



FAS-670A > G gene polymorphism and the risk of allograft rejection after organ transplantation: a systematic review and meta-analysis

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Abstract

The association between the risk of allograft rejection after organ transplantation and FAS gene polymorphism has been evaluated previously. However, inconsistent results have been reported. Hence, we conducted the most up-to-date meta-analysis to evaluate this association. All eligible studies reporting the association between FAS-670A > G polymorphism and the risk of allograft rejection published up to December 2019 were extracted using a comprehensive systematic database search in the Web of Science, Scopus, and PubMed. The pooled odds ratios (OR) and corresponding 95% confidence intervals (CI) were calculated to determine the association strength. This meta-analysis included six case-control studies with 277 patients who experienced allograft rejection and 1,001 patients who did not experience allograft rejection (controls) after organ transplantation. The overall results showed no significant association between FAS-670A > G polymorphism and the risk of allograft rejection in five genetic models (dominant model: OR=0.81, 95% CI=0.58–1.12; recessive model: OR=0.10, 95% CI=0.80–1.53; allelic model: OR=0.96, 95% CI=0.79–1.18; GG vs. AA: OR=0.92, 95% CI=0.62–1.36; and AG vs. AA: OR=0.75, 95% CI=0.52–1.08). Moreover, subgroup analysis according to ethnicity and age did not reveal statistically significant results. Our findings suggest that FAS-670A > G polymorphism is not associated with the risk of allograft rejection after organ transplantation.

Key Words FAS, Allograft rejection, Polymorphism, Meta-analysis

INTRODUCTION

Organ transplantation, such as renal, liver, and heart transplantation, is the best therapeutic option for most patients with end-stage disease [1]. Over the past decades, due to new advances in surgical techniques, expansion of effective immunosuppressive agents, and better recognition of alloimmune response and histocompatibility matching, the short- and long-term graft survival outcomes in transplant recipients have improved [2]. However, immunosuppressive protocols have increased the rates of infection and malignancy in patients undergoing organ transplantation [3]. Therefore, it is important to identify the factors that influence the risk of rejection in such diseases. A growing body of evidence

supports that apoptosis contributes to graft rejection and the establishment of tolerance in transplantation.

FAS is one of the most important inducers of the apoptotic pathway [4]. FAS (also known as CD95/TNFSF6/APO-1) is a cell surface receptor belonging to the tumor necrosis factor receptor (TNF-R) family and is highly expressed in a wide range of cells, including lymphocytes, neutrophils, monocytes, and tissues such as the heart, kidney, and liver [5, 6]. Its gene, located on chromosome 10q24.1, consists of nine exons and eight introns and is highly polymorphic [7]. Apoptosis plays a pivotal role in the deletion of self-reactive lymphocytes, including immature T cells and peripheral mature T cells [8], and death of target cells by effector cytotoxic T lymphocytes (CTLs) [9]. Significant depletion of renal tubular epithelial cells by apoptosis in kidney recipi-

ents experiencing acute or chronic allograft rejection has been described [10]. Moreover, hepatocyte apoptosis has been detected in acute liver graft rejection [11]. However, some studies have shown that apoptosis of activated T cells within accepted grafts plays a significant role in inducing hepatic tolerance [12].

Some studies have suggested that the FAS gene is controlled by various genetic elements positioned in the 5-upstream promoter regions of the gene, especially in the transcription factor binding sites [13]. However, a functional polymorphism involving an A→G transition at position -670 in the enhancer region (Fas-670A>G, rs1800682) of FAS has been reported. This polymorphism destroys signal transducer activator of transcription 1 (STAT1), consequently reducing promoter activity and diminishing FAS expression [14, 15]. Because of the importance of this single nucleotide polymorphism in the susceptibility of recipient T cells to FASL-mediated apoptosis [16], we performed a meta-analysis to determine whether Fas-670A>G gene polymorphism is associated with the risk of allograft rejection after organ transplantation.

METHODS

The present meta-analysis was performed in accordance with the Preferred Reporting Items for Systematic reviews and Meta-Analyses (PRISMA) statement [17], including search strategy, inclusion and exclusion criteria, data ex-

traction and quality assessment, and statistical analysis.

