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Viral Gastroenteritis

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Viral agents are the most common causes of acute gastroenteritis, a syndrome of acute vomiting and diarrhea associated with inflammation of the stomach, small intestines, or large intestines. Among children, viral gastroenteritis remains a leading cause of pediatric morbidity and mortality worldwide.^{1,2} With the discovery of norovirus³ and rotavirus⁴ in the early 1970s, and subsequent development of improved diagnostic strategies for these and other enteric viruses,^{5,6} the importance of viral agents as causes of diarrheal disease has been increasingly appreciated. The global introduction of highly effective rotavirus gastroenteritis and recognition of norovirus as a major cause of acute gastroenteritis.^{7–12}

ETIOLOGIC AGENTS

A small group of viruses accounts for most cases of acute gastroenteritis among children. These include group A rotaviruses, human caliciviruses (including noroviruses and sapoviruses), enteric adenoviruses (types 40 and 41), and human astroviruses.⁷ Other viruses frequently are detected in children suffering from diarrheal illnesses; however, the prevalence of many of these other viruses in healthy children has not been determined, and therefore a causal association with diarrhea has not been established.

Rotaviruses (family Reoviridae) are 100-nm particles consisting of an outer capsid, an inner capsid, and a core.¹³ The double-stranded RNA genome is composed of 11 segments that code for 6 structural proteins and 6 nonstructural proteins. The outer capsid is composed of 2 proteins: VP7 (G protein, for glycoprotein) and VP4 (P protein, for protease-cleaved protein). These proteins are the principal antigens to which neutralizing antibodies are directed and account for the classification scheme for rotavirus strains.¹⁴

Caliciviruses are nonenveloped, 27- to 40-nm, single-stranded RNA viruses in the family Caliciviridae.¹⁵ Human caliciviruses are divided into 2 genera: norovirus and sapovirus. Noroviruses include numerous genetically related viruses, further divided into 6 genogroups (I–VI), 3 of which (I, II, and, rarely, IV) are associated with human disease. At least 9 and 22 genotypes, respectively, have been identified for genogroups I and II.¹⁶ Despite this extensive genetic diversity among noroviruses, genogroup II genotype 4 (GII.4) viruses are the most common causes of norovirus outbreaks worldwide¹⁷; the emergence of new GII.4 strains occurs every 2 to 4 years and can be associated with a global increase in gastroenteritis outbreaks.^{18–20} Efforts to cultivate noroviruses in available cell culture systems or to develop an animal model have been unsuccessful, and this failure has hampered the development of simple diagnostic tests and the evaluation of disinfectants.²¹ Important progress has been made by using B cells, but this system is not widely available.²²

Astroviruses are nonenveloped, 28-nm, single-stranded RNA viruses in the family Astroviridae. Until 2008, human astroviruses were thought to be limited to 8 closely related genotypes (serotypes), now referred to as the "classic" human astroviruses (HAstV 1–8). HAstV genotype 1 is most commonly detected, followed by types 2 to 5; however, more than a single serotype usually circulates in communities during each season.²³ Non–type 1 viruses can predominate in a season, and greater serotype diversity can be found in developing countries.^{23–25}

Adenoviruses are 70- to 100-nm, nonenveloped, double-stranded DNA viruses in the family Adenoviridae.²⁶ Although 7 species of adenoviruses (classified as HAdV A–G) containing at least 51 different genotypes can cause human infection, species F (genotypes 40 and 41) adenoviruses typically cause gastroenteritis.²⁷

Many studies report the detection of viruses in stool samples from children with diarrheal illness. A key method to establish a causal association between a virus and a disease is to observe a higher prevalence among children with gastroenteritis compared with asymptomatic children (Table 56.1).

Many other viruses have been found in fecal specimens, but the data are inconclusive regarding the pathogenicity for many of these viruses. Reports of the clinical characteristics of patients infected with the severe acute respiratory syndrome (SARS) coronavirus and with the more recently described Middle East respiratory syndrome (MERS) coronavirus have noted diarrhea in approximately one fourth of cases.^{28,29}

EPIDEMIOLOGY

All the major enteric viruses are transmitted primarily through close person-to-person contact by the fecal-oral route.³⁰ Noroviruses, in addition, are easily spread through contaminated food and water and therefore are major causes of foodborne disease.^{31–33} Noroviruses are present in the vomitus of ill people, and droplet spread through exposure to vomitus has been demonstrated to be a mechanism of spread in both healthcare and public settings.^{34–36} Contact with a symptomatic child is a major

TABLE 56.1	Positivity Rates of Viral Pathogens Among Children <5
Years of Age	With Acute Gastroenteritis and Healthy Controls: United
States, 2008	to 2009 ⁷

	Acute Gastroenteritis (N = 1564)	Healthy Controls (N = 1000)	
Agent	No. (%)	No. (%)	
Group A rotaviruses	282 (18)	2 (<1)	
Noroviruses	334 (21)	40 (4)	
Adenoviruses 40/41	186 (12)	18 (2)	
Sapoviruses	80 (5)	42 (4)	
Parechoviruses	76 (5)	44 (4)	
Astroviruses	75 (5)	30 (3)	
Bocaviruses	22 (1)	24 (2)	
Aichiviruses	4 (<1)	0 (0)	

determinant of norovirus infection. Children <5 years of age seem to be more contagious than older children and adults and therefore play key roles in norovirus transmission to all age groups.³⁷ The modes of transmission of adenoviruses are less well understood, but transmission is presumed to occur primarily through close contact by fecal-oral spread. Spread through fomites is possible for each of the agents, and it can play an important role in disease acquired in institutional and group childcare settings.³⁸

Common viral gastroenteritis viruses are globally distributed. Although traditionally temperate climates are associated with a strong winter peak of rotavirus, a meta-analysis showed that the level of country development was a stronger predictor of seasonality than was geographic location or climate.³⁹ Since the introduction of routine vaccination in the United States, the rotavirus season has diminished substantially in magnitude, and it has also been delayed in some years.^{12,40} A biennial pattern of lower, delayed rotavirus seasons followed by slightly higher peaks has emerged in the US. (Fig. 56.1).^{12,40,41} In low-income and tropical settings, rotaviruses typically circulate year-round.39,42,43 Noroviruses circulate year-round in most areas, but a clear wintertime seasonality to outbreaks is present, particularly in healthcare settings.^{31,44,45} Astroviruses typically peak during winter and sapoviruses peak during early spring months in temperate countries.^{7,46} The seasonality of adenoviruses is less distinct, and transmission has been described year-round, with summertime epidemics.7,47

The highest rates of severe rotavirus infections occur in the first 2 years of life; most hospitalizations and severe dehydrating disease occur between 4 and 23 months of age.^{48–50} Infections in the first 3 months are less common and are often asymptomatic, probably because of protection from maternally acquired antibodies.^{48,51} Rotaviruses can infect children more than once, with each subsequent infection less severe as a result of immunity that develops following infection.⁴⁸ Severe rotavirus disease among older children and adults is less common, but infection can occur in people exposed to younger children in childcare settings and schools. Outbreaks also occur in long-term care facilities.⁵²

