



Research article

Microbial diversity in relation to physico-chemical properties of hot water ponds located in the Yamunotri landscape of Garhwal Himalaya



Rahul Kumar*, Ramesh C. Sharma

Laboratory of Environmental Microbiology and Biotechnology, Department of Environmental Sciences, H.N.B. Garhwal University (A Central University), Srinagar Garhwal, 246174, Uttarakhand, India

ARTICLE INFO

Keywords:
 Environmental health
 Nature conservation
 Water pollution
 Bacteria
 Microorganism
 Thermophilic bacteria
 Hot water ponds
 Physico-chemical parameters
 MALDI-TOF-MS
 Yamunotri shrine

ABSTRACT

The current study includes the assessment of physico-chemical characteristics along with the microbial diversity of hot water samples collected from three important sacred hot water springs of the Uttarakhand Himalaya close to the world-famous Hindu shrine Yamunotri temple. Hot water samples were collected for two consecutive years 2015–2016 in two sampling operations each year. A total of fifteen physico-chemical attributes of hot water were recorded. Microbes of hot water ponds were characterized by morphological, biochemical, MALDI-TOF MS, and molecular approaches. A total of twenty-two microbial strains were identified from the Surya Kund with water temperature ranging between 86 °C and 89 °C; twenty-two microbial strains were identified from the Draupadi Kund with water temperature ranging between 65 °C and 69 °C and twenty-one microbial strains were identified from the Yamunotri Tapt Kund with water temperature ranging between 45 °C and 48 °C during the study period. The present study on the assessment of physico-chemical characteristics and thermophilic microbial diversity of all the three hot water springs can be a useful reference for further studies on similar aspects in other parts of the Himalaya. The available data will also help to understand the reasons behind the curative properties of these hot water springs. This baseline information will also be instrumental for the conservation and management of these hot water springs.

1. Introduction

Microbes are one of the most important components of any extreme environment. These microbes can take nutrition from these harsh environmental conditions. Thus, these extremophiles can survive and grow in harsh environmental conditions (Oarga, 2009). Among extremophiles, the thermophiles or heat-loving microbes can survive at extreme temperatures (Beg et al., 2000; Akmar et al., 2011). Hot water ponds have water temperatures higher than the atmospheric temperature in surroundings (Sen et al., 2010). Hot water ponds represent harsh environmental conditions available across the entire Himalayan region (Kumar et al., 2004). Hot water ponds are an important source of heat-loving or thermophilic microorganisms useful and effective even at the extremely high temperature ranges. These microorganisms are beneficial for various industries including the pharmaceutical sector (Tekere et al., 2015).

A considerable research has been undertaken on multiple aspects of hot water ponds. This includes the work of Baker et al. (2001) on the isolation and identification of bacterial diversity in the hot springs of

Indonesia; Kumar et al. (2004) on identification of microbiological frequency of soil of two distinct hot water ponds in the Uttarakhand Himalaya; Kvist et al. (2007) on archaeal diversity in Icelandic hot water spring; Sharma et al. (2008) on isolation and characterization of *Geobacillus* spp; Akmar et al. (2011) on identification of a novel heat loving bacterial strain from hot water pond; Bhusare and Wakte (2011) on microbial diversity of Unkeshwar thermal spring; Huang et al. (2011) on diversity of archaea and bacteria in the hot springs of Tibetan Plateau; Sharma et al. (2012) on the identification of Manikaran hot spring microbial community; Pagaling et al. (2012) on diversity of bacteria and archaea in microbial mats obtained from two important thermal springs located in the geothermal region of Tengchong; Jiang et al. (2012) on diversity of actinomycetes in the thermal springs located in Central and Central-Eastern Tibet, an autonomous region of China; Wemheuer et al. (2013) on microorganisms and their biochemical potential; while, Lopez et al. (2013) on hot spring metagenomics; Ghati et al. (2013) on Esterolytic heat loving bacterial strain from a thermal water pond in India; Bandyopadhyay et al. (2013) on a bacterium having a tolerant capacity against the high arsenic concentration that was isolated from a thermal

* Corresponding author.

E-mail address: rahul.khadwalia@gmail.com (R. Kumar).

spring in India; Meyer-Dombard and Amend (2014) on geochemistry and microbial ecology in a hot springs having alkaline pH located at Ambitle Island in Papua New Guinea; Pathak and Rathod (2014) on cultivable bacterial frequency of hot water pond located at Unkeshwar in Maharashtra; Rawat (2015) on bacterial enumeration from a thermal spring of Uttarakhand. Chan et al. (2015) on the enumeration of heat-loving microbial diversity in Malaysia through sequencing and metagenomic approaches; Tekere et al. (2015) on the diversity of bacteria dwelling from some African hot water ponds; Kumar and Sharma (2019a,b) on microbes and physico-chemical characteristics of two Garhwal Himalayan thermal springs; and Kumar and Sharma (2020) on the determination of microbes and physico-chemical characteristics of thermal springs located close to the Badrinath temple. However, no information is available in the public domain regarding the diversity of microbes and physico-chemical characteristics of the hot waters of Surya Kund, Draupadi Kund and Yamunotri Tapt Kund located in the vicinity of world-famous Hindu shrine Yamunotri in Uttarakhand Himalaya. Therefore, it was felt desirable to undertake the work on the diversity of microbes and physico-chemical features of all the three hot water bodies Surya Kund, Draupadi Kund and Yamunotri Tapt Kund hot water. Thus, the present obtained baseline data will have both academic and applied values.

2. Materials and methods

2.1. Study area

All the hot water ponds, Surya Kund, Draupadi Kund and Yamunotri Tapt Kund are located just near to the World famous Hindu's shrine Yamunotri Temple in the Uttarakhand Himalaya (Figure 1). Surya Kund or Suraj Kund is located at a height of 3,291 m above m.s.l. at latitude $31^{\circ}1'0.12''$ N and longitude $78^{\circ}27'0''$ E. Draupadi Kund is situated next to Surya Kund at a height of 3,270 m above m.s.l. This hot water of Draupadi Kund is known for its curative and medicinal properties. The maximum *in-situ* water temperature of Draupadi Kund is 67 °C. However, Yamunotri Tapt Kund is also situated in the vicinity of Surya Kund at a height of 3,262 m above m.s.l. The maximum *in-situ* water temperature of Yamunotri Tapt Kund is 45 °C.

