

STUDY PROTOCOL

Open Access

Efficacy and safety of active negative pressure peritoneal therapy for reducing the systemic inflammatory response after damage control laparotomy (the Intra-peritoneal Vacuum Trial): study protocol for a randomized controlled trial

Derek J Roberts^{1,2}, Craig N Jenne^{3,8}, Chad G Ball^{1,4,5}, Corina Tiruta⁵, Caroline Léger^{3,8}, Zhengwen Xiao⁵, Peter D Faris^{2,6}, Paul B McBeth⁹, Christopher J Doig^{2,3}, Christine R Skinner³, Stacy G Ruddell³, Paul Kubes^{3,7,8} and Andrew W Kirkpatrick^{1,3,5*}

Abstract

Background: Damage control laparotomy, or abbreviated initial laparotomy followed by temporary abdominal closure (TAC), intensive care unit resuscitation, and planned re-laparotomy, is frequently used to manage intra-abdominal bleeding and contamination among critically ill or injured adults. Animal data suggest that TAC techniques that employ negative pressure to the peritoneal cavity may reduce the systemic inflammatory response and associated organ injury. The primary objective of this study is to determine if use of a TAC dressing that affords active negative pressure peritoneal therapy, the ABThera Open Abdomen Negative Pressure Therapy System, reduces the extent of the systemic inflammatory response after damage control laparotomy for intra-abdominal sepsis or injury as compared to a commonly used TAC method that provides potentially less efficient peritoneal negative pressure, the Barker's vacuum pack.

Methods/Design: The Intra-peritoneal Vacuum Trial will be a single-center, randomized controlled trial. Adults will be intraoperatively allocated to TAC with either the ABThera or Barker's vacuum pack after the decision has been made by the attending surgeon to perform a damage control laparotomy. The study will use variable block size randomization. On study days 1, 2, 3, 7, and 28, blood will be collected. Whenever possible, peritoneal fluid will also be collected at these time points from the patient's abdomen or TAC device. Luminex technology will be used to quantify the concentrations of 65 mediators relevant to the inflammatory response in peritoneal fluid and plasma. The primary endpoint is the difference in the plasma concentration of the pro-inflammatory cytokine IL-6 at 24 and 48 h after TAC dressing application. Secondary endpoints include the differential effects of these dressings on the systemic concentration of other pro-inflammatory cytokines, collective peritoneal and systemic inflammatory mediator profiles, postoperative fluid balance, intra-abdominal pressure, and several patient-important outcomes, including organ dysfunction measures and mortality.

(Continued on next page)

* Correspondence: Andrew.Kirkpatrick@albertahealthservices.ca

¹Department of Surgery, University of Calgary and the Foothills Medical Centre, North Tower 10th Floor, 1403-29th Street Northwest, Calgary, Alberta T2N 2T9, Canada

³Department of Critical Care Medicine, University of Calgary and the Foothills Medical Centre, Ground Floor McCaig Tower, 3134 Hospital Drive Northwest, Calgary, Alberta T2N 5A1, Canada

Full list of author information is available at the end of the article

(Continued from previous page)

Discussion: Results from this study will improve understanding of the effect of active negative pressure peritoneal therapy after damage control laparotomy on the inflammatory response. It will also gather necessary pilot information needed to inform design of a multicenter trial comparing clinical outcomes among patients randomized to TAC with the ABThera *versus* Barker's vacuum pack.

Trial registration: ClinicalTrials.gov identifier NCT01355094

Keywords: Abdominal injuries, Damage control laparotomy, Inflammation, Negative pressure wound therapy, Randomized controlled trial, Sepsis, Temporary abdominal closure, Wounds and injuries

Background

Intra-abdominal sepsis and abdominal trauma constitute major international public health problems [1]. As limited treatments aside from source control and antimicrobial therapy exist for severe intra-abdominal infections, intra-peritoneal sepsis is associated with significant morbidity and mortality [2]. Moreover, trauma affects approximately 700 million people, including 30 million North Americans and 2 million Canadians, worldwide each year [1,3]. These injuries result in 5 million deaths, with blunt and penetrating abdominal trauma constituting a substantial proportion of trauma-related mortality [1,3,4].

Deranged patient physiology as a result of massive hemorrhage and/or contamination from hollow viscus or pancreaticobiliary injuries is an important contributor to poor outcomes after intra-abdominal sepsis or injury [5,6]. In an attempt to limit this deranged physiology, damage control laparotomy, or abbreviated initial laparotomy followed by temporary abdominal closure (TAC) and planned re-operation after intensive care unit (ICU) resuscitation, is increasingly used to manage intra-abdominal bleeding and contamination among these patients [5-10]. The stages of damage control laparotomy broadly include: (1) limited initial operation with temporary control of hemorrhage and contamination; (2) application of a TAC device; (3) ICU resuscitation; and (4) re-operation with attempted completion of definitive surgical repairs after normalization of patient physiology [5,7-9].

A common component of the deranged physiologic response after intra-abdominal sepsis or injury is inflammation. Although intra-abdominal sepsis or injury also results in a systematic inflammatory response, evidence suggests that this response is exaggerated in the peritoneum [11,12]. Microcirculatory disruption after septic or hemorrhagic shock may lead to loss of intestinal barrier function, bowel edema, and formation of pro-inflammatory-mediator-rich ascites [13-18]. This inflammatory ascites may serve as a motor for sepsis, which may perpetuate systemic inflammation and result in multi-organ dysfunction syndrome [11,12,14,18]. In support of this, the levels of select peritoneal cytokines have been reported to be significantly different between animals that survived as compared to those who died in animal models of intra-abdominal sepsis

[19]. However, although a number of mediators have been associated with sepsis or injury, relatively little is known about their temporal peritoneal and systemic expression after intra-abdominal sepsis or injury, including abdominal surgery (Table 1) [16,20-43].

Importantly, animal data suggest that TAC techniques that employ constant negative pressure to the peritoneal cavity may remove inflammatory ascites, reduce the systemic inflammatory response, and improve organ injury and potentially outcomes [10,18]. One preclinical study randomly allocated animals with intra-abdominal sepsis to TAC with vacuum-assisted drainage of the peritoneal cavity after laparotomy and observed significantly reduced levels of the pro-inflammatory cytokines tumor necrosis factor- α (TNF- α) and interleukin (IL)-1 β , -6, and -12 as compared to those who received a TAC that afforded only passive peritoneal drainage [18]. Animals with septic open abdominal wounds treated with negative pressure peritoneal therapy also demonstrated significantly reduced intestinal edema and improved pulmonary, renal, and hepatic histological features; intra-abdominal pressure (IAP); cardiac and renal function; and pulmonary compliance, with a trend towards a lower incidence of acute lung injury [18]. These animal data are interesting as several studies have reported that the pro-inflammatory cytokine IL-6 is associated with an increased risk of organ dysfunction, adverse complications, and/or mortality among trauma victims and patients with sepsis [44-50].

However, very little relevant clinical data yet exist to support that negative pressure peritoneal therapy may improve outcomes among trauma and acute care surgery patients after damage control laparotomy [10]. One systematic review and meta-analysis of largely uncontrolled case series of damage control laparotomy suggested that vacuum-assisted TAC techniques may be associated with improved outcomes among critically ill adults with contaminated open abdominal wounds [51]. However, a systematic review conducted by our group in 2012 found only 11 comparative studies, including 2 randomized controlled trials (RCTs) and 9 cohort studies, examining the efficacy and safety of negative pressure peritoneal therapy *versus* alternate TAC methods among critically ill or injured adults [10]. As only one RCT compared negative pressure

Table 1 Inflammatory mediators associated with intra-abdominal sepsis or injury, including abdominal surgery, among studies of animals or humans

Mediator	Potential role in intra-abdominal sepsis or injury	Reference(s)
CRP	A serum marker of sepsis that increases in concentration in plasma following abdominal surgery	[20-22]
Haptoglobin	Elevated expression in blood leukocytes following severe blunt trauma	[23]
IL-1ra	Elevated in plasma following trauma; elevated expression in blood leukocytes following severe blunt trauma	[23,24]
IL-6	Potent inflammatory mediator and marker of sepsis; elevated levels correlate with length of hospital stay, complications, and mortality among patients with intra-abdominal sepsis; elevated levels in peritoneal fluid in a porcine model of intra-abdominal sepsis; elevated plasma/serum levels following abdominal surgery; elevated plasma levels following severe trauma associate with injury severity, development of organ dysfunction, and poor outcomes, including mortality	[21-23,25-28,44-50]
IL-8	Potent neutrophil chemoattractant; elevated expression in blood leukocytes following severe blunt trauma	[23,29]
IL-10	Elevated serum levels during intra-abdominal sepsis; blocks pro-inflammatory cytokine release; elevated after abdominal surgery	[30-32]
IL-15	Elevated levels in serum correlate with organ dysfunction and poor patient prognosis	[33]
IL-17	Potent pro-inflammatory mediator; promotes neutrophil recruitment to the peritoneal cavity and enhanced bacterial clearance in a mouse model of intra-abdominal sepsis; elevated plasma levels in select patients following severe trauma	[28,34,35]
IL-22	Elevated serum levels during intra-abdominal sepsis	[30]
IL-33	Mediates neutrophil recruitment to peritoneum; promotes bacterial clearance and reduces mortality in a mouse model of intra-abdominal sepsis	[36]
MCP-1 (CCL2)	Potent monocyte chemoattractant; serum levels elevated in a rat model of intra-abdominal sepsis; elevated expression in blood leukocytes following severe blunt trauma	[23,37]
M-CSF	Elevated in plasma following trauma	[24]
MIF	Present early in sepsis and remains elevated for a prolonged time period; significantly higher levels in non-survivors of sepsis compared to survivors; MIF neutralization reduces mortality in a mouse model of intra-abdominal sepsis	[38,39]
PDGF	Elevated in plasma following trauma	[24]
Procalcitonin	Marker of infectious complications following abdominal surgery and negatively associated with survival; elevated after abdominal surgery	[32,40]
TNF- α	Serum levels elevated in a rat model of intra-abdominal sepsis	[37]
tPA	Enhances bacterial clearance, reduces cellular influx, increases plasma and peritoneal IL-12 and IL-10 levels, and reduces lung and liver damage in a mouse model of intra-abdominal sepsis	[16]
TRAIL	Promotes inflammatory cell recruitment to the peritoneum, enhances bacterial clearance, and reduces mortality in a mouse model of intra-abdominal sepsis; modulates apoptosis	[41,42]