Search strategy

All studies reporting the association between FAS-670A>G polymorphism and susceptibility to allograft rejection after organ transplantation until December 2019 were retrieved by a systematic search of PubMed, Scopus, and Web of Science. The following combinations of keywords were used: (“rejection” or “graft failure”) and (“APO-1” or “TNFSF6” or “CD95” or “FAS”) and (“polymorphism” or “variation” or “mutation” or “allele” or “genotype” or “SNP” or “single nucleotide polymorphism”). Furthermore, we manually screened the reference lists of eligible studies and relevant reviews to identify missing data during the electronic search.

Inclusion and exclusion criteria

Studies were considered eligible if they met the following criteria: a) studies that evaluated the association between allograft rejection and FAS-670A>G polymorphism; b) studies providing adequate data to calculate the odds ratio (OR) and its 95% confidence interval (CI), and c) studies including two comparison groups (rejection group vs. non-rejection group). Other studies, such as review articles, book chapters, editorials, comments, abstracts, duplicated data, and republished articles, were excluded.

Data extraction and quality assessment

Two authors independently extracted the following data according to an extraction checklist: first author’s name,

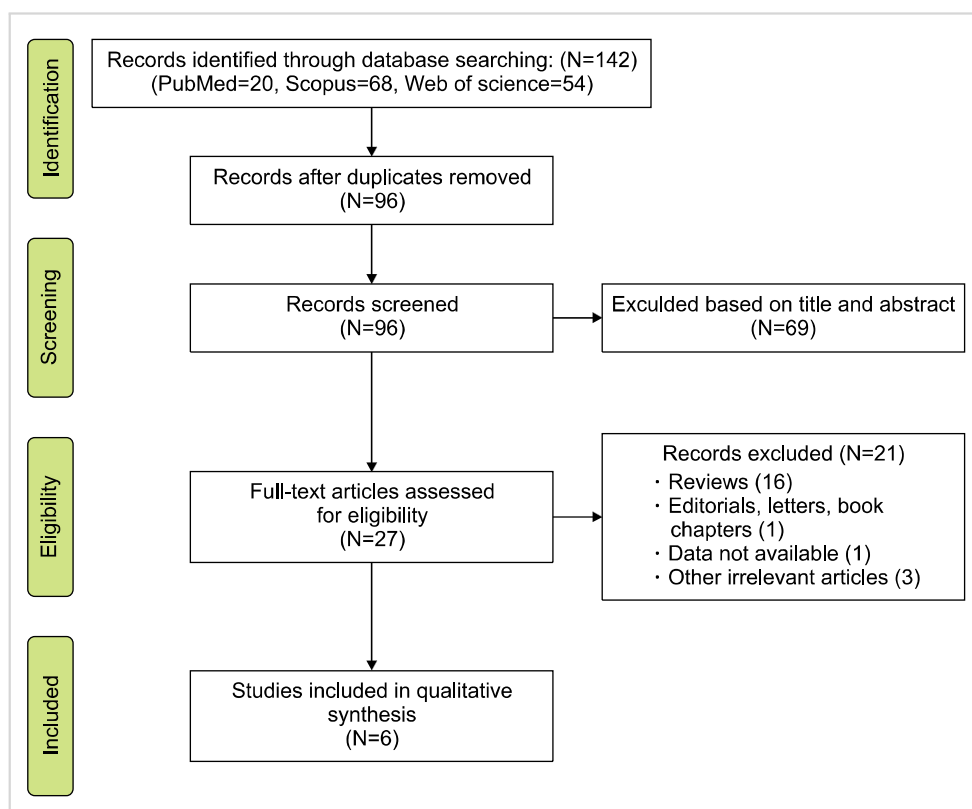


Fig. 1. Flow diagram of the study selection process.

journal and year of publication, ethnicity, country of origin, mean of age, methods for genotyping, sample size of cases and controls, and the number of cases and controls for each genotype. Any discrepancies between the two reviewers were discussed and resolved by consensus. The quality of each study was assessed using the Newcastle-Ottawa Scale (NOS) criteria [18]. Studies with scores of 0-3, 4-6, or 7-9 were considered low-, moderate-, or high-quality studies, respectively.

