



Genome Sequence of Mycobacterium Phage LilHazelnut

Ryan A. Shanks,^a Ashley N. Hazel,^a William H. Jones,^a Miriam Segura-Totten^a

^aDepartment of Biology, University of North Georgia, Dahlonega, Georgia, USA

ABSTRACT Here, we describe LilHazelnut, a novel mycobacteriophage that infects *Mycobacterium smegmatis* mc²155. LilHazelnut is a cluster Q phage that shares 99% nucleotide identity with phage Giles, is 53,746 bp in length, and has a G+C content of 67.5%. LilHazelnut is a temperate *Siphoviridae* virus, as is typical of cluster Q family members.

The increasing number of isolated and characterized bacteriophage provides a valuable database of genetic diversity allowing for the identification of both shared and novel gene products between bacteriophage species (1). Here, we describe a new mycobacteriophage, LilHazelnut, isolated as part of the Science Education Alliance Phage Hunters Advancing Genomics and Evolutionary Science (SEA-PHAGES) program (2) from a single plaque in an enrichment of a 0.22- μ m filtered 7H9 liquid mediumwashed soil sample from North Georgia (coordinates 24.527904, 83.987719) using the host *Mycobacterium smegmatis* mc²155 and purified using three consecutive serial dilutions. LilHazelnut, a *Siphoviridae* virus, has an icosahedral head and a 195-nm tail, and it produces clear round plaques of 0.3 mm with hazy edges.

Genomic DNA was isolated from LilHazelnut lysates with a Wizard DNA extraction kit (Promega). A NEBNext Ultra II FS kit with dual-indexed barcoding was used to prepare a sequencing library from genomic DNA. This library plus those from 47 other phages were pooled and run on an Illumina MiSeq instrument. This assay yielded ~895,000 single-end 150-base reads from the LilHazelnut library, and when assembled, these reads provided ~2,384-fold coverage of the LilHazelnut genome. These raw reads were assembled using Newbler v2.9 with default settings. The resulting single phage contig was checked for completeness, accuracy, and phage genomic termini using Consed v29 as previously described (3). LilHazelnut contains a circularly permuted genome consisting of 53,746 bp with a G+C content of 67.5%.

The LilHazelnut genome was annotated using Glimmer v3.02 (4), GeneMark v2.5p (5), BLASTP v2.7.1 (https://blast.ncbi.nlm.nih.gov/), HHPred v3.0beta (6), and Phamerator (7). The E value cutoff used for BLASTP and HHPred was 10 e-4. Automatic genome annotation followed by manual refinement revealed that LilHazelnut is part of the cluster Q phages and is predicted to have no tRNAs/transfer-messenger RNA (tmRNA) genes but has 85 protein-coding genes and 35 genes with assigned functions. Functional assignments for predicted proteins included virion structure and assembly proteins, as well as terminase, integrase, excise, and lysin A and B proteins. Except for genes 2 to 4, both ends of the phage genome are transcribed in the forward direction, while the genes in the middle portion of the genome are transcribed in the reverse direction.

LilHazelnut displays genome-wide similarity to the other 11 temperate phages in this cluster, sharing 99.99% nucleotide identity with its closest relative Giles (GenBank accession no. NC_009993). Identity to Giles was determined through a nucleotide BLAST search (https://blast.ncbi.nlm.nih.gov/Blast.cgi). Genes *gp29* and *gp30*, which form part of the integration cassette in LilHazelnut, are conserved in phages Kinbote (GenBank accession no. KT22940) and Giles. The attachment site (*attP*) in LilHazelnut is identical to that

Segura-Totten M. 2019. Genome sequence of Mycobacterium phage LilHazelnut. Microbiol Resour Announc 8:e00431-19. https://doi.org/ 10.1128/MRA.00431-19.

Citation Shanks RA, Hazel AN, Jones WH,

Editor Simon Roux, DOE Joint Genome Institute

Copyright © 2019 Shanks et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Miriam Segura-Totten, mstotten@ung.edu.

