

research article

Perioperative increase in neutrophil CD64 expression is an indicator for intra-abdominal infection after colorectal cancer surgery

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Background. Colorectal surgery is associated with a high incidence of postoperative infections. Early clinical signs are difficult to distinguish from the systemic inflammatory response related to surgical trauma. Timely diagnosis may significantly improve the outcome. The objective of this study was to compare a new biomarker index CD64 for neutrophils (iCD64n) with standard biomarkers, white blood cell (WBC) count, neutrophil/lymphocyte ratio (NLR), C-reactive protein (CRP) and procalcitonin (PCT) for the early detection of postoperative infection.

Methods. The prospective study included 200 consecutive patients with elective colorectal cancer surgery. Postoperative values of biomarkers from the postoperative day (POD) 1 to POD5 were analysed by the receiver operating characteristic (ROC) analysis to predict infection. The Cox regression model and the Kaplan-Meier method were used to assess prognostic factors and survival.

Results. The increase of index CD64n (iCD64n) after surgery, expressed as the ratio iCD64n after/before surgery was a better predictor of infection than its absolute value. The best 30-day predictors of all infections were CRP on POD4 (AUC 0.72, 99% CI 0.61–0.83) and NLR on POD5 (AUC 0.69, 99% CI 0.57–0.80). The best 15-day predictors of organ/space surgical site infection (SSI) were the ratio iCD64n on POD1 (AUC 0.72, 99% CI 0.58–0.86), POD3 (AUC 0.73, 99% CI 0.59–0.87) and CRP on POD3 (AUC 0.72, 99% CI 0.57–0.86), POD4 (AUC 0.79, 99% CI 0.64–0.93). In a multivariate analysis independent risk factors for infections were duration of surgery and perioperative transfusion while the infection itself was identified as a risk factor for a worse long-term survival.

Conclusions. The ratio iCD64n on POD1 is the best early predictor of intra-abdominal infection after colorectal cancer surgery. CRP predicts the infection with the same predictive value on POD3.

Key words: colorectal surgery; index CD64n; postoperative infection

Introduction

Colorectal cancer surgery is often followed by postoperative complications. They appear in 24–38%^{1,2}, prolong hospitalization and increase hospital cost. The perioperative mortality rate has been reported to be 3–4%.^{2,3} The most common are infectious complications, especially surgical site infections (SSIs). SSIs are divided into incisional (superficial and deep) wound infections and organ/space infec-

tions, which are mostly the result of anastomotic leak.⁴ Intra-abdominal infection can be manifested as abscess, local or diffuse peritonitis.⁵ The incidence of SSI after elective colorectal resection is 5–30%.^{6,7} Rectal surgery has a higher risk for infection because of longer duration and greater bacterial contamination compared with colon surgery.^{8,9} Postoperative infectious complications, particularly severe infections influence patient outcomes and worsen long-term survival.¹⁰⁻¹² The most common

mechanisms causing this are deregulated host immune response during the infection and extraluminal implantation of malignant cells in anastomotic leakage.¹³

Early clinical signs of postoperative infections are nonspecific and difficult to distinguish from the systemic inflammatory response syndrome (SIRS) triggered by surgical trauma. SIRS is usually self-limiting or may progress to infection, sepsis and septic shock.¹⁴ The median time to diagnosis of infection has been reported to be from POD (postoperative day) 7 to POD9.^{4,7,15-19} Organ/space SSIs have been diagnosed significantly later than incisional SSIs.²⁰ Most causes of infection, such as anastomotic leak, can appear much earlier.⁵ Early identification of patients with a high probability of infections is necessary so that clinicians may focus on additional diagnostic investigations. Pre-emptive antibiotic therapy decreases the incidence and severity of postoperative infections and significantly improves the outcome.²¹

The most commonly used laboratory test during the postoperative period, namely white blood cell (WBC) count is neither very sensitive nor specific.^{22,23} Many studies affirmed the predictive value of a non-specific C-reactive protein (CRP) for infection after surgery, but it is more reliable if analysed together with the clinical assessment.²⁴⁻²⁶ The results of procalcitonin (PCT) studies have been contradictory. In some studies PCT proved to be as good as or even better predictor of infections than CRP^{5,23,27,28}, but in others worse than CRP.^{29,30} Neutrophil/lymphocyte ratio (NLR) is a marker of immunosuppression and is increased in SIRS after major surgery, polytrauma, endotoxaemia and sepsis.³¹ In some studies it proved to be a predictor of all complications after abdominal surgery.^{32,33}

A biological marker which could predict infections before the development of clinical signs and symptoms develop is needed. Therefore we studied a new biomarker neutrophil CD64 (CD64n), in laboratory analysis expressed as an index CD64n (iCD64n). CD64 is a high-affinity Fc receptor for IgG1 and IgG3 subclasses of immunoglobulins (Fc γ RI), expressed on macrophages, monocytes, less on eosinophils and very weakly on non-activated neutrophils.^{34,35} Neutrophil expression of CD64 is down-regulated or lost with cell maturation and strongly up-regulated in response to pro-inflammatory cytokines in SIRS and sepsis.³⁶⁻³⁹ The main functions triggered by Fc γ Rs include phagocytosis, enzyme release and clearance of immune complexes.⁴⁰ The expression of CD64n can be induced by bacteria as well as viruses.^{41,42} Two me-

ta-analyses by Cid *et al.*⁴³ and Li *et al.*⁴⁴ concluded iCD64n could be a promising diagnostic biomarker for bacterial infections. Another meta-analysis reported iCD64n is a helpful marker for early diagnosis of sepsis in critically ill adult patients⁴⁵ and in neonates.⁴⁶⁻⁴⁸ It can differentiate systemic infection from disease flare in patients with inflammatory autoimmune diseases.⁴⁹

A new biomarker iCD64n has up to now been investigated very scarcely after a major surgery.^{34,50-54} The objective of this study was to compare iCD64n with standard predictive markers of infections - WBC count, NLR, CRP and PCT - after colorectal cancer resection. We investigated the risk factors of infection and their impact on survival.

Patients and methods

In this prospective study 200 consecutive patients with elective colorectal carcinoma surgery were included. The study was conducted at the surgery department of the Institute of Oncology Ljubljana from September 2010 to March 2013. The study protocol was approved by the Republic of Slovenia National Medical Ethics Committee. All the patients provided written consent for data collection and publication.

The exclusion criteria were preoperative infection, preoperative ileus and palliative surgical procedure. Neo-adjuvant chemotherapy (CTX) and/or radiotherapy (RT) were carried out in 118 (59%) patients and finished six to eight weeks before surgery. The stage of tumour was evaluated clinically according to the nuclear magnetic resonance (NMR) investigation before the beginning of the treatment. The stage of tumour without neo-adjuvant CTX/RT was diagnosed by histopathological examination. The TNM (Tumour, Node, Metastasis) classification was used for staging colorectal cancer disease.⁵⁵ Each patient was assessed preoperatively according to American Society of Anesthesiologists (ASA) physical status classification which accurately predicts morbidity and mortality.^{56,57} Bioelectric impedance analysis (BIA) measurement was performed on the day before the surgery. Body mass index (BMI) was calculated according to the formula in which body mass (kg) is divided by the square of the body height (cm).

On the day of the surgery blood samples for inflammation markers, albumins and haematocrit analysis were taken in the operating room before intravenous fluid application started. Each patient received systemic antibiotic prophylaxis for Gram

negative and anaerobic bacteria on induction of anaesthesia and prior to skin incision. A standardized protocol for general and epidural anaesthesia was used. During the surgery we recorded the length of the procedure, the blood loss volume and the volume of blood transfusion. The body temperature was measured at the end of the procedure.

