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Performance of photocatalytic oxidation of tetracycline in aqueous solution by TiO₂ nanofibers

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Abstract

The presence of pharmaceutical compounds in water and soil has become an environmental concern. The aim of this study was to evaluate the performance of TiO₂ nanofiber in the oxidation of the antibiotic tetracycline. TiO₂ nanofiber was fabricated by electro-spinning method, and then was calcined at 560°C for 2 h. Central composite design (CCD) statistic model was used to optimize tetracycline concentration, time and pH for TiO₂ catalyst. A tubular Pyrex glass reactor with diameter of 15 cm and height of 30 cm was designed and a 125W Philips HPLN lamp (UV, $\lambda > 254$ nm) was used as light source. Samples were measured by high-performance liquid chromatography (HPLC). Equation of model suggests a direct relationship between pH and time with efficiency of tetracycline removal. The observations indicated that time is the most significant (scaled estimate = + 28.04) parameter in efficiency of tetracycline removal. The application of response surface methodology yielded the equation of Y = 65.82 + 5.74 pH + 28.04 time + (-3.07)(pH)² + (-6.6)(time)², with R² = 0.986 which represents good reliability of model. Based on the response surface plots optimum conditions for degradation of tetracycline with maximum efficiency of around 95% was attained. These conditions are as follow; concentration: 50 mg/l, pH= 8.3, time= 15 min.

Keywords: Antibiotic, Photocatalytic, Response surface methodology, TiO₂ nanofibers, Tetracycline

Introduction

In recent years the use of antibiotics for veterinary and medical purposes has been widely increased (100,000– 200,000 ton/year) and therefore the possibility of water contamination with these compounds has increased [1]. The presence of antibiotics in water and soil can cause some allergies and toxicity [2,3]. Additionally, a few studies on the presence of antibiotic residues in surface water, ground water, sea and drinking water have been reported [4-6]. In addition, the presence of antibiotic residues in wastewater treatment plant effluents and hospital wastewaters have been detected [7-9]. Tetracycline (TC) is used mostly in aquaculture and veterinary medicine. The amount of drug use in animals is higher, for example, in poultry: 20–300 mg per kg of body weight is consumed. Like other antibiotics, tetracycline is discharged into water resources through drug manufacturing wastewater, disposal of non-consumable compounds, expired drugs containing tetracycline and from animal and agricultural wastes [10,11].

Various technologies are used to degrade tetracycline; one group of these technologies is comprised of Advanced Oxidation/Reduction Processes (AO/RPs) which have potential for degradation of contaminated waters by antibiotics. When these technologies operate, they produce radicals that can react and destroy contaminants in the water. AO/RP methods typically generate hydroxyl radicals (OH[•]) which include O_3/UV , O_3/H_2O_2 , TiO₂/UV, ZnO/UV, H_2O_2/UV , and UV/Photo-Fenton [12].

As in recent studies reported, heterogeneous photocatalysis for TC degradation by TiO_2 has been costeffective and environmentally sustainable treatment



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alternative. TiO₂ is one of most suitable photocatalysts for water treatment, because of its high photocatalytic activity, good chemical and biological stability, high energy efficiency, minimum waste production, relatively low cost, and non-toxicity [13]. Several studies have investigated photocatalytic oxidation of the antibiotic tetracycline on TiO₂ particles and TiO₂ coated onto the fixed bed [13-16]. In a report the effect of UV type (UV at 254 and 365 nm and solarium device at 300–400 nm) with TiO₂ nanoparticle on TC degradation was studied [16]. Also, recently a response surface model for predicting the efficiency of TC degradation by TiO2 and ZnO was developed using The Central Composite Circumscribed (CCC) statistic model to optimize the amount of catalyst and the pH of the reaction [15].

In that work, two photocatalitic factors of catalyst and pH were used in the CCC design. In this study, for the first time the effects of three factors were studied by using Central Composite Design (CCD) and TiO_2 nanofiber was used as catalyst TC degradation.

The performance of TiO_2 nanofiber in the oxidation of antibiotic tetracycline has been evaluated. The Response Surface Methodology (RSM) was used to optimize pH, tetracycline concentration and time. The degradation processes were monitored by measuring tetracycline disappearance.

Material and methods

Preparation of precursor solution for electrospinning

Polyvinylpyrrolidone (PVP) (MW=1,300,000 g/mol purchased from Aldrich Co., code: 437190) was dissolved in the mixture of ethanol/acetic acid (2.7:1w:w; Merck Co.) at 60°C for 3 h. Titanium isopropoxide (TTIP) (Aldrich) was added to the PVP solution with vigorous stirring for 6 h. A homogeneous solution including 6.5 wt. % PVP and 10 wt. % TTIP was prepared. The obtained mixed solution was used as the working fluid for electrospinning [17].