2.2. Water sampling

Water samples were collected from all the three hot water bodies (Surya Kund, Draupadi Kund and Yamunotri Tapt Kund) in two sampling operations (June and August) for the period of two years (2015; 2016). The hot water ponds were only accessible during the period from June to

October every year due to their locations in the high altitude Himalayan range. Water temperatures, pH, free CO₂, the concentration of dissolved oxygen were recorded at the collection sites. For rest of the parameters, water samples were collected in autoclaved thermosteel flask and transported to the Laboratory of Environmental Microbiology and Biotechnology (L.E.M.B.), Department of Environmental Sciences, H.N.B. Garhwal University, Srinagar Garhwal, Uttarakhand. The physico-chemical parameters and microbial analyses were made within 24 h of the sample collection following the standard methodology outlined in APHA (2012); Harley and Prescott (2002) and Morello et al. (2003). The samples of water collected from the sampling sites were inoculated in the desired media and incubated under the similar environmental conditions available at the sampling sites like temperature and pH. The inoculated plates were incubated at the same temperature observed at the sampling site for 48–72 h in case of bacteria and 3–5 days in the case of fungi and actinomycetes. All the unique microbial colonies were cultured in similar growth media to obtain a pure culture for further growth and identification (Davis et al., 2005; Huq et al., 2012; Anand and Aoyagi, 2019).

2.3. Physical and chemical characteristics of hot water

Samples of water from all the three hot water ponds were collected for the analysis of a predefined set of fifteen physico-chemical parameters. Water temperature was observed by submerging the digital Centigrade Thermometer having a temperature range of -50 to +300 °C up to 10 cm in water. The pH was recorded on-site by using portable pH meter (Electronics India Model No. 7011) and off-site by using a benchtop pH meter (Toshcon, Model No. TPC-17). The concentration of DO was estimated on-site following the Modified Winkler's Method. Free CO₂ was also recorded at the sites. Electronic conductivity, the concentration of salinity and TDS were recorded with the help of Multiparameter Analyzer (Toshcon, Model No. TPC-17). However, the concentration of total alkalinity, chlorides, total hardness, Calcium and Magnesium were recorded at the Laboratory using the standard methodology available in APHA (2012). The concentrations of nitrates, sulfates and phosphates were analyzed with the help of a UV-VIS Spectrophotometer (Systronic, Model No. 117) at the Laboratory.

2.4. Microbial isolation and enumeration

Bacterial colonies were isolated by using the Nutrient Agar Media (HiMedia, India, Bangalore) commonly known as NA Media. The fungal strains were isolated by using the Sabaroud Dextrose Agar (SDA) Media (HiMedia, India, Bangalore). To avoid any type of bacterial

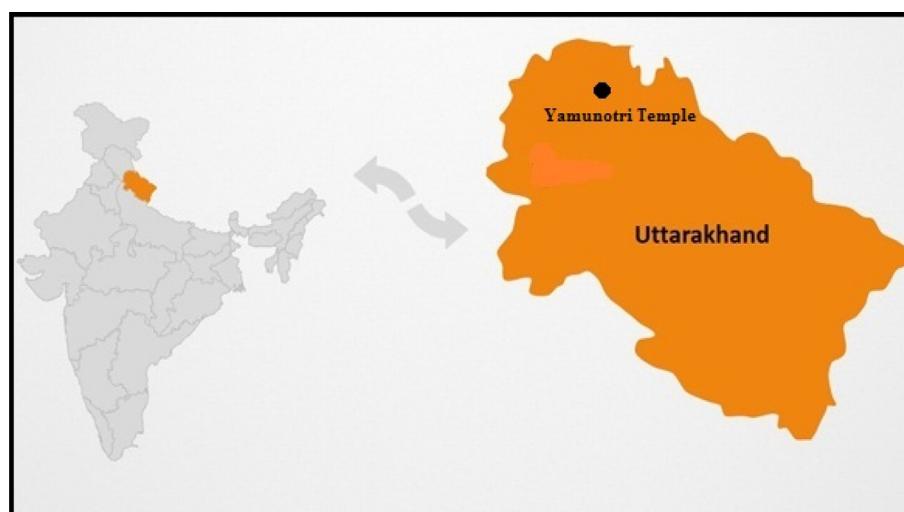


Figure 1. The Study area (Surya Kund; Draupadi Kund and Yamunotri Tapt Kund located near Yamunotri temple).

contamination during the isolation of fungal strains, the SDA media was enriched with 50 mg l⁻¹ of each Ampicillin (HiMedia, India, Bangalore) and Streptomycin (HiMedia, India, Bangalore). For isolation of Actinomycetes, the Actinomycetes Isolation Agar (AIA) media (HiMedia, India, Bangalore) was used. The pH of each culture medium was adjusted matching to the pH recorded at the sampling site. After the enumeration of microorganisms, the isolated colonies were streaked on separate suitable media plates for getting the pure cultures of each microbe (Clesceri et al., 1998).

2.5. Morphological and biochemical characteristics of microbes

A total of twenty-seven morphological and biochemical characteristics were analyzed during the study period. During analysis, pure cultures of microorganisms were carefully recognized by unaided eyes and under the Phase Contrast Microscope (Nikon Eclipse TS100) for studying the morphological structures and arrangements of the bacterial strains. The morphological characteristics of microorganisms included shape, size, margin, elevation, color, cell shape, motility, spore formation, flagellation, and gram staining. However, various biochemical tests were performed to identify a microbial strain up to the lowest possible taxon (Rohomania et al., 2015; Aytiso and Onyango, 2016). Biochemical tests included the assessment of catalase, citrate, urease, fructose, MR-VP, indole test, and maltose, etc.

2.6. Identification of microbial isolates by using MALDI-TOF MS

Pure culture of each isolate was sent to the National Centre for Microbial Resources (NCMR), Pune for final confirmation using the MALDI-TOF mass spectrometry technique. Mass spectrometry is an analytical technique in which samples are ionized into charged molecules and the ratios of their mass-to-charge (*m/z*) can be measured. In MALDI-TOF mass spectrometry, the ion source is matrix-assisted laser desorption/ionization (MALDI), and the mass analyzer is time-of-flight (TOF) analyzer. MALDI-TOF MS is an emerging tool used for high-throughput and rapid microbial identification. The principle behind microbial identification using MALDI-TOF MS is based on the fact that every microorganism has a specific protein composition, which gives its characteristic and unique mass spectra (Risch et al., 2010; Marko et al., 2012). The procedure of identification of microorganisms by using MALDI-TOF MS had two sections: the first section involves the preparation of the HCCA (α -Cyano-4-hydroxycinnamic acid) matrix solution; and the second section involves the direct transfer method, in which the single microbial colony is transferred to the MALDI target plate and later using the HCCA matrix solution and MALDI-TOF MS instrument for the identification of microbial colonies based on the mass spectra.

