crisis after TBI influences outcome despite adequate hemodynamic resuscitation. *Neurocrit Care* (2012) 17:49–57. doi:10.1007/s12028-012-9708-y
- Marcoux J, McArthur DA, Miller C, Glenn TC, Villablanca P, Martin NA, et al. Persistent metabolic crisis as measured by elevated cerebral microdialysis lactate-pyruvate ratio predicts chronic frontal lobe brain atrophy after traumatic brain injury. *Crit Care Med* (2008) 36:2871–7. doi:10.1097/CCM. 0b013e318186a4a0
- Xu Y, Mc Arthur DL, Alger JR, Etchepare M, Hovda DA, Glenn TC, et al. Early non ischemic oxidative metabolic dysfunction leads to chronic brain atrophy in traumatic brain injury. *J Cereb Blood Flow Metab* (2010) 30:883–94. doi:10.1038/jcbfm.2009.263
- McLeod AD, Igielman F, Elwell C, Cope M, Smith M. Measuring cerebral oxygenation during normobaric hyperoxia: a comparison of tissue microprobes, near-infrared spectroscopy, and jugular venous oximetry in head injury. *Anesth Analg* (2003) 97(3):851–6. doi:10.1213/01.ANE.0000072541. 57132.BA
- Severinghaus JW. Simple, accurate equations for human blood O₂ dissociation computations. *J Appl Physiol Respir Environ Exerc Physiol* (1979) 46: 599–602. doi:10.1152/jappl.1979.46.3.599
- Magnoni S, Ghisoni L, Locatelli M, Caimi M, Colombo A, Valeriani V, et al. Lack of improvement in cerebral metabolism after hyperoxia in severe head injury: a micro-dialysis study. *J Neurosurg* (2003) 98:952–8. doi:10.3171/jns.2003. 98.5.0952
- Diringer MN. Hyperoxia: good or bad for the injured brain? Curr Opin Crit Care (2008) 14:167–71. doi:10.1097/MCC.0b013e3282f57552
- Diringer MN, Aiyagari V, Zazulia AR, Videen TO, Powers WJ. Effect of hyperoxia on cerebral metabolic rate for oxygen measured using positron emission tomography in patients with acute severe head injury. *J Neurosurg* (2007) 106:526–9. doi:10.3171/jns.2007.106.4.526
- Xu F, Liu P, Pascual JM, Xiao G, Lu H. Effect of hypoxia and hyperoxia on cerebral blood flow, blood oxygenation and oxidative metabolism. *J Cereb Blood Flow Metab* (2012) 32:1909–18. doi:10.1038/jcbfm.2012.93
- Ghosh A, Highton D, Kolyva C, Tachtsidis I, Elwell CE, Smith M. Hyperoxia results in increased aerobic metabolism following acute brain injury. J Cereb Blood Flow Metab (2017) 37:2910–20. doi:10.1177/0271678X16679171
- 16. Sahoo S, Sheshadri V, Sriganesh K, Madhsudana Reddy KR, Radhakrishnan M, Umamaheswara Rao GS. Effect of hyperoxia on cerebral blood flow velocity and regional oxygen saturation in patients operated on for severe traumatic brain injury-the influence of cerebral blood flow autoregulation. World Neurosurg (2017) 98:211–6. doi:10.1016/j.wneu.2016.10.116
- Shen Q, Huang S, Du F, Duong TQ. Probing ischemic tissue fate with BOLD fMRI of brief oxygen challenge. *Brain Res* (2011) 1425:132–41. doi:10.1016/j. brainres.2011.09.052
- Christen T, Bouzat P, Pannetier N, Coquery N, Moisan A, Lemasson B, et al. Tissue oxygen saturation mapping with magnetic resonance imaging. *J Cereb Blood Flow Metab* (2014) 34(9):1550–7. doi:10.1038/jcbfm.2014.116