Values of WBC count, WBC differential, CRP, PCT and iCD64n were recorded just before surgery and after surgery daily from POD1 to POD5. NLR was calculated by dividing the number of neutrophils by the number of lymphocytes. The ratio iCD64 was calculated by dividing a postoperative value of iCD64n by its preoperative value. SIRS criteria from POD1 to POD5 and postoperative infections up to 30 days after surgery were recorded. SSIs (incisional, organ/space), pneumonia, central venous catheter related bloodstream infections (CRBSIs), urinary tract infections (UTIs) and enterocolitis were recorded. Microbial cultures were used as the gold standard to identify the source of infection. Criteria for SIRS reconsidered in the year 2001 by SCCM/ESICM/ACCP/ATS/SIS International Sepsis Definitions Conference⁵⁸ and criteria for nosocomial infections defined by Centres for Disease Control and Prevention (CDC) were used.⁵⁹ The perioperative transfusion of red blood cells was considered as a risk factor for infection until the infection has developed and as a risk factor for survival 30 days after surgery. The duration of hospitalization was recorded. After discharge each patient was monitored by outpatient clinic examinations 30 days after surgery. The data from medical records and the data of the Cancer Registry of the Republic of Slovenia were used to follow survival.

Biomarkers measurement

WBC count (reference range $4\text{--}10 \times 10^9/\text{L}$), WBC differential (neutrophil count $1.50\text{--}7.40 \times 10^9/\text{L}$, lymphocyte count $1.10\text{--}3.50 \times 10^9/\text{L}$) and haematocrit (reference range 0.390–0.500) were analysed with a haematological blood analyser LH75 (Beckman Coulter). The immunobiochemic analyser Modular Analytics SWE (Roche Diagnostics) was used for serum samples analysis. Serum concentration of CRP (reference range 0–5mg/L) was measured by immunoturbidimetric method, PCT (reference range 0–0.5µg/L) by electrochemiluminescence method and albumins (reference range 35–52g/L) by bromocresol green method.

Neutrophil CD64 was quantified by flow cytometry using the Leuko64 assay (Trillium Diagnostics, LLC, Maine, USA) according to the manufacturer's

instructions. Whole blood EDTA-anticoagulated samples were used for analysis. During working days samples were immediately transported to the laboratory and analysed on the same day. The results were available after a few hours. During weekends samples were stored up to 48 hours in refrigerator at 4°C. 100 µL of the whole blood was incubated for 15 minutes in the dark at room temperature with a mixture of murine monoclonal antibodies. Fluorescence beads were then added and flow cytometer analysis was performed. The iCD64 was derived by the ratio of linearized mean fluorescence intensity (MFI) of the cell population to the fluorescein isothiocyanate (FITC) signal from the beads. An internal negative control was the automatically measured lymphocyte iCD64 (< 1.0) and an internal positive control was the automatically measured monocyte iCD64 (> 3.0). The limit value of iCD64n for probable sepsis was > 1.5.

Bioelectric impedance analysis (BIA)

BIA was performed using a portable bioelectrical impedance analyser BodyStat QuadScan 4000 (Douglas, Great Britain). Phase angle (PA) is a ratio between the reactance (X_c) and resistance (R).⁶⁰ The normal range for men is from 7.90° to 6.19° and for women 7.04° to 5.64°.⁶¹ The illness marker (IM) is the ratio between the impedance measurement at 200kHz and 5kHz. A ratio closer to 1.00 indicates poor cellular health or extreme fluid overload.

Statistical analysis

The t-test was used to compare numeric variables (age, BMI, temperature, PA, IM and albumins) between infected and non-infected patients. Non-normally distributed continuous variables were summarised using medians and interquartile ranges and Mann-Whitney U test was used to compare these variables (infected versus non-infected patients). Categorical variables were analysed with the Pearson's chi-square test and the Fisher's exact test. To counteract the problem of multiple comparisons the Holm-Bonferroni corrected p values were used.⁶²

The predictive values of biomarkers WBC, NLR, CRP, PCT, iCD64 and the ratio iCD64n for postoperative infection were assessed by the receiver operating characteristic (ROC) curve. Each cut-off value was determined by using the maximum value of the Youden index. The Bonferroni correction for AUC confidence intervals was used because multiple statistical tests were performed. The cal-

TABLE 1. Comparison of patient and surgical characteristics between infected and non- infected group

Characteristic		Total N=200	No infection N=132	Infection N=68	p-value	Corrected ^a p-value
Age (years)		62.8 (11.3)	62.2 (11.0)	63.8 (11.9)	0.3787 ^f	1
Gender	male	131 (65.5%)	86 (65.2%)	45 (66.2%)	1 ^g	1
	female	69 (34.5%)	46 (34.8%)	23 (33.8%)		
ASA score	I	14 (7%)	12 (9.1%)	2 (2.9%)	0.0503 ^h	0.8546
	II	105 (52.5%)	74 (56.1%)	31 (45.6%)		
	III	76 (38%)	42 (31.8%)	34 (50%)		
	IV	5 (2.5%)	4 (3%)	1 (1.5%)		
Diabetes mellitus	no	161 (80.5%)	110 (83.3%)	51 (75%)	0.2222 ^g	1
	yes	39 (19.5%)	22 (16.7%)	17 (25%)		
BMI (kg/m ²)		27.2 (4.3)	27.3 (4.1)	27.2 (4.6)	0.8461 ^f	1
Phase angle ^a (°)		5.4 (1.0)	5.5 (1.0)	5.3 (1.1)	0.4632 ^f	1
Illness marker ^a		0.81 (0.04)	0.81 (0.04)	0.81 (0.04)	0.8567 ^f	1
Dry lean body mass ^a (kg)		12.2 (9.8-16.5)	12.8 (9.9-16.5)	11.3 (8.8-16.5)	0.1852 ^f	1
Hematocrit ^b	≥ 38%	100 (50%)	66 (50%)	34 (50%)	1 ^h	1
	30-37%	87 (43.5%)	57 (43.2%)	30 (44.1%)		
	26-29%	12 (6%)	8 (6.0%)	4 (5.9%)		
	21-25%	1 (0.5%)	1 (0.8%)	0 (0%)		
Albumin ^b (g/l)		42.1 (3.6)	42.4 (3.3)	41.6 (4.0)	0.1583 ^f	1
Tumour site	rectum	137 (68.5%)	84 (63.6%)	53 (77.9%)	0.0909 ^h	1
	colon	60 (30%)	46 (34.8%)	14 (20.6%)		
	rectum+ colon	3 (1.5%)	2 (1.5%)	1 (1.5%)		
Stage (TNM)	0	3 (1.5%)	2 (1.5%)	1 (1.5%)	0.0265 ^h	0.477
	I	27 (13.5%)	21 (15.9%)	6 (8.8%)		
	II	42 (21%)	34 (25.8%)	8 (11.8%)		
	III	109 (54.5%)	66 (50%)	43 (63.2%)		
	IV	19 (9.5%)	9 (6.8%)	10 (14.7%)		
Preoperative RT/CTX	no	82 (41%)	62 (47%)	20 (29.4%)	0.0251 ^g	0.477
	yes	118 (59%)	70 (53%)	48 (70.6%)		
Antibiotic prophylaxis	< 24 hours	54 (27%)	35 (26.5%)	19 (27.9%)	0.9404 ^h	1
	24 hours	132 (66%)	87 (65.9%)	45 (66.2%)		
	> 24 hours	14 (7%)	10 (7.6%)	4 (5.9%)		
Surgical procedure						
Rectum resection	LAR	86 (43%)	62 (47%)	24 (35.3%)	0.0034 ^h	0.0749
	Miles + Hartmann	50 (25%)	23 (17.4%)	27 (39.7%)		
Colon resection		64 (32%)	47 (35.6%)	17 (25%)		
Synchronous resection of liver metastases	no	186 (93%)	126 (95.5%)	60 (88.2%)	0.0784 ^h	1
	yes	14 (7%)	6 (4.5%)	8 (11.8%)		
Duration of surgery (min)		170 (130-220)	160 (120-196.2)	200 (150-242.5)	< 0.0001 ^f	0.0004
Loss of blood (ml)		500 (300-800)	400 (200-675)	600 (400-1000)	< 0.0001 ^f	0.0003
Temperature ^c (°C)		35.3 (0.6)	35.3 (0.5)	35.3 (0.7)	0.8719 ^f	1
Perioperative ^d transfusion of PRBC (ml)		0 (0-606.2)	0 (0-326.2)	345 (0-842.5)	< 0.0001 ^f	0.0009
SIRS 1 (POD 1)	no	138 (71.1%)	95 (74.8%)	43 (64.2%)	0.1657 ^g	1
	yes	56 (28.9%)	32 (25.2%)	24 (35.8%)		
SIRS 2 (POD 2)	no	147 (75.4%)	100 (78.7%)	47 (69.1%)	0.1895 ^g	1
	yes	48 (24.6%)	27 (21.3%)	21 (30.9%)		
SIRS 3 (POD 3)	no	161 (82.6%)	107 (84.3%)	54 (79.4%)	0.5151 ^g	1
	yes	34 (17.4%)	20 (15.7%)	14 (20.6%)		
SIRS 4 (POD 4)	no	149 (77.2%)	104 (83.2%)	45 (66.2%)	0.012 ^g	0.2392
	yes	44 (22.8%)	21 (16.8%)	23 (33.8%)		
SIRS 5 (POD 5)	no	144 (80.9%)	99 (87.6%)	45 (69.2%)	0.005 ^g	0.1055
	yes	34 (19.1%)	14 (12.4%)	20 (30.8%)		
Re-operation	no	188 (94%)	132 (100%)	56 (82.4%)	< 0.0001 ^h	< 0.0001
	yes	12 (6%)	0 (0%)	12 (17.6%)		

