

Oxford Nanopore Sequencing Reveals an Exotic *Broad bean mottle virus* Genome within Australian Grains Post-Entry Quarantine

Microbiology[®]

Resource Announcements

S. Maina,^{a,b} S. L. Norton,^a V. L. McQueen,^a S. King,^a B. Rodoni^{c,d}

AMERICAN SOCIETY FOR

MICROBIOLOGY

^aAustralian Grains Genebank, Agriculture Victoria Research, Horsham, Victoria, Australia ^bElizabeth Macarthur Agricultural Institute, NSW Department of Primary Industries, Menangle, New South Wales, Australia ^cMicrobial Pests & Diseases, Agriculture Victoria Research, AgriBio, Bundoora, Victoria, Australia ^dSchool of Applied Systems Biology, La Trobe University, Bundoora, Victoria, Australia

ABSTRACT A *Broad bean mottle virus* (BBMV) isolate (S52) obtained from an infected *Vicia faba* leaf sample from Syria was sequenced using Oxford Nanopore long-read sequencing at the Australian border. The genome had 95.6%, 98.2%, and 93.4% nucleotide sequence identity to BBMV strains RNA1 (Bawden), RNA2 (Mo), and RNA3 (Bawden).

B^{road} bean mottle virus (BBMV) (Bromovirus, Bromoviridae) is a tripartite linear singlestranded RNA (ssRNA) (+) seed-borne virus that infects several temperate pulses. It is found in Africa, Asia, Europe, and the Middle East, where it causes severe damage in pulses (1–3) and constitutes a threat to crop improvement programs (4). BBMV is a quarantinable pathogen in Australia; consequently, it is mandatory for the imported seeds to be screened within post-entry quarantine (PEQ) (5).

Isolate S52 was obtained from *Vicia faba* (faba bean) material imported from Syria in 2003 and mechanically inoculated into faba bean plants in a PEQ controlled glasshouse. The inoculated plants revealed mottling and chlorotic blotch symptoms; the leaves were sampled and tested positive for BBMV by tissue blot immunoassay (6). Total RNA was extracted from the positive leaf material using a ZR plant RNA miniprep kit (Zymo Research), and a quality control check was conducted as previously described (7). The RNA was converted to cDNA using random primers (8), followed by library preparation using a ligation sequencing kit (SQK-LSK109; ONT); the sequencing was conducted on the MinION platform using a FLO-MIN106D (R9.5) flow cell. Base calling was performed using MinKNOW version 20.10.3 in high accuracy mode. A total of 101,000 reads were generated, ranging in length from 1.2 to 5.3 kb, and imported into CLC Genomics Workbench (CLCGW) version 20 (CLC bio; Qiagen) for quality control, followed by *de novo* assembly using default settings with double polishing. In addition, the reads were mapped onto a reference BBMV genome using Minimap2 version 2.0.0 (9).

The consensus genome segments had 351 to 610 reads mapping onto each entire coding genome segment, with average depths of $22 \times$, $31 \times$, and $48 \times$ and GC contents of 43%, 42%, and 44% for RNA1 to RNA3, respectively. The consensus sequence was subjected to a BLASTN search using BLAST+ version 2.7 and MUSCLE (10, 11). The new genome had 95.6%, 98.2%, and 93.4% nucleotide identity (nt) to BBMV strains RNA1 (Bawden), RNA2 (Mo), and RNA3 (Bawden), respectively (12, 13). Primers were designed from the newly generated BBMV RNA3 coat protein (CP) region, targeting 250 to 500 bp, and amplified the expected amplicon band size. Annotation of the BBMV genome (5) revealed that both RNA1 and RNA2 encoded open reading frames 1a and 2a, typical of the genus *Bromovirus*, to which BBMV belongs (13). Further, RNA3 encoded the CP gene, which is required for virus systemic movement and associated with cell-to-cell spread (14). The 93.4% nt between S52 and Bawden suggests that the CP might be a hotspot for BBMV genome variability. Nevertheless,

Editor Jelle Matthijnssens, KU Leuven Copyright © 2022 Maina et al. This is an openaccess article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to S. Maina, solomon.maina@dpi.nsw.gov.au. The authors declare no conflict of interest. Received 28 February 2022 Accepted 11 May 2022 Published 31 May 2022 considering the data quality score (15), this deduction should be confirmed further by Illumina reads.

