



Draft Genome Sequences of Four Bacterial Strains of Heterotrophic *Alteromonas macleodii* and *Marinobacter*, Isolated from a Nonaxenic Culture of Two Marine *Synechococcus* Strains

 Patricia Arias-Orozco,^a  Yunhai Yi,^{a,b}  Oscar P. Kuipers^a

^aDepartment of Molecular Genetics, Groningen Biomolecular Sciences and Biotechnology Institute, University of Groningen, Groningen, The Netherlands

^bBGI Education Center, University of Chinese Academy of Sciences, Shenzhen, China

Patricia Arias-Orozco and Yunhai Yi contributed equally to this work. Author order was determined based on the extent of the contribution and the responsibility for the data in the manuscript. Patricia Arias-Orozco requested the strain, isolated the DNA, helped with annotation, and wrote the first draft of the manuscript. Yunhai Yi was involved in annotation and bioinformatic analyses, submitting GEO data, making a results table, and writing parts of the manuscript.

ABSTRACT Species of the *Alteromonas* and *Marinobacter* genera are heterotrophic *Gammaproteobacteria* that are part of the marine microbial ecosystem. In this study, four strains were isolated from two nonaxenic *Synechococcus* cultures and were sequenced. Few studies of these two genera have been reported. Therefore, genomic data of *Alteromonadaceae* are valuable for the study of heterotroph-phototroph dynamics in marine bacterial communities.

Marine interactions between heterotrophic and phototrophic organisms play an essential role in the nutrient cycle of marine microbial ecosystems (1). *Synechococcus* cultures are often found together with associated heterotrophic bacteria. It was previously observed that these nonaxenic cultures are more stable, possibly due to their mutually dependent relationship (2). Understanding which interactions can induce the expression of specific secondary metabolites can help in elucidating their unknown function (3).

In this work, four strains were isolated from two laboratory stocks, marine *Synechococcus* sp. strain CC9311 and *Synechococcus* sp. strain WH8102, that were a kind gift from the Department of Molecular Sciences at Macquarie University in Australia. For the isolation of the heterotrophic bacteria, both nonaxenic *Synechococcus* cultures were spread onto Difco marine agar plates and incubated at room temperature. Then, a single colony of each strain growing in Difco marine agar 2216 was selected and grown in 5 ml of marine broth 2216 without shaking and incubated for 24 h (*Alteromonas* sp.) or 48 h (*Marinobacter* sp.) at 25°C. Cells from the cultures were harvested by centrifugation at 12,000 rpm for 3 min in a Microfuge 16 centrifuge (Beckman Coulter, Woerden, The Netherlands). Genomic DNA was extracted using a GenElute bacterial genomic DNA kit (Sigma-Aldrich, Munich, Germany) according to the manufacturer's instructions. To determine the genus of the isolated strains, we amplified and sequenced the 16S rRNA genes. According to the sequencing results, both *Synechococcus* strains coexist with one *Alteromonas* sp. and one *Marinobacter* sp.

The genomes of the four isolated strains were paired-end sequenced by the Beijing Genomics Institute European Genome Center in Denmark on a BGISEQ-500 platform. Whole-genome sequencing libraries were constructed with the MGIEasy universal DNA library prep set (MGI Tech Co., Ltd., Shenzhen, China), which is specifically designed for MGI high-throughput sequencing platform series. A total of 45 million paired-end clean reads (150 bp) were acquired, after adaptor sequences, and contamination and low-quality reads were removed from the raw reads using Trimmomatic version 0.38 (4). FastQC version 0.11.9 (5) was used to examine the quality of the reads. Subsequently, the PATRIC (6)

Citation Arias-Orozco P, Yi Y, Kuipers OP. 2021. Draft genome sequences of four bacterial strains of heterotrophic *Alteromonas macleodii* and *Marinobacter*, isolated from a nonaxenic culture of two marine *Synechococcus* strains. *Microbiol Resour Announc* 10:e00116-21. <https://doi.org/10.1128/MRA.00116-21>.

Editor Frank J. Stewart, Georgia Institute of Technology

Copyright © 2021 Arias-Orozco et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Oscar P. Kuipers, o.p.kuipers@rug.nl.

Received 2 February 2021

Accepted 23 April 2021

Published 13 May 2021

TABLE 1 Genome features and accession numbers for the four *Alteromonadaceae* strains

Characteristic	Data for:			
	<i>Marinobacter</i> sp. MC3	<i>Marinobacter</i> sp. MW3	<i>Alteromonas macleodii</i> MC7	<i>Alteromonas macleodii</i> MW7
Genome size (bp)	4,732,670	4,732,670	4,771,423	4,771,559
Coverage (×)	354	352	345	352
No. of reads	11,323,466	11,316,028	11,440,920	11,459,196
No. of contigs	53	54	25	25
N_{50} (bp)	308,870	308,871	406,450	406,450
L_{50}	5	6	4	4
GC content (%)	57.04	57.04	44.64	44.64
No. of genes	4,426	4,426	4,112	4,110
No. of coding sequences	4,332	4,332	4,009	4,008
No. of tRNAs	48	48	66	66
No. of rRNAs	3	3	6	5
GenBank accession no.	JAEMVF000000000	JAEMVG000000000	JAEMVH000000000	JAEMVI000000000
BioSample accession no.	SAMN17124544	SAMN17124543	SAMN17124542	SAMN17124541
SRA accession no.	SRR13297789	SRR13297790	SRR13297791	SRR13297792