ASA = American Society of Anesthesiologists; BMI = body mass index; CTX – chemotherapy; Hartmann = proctosigmoidectomy; LAR = low anterior rectum resection; Miles = abdominoperineal rectum resection; POD = postoperative day; PRBC = packed red blood cells; RT = radiotherapy; SIRS = systemic inflammatory response syndrome; TNM = classification of malignant tumors (Tumour, Nodes, Metastasis)

^a measured one day before surgery; ^b in the morning before surgery; ^c at the end of surgery; ^d during the surgery and 30 days after the surgery or until the infection develops; ^e Holm-Bonferroni correction; ^f T-test (mean, standard deviation); ^g Chi-square test; ^h Fisher's exact test; ⁱ Mann-Whitney U test (median, interquartile range)

culated confidence interval (CI) for AUC was 99%, but interpreted as the 95% one.⁶³ Multivariate logistic regression analysis was used to explore the prediction of the probability of infection. Considering survival of the patients, Kaplan-Meier curves between groups of infected and non-infected patients were compared by the log-rank test.⁶⁴ Prognostic factors were investigated by univariate and multivariate Cox proportional hazard model. A p-value ≤ 0.05 was considered statistically significant. All analyses were performed with R statistical software, version 3.2.1.

Results

A total of 200 patients were included in the study, 131 males (65.5%) and 69 females (34.5%). The characteristics of the patients, cancer disease and surgical procedure are shown in Table 1.

Infectious complications

Sixty-eight patients (34%) developed infectious complications, 132 patients did not. Sepsis was diagnosed in 47 patients (23.5%). Two infections were diagnosed in 10 patients. The most frequent postoperative infection was SSI in 61 patients (30.5%). Incisional SSI was diagnosed as the first infection in 28 patients, organ/space SSI in 30 patients, both of them in 3 patients. Most of the incisional SSIs (58%) were diagnosed in patients with abdominoperineal rectum resection and most of organ/space SSIs (52%) in patients with LAR (low anterior rectum resection). Other infections were quite rare (8 UTIs, 5 pneumonias, 1 *Clostridium difficile* enterocolitis). The median number of days until the first infection occurred was seven with interquartile range (IQR) 6–9. The median time to organ/space SSI occurrence was 8 days (IQR 6–12). Antibiotic therapy was applied in 116 patients. In the cases of negative microbiologic results and clinical signs not concordant with infection it was stopped after a few days. Reoperation was necessary in 12 patients (6%), because of infection in 7 and for other reasons in 5 patients.

Risk factors for any perioperative infection as shown by univariate analysis were: duration of surgery (corrected $p = 0.0004$), loss of blood (corrected $p = 0.0003$) and perioperative transfusion of red blood cells (corrected $p = 0.0009$). Surgical procedure and SIRS on POD5 were on the threshold of statistical significance for postoperative infections. Albumins before surgery, PA, IM and dry lean body

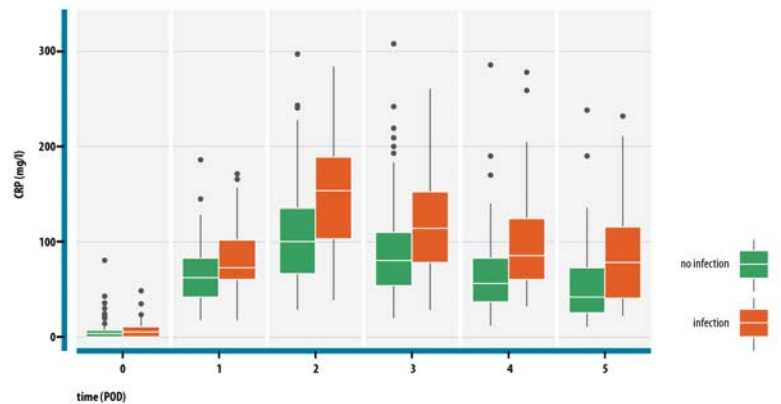


FIGURE 1. The dynamics of CRP shown with median for infected/non-infected group 5 days after colorectal surgery. The width of the box shows the interquartile range. The distal points of vertical line show the highest and the lowest values of CRP.

CRP = C-reactive protein; POD = postoperative day

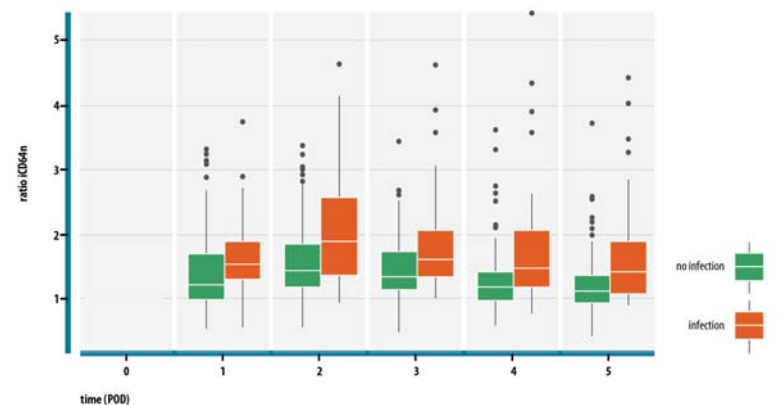


FIGURE 2. The dynamics of the ratio iCD64n shown with median 5 days after colorectal surgery. The width of the box shows the interquartile range. The distal points of vertical line show the highest and the lowest values of the ratio iCD64n.

iCD64n = index CD64n

mass were not found to be risk factors for postoperative infections. A risk factor for organ/space SSI was perioperative transfusion (corrected $p < 0.0001$) while the blood loss was at the border of statistical significance (corrected $p = 0.07$). The hospital stay of 19 days (IQR 14–24) in the infected group was significantly longer as compared to 10 days (IQR 7–12) in the non-infected group ($p < 0.0001$).

