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Original article

# One-step preparation, characterization, and anticancer potential of ZnFe<sub>2</sub>O<sub>4</sub>/RGO nanocomposites

ZabnAllah M. Alaizeri\*, Hisham A. Alhadlaq, Saad Aldawood, Mohd Javed Akhtar, Maqusood Ahamed\*

Department of Physics and Astronomy, College of Science, King Saud University, Riyadh 11451, Saudi Arabia

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## ABSTRACT

Zinc ferrite nanoparticles (ZnFe<sub>2</sub>O<sub>4</sub> NPs) have attracted extensive attention for their diverse applications including sensing, waste-water treatment, and biomedicine. The novelty of the present work is the fabrication of ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs by using a one-step hydrothermal process to assess the influence of RGO doping on the physicochemical properties and anticancer efficacy of ZnFe<sub>2</sub>O<sub>4</sub> NPs. X-ray diffraction (XRD), Scanning electron microscopy (SEM), Energy-dispersive X-ray (EDX), X-ray photoelectron spectroscopy (XPS), Fourier-transform infrared spectroscopy (FTIR), UV-vis spectroscopy, and Photoluminescence (PL) spectroscopy were employed to characterize prepared pure ZnFe<sub>2</sub>O<sub>4</sub> NPs and ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs. XRD results showed that the synthesized samples have high crystallinity. Furthermore, the average crystal sizes of ZnFe<sub>2</sub>O<sub>4</sub> nanoparticles (NPs) and ZnFe<sub>2</sub>O<sub>4</sub>/RGO nanocomposites (NCs) were 51.08 nm and 54.36 nm, respectively. SEM images revealed that pure ZnFe<sub>2</sub>O<sub>4</sub> NPs were spherical in shape with uniformly loaded on the surface of the RGO nanosheet. XPS and EDX analysis confirmed the elemental compositions of ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs. Elemental mapping of SEM shows that the elemental compositions (Zn, Fe, O, and C) were homogeneously distributed in ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs. The intensity of FT-IR spectra depicted that pure ZnFe<sub>2</sub>O<sub>4</sub> NPs were successfully anchored into the RGO nanosheet. An optical study suggested that the band gap energy of ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs (1.61 eV) was lower than that of pure ZnFe<sub>2</sub>O<sub>4</sub> NPs (1.96 eV). PL spectra indicated that the recombination rate of the ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs was lower than ZnFe<sub>2</sub>O<sub>4</sub> NPs. MTT assay was used to evaluate the anticancer performance of ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs and pure ZnFe<sub>2</sub>O<sub>4</sub> NPs against human cancer cells. In vitro study indicates that ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs have higher anticancer activity against human breast (MCF-7) and lung (A549) cancer cells as compared to pure form ZnFe<sub>2</sub>O<sub>4</sub> NPs. This work suggests that RGO doping enhances the anticancer activity of ZnFe<sub>2</sub>O<sub>4</sub> NPs by tuning its optical behavior. This study warrants future research on potential therapeutic applications of these types of nanocomposites.

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## 1. Introduction

Cancer remains one of the most significant health challenges worldwide, necessitating the continuous exploration of novel therapeutic strategies. However, it is recently a complex and deadly disease that affects millions of people worldwide (Bray et al., 2018). Despite progress in cancer treatment, there is an urgent requirement

for the creation of new therapeutic approaches that are both efficient and safe (Shyamala and Gomathi Devi, 2020; Tabrez et al., 2022a; Zughaihi et al., 2022). One promising approach is applied of nanocomposites (NCs) which have shown great potential in enhancing anticancer performance (Ahamed et al., 2023a; Saranya et al., 2023). Nowadays, some challenges of cancer treatment are the synthesis and characterization of nanocomposites with optimal physicochemical properties (Biswal and Yusoff, 2017).

To solve these challenges, previous studies have focused on the synthesis of nanocomposites (NCs) using a variety of methods (Kokila et al., 2022; Saranya et al., 2023; Somwanshi et al., 2020). Among the various types of NCs, metal oxide/reduced graphene oxide (RGO) nanocomposites (NCs) are being applied in potential applications owing to their outstanding properties such as high surface area and good biocompatibility (Aarti et al., 2022). For example, ZnFe<sub>2</sub>O<sub>4</sub> NPs have been combined with reduced graphene

\* Corresponding authors.

E-mail addresses: [zalaizeri@ksu.edu.sa](mailto:zalaizeri@ksu.edu.sa) (ZabnAllah M. Alaizeri), [mahamed@ksu.edu.sa](mailto:mahamed@ksu.edu.sa) (M. Ahamed).

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oxide (RGO) as NCs to enhance the properties of  $\text{ZnFe}_2\text{O}_4$  NPs, which can be performed in potential applications (Khadgi et al., 2016; Wu et al., 2015). However, the nanocomposites (NCs) have been prepared using several approaches including, Physical Vapor Deposition (PVD) (Nguyen et al., 2019), Chemical Vapor Deposition (CVD) (Nguyen et al., 2019), green approach (Prasad et al., 2017), sol-gel method (Banerjee et al., 2018), and hydrothermal/solvothermal method (Khadgi et al., 2016). In another instance, (Ahamed et al., 2022) synthesized  $\text{ZnO/RGO}$  NCs by a green approach. This study revealed that prepared  $\text{ZnO/RGO}$  NCs have been shown to enhance the therapeutic efficacy of doxorubicin in breast cancer cells.

Interestingly, zinc ferrite ( $\text{ZnFe}_2\text{O}_4$ ) NPs have been extensively studied in potential biomedical applications due to their unique physicochemical properties (Aarti et al., 2022). Krishnan et al (Krishnan et al., 2021) prepared  $\text{ZnFe}_2\text{O}_4/\text{RGO}$  NCs via a facile green approach. This study showed that the  $\text{ZnFe}_2\text{O}_4/\text{RGO}$  NCs have a high cytotoxicity effect against human lung cancer (A549) cells. In another study,  $\text{Au/RGO}$  NCs exhibited high cytotoxicity against human colon cancer cell lines (HT-29 and SW-948), while it has high cytocompatibility toward normal human colon (CCD-841) (Al-Ani et al., 2019). Similarly,  $\text{TiO}_2/\text{RGO}$  NCs have been applied as fluorescent probes for the imaging and detection of cancer cells (Umekar et al., 2021). Lee et al (Lee et al., 2015) showed that the  $\text{Fe}_3\text{O}_4/\text{RGO}$  NCs exhibit as contrast agents for magnetic resonance imaging (MRI).