2.7. DNA extraction and gene amplification

Almost all the microbial isolates were identified by using the technique of MALDI-TOF MS. However, few microbial isolates were identified through molecular or sequencing approaches, as their identification was not successfully made through the MALDI-TOF-MS technique. The DNA was extracted from these selected microbial isolates using HiPurA Bacterial Genomic DNA Extraction Kit (HTBM008; HiMedia, India, Bangalore) for bacteria and GSure Fungal DNA Extraction Kit (G45331; GCC Biotech, India, New Delhi) for fungal isolates. Genes of 16S rRNA were amplified by PCR using specific universal primers, forward primer 8F (AGAGTTGATCCTGGCTAG) and reverse primer 1492R (TACGGYTACCTTGTACGACTT) (GCC Biotech, India, New Delhi) (Takahashi et al., 2014); whereas, the genes of 18S rRNA were amplified by PCR using specific universal ITS primers, forward primer ITS 1F (TCCGTAGGTGAACCTGCGG) and ITS 4R (TCCTCCGTTATTGATATGC) (GCC Biotech, India, New Delhi) (Pryce et al., 2003; Raja et al., 2017; Usyk et al., 2017).

2.8. Identification of microbial isolates by using rRNA sequencing approach

Few of the microbial isolates, whose identifications could not be possible through MALDI-TOF MS, were sent for 16S rRNA and 18S rRNA sequencing at the same facilitation center. The facilitation center provided an accession number only to those microbial strains whose identification was done through the gene sequencing method. The accession numbers of such microbial strains are given in Table 1.

2.9. Statistical treatment of data

MS Office 2013 was used for the statistical treatment (minimum; maximum; mean; standard deviation) of the data related with the physico-chemical characteristics of hot water samples of all the hot water ponds.

3. Results and discussion

Results of all the physico-chemical parameters of hot waters of Surya Kund, Draupadi Kund and Yamunotri Tapt Kund have been presented in Table 2. Detailed biochemical and morphological characteristics of few isolates selected randomly from all the examined strains are given in Table 3. The majority of the isolated and identified bacteria were found to be gram-positive and such bacteria are known to be comparatively stress-resistant and long-range migrants, especially the Actinobacteria (Cerritos et al., 2011; Kumar et al., 2014).

3.1. Physico-chemical characteristics and microbial diversity of Surya Kund

The water temperature of Surya Kund ranged from 86 °C to 89 °C with a mean value of 87.8 °C. High temperature is important for the growth of thermophiles. Those microorganisms which can grow at a temperature of more than 80 °C are known as hyperthermophiles. The pH of the hot water was recorded within a range from 8.4 to 8.8, indicating the slightly alkaline nature of water of Surya Kund. An acidic nature of one hot water spring of Kullu district, Himachal Pradesh was recorded by Kumar et al. (2013). The same observation (pH 6.9–9.5) of acidic nature of hot water was reported by Singh et al. (2015) on the thermal springs of Jharkhand and West Bengal. A very low concentration of DO (1.8 mg l⁻¹) was recorded in the hot water of Surya Kund during all sampling operations. Kumar et al. (2013) also observed low concentration of DO (2.52 mg l⁻¹) in hot water springs of Himachal Pradesh. The water temperature is inversely proportional to the concentration of DO. As the water temperature increases, the amount of DO decreases (Rana et al., 2018). Fazlzadeh et al. (2016) observed 3.25 mg l⁻¹ to 3.57 mg l⁻¹ concentration of dissolved oxygen in Iran. Electrical conductivity (EC) ranged between 12.0 mS/cm to 12.4 mS/cm with 12.2 mS/cm as mean value in the hot water of Surya Kund. Haki and Gezmu (2012) reported almost an identical range of electrical conductivity in Ethiopian hot spring. The salinity of water of Surya Kund varied from 6.6^{SAL} to 6.8^{SAL} during the study period. High salinity of thermal springs was observed by Hamzah et al. (2013) from Malaysia. TDS in the hot water of Surya Kund was recorded between 6.5 mg l⁻¹ to 6.8 mg l⁻¹. However, a high concentration of TDS was observed by Hamzah et al. (2013) in Malaysia. Free CO₂ was recorded between 22.0 mg l⁻¹ to 26.4 mg l⁻¹ during the study period. Total hardness of hot water of Surya Kund ranged between 164 mg l⁻¹ to 168 mg l⁻¹. Concentration of calcium ranged from 21.60 mg l⁻¹ to 24.05 mg l⁻¹ in the hot water of Surya Kund. Magnesium concentration was recorded between 26.34 mg l⁻¹ to 27.34 mg l⁻¹ Singh et al. (2015) recorded the concentration of Magnesium in the range of 12.60 mg l⁻¹ to 15.62 mg l⁻¹ in the thermal springs of Jharkhand and West Bengal. Chlorides concentration was recorded between 157.6 mg l⁻¹ to 166.1 mg l⁻¹ with an average of 161.9 mg l⁻¹. Total alkalinity of hot water ranged from 130 mg l⁻¹ to 150 mg l⁻¹ in the water of Surya Kund

Table 3. Morphological and Biochemical characterization of bacterial isolates from hot water ponds located in the Yamunotri landscape.

| Characteristics | Bacterial Isolates | | | | | | | |
|----------------------|-------------------------------|----------------------------|-----------------------------------|--------------------------|-------------------------------|----------------------------|-------------------------------|---------------------------------------|
| | <i>Bacillus licheniformis</i> | <i>Bacillus megaterium</i> | <i>Streptococcus thermophilus</i> | <i>Bacillus subtilis</i> | <i>Actinobacillus hominis</i> | <i>Paenibacillus alvei</i> | <i>Streptococcus pyogenes</i> | <i>Geobacillus stearothermophilus</i> |
| Shape | Round | Round | Round | Round | Round | Circular | Spherical | Circular |
| Size | 4 mm | 3 mm | 2 mm | 2–3 mm | 1 mm | 1 mm | 1 mm | 2–3 mm |
| Margin | Undulate | Undulate | Entire | Lobate | Regular | Regular | Smooth | Entire |
| Elevation | Semi-raised | Elevated | Convex | Flat | Flat | Semi-raised | Convex | Convex |
| Color | White | Pale-white | Cream | White | White | Pale-white | Translucent | Cream |
| Cell shape | Straight rods | Straight rods | Cocci in chain | Rod | Pleomorphic | Rod | Cocci | Rod |
| Spore formation | + | + | - | - | - | + | - | + |
| Motility | Motile | Motile | Non-motile | Motile | Non-motile | Motile | Non-motile | Motile |
| Grams staining | + | + | + | + | - | + | + | + |
| Flagella | Peritrichous | Peritrichous | Atrichous | Peritrichous | Atrichous | Peritrichous | Atrichous | Monotrichous |
| Catalase | + | + | + | + | + | + | - | - |
| Citrate | - | + | + | + | - | - | + | v |
| Urease | + | + | + | - | + | + | - | - |
| Methyl Red (MR) | - | - | - | - | - | - | - | - |
| Voges Proskauer (VP) | - | - | - | + | - | - | - | - |
| Fructose | - | + | + | + | - | + | + | - |
| Indole Test | - | - | - | - | - | - | - | - |
| Raffinose | - | + | - | + | + | - | - | - |
| Ribose | + | + | - | + | + | - | - | + |
| Sorbitol | + | - | - | + | - | + | - | - |
| Sucrose | + | - | + | + | + | - | + | - |
| Xylose | - | - | - | + | + | + | - | + |
| Trehalose | + | - | - | + | - | - | + | - |
| Mannose | + | - | + | + | - | - | + | - |
| Mannitol | + | + | - | + | + | - | - | + |
| Lactose | + | + | + | v | - | - | + | - |
| Maltose | + | - | - | + | - | - | + | - |