Multivariate analysis

The multiple logistic regression analysis was made for prediction of all postoperative infections. It included ASA score, type of surgical procedure, duration of surgery and perioperative transfusion. Independent risk factors for infections were found to be duration of surgery (odds ratio [OR] 1.63, 95%

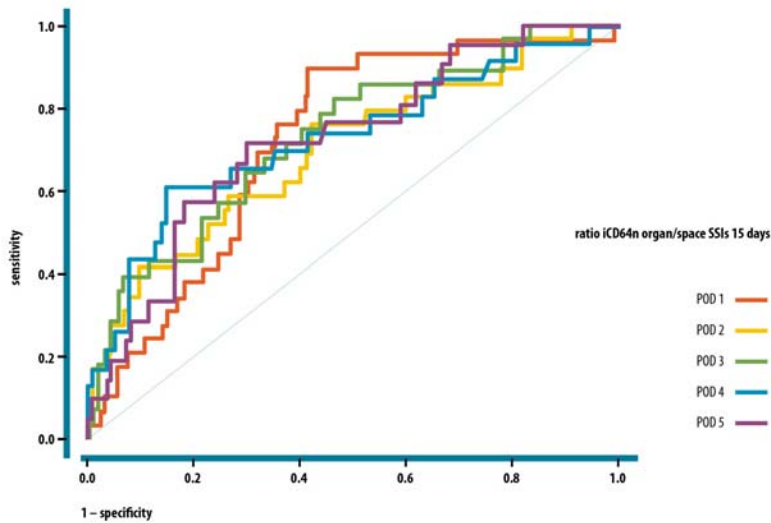


FIGURE 3. ROC curve for the ratio iCD64n POD1-POD5 as a predictor of organ/space SSIs 15 days after colorectal surgery.

iCD64n = index CD64n; POD = postoperative day; ROC = receiver operating characteristic; SSIs = surgical site infections

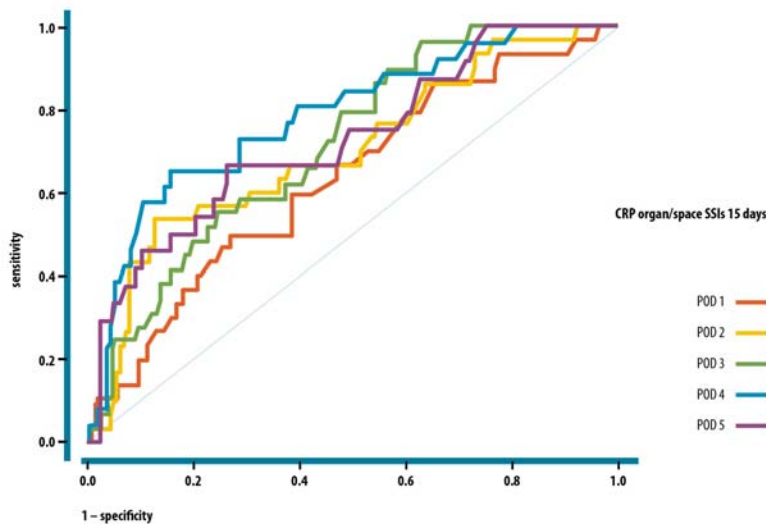


FIGURE 4. ROC curve for CRP POD1-POD5 as a predictor of organ/space SSIs 15 days after colorectal surgery.

CRP = C-reactive protein; POD = postoperative day; ROC = receiver operating characteristic; SSIs = surgical site infections

CI 1.14–2.40, $p = 0.01$) and perioperative transfusion (OR 1.10, 95% CI 1.02–1.19, $p = 0.014$). The expected odds for infection increased with every hour of surgery by 63% (95% CI 14–140%) or with every 100 mL of transfusion by 10% (95% CI 2–19%).

Biomarkers analysis

The values of all biomarkers after surgical procedures were elevated, reaching the peak values on the first day (WBC count, NLR and PCT) or on the

second day (CRP, iCD64n, ratio iCD64n) and later slowly decreasing. The increase of biomarkers was greater in the infection group than in the non-infection group (Figure 1,2).

The ROC analysis was used to compare biomarkers WBC count, NLR, CRP, PCT, iCD64n and the ratio iCD64n from POD1 to POD5 as early predictors of postoperative infections. Predictions for all infections in 15 and 30 days and for organ/space SSIs in 15 days were made. The 15-day prediction was made because the great majority (97%) of the first infections had been diagnosed up to and including this day. The highest diagnostic accuracy for 30-day prediction of all infections was for CRP observed on POD4 (AUC 0.72, 99% CI 0.61–0.83), POD2 (AUC 0.70, 99% CI 0.59–0.80) and POD3 (AUC 0.69, 99% CI 0.58–0.80). The cut-off value for CRP POD4 was 69 mg/L. Two other, rather good predictors were NLR on POD4 (AUC 0.65, 99% CI 0.54–0.77), POD5 (AUC 0.69, 99% CI 0.57–0.80) and the ratio iCD64n on POD2 (AUC 0.67, 99% CI 0.56–0.78) with the cut-off value 1.74. The results of ROC analysis for 15-day prediction of all infections were similar.

In the 15-day prediction of organ/space infections the diagnostic accuracies of CRP and the ratio iCD64n were better than in the two previously mentioned analyses for all infections. The ratio iCD64n was a better predictor on POD3 (AUC 0.73, 99% CI 0.59–0.87) and POD1 (AUC 0.72, 99% CI 0.58–0.86), followed by POD4 (AUC 0.72, 99% CI 0.57–0.88) and POD2 (AUC 0.70, 99% CI 0.55–0.84). The cut-off value for POD1 was 1.37 and POD3 1.40 (Figure 3). The predictive value of CRP was the best on POD4 (AUC 0.79, 99% CI 0.64–0.93) followed by POD5 (AUC 0.73, 99% CI 0.57–0.88), POD3 (AUC 0.72, 99% CI 0.57–0.86) and POD2 (AUC 0.70, 99% CI 0.56–0.85). The cut-off value for CRP on POD4 was 103 mg/L (Figure 4). The prognostic value of PCT for 15-day prediction of organ/space infection was most sensitive on POD4 (AUC 0.72, 99% CI 0.57–0.88) (Figure 5) and for iCD64n on POD5 (AUC 0.69, 99% CI 0.53–0.85).

Survival

The thirty-day mortality was 0% and the ninety-day mortality 0.5%. The Kaplan Meier analysis showed a one-year survival rate in the non-infected group was 97.7% (95% CI 93.1–99.3) and in the infected group 92.6% (95% CI 83.2–96.9). A two-year survival was significantly higher in the non-infected group 92.4% (95% CI 86.4–95.9) as compared to the infected group 80.9% (95% CI 69.4–88.4). P value in log-rank test was 0.0134 (Figure 6). A me-

dian follow-up of patients was 40.9 months (range 32.4–51.5).