Fabrication of TiO₂ nanofibers

The prepared precursor solution (volume: 10 mL) was applied to a droplet of PVP/TTIP precursor solution at the tip of a syringe needle with 0.4 mm inner diameter. The flow rate of the solution was 9 μ L/min controlled by a syringe pump. A voltage of 18 kV was applied between the needle and the aluminum foil, which was placed at 20–25 cm from the syringe tip and used to collect the nanofibers. A schematic diagram of the electrospinning apparatus is shown in Figure 1. After electrospinning, the prepared nanofibers were calcined at 560°C for 2 h. Scanning Electron Microscope (SEM), from Philips Co. (Holland), was used for measuring the fiber diameters and observation of fiber morphologies. TiO₂ nanofibers were characterized by X-ray diffraction (X-ray diffractometer, Philips, PW 1800).

Response surface methodology

The Central Composite Design (CCD) statistic model was used in order to optimize the amount of TC concentration, time and pH of the reaction for TiO_2 catalyst. The optimization procedure for photocatalytic reactions consisted of an experimental design to identify the importance of each variable involved in the reaction with a minimum set of experiments. The variables under study were codified for their higher and lower values (+1, -1), simultaneously changed in a defined range and processed with the adequate statistical software. Multivariate analysis was used to determine the weight and interaction between the variables, obtaining a polynomial and a graphical representation, or response surface, which provides the region where the optimized reaction occurs [17]. In the present study, The

Table 1 The variables and their levels for the centralcomposite experimental design

Independent variable	Coded levels					
	-α	-1	0	+1	+α	
Concentration (mgL ⁻¹)	7.95	25	50	75	92	
рН	2	4	7	10	12	
Time (min)	0	3	7.5	12	15	



variables included concentration of TC, reaction time and pH at five coded levels, $-\alpha$, -1, 0, +1, $+\alpha$ (the number of is 1.68), as shown in Table 1 in which the actual levels of variables for CCD experiments are also given. The CCD included 20 experimental trials with five trials as replication of the central points.

Photocatalytic activity test

A tubular Pyrex glass reactor with diameter of 15 cm and height of 30 cm was used in this study. The reaction was always maintained under cool condition by circulating water (all photocatalytic experiments were performed at ambient temperature). A 125W Philips HPLN lamp (UV, $\lambda \ge 254$ nm) was used for light sources. To avoid the effects of the sunlight, the reactor was covered with aluminum foil. UV light lamp was encapsulated in a quartz jacket and positioned in the middle of the reactor. In a typical experiment, catalyst (TiO₂ nanofiber) was added into TC solution with an initial concentration of (500 mg/L).

Analytical methods

The samples were taken by syringe and filtered through 0.45 μm membranes at pre-selected time intervals, and

Table 2 Experimental design and results of the central composite design

Run	X ₁	X ₂	X ₃	Con. (mg/L)	рН	Time (min)	Efficiency (%)
1	0	0	0	50	7	7.5	65.8
2	1	1	-1	75	10	3	40.0
3	0	0	0	50	7	7.5	65.4
4	1	-1	1	75	4	12	76.6
5	-1	-1	1	25	4	12	75.0
6	0	0	0	50	7	7.5	66.2
7	-1.68	0	0	7.95	7	7.5	59.0
8	0	0	+1.68	50	7	15	95.0
9	1	-1	-1	75	4	3	15.2
10	1	1	1	75	10	12	82.6
11	0	0	0	50	7	7.5	65.8
12	-1	1	1	25	10	12	90.0
13	-1	-1	-1	25	4	3	15.0
14	0	+1.68	0	50	12	7.5	62.5
15	0	0	-1.68	5	7	0	0.0
16	+1.68	0	0	92	7	7.5	58.1
17	0	0	0	50	7	7.5	66.0
18	0	0	0	50	7	7.5	65.6
19	-1	1	-1	25	10	3	30.8
20	0	-1.68	0	50	2	7.5	52.5



then measured using high-performance liquid chromatography (HPLC). HPLC consisted of a Knauer LPG pump, an EZ-chrom HPLC system manager program and a UV detector (k-2500). The UV–detector was set at the maximum absorption wavelength for 365 nm. Aliquots of 100 μ L were injected manually using a model SGE injection valve (SGE. Australia). A column of MZanalysentechnik ODS-3 C18 (4.6 mm×250 mm) packed with 5 μ m spherical particles was used for separation. An acetonitrile/0.01 mol/L aqueous oxalic acid (31:69, v/v) mixture was used as mobile phase at 30°C with a constant flow rate of 1.0 mLmin⁻¹ [18]. The TC retention time in HPLC analysis was 3.83 min.

