

EDITORIAL

An oversimplification of physiological principles leads to flawed macroecological analyses

In light of the rapidly changing climate, there is an urgent need to develop a mechanistic understanding of how physiological functioning mediates ecological patterns. Recently, there has been a spate of papers using analyses that scale up from a standard physiological model, the Scholander–Irving model, to make predictions about range constraints on endothermic vertebrates (Buckley, Khaliq, Swanson, & Hof, 2018; Fristoe et al., 2015; Khaliq, Böhning-Gaese, Prinzing, Pfenninger, & Hof, 2017; Khaliq, Hof, Prinzing, Böhning-Gaese, & Pfenninger, 2014). Here, we argue that oversimplifications of the Scholander–Irving model and the use of questionable datasets lead to questionable macrophysiological analyses. Many of these problems have been addressed elsewhere, directly and indirectly (e.g., McKechnie, Coe, Gerson, & Wolf, 2017; Mitchell et al., 2018), although the focus has largely been on the applicability of the Scholander–Irving model to warm environmental temperatures, which are often seen as more relevant to climate change. However, one specific aspect of the Scholander–Irving model, the assumption that energy expenditure of an endotherm below the thermoneutral zone (TNZ) can be described by basic Newtonian physics, has been used incorrectly in several papers. While not the only paper based on this assumption, the recent work by Buckley et al. (2018) reinvigorated discussions among physiologists about improper interpretations of the Scholander–Irving model. Our concerns are not new and have been voiced repeatedly in the past (Calder & Schmidt-Nielsen, 1967; King, 1964; Tracy, 1972), but many of these ideas seem to have been buried by time. Our goal here is to bring these concerns back to the forefront in the context of modern large-scale macrophysiological analyses, using the work of Buckley et al. (2018) as an example where relevant. We detail these ideas below, but King (1964) provided a scathing, yet technically accurate summary of our position over five decades ago: “The convenience of the Newtonian model as a heuristic or pedagogical device is readily apparent; but its use as an analytical instrument to reveal relatively small interspecific or seasonal adaptations in metabolism is a practice which is beset by many uncertainties, and which occasionally appears to encourage a Procrustean fitting of data.”

In practice, endothermic vertebrates pose significant challenges in large-scale ecological analyses because the relationship between environmental conditions and functional energetics is mediated by complex metabolic and thermoregulatory control (Levesque,

Nowack, & Stawski, 2016). At the heart of the issue is the reliance on the Scholander–Irving model (Scholander, Hock, Walters, Johnson, & Irving, 1950), a classic descriptive model of the relationship between ambient temperature and metabolic rate *in strict homeotherms* (i.e., species that maintain their body temperature within a somewhat narrow, although undefined range). Under this model, homeothermic endotherms are assumed to maintain a constant basal metabolic rate within the TNZ, a constant body temperature (T_b), and a constant thermal conductance. At ambient temperatures below the lower critical temperature (T_{lc} , the lower boundary of the TNZ), metabolic rate increases to compensate for increased heat loss and to maintain constant T_b . Importantly, these relationships vary with many factors, including season, so values measured during summer are inappropriate for analyses of cold tolerance during winter. While the Scholander–Irving model is important for descriptive analyses of energetic function in homeothermic endotherms, its direct application to modeling environmental temperature thresholds for most endotherms is questionable (Levesque et al., 2016; Mitchell et al., 2018; Porter & Kearney, 2009). To generalize across large geographic scales, many analyses of endothermic energetics rely on a series of simplifying—and often unjustified—assumptions (reviewed by Mitchell et al., 2018). These simplifying assumptions are common, but an overreliance on them has inhibited a mechanistic understanding of global patterns in endothermic physiology. Relevant to the current discussion, the Scholander–Irving model predicts that a regression describing the relationship between metabolic rate and ambient temperatures below the T_{lc} extrapolates to an ambient temperature equal to T_b , if metabolic heat production was to reach zero (Scholander et al., 1950). This idea essentially requires that heat balance in endotherms follows Newton's laws of cooling, which may be a reasonable simplification in a small number of homeothermic species (usually mammals, but not birds), but is clearly not universal (King, 1964; McNab, 1979). There are numerous problems, both biological and computational, with this approach. First, a line fit through metabolic rate data rarely predicts T_b accurately, often overestimating it by as much as 10°C (Calder & Schmidt-Nielsen, 1967; McNab, 1979). As a simple example, we calculated T_b using the relationship between metabolic rate and T_{lc} for the rock pocket mouse (*Chaetodipus intermedius*), one of the species included in Buckley et al. (2018). In the source paper (Bradley, Yousef, & Scott, 1975), T_b is