Before the introduction of rotavirus vaccines, rotaviruses accounted for 25% to 50% of hospitalizations for gastroenteritis among children <5 years of age and for 5% to 20% of milder cases in people who sought care in clinics.^{49,53} Following vaccine introduction, the rates of rotavirus-associated healthcare use substantially decreased (see Fig. 56.1).^{11,12} Population-based surveillance in the US detected rotavirus in 13% of children hospitalized for gastroenteritis and in 6% of pediatric outpatient visits for gastroenteritis.¹¹ Globally, rotavirus causes approximately 500,000 deaths/year in children less than 5 years of age, and deaths among children in the poorest countries account for more than 85% of the total.¹⁵⁴ In middle-income countries (Mexico and Brazil) where rotavirus vaccines were introduced in 2006, sustained reductions were documented in diarrhea-associated pediatric mortality rates (Fig. 56.2).^{9,10} Rotavirus-associated death is rare in developed countries.^{55,56}

Noroviruses now are recognized as the most common causes of both endemic disease and outbreaks of gastroenteritis.^{11,32,57} Noroviruses are estimated to cause 19 to 21 million illnesses a year in the US that result in 1.7 to 1.9 million outpatient visits, 400,000 emergency department visits, 56,000 to 71,000 hospitalizations, and 570 to 800 deaths.³² Globally, norovirus is estimated to cause 18% of the cases of severe diarrheal disease in children <5 years old (17% of inpatient cases and 24% of community episodes).⁵⁸ Following the introduction of rotavirus vaccines and the subsequent decline in rotavirus disease burden, noroviruses have overtaken rotavirus to become the predominant causes of severe gastroenteritis in the pediatric population.^{11,59,60} Among children <5 years old who were studied in an active surveillance network in the US, norovirus accounted for 21% of gastroenteritis cases requiring medical attention, including 17% of children admitted to the hospital and 28% of outpatients (see Table 56.1).¹¹

All age groups are affected by noroviruses, but the disease incidence is highest in children <5 years of age.⁶¹ Noroviruses are the most commonly reported causes of foodborne disease in the US, whereas internationally, estimates vary widely.^{32,62} Common foods associated with outbreaks include those that are uncooked or handled after cooking (which can be contaminated by ill food handlers), as well as shellfish harvested from contaminated water. Healthcare facilities, including nursing homes and hospitals, are the most common settings of norovirus outbreaks; other



FIGURE 56.1 Diarrhea-associated healthcare use rates among United States children <5 years of age, 2001 to 2011. (From Leshem E, Moritz RE, Curns AT, et al. Rotavirus vaccines and health care utilization for diarrhea in the United States (2007–2011). Pediatrics 2014;134(1):15–23.)

environments such as childcare facilities and cruise ships are frequently affected.

Sapoviruses are most commonly associated with sporadic gastroenteritis, usually among young children.^{63,64} Outbreaks caused by sapoviruses are less common and tend to occur in older adults.⁶⁵ Sapoviruses were detected in approximately 10% of all gastroenteritis episodes in England and Finland and in 4% of hospitalized cases in Finland among children <2 years of age.^{66,67} In the US, sapoviruses were detected in 5% of pediatric diarrhea cases compared with 4% among healthy controls (P < 0.01) (see Table 56.1).⁷ Sapovirus infections tend to be less severe than norovirus infections; vomiting is common, and diarrhea may not be present.^{7,67}

Although astroviruses have been detected in all age groups, most infections are in children <2 years of age and tend to be less severe than



FIGURE 56.2 Number of diarrhea-related deaths among children aged <5 years from July 2002 through December 2011 in Mexico according to age group. (From Gastanaduy PA, Sanchez-Uribe E, Esparza-Aguilar M, et al. Effect of rotavirus vaccine on diarrhea mortality in different socioeconomic regions of Mexico. Pediatrics 2013;131(4): e1115–1120.)

rotavirus infections.⁶⁸ Serosurveys in the US have shown that >90% of children have antibodies to human astroviruses by 6 to 9 years of age.⁶⁹ Disease in adults is uncommon, but it can occur in outbreak settings.⁷⁰ Astroviruses are usually detected in <10% of young children treated for gastroenteritis in outpatient clinics or in hospitals, but these viruses are occasionally found at higher frequencies.^{7,47,71} Although astroviruses primarily cause sporadic disease, outbreaks have been reported in a range of settings, and astroviruses can be common causes of nosocomial gastroenteritis in children's hospitals.⁷²

Most enteric adenovirus infections occur in children <2 years of age, and these viruses appear to be less important causes of gastroenteritis among adults.^{7,73,74} Enteric adenoviruses were shown to account for 5% to 12% of hospitalizations for acute gastroenteritis in children and can be common causes of healthcare-associated diarrhea.^{7,75} Enteric adenoviruses are detected in 1% to 12% of children with community-associated diarrhea.^{7,76,77}

PATHOGENESIS

After oral inoculation, viruses infect cells of the small intestinal villi.⁷⁸ Infection of the mature villous enterocytes, which have both digestive and absorptive functions, leads to cell death and sloughing of the villus cells. In a normal host, infection resolves as the number of susceptible mature enterocytes decreases secondary to cell death and as the host generates an immune response. Viruses are shed in the stool during the acute illness in large quantities. Rotaviruses, noroviruses, and astroviruses can also be shed for 1 to 2 days before illness and for several days following resolution of symptoms.^{30,38} Although viral gastrointestinal infections are generally confined to the intestine, rotavirus and norovirus infections can result in antigenemia and the presence of nucleic acid in blood of ill patients, but extraintestinal disease is rare.^{79,80} Asymptomatic infection is common, especially for norovirus,⁶⁶ and is thought to play a role in disease transmission.⁸¹

Immunity to rotavirus is acquired, and multiple infections typically are required until the child is fully protected against disease.^{48,82,83} After a primary infection, homotypic immunity is stronger, but immunity seems to broaden to other serotypes with subsequent infections.⁴⁸ Wide estimates of the duration of immunity to norovirus range from 6 months to 4 to 9 years after natural infection occurs; however, heterotypic immunity may be limited.^{37,84} Because diarrheal disease caused by astrovirus, adenovirus, and sapovirus is largely restricted to children, immunity is believed to be long-lasting.

