Case Report

A Huge Pelvic-Abdominal Malignant GIST Tumour in a Patient with Neurofibromatosis Type 1: Case Report and Literature Review

Islam Omar (D),^{1,2} Hani Alsaati,² and Ejaz Waris²

¹Furness General Hospital, UHMBT, NHS, UK ²Oncology Center, King Hamad University Hospital, Bahrain

Correspondence should be addressed to Islam Omar; islamfawzyomar@hotmail.com

Received 29 May 2019; Revised 25 August 2019; Accepted 23 December 2019; Published 4 January 2020

Academic Editor: Mauro Cives

Copyright © 2020 Islam Omar et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Gastrointestinal stromal tumours are rare tumours of the gastrointestinal tract (GIT) accounting for 0.1%-3% of all gastrointestinal tumours. The most common location is the stomach (55%) followed by the small bowel (31.8%), colon (6%), other various locations (5.5%), and the oesophagus (0.7%). They may also occur in extraintestinal locations. The signs and symptoms of GIST depend on the tumour's location and size. Gastrointestinal bleeding is one of the most common symptoms. Other signs and symptoms include abdominal discomfort, pain or distention; intestinal obstruction, and weight loss. The association between the development of GISTs and neurofibromatosis 1 (NF1) has been established. NF1-associated GISTs tend to have a distinct phenotype, and the absence of KIT/PDGRF α mutations in turn has implications on further management when they do not respond well to imatinib treatment. Here, we present one of the largest GISTs reported in the literature with a total volume of $25.3 \times 20 \times 14$ cm + $27.9 \times 23 \times 8$ cm and an overall weight of 7.3 kg, which developed in a 43-year-old female patient with NF1 and was resected on an emergency basis due to the rapid deterioration and development of abdominal compartment syndrome. Pathology assessment showed a malignant GIST composed of spindle cells with elongated nuclei with necrosis, marked pleomorphism and numerous giant cell. The mitotic count was >15/50 HPF, Ki 67 was 80%, and the lymphovascular invasion was clear. Immunohistochemistry investigations showed that Vimentin, CD117, and DOG1 were positive, while BCL-2 and CD99 were focal positives. Pan-CK, S-100, CD34, Desmin, SMA, and HMB-45 were negatives.

1. Introduction

Gastrointestinal stromal tumours are rare tumours of the GIT (see comment above) accounting for 0.1%-3% of all gastrointestinal tumours [1, 2]. The most common location is gastric (55%) followed by the small bowel (31.8%), colon (6%), other various locations (5.5%), and the oesophagus (0.7%) [3]. They may also occur in the extraintestinal locations like the mesentery and omentum and in exceptionally rare sites like the gallbladder, urinary bladder, and prostate [4].

The tumour shows no strong sex predilection and usually occurs in adults [5], with a mean reported age of 50.6 years [6]. GIST is extremely uncommon in children and adolescents [7]. Some instances of GISTs show a pattern of familial incidence suggestive of genetic predisposition, whereas others are associated with neurofibromatosis and Carney's triad [5].

GISTs are believed to arise from the interstitial cells of Cajal or related stem cells [8]. In most cases of GISTs, there is a mutation of C-KIT oncogene, which is responsible for the formation of a protein called KIT. Accordingly, KIT (CD117) positivity is observed in 95% of cases [2]. Plateletderived growth factor receptor alpha (PDGFRA α) mutations play a role in GIST pathogenesis [9].

Benign GISTs outnumber the malignant ones by a margin of 10:1. Clinical presentation depends on the site and size of the tumour and may include abdominal pain, gastrointestinal bleeding from ulceration of the overlying mucosa, or signs of obstruction. However, small tumours



FIGURE 1: External compression on the sigmoid and descending colon with neither obstruction nor intraluminal lesion.

may be asymptomatic. The major clinical findings are upper abdominal ulcer-like pain, dyspepsia, iron-deficiency anaemia, gastrointestinal bleeding, nausea, vomiting, palpable abdominal mass, and weight loss [10, 11]. Also, GISTs with an intra-abdominal abscess have been reported [12, 13].

Many studies confirmed the association between the development of GISTs and neurofibromatosis 1 (NF1) [14–17]. GISTs associated with NF1 seem however to have a distinct phenotype and the absence of KIT/PDGRF α mutations which has, in turn, an implication on further management where they do not respond well to Imatinib treatment [18–20].

Here, we present a case of a huge pelvic-abdominal GIST in a known patient of NF1 who showed rapid progression over a short time mounting to abdominal compartment syndrome. Consequently, she underwent emergency surgical excision.