Prognostic factors were investigated by univariate and multivariate Cox proportional hazard model. Predictive factors for shorter survival in univariate analysis were: age ($p < 0.0001$), ASA score ($p = 0.0006$), tumour stage ($p = 0.027$), PA ($p = 0.0024$), dry lean body mass ($p = 0.015$), perioperative transfusion ($p = 0.0004$) and postoperative infection ($p = 0.016$). Multivariate analysis showed independent factors associated with shorter survival were: age (hazard ratio [HR] 1.06, 95% CI 1.02–1.10, $p = 0.0044$) and postoperative infection (HR 1.96, 95% CI 1.03–3.73, $p = 0.04$).

Discussion

Colorectal surgery is associated with an intensive release of pro-inflammatory cytokines followed by an anti-inflammatory response and immuno-paralysis. Cellular immunity, crucial for defence against cancer cells in the perioperative period is significantly suppressed.⁶⁵⁻⁶⁷ Postoperative immunosuppression can be further exacerbated by blood transfusion. These immune changes predispose the host to infection, sepsis and even multiple organ dysfunction syndrome (MODS).⁶⁸

In the present study we compared a new biomarker iCD64n and the ratio iCD64n with other biomarkers - WBC count, NLR, CRP and PCT. We found out the best early predictor of organ/space SSIs was the ratio iCD64n. CRP and PCT predicted these infections with the same AUC later, on POD3 and POD4. Other biomarkers as predictive factors for infections after colorectal surgery had already been much studied widely whereas iCD64n had not yet been.

Warschkow *et al.* reported in a diagnostic meta-analysis of 1832 patients the best CRP predictive value for postoperative complications after colorectal surgery was 135 mg/L on POD4 with a negative predictive value (NPV) 89%.⁶⁹ In another retrospective study with 1187 patients Warschkow *et al.* concluded CRP on POD4 with the cut-off 123 mg/L had the highest diagnostic accuracy for the early detection of infections (sensitivity 66%, specificity 77%).²² In a meta-analysis with 2215 patients Gans *et al.* reported infectious complications after major abdominal surgery were very unlikely in patients with CRP below 159 mg/L on POD3 (pooled values: AUC 0.87, sensitivity 77%, specificity 77%, NPV 90%). Maximum predictive values were reached on POD5 (pooled values: AUC 0.83, sen-

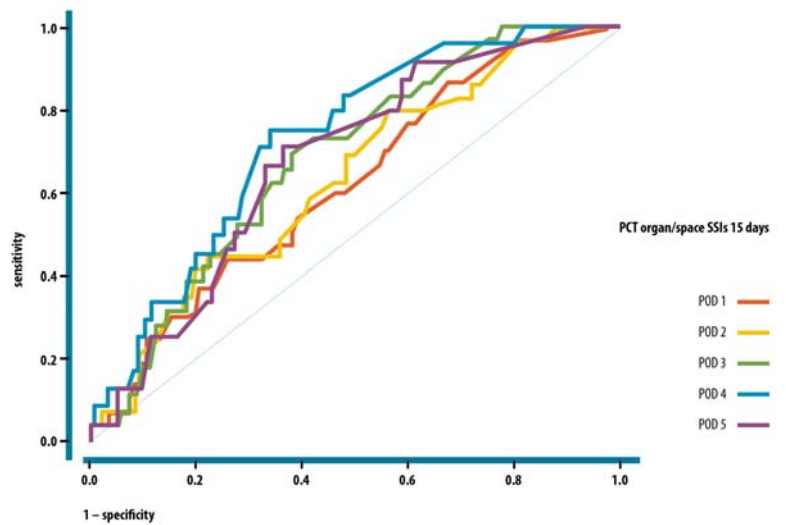


FIGURE 5. ROC curve for PCT POD1-POD5 as a predictor of organ/space SSIs 15 days after colorectal surgery.

POD = postoperative day; PCT = procalcitonin; ROC = receiver operating characteristic; SSIs = surgical site infections

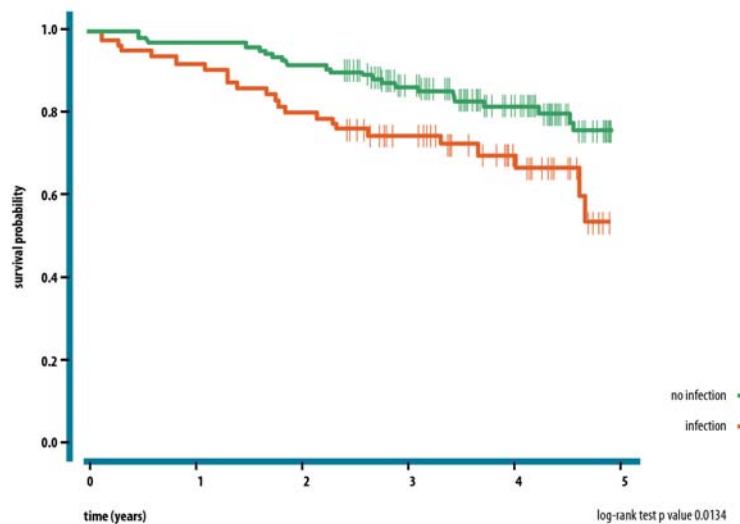


FIGURE 6. Kaplan-Meier survival curve for infected and non-infected group. Patients have been followed up for a minimum of 28 months.

sitivity 86%, specificity 86%, NPV 92%).⁷⁰ The conclusion of the pooled analysis with 1427 patients made by Straatman *et al.* was that CRP on POD3 below 75 mg/L may be a safe discharge criterion after major abdominal surgery with a NPV 97.2%. The probability of major postoperative complications for CRP cut-off 215 mg/L was 20% (95% CI 14.7–25.60%).⁷¹ In our study CRP on POD4 had the best predictive value in the 30-day prediction of all infections and even better on POD4 in the 15-day prediction of organ/space SSIs.

Some recent studies have compared the diagnostic accuracy of PCT and CRP for infection after colorectal surgery. Takakura *et al.* reported in the study which included 114 patients with colorectal resection PCT on POD1 and POD3 (AUC 0.76 and 0.77) was a more relevant predictor for surgical site infection than CRP (AUC 0.71).²⁷ Garcia-Granero *et al.* reported that in 205 patients undergoing surgery for colorectal cancer PCT was better than CRP in the early prediction of major anastomotic leak on POD3, POD4 and POD5. The best AUC values for PCT were 0.87 and for CRP 0.85 on POD5.²³ The results of study, made by Lagoutte *et al.* were different. It included 100 patients and showed PCT is neither earlier nor more accurate than CRP for the detection of anastomotic leakage after colorectal surgery. The best accuracy for CRP and PCT was obtained on POD4 (AUC 0.87 and 0.75).²⁹ Oberhofer *et al.* reported in a study with 79 colorectal surgical patients PCT on POD2 and CRP on POD3 had similar predictive values for infections (AUC 0.75 and 0.75).²⁸ In the present study we found out that PCT on POD4 (AUC 0.64) was a worse predictor than CRP (AUC 0.72) in the 30-day prediction of all infections. In the 15-day prediction of organ/space SSIs PCT on POD4 (AUC 0.72) was worse than CRP (AUC 0.79) again.

As in other studies WBC count proved to be a poor early diagnostic marker of postoperative infections. The best predictive value of NLR was on POD5 (AUC 0.69) in the 30-day prediction of all infections and was as good as for CRP (AUC 0.68).