A correlation exists between the expression of histo-blood group antigens (HBGAs) and susceptibility to norovirus and possibly rotavirus infections.⁸⁵⁻⁹² HBGAs comprise a diverse family of carbohydrates expressed on the mucosal epithelia of the respiratory, genitourinary, and

digestive tracts; HBGAs are recognized as receptors allowing norovirus attachment and cellular entry. The expression of HBGAs is determined by 3 gene families expressing the ABO (A/B enzymes), secretor (FUT2) gene, and Lewis-type (FUT3) gene. Single nucleotide gene polymorphisms can inactivate the expression of these gene products, thereby breaking a link in the norovirus binding and infection process. Mutations in the FUT2 gene leading to the absence of HBGA expression (nonsecretor phenotype) have been associated with resistance to infection, in particular to GII.4 noroviruses, which are the predominant global genotype.^{85,86,88,89,93-96} HGBA expression appears to vary by ethnicity; 20% to 25% of the US. white population consists of nonsecretors compared with 2% of persons of Meso-American or Hispanic ancestry. In East Asia, different secretor mutations (A385T and FUT3) observed in Chinese and Vietnamese children confer lower risk of norovirus infections.91,97 Evidence points to lower susceptibility of nonsecretors to rotavirus infections in various populations.^{90–92}

CLINICAL MANIFESTATIONS

Although these viruses differ genetically and structurally, the clinical presentations of acute gastroenteritis caused by these agents are indistinguishable. After a short incubation period, infections with any of the viruses lead to an acute onset of gastroenteritis (Table 56.2). Vomiting is often an early sign, common in rotavirus infection, and particularly pronounced in norovirus and sapovirus infections.^{7,98} Typically diarrhea is watery, and without blood, mucus, or fecal leukocytes. Fever occurs in approximately half of children with rotavirus infection and in approximately one fourth of children with nonrotavirus viral gastroenteritis, and it is often an early sign. Vomiting and fever frequently cease within 1 to 3 days, whereas diarrhea can persist longer, especially in rotavirus infections. Other symptoms include abdominal cramps, malaise, and seizures.^{7,99}

The most important and common complication of viral gastroenteritis is dehydration, often associated with electrolyte abnormalities. Malabsorption can occur during the illness and persist for weeks following infection. Extraintestinal complications are rare, but encephalitis, seizures, acute myositis, hemophagocytic lymphohistiocytosis, acute flaccid paralysis, and sudden infant death syndrome have been described in children with rotavirus infections.^{13,99,100} The association of these complications with rotavirus infection remains unclear. Severe and prolonged diarrhea associated with each agent has been reported among children with malnutrition and among children with congenital or acquired immunodeficiencies.¹⁰¹⁻¹⁰⁵

Although viral causes of gastroenteritis are not distinguishable by clinical signs and symptoms, clinical characteristics of cases in outbreak settings have been helpful in predicting the presence of noroviruses.

TABLE 56.2 Epidemiologic Features of Viral Causes of Gastroenteritis								
Feature	Rotavirus	Noroviruses	Sapoviruses	Astroviruses	Adenoviruses			
Predominant age of illness	<5 yr	All ages	<5 yr	<2 yr	<2 yr			
Mode of transmission	Person-to-person through fecal-oral route, fomites	Person-to- person through fecal-oral and vomitus-oral, fomites, food or water	Person-to- person through fecal-oral route	Person-to-person through fecal-oral route	Person-to- person through fecal-oral route			
Incubation period	1–3 days	12–48 hr	12–48 hr	1–4 days	3–10 days			
Symptoms								
Diarrhea	Explosive, watery	Watery with acute onset	Watery; milder than rotavirus	Watery; milder than rotavirus	Watery; milder than rotavirus; can be prolonged			
Vomiting	80%–90%	>50%; often dominant symptom	Less common than rotavirus	Less common than rotavirus	Less common than rotavirus			
Fever	Frequent	Less common, usually mild	Less common, usually mild	Less common, usually mild	Less common, usually mild			
Illness duration	2–8 days	1–5 days	1–4 days	1–5 days	3–10 days			
Clinical diagnosis	Stool EIA or LPA	RT-PCR	RT-PCR	Stool EIA (not available in United States)	Stool EIA			

EIA, enzyme immunoassay; LPA, latex particle agglutination; RT-PCR, reverse transcriptase-polymerase chain reaction.

Modified from Peck AJ, Bresee JS. Viral gastroenteritis. In: McMillan JA, Feigin RD, De Angelis CD, Jones MD Jr (eds). Oski's Pediatrics, 4th ed. Philadelphia, Lippincott Williams & Wilkins, 2006, pp. 1288–1294.

Kaplan and colleagues⁹⁸ found that outbreaks that met simple epidemiologic and clinical criteria were likely to have been caused by noroviruses. These criteria included (1) failure to detect a bacterial or parasitic pathogen in stool specimens, (2) the occurrence of vomiting in >50% of patients, (3) mean duration of illness of 12 to 60 hours, and (4) mean incubation period of 24 to 48 hours. The "Kaplan criteria" have been validated and are used by local health departments for the diagnosis of outbreaks in the absence of laboratory testing for norovirus.¹⁰⁶

DIAGNOSIS

Laboratory diagnosis of viral gastroenteritis is made by detection of viral antigen or nucleic acid in fresh, whole stool samples obtained during the acute illness. Commercially available assays to detect rotavirus antigen in stools offer easy and inexpensive methods to diagnose infection in children. These tests are available as either enzyme immunoassay (EIA) or a latex particle agglutination test for group A rotaviruses, designed to detect the VP6 protein.¹³ Antigen detection tests generally have high (90% to 95%) sensitivity and specificity.⁹³ Other methods of rotavirus detection include electron microscopy, viral isolation, and RNA poly-acrylamide gel electrophoresis. Reverse transcription–polymerase chain reaction (RT-PCR) is rarely used in clinical practice because of the test's high sensitivity; it may detect virus when that virus is not causing disease.¹⁰⁷ Serologic testing for rotavirus infection is possible but impractical and thus is not widely available in clinical care settings.

Real-time RT-PCR is the preferred laboratory method for detecting noroviruses. This assay is highly sensitive and can detect as few as 10 norovirus copies per reaction and provide a semiquantitative estimate of viral load. The assay is generally used to detect norovirus in stool, but it can also be used for vomitus, foods, water, and environmental specimens for outbreak investigations, although with reduced sensitivity. Conventional PCR followed by sequence analysis of the PCR products is used for norovirus and the high frequency that the virus can be found in healthy persons,^{11,66,109} diagnostic results should be interpreted in light of clinical characteristics and, if available, the background level of detection in a control population. A commercial EIA for norovirus was approved in the US in 2011; however, this assay has low sensitivity and is not recommended for diagnosing norovirus infection in sporadic cases. Norovirus EIA can be useful in outbreak investigations.^{110–112}

Commercial EIAs for detection of astrovirus viral antigen in stool are available in Europe, but not in the US. RT-PCR is a sensitive and specific method for detection of astroviruses.¹¹³ RT-PCR, serologic assays, and electron microscopy are used primarily in research settings. Similarly, RT-PCR, EIA, and latex particle agglutination kits are available commercially and provide highly sensitive and specific antigen detection of enteric adenoviruses.²⁶ All viral agents of gastroenteritis are detectable by electron microscopy and immune electron microscopy, but these tests are seldom used because of their relatively low sensitivity and specificity, expense, and required expertise.