Statistical analysis

For each case-control study, deviation from the Hardy-Weinberg equilibrium was analyzed using the χ^2 test in the control group. The pooled OR and 95% CI were computed to evaluate the strength of associations between FAS-670A >G gene polymorphism and the risk of rejection after organ transplantation. Different possible comparison models for FAS-670A >G gene single-nucleotide polymorphism (SNP) included the dominant model (GG+AG vs. AA), recessive model (GG vs. AG+AA), allelic model (G vs. A), homozygote (GG vs. AA), and heterozygote (AG vs. AA). Heterogeneity among the included studies was measured using Q statistics ($P < 0.1$ was considered statistically significant) and I^2 test (I^2 values of 25%, 50%, and 75%

were described as low, moderate, and high heterogeneity, respectively) [19, 20]. If heterogeneity was detected, a random effects model (Der Simonian-Laird approach) was used; otherwise, the fixed effects model (Mantel-Haenszel approach) was used (Q statistic $P > 0.1$ or $I^2 < 50\%$) [21]. Sensitivity analysis was used to evaluate the stability of our results. Publication bias was estimated using funnel plots and Begg's and Egger's tests [22, 23] ($P < 0.05$ was considered statistically significant). This meta-analysis was performed using STATA 14.0 software (State Corporation, College Station, TX, USA).

RESULTS

Characteristics of the studies included in the meta-analysis

Fig. 1 shows the flow diagram of the study selection process according to the PRISMA guidelines. In total, 142 studies were identified during the primary search. Subsequently, duplicates (N=46) were omitted, and other studies were excluded either by title and abstract (N=69) or full-text (N=21) screening. Eventually, six publications reporting the association between FAS-670A >G gene polymorphism and the risk of rejection were included in the quantitative analysis

Table 1. Characteristics of studies included in meta-analysis of overall FAS-670A >G.

Study author	Year	Country	Ethnicity	Sex cases/controls	Total cases/control	Age case/control (mean)	Genotyping method	Quality score
Cappellesso <i>et al.</i> [24]	2002	France	Caucasian	M=NR F=NR	20/77	NR/NR	RFLP-PCR	6
Marín <i>et al.</i> [25]	2006	Spain	Caucasian	M=NR F=NR	53/227	49±12/NR	RFLP-PCR	7
Jahadi Hosseini <i>et al.</i> [26]	2009	Iran	Caucasian	M=NR F=NR	47/225	43.67±22.18/40.08±22.18	ASO-PCR	7
Ertan <i>et al.</i> [16]	2010	Turkey	Caucasian	M=NR F=NR	16/37	12.3±0.6/12.3±0.6	RFLP-PCR	7
Girnita <i>et al.</i> [27]	2011	Multicenter	Mixed	M=NR F=NR	124/405	NR/NR	PCR	6
Fadel <i>et al.</i> [28]	2016	Egypt	Arab	M=10/19 F=7/11	17/30	9.37±3.56/10.09±2.95	RFLP-PCR	8

Abbreviations: F, female; M, male; NR, not reported.

Table 2. Distribution of genotype and allele among FAS 670A/G patients and controls.

Study author	Rejection cases					Non-rejection control					P-HWE	MAF
	AA	AG	GG	A	G	AA	AG	GG	A	G		
Cappellesso <i>et al.</i> [24]	8	9	3	25	15	25	40	12	90	64	0/54	0/415
Marín <i>et al.</i> [25]	15	24	14	54	52	65	106	56	236	218	0/33	0/48
Jahadi Hosseini <i>et al.</i> [26]	12	20	15	44	50	77	73	75	227	223	≤0.001	0/495
Ertan <i>et al.</i> [16]	4	10	2	18	14	11	23	3	45	29	0/06	0/391
Girnita <i>et al.</i> [27]	40	46	38	126	122	81	213	111	375	435	0/24	0/537
Fadel <i>et al.</i> [28]	4	3	10	11	23	11	3	16	25	35	≤0.001	0/583

Abbreviations: MAF, minor allele frequency of the control group; P-HWE, P-value for Hardy-Weinberg equilibrium.

[16, 24-28]. The studies were performed in different countries, including Iran, France, Spain, Turkey, and Egypt. All eligible studies had good overall methodological scores, ranging from 6 to 8. Restriction fragment length polymorphism was the common genotyping method used in the included studies. The characteristics, allele frequency, and genotype

distributions of the included studies are summarized in Tables 1 and 2.

