

Since January 2020 Elsevier has created a COVID-19 resource centre with free information in English and Mandarin on the novel coronavirus COVID-19. The COVID-19 resource centre is hosted on Elsevier Connect, the company's public news and information website.

Elsevier hereby grants permission to make all its COVID-19-related research that is available on the COVID-19 resource centre - including this research content - immediately available in PubMed Central and other publicly funded repositories, such as the WHO COVID database with rights for unrestricted research re-use and analyses in any form or by any means with acknowledgement of the original source. These permissions are granted for free by Elsevier for as long as the COVID-19 resource centre remains active.



Available online at www.sciencedirect.com







www.elsevier.com/locate/vaccine

# The development of vaccines against SARS corona virus in mice and SCID-PBL/hu mice

Masaji Okada<sup>a,\*</sup>, Yuji Takemoto<sup>a</sup>, Yoshinobu Okuno<sup>b</sup>, Satomi Hashimoto<sup>a</sup>, Shigeto Yoshida<sup>c</sup>, Yukari Fukunaga<sup>a</sup>, Takao Tanaka<sup>a</sup>, Yoko Kita<sup>a</sup>, Sachiko Kuwayama<sup>a</sup>, Yumiko Muraki<sup>a</sup>, Noriko Kanamaru<sup>a</sup>, Hiroko Takai<sup>a</sup>, Chika Okada<sup>a</sup>, Yayoi Sakaguchi<sup>a</sup>, Izumi Furukawa<sup>a</sup>, Kyoko Yamada<sup>a</sup> , Makoto Matsumoto<sup>d</sup>, Tetsuo Kase<sup>b</sup>, Daphne E. deMello<sup>e</sup>, J.S.M. Peiris<sup>f</sup>, Pei-Jer Chen<sup>g</sup>, Naoki Yamamoto<sup>h</sup>, Yoshiyuki Yoshinaka<sup>h</sup>, Tatsuji Nomura<sup>i</sup>, Isao Ishida<sup>j</sup>, Shigeru Morikawa<sup>k</sup>, Masato Tashiro<sup>k</sup>, Mitsunori Sakatani<sup>a</sup>

<sup>a</sup> Clinical Research Center, National Hospital Organization Kinki-Chuo Chest Medical Center, 1180 Nagasone, Sakai, Osaka 591-8555, Japan <sup>b</sup> Department of Infectious Diseases, Osaka Prefectural Institute of Public Health, 3-69 Nakamichi 1-chome Higashinari-ku, Osaka 537-0025, Japan

<sup>c</sup> Department of Infection and Immunity, Jichi Medical School, 3311-1 Yakushiji, Minamikawachi-machi, Tochigi 329-0498, Japan

<sup>d</sup> Microbiological Research Institute, Otsuka Pharmaceutical Co., Ltd., 463-10, Kagasuno, Kawauchi-cho, Tokushima 771-019, Japan

<sup>e</sup> Department of Pathology Cardinal Glennon Children's Hospital, St. Louis University Health Science Center,

1465 South Grand Blvd. St. Louis, MO 63104, USA

<sup>f</sup> Department of Microbiology, The University of Hong Kong, Pokfulam Road, Hong Kong

<sup>g</sup> Hepatitis Research Center, National Taiwan University College of Medicine, Room 328, 3F, No.1, Sec. 1,

Ren-ai Rd., Jhongjheng District 100, Taipei, Taiwan

<sup>h</sup> Tokyo Medical and Dental University, 1-5-45 Yushima, Bunkyo-ku, Tokyo 113-8549, Japan

<sup>i</sup> Central Institute for Experimental Animals, 1430 Nogawa, Miyamae, Kawasaki, Kanagawa 216-0001, Japan

<sup>j</sup> Pharmaceutical Division, Kirin Brewery Co., 6-26-1 Jingumae, Shibuya, Tokyo 150-8011, Japan

<sup>k</sup> National Institute of Infectious Diseases, 1-23-1 Toyama, Shinjuku-ku, Tokyo 162-8640, Japan