| Characteristics | Bacterial Isolates | | | | | | | |
|----------------------|--------------------------|---------------------------------|-----------------------------|------------------------------|----------------------------------|-------------------------------|----------------------------------|-------------------------|
| | <i>Bacillus mycoides</i> | <i>Bacillus thermoamylorans</i> | <i>Streptomyces rangoon</i> | <i>Streptomyces clavifer</i> | <i>Lysinibacillus sphaericus</i> | <i>Actinobacillus seminis</i> | <i>Brevibacillus thermoruber</i> | <i>Bacillus simplex</i> |
| Shape | Round | Round | Round | Round | Round | Round | Circular | Round |
| Size | 3 mm | 2–3 mm | 1–3 mm | 1–3 mm | 3 mm | 2–4 mm | 2 mm | 3 mm |
| Margin | Rhizoid | Smooth | Entire | Entire | Regular | Entire | Undulate | Regular |
| Elevation | Raised | Flat | Centrally raised | Centrally raised | Raised | Convex | Convex | Semi-raised |
| Color | Cream | White | Cream | Sandy yellow | Pale-white | White | Cream | Cream |
| Cell shape | Rod | Rod | Rod | Rod | Rod | Coco-bacillary | Rod | Rod |
| Spore formation | + | - | + | + | + | - | + | + |
| Motility | Non-motile | Motile | Motile | Motile | Motile | Non-motile | Non-motile | Motile |
| Grams staining | + | + | + | + | + | - | + | + |
| Flagella | Peritrichous | Peritrichous | Peritrichous | Peritrichous | Peritrichous | Atrichous | Peritrichous | Peritrichous |
| Catalase | + | + | - | - | + | + | + | + |
| Citrate | v | - | + | + | + | - | - | - |
| Urease | v | - | - | - | - | - | - | + |
| Methyl Red | + | - | - | - | - | - | - | - |
| Voges Proskauer (VP) | + | + | + | + | - | - | + | - |
| Fructose | v | + | + | + | - | - | + | - |
| Indole Test | - | - | - | - | - | - | - | - |
| Raffinose | - | - | - | - | - | - | + | - |
| Ribose | v | - | - | - | - | - | - | + |
| Sorbitol | - | - | + | + | - | - | - | - |
| Sucrose | v | + | + | + | - | - | + | + |
| Xylose | - | + | - | - | - | - | - | + |
| Trehalose | + | + | - | - | - | - | + | - |

(continued on next page)

Table 3 (continued)

| Characteristics | Bacterial Isolates | | | | | | | |
|-----------------|--------------------------|---------------------------------|-----------------------------|------------------------------|----------------------------------|-------------------------------|----------------------------------|-------------------------|
| | <i>Bacillus mycoides</i> | <i>Bacillus thermoamylorans</i> | <i>Streptomyces rangoon</i> | <i>Streptomyces clavifer</i> | <i>Lysinibacillus sphaericus</i> | <i>Actinobacillus seminis</i> | <i>Brevibacillus thermoruber</i> | <i>Bacillus simplex</i> |
| Mannose | - | + | - | - | - | - | - | - |
| Mannitol | - | - | + | - | + | - | - | - |
| Lactose | - | + | - | - | - | - | - | - |
| Maltose | - | + | - | - | - | - | - | + |

Abbreviations: +: positive; -: negative; v: variable.

Table 4. Microbial diversity present in hot water ponds located in the Yamunotri landscape of Garhwal Himalaya.

| S. No. | Microorganisms | Surya Kund | Draupadi Kund | Yamunotri Tapt Kund |
|-------------------------|--|------------|---------------|---------------------|
| A. Bacteria | | | | |
| 1 | <i>Bacillus subtilis</i> | + | + | + |
| 2 | <i>Brevibacillus borstelensis</i> | - | + | - |
| 3 | <i>Bacillus licheniformis</i> (AE017333) | + | + | + |
| 4 | <i>Streptococcus thermophilus</i> | - | - | + |
| 5 | <i>Actinobacillus seminis</i> | - | - | + |
| 6 | <i>Bacillus mycoides</i> | + | - | + |
| 7 | <i>Paenibacillus dendritiformis</i> | + | - | - |
| 8 | <i>Paenibacillus ehimensis</i> | + | - | - |
| 9 | <i>Bacillus simplex</i> | + | - | - |
| 10 | <i>Bacillus thermoamylorans</i> | + | + | - |
| 11 | <i>Geobacillus stearothermophilus</i> | + | + | - |
| 12 | <i>Bacillus megaterium</i> | + | + | - |
| 13 | <i>Paenibacillus alvei</i> | + | + | - |
| 14 | <i>Actinobacillus hominis</i> | + | + | + |
| 15 | <i>Bacillus tequilensis</i> (AYTO01000043) | + | + | - |
| 16 | <i>Brevibacillus thermoruber</i> | - | + | - |
| 17 | <i>Brevibacillus choshinensis</i> | - | + | + |
| 18 | <i>Streptococcus pyogenes</i> | - | + | 5 |
| 19 | <i>Thiobacillus denitrificans</i> | - | - | + |
| 20 | <i>Paenibacillus thiominolyticus</i> | - | - | + |
| 21 | <i>Lysinibacillus sphaericus</i> | - | - | + |
| 22 | <i>Bacillus paralicheniformis</i> (LBMN01000156) | - | + | - |
| B. Actinomycetes | | | | |
| 1 | <i>Streptomyces ruber</i> | + | + | + |
| 2 | <i>Nocardiopsis prasina</i> | + | + | + |
| 3 | <i>Streptomyces thermophilaceus</i> | + | - | - |
| 4 | <i>Thermobifida fusca</i> | + | + | + |
| 5 | <i>Streptomyces erumpens</i> | - | + | + |
| 6 | <i>Streptomyces clavifer</i> | - | - | + |
| 7 | <i>Streptomyces rangoon</i> | - | - | + |
| C. Fungi | | | | |
| 1 | <i>Aspergillus nidulans</i> | + | - | + |
| 2 | <i>Thermoascus crustaceus</i> | + | + | - |
| 3 | <i>Thermoascus viride</i> | + | - | - |
| 4 | <i>Cladosporium cladosporioides</i> | + | - | - |
| 5 | <i>Cladosporium herbarum</i> | + | - | - |
| 6 | <i>Penicillium soppii</i> | + | - | - |
| 7 | <i>Canariomyces thermophila</i> | - | + | - |
| 8 | <i>Chaetomium thermophile</i> | - | + | - |
| 9 | <i>Talaromyces thermophilus</i> | - | + | - |
| 10 | <i>Sporotrichum thermophile</i> | - | + | - |
| 11 | <i>Aspergillus niger</i> | - | - | + |
| 12 | <i>Aspergillus flavus</i> | - | - | + |
| 13 | <i>Aspergillus costaricensis</i> (NR_103604.1) | - | - | + |
| 14 | <i>Thermomyces lanuginosus</i> | - | - | + |
| Total | | 22 | 22 | 21 |