TREATMENT

No specific therapies are available for viral gastroenteritis. Case management depends on accurate and rapid assessment of the severity of dehydration, correction of fluid loss and electrolyte disturbances, and maintenance of adequate hydration and nutrition.¹¹³ Treatment with oral rehydration solution (ORS) containing appropriate glucose-electrolyte solutions is sufficient for most patients. ORS use a protein sodium-glucose cotransporter 1 (SGALT1) that facilitates solute absorption across the luminal membrane. Water passively follows the osmotic gradient generated by transcellular transport of electrolytes and nutrients. In 2004, the World Health Organization recommended use of low-osmolarity ORS (245 mOsm/L), which has been shown to decrease stool output and vomiting in comparison with children treated with traditional ORS (311 mOsm/L). Intravenous rehydration can be required for children with severe dehydration (≥10% fluid deficit, shock, or near shock), intractable vomiting, or ORS treatment failure. Factors such as young age, unusual irritability or drowsiness, progressive course of symptoms, or uncertainty of diagnosis can indicate a need for close observation.

Breastfed infants should continue to nurse on demand. In children who are not breastfed, change to a lactose-free diet may result in earlier resolution of acute diarrhea and reduce treatment failure.¹¹⁴ Children taking solid foods should continue to receive their usual diet during episodes of diarrhea, although substantial amounts of foods high in simple sugars should be avoided because the osmotic content may worsen diarrhea. Evidence on the use of oral probiotics (e.g., *Lactobacillus* spp.) to reduce the duration of diarrhea caused by rotavirus is conflicting.^{115–117} Zinc, used both as supplement and treatment, reduces the severity, duration, and incidence of diarrhea in low- to middle-income settings and is considered one of the mainstays of treatment in developing countries^{118,119}; however, the role of zinc supplementation in high-income settings has not been extensively studied. Because viral agents account for most cases of infectious gastroenteritis in children,

Key Points: Epidemiology, Clinical Features, Diagnosis, Treatment, and Prevention of Viral Gastroenteritis

EPIDEMIOLOGY

- Viruses account for most cases of acute gastroenteritis among children. These include rotaviruses, noroviruses, enteric adenoviruses (types 40 and 41), astroviruses, and sapoviruses.
- Viral gastroenteritis occurs in endemic and epidemic forms. Rotaviruses and noroviruses are the major causes of endemic disease, with seasonal peaks during the winter months in temperate climates. Noroviruses are the main causes of epidemic disease.
- Following introduction of rotavirus vaccine, rates of rotavirus gastroenteritis have decreased substantially, and currently noroviruses are the major causes of acute gastroenteritis in the United States.

CLINICAL FEATURES

- The clinical characteristics of illnesses caused by different viruses generally are indistinguishable. These include watery diarrhea and, in some cases, vomiting and fever.
- The most common severe complication of viral gastroenteritis is dehydration, often with electrolyte abnormalities.
- Prolonged diarrhea has been reported among children with malnutrition and among immunocompromised patients.

DIAGNOSIS

 Laboratory diagnosis of viral gastroenteritis is best made by detection of viral antigen or nucleic acid in fresh, whole stool samples obtained during the acute illness.

antimicrobial therapy is not usually indicated for children (see Chapter 55). $^{\rm 120}$

PREVENTION

The best public health intervention to reduce the incidence of severe viral gastroenteritis is the implementation of live, oral rotavirus vaccines in routine immunization programs. Rotavirus vaccines are attenuated strains given in multiple doses designed to replace a child's first exposure to wild-type rotavirus with strains that will not cause disease but will generate an adequate immune response to confer protection. Currently, 2 rotavirus vaccines are licensed in the US.¹²¹ A third vaccine has been evaluated in India.¹²² Additional vaccines are in development and may be available within several years.

As of 2015, rotavirus vaccines have been added to routine immunization programs in more than 75 countries, and these vaccines exert a dramatic impact with rapid and sustainable reductions in diarrheaassociated healthcare use and acute gastroenteritis–associated mortality rates.^{9,10,12,40,123} Vaccine efficacy was shown to be very high in high- and middle-income settings and moderate in low-income populations^{124–128}; however, vaccines may have their greatest impact in these low-income settings because of the higher burden of disease. The World Health Organization issued a global recommendation for the use of rotavirus vaccines.¹²⁹ Vaccine effectiveness in real-life routine use across various demographic and geographic settings has been good.^{130,131}

Except for rotavirus, prevention of viral gastroenteritis is limited to nonspecific strategies. Breastfeeding confers some protection against rotavirus infection, and probably other viral infections, in young infants. Good hygiene, including meticulous handwashing practices, is an effective prevention strategy and should be encouraged, particularly in institutional settings, such as childcare centers and hospitals.¹³² Significantly reducing transmission of viral agents of gastroenteritis requires strict adherence to recommendations because of the low infectious dose

 Commercially available assays to detect rotavirus antigen in stools offer an easy and inexpensive method to diagnose infection in children.

TREATMENT

- No specific therapies are available for viral gastroenteritis.
- The mainstays of therapy include assessment of dehydration severity, correction of fluid loss and electrolyte disturbances, and maintenance of adequate nutrition.
- Oral rehydration using oral rehydration solution with appropriate glucose-electrolyte solutions is sufficient for most patients.
- Zinc is used for treatment in low-income countries.

PREVENTION

- Rotavirus vaccines have been added to routine immunization programs in more than 75 countries and have substantially reduced diarrhea-associated healthcare use and acute gastroenteritis-associated mortality rates.
- Except for rotavirus, prevention of viral gastroenteritis is limited to nonspecific strategies including breastfeeding, hand hygiene, and strict adherence to hygiene guidelines including use of Environmental Protection Agency–approved cleaners for food preparation areas and contaminated surfaces.

and the high quantity of viruses excreted in stool (and often vomitus) from infected persons and because these viral agents are quite stable in the environment. Noroviruses are relatively resistant to environmental disinfection; cleaning contaminated surfaces and food preparation areas with Environmental Protection Agency–approved cleaners is effective in settings where viral gastroenteritis outbreaks occur.^{132,133}

Although experimental vaccines against noroviruses are in development, proof that these vaccines could protect against natural infection remains to be established.^{134,135} These vaccines are based on virus-like particles and showed promise in volunteer challenge studies when they were administered both intranasally and intramuscularly. No vaccines against other viral gastroenteritis agents are yet in human trials.

All references are available online at www.expertconsult.com.

KEY REFERENCES

- Chhabra P, Payne DC, Szilagyi PG, et al. Etiology of viral gastroenteritis in children <5 years of age in the United States, 2008–2009. J Infect Dis 2013;208(5):790–800.
- 9. Gastanaduy PA, Sanchez-Uribe E, Esparza-Aguilar M, et al. Effect of rotavirus vaccine on diarrhea mortality in different socioeconomic regions of Mexico. Pediatrics 2013;131(4):e1115–e1120.
- 11. Payne DC, Vinje J, Szilagyi PG, et al. Norovirus and medically attended gastroenteritis in U.S. children. N Engl J Med 2013;368(12):1121–1130.
- Leshem E, Moritz RE, Curns AT, et al. Rotavirus vaccines and health care utilization for diarrhea in the United States (2007–2011). Pediatrics 2014;134(1):15–23.
- Jones MK, Watanabe M, Zhu S, et al. Enteric bacteria promote human and mouse norovirus infection of B cells. Science 2014;346:755–759.
- Ahmed SM, Hall AJ, Robinson AE, et al. Global prevalence of norovirus in cases of gastroenteritis: a systematic review and meta-analysis. Lancet Infect Dis 2014;14(8):725–730.
- 122. Bhandari N, Rongsen-Chandola T, Bavdekar A, et al. Efficacy of a monovalent human-bovine (116E) rotavirus vaccine in Indian infants: a randomised, doubleblind, placebo-controlled trial. Lancet 2014;383:2136–2143.
- Atmar RL, Bernstein DI, Harro CD, et al. Norovirus vaccine against experimental human Norwalk Virus illness. N Engl J Med 2011;365(23):2178–2187.