Only few studies have investigated the dynamics of iCD64n in the postoperative period. The first have been done in cardiovascular and orthopedic surgery. Two studies done by Kolackova *et al.* with 40 cardiac patients³⁴ and Katoh *et al.*⁵⁰ with 41 orthopaedic patients reported the expression of CD64n after surgery was significantly increased with the peak on POD3. Studies by Fjaertoft *et al.*⁵¹ and Gerrits *et al.*⁵² reported that iCD64n was significantly higher in septic patients compared to patients with SIRS after surgery and control group. Unlike clean surgical procedures, the iCD64n in clean-contaminated surgery has only recently been explored. In the study Janež *et al.*⁵³ included 77 patients. They compared postoperative differences in inflammatory and immunological response between opened and laparoscopically assisted colorectal surgery. There was a considerable increase of iCD64n in both groups of patients on POD1 (1.42 in open surgery group versus 1.24 in laparoscopic surgery group). But they did not observe difference in infectious complications in these two

groups. In a very recent study, which included 189 patients with colorectal, 17 with maxillofacial and 23 with open heart surgery, Jukic *et al.*⁵⁴ reported that iCD64n is the best predictor of postoperative infections in the first 48 hours after major surgery compared to WBC count, neutrophils and CRP. The AUC value after 24 hours was 0.89 and after 48 hours 0.82. The most frequent infection was the respiratory tract infection (40%).

In the present study index CD64n was not found to be a good early predictor for any infection including organ/space SSIs. However, we found that the ratio iCD64n was a better predictor of infection than its absolute value. In the 30-day prediction of all infections the ratio iCD64n on POD2 (AUC 0.67) was a worse predictor than CRP (AUC 0.70). However, for the 15-day prediction of organ/space SSI the ratio iCD64n on POD1 (AUC 0.72) was the best early predictor among all studied biomarkers (AUC on POD1 for iCD64n and CRP was 0.63 and for PCT 0.61). Patients with the ratio iCD64n higher than the cut-off value 1.37 on POD1 should be closely monitored and additional diagnostic measures should be taken to confirm or exclude infection. Our study group was homogeneous and it is comprised of patients with clean-contaminated colorectal surgery. SSIs were the most common postoperative infections. We explain this with a high number of rectum surgeries (68%), a high stage of the disease (64% stage III and IV) and a high proportion of patients preoperatively treated with neo-adjuvant CTX and/or RT (59%). Risk factors for infections were analysed. It was shown that duration of surgery and transfusion of red blood cells were independent risk factors for infections. The length of surgery procedure is closely correlated with perioperative immunoparalysis and predisposition to infection.⁷¹

We found out that age, ASA score, tumour stage, PA, dry lean body mass, perioperative transfusion and postoperative infection correlated with the length of survival. Multivariate analysis showed that the only independent factors associated with shorter survival were age and postoperative infection. In our study a perioperative transfusion of red blood cells was not an independent prognostic marker for survival. However, in many other studies postoperative infections and perioperative transfusion were independent risk factors for worse long-term survival.⁷³⁻⁷⁷ Both of them aggravate cytokine response after surgery, suppress cell mediated immunity and can facilitate the growth of tumour cells. Due to the synergistic effect patients with both risk factors represent a group with

a particularly poor prognosis.⁷⁸ The efforts to reduce postoperative infections may have a favourable effect on cancer prognosis.

Conclusions

In the present study we found the ratio iCD64n on POD1 was the earliest predictor of intra-abdominal infection after colorectal cancer surgery. CRP predicts the infection with the same predictive value later, not before POD3. Further research is needed to evaluate the role of neutrophil CD64 expression in infection diagnosis after major surgery. Postoperative infection was found to be an independent predictive factor of shorter long-term survival.