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2019 The Authors. *Ecology and Evolution* published by John Wiley & Sons Ltd.

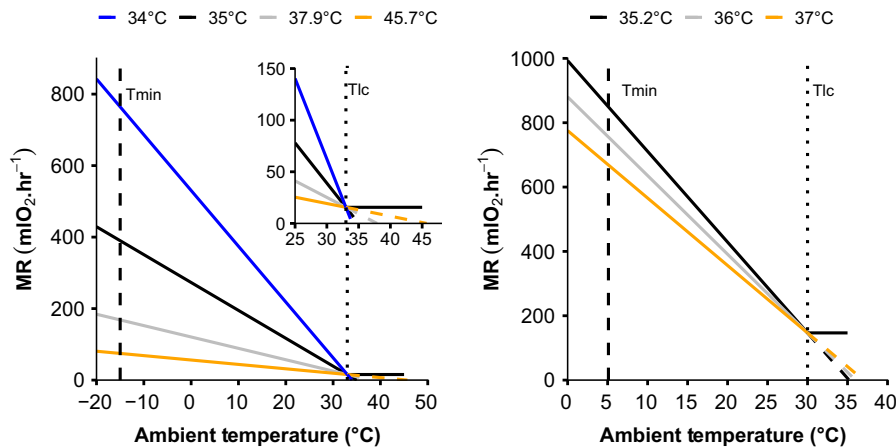


FIGURE 1 Demonstration of the inherent limitations of using body temperature (T_b) and the lower critical temperature (T_{lc}) of the thermal neutral zone to calculate thermal conductance (C) when T_b and T_{lc} are poorly defined. Buckley et al. (2018) analyzed metabolic expansibility, which is the metabolic rate at the range boundary (MR_{RB}) divided by basal metabolic rate (BMR). MR_{RB} , and therefore ME, changes drastically depending on assumptions of T_b and T_{lc} used to calculate C . Data are for a rock mouse (*Chaetodipus intermedius*, left; Bradley et al., 1975) and a hedgehog (*Aterix albiventris*, right; McNab, 1980). In each case, black lines represent the T_b value used by Buckley et al and the various other lines represent plausible assumptions about T_b , assuming T_{lc} is constant. Dashed lines represent the extrapolation of T_b from the MR line assuming the animal is perfectly following Newton's Laws of Cooling. The inset in (a) is an expanded view encapsulating the T_{lc} and T_b of the rock mouse

estimated at $\sim 35^\circ\text{C}$ and metabolic data are provided for two periods: January and April. By extrapolating the metabolic rate line to zero on the y-axis, T_b is estimated at 37.9°C in January and 45.7°C in April (Figure 1). Clearly, the April value is unrealistic for a mammal. The inverse assumption that T_b can be used to predict metabolic rate is also problematic. For example, a difference in estimate of only 1°C (the black line vs. the gray line on the left panel of Figure 1) for the T_b of the four-toed hedgehog (*Aterix albiventris*) would lead to a difference in the metabolic rate at the cold range boundary (MR_{CRB}) described by Buckley et al. (2018) of $\sim 12\%$.