CCL2, Chemokine (C-C motif) ligand 2; CRP, C-reactive protein; IL-1ra, Interleukin-1 receptor antagonist; IL-6, Interleukin-6; IL-8, Interleukin-8; IL-10, Interleukin-10; IL-15, Interleukin-15; IL-17, Interleukin-17; IL-22, Interleukin-22; IL-33, Interleukin-33; MCP-1, Monocyte chemoattractant protein-1; M-CSF, Macrophage colony-stimulating factor; MIF, Macrophage migration inhibitory factor; PDGF, Platelet-derived growth factor; TNF- α , Tumor necrosis factor-alpha; tPA, Tissue plasminogen activator; TRAIL, Tumor necrosis factor-related apoptosis-inducing ligand.

peritoneal therapy with an alternate type of TAC technique [52], and all studies were associated with at least a moderate risk of bias and significant clinical heterogeneity, we concluded that there was insufficient evidence to support the preferential use of negative pressure peritoneal therapy after damage control laparotomy [10].

The primary objective of this study is to determine if use of a TAC dressing that may afford active negative pressure peritoneal therapy, the ABThera™ Open Abdomen Negative Pressure Therapy System (Kinetic Concepts Inc. (KCI), San Antonio, TX, USA) [10,53], reduces the extent of the systemic inflammatory response after damage control laparotomy for intra-abdominal sepsis or injury as

compared to a commonly used TAC method that provides potentially less efficient peritoneal negative pressure, the Barker's vacuum pack [54-56].

Methods/Design

Overview

The Intra-peritoneal Vacuum Trial is a single-center RCT, which will intraoperatively allocate critically ill or injured adults to TAC with either the ABThera™ or Barker's vacuum pack after damage control laparotomy for intra-abdominal sepsis or injury. Although the trial in itself may not be sufficient to establish superiority of the ABThera™ over the Barker's vacuum pack, we hypothesize superiority,

and will examine for evidence of this in hypothesis tests. The CONSORT flow diagram [57] and an overview of the study design are presented in Figures 1 and 2, respectively.

Intervention and comparator choice rationale

The ABThera™ was chosen as the intervention as its design may allow for active, and potentially equally distributed vacuum pressures throughout the peritoneal cavity and across the viscera [58], theoretically enabling effective removal of pro-inflammatory-mediator-rich intra-peritoneal fluid. The Barker's vacuum pack was chosen as the comparator as it is a commonly used TAC technique, which has been recommended by the Eastern Association for the Surgery of Trauma as the current standard by which to measure other devices [59].

Setting

The trial will be set at the Foothills Medical Centre (FMC) in Calgary, Alberta, Canada. The FMC is a University of Calgary-affiliated, adult, tertiary care, level one regional trauma center that affords trauma and emergency general surgical services for southern Alberta, southwest British Columbia, and southeast Saskatchewan. Postoperative critically ill or injured adults at the FMC are cared

for in a 30-bed, closed, medical/surgical/neurosurgical ICU staffed by fellowship-trained intensivists.

Patient recruitment

Recruitment into the trial will occur in the operating room once the decision has been made by the attending trauma or general surgeon to perform a damage control laparotomy and TAC. We will define an open abdomen as that requiring a TAC due to the abdominal skin and fascia not being closed after laparotomy.

Although no objective, evidence-based indications for damage control laparotomy or open abdominal management exist [60,61], the Eastern Association for the Surgery of Trauma suggest that this technique may be considered in cases of severe abdominal trauma involving hepatic, non-hepatic, or vascular injuries with intra-abdominal packing; among trauma patients with acidosis ($\text{pH} \leq 7.2$), hypothermia (temperature $\leq 35^\circ\text{C}$), and clinical coagulopathy or if the patient is receiving massive transfusion; and in patients with severe intra-abdominal infection/peritonitis or necrotizing pancreatitis [59]. Other indications for the procedure suggested in 2005 by a multidisciplinary expert advisory panel on management of the open abdomen included bowel edema and a significant risk of developing abdominal compartment syndrome [62].

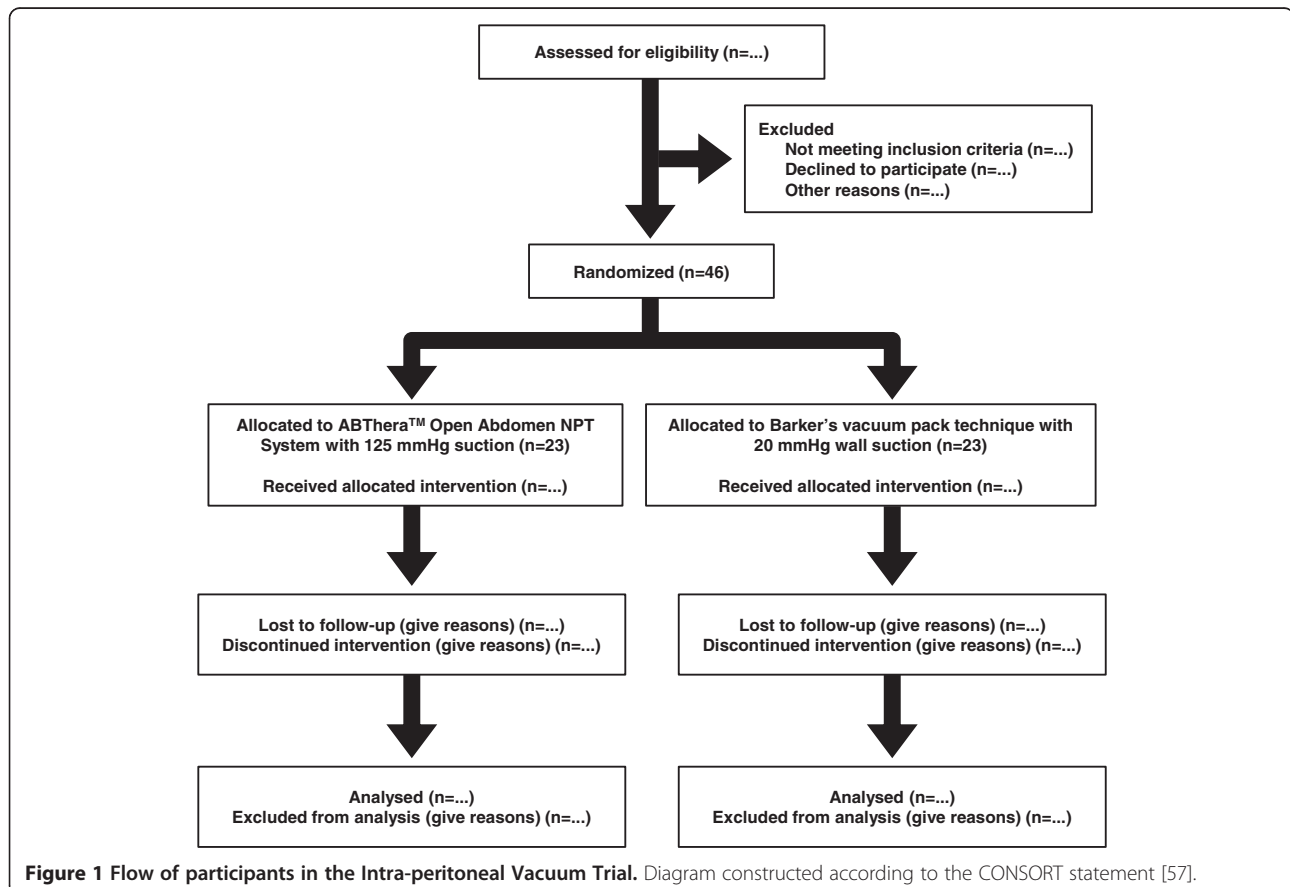
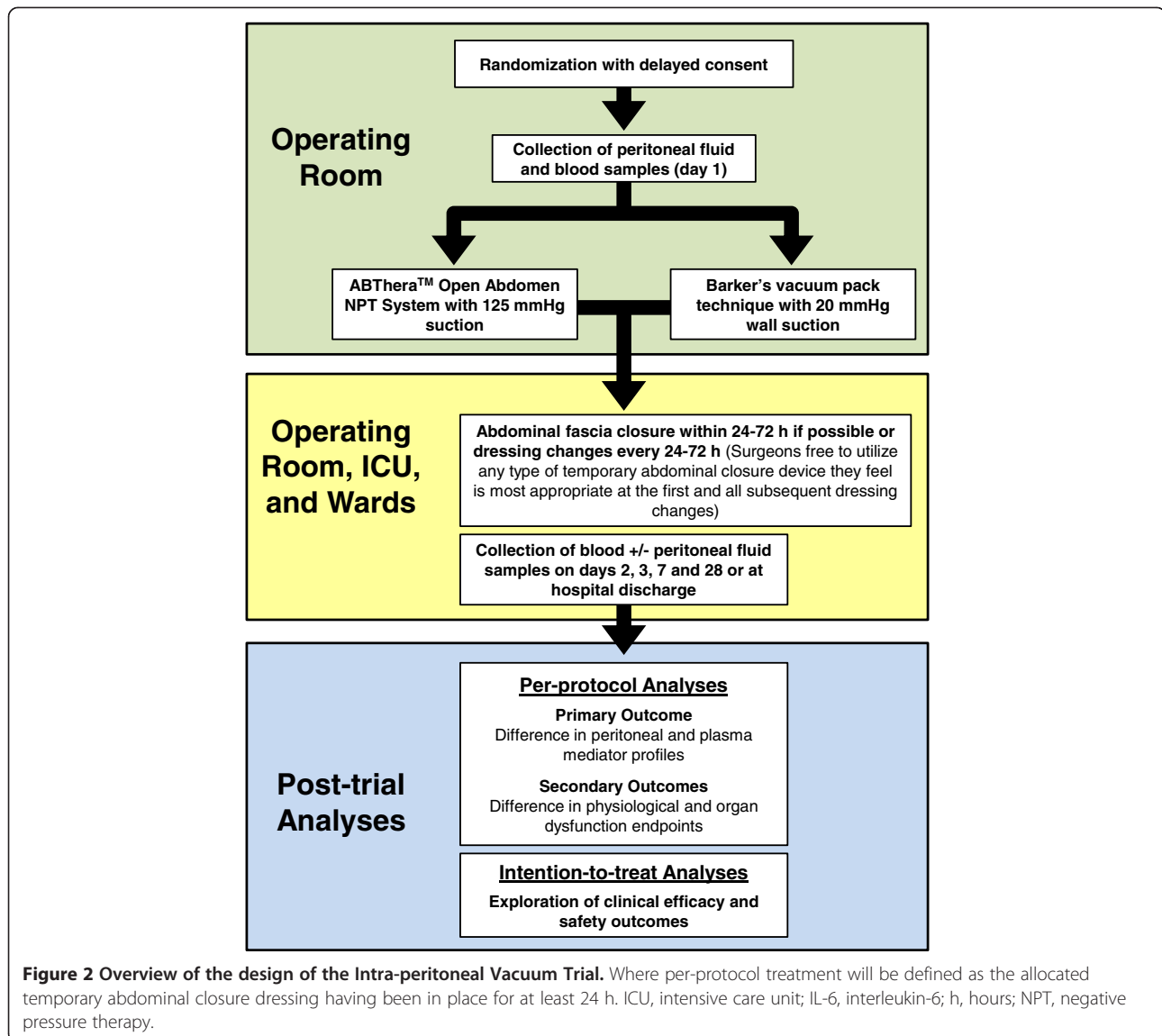