Abbreviations: +: present; -: absent.

Draupadi Kund ranged from 0.137 mg l⁻¹ to 0.142 mg l⁻¹. Sulfates concentration of hot water of Draupadi Kund ranged from 1.201 mg l⁻¹ to 1.213 mg l⁻¹. The high concentration of sulfates and nitrates in hot water ponds of Sikkim was reported by [Sherpa et al. \(2013\)](#). Phosphates concentration in hot water of Draupadi Kund was recorded between 0.022 mg l⁻¹ to 0.028 mg l⁻¹.

A total of twenty-two species of microorganisms with thirteen species of bacteria, four species of actinomycetes and five species of fungi were isolated and identified from the hot water of Draupadi Kund ([Table 4](#)). *Geobacillus stearothermophilus* was also reported by [Ghati et al. \(2013\)](#) from Indian hot spring. [Reyensbach et al. \(2000\)](#) also reported the identification of *Bacillus subtilis* in the Yellowstone National Park.

3.3. Physico-chemical characteristics and microbial diversity of Yamunotri Tapt Kund

The water temperature of Yamunotri Tapt Kund ranged from 45 °C to 48 °C with a mean value of 46.8 °C. The pH of Yamunotri Tapt Kund water ranged from 7.3 to 7.8, showing slightly alkaline nature of water. [Singh et al. \(2015\)](#) reported wide variation (6.9–9.5) in pH of hot water springs located in West Bengal and Jharkhand. Considerable concentration of DO (5.8 mg l⁻¹ to 6.2 mg l⁻¹) as recorded in Yamunotri Tapt Kund. [Fazlzadeh et al. \(2016\)](#) reported low concentration (3.25 mg l⁻¹ to 3.57 mg l⁻¹) of dissolved oxygen from hot water springs located in Iran; [Kumar et al. \(2013\)](#) also reported low concentration of DO (2.52 mg l⁻¹) from thermal springs of Himachal Pradesh. Electrical conductivity of water of Yamunotri Tapt Kund ranged from 8.19 mS/cm to 8.28 mS/cm. [Homma and Tsukahara \(2008\)](#) also reported the same range of electrical conductivity from the thermal springs of the Itoigawa Shizuoka Tectonic Line. Salinity of water of Yamunotri Tapt Kund ranged from 4.1^{SAL} to 4.5^{SAL}. Total Dissolved Solids (TDS) ranged from 4.51 mg l⁻¹ to 4.59 mg l⁻¹ in hot water of Yamunotri Tapt Kund. A high concentration of TDS was recorded by [Hamzah et al. \(2013\)](#) in Malaysia. Free CO₂ concentration in Yamunotri Tapt Kund was recorded within a range of 92.4 mg l⁻¹ to 96.8 mg l⁻¹. Total hardness of hot water of Yamunotri Tapt Kund ranged from 128 mg l⁻¹ to 132 mg l⁻¹. Calcium concentration ranged from 27.2 mg l⁻¹ to 31.2 mg l⁻¹ with an average of 29.2 mg l⁻¹. Magnesium concentration fluctuated between 12.71 mg l⁻¹ to 15.64 mg l⁻¹. [Singh et al. \(2015\)](#) reported magnesium concentration from 12.60 mg l⁻¹ to 15.62 mg l⁻¹ for hot water springs of Jharkhand and West Bengal, India. Chlorides concentration was recorded between 92.3 mg l⁻¹ to 96.56 mg l⁻¹ in water samples of Yamunotri Tapt Kund. Total alkalinity ranged from 95 mg l⁻¹ to 120 mg l⁻¹ during the period of study. Nitrates concentration was found within the range of 0.092 mg l⁻¹ to 0.098 mg l⁻¹. Sulfates concentration ranged from 0.241 mg l⁻¹ to 0.244 mg l⁻¹. Phosphates concentration in hot water of Yamunotri Tapt Kund was recorded from 0.081 mg l⁻¹ to 0.090 mg l⁻¹.

An overall, ten species of bacteria, six species of actinomycetes and five species of fungi were isolated and identified from the Yamunotri Tapt Kund. The α -diversity of microbes in hot water of Yamunotri Tapt Kund was recorded to be twenty-one during the study period ([Table 4](#)). *Bacillus licheniformis* was also reported by [Sharma et al. \(2012\)](#) in Manikaran hot water spring of Himachal Pradesh. Three species of bacteria (*Bacillus subtilis*, *Bacillus licheniformis*, and *Actinobacillus hominis*) and three species of actinomycetes (*Streptomyces ruber*, *Nocardiopsis prasina*, and *Thermobifida fusca*) were found common from the hot water of all the three hot water springs.

Temperature is considered, as one of the major environmental factors that control the activities and survival of microbes ([Kumar and Sharma, 2020](#); [Kumar et al. 2013](#); [Takacs-Vesbach et al. 2008](#); [Abou-Shanab, 2007](#)). Most of the thermophiles belong to the domain of Archaea. *Methanopyrus kandleri* is a thermophilic microorganism that can grow and reproduce at a temperature in excess of 250°F ([Vieille and Zeikus, 2001](#)). The other thermophilic microorganisms are *Thermus aquaticus*, *Thermococcus litoralis*, *Bacillus stearothermophilus*, etc. A broad range of thermophilic microorganisms has been isolated from thermal springs, mud pots,

geysers, hot spring algal mat, active volcanoes of the world. These thermophilic habitats are the home of a wide range of thermophilic and thermotolerant microbes. Protein stability, DNA stability, Lipid stability are the three main mechanisms that are responsible for the growth and development of thermophiles at such a high temperature.