REFERENCES

- Liu L, Oza S, Hogan D, et al. Global, regional, and national causes of child mortality in 2000–13, with projections to inform post-2015 priorities: an updated systematic analysis. Lancet 2015;385:430–440.
- Walker CL, Rudan I, Liu L, et al. Global burden of childhood pneumonia and diarrhoea. Lancet 2013;381:1405–1416.
- Kapikian AZ, Wyatt RG, Dolin R, et al. Visualization by immune electron microscopy of a 27-nm particle associated with acute infectious nonbacterial gastroenteritis. J Virol 1972;10(5):1075–1081.
- Bishop RF, Davidson GP, Holmes IH, Ruck BJ. Virus particles in epithelial cells of duodenal mucosa from children with acute non-bacterial gastroenteritis. Lancet 1973;2:1281–1283.
- 5. Robilotti E, Deresinski S, Pinsky BA. Norovirus. Clin Microbiol Rev 2015;28(1):134–164.
- Buss SN, Leber A, Chapin K, et al. Multicenter evaluation of the BioFire FilmArray gastrointestinal panel for the etiologic diagnosis of infectious gastroenteritis. J Clin Microbiol 2015;53(3):915–925.
- Chhabra P, Payne DC, Szilagyi PG, et al. Etiology of viral gastroenteritis in children <5 years of age in the United States, 2008–2009. J Infect Dis 2013;208(5):790–800.
- 8. Patel MM, Glass R, Desai R, et al. Fulfilling the promise of rotavirus vaccines: how far have we come since licensure? Lancet Infect Dis 2012;12(7):561–570.
- Gastanaduy PA, Sanchez-Uribe E, Esparza-Aguilar M, et al. Effect of rotavirus vaccine on diarrhea mortality in different socioeconomic regions of Mexico. Pediatrics 2013;131(4):e1115–e1120.
- do Carmo GM, Yen C, Cortes J, et al. Decline in diarrhea mortality and admissions after routine childhood rotavirus immunization in Brazil: a time-series analysis. PLoS Med 2011;8(4):e1001024.
- Payne DC, Vinje J, Szilagyi PG, et al. Norovirus and medically attended gastroenteritis in U.S. children. N Engl J Med 2013;368(12):1121–1130.
- Leshem E, Moritz RE, Curns AT, et al. Rotavirus vaccines and health care utilization for diarrhea in the United States (2007–2011). Pediatrics 2014;134(1):15–23.
- Kapikian A, Chanock RM. Rotaviruses. In: Field BN, Knipe DM, Howley PM (eds) Fields Virology, 3rd ed. Philadelphia, Lippincott-Raven, 1996, pp 1657–1708.
- Matthijnssens J, Ciarlet M, Rahman M, et al. Recommendations for the classification of group A rotaviruses using all 11 genomic RNA segments. Arch Virol 2008;153(8):1621–1629.
- Kapikian A, Estes M, Chanock RM. Norwalk group of viruses. In: Fields BN, Knipe DM, Howley PM (eds) Fields Virology, 3rd ed. Philadelphia, Lippincott-Raven, 1996.
- Vinje J. Advances in laboratory methods for detection and typing of norovirus. J Clin Microbiol 2015;53(2):373–381.
- Siebenga JJ, Vennema H, Zheng DP, et al. Norovirus illness is a global problem: emergence and spread of norovirus GII.4 variants, 2001–2007. J Infect Dis 2009;200(5):802–812.
- Lopman B, Vennema H, Kohli E, et al. Increase in viral gastroenteritis outbreaks in Europe and epidemic spread of new norovirus variant. Lancet 2004;363:682–688.
- 19. Siebenga JJ, Vennema H, Renckens B, et al. Epochal evolution of GGII.4 norovirus capsid proteins from 1995 to 2006. J Virol 2007;81(18):9932–9941.
- Leshem E, Wikswo M, Barclay L, et al. Effects and clinical significance of GII.4 Sydney norovirus, United States, 2012–2013. Emerg Infect Dis 2013;19(8):1231–1238.
- Patel MM, Widdowson MA, Glass RI, et al. Systematic literature review of role of noroviruses in sporadic gastroenteritis. Emerg Infect Dis 2008;14(8):1224–1231.
- Jones MK, Watanabe M, Zhu S, et al. Enteric bacteria promote human and mouse norovirus infection of B cells. Science 2014;346(6210):755–759.
- De Benedictis P, Schultz-Cherry S, Burnham A, Cattoli G. Astrovirus infections in humans and animals: molecular biology, genetic diversity, and interspecies transmissions. Infect Genet Evol 2011;11(7):1529–1544.
- Tseng WC, Wu FT, Hsiung CA, et al. Astrovirus gastroenteritis in hospitalized children of less than 5 years of age in Taiwan, 2009. J Microbiol Immunol Infect 2012;45(4):311–317.
- De Grazia S, Platia MA, Rotolo V, et al. Surveillance of human astrovirus circulation in Italy 2002-2005: emergence of lineage 2c strains. Clin Microbiol Infect 2011;17(1):97–101.
- Lion T. Adenovirus infections in immunocompetent and immunocompromised patients. Clin Microbiol Rev 2014;27(3):441–462.
- 27. de Jong JC, Wigand R, Kidd AH, et al. Candidate adenoviruses 40 and 41: fastidious adenoviruses from human infant stool. J Med Virol 1983;11(3):215–231.
- Wong TW, Lee CK, Tam W, et al. Cluster of SARS among medical students exposed to single patient, Hong Kong. Emerg Infect Dis 2004;10(2):269–276.
- Assiri A, Al-Tawfiq JA, Al-Rabeeah AA, et al. Epidemiological, demographic, and clinical characteristics of 47 cases of Middle East respiratory syndrome coronavirus disease from Saudi Arabia: a descriptive study. Lancet Infect Dis 2013;13(9):752–761.
 Kapikian AZ. Viral gastroenteritis. JAMA 1993;269(5):627–630.
- Lopman BA, Adak GK, Reacher MH, Brown DW. Two epidemiologic patterns of norovirus outbreaks: surveillance in England and wales, 1992–2000. Emerg Infect Dis 2003;9(1):71–77.
- Hall AJ, Lopman BA, Payne DC, et al. Norovirus disease in the United States. Emerg Infect Dis 2013;19(8):1198–1205.
- Hall AJ, Wikswo ME, Pringle K, et al. Vital signs: foodborne norovirus outbreaks— United States, 2009–2012. MMWR Morb Mortal Wkly Rep 2014;63:491–495.
- 34. Marks PJ, Vipond IB, Carlisle D, et al. Evidence for airborne transmission of Norwalklike virus (NLV) in a hotel restaurant. Epidemiol Infect 2000;124(3):481–487.
- Marks PJ, Vipond IB, Regan FM, et al. A school outbreak of Norwalk-like virus: evidence for airborne transmission. Epidemiol Infect 2003;131(1):727–736.