Results and discussion

Characterizations of synthesized TiO₂ nanofiber

Figure 2a shows a typical SEM image of the synthesized nanofibers. The image was taken from a sample containing 6.5% PVP and 10% TTIP, distance= 20 cm, applied voltage=18 kV and flow rate=0.6 ml/h. This image shows that diameter of nanofiber is between 29–90 nm. Figure 2b shows X-ray diffraction patterns of the

synthesized TiO₂ nanofiber after calcinations at 560°C and 685°C for 2 h under air atmosphere. As shown in Figure 2b the formation of pure anatase TiO₂ phase attained after calcinations at 560°C. Increasing the temperature to 685°C led to formation of rutile TiO₂ as well. Figure 2a displays SEM images of TiO₂ nanofibers calcined at 560°C in an air atmosphere for 2 h.

Optimization of the reaction

Central Composite Design matrix of three variables in coded and natural units with the observed responses is shown in Table 2. A typical response surface function for three input variables is in the form of the following equation:

$$Y = b_0 + b_1 x_1 + b_2 x_2 + b_3 x_3 + b_{12} x_1 x_2 + b_{13} x_1 x_3 + b_{23} x_2 x_3 + b_{11} x_1^2 + b_{22} x_2^2 + b_{33} x_3^2$$
(1)

The parameters included TC concentration, pH and time. The levels of input variables are shown in Table 1.





The efficiency of correlated model was checked by the coefficient of determination (\mathbb{R}^2). In this case, the value of the determination coefficient was indicated as ($\mathbb{R}^2 = 0.986$). The value of the adjusted determination coefficient (Adj. $\mathbb{R}^2 = 0.976$) is a high significance of the model. The application of response surface methodology yielded the Eq. (2), which is an empirical relationship between the efficiency of TC removal and the test variables in coded unit:

$$Y = 65.82 + 5.74 X_2 + 28.04X_3 + (-3.07)X_2^2 + (-6.6)X_3^2$$
(2)

Where Y is the response (efficiency of TC removal) and X_2 , X_3 are the coded values of the test variables that are shown in Table 2. This equation suggests a direct relationship between pH and time with efficiency of TC removal (P < 0.05). These observations indicate that time is the most significant (scaled estimate=+28.04) parameter in the efficiency of TC removal. Except for the linear term X_2 (pH), X_3 (time), (P < 0.05) and quadratic term X_2^2 and X_3^{-2} (p < 0.05), none of the other linear, quadratic and interaction terms were statistically significant.

The quadratic effect of pH indicated that at both high and low pH values, the reaction will be less efficient, where this negative effect is more pronounced in acidic conditions, as shown in the response surface and contour plots (Figure 3). Under acidic pH (<3.3), both

Table 3 Kinetic order degradation of TC at various pH, subjected to UVC/TiO₂ nanofiber reactor

-	_			
C0 (mg/l)	рН	k (1/min)	Reaction order	
75	10	0.2531	First order	
75	7	0.1378	First order	
75	4	0.1568	First order	
50	10	0.2296	First order	
50	7	0.1250	First order	
50	4	0.1423	First order	
25	10	0.2055	First order	
25	7	0.1119	First order	
25	4	0.1273	First order	

tetracycline and Titania are positively charged provoking a repulsive effect. At pH=8.7, TC shows a negative charge increasing the repulsive effect with the Titania surface, which is also partially negatively charged. At higher pH (>9.7) the repulsive effect increases because TC shows a double negative charge [15].

The predicted values of efficiency of TC removal based on the range of time were estimated as contour and response surface plots (Figure 4). In this contour and response surface plot, the effect of two variables on efficiency of TC removal is shown. Based on Response surface plots optimum conditions for degradation of TC with maximum efficiency around 95% was achaived. These conditions are as follows; concentration=50 mg/l, pH=8.3, time=15 min (Figure 5). The optimal pH for degradation of TC 8.7 has been reported by Rodrigo et al. [15].

Degradation of TC in the UVC/TiO₂ nanofiber reactor in different pH_S in all concentrations followed first order kinetics model (Table 3). Most studies previously reported the kinetics of photolysis of pollutants to be first order [14,16].

Conclusion

The application of response surface methodology yielded the equation of Y = 65.82 + 5.74 pH + 28.04 time + (-3.07) (pH)² + (-6.6)(time)² with R² = 0.986 which represents good reliability of model. The effect of various parameters on degradation efficiency of tetracycline was studied via concentration of TC analysis by HPLC. For instance with increasing pH value from 4 to 10, the degradation efficiency of tetracycline increased from 75.8% ± 0.8 to 86.3% ± 3.7 (time = 15 min). Based on Response surface plot, optimum conditions for degradation of TC with maximum efficiency around 95% were attained. These conditions are as follow: concentration= 50 mg/l, pH= 8.3, time= 15 min.

