

# Resistance to cellular HIV infection

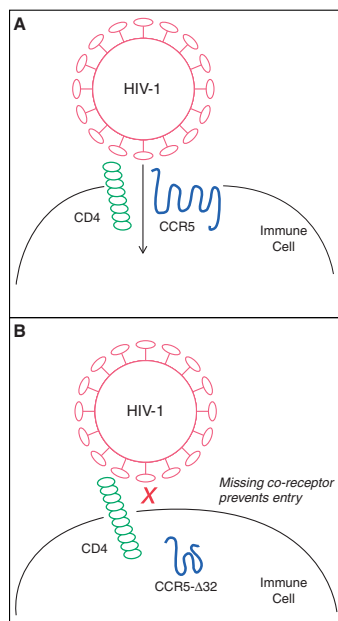
Alice M. Clomegah<sup>1</sup> and Stephen J. Chapman<sup>2</sup>

<sup>1</sup>NYU College of Global Public Health New York, NY 10003 USA; <sup>2</sup>Oxford University Hospitals, Oxford OX3 7LE, UK



## GENETIC RESISTANCE TO HIV

Infection with human immunodeficiency virus-1 (HIV-1) remains a major cause of premature death worldwide. In order to enter and infect immune cells, HIV-1 binds to cell surface receptors including the CCR5 chemokine receptor (Fig. 1A). A well-described functional polymorphism in the CCR5 gene comprises a 32-bp deletion (called CCR5-Δ32) which results in a lack of the last three transmembrane domains of the CCR5 protein [1]. The mutated protein is contained in the cytoplasm, resulting in a complete lack of CCR5 cell surface receptor. In the absence of this co-receptor binding site, HIV-1 is unable to enter the cell (Fig. 1B); individuals homozygous for CCR5-Δ32 display complete resistance to HIV-1, whereas heterozygotes have a delayed onset of AIDS [1].



**Figure 1.** HIV-1 entry into immune cells

## EVOLUTIONARY PERSPECTIVES

Chemokines and their receptors play a central role in the trafficking and activation of lymphocytes, but perhaps surprisingly there are no apparent pathological consequences of CCR5-Δ32 homozygosity. Intriguingly, this variant displays marked population differentiation which would not be explained by the very recent emergence of HIV-1 in sub-Saharan Africa: the CCR5-Δ32 allele is highly prevalent in Europeans (frequency up to 14%), while very rare or absent among African and Asian populations [2].

Early studies suggested that the CCR5-Δ32 allele arose relatively recently (~700 years ago) and was subject to strong positive selection [2]. Infection with *Yersinia pestis*—the cause of the bubonic plague in Europe during this time period—or smallpox were suggested as potential selective factors favoring CCR5-Δ32 [2]. HIV and smallpox both cause cellular immune dysfunction and both enter leukocytes using chemokine receptors. If exposure to smallpox provided selective pressure favoring CCR5-Δ32, it is plausible that exposed populations now have an evolutionary advantage in facing HIV [3]. Recently, however, high-density genetic maps have questioned the evidence for a recent origin of CCR5-Δ32, suggesting instead that this allele may have arisen ~5000 years ago [4], under neutral evolution, or subject to an ancient, currently unknown positive selective pressure.

## REFERENCES

- Chapman SJ, Hill AVS. Human genetic susceptibility to infectious disease. *Nat Rev Genet* 2012;**13**:175–88.
- Stephen JC, Reich DE, Goldstein DB *et al*. Dating the origin of the CCR5-Δ32 AIDS-resistance allele by the coalescence of haplotypes. *Am J Hum Genet* 1998;**62**:1507–15.

## FUTURE IMPLICATIONS

Antagonism of CCR5 interferes with HIV-1 cell entry, and indeed the CCR5 antagonist drug maraviroc is approved by the FDA for treatment of exclusively CCR5-tropic viral strains of HIV-1 [5]. The clinical relevance of CCR5-Δ32 is further illustrated by a case of long-term HIV control following stem-cell transplantation from a CCR5-Δ32 homozygous donor [6], although viral escape through chemokine receptors other than CCR5 may limit the success of this approach [7]. Of broader relevance, development of a therapy that eliminates the transmembrane portion of the CCR5 protein to mimic the advantage conferred by the natural CCR5-Δ32 variant might constitute a functional HIV-1 cure. A 'gene editing' approach using infusion of autologous CD4 T cells which have been modified by zinc finger nucleases to disrupt CCR5 (mimicking CCR5-Δ32) appears safe and confers partial disease resistance [8].

- Galvani AP, Slatkin M. Evaluating plague and smallpox as historical selective pressures for the CCR5-Δ32 HIV-resistance allele. *Proc Natl Acad Sci U S A* 2003;**100**:15276–9.
- Sabeti PC, Walsh E, Schaffner SF *et al*. The case for selection at CCR5-D32. *PLoS Biol* 2005;**3**:e378.
- Wilkin TJ, Gulick RM. CCR5 Antagonism in HIV infection: current concepts and future opportunities. *Annu Rev Med* 2012;**63**:81–93.
- Hütter G, Nowak D, Mossner M *et al*. Long-term control of HIV by CCR5 Delta32/Delta32 stem-cell transplantation. *N Engl J Med* 2009;**360**:692–8.
- Kordelas L, Verheyen J, Esser S. Shift of HIV tropism in stem-cell transplantation with CCR5 delta32 mutation. *N Engl J Med* 2014;**371**:880–2.
- Tebas P, Stein D, Tang WW *et al*. Gene editing of CCR5 in autologous CD4 T cells of persons infected with HIV. *N Engl J Med* 2014;**370**:901–10.