- 36. Kirking HL, Cortes J, Burrer S, et al. Likely transmission of norovirus on an airplane, October 2008. Clin Infect Dis 2010;50(9):1216–1221.
- Simmons K, Gambhir M, Leon J, Lopman B. Duration of immunity to norovirus gastroenteritis. Emerg Infect Dis 2013;19(8):1260–1267.
- Mitchell DK, Van R, Morrow AL, et al. Outbreaks of astrovirus gastroenteritis in day care centers. J Pediatr 1993;123(5):725–732.
- Patel MM, Pitzer VE, Alonso WJ, et al. Global seasonality of rotavirus disease. Pediatr Infect Dis J 2013;32(4):e134–e147.
- 40. Tate JE, Haynes A, Payne DC, et al. Trends in national rotavirus activity before and after introduction of rotavirus vaccine into the national immunization program in the United States, 2000 to 2012. Pediatr Infect Dis J 2013;32(7):741–744.
- 41. Pitzer VE, Viboud C, Simonsen L, et al. Demographic variability, vaccination, and the spatiotemporal dynamics of rotavirus epidemics. Science 2009;325:290–294.
- Mwenda JM, Ntoto KM, Abebe A, et al. Burden and epidemiology of rotavirus diarrhea in selected African countries: preliminary results from the African Rotavirus Surveillance Network. J Infect Dis 2010;202:S5–S11.
- Hashizume M, Armstrong B, Wagatsuma Y, et al. Rotavirus infections and climate variability in Dhaka, Bangladesh: a time-series analysis. Epidemiol Infect 2008;136(9):1281–1289.
- Mounts AW, Ando T, Koopmans M, et al. Cold weather seasonality of gastroenteritis associated with Norwalk-like viruses. J Infect Dis 2000;181(suppl 2):S284–S287.
- Ahmed SM, Lopman BA, Levy K. A systematic review and meta-analysis of the global seasonality of norovirus. PLoS ONE 2013;8(10):e75922.
- Lee TW, Kurtz JB. Prevalence of human astrovirus serotypes in the Oxford region 1976–92, with evidence for two new serotypes. Epidemiol Infect 1994;112(1):187–193.
- Bates PR, Bailey AS, Wood DJ, et al. Comparative epidemiology of rotavirus, subgenus F (types 40 and 41) adenovirus and astrovirus gastroenteritis in children. J Med Virol 1993;39(3):224–228.
- Velazquez FR, Matson DO, Calva JJ, et al. Rotavirus infections in infants as protection against subsequent infections. N Engl J Med 1996;335(14):1022–1028.
- Parashar UD, Hummelman EG, Bresee JS, et al. Global illness and deaths caused by rotavirus disease in children. Emerg Infect Dis 2003;9(5):565–572.
- Kotloff KL, Nataro JP, Blackwelder WC, et al. Burden and aetiology of diarrhoeal disease in infants and young children in developing countries (the Global Enteric Multicenter Study, GEMS): a prospective, case-control study. Lancet 2013;382:209–222.
- Oyejide CO, Fagbami AH. An epidemiological study of rotavirus diarrhoea in a cohort of Nigerian infants: II. Incidence of diarrhoea in the first two years of life. Int J Epidemiol 1988;17(4):908–912.
- Cardemil CV, Cortese MM, Medina-Marino A, et al. Two rotavirus outbreaks caused by genotype G2P[4] at large retirement communities: cohort studies. Ann Intern Med 2012;157(9):621–631.
- Bresee J, Fang ZY, Wang B, et al. First report from the Asian Rotavirus Surveillance Network. Emerg Infect Dis 2004;10(6):988–995.
- 54. Tate JE, Burton AH, Boschi-Pinto C, et al. 2008 estimate of worldwide rotavirusassociated mortality in children younger than 5 years before the introduction of universal rotavirus vaccination programmes: a systematic review and meta-analysis. Lancet Infect Dis 2012;12(2):136–141.
- Harris JP, Jit M, Cooper D, Edmunds WJ. Evaluating rotavirus vaccination in England and Wales. Part I. Estimating the burden of disease. Vaccine 2007;25:3962–3970.
- Esposito DH, Holman RC, Haberling DL, et al. Baseline estimates of diarrheaassociated mortality among United States children before rotavirus vaccine introduction. Pediatr Infect Dis J 2011;30:942–947.
- Glass RI, Parashar UD, Estes MK. Norovirus gastroenteritis. N Engl J Med 2009;361:1776–1785.
- Ahmed SM, Hall AJ, Robinson AE, et al. Global prevalence of norovirus in cases of gastroenteritis: a systematic review and meta-analysis. Lancet Infect Dis 2014;14(8):725–730.
- Becker-Dreps S, Bucardo F, Vilchez S, et al. Etiology of childhood diarrhea after rotavirus vaccine introduction: a prospective, population-based study in Nicaragua. Pediatr Infect Dis J 2014;33:1156–1163.
- 60. Hemming M, Rasanen S, Huhti L, et al. Major reduction of rotavirus, but not norovirus, gastroenteritis in children seen in hospital after the introduction of RotaTeq vaccine into the National Immunization Programme in Finland. Eur J Pediatr 2013;172(6):739–746.
- Phillips G, Tam CC, Conti S, et al. Community incidence of norovirus-associated infectious intestinal disease in England: improved estimates using viral load for norovirus diagnosis. Am J Epidemiol 2010;171(9):1014–1022.
- Lopman BA, Reacher MH, van Duijnhoven Y, et al. Viral gastroenteritis outbreaks in Europe, 1995–2000. Emerg Infect Dis 2003;9(1):90–96.
- Pang XL, Joensuu J, Vesikari T. Human calicivirus–associated sporadic gastroenteritis in Finnish children less than two years of age followed prospectively during a rotavirus vaccine trial. Pediatr Infect Dis J 1999;18(5):420–426.
- 64. Sakai Y, Nakata S, Honma S, et al. Clinical severity of Norwalk virus and Sapporo virus gastroenteritis in children in Hokkaido, Japan. Pediatr Infect Dis J 2001;20(9):849–853.
- Lopman BA, Brown DW, Koopmans M. Human caliciviruses in Europe. J Clin Virol 2002;24(3):137–160.
- 66. Amar CF, East CL, Gray J, et al. Detection by PCR of eight groups of enteric pathogens in 4,627 faecal samples: re-examination of the English case-control Infectious Intestinal Disease Study (1993–1996). Eur J Clin Microbiol Infect Dis 2007;26(5):311–323.
- Pang XL, Honma S, Nakata S, Vesikari T. Human caliciviruses in acute gastroenteritis of young children in the community. J Infect Dis 2000;181(suppl 2):S288–S294.