Available online 21 January 2005

#### Abstract

We have investigated to develop novel vaccines against SARS CoV using cDNA constructs encoding the structural antigen; spike protein (S), membrane protein (M), envelope protein (E), or nucleocapsid (N) protein, derived from SARS CoV. Mice vaccinated with SARS-N or -M DNA using pcDNA 3.1(+) plasmid vector showed T cell immune responses (CTL induction and proliferation) against N or M protein, respectively. CTL responses were also detected to SARS DNA-transfected type II alveolar epithelial cells (T7 cell clone), which are thought to be initial target cells for SARS virus infection in human. To determine whether these DNA vaccines could induce T cell immune responses in humans as well as in mice, SCID-PBL/hu mice was immunized with these DNA vaccines. As expected, virus-specific CTL responses and T cell proliferation were induced from human T cells. SARS-N and SARS-M DNA vaccines and SCID-PBL/hu mouse model will be important in the development of protective vaccines.

© 2005 Elsevier Ltd. All rights reserved.

Keywords: SARS DNA vaccine; SCID-PBL/hu; Human CTL

# 1. Introduction

The causative agent of severe acute respiratory syndrome (SARS) has been identified as a new type of corona virus,

SARS corona virus (SARS CoV) [1–3]. SARS has infected more than 8400 patients in about 7 months in over 30 countries and caused more than 800 deaths. The deadly epidemic has had significant impacts on many health, social, economic and political aspects. SARS is assumed to resurge in the near future. However, no SARS vaccine is currently available for clinical use. Therefore, we have developed novel vaccine

<sup>\*</sup> Corresponding author. Tel.: +81 72 252 3021; fax: +81 72 251 2153. *E-mail address:* okm@kch.hosp.go.jp (M. Okada).

<sup>0264-410</sup>X/\$ – see front matter © 2005 Elsevier Ltd. All rights reserved. doi:10.1016/j.vaccine.2005.01.036

candidates against SARS CoV using cDNA constructs encoding the structural antigens; S, M, E, or N protein. In immunized mice, neutralizing antibodies against the virus and T cell immunity against virus-infected-cells were studied, since these immunities play important roles in protection against many virus infections. In particular, CD8<sup>+</sup> CTL plays an important role in T cell immunity dependent protection against virus infections and the eradication of murine and human cancers [4,5]. In the present study, a type II alveolar epithelial cell clone, T7, was used for analyzing precise mechanism of CTL against SARS CoV membrane antigens, as the SARS-CoV infects alveolar epithelial cell in the lungs [6]. Furthermore, the SCID-PBL/hu model, which is capable of analyzing in vivo human immune response, was also used because it is a more relevant translational model for human cases [4].

#### 2. Materials and methods

Three kinds of SARS CoV strains: HKU39849(1), TW-1 and FFM-1(2) and their cDNAs were used. S, M, N or E cDNA was transferred into pcDNA 3.1(+) vector and pcDNA 3.1(+)/vs-His Topo (QIAGEN K K, Tokyo, Japan). These genes were expressed in eukaryotic cells and Escherichia coli. pcDNA 3.1(+) vector, 50 µg each, containing SARS S, M, N, or E DNA was injected i.m. (M.tibia anterior) into C57BL/6 mice (female, 8 weeks CLEA Japan Inc, Japan) and BALB/c mice (female, 8 weeks) three times, at an interval of 7 days. Neutralizing antibodies against SARS CoV in the serum from the mice immunized with SARS S, M, N or -E DNA vaccines were assayed by use of Vero-E6 cell. CTL activity against SARS CoV was studied using human type II alveolar epithelial cells, T7, expressing SARS antigens [6]. PBL from healthy human volunteers were administered i.p. into IL-2 receptor y-chain disrupted NOD SCID mice [IL-2R(-/-) NOD-SCID], and SCID-PBL/hu mice were constructed [4]. SARS DNA vaccines at 50 µg were injected i.m. into the SCID-PBL/hu mice. CTL activity of human CD8-positive lymphocytes in the spleen from SCID-PBL/hu was assessed using IFN- $\gamma$  production and 51Cr-release assay [4,5].