The present study aimed to investigate the novel application of RGO to improve the physicochemical properties of  $\text{ZnFe}_2\text{O}_4$  NPs and their potential in anticancer therapy. Pure  $\text{ZnFe}_2\text{O}_4$  NPs and  $\text{ZnFe}_2\text{O}_4/\text{RGO}$  NCs were successfully prepared by using One-step hydrothermal synthesis. X-ray diffraction (XRD), scanning electron microscopy (SEM), energy-dispersive X-ray (EDX), X-ray photoelectron (XPS), Fourier Infrared (FTIR), UV-vis spectroscopy, and photoluminescence (PL) spectrometer were applied to characterize the physicochemical properties. Cytotoxicity study showed that the  $\text{ZnFe}_2\text{O}_4/\text{RGO}$  NCs induced high cytotoxicity against MCF-7 and A549 cancer cells compared with pure  $\text{ZnFe}_2\text{O}_4$  NPs.

## 2. Experimental part

### 2.1. Chemicals

Iron (III) nitrate nonahydrate ( $\text{Fe}(\text{NO}_3)_3 \cdot 9\text{H}_2\text{O}$ ), Zinc nitrate hexahydrate ( $\text{Zn}(\text{NO}_3)_2 \cdot 6\text{H}_2\text{O}$ ), Graphene oxide (GO), Sodium hydrox-

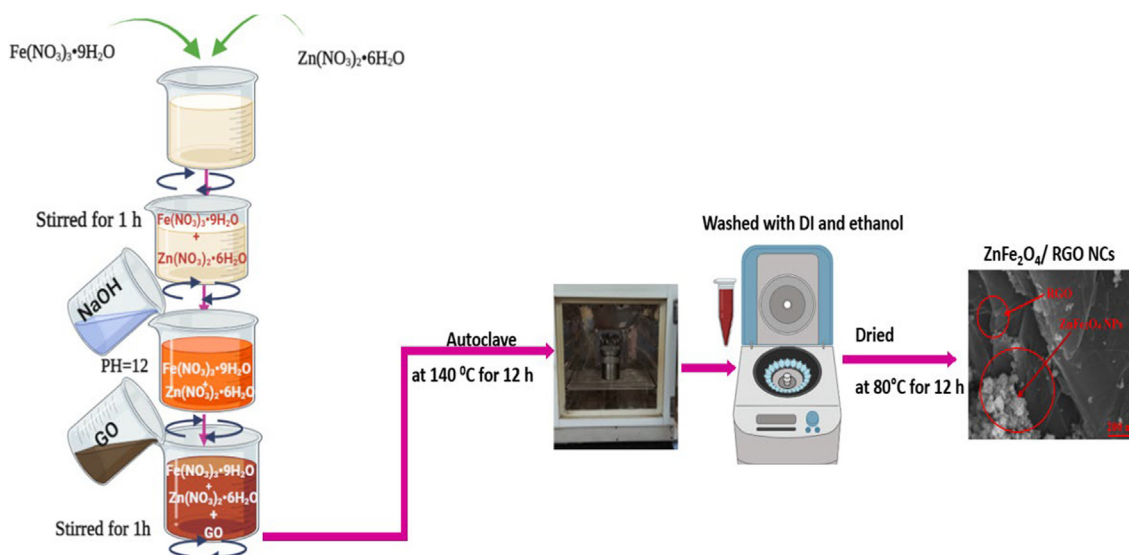
ide (NaOH), and Ethylene glycol (EG) were supplied from Sigma Aldrich. The preparation was carried out using distilled water (DW) as the medium. These chemicals were used as received without undergoing any additional purification.

### 2.2. Synthesis of pure $\text{ZnFe}_2\text{O}_4$ NPs and $\text{ZnFe}_2\text{O}_4/\text{RGO}$ NCs

Pure  $\text{ZnFe}_2\text{O}_4$  NPs and  $\text{ZnFe}_2\text{O}_4/\text{RGO}$  NCs were successfully prepared by one-step hydrothermal approach (Behera et al., 2019). The 1 g of  $\text{Fe}(\text{NO}_3)_3 \cdot 9\text{H}_2\text{O}$  and 0.5 g of  $\text{Zn}(\text{NO}_3)_2 \cdot 6\text{H}_2\text{O}$  were dissolved in 50 ml of EG under continuous stirring for 1 h. Then, the 800 mg of GO was also dispersed in DI water and added to the above solution under sonication for 30 min. Next, 10 ml of NaOH (2 M) was gradually added drop into the mixture to reach  $\text{pH} = 12$ . The mixture solution was stirred for 1 h and further transferred into a Teflon-lined autoclave, and heated at  $140^\circ\text{C}$  for 12 h. After that, the precipitate was collected by centrifugation and washed several times with DI water and ethanol. Finally, this precipitate was dried in the oven at  $80^\circ\text{C}$  for 12 h. The dried precipitate was crushed into powder using a mortar and pestle. Pure  $\text{ZnFe}_2\text{O}_4$  NPs were synthesized using the same above procedures without GO. A similar procedure was further used for the preparation of RGO without mixing ( $\text{Fe}(\text{NO}_3)_3 \cdot 9\text{H}_2\text{O}$ ,  $\text{Zn}(\text{NO}_3)_2 \cdot 6\text{H}_2\text{O}$ ). The synthesis protocol of  $\text{ZnFe}_2\text{O}_4/\text{RGO}$  NCs was depicted in Scheme 1.

### 2.3. Characterization

Different analytical techniques were employed to characterize the prepared  $\text{ZnFe}_2\text{O}_4$  NPs and  $\text{ZnFe}_2\text{O}_4/\text{RGO}$  NCs. X-ray diffraction (XRD) (PanAnalytic X'Pert Pro, Malvern Instruments, Malvern, UK) was used to examine their crystal structure and phase purity. Morphological analysis of prepared samples was carried out using scanning electron microscopy (SEM) (SEM, JSM-7600F, JEOL, Inc. Tokyo, Japan). Energy-dispersive X-ray spectroscopy (EDX) and X-ray photoelectron spectroscopy (XPS) (PHI-5300 ESCA PerkinElmer, Boston, MA, USA) were utilized to determine the chemical composition and oxidation states of the prepared samples. Fourier transform infrared spectroscopy (FTIR) was used to study their functional groups. To investigate their optical properties, UV-Vis (Hitachi U-2600) and photoluminescence (PL) Hitachi F-4600) spectroscopy were employed to determine the band gap energy of the NPs and NCs.