4. Importance to the society

4.1. Importance of thermal springs

Natural hot water contains Na⁺, K⁺, Ca²⁺, S, SO₄²⁻ and Cl⁻ which are good for balneotherapy. The chemical composition of the hot water of these natural water bodies contributes to the well being and health of the people. It generally increase metabolism, accelerate healing, soothe muscles, improve blood circulation and detoxify the body's lymphatic system ([Javed et al., 2009](#); [Hamzah et al., 2013](#)). The collected data from the thermal springs shows that water of these springs is suitable for bathing purposes but not for human consumption. The major benefits of bathing in these hot water bodies are: (i) It enhances the circulation of blood; (ii) it reduces stress and promotes sleep; (iii) gives relief from pain or body ache naturally; (iv) it cures skin ailments like rough and dry skin, etc; (v) helps in removal of toxic compounds or elements out of the body; (vii) anti-aging; and (viii) other therapeutic effects such as positive mindset, etc. These possible benefits have been discussed by [Joshi \(2015\)](#).

4.2. Importance of thermophilic microorganisms

Enzymes obtained from heat-loving microorganisms are of paramount importance as they act as biocatalysts for various industrial applications. Such enzymes can work effectively even at high temperatures ([Lopez et al., 2013](#)). The bacterial species, *Bacillus subtilis* has been isolated from Surya Kund, Draupadi Kund and Yamunotri Tapt Kund act as an antidiarrhoeal microorganism, which has an ability to treat diarrhea. This species is also used to produce fermented soybeans, yogurt, ice cream, milk and cheese ([Earl et al., 2008](#); [Shahcheraghi et al., 2015](#); [Ulrich et al., 2018](#)). The bacterial species *Streptococcus thermophilus* has been isolated from Yamunotri Tapt Kund has a property to boost the immunity of a human body and can also be used in the fermentation of cheese. *Brevibacillus borstelensis* can be used for the degradation of polyethylene and long-chain hydrocarbons. This has also been reported by [Hadad et al. \(2005\)](#) and [Khalil et al. \(2018\)](#). *Geobacillus stearothermophilus* showed significant esterase producing ability, hence it can be used for the production of thermostable esterase ([Ghati et al., 2013](#)). *B. licheniformis*, *B. megaterium*, *A. hominis*, *L. sphaericus*, *P. alvei*, *B. simplex*, *A. seminis*, *P. fragii* and *S. cohnii* belong to class Firmicutes and Gamma proteobacteria and showed the production of amylase, gelatinase, lipase ([Pathak and Rathod, 2014](#)).

5. Conclusion

The present study on the physico-chemical characteristics of hot water of Surya Kund, Draupadi Kund and Yamunotri Tapt Kund revealed that the water of all the three hot water ponds is highly alkaline. All the three hot water ponds are the rich sources of thermophilic microbial diversity (bacteria, actinomycetes and fungi). *Bacillus subtilis*, *Bacillus licheniformis*, *Actinobacillus hominis*, *Streptomyces ruber*, *Nocardiopsis prasina*, *Thermobifida fusca* are the common microorganisms present in all the three hot water bodies. Almost all bacterial species were gram-positive. The present study can provide baseline data for further studies on similar aspects and also useful in providing characteristics of indigenous thermophilic microbial isolates that produce thermostable enzymes. Keeping in view the importance of these natural hot water bodies and their rich microbial diversity, an attempt should be made by the Government of Uttarakhand for the conservation and management of these high altitude natural hot water bodies.

Declarations

Author contribution statement

Rahul Kumar: Conceived and designed the experiments; Performed the experiments; Analyzed and interpreted the data; Contributed reagents, materials, analysis tools or data; Wrote the paper.

Ramesh C. Sharma: Conceived and designed the experiments; Analyzed and interpreted the data.

Funding statement

This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

Competing interest statement

The authors declare no conflict of interest.

Additional information

No additional information is available for this paper.

Acknowledgements

One of the authors (Rahul Kumar) is thankfully acknowledge for the fellowship given by the University Grant Commission, New Delhi through Hemvati Nandan Bahuguna Garhwal University (A Central University), Srinagar-Garhwal, Uttarakhand, India for undertaking the present work.