BBMV vectors have not been reported in Australia, and imported germplasm remains the major introduction pathway. Imported seeds serve as a source of genetic diversity for Australian grain-breeding programs. Therefore, it is plausible that the importation of this new germplasm into Australia poses a significant threat in introducing damaging exotic viruses and their variants. As such, persistent surveillance and the integration of robust diagnostic genomics tools at the PEQ border are vital in safeguarding the Australian grains industry.

Data availability. The data described here were deposited in the DDBJ under accession numbers LC683787 to LC683789. The raw reads were deposited at the SRA under accession number SRR16883245 and BioProject accession number PRJNA778895.

ACKNOWLEDGMENTS

This research was funded by the Grains Research and Development Corporation and the Department of Jobs, Precincts and Regions, Victoria.

REFERENCES

- Bawden FC, Chaudhuri RP, Kassanis B. 1951. Some properties of broad bean mottle virus. Ann Applied Biology 38:774–784. https://doi.org/10.1111/ j.1744-7348.1951.tb07849.x.
- Walters HJ, Surin P. 1973. Transmission and host range studies of broad bean mottle virus. Plant Dis Reporter 57:833–836.
- Makkouk KM, Bos L, Azzam O, Koumari S, Rizkallah A. 1988. Survey of viruses affecting faba bean in six Arab countries. Arab J Plant Prot 6:61–63.
- Fortass M, Diallo S. 1993. Broad bean mottle virus in Morocco; curculionid vectors, and natural occurrence in food legumes other than faba bean (*Vicia faba*). Netherlands J Plant Pathol 99:219–226. https://doi.org/10.1007/BF01974666.
- Maina S, Zheng L, King S, McQueen VL, Norton SL, Rodoni B. 2020. Transcriptome sequencing reveals the genome sequence of *Pea early browning virus* from a 29-year-old faba bean sample. Microbiol Resour Announc 9:e00673-20. https://doi.org/10.1128/MRA.00673-20.
- Makkouk KM, Rizkallah L, Madkour M, El-Sherbeeny M, Kumari SG, Amriti AW, Solh MB. 1994. Survey of faba bean (*Vicia faba*) for viruses in Egypt. Phytopathol Mediterr 33:207–211.
- Maina S, Coutts BA, Edwards OR, de Almeida L, Ximenes A, Jones RAC. 2017. Papaya ringspot virus populations from East Timorese and northern Australian cucurbit crops: biological and molecular properties, and absence of genetic connectivity. Plant Dis 101:985–993. https://doi.org/10.1094/PDIS-10-16-1499-RE.
- Maina S, Zheng L, Rodoni BC. 2021. Targeted genome sequencing (TG-Seq) approaches to detect plant viruses. Viruses 13:583. https://doi.org/10.3390/ v13040583.

- Li H. 2018. Minimap2: pairwise alignment for nucleotide sequences. Bioinformatics 34:3094–3100. https://doi.org/10.1093/bioinformatics/bty191.
- Altschul SF, Madden TL, Schäffer AA, Zhang J, Zhang Z, Miller W, Lipman DJ. 1997. Gapped BLAST and PSI-BLAST: a new generation of protein database search programs. Nucleic Acids Res 25:3389–3402. https://doi.org/10.1093/ nar/25.17.3389.
- 11. Edgar RC. 2004. MUSCLE: a multiple sequence alignment method with reduced time and space complexity. BMC Bioinformatics 5:113. https://doi .org/10.1186/1471-2105-5-113.
- Dzianott AM, Bujarski JJ. 1991. The nucleotide sequence and genome organization of the RNA-1 segment in two bromoviruses: broad bean mottle virus and cowpea chlorotic mottle virus. Virology 185:553–562. https://doi.org/10.1016/ 0042-6822(91)90525-G.
- Sandoval C, Pogany J, Bujarski J, Romero J. 2008. Use of a defective RNA of broad bean mottle bromovirus for stable gene expression in legumes. Arch Virol 153:1755–1758. https://doi.org/10.1007/s00705-008-0174-y.
- Bujarski J, Gallitelli D, García-Arenal F, Pallás V, Palukaitis P, Reddy MK, Wang A. 2019. ICTV virus taxonomy profile: Bromoviridae. J Gene Virol 100:1206–1207. https://doi.org/10.1099/jgv.0.001282.
- Laver T, Harrison J, O'Neill PA, Moore K, Farbos A, Paszkiewicz K, Studholme DJ. 2015. Assessing the performance of the Oxford Nanopore Technologies MinION. Biomol Detect Quantif 3:1–8. https://doi.org/10.1016/j.bdq.2015 .02.001.