Figure 1 Flow of participants in the Intra-peritoneal Vacuum Trial. Diagram constructed according to the CONSORT statement [57].



Inclusion criteria

- Age ≥ 18 years
- Decision made by the attending trauma or general surgeon to perform a damage control laparotomy with application of a TAC device during the index laparotomy
- Requirement for postoperative ICU admission

Exclusion criteria

- Age < 18 years
- Decision made to perform a definitive or single-stage laparotomy with closure of the abdominal fascia during the index laparotomy
- Administration of intraperitoneal chemotherapy for treatment of intra-abdominal malignancies
- Pregnancy

Eligible, non-randomized patients

Dedicated research coordinators will screen all ICU admissions daily and identify and record patients admitted with an open abdominal wound. When potentially eligible patients are identified that were not recruited, the principal investigator, a trauma and acute care surgeon (AWK), will be informed such that he can ascertain and record reason(s) why these patients were not randomized. These reasons and the list of eligible, non-randomized patients will be used to populate the CONSORT flow diagram and estimate trial feasibility (see below).

Patient randomization

To preserve allocation concealment, randomization will be performed using a random treatment generator hosted on a dedicated, publicly available trial website (<http://peritonealvac.com/>). When an eligible patient is identified

by the attending surgeon, this website will be accessed by a member of the operating room team and the patient's last name and hospital number will be entered. The random treatment allocation will then be announced such that the attending surgeon can perform the indicated method of TAC. Variable block size randomization will be performed to ensure equal numbers of patients in each treatment group. Day 1 is considered the day of randomization.

Consent procedures

We were granted a waiver of immediate consent for patient enrollment in the trial because: (1) all of the eligible patients will be intubated, critically ill or injured adults under general anesthesia at the time the decision to perform a damage control laparotomy and TAC is made; (2) it will be impractical for the attending surgeon or investigators to contact surrogate decision-makers as the decision to perform open abdominal management is frequently made intraoperatively, and many trauma patients arrive unidentified (as an 'unknown male/female'); (3) the TAC dressing must be placed emergently after the decision is made to leave the abdomen open [59]; (4) the ABThera™ and Barker's vacuum pack technique are commonly utilized for TAC at the FMC and across North America [10]; and (5) a recent systematic review by our group found insufficient evidence to suggest that the ABThera™ was superior to the vacuum pack for TAC among critically ill or injured adults requiring damage control laparotomy [10]. Trained surgical investigators will therefore approach recruited patients after their recovery to obtain delayed, written, informed consent to utilize their data for the study. These investigators will provide a written information sheet and explain the study procedures and that continued enrollment in the trial is voluntary. One copy of the consent form will be given to the patient, one will be kept in their chart, and one will be filed as trial documentation.

Study interventions overview

The first application of the ABThera™ or Barker's vacuum pack will be performed in the operating room by the attending trauma or general surgeon while the patient is under general anesthesia.

ABThera™ application

The ABThera™ (Figure 3) will be applied according to manufacturer's recommendations [53]. The visceral protective layer will first be folded or cut to size to ensure full coverage of all viscera. It will then be gently inserted into the abdominal cavity, beneath the peritoneal lining of the abdominal wall, ensuring to work the dressing down evenly into both pericolic gutters. The more superficial perforated foam layer will then be torn to size and

inserted into the open abdominal wound directly over the visceral protective layer. The adjacent skin will subsequently be sponge-dried, after which the open abdomen drape will be placed adhesive-side down onto the foam and intact skin, covering at least an 8- to 10-cm border of periwound tissue. A 2.5-cm hole will then be cut through the drape, after which the interface pad will be applied. The tubing set will subsequently be connected between the interface pad and the canister located in the negative pressure source. Finally, active negative pressure peritoneal therapy will be initiated at 125 mmHg of negative suction.

Barker's vacuum pack application

The Barker's vacuum pack (Figure 4) will be applied according to our local institutional guideline and previously published descriptions [54-56]. At the completion of abdominal operation, a polyurethane sheet will be opened and may be perforated several times with a scalpel or surgical scissors. This sheet will then be placed evenly over the viscera, beneath the peritoneal cavity of the abdominal wall, extending deep into the pericolic gutters. Moistened surgical towels will subsequently be placed over the polyurethane sheet. Two closed-suction, 10-French, flat, silicone, Jackson-Pratt drains will then be placed over the moistened surgical towels. Thereafter, their drainage tubing will be tunneled approximately 3 to 5 cm below the skin in order to exit away from the laparotomy wound. The adjacent skin will subsequently be sponge-dried and covered by a large Opsite™ (Smith & Nephew Inc., St. Petersburg, FL, USA) adhesive film dressing. Finally, the drainage tubing from the two Jackson-Pratt drains will be connected to closed drain bulb suction. The Jackson-Pratt drainage tubing will be removed from these drainage bulbs after the patient arrives in the ICU, and connected to 20 mmHg of negative wall suction.

Reoperation and TAC dressing changes

Although the allocated TAC dressings will be left in place for a duration determined by the attending surgeon, local FMC and international guidelines suggest re-operation with attempts at abdominal fascia closure between 24 and 72 h after initial application [53,59,62]. If abdominal fascial closure is not safe or possible at this time, then a TAC dressing change is recommended [53,62]. Surgeons will be free to utilize any TAC device that they feel is most appropriate at the time of the first and all subsequent dressing changes.

Data collection

Peritoneal fluid and blood

On study day 1, just before application of the allocated TAC dressing, 4 mL of peritoneal fluid and 16 mL of blood will be collected. Peritoneal fluid will be taken directly from the peritoneal cavity and then transferred

ABThera™ Open Abdomen NPT System

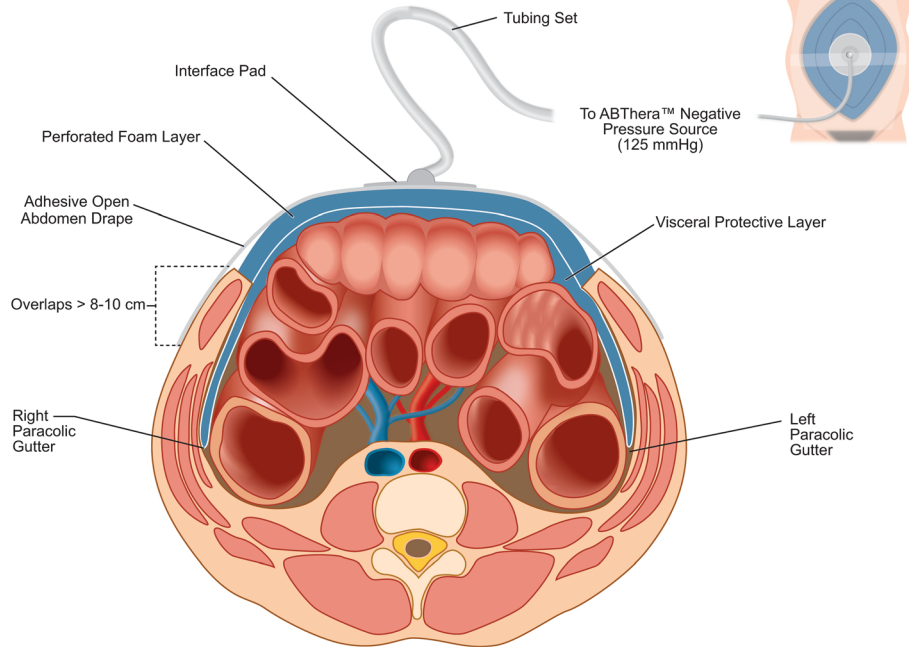


Figure 3 The ABThera™ Open Abdomen NPT System. NPT, Negative pressure therapy.