# 3. Results

# 3.1. Induction of CTL against SARS CoV by SARS (N) DNA and SARS (M) DNA vaccine

Spleen cells from C57BL/6 mice immunized with SARS-S, -M, -N or -E DNA vaccine were cultured with syngeneic T7 lung cells transfected with S, M, N or E cDNA. pcDNA 3.1(+) SARS (N) DNA vaccine induced significantly CTL activity (IFN- $\gamma$  production) against N cDNA transfected T7 cells (Fig. 1A). Similarly, SARS M DNA vaccine induced SARS antigen M-specific CTL against T7 cells transfected with SARS M DNA (data not shown).



Fig. 1. Induction of CTL and T cell proliferation against SARS (N). (A) Induction of CTL against SARS (N) antigen in the spleen cells from C57BL/6 mice immunized with SARS (N) DNA vaccine. SARS (N) DNA using pcDNA3.1(+) vector was injected i.m. into C57BL/6 mice three times, at an interval of 7 days. CTL activity was assessed by IFN- $\gamma$  production in the culture of 1 × 10<sup>6</sup> spleen cells and 1 × 10<sup>4</sup> T7 lung alveolar type II epithelial cells transfected with SARS (N) DNA at the E/T ratio of 100:1. IFN- $\gamma$  production was assessed by ELISA assay. (B) Augmentation of lymphocyte proliferation specific for SARS (N) DNA vaccine. 1 × 10<sup>5</sup> responder cells from vaccinated mice were cultured with Mitomycin C treated 1 × 10<sup>4</sup> T7 cells transfected with SARS (N) DNA for 48 h and then Bromodeoxy Uridine (BrdU) was added. Proliferative responses were assessed by BrdU assay.

# 3.2. Augmentation of lymphocyte proliferation specific for SARS CoV antigens by the immunization with SARS (M) DNA and SARS (N) DNA vaccine

The proliferation of splenic T cells stimulated by coculture either with T7 cells transfected with M DNA or SARS M peptide (TW1 M102-116) was strongly augmented by M DNA vaccine (data not shown). SARS N DNA vaccine also induced proliferation of splenic T cells in the presence of recombinant N protein as well as N DNA-transfected T7 cells (Fig. 1B). Thus, both SARS N DNA vaccine and



Fig. 2. SARS (M) DNA vaccine induces in vivo human T cell proliferation against SARS CoV in the SCID-PBL/hu human immune systems.  $4 \times 10^7$  PBL from healthy human volunteers were administered i.p. into IL-2 receptor  $\gamma$ -chain disrupted NOD SCID mice [IL-2R (-/-) NOD-SCID], and SCID-PBL/hu mice were constructed. Fifty micrograms of SARS DNA vaccine was injected i.m. into these SCID-PBL/hu mice.  $1 \times 10^5$  spleen cells from these vaccinated mice were cultured with  $10{\sim}50\,\mu g$  of SARS M peptide for 3 days. Proliferation was assayed by BrdU.

M DNA vaccine were shown to induce T cell immune responses against the relevant SARS CoV antigens.

# 3.3. SARS M DNA and N DNA vaccines induced human T cell immune responses (CTL and proliferation) in SCID-PBL/hu model

The M DNA vaccine enhanced the CTL activity and proliferation in the presence of M peptide in SCID-PBL/hu mice (Fig. 2). Furthermore, the SARS N DNA vaccine induced CTL activity (IFN- $\gamma$  production by recombinant N protein or N protein pulsed-autologous B blast cells) and proliferation of spleen cells in SCID-PBL/hu mice (Fig. 3). From these results, it was demonstrated that SARS M DNA vaccine and N DNA vaccine induced human CTL and human T cell proliferative responses.

### 4. Discussion

We have demonstrated that SARS (M) DNA and (N) DNA vaccines induce virus-specific immune responses (CTL and T cell proliferation) in the mouse systems using type II lung alveolar T cell lines in clone target models [6]. These DNA vaccines induced SARS-CoV-specific CTL and T cell proliferation in vivo human immune systems using SCID-PBL/hu. Gao et al. developed adenovirus based a SARS DNA vaccine encoding S1 polypeptide was capable of inducing neutralizing antibody, while another SARS DNA vaccine encoding N protein generated IFN- $\gamma$  producing T cells in rhesus monkeys [7]. SARS S DNA vaccine which elicits effective neutralizing antibody responses that generate protective immunity