Scheme 1. Synthesis protocol of  $\text{ZnFe}_2\text{O}_4/\text{RGO}$  NCs.

## 2.4. Cell culture

Human breast cancer (MCF-7) and human lung cancer (A549) cells were supplied from America (ATTC, Manassas, WV, USA). These cell lines were cultured in Dulbecco's Modified Eagle's Medium (DMEM) with high glucose, along with 10% fetal bovine serum (FBS), 1% L-glutamine, and 1% penicillin-streptomycin supplementation. The cells were then incubated in a humidified environment at 37 °C with 5% CO<sub>2</sub>.

## 2.5. Cytotoxicity assay

The MTT assay was utilized to assess the cytotoxicity of the prepared samples. To further implement the MTT assay on pure ZnFe<sub>2</sub>O<sub>4</sub> NPs and ZnFe<sub>2</sub>O<sub>4</sub>/rGO NCs, the following procedures were carried out. Initially, MCF-7 and A549 cells were seeded at a density of 1×10<sup>4</sup> cells per well in 96-well plates and then incubated at 37 °C with 5% CO<sub>2</sub> for 24 h. Following that, the cells were exposed to various concentrations (ranging from 1.5 µg/ml to 200 µg/ml) of the synthesized samples in an incubator at 37 °C with 5% CO<sub>2</sub> for 24 h. Subsequently, 100 µL of MTT solution was added to each well and then incubated for 3 h. Finally, 100 µL of DMSO was added to dissolve the formazan crystals in each well. The absorbance of the final solution was measured at 570 nm utilizing a microplate reader.

## 2.6. Statistical analysis

The analysis of biological data was carried out using one-way ANOVA and Dunnett's multiple comparison test. Statistical significance was further  $p < 0.05$ .

## 3. Results

### 3.1. XRD study

Fig. 1 depicts the XRD spectra of RGO, ZnFe<sub>2</sub>O<sub>4</sub> NPs, and ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs. XRD spectra of RGO (Fig. 1a) showed that the RGO has a high peak at 26° and 43.02° which corresponds to (002) and (102) planes. It can be seen in Fig. 1b that the diffraction peaks of the ZnFe<sub>2</sub>O<sub>4</sub> NPs at 2θ values of 17.29, 35°, 42.64°, 52.90°, 56.50°, 62.12°, and 73.40° correspond to the (220), (311), (400), (422), (511), (440), and (533) planes of the spinel structure of ZnFe<sub>2</sub>O<sub>4</sub> (JCPDS card no. 01-79-1905). Fig. 1c showed that the

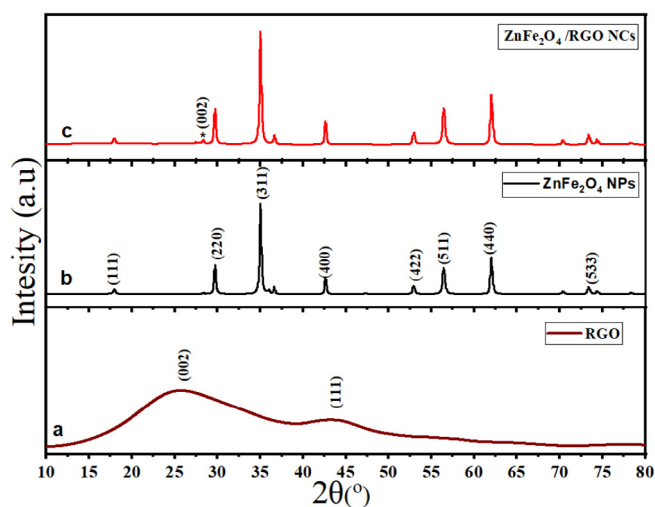


Fig. 1. XRD spectra of RGO (a), ZnFe<sub>2</sub>O<sub>4</sub> NPs (b), and ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs (c).

diffraction peaks were assigned at 2θ values of 30.02°, 35.48°, 43.24°, 53.64°, 57.00°, 62.72°, and 74.00° correspond to the (220), (311), (400), (422), (511), (440), and (533) crystal planes respectively. Furthermore, it was observed that the average crystal sizes of ZnFe<sub>2</sub>O<sub>4</sub> NPs and ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs were 51.08 nm and 54.36 nm, respectively, which were calculated by the Scherrer equation.

### 3.2. SEM analysis

SEM and EDX analysis of the synthesized samples provide valuable information about their morphology, structure, and elemental composition. The SEM images and EDX data of pure ZnFe<sub>2</sub>O<sub>4</sub> NPs and ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs were illustrated in Fig. 2. However, the SEM image of ZnFe<sub>2</sub>O<sub>4</sub> NPs displayed uniform distributions of spherical particles with agglomerations. The EDX spectra (Fig. 2b) of ZnFe<sub>2</sub>O<sub>4</sub> NPs confirmed the presence of zinc (Zn), iron (Fe), and oxygen (O) elements. Moreover, SEM images of ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs (Fig. 2c) showed that the presence of ZnFe<sub>2</sub>O<sub>4</sub> NPs was successfully anchored onto the RGO nanosheet. ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs were prepared by integrating ZnFe<sub>2</sub>O<sub>4</sub> NPs onto the RGO nanosheet. Furthermore, the elemental composition (Zn, Fe, O, and C) of both ZnFe<sub>2</sub>O<sub>4</sub> NPs and RGO were further confirmed in EDX spectra (Fig. 2d). Fig. 3b-e demonstrated the SEM elemental mapping of ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs, which confirmed the uniform distribution of all elements (Zn, Fe, O, and C) in the synthesized samples.