The Scholander–Irving model was groundbreaking in 1950 and has proven to be useful in shaping our basic understanding of endothermic thermoregulation (Somero, 2013). However, this model requires that (a) thermal conductance below the TNZ is constant, (b) that endotherms maintain a relatively constant T_b , and (c) that T_{lc} can be estimated at a single value. The idea of a constant thermal conductance has long been discounted (Tracy, 1972) and has repeatedly been shown to be unrealistic (e.g., Calder & Schmidt-Nielsen, 1967; Noakes, Smit, Wolf, & McKechnie, 2013). Large-scale analyses assuming Newtonian cooling are on shaky theoretical footing if the assumption of constant thermal conductance is violated (King, 1964). Scholander et al. (1950) also assume that most mammals maintain T_b within $\pm 1^\circ\text{C}$. While the data are not strictly analogous, such homeothermy likely occurs in $<30\%$ of mammals with quality T_b data collected during winter (Boyles et al., 2013). Even strictly homeothermic humans show a decrease of $\sim 1^\circ\text{C}$ during sleep (Wright, Badia, Myers, Plenzler, & Hakel, 1997) and T_b of marsupials and monotremes can be lowered by $>8^\circ\text{C}$ during normothermic resting. Further, describing endothermic T_b is surprisingly difficult, and the most commonly used metric, mean T_b , is usually a poor descriptor of regulated endothermic T_b (Boyles, 2019; Hetem, Maloney, Fuller, & Mitchell, 2016).

Finally, T_{lc} is often difficult to delineate at a single temperature using standard respirometry techniques, because no clear “breakpoint” in metabolic rate exists for many species (McNab, 1995), and T_{lc} may change seasonally (Kobbe, Nowack, & Dausmann, 2014). Even ignoring these theoretical concerns, extreme care must be taken in extrapolating from such variable and hard-to-describe values.

If one begins with the assumption the Scholander–Irving model is static and easily fit for all endotherms, data quality issues are nearly inevitable. Here, we use the dataset of Buckley et al. (2018) as an example to demonstrate how poor assumptions can be manifested in poor data quality. Specifically, we traced T_b and T_{lc} data for a subset of their dataset back to the original sources. As with previous critiques of the upper critical temperature in similar datasets (McKechnie et al., 2017), we found considerable variability in the quality of the data used (Table 1). For example, many of the values presented were simply one value within the range of T_b or T_{lc} listed in the paper, but there was little consistency in how the value was chosen. This error stems from the assumption that endotherms maintain single, constant T_b s. Likewise, many older papers calculated T_{lc} by eyeballing lines through metabolic data and estimating the intersection with basal metabolic rate. Therefore, the values are approximations (in many cases, they are different than what we would estimate from the same data). This error stems from the assumption of a single, unchanging T_{lc} . Two of us (JGB and DLL) independently judged the quality of the T_b and T_{lc} included in the first 20 mammal species included by Buckley et al. (2018) (excluding species with partial data and including only one species from a genus; Table 1). Of those 20 datasets, we classified five as appropriate for inclusion in the analysis (i.e., data generally followed the Scholander–Irving model, and T_b variation was relatively low). Note that even among these “appropriate” datasets, data for three of

TABLE 1 Evaluation of data quality of the first 20 mammal species included in the analysis of Buckley et al. (2018)