Barker's Vacuum Pack

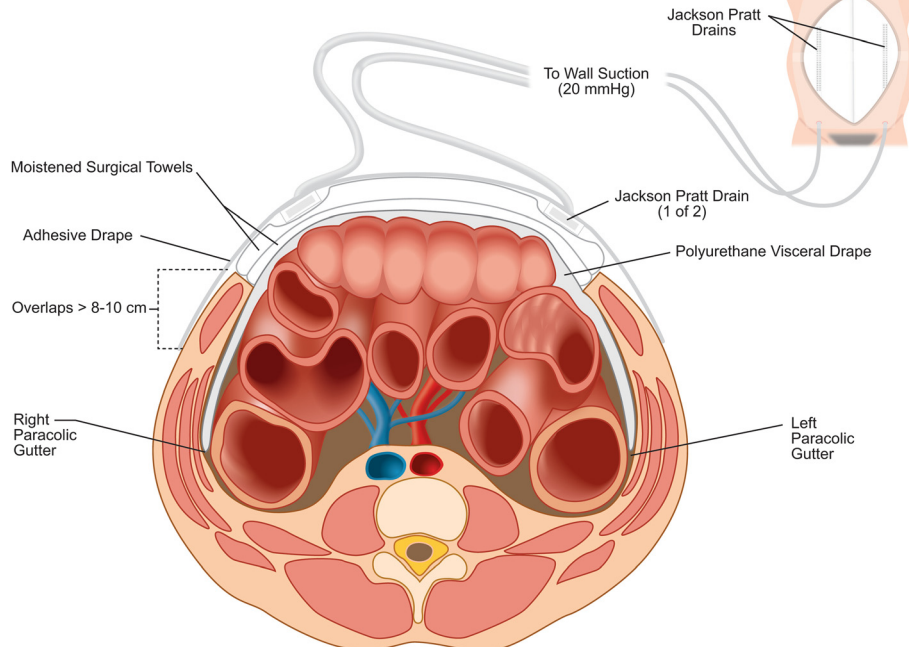


Figure 4 The Barker's vacuum pack.

into sodium-heparin-containing vacutainer tubes (Benton Dickinson Biosciences, Oakville, ON, Canada). Blood will be collected from the patient's indwelling intravascular catheter or via venipuncture into sodium-heparin containing and clot-activating (serum collection) vacutainer tubes (Benton Dickson Biosciences, Oakville, ON, Canada). Sixteen mL of blood will also be drawn on study days 2, 3, and 7, as well as on study day 28 or at hospital discharge (whichever comes first). Whenever possible, 4 mL of peritoneal fluid will also be removed at these times from the tubing set or Jackson-Pratt drainage tubing among patients fitted with an ABThera™ or Barker's vacuum pack, respectively. Immediately after collection, blood and peritoneal fluid samples will be gently inverted several times and then immediately spun at 1,200 g for 15 min at 4°C in a swinging bucket centrifuge to remove cellular debris. The cell-free supernatants will subsequently be collected, aliquoted into labeled cryopreservation tubes, and frozen at -80°C until laboratory analyses can be performed.

Clinical data

Clinical data will be prospectively recorded at the time of enrollment and until study day 28, hospital discharge, or death. We will collect data on baseline patient demographics; past history; indication for damage control laparotomy; injury or illness severity scores (Acute Physiology and Chronic Health Evaluation-II [APACHE-II] [63]; Sequential Organ Failure Assessment [SOFA] [64], and Injury Severity Score [ISS] [65,66]); fluid balance; need for renal replacement therapy and the Risk, Injury, Failure, Loss, and End-stage-kidney-disease (ESKD) (RIFLE) criteria for acute renal dysfunction [67]; and patient physiology (arterial pH, base deficit, and lactate levels; oxygenation indices and partial pressure of arterial oxygen [PaO₂]/fraction of inspired oxygen [FIO₂] ratios; and gastric residual volumes). IAP will also be measured via an indwelling bladder catheter as recommended by the World Society of the Abdominal Compartment Syndrome (WSACS) [68] every 6 h while the abdomen is open and every 12 h once it has been closed.

Laboratory analyses

We will quantify the peritoneal and plasma concentrations of 65 different mediators relevant to the inflammatory response (Table 2). This broad-based approach will be taken in order to minimize bias with regards to which mediators are measured and to gain a better understanding of which inflammatory pathways are activated after damage control laparotomy and intra-abdominal sepsis or injury.

The levels of the mediators listed in Table 2 will be determined using Luminex® technology (EMD Millipore) by an investigator blinded to TAC dressing allocation

status. All samples will be screened using a unique combination of the following multiplex kits: Bio-Plex Pro™ Human Cytokine 21-plex Assay, Bio-Plex Pro™ Human Cytokine 27-plex Assay, and Bio-Plex Pro™ Human Acute Phase 5 + 4-plex Panel Complete (Bio-Rad Laboratories). In addition, a Bio-Plex Pro™ Human Custom 8-Plex (IL-17F, -21, -22, -23, -25, -31, -33, and sCD-40L) (Bio-Rad Laboratories) will also be used to provide broadened coverage of Th17 cytokines.

Luminex® combines the sensitivity, specificity, and reproducibility of the Enzyme-Linked ImmunoSorbent Assay (ELISA) with the multiplexing capacity of fluorescent bead immunoassays, thus allowing multiple biomarkers to be measured simultaneously. Using this approach, we will be able to measure, in a cost- and time-efficient manner, a host of analytes using <200 µL of sample (as compared to an ELISA-based approach, which would require >5 mL of sample), while minimizing inter-assay variability. This technology, although new, has been thoroughly tested and validated using human samples [69]. The biomarkers listed in Table 2 will be grouped into four panels based on typical sample dilution and assay buffer compatibilities. Standard curves will then be generated and the data analyzed using Bio-Plex Manager Software version 6.0 (Bio-Rad Laboratories).

Study endpoints

Primary endpoints

As the pro-inflammatory cytokine IL-6 is consistently upregulated following intra-abdominal sepsis or injury and has been reported to correlate with outcomes (Table 2), the primary endpoint is the difference in the plasma concentration of IL-6 at 24 and 48 h after dressing application between patients randomized to TAC with the ABThera™ *versus* Barker's vacuum pack after damage control laparotomy.

Secondary endpoints and exploration of clinical efficacy and safety

Secondary endpoints of interest include trial feasibility (number of patients enrolled/number of eligible candidates) and the differential effects of the allocated TAC dressings on several physiological and organ dysfunction outcomes, including: (1) the plasma concentration of TNF-α and IL-1, -8, -10, and -12 at 24 and 48 h; (2) the collective peritoneal and systemic mediator profiles (see below); (3) the activation potential of peritoneal fluid [70,71]; (4) peritoneal fluid drainage volume; (5) postoperative daily fluid balance; (6) SOFA scores and individual organ system components of these scores; (7) the PaO₂/FIO₂ ratio; (8) vasopressor requirements and arterial lactate levels; (9) need for renal replacement therapy; (10) RIFLE criteria; (11) APACHE-II scores; and (12) 24 h

Table 2 Groups of mediators to be measured using Luminex® bead-based multiplexing technology

Bio-Plex Pro™ Human Cytokine 21-plex Assay (Bio-Rad Laboratories)	Bio-Plex Pro™ Human Cytokine 27-plex Assay (Bio-Rad Laboratories)	Bio-Plex Pro™ Human Acute Phase 5- + 4-Plex Panel Complete (Bio-Rad Laboratories)	Bio-Plex Pro™ Human Custom 8-Plex (Bio-Rad Laboratories)
IL-1α	IL-1β	Ferritin	IL-17F
IL-2Ra	IL-1ra	Fibrinogen	IL-21
IL-3	IL-2	Procalcitonin	IL-22
IL-12 (p40)	IL-4	Serum amyloid A	IL-23
IL-16	IL-5	tPA	IL-25
IL-18	IL-6	α ₂ -macroglobulin	IL-31
CTACK (CCL27)	IL-7	CRP	IL-33
GRO-α (CXCL1)	IL-8	Haptoglobin	sCD-40L
HGF	IL-9	Serum amyloid P	
IFN-α2	IL-10		
LIF	IL-12 (p70)		
MCP-3 (CCL7)	IL-13		
M-CSF	IL-15		
MIF	IL-17		
MIG (CXCL9)	Basic FGF		
β-NGF	Eotaxin (CCL11)		
SCF	G-CSF		
SCGF-β	GM-CSF		
SDF-1α	IFN-γ		
TNF-β	IP-10 (CXCL10)		
TRAIL	MCP-1 (CCL2)		
	MIP-1α (CCL3)		
	MIP-1β (CCL4)		
	PDGF-BB		
	RANTES (CCL5)		
	TNF-α		
	VEGF		