- Powers WJ, Grubb RL Jr, Darriet D, Raichle ME. Cerebral blood flow and cerebral metabolic rate of oxygen requirements for cerebral function and viability in humans. J Cereb Blood Flow Metab (1985) 5:600–8. doi:10.1038/jcbfm.1985.89
- Gupta AK, Hutchinson PJ, Fryer T, Al-Rawi PG, Parry DA, Minhas PS, et al. Measurement of brain tissue oxygenation performed using positron emission tomography scanning to validate a novel monitoring method. *J Neurosurg* (2002) 96(2):263–8. doi:10.3171/jns.2002.96.2.0263
- Johnston AJ, Steiner LA, Gupta AK, Menon DK. Cerebral oxygen vasoreactivity and cerebral tissue oxygen reactivity. *Br J Anaesth* (2003) 90(6):774–86. doi:10.1093/bja/aeg104
- Vilalta A, Sahuquillo J, Merino MA, Poca MA, Garnacho A, Martinez-Valverde T, et al. Normobaric hyperoxia in traumatic brain injury. Does brain metabolic state influence the response to hyperoxic challenge? *J Neurotrauma* (2011) 28:1139–48. doi:10.1089/neu.2010.1720
- Lindskog S. Structure and mechanism of carbonic anhydrase. *Pharmacol Ther* (1997) 74(1):1–20. doi:10.1016/S0163-7258(96)00198-2
- 24. Voet D, Voet JG. *Biochemistry*. 4th ed. United States of America: John Wiley & Sons (2010).
- Gutierrez G, Pohil R. Oxygen consumption is linearly related to O₂ supply in critically ill patients. J Crit Care (1986) 1(1):45–53. doi:10.1016/ S0883-9441(86)80116-2
- Steiner T, Pilz J, Schellinger P, Wirtz R, Friederichs V, Aschoff A, et al. Multimodal online monitoring in middle cerebral artery territory stroke. *Stroke* (2001) 32:2500–6. doi:10.1161/hs1101.097400
- Gupta AK, Al-Rawi PG, Hutchinson PJ, Kirkpatrick PJ. Effect of hypothermia on brain tissue oxygenation in patients with severe head injury. *Br J Anesth* (2002) 88:188–92. doi:10.1093/bja/88.2.188
- Stiefel MF, Heuer GG, Abrahams JM, Bloom S, Smith MJ, Maloney-Wilensky E, et al. The effect of nimodipine on cerebral oxygenation in patients with poorgrade subarachnoid hemorrhage. *J Neurosurg* (2004) 101:594–9. doi:10.3171/ jns.2004.101.4.0594
- Zygun DA, Nortje J, Hutchinson PJ, Timofeev I, Menon DK, Gupta AK. The effect of red blood cell transfusion on cerebral oxygenation and metabolism after severe traumatic brain injury. *Crit Care Med* (2009) 37:1074–8. doi:10.1097/CCM.0b013e318194ad22
- Sakowitz OW, Stover JF, Sarrafzadeh AS, Unterberg AW, Kiening KL. Effects of mannitol bolus administration on intracranial pressure, cerebral extracellular metabolites, and tissue oxygenation in severely head-injured patients. *J Trauma* (2007) 62:292–8. doi:10.1097/01.ta.0000203560.03937.2d
- LeRoux PD, Oddo M. Parenchymal brain oxygen monitoring in the neurocritical care unit. *Neurosurg Clin N Am* (2013) 24:427–39. doi:10.1016/j. nec.2013.03.001
- 32. Sahuquillo J, Amoros S, Santos A, Poca MA, Panzardo H, Domínguez L, et al. Does an increase in cerebral perfusion pressure always mean a better oxygenated brain? A study in head-injured patients. *Brain Edema XI. Acta Neurochirurgica Supplements.* (Vol. 76), Springer, Vienna (2000).
- Coles JP, Fryer TD, Coleman MR, Smielewski P, Gupta AK, Minhas PS, et al. Hyperventilation following head injury: effect on ischemic burden and cerebral oxidative metabolism. *Crit Care Med* (2007) 35(2):568–78. doi:10.1097/01.CCM.0000254066.37187.88
- Akça O, Doufas AG, Morioka N, Iscoe S, Fisher J, Sessler DI. Hypercapnia improves tissue oxygenation. *Anesthesiology* (2002) 97(4):801–6. doi:10.1097/ 00000542-200210000-00009
- Salim A, Hadjizacharia P, DuBose J, Brown C, Inaba K, Chan L, et al. Role of anemia in traumatic brain injury. *J Am Coll Surg* (2008) 207(3):398–406. doi:10.1016/j.jamcollsurg.2008.03.013
- Sekhon MS, McLean N, Henderson WR, Chittock DR, Griesdale DE. Association of hemoglobin concentration and mortality in critically ill patients with severe traumatic brain injury. *Crit Care* (2012) 16(4):R128. doi:10.1186/ cc1143
- 37. Steyerberg EW, Mushkudiani N, Perel P, Butcher I, Lu J, McHugh GS, et al. Predicting outcome after traumatic brain injury: development and international validation of prognostic scores based on admission characteristics. *PLoS Med* (2008) 5(8):165. doi:10.1371/journal.pmed.0050165
- Naidech AM, Jovanovic B, Wartenberg KE, Parra A, Ostapkovich N, Connolly ES, et al. Higher hemoglobin is associated with improved outcome after subarachnoid hemorrhage. *Crit Care Med* (2007) 35(10):2383–9. doi:10.1097/01. CCM.0000284516.17580.2C