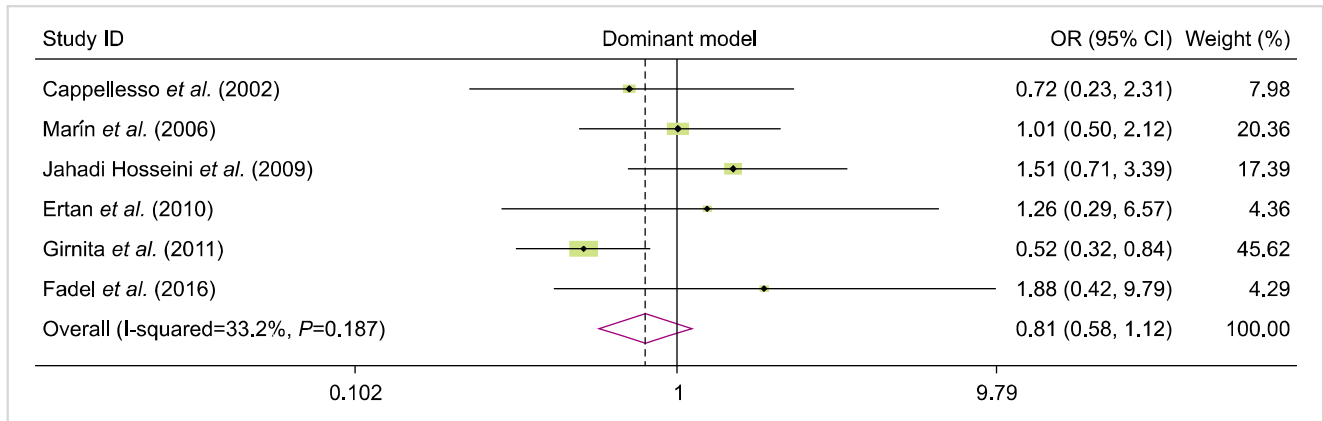


Fig. 2. Forest plot of the association between FAS-670A>G gene single-nucleotide polymorphism and the risk of allograft rejection in the dominant model.

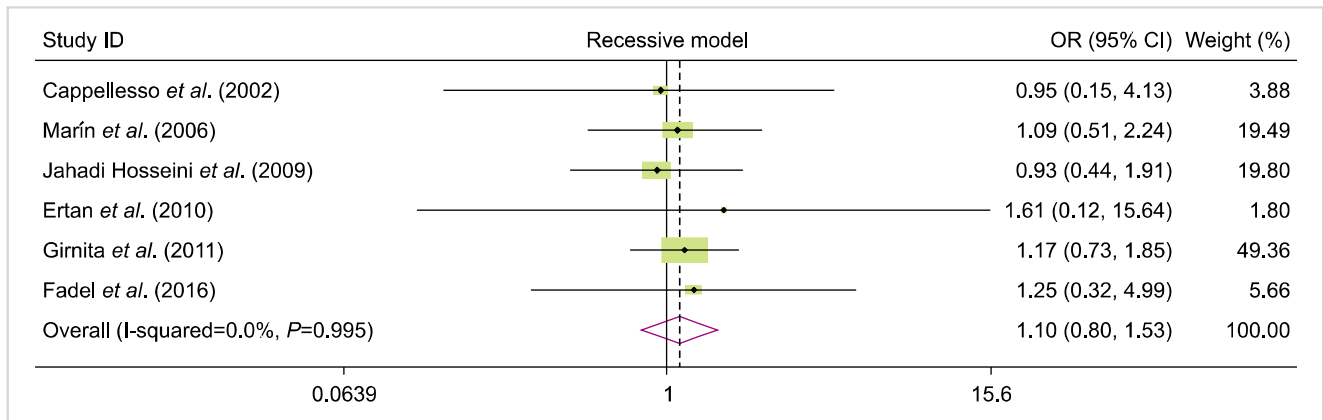


Fig. 3. Forest plot of the association between FAS-670A>G gene single-nucleotide polymorphism and the risk of allograft rejection in the recessive model.