Basic FGF, Basic fibroblast growth factor; β-NGF, Beta-nerve growth factor; CCL2, Chemokine (C-C motif) ligand 2; CCL3, Chemokine (C-C motif) ligand 3; CCL4, Chemokine (C-C motif) ligand 4; CCL7, Chemokine (C-C motif) ligand 7; CCL11, Chemokine (C-C motif) ligand 11; CCL27, Chemokine (C-C motif) ligand 27; CRP, C-reactive protein; CTACK, Cutaneous T-cell-attracting chemokine; CXCL1, Chemokine (C-X-C motif) ligand 1; CXCL9, Chemokine (C-X-C motif) ligand 9; CXCL10, Chemokine (C-X-C motif) ligand 10; eotaxin, Eosinophil chemotactic protein; G-CSF, Granulocyte colony-stimulating factor; GM-CSF, Granulocyte macrophage colony-stimulating factor; GRO-α, Growth-related oncogene-alpha; HGF, Hepatocyte growth factor; IFN-α2, Interferon alpha-2; IFN-γ, Interferon-gamma; IL-1α, Interleukin-1-alpha; IL-1β, Interleukin-1-beta; IL-1ra, Interleukin-1 receptor antagonist; IL-2, Interleukin-2; IL-2Ra, Interleukin-2 receptor-alpha; IL-3, Interleukin-3; IL-4, Interleukin-4; IL-5, Interleukin-5; IL-6, Interleukin-6; IL-7, Interleukin-7; IL-8, Interleukin-8; IL-9, Interleukin-9; IL-10, Interleukin-10; IL-12 (p40), Interleukin-12 beta subunit; IL-12 (p70), Interleukin-12 (active heterodimer of IL-12 (p40) and IL-12 (p35)); IL-13, Interleukin-13; IL-15, Interleukin-15; IL-16, Interleukin-16; IL-17, Interleukin-17; IL-17F, Interleukin-17F; IL-18, Interleukin-18; IL-21, Interleukin-21; IL-22, Interleukin-22; IL-23, Interleukin-23; IL-25, Interleukin-25; IL-31, Interleukin-31; IL-33, Interleukin-33; IP-10, Interferon gamma-induced protein 10; LIF, Leukemia inhibitory factor; M-CSF, Macrophage colony-stimulating factor; MCP-1, Monocyte chemoattractant protein-1; MCP-3, Monocyte-specific chemokine 3; MIF, Macrophage migration inhibitory factor; MIG, Monokine induced by gamma interferon; MIP-1α, Macrophage inflammatory protein-1-alpha; MIP-1β, Macrophage inflammatory protein-1-beta; PDGF-BB, Platelet-derived growth factor-B homodimer; RANTES, Regulated and normal T cell expressed and secreted; sCD-40L, Soluble cluster of differentiation 40 ligand; SCF, Stem cell factor; SCGF-β, Stem cell growth factor-beta; SDF-1α, Stromal cell-derived factor-1; TNF-α, Tumor necrosis factor-alpha; TNF-β, Tumor necrosis factor-beta; tPA, Tissue plasminogen activator; TRAIL, Tumor necrosis factor-related apoptosis-inducing ligand; VEGF, Vascular endothelial growth factor.

enteral tolerance (if no gastrointestinal anastomosis was performed).

Secondary clinical efficacy and safety endpoints of interest include: (1) in-hospital death; (2) fascial closure rate and days with fascial closure for the month after admission to hospital; (3) ventilator-free days for the month

after admission to hospital; (4) ICU-free days from the month after admission to hospital; (5) hospital-free days from the month after admission to hospital; and (6) risk of renal replacement therapy and days free of renal replacement therapy from the month after admission to hospital. Further, as some have suggested that negative

pressure peritoneal therapy may be associated with an increased risk of abdominal fistula formation or development of intra-abdominal hypertension/recurrent abdominal compartment syndrome [72-74], additional safety outcomes include: (1) intestinal and enteroatmospheric fistula formation; (2) daily IAP; (3) daily WSACS intra-abdominal hypertension grading classification; and (4) risk of abdominal compartment syndrome development.

Sample size

No clinical data yet exist on the effect of negative pressure peritoneal therapy on the inflammatory response after damage control laparotomy for intra-abdominal sepsis or injury. Thus, estimates are not available to allow accurate sample size estimation. We therefore propose to undertake a study among a convenience sample of 46 adults. As there are approximately 40 to 50 patients who undergo damage control laparotomy and open abdominal management per year at our institution, this trial should be able to be completed within a 2-year time period.

Statistical analyses

All continuous variables will be summarized using histograms and measures of central tendency to determine their underlying distribution before statistical descriptions or analyses are conducted. Means (with standard deviations) and medians (with interquartile ranges) will be used to summarize normal and skewed data, respectively. Non-correlated means and medians will be compared using t-tests and Wilcoxon rank sum tests while dependent means and medians will be compared using paired t-tests and Wilcoxon signed rank tests. Differences in proportions will be compared using Fisher's exact test and risk ratios with associated 95% confidence intervals.

For the analysis of the primary endpoint, we will use a mixed-effects repeated measures linear model to compare the effects of the ABThera™ and Barker's vacuum pack on changes in plasma IL-6 concentration at 24 and 48 h after TAC dressing application [75,76]. For longitudinal studies, mixed-effects models have previously been shown to be superior to conventional last-observation-carried-forward approaches for handling missed observations during follow-up [77,78]. This approach will also be used to compare the plasma concentrations of TNF- α and IL-1, -8, -10, and -12 between groups at 24 and 48 h in the analysis of secondary endpoints.

Multidimensional analysis will be used to assess whether the allocated TAC dressing has an effect on the collective peritoneal and plasma mediator profiles of the study patients. Principal components analysis will be used to reduce the mediator concentration variables into a smaller set of components that account for most of the variability in the data. Component scores will subsequently be calculated along the reduced number dimensions, after which

we will attempt to distinguish between patients treated with the ABThera™ and Barker's vacuum pack using discriminant or latent class analysis. A similar approach will be performed in order to relate inflammatory mediator concentrations to outcomes.

Results will be stratified according to the primary indication for damage control laparotomy (intra-abdominal sepsis or abdominal trauma). While clinical efficacy and safety data will be analyzed according to intention-to-treat methods, mediator data will be examined using per-protocol techniques, with per-protocol treatment being defined as the allocated TAC dressing having been in place for at least 24 h. As the number of conducted statistical tests will be large, we will use the false discovery rate (FDR) procedure developed by Benjamini and Hochberg to restrict the proportion of incorrectly rejected null hypotheses to 0.05 [79-81]. All tests will be two-sided, and only those with an FDR-corrected *P* value (that is *q*-value) <0.05 will be considered statistically significant [79]. Stata version 12.0 (Stata Corp., College Station, TX, USA) will be used for all analyses.

Ethics approval, trial registration, and role of the sponsor

The study protocol has been approved by the Conjoint Health Research Ethics Board (CHREB) at the University of Calgary, and is registered online at ClinicalTrials.gov (identifier NCT01355094). Costs of the study will be covered by an investigator-initiated trial funding agreement between the principal investigator (AWK), KCI USA (the manufacturer of the ABThera™ Open Abdomen NPT System), and the Governors of the University of Calgary (KCI contract number: KCI Clinical/UniversityCalgaryAlbertaHealth/082611-000/7). KCI USA has had no role in the design or conduct of the study and will have no role in the collection, management, analysis, or interpretation of the data or preparation, review, or approval of the final manuscript.

Discussion

This will be the first study to compare the systemic inflammatory response among critically ill or injured adults fitted with a TAC that may provide active negative pressure peritoneal therapy with a commonly used TAC technique that provides potentially less efficient peritoneal negative pressure, the Barker's vacuum pack [10]. This study aims to afford a robust prospective description of the peritoneal and systemic inflammatory response after damage control laparotomy for intra-abdominal sepsis or injury. A secondary aim is to gather necessary pilot information related to trial feasibility, intervention safety, and design deficiencies needed to inform construction of a multicenter RCT comparing clinical outcomes among patients fitted with the ABThera™ *versus* Barker's vacuum pack.

In this trial, study patients in both treatment arms will undergo planned re-operation with attempts at abdominal

fascial closure approximately 24 to 72 h after damage control laparotomy. Although one RCT (the RELAP trial) reported that planned re-laparotomy after an initial emergency laparotomy for secondary peritonitis resulted in increased healthcare utilization and costs as compared to re-laparotomy on demand [82,83], these findings likely cannot be generalized to our study's source population. In the RELAP trial, the investigators excluded patients that were managed with temporary operative techniques such as intra-abdominal gauze packing and stapled intestinal resections without reanastomosis [82]. Moreover, 89% of the patients allocated to planned re-laparotomy received intraoperative primary abdominal fascial closure [82]. Thus, very few, if any, of the patients in the planned re-laparotomy group likely received damage control laparotomy.

Several difficulties were encountered in designing the Intra-peritoneal Vacuum trial. The trial is demanding for study investigators as damage control laparotomy must be performed emergently and is often done at night in our center when research coordinators and laboratory members are unavailable. For these reasons, the randomization interface and enrollment process were simplified in order to prevent delays in application of the TAC device, which could alter patient outcomes. Further, as no objective, evidence-based consensus indications for damage control laparotomy or open abdominal management have yet been described, we were forced to utilize a pragmatic study design, which attempts to mimic actual surgical practice. However, in order to increase understanding, we planned *a priori* to conduct a nested prospective cohort study that will explore indications used for open abdominal management among attending trauma and general surgeons at our center.

As compared to previous translational human studies of physical therapies to reduce peritoneal and systemic inflammation among critically ill adults, this trial will utilize novel laboratory analyses and rigorous statistical techniques. In order to avoid bias with regards to which mediators are measured, we will use the validated Luminex® multiplexing technology to assay a host of pro-inflammatory mediators simultaneously in a cost- and time-efficient manner, maximizing use of the obtained patient samples [69]. Further, although many authors have conducted comparisons of cytokine concentrations with study endpoints, they frequently ignore the correlation between mediators measured within individual patients over time. Thus, when examining the relationship between cytokines and TAC method, we will use validated statistical techniques that estimate and incorporate this correlation, and which appropriately handle missed observations during follow-up. Finally, in order to collectively analyze the difference in multiple measured mediators between TAC treatment groups, we will also use multidimensional analyses, including principal components analysis.

This trial has limitations. Although we will be combining the inflammatory mediator concentrations and outcome data for patients with both intra-abdominal sepsis and injury, which may be different, we will stratify our analyses to examine if differences exist between patient diagnoses. Moreover, as the primary objective of this trial is to explore the influence of active abdominal therapy on the systemic inflammatory response among critically ill or injured adults, we will principally use a per-protocol method of analysis. As this method of analysis may not be appropriate or as clinically useful for the exploration of safety and efficacy, we will also utilize an intention-to-treat method of analysis when comparing these endpoints. Importantly, however, as we were unable to estimate the required sample size, we may be underpowered to detect differences in the plasma pro-inflammatory mediator concentrations (or clinical and safety endpoints) among patients randomized to TAC with the ABThera™ *versus* Barker's vacuum pack.