- SOS-KANTO Study Group. Relationship between the hemoglobin level at hospital arrival and post–cardiac arrest neurologic outcome. *Am J Emerg Med* (2012) 30:770–4. doi:10.1016/j.ajem.2011.03.031
- Wang CH, Huang CH, Chang WT, Tsai MS, Yu PH, Wang AY, et al. Association between hemoglobin levels and clinical outcomes in adult patients after in-hospital cardiac arrest: a retrospective cohort study. *Intern Emerg Med* (2016) 11:727–36. doi:10.1007/s11739-015-1386-2
- Johnson NJ, Rosselot B, Perman SM, Dodampahala K, Goyal M, Gaieski DF, et al. The association between hemoglobin concentration and neurologic outcome after cardiac arrest. *J Crit Care* (2016) 36:218–22. doi:10.1016/j. jcrc.2016.07.012
- Warner MA, O'Keeffe T, Bhavsar P, Shringer R, Moore C, Harper C, et al. Transfusions and long-term functional outcomes in traumatic brain injury. *J Neurosurg* (2010) 113(3):539–46. doi:10.3171/2009.12.JNS091337
- Stan A, Zsigmond E. The restoration in vivo of 2,3-diphosphoglycerate (2,3-DPG) in stored red cells, after transfusion. The levels of red cells 2,3-DPG. *Rom J Intern Med* (2009) 47(2):173–7.
- 44. Vidal-Jorge M, Sánchez-Guerrero A, Mur-Bonet G, Castro L, Rădoi A, Riveiro M, et al. Does normobaric hyperoxia cause oxidative stress in the injured brain? A microdialysis study using 8-iso-prostaglandin F2α as a biomarker. *J Neurotrauma* (2017) 34:2731–42. doi:10.1089/neu.2017.4992
- 45. Reinert M, Barth A, Rothen HU, Schaller B, Takala J, Seiler RW. Effects of cerebral perfusion pressure and increased fraction of inspired oxygen on brain tissue oxygen, lactate and glucose in patients with severe head injury. Acta Neurochir (Wien) (2003) 145:341–9. doi:10.1007/s00701-003-0027-0
- 46. Tolias CM, Reinert M, Seiler R, Gilman C, Scharf A, Bullock MR. Normobaric hyperoxia-induced improvement in cerebral metabolism and reduction

in intracranial pressure in patients with severe head injury: a prospective historical cohort-matched study. *J Neurosurg* (2004) 101:435–44. doi:10.3171/jns.2004.101.3.0435

- Holmes WM, Lopez-Gonzalez MR, Gallagher L, Deuchar GA, Macrae IM, Santosh C, et al. Detection of the ischemic penumbra: direct assessment of metabolic integrity. NMR Biomed (2012) 25:295–304. doi:10.1002/ nbm.1748
- Wu O, Lu J, Mandeville JB, Murata Y, Egi Y, Dai G, et al. Dynamic functional cerebral blood volume responses to normobaric hypoxia in acute ischemic stroke. J Cereb Blood Flow Metab (2012) 32:1800–9. doi:10.1038/ jcbfm.2012.87
- Martini RP, Deem S, Yanez ND, Chesnut RM, Weiss NS, Daniel S, et al. Management guided by brain tissue oxygen monitoring and outcome following severe traumatic brain injury. *J Neurosurg* (2009) 111(4):644–9. doi:10.3171/2009.2.JNS08998

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2018 Harutyunyan, Harutyunyan and Mkhoyan. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.