| Species | T_b | T_{ic} | Original source | Comments | n | Wild/Zoo/Lab | Data quality |
|---------------------------------|-------|----------|--------------------------------|--|---------|--------------|--------------|
| <i>Abrothrix andinus</i> | 39.3 | 26.8 | Bozinovic and Rosenmann (1988) | T_b calculated working backwards estimates of MR. Actual measured T_b differs (and has a range) | 8 | Wild | Questionable |
| <i>Ailurus fulgens</i> | 37.6 | 25 | McNab (1988) | McNab argues that they do not follow standard homeotherm energetics; Scholander-Irving (S-I) model likely a bad model for this species | 2 | Zoo | Questionable |
| <i>Akodon azarae</i> | 37.7 | 30 | Dalby and Heath (1976) | T_b continues to drop with T_a , suggesting the S-I model might be inappropriate | 8 | Lab | Questionable |
| <i>Amospermophilus leucurus</i> | 37.2 | 31 | Dawson (1955) | T_b used was measured at T_a below T_{ic} presented; Range was 35.6–38.1°C, and taken during active period instead of the inactive period; animals were excitable and did not cope well with captivity | 12 | Wild | Questionable |
| <i>Anoura caudifer</i> | 36.3 | 26 | McNab (1969) | T_b in fig. 10 range from roughly 33 to 42°C with c.v. of 4.0% around the mean. T_{ic} was not referenced in the original paper, probably estimated from fig. 10. | Unknown | Wild | Questionable |
| <i>Aplodontia rufa</i> | 38 | 26.5 | McNab (1979) | Appears to follow the S-I model. There is some minor variation in T_b | 3 | Wild | Good |
| <i>Apodemus mystacinus</i> | 38.3 | 28 | Haim, Rubal, and Harari (1993) | Appears to follow the S-I model. Relatively low variation in T_b | 6 | Wild | Good |
| <i>Arctictis binturong</i> | 36 | 27 | McNab (1995) | McNab argues the core temperature is variable (~32–41°C) and decreases with ambient temperature; McNab also says "the zone of thermoneutrality... was especially difficult to define" | 2 | Zoo | Questionable |
| <i>Arctogalidia trivirgata</i> | 36.2 | 19 | McNab (1995) | The S-I model is inappropriate for this species and T_{ic} is impossible to define with the presented data. McNab says "the variability in rate of metabolism made the zone of thermoneutrality difficult to define with clarity" | 4 | Zoo | Poor |
| <i>Artibeus concolor</i> | 35 | 29 | McNab (1969) | Paper lists T_b of 35.3°C with a c.v. of 4.1%. No reference to T_{ic} , so probably estimated from fig. 17, and we would estimate it lower (probably about 28°C). | ? | Wild | Questionable |
| <i>Atelerix albigentris</i> | 35.2 | 30 | McNab (1980) | S-I model is probably appropriate, given the data presented herein | 2 | Zoo | Good |
| <i>Baiomys taylori</i> | 36 | 29 | Hudson (1965) | Original paper gives a wide range for T_b (32–36°C) and mentions T_{ic} at 29°C in the summary and 30–33°C in the text (but does not define how it was measured) | 10 | Lab | Poor |
| <i>Blarina brevicauda</i> | 38 | 25 | Neal and Lustick (1973) | Thermoneutral zone analyzed as the range within 95% confidence intervals, but the authors say metabolic rate increases below 30°C. T_b measured in separate experiments | 12 | Wild | Questionable |
| <i>Cabassous centralis</i> | 33.6 | 27.5 | McNab (1980) | S-I model is probably appropriate, given the data presented herein | 3 | Zoo | Good |
| <i>Caluromys derbianus</i> | 34 | 26.36 | McNab (1978) | Data were presented for 1 wild animal and 3 larger animals from a zoo. T_b is from the zoo animals (wild individual was 36°C). T_{ic} is given as a value somewhere between the zoo and wild individuals and is not mentioned in the paper | 4 | Wild/Zoo | Poor |
| <i>Canis latrans</i> | 36 | 22 | Golightly and Ohmart (1983) | T_b is difficult to determine in the original paper. Data come from a desert population. The original authors mention that animals near the northern edge of the range (which is far more relevant in the current study) have T_{ic} of ~10°C (Shield, 2009) | 3 | Wild | Poor |

(Continues)