In conclusion, the Intra-peritoneal Vacuum Trial will be a single-center RCT. Results from this study will lead to an improved understanding of the effect of active negative pressure peritoneal therapy on the systemic inflammatory response to intra-abdominal sepsis or injury after damage control laparotomy. The study will also gather pilot information needed to inform creation of a future multicenter RCT comparing clinical outcomes among those undergoing TAC with the ABThera™ *versus* the Barker's vacuum pack after damage control laparotomy.

Trial status

The first patient was enrolled into the Intra-peritoneal Vacuum Trial on 29 September 2011, and we are still actively recruiting patients. At the time of writing this manuscript, 37 patients had been enrolled into the trial, the majority of which have already given delayed informed consent for use of their data. Final results of the study are expected to be prepared for publication near the conclusion of 2013.

Abbreviations

APACHE-II: Acute physiology and chronic health evaluation-II; ELISA: Enzyme-linked ImmunoSorbent assay; ESKD: End stage kidney disease; FIO₂: Fraction of inspired oxygen; FMC: Foothills medical center; IAP: Intra-abdominal pressure; KCI: Kinetic Concepts Inc; ICU: Intensive care unit; ISS: Injury severity score; PaO₂: Partial pressure of arterial oxygen; RCT: Randomized controlled trial; RIFLE: Risk injury, failure, loss, and end-stage-kidney-disease; SOFA: Sequential organ failure assessment; TAC: Temporary abdominal closure; WSACS: World Society of the Abdominal Compartment Syndrome.

Competing interests

AWK has an investigator-initiated trial funding agreement with Kinetic Concepts Incorporated (KCI) USA and the Governors of the University of Calgary for the clinical and laboratory costs of conducting the study (KCI contract number: KCI Clinical/UniversityCalgaryAlbertaHealth/082611-000/7). KCI USA will have no role in the design or conduct of the study; collection, management, analysis, or interpretation of the data; or preparation, review, or approval of the manuscript. The remaining authors have no competing interests to declare.

Authors' contributions

AWK conceived the study. CGB, PBM, CT, CJD, and AWK designed the inclusion/exclusion criteria, specified the interventions, developed the randomization scheme, and submitted the protocol for ethics approval. DJR, CNJ, CT, CJD, CS, SGR, PK, and AWK developed and refined the sample collection procedures. CNJ, CL, PK, and AWK developed the laboratory analyses plan and all authors selected the mediators for measurement. CT and JX developed the trial website while CT developed the online randomization tool. All authors selected the primary and secondary endpoints of interest. DJR and PDF designed the statistical analysis plan. DJR, CNJ, CGB, CT, JX, CJD, CS, SGR, PK, and AWK assist in patient enrollment and revised protocol drafts and data collection forms. DJR, CNJ, CT, JX, CS, SGR, and AWK provide trial oversight on trial coordination and implementation. DJR and CNJ drafted the protocol for publication. All authors afforded critical input on the manuscript and saw and approved the final version before submission for publication.

Authors' information

DJR is a surgery and Clinician Investigator Program resident who is presently conducting a Doctor of Philosophy in epidemiology with a thesis on damage control surgery at the University of Calgary. CGB and AWK are academic trauma and acute care surgeons while AWK is also an intensivist at the Foothills Medical Center. AWK is also the past President of the Trauma Association of Canada and a member of the Executive Committee of the World Society of the Abdominal Compartment Syndrome. PDF is a biostatistician with experience in randomized trial design and analysis of complex correlated data sets. CNJ, CL, and PK are basic scientists with an interest in inflammation and infection while PK is the Director of the Calvin, Phoebe and Joan Snyder Institute of Chronic Diseases, which is renowned for its study of sepsis and infection. PK is also the founding member of the Alberta Sepsis Network, an Alberta Innovates - Health Solutions-funded team grant focusing on the development of novel science and technology, including the conduct of randomized controlled trials, for the understanding and treatment of sepsis.

Acknowledgments

DJR is supported by an Alberta Innovates - Health Solutions Clinician Fellowship Award, a Knowledge Translation Canada Strategic Training in Health Research Fellowship, and funding from the Clinician Investigator and Surgeon Scientist Programs at the University of Calgary. CL and CNJ are supported by the Calvin, Phoebe and Joan Snyder Chair in Critical Care Medicine based at the University of Calgary and awarded to PK. AWK has an investigator-initiated trial funding agreement with Kinetic Concepts Incorporated (KCI) USA and the Governors of the University of Calgary for the clinical and laboratory costs of conducting the study (KCI contract number: KCI Clinical/UniversityCalgaryAlbertaHealth/082611-000/7). KCI USA will have no role in the design or conduct of the study; collection, management, analysis, or interpretation of the data; or preparation, review, or approval of the manuscript. The remaining authors have no competing interests to declare. We would like to thank Anita Verdonk, RN, BScN, CEN, ET, who was formerly a clinical therapies specialist with KCI Medical Canada Inc., for providing technical information on the ABThera™ device. We would also like to thank Sandy Cochrane at Multimedia Services, University of Calgary for assistance with creation of the illustrations of the ABThera™ and Barker's vacuum pack.

Author details

¹Department of Surgery, University of Calgary and the Foothills Medical Centre, North Tower 10th Floor, 1403-29th Street Northwest, Calgary, Alberta T2N 2T9, Canada. ²Department of Community Health Sciences (Divisions of Epidemiology and Biostatistics), University of Calgary, TRW (Teaching, Research, and Wellness) Building, 3rd Floor, 3280 Hospital Drive Northwest, Calgary, Alberta T2N 4Z6, Canada. ³Department of Critical Care Medicine, University of Calgary and the Foothills Medical Centre, Ground Floor McCaig Tower, 3134 Hospital Drive Northwest, Calgary, Alberta T2N 5A1, Canada. ⁴Department of Oncology, University of Calgary and the Foothills Medical Centre, 1403-29th Street Northwest, Calgary, Alberta T2N 2T9, Canada. ⁵Regional Trauma Program, University of Calgary and the Foothills Medical Centre, 1403-29th Street Northwest, Calgary, Alberta T2N 2T9, Canada. ⁶Alberta Health Services - Research Excellence Support Team, University of Calgary and the Foothills Medical Centre, South Tower, 1403-29th Street

Northwest, Calgary, Alberta T2N 2T9, Canada. ⁷Hotchkiss Brain Institute, University of Calgary and the Foothills Medical Centre, Health Research Innovation Centre, 3330 Hospital Drive Northwest, Calgary, Alberta T2N 4N1, Canada. ⁸Calvin, Phoebe and Joan Snyder Institute for Chronic Diseases, University of Calgary and the Foothills Medical Centre, Health Research Innovation Centre, 3280 Hospital Drive Northwest, Calgary, Alberta T2N 4N1, Canada. ⁹Division of Critical Care Medicine, Department of Medicine, University of British Columbia and St. Paul's Hospital, 1081 Burrard Street, Vancouver, British Columbia V6Z 1Y6, Canada.

Received: 5 December 2012 Accepted: 30 April 2013

Published: 16 May 2013

References

1. Murray CJ, Lopez AD: **Mortality by cause for eight regions of the world: global burden of disease study.** *Lancet* 1997, **349**:1269-1276.
2. Chow AW, Evans GA, Nathens AB, Ball CG, Hansen G, Harding GK, Kirkpatrick AW, Weiss K, Zhanel GG: **Canadian practice guidelines for surgical intra-abdominal infections.** *Can J Infect Dis Med Microbiol* 2010, **21**:11-37.
3. Stelfox HT, Bobranska-Artiuch B, Nathens A, Straus SE: **Quality indicators for evaluating trauma care: a scoping review.** *Arch Surg* 2010, **145**:286-295.
4. Nishijima DK, Simel DL, Wisner DH, Holmes JF: **Does this adult patient have a blunt intra-abdominal injury?** *JAMA* 2012, **307**:1517-1527.
5. Chovanes J, Cannon JW, Nunez TC: **The evolution of damage control surgery.** *Surg Clin North Am* 2012, **92**:859-875. vii-viii.
6. Waibel BH, Rotondo MF: **Damage control for intra-abdominal sepsis.** *Surg Clin North Am* 2012, **92**:243-257. vii.
7. Stone HH, Strom PR, Mullins RJ: **Management of the major coagulopathy with onset during laparotomy.** *Ann Surg* 1983, **197**:532-535.
8. Burch JM, Ortiz VB, Richardson RJ, Martin RR, Mattox KL, Jordan GL Jr: **Abbreviated laparotomy and planned reoperation for critically injured patients.** *Ann Surg* 1992, **215**:476-483. discussion 483-484.
9. Rotondo MF, Schwab CW, McGonigal MD, Phillips GR III, Fruchterman TM, Kauder DR, Latenser BA, Angood PA: **'Damage control': an approach for improved survival in exsanguinating penetrating abdominal injury.** *J Trauma* 1993, **35**:375-382. discussion 382-383.
10. Roberts DJ, Zygun DA, Grendar J, Ball CG, Robertson HL, Ouellet JF, Cheatham ML, Kirkpatrick AW: **Negative-pressure wound therapy for critically ill adults with open abdominal wounds: a systematic review.** *J Trauma Acute Care Surg* 2012, **73**:629-639.
11. Holzheimer RG, Schein M, Wittmann DH: **Inflammatory response in peritoneal exudate and plasma of patients undergoing planned relaparotomy for severe secondary peritonitis.** *Arch Surg* 1995, **130**:1314-1319. discussion 1319-1320.
12. Scheingraber S, Bauerfeind F, Bohme J, Dralle H: **Limits of peritoneal cytokine measurements during abdominal lavage treatment for intraabdominal sepsis.** *Am J Surg* 2001, **181**:301-308.
13. Mayberry JC, Welker KJ, Goldman RK, Mullins RJ: **Mechanism of acute ascites formation after trauma resuscitation.** *Arch Surg* 2003, **138**:773-776.
14. Fink MP, Delude RL: **Epithelial barrier dysfunction: a unifying theme to explain the pathogenesis of multiple organ dysfunction at the cellular level.** *Crit Care Clin* 2005, **21**:177-196.
15. Ince C: **The microcirculation is the motor of sepsis.** *Crit Care* 2005, **Suppl 4**:S13-S19.
16. van Veen SQ, Meijers JC, Levi M, van Gulik TM, Boermeester MA: **Effects of intra-abdominal administration of recombinant tissue plasminogen activator on coagulation, fibrinolysis and inflammatory responses in experimental polymicrobial peritonitis.** *Shock* 2007, **27**:534-541.
17. el Zakaria R, Li N, Garrison RN: **Mechanisms of direct peritoneal resuscitation-mediated splanchnic hyperperfusion following hemorrhagic shock.** *Shock* 2007, **27**:436-442.
18. Kubiak BD, Albert SP, Gatto LA, Snyder KP, Maier KG, Vieau CJ, Roy S, Nieman GF: **Peritoneal negative pressure therapy prevents multiple organ injury in a chronic porcine sepsis and ischemia/reperfusion model.** *Shock* 2010, **34**:525-534.
19. Hendriks T, Bleichrodt RP, Lomme RM, De Man BM, van Goor H, Buyne OR: **Peritoneal cytokines predict mortality after surgical treatment of secondary peritonitis in the rat.** *J Am Coll Surg* 2010, **211**:263-270.
20. Ravishankaran P, Shah AM, Bhat R: **Correlation of interleukin-6, serum lactate, and C-reactive protein to inflammation, complication, and**