### 3.3. XPS study

Fig. 4 shows the full scan of XPS spectra of ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs, with high-resolution spectra of Zn 2p, Fe 2p, O1s, and C1s. The binding energy (B.E) of Zn in the ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs was shown in the high-resolution XPS spectra of Zn 2P (Fig. 4 b). As shown in Fig. 4b, the Zn 2P peaks were observed at 1021.68 eV and 1045 eV for Zn 2P<sub>3/2</sub> and Zn 2P<sub>1/2</sub>, respectively (Lai et al., 2021). In Fig. 4 c, the high-resolution XPS spectra of Fe 2P display peaks at 711.31 eV and 725.32 eV for the tetrahedral Fe 2p<sub>3/2</sub> and octahedral Fe 2p<sub>1/2</sub>, respectively. The peaks of C 1 s (Fig. 4d) were detected at 288.01 eV, 286.43 eV, 284.84 eV, and 283.57 eV, which corresponded to the C = O bond, C-O bond (epoxy and hydroxyl), C-C bond, and C = C bond, respectively. Similarly, Fig. 4e displays the high-resolution XPS spectra of O 1 s, which reveals a peak at 529.3 due to the interaction between Fe-O of the ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs. In addition, the peak located at 531.1 eV indicates the presence of O-C-O bonding, while the peak at 532.2 eV indicates the presence of Fe-C-O bonding.

### 3.4. FTIR study

The functional group of ZnFe<sub>2</sub>O<sub>4</sub> NPs and ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs are shown in FT-IR spectra (Fig. 5). It can be seen in Fig that the ZnFe<sub>2</sub>O<sub>4</sub> NPs exhibit absorption peaks at 558.82 and 402.10 cm<sup>-1</sup>, which were assigned to Zn-O and Fe-O bonds, respectively. These peaks could be attributed to different vibrational modes of the metal oxide NPs. The C = O and C = C stretching vibrations were associated at 1964.31 and 2138 cm<sup>-1</sup> for ZnFe<sub>2</sub>O<sub>4</sub> NPs and ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs, respectively. Additionally, the RGO shows characteristic peaks at around 1613.90 cm<sup>-1</sup> that can be attributed to C = C stretching vibrations. Moreover, the absorption bands of ZnFe<sub>2</sub>O<sub>4</sub> NPs and ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs at around 3551.11 cm<sup>-1</sup> correspond to the stretching vibration of O-H groups.

### 3.5. UV analysis

UV spectroscopy is a commonly employed analytical technique to study the optical properties of synthesized samples. As shown in



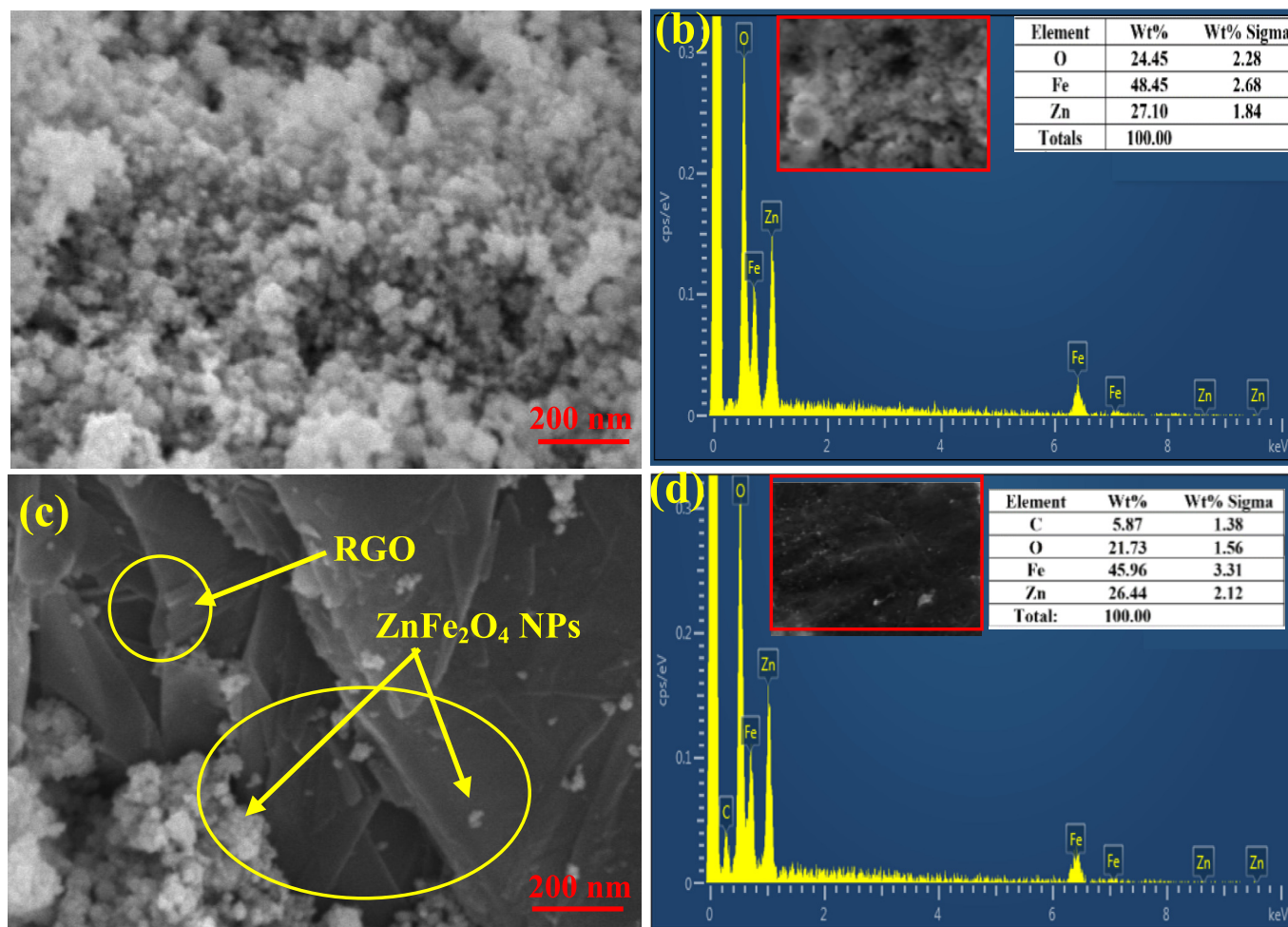


Fig. 2. (a) Image SEM of ZnFe<sub>2</sub>O<sub>4</sub> NPs, (b)EDX of ZnFe<sub>2</sub>O<sub>4</sub> NPs, (c) Image SEM of ZnFe<sub>2</sub>O<sub>4</sub>/ RGO NCs, (b)EDX of ZnFe<sub>2</sub>O<sub>4</sub>/ RGO NCs.