TABLE 1 (Continued)

| Species | T_b | T_{lc} | Original source | Comments | n | Wild/Zoo/Lab | Data quality |
|-------------------------------|-------|----------|----------------------------------|--|---------|--------------|--------------|
| <i>Cannomys badius</i> | 36 | 26.74 | McNab (1979) | Original paper lists T_{lc} at 27°C. Otherwise, S-I model is probably appropriate, given the data presented herein | 5 | Zoo | Good |
| <i>Carollia perspicillata</i> | 36.6 | 29 | McNab (1969) | Paper lists T_b of 36.4°C with a c.v. of 2.8%. T_{lc} probably estimated from fig. 11 and we would estimate it lower (probably about 28°C from fig. 11) | Unknown | Wild | Questionable |
| <i>Cercartetus nanus</i> | 34.9 | 31 | Unknown | The citation given is incorrect, and we were unable to track the values back to a paper on this species | Unknown | Unknown | Poor |
| <i>Cercopithecus mitis</i> | 37 | 5 | Müller, Kamau, and Maloiy (1983) | T_b given as 36–38.5°C in the original paper, but otherwise unclear where the presented value comes from; one individual maintained T_b of around 37°C during the active period; 5°C is given as the approximate T_{lc} . 5°C was the lowest T_a measured, so it is impossible to determine T_{lc} | 2 | Zoo | Poor |

these five species were collected on zoo animals and may therefore not represent wild animals. We classified nine as marginal or questionable for inclusion in the dataset (i.e., T_b varied by 3°C or T_{lc} was difficult to determine). Ultimately, we classified six as inappropriate for inclusion (i.e., there were clear violations of the Scholander-Irving model or the citation was incorrect and impossible to track). Importantly, authors of several of the original papers commented that either T_b or T_{lc} was difficult or impossible to establish for several of these species (e.g., *Arctictis binturong*, McNab, 1995). Such datasets are too often compiled by ignoring biologically important variation and shoehorning incompatible data into a highly conceptualized model (King, 1964).

Finally, even if one accepts the assumptions of constant T_b , thermal conductance, and T_{lc} , there are analytical concerns with using these values to make predictions of organismal responses to conditions far beyond the measured values. For example, one could use these values to estimate energetic expenditure at temperatures below the range empirically measured or to estimate range boundaries (Buckley et al., 2018; Root, 1988). However, such extrapolations are highly sensitive to data quality because they often extend far beyond the range of empirically measured data. Such extrapolations mean that small errors in estimating T_b or T_{lc} (which we reiterate are very hard to estimate and rarely a single value) can lead to large errors in the estimated metabolic rates at cold temperatures. Again, we return to the dataset of Buckley et al. (2018) to demonstrate the scope of the possible error in extrapolating far beyond the known data. In some species, the metabolic rate at the range boundary (MR_{RB}) may vary by an order of magnitude depending on the chosen T_b value (Figure 1). At the extreme, a 1°C change in the T_b value chosen for these extrapolations can lead to differences of over 400% in calculated metabolic expansibility values for some species (although the median difference for all mammals included in the Buckley et al. dataset is 10%). Unfortunately, it is difficult to predict how these errors will be manifested in the interpretations of data. On one hand, we might predict that errors will be largest in high latitude species because the minimum environmental temperature is far below T_{lc} (i.e., the extrapolation is more extreme). On the other hand, the calculated thermal conductance value varies strongly when T_{lc} and T_b are close together, which is most likely to occur in tropical and subtropical species. The results of these analyses often seem to corroborate previous analyses using similar methodologies, but as King (1964) and others have recognized, oversimplifying assumptions of the Scholander-Irving model might result in overly simplified and generic results.

Although we strongly encourage the continued integration of physiological ecology and macroecology, we contend that oversimplification of physiological principles can lead to unreliable analyses. In addition, data quality is of utmost important in these analyses, especially in cases where analyses are highly sensitive to variance in input variables. Understanding the thermophysiology of endotherms relies on recognizing a number of caveats which have unfortunately not been widely adopted by nonspecialists (Mitchell et al.,

2018). While we recognize the importance of broader macroscale analyses, such studies would benefit from closer collaborations between macroecologists and physiological ecologists as each could help the other better understand the hidden nuances in their respective analyses and move toward a more comprehensive understanding of global patterns.

ACKNOWLEDGMENTS

We thank many unnamed colleagues for discussing the ideas in this commentary and focusing our argument. After JGB and DLL, author order was determined by world football rankings.