- outcome during the surgical course of patients with acute abdomen. *J Interferon Cytokine Res* 2011, **31**:685–690.
21. Li P, Xu Q, Ji Z, Gao Y, Zhang X, Duan Y, Guo Z, Zheng B, Guo X, Wu X: **Comparison of surgical stress between laparoscopic and open appendectomy in children.** *J Pediatr Surg* 2005, **40**:1279–1283.
 22. Roelofsens H, Alvarez-Llamas G, Dijkstra M, Breitling R, Havenga K, Bijzet J, Zandbergen W, de Vries MP, Ploeg RJ, Vonk RJ: **Analyses of intricate kinetics of the serum proteome during and after colon surgery by protein expression time series.** *Proteomics* 2007, **7**:3219–3228.
 23. Xiao W, Mindrinos MN, Seok J, Cuschieri J, Cuenca AG, Gao H, Hayden DL, Hennessy L, Moore EE, Minei JP, Bankey PE, Johnson JL, Sperry J, Nathens AB, Billiar TR, West MA, Brownstein BH, Mason PH, Baker HV, Finnerty CC, Jeschke MG, Lopez MC, Klein MB, Gamelli RL, Gibran NS, Arnoldo B, Xu W, Zhang Y, Calvano SE, McDonald-Smith GP, Inflammation and Host Response to Injury Large-Scale Collaborative Research Program, et al: **A genomic storm in critically injured humans.** *J Exp Med* 2011, **208**:2581–2590.
 24. Liu T, Qian WJ, Gritsenko MA, Xiao W, Moldawer LL, Kaushal A, Monroe ME, Vamum SM, Moore RJ, Purvine SO, Maier RV, Davis RW, Tompkins RG, Camp DG III, Smith RD, Inflammation and the Host Response to Injury Large Scale Collaborative Research Program: **High dynamic range characterization of the trauma patient plasma proteome.** *Mol Cell Proteomics* 2006, **5**:1899–1913.
 25. Hack CE, De Groot ER, Felt-Bersma RJ, Nuijens JH, Strack Van Schijndel RJ, Eerenberg-Belmer AJ, Thijs LG, Aarden LA: **Increased plasma levels of interleukin-6 in sepsis.** *Blood* 1989, **74**:1704–1710.
 26. Latifi SQ, O'Riordan MA, Levine AD, Stallion A: **Persistent elevation of serum interleukin-6 in intraabdominal sepsis identifies those with prolonged length of stay.** *J Pediatr Surg* 2004, **39**:1548–1552.
 27. Gletsu N, Lin E, Zhu JL, Khaitan L, Ramshaw BJ, Farmer PK, Ziegler TR, Papanicolaou DA, Smith CD: **Increased plasma interleukin 6 concentrations and exaggerated adipose tissue interleukin 6 content in severely obese patients after operative trauma.** *Surgery* 2006, **140**:50–57.
 28. Frangen TM, Bogdanski D, Schinkel C, Roetman B, Kalicke T, Muhr G, Koller M: **Systemic IL-17 after severe injuries.** *Shock* 2008, **29**:462–467.
 29. Zeilhofer HU, Schorr W: **Role of interleukin-8 in neutrophil signaling.** *Curr Opin Hematol* 2000, **7**:178–182.
 30. Bingold TM, Ziesche E, Scheller B, Sadik CD, Franck K, Just L, Sartorius S, Wahrmann M, Wissing H, Zwissler B, Pfeilschifter J, Muhl H: **Interleukin-22 detected in patients with abdominal sepsis.** *Shock* 2010, **34**:337–340.
 31. Rongione AJ, Kusske AM, Ashley SW, Reber HA, McFadden DW: **Interleukin-10 prevents early cytokine release in severe intraabdominal infection and sepsis.** *J Surg Res* 1997, **70**:107–112.
 32. van Berge Henegouwen MI, van der Poll T, van Deventer SJ, Gouma DJ: **Peritoneal cytokine release after elective gastrointestinal surgery and postoperative complications.** *Am J Surg* 1998, **175**:311–316.
 33. Kimura A, Ono S, Hiraki S, Takahata R, Tsujimoto H, Miyazaki H, Kinoshita M, Hatsuse K, Saitoh D, Hase K, Yamamoto J: **The postoperative serum interleukin-15 concentration correlates with organ dysfunction and the prognosis of septic patients following emergency gastrointestinal surgery.** *J Surg Res* 2012, **175**:e83–e88.
 34. Pappu R, Ramirez-Carrozzi V, Sambandam A: **The interleukin-17 cytokine family: critical players in host defence and inflammatory diseases.** *Immunology* 2011, **134**:8–16.
 35. Freitas A, Alves-Filho JC, Victori T, Secher T, Lemos HP, Sonogo F, Cunha FQ, Ryffel B: **IL-17 receptor signaling is required to control polymicrobial sepsis.** *J Immunol* 2009, **182**:7846–7854.
 36. Alves-Filho JC, Sonogo F, Souto FO, Freitas A, Verri WA Jr, Auxiliadora-Martins M, Basile-Filho A, McKenzie AN, Xu D, Cunha FQ, Liew FY: **Interleukin-33 attenuates sepsis by enhancing neutrophil influx to the site of infection.** *Nat Med* 2010, **16**:708–712.
 37. Dib M, Zhao X, Wang X, Andersson E, Drewsen G, Andersson R: **Acute phase response in acute pancreatitis: a comparison with abdominal sepsis.** *Scand J Gastroenterol* 2003, **38**:1072–1077.
 38. Brenner T, Hofer S, Rosenhagen C, Steppan J, Lichtenstern C, Weitz J, Bruckner T, Lukic IK, Martin E, Bierhaus A, Hoffmann U, Weigand MA: **Macrophage migration inhibitory factor (MIF) and manganese superoxide dismutase (MnSOD) as early predictors for survival in patients with severe sepsis or septic shock.** *J Surg Res* 2010, **164**:e163–e171.
 39. Calandra T, Echtenacher B, Roy DL, Pugin J, Metz CN, Hultner L, Heumann D, Mannel D, Bucala R, Glauser MP: **Protection from septic shock by neutralization of macrophage migration inhibitory factor.** *Nat Med* 2000, **6**:164–170.
 40. Reith HB, Mittelkotter U, Wagner R, Thiede A: **Procalcitonin (PCT) in patients with abdominal sepsis.** *Intensive Care Med* 2000, **Suppl 2**:S165–S169.
 41. Cziupka K, Busemann A, Pardecke LI, Potschke C, Rath M, Traeger T, Koerner P, von Bernstorff W, Kessler W, Diedrich S, Weiss FU, Maier S, Broker BM, Heidecke CD: **Tumor necrosis factor-related apoptosis-inducing ligand (TRAIL) improves the innate immune response and enhances survival in murine polymicrobial sepsis.** *Crit Care Med* 2010, **38**:2169–2174.
 42. Wang S: **TRAIL: a sword for killing tumors.** *Curr Med Chem* 2010, **17**:3309–3317.
 43. McBeth PB, Leger C, Ball CG, Ouellet JF, Tiruta C, Laupland KB, Kubes P, Roberts DJ, Shahpori R, Kirkpatrick AW: **Intra-abdominal hypertension and intra-abdominal sepsis: critical concepts and possibilities.** *Int J Intensive Care* 2011, **18**:10–19.
 44. Casey LC, Balk RA, Bone RC: **Plasma cytokine and endotoxin levels correlate with survival in patients with the sepsis syndrome.** *Ann Intern Med* 1993, **119**:771–778.
 45. Roumen RM, Hendriks T, van der Ven-Jongekrijg J, Nieuwenhuijzen GA, Sauerwein RW, van der Meer JW, Goris RJ: **Cytokine patterns in patients after major vascular surgery, hemorrhagic shock, and severe blunt trauma. Relation with subsequent adult respiratory distress syndrome and multiple organ failure.** *Ann Surg* 1993, **218**:769–776.
 46. Pinsky MR, Vincent JL, Deviere J, Alegre M, Kahn RJ, Dupont E: **Serum cytokine levels in human septic shock. Relation to multiple-system organ failure and mortality.** *Chest* 1993, **103**:565–575.
 47. Svoboda P, Kantorova I, Ochmann J: **Dynamics of interleukin 1, 2, and 6 and tumor necrosis factor alpha in multiple trauma patients.** *J Trauma* 1994, **36**:336–340.
 48. Pettila V, Hynninen M, Takkunen O, Kuusela P, Valtonen M: **Predictive value of procalcitonin and interleukin 6 in critically ill patients with suspected sepsis.** *Intensive Care Med* 2002, **28**:1220–1225.
 49. Spindler-Vesel A, Wraber B, Vovk I, Kompan L: **Intestinal permeability and cytokine inflammatory response in multiply injured patients.** *J Interferon Cytokine Res* 2006, **26**:771–776.
 50. Jawa RS, Anillo S, Huntoon K, Baumann H, Kulaylat M: **Interleukin-6 in surgery, trauma, and critical care part II: clinical implications.** *J Intensive Care Med* 2011, **26**:73–87.
 51. Quyn AJ, Johnston C, Hall D, Chambers A, Arapova N, Ogston S, Amin AI: **The open abdomen and temporary abdominal closure systems—historical evolution and systematic review.** *Colorectal Dis* 2012, **14**:e429–e438.
 52. Bee TK, Croce MA, Magnotti LJ, Zarzar BL, Maish GO III, Minard G, Schroepel TJ, Fabian TC: **Temporary abdominal closure techniques: a prospective randomized trial comparing polyglactin 910 mesh and vacuum-assisted closure.** *J Trauma* 2008, **65**:337–342. discussion 342–344.
 53. *Clinical guidelines for the management of the open abdomen with ABThera™ Open Abdomen Negative Pressure Therapy System for active abdominal therapy.* [<http://peritonealvac.com/ABThera%20Clinical%20Guidelines.pdf>].
 54. Brock WB, Barker DE, Burns RP: **Temporary closure of open abdominal wounds: the vacuum pack.** *Am Surg* 1995, **61**:30–35.
 55. Smith LA, Barker DE, Chase CW, Somberg LB, Brock WB, Burns RP: **Vacuum pack technique of temporary abdominal closure: a four-year experience.** *Am Surg* 1997, **63**:1102–1107. discussion 1107–1108.
 56. Barker DE, Kaufman HJ, Smith LA, Ciraulo DL, Richart CL, Burns RP: **Vacuum pack technique of temporary abdominal closure: a 7-year experience with 112 patients.** *J Trauma* 2000, **48**:201–206. discussion 206–207.
 57. Schulz KF, Altman DG, Moher D, CONSORT Group: **CONSORT 2010 Statement: updated guidelines for reporting parallel group randomised trials.** *Trials* 2010, **11**:32.
 58. Sammons A, Delgado A, Cheatham ML: *In-vitro pressure manifold distribution evaluation of the ABThera Open Abdomen Negative Pressure Therapy System, V.A.C. Abdominal Dressing System, and Barker's vacuum-pack technique, conducted under dynamic conditions.* San Antonio, TX: [Abst P 078] Clinical Symposium on Advances in Skin & Wound Care; 2009.
 59. Diaz JJ Jr, Cullinane DC, Dutton WD, Jerome R, Bagdonas R, Bilaniuk JW, Collier BR, Como JJ, Cumming J, Griffen M, Gunter OL, Kirby J, Lottenburg L, Mowery N, Riordan WP Jr, Martin N, Platz J, Stassen N, Winston ES: **The management of the open abdomen in trauma and emergency general surgery: part 1—damage control.** *J Trauma* 2010, **68**:1425–1438.
 60. Asensio JA, Petrone P, O'Shanahan G, Kuncir EJ: **Managing exsanguination: what we know about damage control/bailout is not enough.** *Proc (Bayl Univ Med Cent)* 2003, **16**:294–296.
 61. Ivatury RR, Malhotra AK: **Damage Control.** In *Trauma: Emergency Resuscitation, Perioperative Anesthesia, Surgical Management.* Edited by