Competing interests

The authors declare that they no competing interests.

Authors' contributions

AJ was the main investigator, collected the data, performed the statistical analysis, and drafted the manuscript. SN and AM supervised the study. AHM, MA, RMA and NR were advisors of the study. All authors read and approved the final manuscript.

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References

- Xu W-H, Zhang G, Zou S-C, Li X-D, Liu Y-C: Determination of selected antibiotics in the Victoria harbour and the pearl river, south china using ighperformance iquid chromatographyeelectrospray ionization tandem mass spectrometry. Environ Pollut 2007, 145:672–679.
- Kummerer K: The presence of pharmaceuticals in the environment due to human use - present knowledge and future challenges. J Environ Manage 2009, 90:2354–2366.
- Rozas O, Contreras D, Mondacab MA, Perez-Moyac M, Mansillaa HD: Experimental design of Fenton and photo-Fenton reactions for the treatment of ampicillin solutions. J Hazard Mater 2010, 177:1025–1030.
- Sacher F, Lange F, Brauch H, Blankenhorn I: Pharmaceuticals in groundwaters analytical methods and results of a monitoring program in Baden-Württemberg. Germany J Chromatogr A 2001, 938(1–2):199–210.
- Sarmah AK, Meyer MT, Boxall BA: A global perspective on the use, sales, exposure pathways, occurrence, fate and effects of veterinary antibiotics (VAs) in the environment. *Chemosphere* 2006, 65:725–759.
- Yiruhan, Wang QJ, Mo CH, Li YW, Gao P, Tai YP, Zhang Y, Ruan ZL, JW X: Determination of four fluoroquinolone antibiotics in tap water in Guangzhou and Macao. Environ Pollut 2010, 158:2350–2358.
- Brown K, Kulis J, Thomson B, Chapman T, Mawhinney D: Occurrence of antibiotics in hospital, residential, and dairy effluent, municipal wastewater, and the Rio Grande in New Mexico. Sci Total Environ 2006, 366(2–3):772–783.
- Homem V, Santos L: Degradation and removal methods of antibiotics from aqueous matrices–a review. J Environ Manage 2011, 92(10):2304–2347.
- Seifrtová M, Pena A, Lino CM PS: Determination of fluoroquinolone antibiotics in hospital and municipal wastewaters in Coimbra by liquid chromatography with a monolithic column and fluorescence detection. *Anal Bioanal Chem* 2008, **391**:799–805.
- 10. Boxall ABA, Kolphin DW, Halling-Sørensen B, Tolls J: Are veterinary medicines causing environmental risks? *Environ Sci Technol* 2003, 37:286–294.
- Mompelat S, LeBot B, Thomas O: Occurrence and fate of pharmaceutical products and by-products, from resource to drinking water. *Environ Int* 2009, 35:803–814.
- Rickman KA, Mezyk SP: Kinetics and mechanisms of sulfate radical oxidation of b-lactam antibiotics in water. Chemosphere 2010, 81:359–365.
- Wang P, Yap P-S, Lim TT: C-N-S tridoped TiO2 for photocatalytic degradation of tetracycline under visible-light irradiation. *Appl Catal Gen* 2011, 399:252–261.
- Addamo M, Augugliaro V, Di Paola A, Gracía-López E, Loddo V, Marcí G, Palmisano L: Removal of drugs in aqueous systems by photoassisted degradation. J Appl Electrochem 2005, 35(7–8):765–774.
- Palominos RA, Mondaca MA, Giraldo A, Penuela G, Perez-Moya M, Mansilla HD: Photocatalytic oxidation of the antibiotic tetracycline on TiO2 and ZnO suspensions. *Catal Today* 2009, 144:100–105.
- Reyes C, Fernandez J, Freer J, Mondaca MA, Zaror C, Malato S, Mansilla HD: Degradation and inactivation of tetracycline by TiO2 photocatalysis. J Photochem Photobiol A: Chemistry 2006, 184:141–146.
- Sarlak N, Nejad MAF, Shakhesi S, Shabani K: Effects of electrospinning parameters on titanium dioxide nanofibers diameter and morphology: an investigation by Box–Wilson central composite design (CCD). *Chem Eng J* 2012, 210:410–416.
- Wang Y, Zhang H, Zhang J, Lu C, Huang Q, Wu J, Liu F: Degradation of tetracycline in aqueous media by ozonation in an internal loop-lift reactor. J Hazard Mater 2011, 192:35–43.

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