website server was employed to perform comprehensive genome analysis. To assemble the short reads, we used Unicycler version 0.4.8 (7) integrated with SPAdes version 3.12.0 (8). The assembled draft genome sequences were evaluated with QUAST version 5.0.2 (9). Genomes were also annotated in the Prokaryotic Genome Annotation Pipeline (PGAP) (10). Default parameters were used for all software unless otherwise noted. The coverages of the four sequenced genomes were all around 350×. The assembly and annotation statistics are described in Table 1. To identify the species, we calculated the average nucleotide identity (ANI, >95% for the same species) and digital DNA-DNA hybridization (dDDH; >70% for the same species) using JSpeciesWS (11) and TYGS (12), respectively. The *Alteromonas* strain was confirmed to be *A. macleodii* (ANI of 98.21% and dDDH of 89.3% compared to *Alteromonas macleodii* ATCC 27126), while the *Marinobacter* strain could not be specified (ANI of 92.69% and dDDH of 68.2% compared to *Marinobacter maroccanus*). Further analysis of these genomes is under way in order to investigate their specialties as symbionts of the marine cyanobacterium *Synechococcus*.

Data availability. The genome sequences of the two *Marinobacter* strains and the two *Alteromonas macleodii* strains have been submitted to NCBI under BioProject PRJNA686772; see details in Table 1.

ACKNOWLEDGMENTS

We thank Deepa Varkey from the Department of Molecular Sciences, Macquarie University, for her help with the *Synechococcus* strains. We thank Anne de Jong (Department of Molecular Genetics, University of Groningen) for advice during genome sequencing and analysis.

P.A.-O. and O.P.K. were supported by the European Union's Horizon 2020 Research and Innovation Program under the Marie Skłodowska-Curie grant agreement (ALERT Program, grant 713482). Y.Y. was supported by the Chinese Scholarship Council.

REFERENCES

- Zheng Q, Wang Y, Xie R, Lang AS, Liu Y, Lu J, Zhang X, Sun J, Suttle CA, Jiao N. 2017. Dynamics of heterotrophic bacterial assemblages within *Synechococcus* cultures. *Appl Environ Microbiol* 84:e01517-17. <https://doi.org/10.1128/AEM.01517-17>.
- Zheng Q, Wang Y, Lu J, Lin W, Chen F, Jiao N. 2020. Metagenomic and metaproteomic insights into photoautotrophic and heterotrophic interactions in a *Synechococcus* culture. *mBio* 11:e03261-19. <https://doi.org/10.1128/mBio.03261-19>.
- Aharonovich D, Sher D. 2016. Transcriptional response of *Prochlorococcus* to co-culture with a marine *Alteromonas*: differences between strains and the involvement of putative infochemicals. *ISME J* 10:2892–2906. <https://doi.org/10.1038/ismej.2016.70>.
- Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics* 30:2114–2120. <https://doi.org/10.1093/bioinformatics/btu170>.
- Andrews S. 2010. FastQC: a quality control tool for high throughput sequence data. <https://www.bioinformatics.babraham.ac.uk/projects/fastqc/>.
- Davis JJ, Wattam AR, Aziz RK, Brettin T, Butler R, Butler RM, Chlenski P, Conrad N, Dickerman A, Dietrich EM, Gabbard JL, Gerdes S, Guard A, Kenyon RW, Machi D, Mao C, Murphy-Olson D, Nguyen M, Nordberg EK, Olsen GJ, Olson RD, Overbeek JC, Overbeek R, Parrello B, Pusch GD, Shukla M, Thomas C, VanOeffelen M, Vonstein V, Warren AS, Xia F, Xie D, Yoo H, Stevens R. 2020. The PATRIC Bioinformatics Resource Center: expanding data and analysis capabilities. *Nucleic Acids Res* 48:D606–D612. <https://doi.org/10.1093/nar/gkz943>.

7. Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Unicycler: resolving bacterial genome assemblies from short and long sequencing reads. *PLoS Comput Biol* 13:e1005595. <https://doi.org/10.1371/journal.pcbi.1005595>.
8. Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <https://doi.org/10.1089/cmb.2012.0021>.
9. Gurevich A, Saveliev V, Vyahhi N, Tesler G. 2013. QUAST: quality assessment tool for genome assemblies. *Bioinformatics* 29:1072–1075. <https://doi.org/10.1093/bioinformatics/btt086>.
10. Li W, O'Neill KR, Haft DH, DiCuccio M, Chetvernin V, Badretdin A, Coulouris G, Chitsaz F, Derbyshire MK, Durkin AS, Gonzales NR, Gwadz M, Lanczycki CJ, Song JS, Thanki N, Wang J, Yamashita RA, Yang M, Zheng C, Marchler-Bauer A, Thibaud-Nissen F. 2021. RefSeq: expanding the Prokaryotic Genome Annotation Pipeline reach with protein family model curation. *Nucleic Acids Res* 49:D1020–D1028. <https://doi.org/10.1093/nar/gkaa1105>.
11. Richter M, Rosselló-Móra R, Oliver Glöckner F, Peplies J. 2016. JSpeciesWS: a Web server for prokaryotic species circumscription based on pairwise genome comparison. *Bioinformatics* 32:929–931. <https://doi.org/10.1093/bioinformatics/btv681>.
12. Meier-Kolthoff JP, Göker M. 2019. TYGS is an automated high-throughput platform for state-of-the-art genome-based taxonomy. *Nat Commun* 10:2182. <https://doi.org/10.1038/s41467-019-10210-3>.