- Wilson WC, Grande CM, Hoyt DB. New York, NY: Informa Healthcare USA, Inc; 2007:405–416.
62. Kaplan M, Banwell P, Orgill DP, Ivatury RR, Demetriades D, Moore FA, Miller P, Nicholas J, Henry S: **Guidelines for the management of the open abdomen.** *Wounds-Compend Clin Res Pract* 2005, **Suppl**:1–24.
63. Knaus WA, Draper EA, Wagner DP, Zimmerman JE: **APACHE II: a severity of disease classification system.** *Crit Care Med* 1985, **13**:818–829.
64. Vincent JL, Moreno R, Takala J, Willatts S, De Mendonca A, Bruining H, Reinhart CK, Suter PM, Thijs LG: **The SOFA (sepsis-related organ failure assessment) score to describe organ dysfunction/failure. On behalf of the working group on sepsis-related problems of the European society of intensive care medicine.** *Intensive Care Med* 1996, **22**:707–710.
65. Baker SP, O'Neill B, Haddon W Jr, Long WB: **The injury severity score: a method for describing patients with multiple injuries and evaluating emergency care.** *J Trauma* 1974, **14**:187–196.
66. Linn S: **The injury severity score—importance and uses.** *Ann Epidemiol* 1995, **5**:440–446.
67. Bellomo R, Ronco C, Kellum JA, Mehta RL, Palevsky P, Acute Dialysis Quality Initiative workgroup: **Acute renal failure - definition, outcome measures, animal models, fluid therapy and information technology needs: the second international consensus conference of the acute dialysis quality initiative (ADQI) group.** *Crit Care* 2004, **8**:R204–R212.
68. Cheatham ML, Malbrain ML, Kirkpatrick A, Sugrue M, Parr M, De Waele J, Balogh Z, Leppaniemi A, Olvera C, Ivatury R, D'Amours S, Wendon J, Hillman K, Wilmer A: **Results from the international conference of experts on intra-abdominal hypertension and abdominal compartment syndrome. II. Recommendations.** *Intensive Care Med* 2007, **33**:951–962.
69. Dupont NC, Wang K, Wadhwa PD, Culhane JF, Nelson EL: **Validation and comparison of luminex multiplex cytokine analysis kits with ELISA: determinations of a panel of nine cytokines in clinical sample culture supernatants.** *J Reprod Immunol* 2005, **66**:175–191.
70. Shah SK, Jimenez F, Walker PA, Aroom KR, Xue H, Feeley TD, Uray KS, Norbury KC, Stewart RH, Laine GA, Cox CS Jr: **A novel mechanism for neutrophil priming in trauma: potential role of peritoneal fluid.** *Surgery* 2010, **148**:263–270.
71. Shah SK, Jimenez F, Walker PA, Xue H, Feeley TD, Uray KS, Norbury KC, Stewart RH, Laine GA, Cox CS Jr: **Peritoneal fluid: a potential mechanism of systemic neutrophil priming in experimental intra-abdominal sepsis.** *Am J Surg* 2012, **203**:211–216.
72. Ouellet JF, Ball CG: **Recurrent abdominal compartment syndrome induced by high negative pressure abdominal closure dressing.** *J Trauma* 2011, **71**:785–786.
73. Rao M, Burke D, Finan PJ, Sagar PM: **The use of vacuum-assisted closure of abdominal wounds: a word of caution.** *Colorectal Dis* 2007, **9**:266–268.
74. Fischer JE: **A cautionary note: the use of vacuum-assisted closure systems in the treatment of gastrointestinal cutaneous fistula may be associated with higher mortality from subsequent fistula development.** *Am J Surg* 2008, **196**:1–2.
75. Ware JH: **Linear models for the analysis of longitudinal studies.** *Am Stat* 1985, **39**:95–101.
76. Laird NM, Donnelly C, Ware JH: **Longitudinal studies with continuous responses.** *Stat Methods Med Res* 1992, **1**:225–247.
77. Liu G, Gould AL: **Comparison of alternative strategies for analysis of longitudinal trials with dropouts.** *J Biopharm Stat* 2002, **12**:207–226.
78. Mallinckrodt CH, Clark WS, David SR: **Accounting for dropout bias using mixed-effects models.** *J Biopharm Stat* 2001, **11**:9–21.
79. Benjamini Y, Hochberg Y: **Controlling the false discovery rate: a practical and powerful approach to multiple testing.** *J R Statist Soc B* 1995, **57**:289–300.
80. Newson R: **Multiple-test procedures and smile plots.** *Stata J* 2003, **3**:109–132.
81. Newson RB: **Frequentist q-values for multiple-test procedures.** *Stata J* 2010, **10**:568–584.
82. van Ruler O, Mahler CW, Boer KR, Reuland EA, Gooszen HG, Opmeer BC, de Graaf PW, Lamme B, Gerhards MF, Steller EP, van Till JW, de Borgie CJ, Gouma DJ, Reitsma JB, Boermeester MA, Dutch Peritonitis Study Group: **Comparison of on-demand vs planned relaparotomy strategy in patients with severe peritonitis: a randomized trial.** *JAMA* 2007, **298**:865–872.
83. Opmeer BC, Boer KR, van Ruler O, Reitsma JB, Gooszen HG, de Graaf PW, Lamme B, Gerhards MF, Steller EP, Mahler CM, Obertop H, Gouma DJ, Bossuyt PM, de Borgie CA, Boermeester MA: **Costs of relaparotomy on-demand versus planned relaparotomy in patients with severe peritonitis: an economic evaluation within a randomized controlled trial.** *Crit Care* 2010, **14**:R97.

doi:10.1186/1745-6215-14-141

Cite this article as: Roberts *et al.*: Efficacy and safety of active negative pressure peritoneal therapy for reducing the systemic inflammatory response after damage control laparotomy (the Intra-peritoneal Vacuum Trial): study protocol for a randomized controlled trial. *Trials* 2013 **14**:141.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