- Kotloff KL, Herrmann JE, Blacklow NR, et al. The frequency of astrovirus as a cause of diarrhea in Baltimore children. Pediatr Infect Dis J 1992;11(7):587–589.
- Mitchell DK, Matson DO, Cubitt WD, et al. Prevalence of antibodies to astrovirus types 1 and 3 in children and adolescents in Norfolk, Virginia. Pediatr Infect Dis J 1999;18(3):249–254.
- 70. Oishi I, Yamazaki K, Kimoto T, et al. A large outbreak of acute gastroenteritis associated with astrovirus among students and teachers in Osaka, Japan. J Infect Dis 1994;170(2):439–443.
- Unicomb LE, Banu NN, Azim T, et al. Astrovirus infection in association with acute, persistent and nosocomial diarrhea in Bangladesh. Pediatr Infect Dis J 1998;17(7):611–614.
- Ford-Jones EL, Mindorff CM, Gold R, Petric M. The incidence of viralassociated diarrhea after admission to a pediatric hospital. Am J Epidemiol 1990;131(4):711–718.
- Kim KH, Yang JM, Joo SI, et al. Importance of rotavirus and adenovirus types 40 and 41 in acute gastroenteritis in Korean children. J Clin Microbiol 1990;28:2279–2284.
- 74. Wheeler JG, Sethi D, Cowden JM, et al. Study of infectious intestinal disease in England: rates in the community, presenting to general practice, and reported to national surveillance. The Infectious Intestinal Disease Study Executive. BMJ 1999;318:1046–1050.
- 75. Yolken RH, Lawrence F, Leister F, et al. Gastroenteritis associated with enteric type adenovirus in hospitalized infants. J Pediatr 1982;101(1):21–26.
- Tompkins DS, Hudson MJ, Smith HR, et al. A study of infectious intestinal disease in England: microbiological findings in cases and controls. Commun Dis Public Health 1999;2(2):108–113.
- Kotloff KL, Losonsky GA, Morris JG Jr, et al. Enteric adenovirus infection and childhood diarrhea: an epidemiologic study in three clinical settings. Pediatrics 1989;84(2):219–225.
- Mavromichalis J, Evans N, McNeish AS, et al. Intestinal damage in rotavirus and adenovirus gastroenteritis assessed by d-xylose malabsorption. Arch Dis Child 1977;52(7):589–591.
- 79. Blutt SE, Kirkwood CD, Parreno V, et al. Rotavirus antigenaemia and viraemia: a common event? Lancet 2003;362:1445–1449.
- Takanashi S, Hashira S, Matsunaga T, et al. Detection, genetic characterization, and quantification of norovirus RNA from sera of children with gastroenteritis. J Clin Virol 2009;44(2):161–163.
- Lopman B, Simmons K, Gambhir M, et al. Epidemiologic implications of asymptomatic reinfection: a mathematical modeling study of norovirus. Am J Epidemiol 2014;179(4):507–512.
- Velazquez FR, Matson DO, Guerrero ML, et al. Serum antibody as a marker of protection against natural rotavirus infection and disease. J Infect Dis 2000;182(6):1602–1609.
- Fischer TK, Valentiner-Branth P, Steinsland H, et al. Protective immunity after natural rotavirus infection: a community cohort study of newborn children in Guinea-Bissau, West Africa. J Infect Dis 2002;186(5):593–597.
- Parrino TA, Schreiber DS, Trier JS, et al. Clinical immunity in acute gastroenteritis caused by Norwalk agent. N Engl J Med 1977;297(2):86–89.
- Hutson AM, Atmar RL, Graham DY, Estes MK. Norwalk virus infection and disease is associated with ABO histo-blood group type. J Infect Dis 2002;185(9): 1335–1337.
- Lindesmith L, Moe C, Marionneau S, et al. Human susceptibility and resistance to Norwalk virus infection. Nat Med 2003;9(5):548–553.
- Marionneau S, Ruvoen N, Le Moullac-Vaidye B, et al. Norwalk virus binds to histo-blood group antigens present on gastroduodenal epithelial cells of secretor individuals. Gastroenterology 2002;122(7):1967–1977.
- Tan M, Jin M, Xie H, et al. Outbreak studies of a GII-3 and a GII-4 norovirus revealed an association between HBGA phenotypes and viral infection. J Med Virol 2008;80(7):1296–1301.
- Payne DC, Parashar UD, Lopman BA. Developments in understanding acquired immunity and innate susceptibility to norovirus and rotavirus gastroenteritis in children. Curr Opin Pediatr 2015;27(1):105–109.
- Nordgren J, Sharma S, Bucardo F, et al. Both Lewis and secretor status mediate susceptibility to rotavirus infections in a rotavirus genotype-dependent manner. Clin Infect Dis 2014;59:1567–1573.
- Van Trang N, Vu HT, Le NT, et al. Association between norovirus and rotavirus infection and histo-blood group antigen types in Vietnamese children. J Clin Microbiol 2014;52(5):1366–1374.
- Imbert-Marcille BM, Barbe L, Dupe M, et al. A FUT2 gene common polymorphism determines resistance to rotavirus A of the P[8] genotype. J Infect Dis 2014;209(8):1227–1230.
- Hutson AM, Atmar RL, Estes MK. Norovirus disease: changing epidemiology and host susceptibility factors. Trends Microbiol 2004;12(6):279–287.
- 94. Larsson MM, Rydell GE, Grahn A, et al. Antibody prevalence and titer to norovirus (genogroup II) correlate with secretor (FUT2) but not with ABO phenotype or Lewis (FUT3) genotype. J Infect Dis 2006;194(10):1422–1427.
- Lindesmith L, Moe C, Lependu J, et al. Cellular and humoral immunity following Snow Mountain virus challenge. J Virol 2005;79(5):2900–2909.
- Rockx B, De Wit M, Vennema H, et al. Natural history of human calicivirus infection: a prospective cohort study. Clin Infect Dis 2002;35(3):246–253.
- Liu P, Wang X, Lee JC, et al. Genetic susceptibility to norovirus GII.3 and GII.4 infections in Chinese pediatric diarrheal disease. Pediatr Infect Dis J 2014;33(11):e305–e309.
- 98. Kaplan JE, Feldman R, Campbell DS, et al. The frequency of a Norwalk-like pattern of illness in outbreaks of acute gastroenteritis. Am J Public Health 1982;72(12):1329–1332.