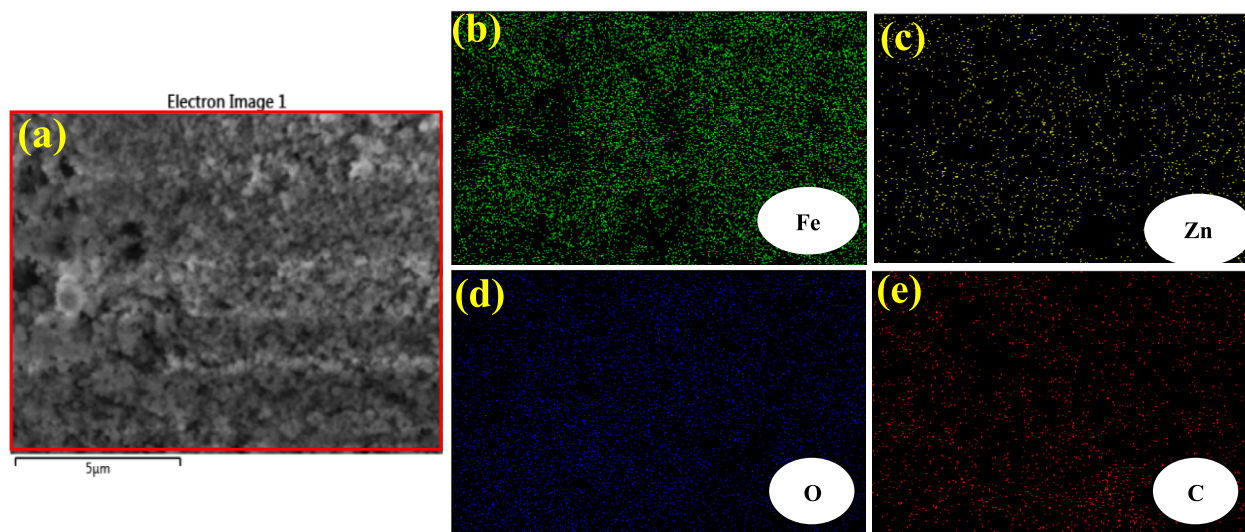
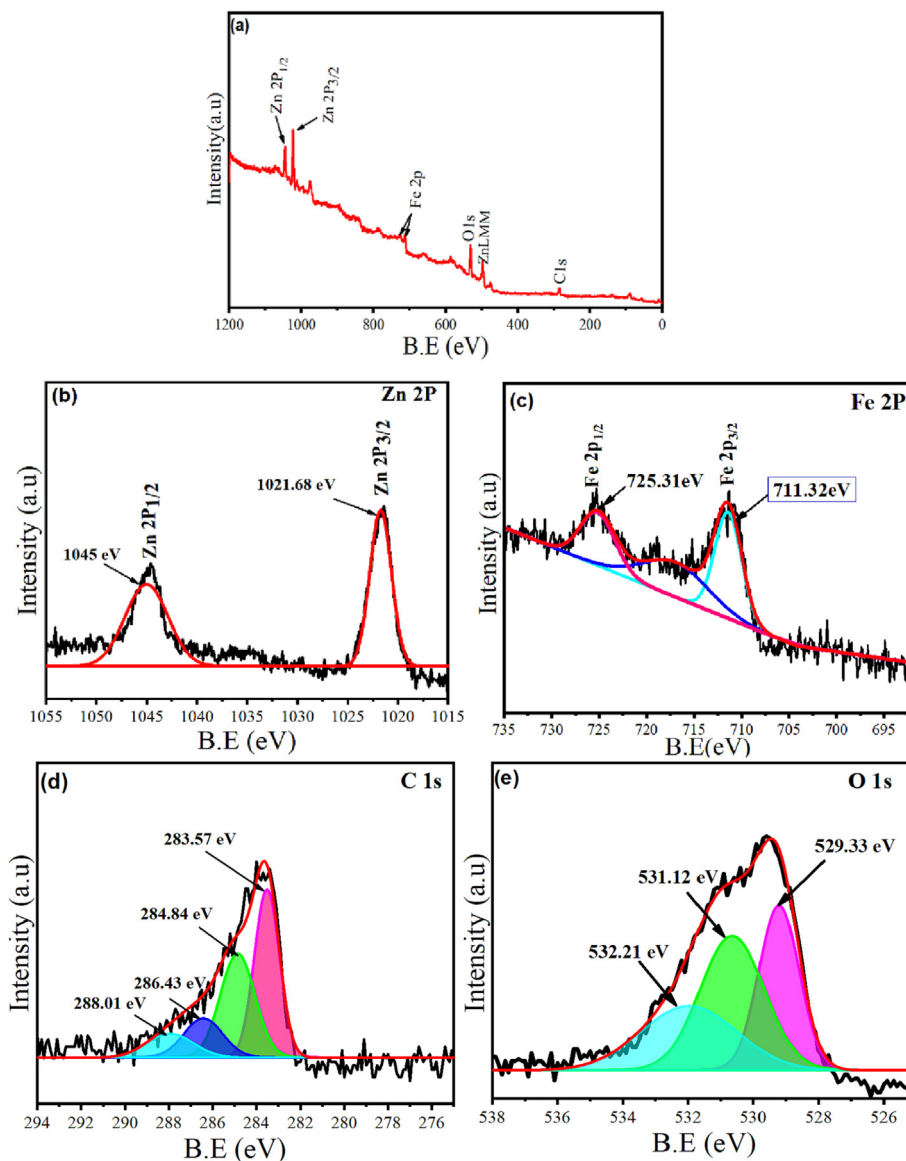


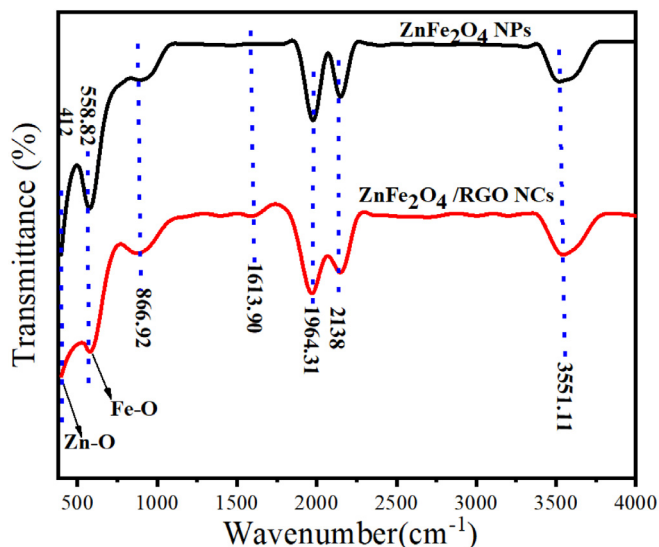
Fig. 3. Elemental mapping of ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs, SEM image (a), Iron (Fe) (b), zinc (Zn)(c), oxygen(O) (d), and carbon (C)(e).