References

- Zoucas E, Lydrup ML. Hospital costs associated with surgical morbidity after elective colorectal procedures: a retrospective observational cohort study in 530 patients. *Patient Saf Surg* 2014; **8**: 2.
- Sjo OH, Larsen S, Lunde OC, Nesbakken A. Short term outcome after emergency and elective surgery for colon cancer. *Colorectal Dis* 2009; **11**: 733-9.
- Alves A, Panis Y, Mathieu P, Mantion G, Kwiatkowski F, Slim K. Postoperative mortality and morbidity in French patients undergoing colorectal surgery: results of a prospective multicentre study. *Arch Surg* 2005; **140**: 278-83.
- Korner H, Nielsen HJ, Soreide JA, Nedrebo BS, Soreide K, Knapp JC. Diagnostic accuracy of C reactive protein for intra-abdominal infections after colorectal resections. *J Gastrointest Surg* 2009; **13**: 1599-606.
- Dominguez-Comesana E, Lopez-Gomez V, Estevez-Fernandez SM, Padin EM, Ballinas-Miranda J, Carrera-Dacosta E, et al. [Procalcitonin and C-reactive protein as early indicators of postoperative intra-abdominal infection after surgery for gastrointestinal cancer]. [Spanish]. *Cir Esp* 2014; **92**: 240-6.
- Tang R, Chen HH, Wang YL, Changchien CR, Chen JS, Hsu KC, et al. Risk factors for surgical site infection after elective resection of the colon and rectum: A single-center prospective study of 2809 consecutive patients. *Ann Surg* 2001; **234**: 181-9.
- Smith RL, Bohl JK, McElearney ST, Friel CM, Barclay MM, Sawyer RG, et al. Wound infection after elective colorectal resection. *Ann Surg* 2004; **239**: 599-607.
- Konishi T, Watanabe T, Kishimoto J, Nagawa H. Elective colon and rectal surgery differ in risk factors for wound infection: results of prospective surveillance. *Ann Surg* 2006; **244**: 758-63.
- Horzic M, Kopljarić M. Postoperative infections in colorectal cancer patients. *Hepatogastroenterology* 2005; **52**: 101-4.
- Nespoli A, Gianotti L, Totis M, Bovo G, Nespoli L, Chiodini P, et al: Correlation between postoperative infections and long term survival after colorectal resection for cancer. *Tumori* 2004; **90**: 485-90.
- Khuri SF, Henderson WG, DePalma RG, Mosca C, Healey NA, Kumbhani DJ, et al. Determinants of long-term survival after major surgery and the adverse effect of postoperative complications. *Ann Surg* 2005; **242**: 326-41.
- Artinyan A, Orcutt ST, Anaya DA, Richardson P, Chen GJ, Berger DH. Infectious postoperative complications decrease long-term survival in patients undergoing curative surgery for colorectal cancer. A study of 12075 patients. *Ann Surg* 2015; **261**: 497-505.
- Tsujimoto H, Ueno H, Hashiguchi Y, Ono S, Ichikura T, Hase K. Postoperative infections are associated with adverse outcome after resection with curative intent for colorectal cancer. *Oncol Lett* 2010; **1**: 119-25.
- Mokart D, Merlin M, Sannini A, Brun JP, Tison A, Delperio JL, et al. Procalcitonin, interleukin 6 and systemic inflammatory response syndrome (SIRS): early markers of postoperative sepsis after major surgery. *Br J Anaesth* 2005; **94**: 767-73.
- Pedersen T, Roikjaer O, Jess P. Increased levels of C-reactive protein and leukocyte count are poor predictors of anastomotic leakage following laparoscopic colorectal resection. *Dan Med J* 2012; **59**: A4552.
- Welsch T, Müller SA, Ulrich A, Kischlat A, Hinz U, Kienle P, et al. C reactive protein as early predictor for infectious postoperative complications in rectal surgery. *Int J Colorectal Dis* 2007; **22**: 1499-507.
- den Dulk M, Noter SL, Hendriks ER, Brouwers MAM, van der Vlies CH, Oostenbroek RJ, et al. Improved diagnosis and treatment of anastomotic leakage after colorectal surgery. *Eur J Surg Oncol* 2009; **35**: 420-6.
- Matthiessen P, Henriksson M, Hallböök O, Grunditz E, Noren B, Arbman G. Increase of serum C-reactive protein is an early indicator of subsequent symptomatic leakage after anterior resection. *Colorectal Dis* 2008; **10**: 75-80.
- Ishikawa K, Kusumi T, Kosokawa M, Nishida Y, Sumikawa S, Furukawa H. Incisional surgical site infection after elective open surgery for colorectal cancer. *Int J Surg Oncol* 2014; **2014**: 419712.
- Blumetti J, Luu M, Sarosi G, Hartless K, McFarlin J, Parker B, et al. Surgical site infections after colorectal surgery: Do risk factors vary depending on the type of infection considered? *Surgery* 2007; **142**: 704-11.
- Chromik AM, Endter F, Uhl W, Thiede A, Reith HB, Mittelkötter U. Preemptive antibiotic treatment vs standard treatment in patients with elevated serum procalcitonin levels after elective colorectal surgery: a prospective randomised pilot study. *Langenbecks Arch Surg* 2006; **391**: 187-94.
- Warschke R, Tarantino I, Torzewski M, Näf F, Lange J, Steffen T. Diagnostic accuracy of C-reactive protein and white blood cell counts in the early detection of inflammatory complications after open resection of colorectal cancer: a retrospective study of 1187 patients. *Int J Colorectal Dis* 2011; **26**: 1405-13.
- Garcia-Granero A, Frasson M, Flor-Lorente B, Blanco F, Puga R Carratala A, et al: Procalcitonin and C-reactive protein as early predictors of anastomotic leak in colorectal surgery: a prospective observational study. *Dis Colon Rectum* 2013; **56**: 475-83.
- Nunes BK, Lacerda RA, Jardim JM. Systematic review and meta-analysis of the predictive value of C-reactive protein in postoperative infections. *Rev Esc Enferm USP* 2011; **45**: 1480-5.
- Neumaier M, Scherer MA. C-reactive protein levels for early detection of postoperative infection after fracture surgery in 787 patients. *Acta Orthop* 2008; **79**: 428-32.
- Welsch T, Frommhold K, Hinz U, Weigand M, Kleeff J, Friess H, et al. Persisting elevation of C-reactive protein after pancreatic resections can indicate developing inflammatory complications. *Surgery* 2008; **143**: 20-8.
- Takakura Y, Hinoi T, Egi H, Shimomura M, Adachi T, Saito Y, et al. Procalcitonin as a predictive marker for surgical site infection in elective colorectal cancer surgery. *Langenbecks Arch Surg* 2013; **398**: 833-9.
- Oberhofer D, Juras J, Pavičić AM, Rančić Žurić I, Rumanjak V. Comparison of C-reactive protein and procalcitonin as predictors of postoperative infectious complications after elective colorectal surgery. *Croat Med J* 2012; **53**: 612-9.
- Lagoutte N, Facy O, Ravoire A, Chalumeau C, Jonval L, Rat P, et al. C-reactive protein and procalcitonin for the early detection of anastomotic leakage after elective colorectal surgery: pilot study in 100 patients. *J Visc Surg* 2012; **149**: e345-9.
- Silvestre J, Rebanda J, Lourenco C, Povoia P. Diagnostic accuracy of C-reactive protein and procalcitonin in the early detection of infection after elective colorectal surgery-a pilot study. *BMC Infect Dis* 2014; **14**: 444.
- Zahorec R. Ratio of neutrophil to lymphocyte counts-rapid and simple parameter of systemic inflammation and stress in critically ill. *Bratisl Lek Listy* 2001; **102**: 5-14.
- Cook EJ, Walsh SR, Farooq N, Alberts JC, Justin TA, Keeling NJ. Post-operative neutrophil-lymphocyte ratio predicts complications following colorectal surgery. *Int J Surg* 2007; **5**: 27-30.
- Forget P, Dinant V, De Kock M. Is the neutrophil to lymphocyte ratio more correlated than C-reactive protein with postoperative complications after major abdominal surgery? *PeerJ* 2015; **3**: e713.
- Kolackova M, Kudlova M, Kunes P, Lonsky V, Mandak J, Andrys C, et al. Early expression of FcγRI (CD64) on monocytes of cardiac surgical patients and higher density of monocyte anti-inflammatory scavenger CD163 receptor in on pump patients. *Mediators Inflamm* 2008; **2008**: 235461.
- Radaev S, Sun P. Recognition of immunoglobulins by Fcγ receptors. *Mol Immunol* 2001; **38**: 1073-83.