References

- Abou-Shanab, R.A.I., 2007. Characterization and 16S rDNA identification of thermotolerant bacteria isolated from hot springs. *J. Appl. Sci. Res.* 3, 994–1000.
- Akmar, H.N., Asma, I., Venugopal, B., Latha, L.Y., Sasidharan, S., 2011. Identification of appropriate sample and culture method for isolation of new thermophilic bacteria from hot pond. *Afr. J. Microbiol. Res.* 5 (3), 217–221.
- Anand, A., Aoyagi, H., 2019. Estimation of microbial phosphate accumulation abilities. *Sci. Rep.* 9, 4879.
- APHA, 2012. Standard methods for the examination of water and wastewater. In: Rice, E.W., Baird, R.B., Eaton, A.D., Clesceri, L.S. (Eds.), American Public Health Association (APHA), American Water Works Association (AWWA) and Water Environment Federation (WEF), Washington, D.C., USA, twenty-second ed.
- Ayitso, A.S., Onyango, D.M., 2016. Isolation and identification by morphological and biochemical methods of antibiotic producing microorganisms from the gut of *Macrotermes michaelensi* in maseno, Kenya. *J. Appl. Biol. Biotechnol.* 4 (1), 27–33.
- Baker, G.C., Gaffar, S., Cowan, D.A., Suharto, A.R., 2001. Bacterial community analysis of Indonesian hot springs. *FEMS Microbiol. Lett.* 200, 103–109.
- Beg, Q.K., Bhushan, B., Kapoor, M., Hoondal, G.S., 2000. Production and characterization of thermostable xylanase and pectinase from *Streptomyces* sp. QG-11-3. *J. Ind. Microbiol. Biotechnol.* 24, 396–402.
- Bandyopadhyay, S., Schumann, P., Das, S.K., 2013. *Pannonobacter indica* sp. nov., a highly arsenate-tolerant bacterium isolated from a hot spring in India. *Arch. Microbiol.* 195, 1–8.
- Bhusare, D.U., Wakte, P.S., 2011. Microbiological and physicochemical attributes of hot water sulphur pond of Unkeshwar. *J. Exp. Sci.* 2 (4), 4–6.
- Cerritos, R., Eguíalar, L.E., Avitia, M., Siebert, J., Travissano, M., 2011. Diversity of culturable thermo-resistant aquatic bacteria along an environmental gradient in Cuatro Cienegas, Mexico. *Antonie van Leeuwenhoek J. Microbiol.* 99, 303–318.
- Chan, C.S., Chan, K.G., Tay, Y.L., Chua, Y.H., Goh, K.M., 2015. Diversity of thermophiles in a Malaysian hot spring determined using 16S rRNA and shotgun metagenome sequencing. *Front. Microbiol.* 6, 1–14.
- Clesceri, L.S., Greenberg, A.E., Eaton, A.D., 1998. Standard Methods for the Examination of Water and Wastewater, twentieth ed. American Public Health Association, American Water Works Association, Water Environmental Federation, Baltimore, MA.
- Davis, K.E.R., Joseph, S.J., Janssen, P.H., 2005. Effects of growth medium, inoculum size, and incubation time on culturability and isolation of soil bacteria. *Appl. Environ. Microbiol.* 71 (2), 826–834.
- Earl, A.M., Losick, R., Kolter, R., 2008. Ecology and genomics of *Bacillus subtilis*. *Trends Microbiol.* 16 (6), 269–275.
- Fazlzadeh, M., Sadeghi, H., Bagheri, P., Poureshg, Y., Rostami, R., 2016. Microbial quality and physical-chemical characteristics of thermal springs. *Environ. Geochem. Health* 38, 413–422.
- Ghati, A., Sarkar, K., Paul, G., 2013. Isolation, characterization and molecular identification of Esterolytic bacteria from an Indian Hot pond. *Curr. Res. Microbiol. Biotechnol.* 1 (4), 196–202.
- Guzman, A.R., Taran, Y., Armienta, M.A., 2004. Geochemistry and origin of high-pH thermal springs in the Pacific coast of Guerrero, Mexico. *Geofisica Int.* 43 (3), 415–425.
- Hadad, D., Geresh, S., Sivan, A., 2005. Biodegradation of polyethylene by the thermophilic bacterium *Brevibacillus borstelensis*. *J. Appl. Microbiol.* 98 (5), 1093–1100.
- Haki, G.D., Gezmu, T.B., 2012. Physico-chemical properties of waters from some Ethiopian hot springs and the risk to the health of the community. *Greener J. Phys. Sci.* 2 (4), 138–140.
- Hamzah, Z., Rani, N.L.A., Saat, A., Wood, A.K., 2013. Determination of hot springs physico-chemical water quality potentially use for Balneotherapy. *Malays. J. Anal. Sci.* 17 (3), 436–444.
- Harley, J.P., Prescott, L.M., 2002. *Laboratory Exercises in Microbiology*, fifth ed. The MacGraw Hill publication, pp. 1–445.
- Homma, A., Tsukahara, H., 2008. Chemical characteristics of hot spring water and geological environment in the northernmost area of the Itoigawa Shizuoka tectonic line. *Bull. Earthq. Res. Inst. Univ. Tokyo* 83, 217–225.
- Huang, Q., Dong, C.Z., Dong, R.M., Jiang, H., Wang, S., Wang, G., Fang, B., Ding, X., Niu, L., Li, X., Zhang, C., Dong, H., 2011. Archaeal and bacterial diversity in hot springs on the Tibetan Plateau, China. *Extremophiles* 15, 549–563.
- Huq, A., Haley, B.J., Taviani, E., Chen, A., Hasan, N.A., Colwell, R.R., 2012. Detection, isolation, and identification of *Vibrio cholerae* from the environment. *Curr. Protocol. Microbiol.* Chapter 6: Unit6A.5.
- Javed, A., Iqbal, J., Asghar, U., Khan, F.A., Munshi, A.B., Siddiqui, I., 2009. A study to evaluate therapeutic properties of minerals of Manghopir Hot Spring, Karachi. *J. Chem. Soc. Pakistan* 31 (3), 396–401.
- Jiang, H., Dong, C., Huang, Q., Wang, G., Fang, B., Zhang, C., Dong, H., 2012. Actinobacterial diversity in microbial mats of five hot springs in Central and Central-Eastern Tibet, China. *Geomicrobiol. J.* 29, 520–527.
- Joshi, V., 2015. Hot Spring Complex. Ph.D. Thesis. L.S. Raheja School of Architecture, Maharashtra, pp. 1–81.
- Khalil, A., Anfoka1, G., Bdour, S., 2003. Isolation of plasmids present in thermophilic strains from hot springs in Jordan. *World J. Microbiol. Biotechnol.* 19, 239–241.
- Khalil, A.B., Sivakumar, N., Arslan, M., Saleem, H., Qarawi, S., 2018. Insights into *Brevibacillus borstelensis* AK1 through whole genome sequencing: a thermophilic bacterium isolated from a hot spring in Saudi arabia. *BioMed Res. Int.* 5862437.
- Kumar, B., Trivedi, P., Mishra, A.K., Pandey, A., Palni, L.M.S., 2004. Microbial diversity of soil from two hot ponds in Utтарanchal Himalaya. *Microbiol. Res.* 159, 141–146.
- Kumar, M., Yadav, A.N., Tiwari, R., Prasanna, R., Saxena, A.K., 2014. Evaluating the diversity of culturable thermotolerant bacteria from four hot springs of India. *Biodivers. Bioprospecting Dev.* 1 (3), 127.
- Kumar, N., Singh, A., Sharma, P., 2013. To study the physico-chemical properties and bacteriological examination of hot spring water from Vashishth region in Distt. Kullu of HP, India. *Int. Res. J. Environ. Sci.* 2 (8), 28–31.
- Kumar, R., Sharma, R.C., 2019a. Microbial diversity and physico-chemical attributes of two hot water springs in the Garhwal Himalaya, India. *J. Microbiol. Biotechnol. Food Sci.* 8 (6), 1249–1253.
- Kumar, R., Sharma, R.C., 2019b. Determination of microbial diversity and physico-chemical characteristics of two hot water ponds near Badrinath Shrine in the Uttarakhand, India. *IWRA (India) J.* 9 (1), 16–23.
- Kumar, R., Sharma, R.C., 2020. Thermophilic microbial diversity and physicochemical attributes of thermal springs in the Garhwal Himalaya. *Environ. Exp. Biol.* 18, 143–152.
- Kvist, T., Ahring, B.K., Westermann, P., 2007. Archaeal diversity in Icelandic hot springs. *FEMS Microbiol. Ecol.* 59, 71–80.
- Lopez, O.L., Cerdan, M.E., Gonzalez-Siso, M.I., 2013. Hot Spring Metagenomics. *Life* 2, 308–320.
- Marko, D.C., Saffert, R.T., Cunningham, S.A., Hyman, J., Walsh, J., Arbefeville, S., Howard, W., Pruessner, J., Safwat, N., Cockerill, F.R., Bossler, A.D., Patel, R., Richter, S.S., 2012. Evaluation of the Bruker Biotyper and Vitek MS matrix-assisted laser desorption ionization-time of flight mass spectrometry systems for identification of non-fermenting Gram-negative bacilli isolated from cultures from cystic fibrosis patients. *J. Clin. Microbiol.* 50, 2034–2039.
- Morello, J.A., Granato, P.A., Mizer, H.E., 2003. *Laboratory Manual and Workbook in Microbiology*, seventh ed. The MacGraw Hill Publication, pp. 1–285.
- Meyer-Dombard, D.R., Amend, J.P., 2014. Geochemistry and microbial ecology in alkaline hot springs of Ambitile Island, Papua New Guinea. *Extremophiles* 18, 763–778.
- Oarga, A., 2009. Life in extreme environments. *Revista De Biología E Ciencias Da Terra* 9 (1), 1–9.
- Pathak, A.P., Rathod, M.G., 2014. Cultivable bacterial diversity of terrestrial hot water pond of Unkeshwar, India. *J. Biochem. Technol.* 5 (4), 814–818.
- Pagaling, E., Grant, W.D., Cowan, D.A., Jones, B.E., Ma, Y., Ventosa, A., Heaphy, S., 2012. Bacterial and archaeal diversity in two hot spring microbial mats from the geothermal region of Tengchong, China. *Extremophiles* 16 (4), 607–618.
- Pryce, T.M., Palladino, S., Kay, I.D., Coombs, G.W., 2003. Rapid identification of fungi by sequencing the ITS 1 and ITS2 regions using an automated capillary electrophoresis system. *Med. Mycol.* 41, 369–381.
- Raja, H.A., Miller, A.N., Pearce, C.J., Oberlies, N.H., 2017. Fungal identification using molecular tools: a primer for the natural products research community. *J. Nat. Products* 80, 756–770.
- Rana, K.S., Sharma, R.C., Tivari, V., Kumar, R., 2018. Assessment of surface water quality of the Himalayan lake Beni Tal, India. *Curr. Res. Hydrol. Water Resources* 1, 1–11.
- Rawat, S., 2015. Bacterial diversity of a sulphur pond in Uttarakhand, India. *Adv. Appl. Sci. Res.* 6 (4), 236–244.