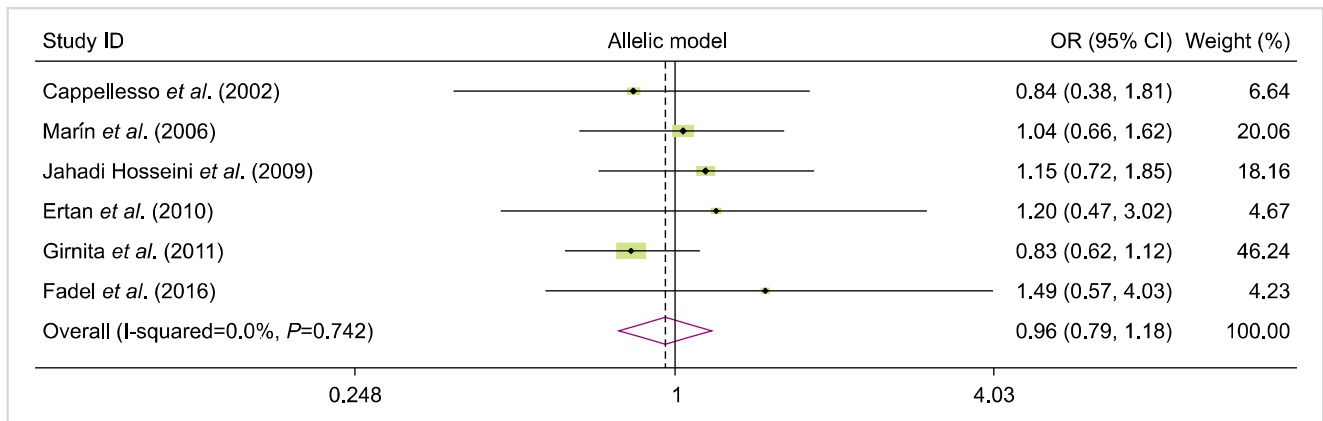


Fig. 4. Forest plot of the association between FAS-670A>G gene single-nucleotide polymorphism and the risk of allograft rejection in the allelic model.

Meta-analysis of FAS-670A > G gene polymorphism and the risk of allograft rejection

We analyzed all eligible studies on the association between FAS-670A > G polymorphism and the risk of allograft rejection after organ transplantation. The pooled effect size indicated that there was no significant association between FAS-670A > G gene polymorphisms and the risk of allograft rejection across the different genotype models—dominant model (OR=0.81, 95% CI=0.58–1.12, *P*=0.19, REM), recessive model (OR=0.10, 95% CI=0.80–1.53, *P*=0.55, REM), allelic model (OR=0.96, 95% CI=0.79, 1.18, *P*=0.7, REM), GG vs. AA model (OR=0.92, 95% CI=0.62–1.36, *P*=0.66, REM), and AG vs. AA model (OR=0.75, 95% CI=0.52–1.08, *P*=0.12, REM) (Figs. 2–6).

Subgroup analysis according to ethnicity and age

We categorized studies according to ethnicity—Caucasians (4 articles), mixed (1 article), and Arabs (1 article). Since there was only one study for the mixed and Arab populations, these studies were excluded from the analysis. The results of subgroup analysis in the Caucasian population did not reveal any significant association between FAS-670A > G

gene polymorphisms and the risk of allograft rejection in all genetic models. Additionally, categorized studies according to age—children (3 articles) and adults (2 articles). The results did not reveal any statistically significant association. The details are listed in Table 3.

Evaluation of heterogeneity and publication bias

No significant heterogeneity was identified in the meta-analysis. Additionally, Egger’s linear regression and Begg’s funnel plot test were used to evaluate publication bias. The shape of the funnel plot did not reveal obvious asymmetry in any of the genotype models of FAS-670A > G gene polymorphism (Table 3).

Sensitivity analysis

The impact of individual studies on the pooled OR was evaluated by sequential omission of each study. The results showed that no individual study significantly affected the pooled OR in all genotype models of FAS-670A > G polymorphism (Fig. 7).