36. Davis BH, Olsen SH, Ahmad E, Bigelow NC. Neutrophil CD64 is an improved indicator of infection or sepsis in emergency department patients. *Arch Pathol Lab Med* 2006; **130**: 654-61.
37. Repp R, Valerius T, Sendler A, Gramatzki M, Iro H, Kalden R, et al. Neutrophils express the high affinity receptor for IgG (FcγRI, CD64) after in vivo application of recombinant human granulocyte colony-stimulating factor. *Blood* 1991; **78**: 885-9.
38. van der Meer W, Pickkers P, Scott CS, van der Hoeven JG, Gunnewiek JK. Hematological indices, inflammatory markers and neutrophil CD64 expression: comparative trends during experimental human endotoxemia. *J Endotoxin Res* 2007; **13**: 94-100.
39. Qureshi SS, Lewis SM, Gant VA, Treacher D, Davis BH, Brown KA. Increased distribution and expression of CD64 on blood polymorphonuclear cells from patients with the systemic inflammatory response syndrome (SIRS). *Clin Exp Immunol* 2001; **125**: 258-65.
40. Hoffmeyer F, Witte K, Schmidt RE. The high affinity FcγRI on PMN: regulation of expression and signal transduction. *Immunology* 1997; **92**: 544-52.
41. Jalava Karvinen P, Hohenthal U, Laitinen I, Kotilainen P, Rajamäki A, Nikoskelainen J, et al. Simultaneous quantitative analysis of FcγRI (CD64) and CR1 (CD35) on neutrophils in distinguishing between bacterial infections, viral infections and inflammatory diseases. *Clin Immunol* 2009; **133**: 314-23.
42. Mokuda S, Doi O, Takasugi K. Simultaneous quantitative analysis of CD64 and CD35 on neutrophils as markers to differentiate between bacterial and viral infections in patients with rheumatoid arthritis. *Mod Rheumatol* 2012; **22**: 750-7.
43. Cid J, Aguinaco R, Sanchez R, Garcia Pardo G, Llorente A. Neutrophil CD64 expression as marker of bacterial infection: A systematic review and meta-analysis. *J Infect* 2010; **60**: 313-9.
44. Li S, Huang X, Chen Z, Zhong H, Peng Q, Deng Y, et al. Neutrophil CD64 expression as a biomarker in the early diagnosis of bacterial infection: a meta-analysis. *Int J Infect Dis* 2013; **17**: e12-e23.
45. Wang X, Li ZY, Zeng L, Zhang AQ, Pan W, Gu W, et al. Neutrophil CD64 expression as a diagnostic marker for sepsis in adult patients: a meta-analysis. *Crit Care* 2015; **19**: 245.
46. Bhandari V, Wang C, Rinder C, Rinder H. Hematologic profile of sepsis in neonates: neutrophil CD64 as a diagnostic marker. *Pediatrics* 2008; **121**: 129-34.
47. Fang DH, Fan CH, Li J, An Q, Yao H, Ji Q, et al. Ratios of CD64 expressed on neutrophils, monocytes, and lymphocytes may be a novel method for diagnosis of neonatal sepsis. *J Infect Dev Ctries* 2015; **9**: 175-81.
48. Du J, Li L, Dou Y, Li P, Chen R, Liu H. Diagnostic utility of neutrophil CD64 as a marker for early onset sepsis in preterm neonates. *PLoS One* 2014; **9**: e102647.
49. Allen E, Bakke AC, Purtzer MZ, Deodhar A. Neutrophil CD64 expression: distinguishing acute inflammatory autoimmune disease from systemic infections. *Ann Rheum Dis* 2002; **61**: 522-5.
50. Katoh N, Nishino J, Nishimura K, Kawabata C, Hotta Y, Matsui T, et al. Normal sequential changes in neutrophil CD64 expression after total joint arthroplasty. *J Orthop Sci* 2013; **18**: 949-54.
51. Fjaertoft G, Hakansson LD, Pauksens K, Sisask G, Venge P. Neutrophil CD64 (FcγRI) expression is a specific marker of bacterial infection: A study on the kinetics and the impact of major surgery. *Scand J Infect Dis* 2007; **39**: 525-35.
52. Gerrits JH, McLaughlin PMJ, Nienhuis BN, Smit JW, Loef B. Polymorphic mononuclear neutrophils CD64 index for diagnosis of sepsis in postoperative surgical patients and critically ill patients. *Clin Chem Lab Med* 2013; **51**: 897-905.
53. Janež J, Korač T, Rebolj Kodre A, Jelenc F, Ihan A. Laparoscopically assisted colorectal surgery provides better short-term clinical and inflammatory outcomes compared to open colorectal surgery. *Arch Med Sci* 2015; **11**: 1217-26.
54. Jukic T, Ihan A, Stubljar D. Dynamics of inflammation biomarkers C-reactive protein, leukocytes, neutrophils, and CD64 on neutrophils before and after major surgical procedures to recognize potential postoperative infection. *Scand J Clin Lab Invest* 2015; e1-8.
55. International Union Against Cancer (UICC). TNM classification of malignant tumours, 7th edition. Sobin LH, Gospodarowicz MK, Wittekind Ch, editors. New York: Wiley; 2009.
56. Fitz-Henry J. The ASA classification and perioperative risk. *Ann R Coll Surg Engl* 2011; **93**: 185-7.
57. Daabis M. American society of anaesthesiologists physical status classification. *Indian J Anaesth* 2011; **55**: 111-5.
58. Levy MM, Fink MP, Marshall JC, Abraham E, Angus D, Cook D, et al. 2001 SCCM/ESICM/ACCP/ATS/SIS international sepsis definitions conference. *Crit Care Med* 2003; **31**: 1250-6.
59. Horan TC, Andrus M, Dudeck MA. CDC/NHSN surveillance definition of health care-associated infection and criteria for specific types of infections in the acute care setting. *Am J Infect Control* 2008; **36**: 309-32.
60. Gupta D, Lis CG, Dahlk SL, King J, Vashi PG, Grutsck JF, et al. The relationship between bioelectrical impedance phase angle and subjective global assessment in advanced colorectal cancer. *Nutr J* 2008; **7**: 19.
61. Barbosa-Silva MCG, Barros AJD, Wang J, Heymsfield SB, Pierson RN. Bioelectrical impedance analysis: population reference values for phase angle by age and sex. *Am J Clin Nutr* 2005; **82**: 49-52.
62. Holm S. A simple sequentially rejective multiple test procedure. *Scand J Statist* 1979; **6**: 65-70.
63. Zweig MH, Campbell G. Receiver-Operating Characteristic (ROC) Plots: A fundamental evaluation tool in clinical medicine. *Clin Chem* 1993; **39**: 561-77.
64. Rich JT, Neely JG, Paniello RC, Voelker CJC, Nussenbaum B, Wang EW. A practical guide to understanding Kaplan-Meier curves. *Otolaryngol Head Neck Surg* 2010; **143**: 331-6.
65. Karanika S, Karantanos T, Theodoropoulos GE. Immune response after laparoscopic colectomy for cancer: a review. *Gastroenterol Rep* 2013; **1**: 85-94.
66. Evans C, Galustian C, Kumar D, Hagger R, Melville DM, Bodman-Smith M, et al. Impact of surgery on immunologic function: comparison between minimally invasive techniques and conventional laparotomy for surgical resection of colorectal tumors. *Am J Surg* 2009; **197**: 238-45.
67. Huang C, Huang R, Jiang T, Huang K, Cao J, Qiu Z. Laparoscopic and open resection for colorectal cancer: an evaluation of cellular immunity. *BMC Gastroenterol* 2010; **10**: 127.
68. Choileain NN, Redmond HP. Cell response to surgery. *Arch Surg* 2006; **141**: 1132-40.
69. Warschkow R, Beutner U, Steffen T, Müller SA, Schmied BM, Güller U, et al. Safe and early discharge after colorectal surgery due to C-reactive protein. A diagnostic meta-analysis of 1832 patients. *Ann Surg* 2012; **256**: 245-50.
70. Gans SL, Atema JJ, van Dieren S, Koerkamp BG, Boermeester MA. Diagnostic value of C-reactive protein to rule out infectious complications after major abdominal surgery: a systematic review and meta-analysis. *Int J Colorectal Dis* 2015; **30**: 861-73.
71. Straatman J, Harmsen AMK, Cuesta MA, Berkhof J, Jansma EP, van der Peet DL. Predictive value of C-reactive protein for major complications after major abdominal surgery: A systematic review and pooled-analysis. *PLoS One* 2015; **10**: e0132995.
72. Menges P, Kessler W, Kloecker C, Feuerherd M, Gaubert S, Diedrich S, et al. Surgical trauma and postoperative immune dysfunction. *Eur Surg Res* 2012; **48**: 180-6.
73. Cho JY, Han HS, Yoon YS, Hwang DW, Jung K. Postoperative complications influence prognosis and recurrence patterns in periampullary cancer. *World J Surg* 2013; **37**: 2234-41.
74. Law WL, Choi HK, Lee YM, Ho JWC. The impact of postoperative complications on long-term outcomes following curative resection for colorectal cancer. *Ann Surg Oncol* 2007; **14**: 2559-66.
75. Miki C, Hiro J, Ojima E, Inoue Y, Mohri Y, Kusunoki M. Perioperative allogeneic blood transfusion, the related cytokine response and long-term survival after potentially curative resection of colorectal cancer. *Clin Oncol* 2006; **18**: 60-6.
76. Amato A, Pescatori M. Perioperative blood transfusion and recurrence of colorectal cancer. *Cochrane Database Sys Rev* 2006; **1**. Art. No.: CD005033.
77. Acheson AG, Brookes MJ, Spahn DR. Effects of allogeneic red blood cell transfusions on clinical outcomes in patients undergoing colorectal cancer surgery, a systematic review and meta-analysis. *Ann Surg* 2012; **256**: 235-44.
78. Mynster T, Christensen IJ, Moesgaard F, Nielsen HJ. Effects of the combination of blood transfusion and postoperative infectious complications on prognosis after surgery for colorectal cancer. *Br J Surg* 2000; **87**: 1553-62.