Fig. 6, the UV-Vis spectra of ZnFe<sub>2</sub>O<sub>4</sub> NPs and ZnFe<sub>2</sub>O<sub>4</sub>/ RGO NCs were investigated. In particular, the UV spectra (Fig. 6a) displayed absorption peaks at 632.65 nm and 770.18 nm for ZnFe<sub>2</sub>O<sub>4</sub> NPs and

ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs, respectively. This process indicates that the band gap energy of the ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs decreased with RGO doping. The band gap energy (E<sub>g</sub>) of synthesized samples was calcu-



**Fig. 4.** (a) The full scan of XPS spectra of ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs, (b) high resolution of XPS of Zn 2P, (c) high resolution of XPS of Fe 2P, (d) high resolution of XPS of O 1s and (e) high resolution of XPS of C 1s in ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs.



**Fig. 5.** FT-IR spectra of ZnFe<sub>2</sub>O<sub>4</sub> NPs and ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs.

lated by Tauc plot analysis (Fig. 6b). Furthermore, ZnFe<sub>2</sub>O<sub>4</sub> NPs and ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs have band gap energies ( $E_g$ ) of 1.96 and 1.61 eV, respectively.

### 3.6. PL analysis

The migration rate of electrons and holes in ZnFe<sub>2</sub>O<sub>4</sub> NPs and ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs was measured using photoluminescence (PL). Fig. 7 illustrates the PL spectra of the synthesized samples at room temperature using an excitation wavelength of 355 nm. The addition of dopant to RGO led to a decrease in the emission peak observed at 481 nm. As shown in Fig. 7, the PL intensity of ZnFe<sub>2</sub>O<sub>4</sub> NPs was slightly reduced in the presence of RGO sheet. This reduction in PL intensity can be attributed to the improved charge transfer that occurs between RGO and ZnFe<sub>2</sub>O<sub>4</sub> NPs. Consequently, the strong electrical conductivity of RGO sheets facilitates effective separation of electron-hole ( $e^-/h^+$ ) pairs. PL spectra indicated that the recombination rate of the ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs was lower than ZnFe<sub>2</sub>O<sub>4</sub> NPs due to the interaction between ZnFe<sub>2</sub>O<sub>4</sub> and RGO.

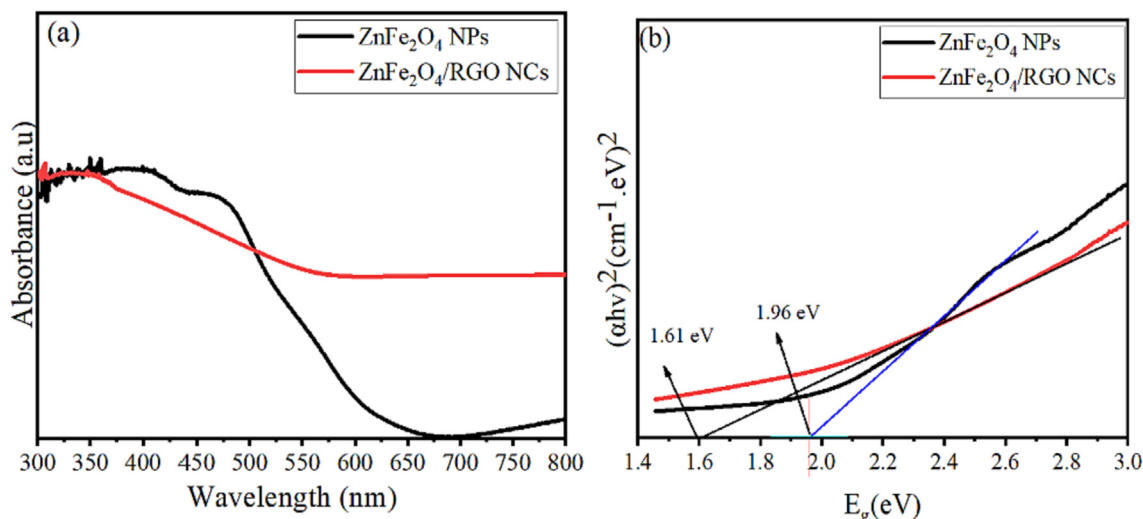


Fig. 6. (a) UV spectra of ZnFe<sub>2</sub>O<sub>4</sub> NPs and ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs, and (b) the band gap energy ( $E_g$ ) of ZnFe<sub>2</sub>O<sub>4</sub> NPs and ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs.

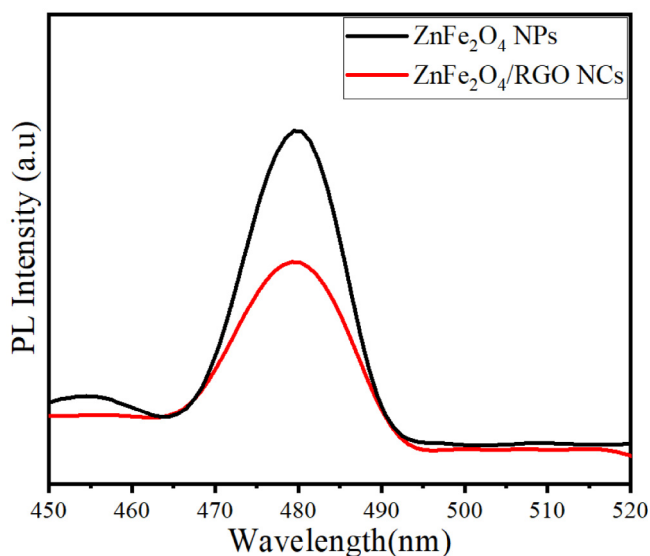


Fig. 7. PL spectra of ZnFe<sub>2</sub>O<sub>4</sub> NPs and ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs.