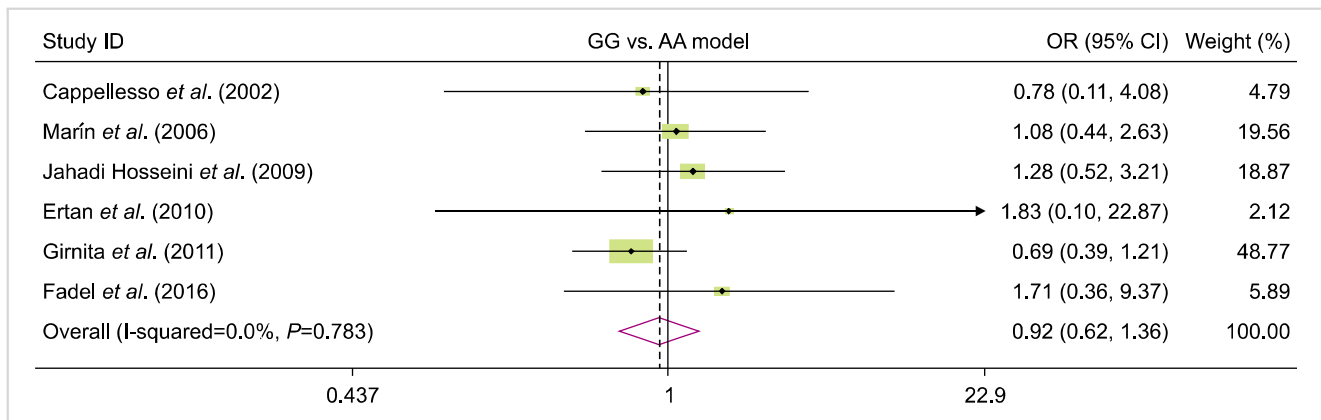


Fig. 5. Forest plot of the association between FAS-670A > G gene single-nucleotide polymorphism and the risk of allograft rejection in the GG vs. AA model.

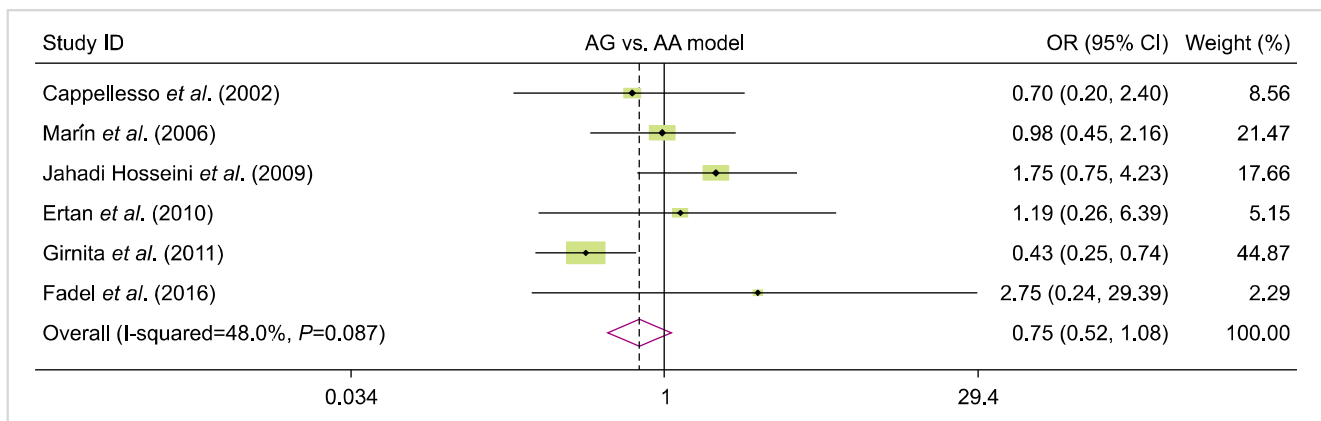
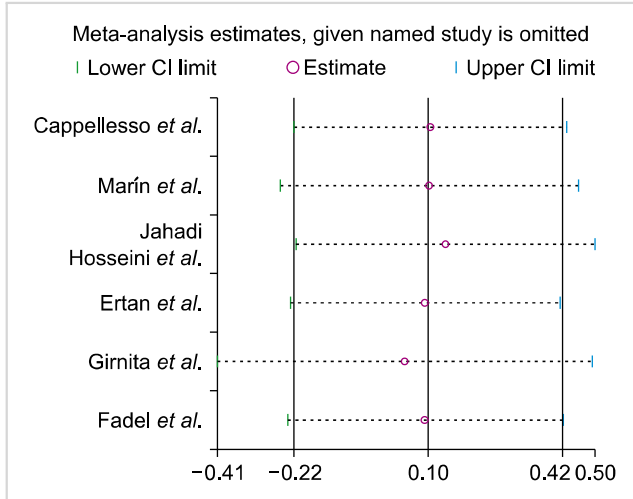


Fig. 6. Forest plot of the association between FAS-670A > G gene single-nucleotide polymorphism and the risk of allograft rejection in the AG vs. AA model.