CONFLICT OF INTEREST

None declared.

AUTHOR CONTRIBUTIONS

JGB and DLL conducted the literature review and analyses. All authors contributed to writing the manuscript.

DATA AVAILABILITY STATEMENT

No original data were collected for this paper. Data used in the analysis can be found associated with Buckley et al. (2018).

Justin G. Boyles¹ 

Danielle L. Levesque² 

Julia Nowack³ 

Michał S. Wojciechowski⁴ 

Clare Stawski⁵ 

Andrea Fuller⁶ 

Ben Smit⁷ 

Glenn J. Tattersall⁸ 

¹Cooperative Wildlife Research Laboratory, Center for Ecology, School of Biological Sciences, Southern Illinois University, Carbondale, IL, USA

²School of Biology and Ecology, University of Maine, Orono, ME, USA

³School of Biological and Environmental Sciences, Liverpool John Moores University, Liverpool, UK

⁴Department of Vertebrate Zoology and Ecology, Faculty of Biology and Environmental Protection, Nicolaus Copernicus University, Toruń, Poland

⁵Department of Biology, Norwegian University of Science and Technology, Trondheim, Norway

⁶Brain Function Research Group, School of Physiology, University of the Witwatersrand, Johannesburg, South Africa

⁷Department of Zoology and Entomology, Rhodes University, Grahamstown, South Africa

⁸Department of Biological Sciences, Brock University, St. Catharines, ON, Canada

Correspondence

Justin G. Boyles, Cooperative Wildlife Research Laboratory, Center for Ecology, and School of Biological Sciences, Southern Illinois University, Carbondale, IL, USA.
Email: jgboyles@siu.edu

ORCID

Justin G. Boyles  <https://orcid.org/0000-0003-1494-4515>

Danielle L. Levesque  <https://orcid.org/0000-0003-0132-8094>

Julia Nowack  <https://orcid.org/0000-0002-4512-5160>

Michał S. Wojciechowski  <https://orcid.org/0000-0001-7765-0720>

Clare Stawski  <https://orcid.org/0000-0003-1714-0301>

Andrea Fuller  <https://orcid.org/0000-0001-6370-8151>

Ben Smit  <https://orcid.org/0000-0003-4160-8242>

Glenn J. Tattersall  <https://orcid.org/0000-0002-6591-6760>

REFERENCES

- Boyles, J. G. (2019). A brief introduction to methods for describing body temperature in endotherms. *Physiological and Biochemical Zoology*, 92, 365–372. <https://doi.org/10.1086/703420>
- Boyles, J. G., Thompson, A. B., McKechnie, A. E., Malan, E., Humphries, M. M., & Careau, V. (2013). A global heterothermic continuum in mammals. *Global Ecology and Biogeography*, 22, 1029–1039. <https://doi.org/10.1111/geb.12077>
- Bozinovic, F., & Rosenmann, M. (1988). Comparative energetics of South American cricetid rodents. *Comparative Biochemistry and Physiology Part A*, 91A, 195–202. [https://doi.org/10.1016/0300-9629\(88\)91616-7](https://doi.org/10.1016/0300-9629(88)91616-7)
- Bradley, W. G., Yousef, M. K., & Scott, I. M. (1975). Physiological studies on the rock pocket mouse, *Perognathus intermedium*. *Comparative Biochemistry and Physiology Part A*, 50A, 331–337. [https://doi.org/10.1016/0300-9629\(75\)90022-5](https://doi.org/10.1016/0300-9629(75)90022-5)
- Buckley, L. B., Khaliq, I., Swanson, D. L., & Hof, C. (2018). Does metabolism constrain bird and mammal ranges and predict shifts in response to climate change? *Ecology and Evolution*, 8, 12375–12385. <https://doi.org/10.1002/ece3.4537>
- Calder, W. A., & Schmidt-Nielsen, K. (1967). Temperature regulation and evaporation in the pigeon and the roadrunner. *American Journal of Physiology*, 213, 883–889. <https://doi.org/10.1152/ajplegacy.1967.213.4.883>
- Dalby, P. L., & Heath, A. G. (1976). Oxygen consumption and body temperature of the Argentine field mouse, *Akodon azarae*, in relation to ambient temperature. *Journal of Thermal Biology*, 1, 177–179. [https://doi.org/10.1016/0306-4565\(76\)90010-3](https://doi.org/10.1016/0306-4565(76)90010-3)
- Dawson, W. R. (1955). The relation of oxygen consumption to temperature in desert rodents. *Journal of Mammalogy*, 36(4), 543–553. <https://doi.org/10.2307/1375808>
- Fristoe, T. S., Burger, J. R., Balk, M. A., Khaliq, I., Hof, C., & Brown, J. H. (2015). Metabolic heat production and thermal conductance are mass-independent adaptations to thermal environment in birds and mammals. *Proceedings of the National Academy of Sciences of the United States of America*, 112(52), 15934–15939. <https://doi.org/10.1073/pnas.1521662112>
- Golightly, R. T. Jr., & Ohmart, R. D. (1983). Metabolism and body temperature of two desert canids: Coyotes and kit foxes. *Journal of Mammalogy*, 64(4), 624–635. <https://doi.org/10.2307/1380518>