### 3.7. Cytotoxicity performance

The cytotoxicity effect of pure ZnFe<sub>2</sub>O<sub>4</sub>NPs and ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs by using the MTT assay toward two cancer cells ((MCF-7 and A549) was presented in Fig. 8a and b. We observed (Fig. 8a) that the cell viability of MCF-7 cells decreased with increasing concentrations of pure ZnFe<sub>2</sub>O<sub>4</sub>NPs and ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs. Similarly, the cell viability of the A549 cells (Fig. 8b) reduced with an increase in the concentrations (1.5–200 µg/mL) of the synthesized samples. Fig. 8a and b showed the cytotoxicity performance of ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs toward the MCF-7 and A549 was greater than pure ZnFe<sub>2</sub>O<sub>4</sub> NPs. The IC<sub>50</sub> values of ZnFe<sub>2</sub>O<sub>4</sub> NPs and ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs toward two types of human cancer cells were presented in Table 1.

The graphene-based nanocomposites have demonstrated promising anticancer activity against human cancer cells in vitro studies through several mechanisms. In the present study, the prepared nanocomposites can interact with cellular membranes and intracellular organelles, disrupting vital cellular processes in cancer cells. Furthermore, these synthesized nanocomposites are

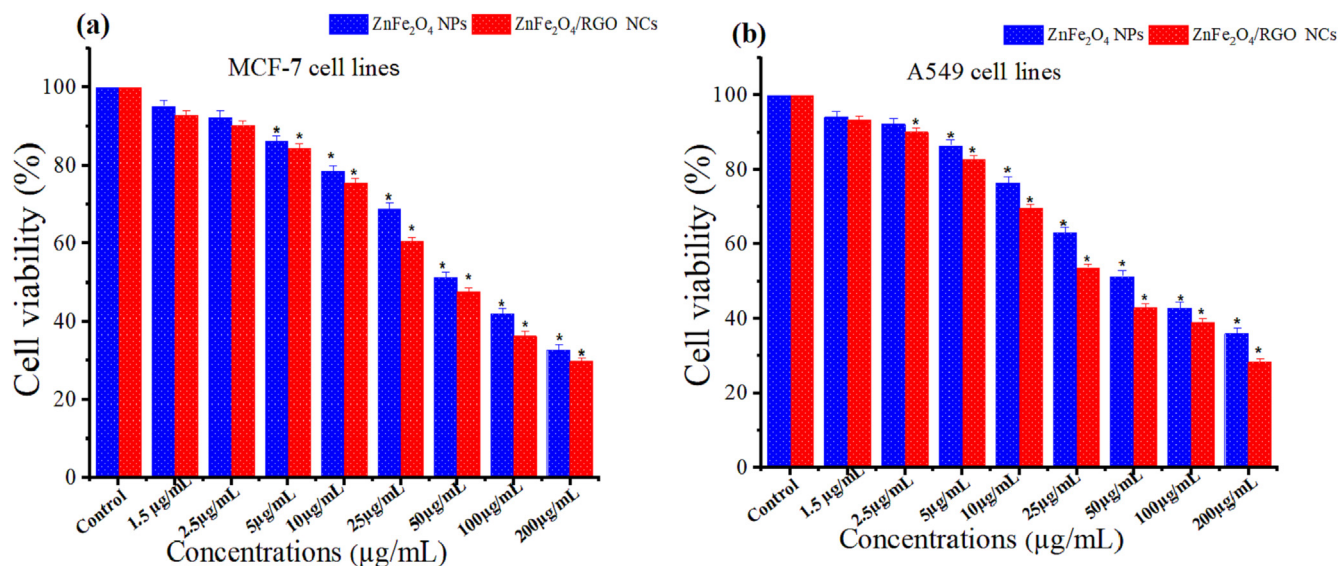
uptake by cancer cells through receptor-mediated endocytosis to produce reactive oxygen species (ROS). These ROS are oxygen radicals capable of death cancer cells by causing oxidative damage to killing cells. For example, synthesized ZrO<sub>2</sub>-ZnO/RGO NCs have been found to induce oxidative stress in cancer cells, leading to DNA damage (Ahamed et al., 2023b).

## 4. Discussion

In the present work, the ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs were synthesized using a simple one-step hydrothermal method and characterized using various techniques such as XRD, SEM, EDX, XPS, FTIR, UV-vis spectroscopy, and PL spectrometer. XRD data indicate that the ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs exhibited high purity and good crystallinity, with the ZnFe<sub>2</sub>O<sub>4</sub> NPs uniformly distributed on the RGO surface. (Hidayah et al., 2017). Furthermore, average crystal sizes were increased with RGO doping, which was calculated by the Scherrer equation (Patterson, 1939). XRD results show the successful synthesis of the ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs, which agrees with previous studies (Kaur et al., 2018; Wang and Shih, 2021). As shown in the SEM image, ZnFe<sub>2</sub>O<sub>4</sub> NPs displayed uniform distributions of spherical particles with agglomerations (Sripriya et al., 2018). EDX analysis confirmed that the elemental composition of both ZnFe<sub>2</sub>O<sub>4</sub> NPs and RGO were Zn, Fe, O, and C elements. Moreover, the uniform distribution of all elements (Zn, Fe, O, and C) in the synthesized ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs was investigated by SEM elemental mapping. The elemental composition and chemical state of the prepared samples were determined by the XPS technique. Importantly, the peaks at 531.1 eV and 532.2 eV represent O-C-O and Fe-C-O bonding, respectively, which were in good agreement with earlier studies (Hou et al., 2015; Shah et al., 2013; Yuan et al., 2015; Zhang et al., 2016; Zhao et al., 2017). These results were supported by EDX data.