- Reyzenbach, A.L., Ehringer, M., Hershberger, K., 2000. Microbial diversity at 83°C in calcite springs, Yellowstone national park: another environment where the aquificles and "Korarchaeota" coexist. *Extremophiles* 4, 61–67.
- Risch, M., Radjenovic, D., Han, J.N., Wydler, M., Nydegger, U., Risch, L., 2010. Comparision of MALDI-TOF with conventional identification of clinically relevant bacteria. *Swiss Med. Wkly.* 140, w13095.
- Rohomania, T., Saha, M.L., Hossain, A., Rahman, M.S., 2015. Morphological and biochemical characterization of bacteria isolated from fresh and salted Hilsa, *Tenualosus ilisha* (Hamilton, 1822). *Bangladesh J. Microbiol.* 32 (1&2), 7–13.
- Saggu, G., Shrivastava, D., 2013. Study on thermophiles reveals the presence of *Actinobacillus lignieresii* in cattle compost. *Int. J. Appl. Sci. Biotechnol.* 1 (2), 33–41.
- Sen, S.K., Mohapatra, S.K., Satpathy, S., Rao, G.T.V., 2010. Characterization of hot water pond source isolated clones of bacteria and their industrial applicability. *Int. J. Chem. Res.* 2, 1–7.
- Shahcheraghi, S.H., Ayatollahi, J., Lofti, M., 2015. Applications of *Bacillus subtilis* as an important bacterium in medical sciences and human life. *Trop. J. Med. Res.* 18, 1–4.
- Sharma, B., Verma, R., Dev, K., Thakur, R., 2012. Molecular characterization of Manikaran hot spring microbial community by 16S rRNA and RAPD analysis. *Biotechnol. Indian J.* 6 (8,9), 254–266.
- Sharma, A., Pandey, A., Shouche, Y.S., Kumar, B., Kulkarni, G., 2008. Characterization and identification of *Geobacillus* spp. isolated from Soldhar hot pond site of Garhwal Himalaya, India. *J. Basic Microbiol.* 48, 1–8.
- Sharma, N., Vyas, G., Pathania, S., 2013. Culturable diversity of thermophilic microorganisms found in hot springs of northern Himalayas and to explore their potential for production of industrially important enzymes. *Scholars Acad. J. Biosci.* 1 (5), 165–178.
- Sherpa, M.T., Das, S., Thakur, N., 2013. Physicochemical analysis of hot water springs of Sikkim-Polok tatopani, borong tatopani and Reshi tatopani. *Recent Res. Sci. Technol.* 5 (1), 63–67.
- Singh, H.K., Chandrasekharam, D., Vaselli, O., Trupti, G., Singh, B., Lashin, A., Arifi, N.A., 2015. Physico-chemical characteristics of Jharkhand and West Bengal thermal springs along SONATA mega lineament, India. *J. Earth System Sci.* 124 (2), 419–430.
- Takahashi, S., Tomita, J., Nishioka, K., Hisada, T., Nishijima, M., 2014. Development of a prokaryotic universal primer for simultaneous analysis of bacteria and archaea using next-generation sequencing. *PLoS One* 9 (8), e105592.
- Takacs-Vesbach, C., Mitchell, K., Jackson-Weaver, O., Reyzenbach, A., 2008. Volcanic calderas delineate biogeographic provinces among Yellowstone thermophiles. *Environ. Microbiol.* 10, 1681–1689.
- Tekere, M., Lötter, A., Olivier, J., Venter, S., 2015. Bacterial Diversity in Some South African Hot Water Ponds: A Metagenomic Analysis. *Proceedings World Geohot water Congress*, Melbourne, Australia, pp. 1–8.
- Ulrich, N., Nagler, K., Laue, M., Cockell, C.S., Setlow, P., et al., 2018. Experimental studies addressing the longevity of *Bacillus subtilis* spores – the first data from a 500-year experiment. *PLoS One* 13 (12), e0208425.
- Usyk, M., Zolnik, C.P., Patel, H., Levi, M.H., Burk, R.D., 2017. Novel ITS1 fungal primers for characterization of the mycobiome. *mSphere* 2 (6), 1–11.
- Vieille, C., Zeikus, G.J., 2001. Hyperthermophilic enzymes: sources, uses, and molecular mechanisms for thermostability. *Microbiol. Mol. Biol. Rev.* 65 (1), 1–43.
- Wemheuer, B., Taube, R., Akyol, P., Wemheuer, F., Daniel, R., 2013. Microbial Diversity and Biochemical Potential Encoded by thermal spring Metagenomes Derived from the Kamchatka Peninsula. *Archaea Article ID* 136714.