A.N.H. and W.H.J. contributed equally.

Received 11 April 2019 **Accepted** 18 April 2019 **Published** 9 May 2019 of Giles, and is also found upstream of the integrase gene (positions 25134 to 25179). Similarly, *gp47*, which in Giles functions as a phage repressor, is conserved in LilHazelnut as *gp49*. A putative DNA binding domain in this repressor is conserved across all cluster Q members (8).

Data availability. LilHazelnut is available at GenBank with accession no. MF919517. Sequencing reads are part of the Sequence Read Archive with SRA accession no. SRX5282798 under BioProject accession no. PRJNA488469.

ACKNOWLEDGMENTS

This research was conducted as part of the Science Education Alliance Phage Hunters Advancing Genomics and Evolutionary Science (SEA-PHAGES) program, supported by the Howard Hughes Medical Institute (HHMI). Funding was provided in part by a UNG College of Science and Mathematics professional development grant.

We thank John Shields and Mary Ard at UGA Georgia Electron Microscopy for imaging support. We thank Debbie Jacobs-Sera, Dan Russell, and Welkin Pope for guidance during manuscript preparation. We thank Amelia Claire Dempsey Arthur, Ethan Noah Strickland, Gabriella Alexandra Fleck, Grant Lemert Zacher, Haley Jordan Polane, John Martin Corbett, Katelyn D. Shook, Lauren Emily Colston, Lauren Nicole Silvers, Reily Amanda Pilcher, and Vanessa Renee Lewis for their assistance in the isolation and genomic annotation of mycobacteriophage LilHazelnut.

REFERENCES

- Hatfull GF. 2018. Mycobacteriophages. Microbiol Spectr 6:GPP3-0226-2018. https://doi.org/10.1128/microbiolspec.GPP3-0026-2018.
- Hanauer DI, Graham MJ, Sea P, Betancur L, Bobrownicki A, Cresawn SG, Garlena RA, Jacobs-Sera D, Kaufmann N, Pope WH, Russell DA, Jacobs WR, Jr, Sivanathan V, Asai DJ, Hatfull GF. 2017. An inclusive Research Education Community (iREC): impact of the SEA-PHAGES program on research outcomes and student learning. Proc Natl Acad Sci U S A 114:13531–13536. https://doi.org/10.1073/pnas.1718188115.
- Russell DA. 2018. Sequencing, assembling, and finishing complete bacteriophage genomes, p 109–125. *In* Clokie MRJ, Kropinski AM, Lavigne R (ed), Bacteriophages: methods and protocols, vol 3. Springer, New York, NY.
- Delcher AL, Bratke KA, Powers EC, Salzberg SL. 2007. Identifying bacterial genes and endosymbiont DNA with Glimmer. Bioinformatics 23:673–679. https://doi.org/10.1093/bioinformatics/btm009.
- Besemer J, Borodovsky M. 2005. GeneMark: Web software for gene finding in prokaryotes, eukaryotes and viruses. Nucleic Acids Res 33: W451–W454. https://doi.org/10.1093/nar/gki487.
- Söding J, Biegert A, Lupas AN. 2005. The HHpred interactive server for protein homology detection and structure prediction. Nucleic Acids Res 33:W244–W248. https://doi.org/10.1093/nar/gki408.
- Cresawn SG, Bogel M, Day N, Jacobs-Sera D, Hendrix RW, Hatfull GF. 2011. Phamerator: a bioinformatic tool for comparative bacteriophage genomics. BMC Bioinformatics 12:395. https://doi.org/10.1186/1471-2105-12 -395.
- Dedrick RM, Marinelli LJ, Newton GL, Pogliano K, Pogliano J, Hatfull GF. 2013. Functional requirements for bacteriophage growth: gene essentiality and expression in mycobacteriophage Giles. Mol Microbiol 88: 577–589. https://doi.org/10.1111/mmi.12210.