Table 3. Main results of pooled OR in meta-analysis of FAS 670A/G gene polymorphisms.

	Genetic model	Sample size	Test of association		Test of heterogeneity		Test of publication bias (Begg's test)		Test of publication bias (Egger's test)	
			Case/control	OR	95% CI (P)	I ² (%)	P	Z	P	T
Overall population	Dominant model	277/1001	0.81	0.58–1.12 (0.19)	33.2	0.18	0.94	0.34	1.8	0.14
	Recessive model	277/1001	1.10	0.80–1.53 (0.55)	0	0.99	0.19	0.85	0.14	0.89
	Allelic model	277/1001	0.96	0.79–1.18 (0.7)	0	0.74	1.69	0.09	1.81	0.14
	GG vs. AA	277/1001	0.92	0.62–1.36 (0.66)	0	0.78	0.56	0.57	1.8	0.14
	AG vs. AA	277/1001	0.75	0.52–1.08 (0.12)	48	0.08	0.94	0.34	1.66	1.17
Subgroup analysis										
Caucasians	Dominant model	136/566	1.12	0.71–1.78 (0.62)	0	0.74	0	1	-0.35	0.76
	Recessive model	136/566	1.02	0.63–1.66 (0.93)	0	0.97	1.36	0.17	0.86	0.48
	Allelic model	136/566	1.06	0.80–1.41 (0.67)	0	0.91	0	1	-0.36	0.75
	GG vs. AA	136/566	1.15	0.64–2.06 (0.64)	0	0.94	0.68	0.49	0.05	0.96
	AG vs. AA	136/566	1.15	0.70–1.89 (0.58)	0	0.64	0	1	-0.38	0.74
Children	Dominant model	157/472	0.62	0.40–1.06 (0.07)	37.7	0.20	1.57	0.11	5.15	0.12
	Recessive model	157/472	1.19	0.77–1.84 (0.43)	0	0.96	1.57	0.11	2.79	0.21
	Allelic model	157/472	0.90	0.68–1.17 (0.42)	0	0.43	1.57	0.11	4.79	0.13
	GG vs. AA	157/472	0.79	0.47–1.33 (0.36)	0	0.48	0.52	0.60	2.29	0.21
	AG vs. AA	157/472	0.72	0.27–1.95 (0.52)	40	0.18	1.57	0.11	53.19	0.01
Adults	Dominant model	100/452	1.22	0.72–2.07 (0.47)	0	0.45	1.0	0.31	*	*
	Recessive model	100/452	1.01	0.60–1.69 (0.98)	0	0.76	1.0	0.31	*	*
	Allelic model	100/452	1.09	0.79–1.51 (0.60)	0	0.76	1.0	0.31	*	*
	GG vs. AA	100/452	1.17	0.62–2.22 (0.62)	0	0.79	1.0	0.31	*	*
	AG vs. AA	100/452	1.27	0.71–2.28 (0.41)	0	0.33	1.0	0.31	*	*

*Was not calculable.

**Fig. 7.** Sensitivity analysis to investigate whether FAS-670A/G gene single nucleotide polymorphism contributes to risk for allograft rejection (Recessive model).

DISCUSSION

To date, several individual case-control replication studies have attempted to investigate the association between the FAS-670A >G gene polymorphism and the risk of allograft rejection after organ transplantation. Due to some differences, however, these dispersed investigations have demon-

strated incongruous reports. However, a meta-analysis is a tool that has the potential to solve the problem of inconsistency by removing the confining issues of insufficient statistical power in individual studies. Therefore, to resolve the mentioned confining factors of the FAS-670A >G gene polymorphism, the most recent meta-analysis was conducted to determine a bona fide estimation of the association between the FAS-670A >G gene polymorphism and allograft rejection after organ transplantation. Our findings indicated that FAS-670A >G gene polymorphism was not associated with the risk of allograft rejection after organ transplantation in the overall population. In addition, subgroup analysis according to ethnicity and age showed no significant association between FAS-670A >G gene polymorphism and the risk of allograft rejection.