- Le Saux N, Bettinger JA, Halperin SA, et al., Canadian Immunization Monitoring Program A. Substantial morbidity for hospitalized children with communityacquired rotavirus infections: 2005–2007 IMPACT surveillance in Canadian hospitals. Pediatr Infect Dis J 2010;29(9):879–882.
- Payne DC, Baggs J, Zerr DM, et al. Protective association between rotavirus vaccination and childhood seizures in the year following vaccination in US children. Clin Infect Dis 2014;58(2):173–177.
- Capizzi T, Makari-Judson G, Steingart R, Mertens WC. Chronic diarrhea associated with persistent norovirus excretion in patients with chronic lymphocytic leukemia: report of two cases. BMC Infect Dis 2011;11:131.
- Grohmann GS, Glass RI, Pereira HG, et al. Enteric viruses and diarrhea in HIVinfected patients. Enteric Opportunistic Infections Working Group. N Engl J Med 1993;329(1):14–20.
- 103. Liakopoulou E, Mutton K, Carrington D, et al. Rotavirus as a significant cause of prolonged diarrhoeal illness and morbidity following allogeneic bone marrow transplantation. Bone Marrow Transplant 2005;36(8):691–694.
- Mans J, Murray TY, Kiulia NM, et al. Human caliciviruses detected in HIVseropositive children in Kenya. J Med Virol 2014;86(1):75–81.
- Green KY. Norovirus infection in immunocompromised hosts. Clin Microbiol Infect 2014;20(8):717–723.
- Turcios RM, Widdowson MA, Sulka AC, et al. Reevaluation of epidemiological criteria for identifying outbreaks of acute gastroenteritis due to norovirus: United States, 1998–2000. Clin Infect Dis 2006;42(7):964–969.
- 107. Tate JE, Mijatovic-Rustempasic S, Tam KI, et al. Comparison of 2 assays for diagnosing rotavirus and evaluating vaccine effectiveness in children with gastroenteritis. Emerg Infect Dis 2013;19(8):1245–1252.
- Vega E, Barclay L, Gregoricus N, et al. Genotypic and epidemiologic trends of norovirus outbreaks in the United States, 2009 to 2013. J Clin Microbiol 2014;52(1):147–155.
- Phillips G, Lopman B, Tam CC, et al. Diagnosing norovirus-associated infectious intestinal disease using viral load. BMC Infect Dis 2009;9:63.
- Duizer E, Pielaat A, Vennema H, et al. Probabilities in norovirus outbreak diagnosis. J Clin Virol 2007;40(1):38–42.
- Costantini V, Grenz L, Fritzinger A, et al. Diagnostic accuracy and analytical sensitivity of IDEIA norovirus assay for routine screening of human norovirus. J Clin Microbiol 2010;48(8):2770–2778.
- 112. Gray JJ, Kohli E, Ruggeri FM, et al. European multicenter evaluation of commercial enzyme immunoassays for detecting norovirus antigen in fecal samples. Clin Vaccine Immunol 2007;14(10):1349–1355.
- King CK, Glass R, Bresee JS, Duggan C. Managing acute gastroenteritis among children: oral rehydration, maintenance, and nutritional therapy. MMWR Recomm Rep 2003;52(RR-16):1–16.
- MacGillivray S, Fahey T, McGuire W. Lactose avoidance for young children with acute diarrhoea. Cochrane Database Syst Rev 2013;(10):CD005433.
- Majamaa H, Isolauri E, Saxelin M, Vesikari T. Lactic acid bacteria in the treatment of acute rotavirus gastroenteritis. J Pediatr Gastroenterol Nutr 1995;20(3):333–338.
- 116. Isolauri E, Joensuu J, Suomalainen H, et al. Improved immunogenicity of oral D x RRV reassortant rotavirus vaccine by *Lactobacillus casei* GG. Vaccine 1995;13(3):310–312.
- Szajewska H, Ruszczynski M, Kolacek S. Meta-analysis shows limited evidence for using *Lactobacillus acidophilus* LB to treat acute gastroenteritis in children. Acta Paediatr 2014;103(3):249–255.
- Tielsch JM, Khatry SK, Stoltzfus RJ, et al. Effect of daily zinc supplementation on child mortality in southern Nepal: a community-based, cluster randomised, placebo-controlled trial. Lancet 2007;370:1230–1239.
- Lazzerini M, Ronfani L. Oral zinc for treating diarrhoea in children. Cochrane Database Syst Rev 2013;(1):CD005436.
- Guerrant RL, Van Gilder T, Steiner TS, et al. Practice guidelines for the management of infectious diarrhea. Clin Infect Dis 2001;32(3):331–351.
- Tate JE, Parashar UD. Rotavirus vaccines in routine use. Clin Infect Dis 2014;59(9):1291–1301.
- 122. Bhandari N, Rongsen-Chandola T, Bavdekar A, et al. Efficacy of a monovalent human-bovine (116E) rotavirus vaccine in Indian infants: a randomised, doubleblind, placebo-controlled trial. Lancet 2014;383:2136–2143.
- PATH. Country Introductions of Rotavirus Vaccines. 2015. http://sites.path.org/ rotavirusvaccine/country-introduction-maps-and-spreadsheet/>.
- 124. Armah GE, Sow SO, Breiman RF, et al. Efficacy of pentavalent rotavirus vaccine against severe rotavirus gastroenteritis in infants in developing countries in sub-Saharan Africa: a randomised, double-blind, placebo-controlled trial. Lancet 2010;376:606–614.
- 125. Zaman K, Dang DA, Victor JC, et al. Efficacy of pentavalent rotavirus vaccine against severe rotavirus gastroenteritis in infants in developing countries in Asia: a randomised, double-blind, placebo-controlled trial. Lancet 2010;376:615–623.
- Madhi SA, Cunliffe NA, Steele D, et al. Effect of human rotavirus vaccine on severe diarrhea in African infants. N Engl J Med 2010;362(4):289–298.
- Vesikari T, Matson DO, Dennehy P, et al. Safety and efficacy of a pentavalent humanbovine (WC3) reassortant rotavirus vaccine. N Engl J Med 2006;354(1):23–33.
- Ruiz-Palacios GM, Perez-Schael I, Velazquez FR, et al. Safety and efficacy of an attenuated vaccine against severe rotavirus gastroenteritis. N Engl J Med 2006;354(1):11–22.
- World Health Organization. Rotavirus vaccines: an update. Wkly Epidemiol Rec 2009;84(50):533–540.
- Payne DC, Boom JA, Staat MA, et al. Effectiveness of pentavalent and monovalent rotavirus vaccines in concurrent use among US children <5 years of age, 2009–2011. Clin Infect Dis 2013;57(1):13–20.

- 131. Leshem E, Lopman B, Glass R, et al. Distribution of rotavirus strains and strainspecific effectiveness of the rotavirus vaccine after its introduction: a systematic review and meta-analysis. Lancet Infect Dis 2014;14(9):847–856.
- 132. Centers for Disease Control and Prevention. Updated norovirus outbreak management and disease prevention guidelines. MMWR Recomm Rep 2011;60(RR-3):1–18.
- Parashar U, Quiroz ES, Mounts AW, et al. Norwalk-like viruses: public health consequences and outbreak management. MMWR Recomm Rep 2001;50(RR-9):1–17.
- 134. Atmar RL, Bernstein DI, Harro CD, et al. Norovirus vaccine against experimental human Norwalk Virus illness. N Engl J Med 2011;365:2178–2187.
- Bernstein DI, Atmar RL, Lyon GM, et al. Norovirus vaccine against experimental human GII.4 virus illness: a challenge study in healthy adults. J Infect Dis 2014;211(6):870–878.