- Haim, A., Rubal, A., & Harari, J. (1993). Comparative thermoregulatory adaptations of field mice of the genus *Apodemus* to habitat challenges. *Journal of Comparative Physiology B*, 163(7), 602–607. <https://doi.org/10.1007/BF00302120>
- Hetem, R. S., Maloney, S. K., Fuller, A., & Mitchell, D. (2016). Heterothermy in large mammals: Inevitable or implemented? *Biological Reviews*, 91, 187–205. <https://doi.org/10.1111/brv.12166>
- Hudson, J. W. (1965). Temperature regulation and torpidity in the pygmy mouse, *Baiomys taylori*. *Physiological Zoology*, 38(3), 243–254. <https://doi.org/10.1086/physzool.38.3.30152836>
- Khaliq, I., Böhning-Gaese, K., Prinzinger, R., Pfenninger, M., & Hof, C. (2017). The influence of thermal tolerances on geographical ranges of endotherms. *Global Ecology and Biogeography*, 26(6), 650–668. <https://doi.org/10.1111/geb.12575>
- Khaliq, I., Hof, C., Prinzinger, R., Böhning-Gaese, K., & Pfenninger, M. (2014). Global variation in thermal tolerances and vulnerability of endotherms to climate change. *Proceedings of the Royal Society B: Biological Sciences*, 281(1789), 20141097. <https://doi.org/10.1098/rspb.2014.1097>
- King, J. R. (1964). Oxygen consumption and body temperature in relation to ambient temperature in the White-Crowned Sparrow. *Comparative Biochemistry and Physiology*, 12, 13–24. [https://doi.org/10.1016/0010-406X\(64\)90044-1](https://doi.org/10.1016/0010-406X(64)90044-1)
- Kobbe, S., Nowack, J., & Dausmann, K. H. (2014). Torpor is not the only option: Seasonal variations of the thermoneutral zone in a small primate. *Journal of Comparative Physiology B*, 184, 789–797. <https://doi.org/10.1007/s00360-014-0834-z>
- Levesque, D. L., Nowack, J., & Stawski, C. (2016). Modelling mammalian energetics: The heterothermy problem. *Climate Change Responses*, 3, 7. <https://doi.org/10.1186/s40665-016-0022-3>
- McKechnie, A. E., Coe, B. H., Gerson, A. R., & Wolf, B. O. (2017). Data quality problems undermine analyses of endotherm upper critical temperatures. *Journal of Biogeography*, 44, 2424–2426. <https://doi.org/10.1111/jbi.12941>
- McNab, B. K. (1969). The economics of temperature regulation in neotropical bats. *Comparative Biochemistry and Physiology*, 31, 227–268.
- McNab, B. K. (1978). The comparative energetics of neotropical marsupials. *Journal of Comparative Physiology B*, 125(2), 115–128. <https://doi.org/10.1007/BF00686747>
- McNab, B. K. (1979). The influence of body size on the energetics and distribution of fossorial and burrowing mammals. *Ecology*, 60, 1010–1021. <https://doi.org/10.2307/1936869>
- McNab, B. K. (1980). Energetics and the limits to a temperate distribution in armadillos. *Journal of Mammalogy*, 61(4), 606–627. <https://doi.org/10.2307/1380307>