The functional groups of prepared samples were further carried out by FTIR spectroscopy. In ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs, new peaks were observed at approximately 1613.90 cm<sup>-1</sup>, which are indicative of the stretching vibrations of C = C bonds in RGO. This peak was not appeared in the FTIR spectra of ZnFe<sub>2</sub>O<sub>4</sub>NPs, indicating the absence of RGO as shown in this study (Sarala et al., 2020). This phenomenon indicated that ZnFe<sub>2</sub>O<sub>4</sub>NPs were successfully anchored on the RGO sheet. FTIR spectra revealed that the intensities of the absorption bands of ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs reduced due to the presence of the RGO sheet (Zamani et al., 2020). FTIR results





**Fig. 8.** (a) Cell viability of MCF-7 cells with different concentrations of ZnFe<sub>2</sub>O<sub>4</sub> NPs and ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs for 24 h and (b) Cell viability of A549 cells with different concentrations of ZnFe<sub>2</sub>O<sub>4</sub> NPs and ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs for 24 h by using the MTT assay. \* Significantly different from control ( $p < 0.05$ ).

**Table 1**

The IC<sub>50</sub> values (µg/mL) of synthesized samples for two types of human cancer cells.

Cell lines	ZnFe <sub>2</sub> O <sub>4</sub> NPs	ZnFe <sub>2</sub> O <sub>4</sub> /RGO NCs
MCF-7	64.25	48.28
A549	65.51	46.70

were excellent and in agreement with XRD results and previous investigators (Li and Zhou, 2020; Riaz et al., 2022).

The optical properties of synthesized samples were performed by UV–vis spectroscopy. Interestingly, the UV spectra of ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs exhibited a shift in the position of the peaks compared to pure ZnFe<sub>2</sub>O<sub>4</sub> NPs due to the interaction between ZnFe<sub>2</sub>O<sub>4</sub> NPs and RGO as supported in these studies (Nadeem et al., 2020; Xie et al., 2013). Furthermore, the band gap energy ( $E_g$ ) of ZnFe<sub>2</sub>O<sub>4</sub> NPs and ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs were 1.96 eV and 1.61 eV, respectively, which were according with earlier research (Wu et al., 2015). The recombination rate of ZnFe<sub>2</sub>O<sub>4</sub> NPs and ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs was further evaluated using a photoluminescence (PL) spectrometer. PL results showed that the reduction of the emission peak at 481 nm of ZnFe<sub>2</sub>O<sub>4</sub> NPs was observed with RGO doping. The PL intensity of the ZnFe<sub>2</sub>O<sub>4</sub> NPs was lower than that of ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs. The reduction of PL intensity could be attributed to the enhanced charge transfer between the RGO and ZnFe<sub>2</sub>O<sub>4</sub> NPs (Khadgi et al., 2017). Additionally, the RGO sheets have strong electrical conductivity as the electron source to effectively separate the electron-hole ( $e^-/h^+$ ) pair (Hou et al., 2013). These results suggested that ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs can be applied in potential applications including, photocatalytic and cancer treatment (Behera et al., 2018; Renukadevi and Pricilla Jeyakumari, 2020).

Due to their unique properties, many researchers have been interested in applying metal oxide NPs and graphene-based NCs in biomedicine and cancer therapy (Alafaleq et al., 2023; Alaizeri et al., 2022; Arunima Rajan et al., 2019; Krishnan et al., 2021; Sarala et al., 2020; Tabrez et al., 2022c, 2022b). In this work, the MTT experiment was carried out on two different cancer cell lines (MCF-7 and A549) to assess the anticancer activity of synthesized samples. It was observed that ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs exhibited higher cytotoxicity towards the cancer cell lines (MCF-7 and A549) compared to pure ZnFe<sub>2</sub>O<sub>4</sub> NPs exhibiting IC<sub>50</sub> around 50 for 24 h. At

the same interval time, previous studies have observed significant cytotoxicity of metal with RGO in various cancer cell types, including HepG2, and A549 cells, with IC<sub>50</sub> concentrations of approximately 100 µg/mL and 250 µg/mL (Alaizeri and Alhadlaq, 2022; Krishnan et al., 2021). Our results suggest that RGO doping in ZnFe<sub>2</sub>O<sub>4</sub> NPs enhances their anticancer performance against human cancer (MCF-7 and A549) cells.

## 5. Conclusion

In the present work, one-step hydrothermal synthesis was successfully used to prepare the ZnFe<sub>2</sub>O<sub>4</sub> NPs and ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs. Physicochemical properties of pure ZnFe<sub>2</sub>O<sub>4</sub> NPs and ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs were further investigated by different analytical techniques such as XRD, SEM, XPS, UV–Vis, FTIR, and PL. XRD results showed that the prepared samples have high crystallinity. Furthermore, the average crystal sizes of ZnFe<sub>2</sub>O<sub>4</sub> NPs increased with the addition of the RGO sheet. SEM images demonstrate that ZnFe<sub>2</sub>O<sub>4</sub> NPs were successfully anchored on the RGO nanosheets. The elemental compositions of ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs were confirmed by XPS and EDX analysis. SEM elemental mapping revealed a uniform distribution of zinc (Zn), iron (Fe), oxygen (O), and carbon (C) within the ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs. FT-IR spectra confirmed that the functional group of prepared samples was changed with RGO adding. UV–vis spectra showed the absorption peak of ZnFe<sub>2</sub>O<sub>4</sub> NPs affected by RGO doping. This process indicates that the band gap energy of ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs (1.96 eV) is lower than pure ZnFe<sub>2</sub>O<sub>4</sub> NPs (1.61 eV). PL results showed that the ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs have a lower recombination rate than ZnFe<sub>2</sub>O<sub>4</sub> NPs. Biological results showed that the ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs exhibit higher anticancer activity in human cancer cells (MCF-7 and A549) than pure ZnFe<sub>2</sub>O<sub>4</sub> NPs. This study suggests that RGO plays a crucial role in improving the anticancer potential of ZnFe<sub>2</sub>O<sub>4</sub> NPs by tuning its optical properties. The results of this study warrant further inquiry into the potential therapeutic applications of nanocomposites created by ZnFe<sub>2</sub>O<sub>4</sub> NPs and RGO sheets. In this work, the one-step preparation, characterization, and anticancer potential of ZnFe<sub>2</sub>O<sub>4</sub>/RGO NCs provides valuable insights into the field of nanomaterials for cancer therapy. However, certain limitations and areas for future research should be acknowledged. One of the limitations of this study is synthesis parameters, such as reaction time, temperature,

and precursor concentrations, which affect improved control over the nanocomposite morphology and properties. Additionally, further in vitro and in vivo studies are required to comprehensively evaluate their therapeutic efficacy and safety profiles.

### Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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