Programmed cell death (apoptosis) is an essential physiological mechanism involved in the development and homeostasis of the immune system. The main mechanism of apoptosis is the extrinsic pathway involving surface molecules known as “death receptors” and their ligands, the best-characterized death receptor including FAS [29]. Engagement of the T-cell receptor/CD3 complex upregulates CD95 expression and induces CD95L expression through antigen stimulation. Through these cell surface molecules, activated T cells undergo activation-induced cell death, which is principally mediated by the CD95/CD95L system to develop spontaneous tolerance to the allograft [30, 31]. Consistent with these theoretical points, Boix *et al.* [32] and Mancebo

et al. [33] reported that patients who experienced liver and kidney rejection, respectively, had higher levels of CD95 in both CD4 and CD8 T cells within the first month after transplantation. In addition, Wang *et al.* [34] observed that CD95 expression on CD3+ T cells in liver transplantation rejection compared to that in stable recipients or healthy individuals was significantly increased. Therefore, in patients experiencing allograft rejection, we cannot deny the role of allograft infiltrated T cells with overexpressed CD95 that induces apoptosis through the establishment of the CD95-CD95L complex and stimulates the rejection process.

FAS-670A>G and FAS-1377G/A are two important SNPs that have been reported in the FAS promoter region [14]. The first one with the G variant disrupts the interferon-gamma binding site for the transcription factor STAT1. FAS was significantly upregulated by interferon-gamma in several reports [35-37]. Therefore, healthy subjects who are homozygous for the 670 A/A major allele have higher levels of FAS expression than those who are homozygous for the 670 G/G variant [14], which in turn could decrease their capability to be depleted by apoptosis. Moreover, FAS and FASL may occur as cell surface proteins or in soluble forms [38]. Various isoforms of soluble FAS (sFAS) are generated by alternative splicing of FAS, and the most frequent sFAS isoform results from the deletion of exon 6, which encodes the last five amino acids of the extracellular domain and 16 of the 17 amino acids of the transmembrane domain, which is thought to prevent the function of FAS [39, 40].

According to this mechanism, FAS-670A>G gene polymorphism probably influences the risk of organ rejection. The FAS/FASL system plays a significant role in progressive renal disease and organ rejection in liver [41], cardiac [39], and renal transplantation [42]. For example, liver transplant recipients carrying the FAS-670AA genotype displayed significantly lower graft survival rates than those carrying the AG genotype [25]. In addition, low levels of soluble FAS are present in the serum of normal individuals, and enhanced serum concentrations of sFAS have been reported in bone marrow transplantation [43], chronic kidney allograft rejection [44], and acute liver allograft rejection [45]. Wang *et al.* [46] reported that overexpression of sFAS in allograft endothelium reduced vascular cell apoptosis, infiltration of the arterial wall by leukocytes, and disruption of the media layer in a rat aortic allograft model of chronic rejection. Further investigation revealed that FAS-670A>G gene polymorphism could regulate sFAS expression, and normal patients carrying the FAS-A/A genotype produced markedly higher levels of sFAS than those carrying the sFAS-G/G genotype [47].

This meta-analysis has some limitations and limitations. First, the analysis was based on a crude estimation of FAS-670A>G gene polymorphism association with allograft rejection, regardless of the effect of confounding factors such as age, sex, environmental factors, and the contribution of other genes in LD with the FAS gene. Second, because of the limited number of studies on other SNPs, we did not analyze other SNPs of the FAS gene that could contribute

to the understanding of FAS SNP involvement in allograft rejection. Third, we were unable to perform subgroup analyses according to sex and clinical or environmental variables. Fourth, although we used a comprehensive search strategy, the number of eligible studies for quantitative analysis was low, and we strongly suggest that our findings should be interpreted with caution. Fifth, since there was no meta-analysis on FAS gene polymorphism and its association with age and ethnicity, we could not compare our findings.

In conclusion, the present meta-analysis demonstrated that there was no significant independent association between Fas-670A>G gene polymorphism and the risk of allograft rejection after organ transplantation. To reach a definitive conclusion, more well-designed studies with larger samples are necessary to clarify the role of this polymorphism in allograft rejection risk.

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Authors' Disclosures of Potential Conflicts of Interest

No potential conflicts of interest relevant to this article were reported.

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