- McNab, B. K. (1988). Energy conservation in a tree-kangaroo (*Dendrolagus matschiei*) and the red panda (*Ailurus fulgens*). *Physiological Zoology*, 61, 280–292. <https://doi.org/10.1086/physzool.61.3.30161241>
- McNab, B. K. (1995). Expenditure and conservation in frugivorous and mixed-diet carnivorans. *Journal of Mammalogy*, 76, 206–222.
- Mitchell, D., Snelling, E. P., Hetem, R. S., Maloney, S. K., Strauss, W. M., & Fuller, A. (2018). Revisiting concepts of thermal physiology: Predicting responses of mammals to climate change. *Journal of Animal Ecology*, 87, 956–973. <https://doi.org/10.1111/1365-2656.12818>
- Müller, E., Kamau, J., & Maloiy, G. (1983). A comparative study of basal metabolism and thermoregulation in a folivorous (*Colobus guereza*) and an omnivorous (*Cercopithecus mitis*) primate species. *Comparative Biochemistry and Physiology Part A*, 74(2), 319–322. [https://doi.org/10.1016/0300-9629\(83\)90608-4](https://doi.org/10.1016/0300-9629(83)90608-4)
- Neal, C. M., & Lustick, S. I. (1973). Energetics and evaporative water loss in the short-tailed shrew *Blarina brevicauda*. *Physiological Zoology*, 46(3), 180–185. <https://doi.org/10.1086/physzool.46.3.30155600>
- Noakes, M. J., Smit, B., Wolf, B. O., & McKechnie, A. E. (2013). Thermoregulation in African Green Pigeons (*Treron calvus*) and a re-analysis of insular effects on basal metabolic rate and heterothermy in columbid birds. *Journal of Comparative Physiology B*, 183, 969–982. <https://doi.org/10.1007/s00360-013-0763-2>
- Porter, W. P., & Kearney, M. (2009). Size, shape, and the thermal niche of endotherms. *Proceedings of the National Academy of Sciences of the United States of America*, 106(Supplement 2), 19666–19672. <https://doi.org/10.1073/pnas.0907321106>
- Root, T. (1988). Energy constraints on avian distributions and abundances. *Ecology*, 69, 330–339. <https://doi.org/10.2307/1940431>
- Scholander, P. F., Hock, R., Walters, V., Johnson, F., & Irving, L. (1950). Heat regulation in some arctic and tropical mammals and birds. *Biological Bulletin*, 99, 237–258. <https://doi.org/10.2307/1538741>
- Shield, J. (2009). Acclimation and energy metabolism of the dingo, *Canis dingo* and the coyote, *Canis latrans*. *Journal of Zoology*, 168(4), 483–501.
- Somero, G. N. (2013). A 1950s classic of thermal adaptation to cold. *Journal of Experimental Biology*, 216, 1759–1761. <https://doi.org/10.1242/jeb.076398>
- Tracy, C. R. (1972). Newton's Law: Its application for expressing heat losses from homeotherms. *BioScience*, 22, 656–659. <https://doi.org/10.2307/1296267>
- Wright, K. P. Jr, Badia, P., Myers, B. L., Plenzler, S. C., & Hakel, M. (1997). Caffeine and light effects on nighttime melatonin and temperature levels in sleep-deprived humans. *Brain Research*, 747, 78–84. [https://doi.org/10.1016/S0006-8993\(96\)01268-1](https://doi.org/10.1016/S0006-8993(96)01268-1)