Fig. 3. SARS (N) DNA vaccine induces in vivo human CTL against SARS CoV in the SCID-PBL/hu human immune systems.  $4 \times 10^7$  PBL from healthy human volunteers were administered i.p. into IL-2 receptor  $\gamma$ -chain disrupted NOD SCID mice [IL-2R (-/-) NOD-SCID], and SCID-PBL/hu mice were constructed 50 µg of SARS (N) DNA vaccine or 50 µg of SARS (S) DNA vaccine.  $1 \times 10^5$  spleen cells from SCID-PBL/hu were cultured with 10 µg of recombinant SARS (N) protein for 72 h. IFN- $\gamma$  production in the culture supernatant was assayed using ELISA.

in a mouse model [8]. However its immunogenicity in humans has yet to be established. Therefore, it is very important to evaluate the efficacy of SARS DNA vaccine in a SCID-PBL/hu mice, which is a highly relevant translational model for demonstrating human immune responsiveness. Recently, SARS DNA vaccines capable of inducing human neutralizing antibodies against SARS CoV have been established by our SCID-PBL/hu model. It has been demonstrated that Angiotensin-converting enzyme 2 (ACE2) is a functional receptor for the SARS CoV [9]. A transgenic mouse with human ACE-2 may be useful as an animal model of SARS. Furthermore, ACE-2 transgenic SCID mice should be useful as a human model for pre-clinical trial for SARS vaccines, since ACE-transgenic SCID-PBL/hu model could analyze the human immune responses against SARS infection in vivo. The effect of combination immunization with such SARS vaccines and neutralizing antibody dependent DNA vaccine is now being studied. These DNA vaccines should provide a useful tool for development of protective vaccines.

### Acknowledgements

This study was supported by Grant-in-Aid for the science and technology and Grant-in-Aid for Scientific Research on Priority Areas from the Ministry of Education Culture Sports, Science and Technology, Japan. This study was also supported by a Heath and Labour Science Research Grant from the Ministry of Health, Labour, and Welfare, Japan.

#### References

 Peiris JS, Lai ST, Poon LL, Guan Y, Yam LY, Lim W, et al. SARS study group. Coronavirus as a possible cause of severe acute respiratory syndrome. Lancet 2003;361(9366):1319–25.

- [2] Drosten C, Gunther S, Preiser W, van der Werf S, Brodt HR, Becker S, et al. Identification of a novel coronavirus in patients with severe acute respiratory syndrome. N Engl J Med 2003;348(20):1967–76.
- [3] Peiris JS, Yuen KY, Osterhaus AD, Stohr K. The severe acute respiratory syndrome. N Engl J Med 2003;349(25):2431–41.
- [4] Tanaka F, Abe M, Akiyoshi T, Nomura T, Sugimachi K, Kishimoto T, et al. The anti-human tumor effect and generation of human cytotoxic T cells in SCID mice given human peripheral blood lymphocytes by the in vivo transfer of the Interleukin-6 gene using adenovirus vector. Cancer Res 1997;57(7):1335–43.
- [5] Okada M, Yoshimura N, Kaieda T, Yamamura Y, Kishimoto T. Establishment and characterization of human T hybrid cells secreting immunoregulatory molecules. Proc Natl Acad Sci USA 1981;78(12):7717–21.
- [6] deMello DE, Mahmoud S, Padfield PJ, Hoffmann JW. Generation of an immortal differentiated lung type-II epithelial cell line from the adult H-2K(b)tsA58 transgenic mouse. In Vitro Cell Dev Biol Anim 2000;36(6):374–82.
- [7] Gao W, Tamin A, Soloff A, D'Aiuto L, Nwanegbo E, Robbins PD, et al. Effects of a SARS-associated coronavirus vaccine in monkeys. Lancet 2003;362(9399):1895–6.
- [8] Yang ZY, Kong WP, Huang Y, Roberts A, Murphy BR, Subbarao K, et al. A DNA vaccine induces SARS coronavirus neutralization and protective immunity in mice. Nature 2004;428(6982):561–4.
- [9] Li W, Moore MJ, Vasilieva N, Sui J, Wong SK, Berne MA, et al. Angiotensin-converting enzyme 2 is a functional receptor for the SARS coronavirus. Nature 2003;426(6965):450–4.