2. Case Presentation

A 42-year-old female is a known case of familial NF1. She and her identical twin in addition to her only brother are all affected by NF1. She has no family history of malignancy, has not had previous surgery, and has no other comorbidities. Initially, she presented with fatigue, cough, and shortness of breath. She sought medical advice, and a low haemoglobin of 4 g/dl was discovered. She was resuscitated and received a blood transfusion. She had a history of constipation and change of stool calibre and melena. Clinical examination showed an abdominal mass which was confirmed by the US ultrasound later. An urgent OGD and a colonoscopy were done. OGD revealed normal upper GI study while the colonoscopy showed external compression on the sigmoid and descending colon with neither obstruction nor intraluminal lesion (Figure 1).

After stabilisation, the patient was then referred to our tertiary oncology centre for further evaluation. A CT scan (Figure 2) was done which revealed a huge cystic mass occupying almost the whole pelvic and abdominal cavity, which was lobulated with enhancing peripheral soft tissue components and septations. Moreover, a mild free-flowing peritoneal fluid was noted with diffuse subcutaneous tissue oedema. Of note, there were no signs of metastasis to the



FIGURE 2: A huge cystic mass occupying almost the whole pelvic and abdominal cavity. The mass is lobulated with enhancing the peripheral soft tissue components and septations, displacing all bowel posteriorly.



FIGURE 3: Whole-body PET CT showing multiple amalgamated masses exhibiting peripheral patchy FDG activity with low-grade FDG avid areas of peritoneal thickening/fat stranding suggesting peritoneal involvement. Low-grade FDG avid small prehepatic node measures 8 mm with diffusely activated bone marrow.

liver, lung, or the bones, and the scanned parts of the bowel loops showed no abnormalities.

Tumour markers were done and showed AFP ($2.8 \mu g/L$), CA 125 (154.7 U/ml), CA 15-3 (10.5 U/ml), CEA ($0.3 \mu g/L$), CA 19-9 (5.4 U/ml), and B-HCG (0.9 mIU/ml). For staging, a whole-body PET CT scan (Figure 3) was done and showed multiple amalgamated, well-defined, oval-to-round soft tissue masses extending from right subhepatic and lumbar regions just behind the anterolateral aspects of the anterior abdominal wall down to the pelvis, compressing the bowel loops. These masses exhibited heterogeneous fluid and soft tissue densities as well as peripheral patchy FDG activity.

They were associated with low-grade FDG, avid areas of peritoneal thickening, and fat stranding with pelvic ascites,



FIGURE 4: Intraoperative findings showing a huge lobulated mass, occupying most of the pelvic and abdominal cavities with the involvement of the sigmoid colon and small bowel. The weight of the resected mass exceeded 7.3 kg.

suggesting peritoneal involvement. Also, a low-grade FDG avid small prehepatic node was detected which measured 8 mm in diameter. Additionally, features of a diffusely activated bone marrow were noted, likely proliferative in response to anaemia.

Given the previous scans, the origin of the mass was not clear and was thought to be of ovarian origin. The patient was being optimised and prepared for surgery. At that time, melena was stopped, but the patient continued to complain of shortness of breath, constipation, abdominal pain, and distention.

In an attempt to fully optimize the patient for definitive surgery, all the efforts were carried out to ameliorate the abdominal distention and the mass effects through NGT, urinary catheterization, and strict fluid balance management. Unfortunately, the mass effect was out of control due to the huge size of the tumour and the patient started to experience tachycardia and tachypnea. Abdominal pressure measurement showed a pressure approaching 26 mmHg, a value that met a grade IV ACS [21].

At that stage, the decision was taken to do emergency surgery to avoid the expected consequences of ACS [22]. Exploratory laparotomy was performed after bilateral ureteral stenting. A huge intra-abdominal mucinous mass was found (Figure 4) coming from the pelvis with no clear origin. There was involvement of the peritoneum, small bowel, sigmoid colon, and upper rectum. Moreover, multiple areas of the small bowel were involved, around 80 cm from the ileocaecal junction. Therefore, excision of the mass was performed with bilateral salpingo-oophorectomy. Since the sigmoid colon was found adherent to the tumour, resection of the sigmoid colon and colostomy creation were done. Moreover, the involved segments of the small bowel were resected and primarily anastomosed. The grossly involved peritoneum was resected.

On gross pathological examination, the overall weight of the specimen was 7.3 kg. The first specimen (Figure 5(a)) was a huge mass with dimensions of $25.3 \times 20 \times 14$ cm. The mass was firm, greyish white with nodular surface, and partially capsulated. There were multiple foci of haemorrhage, congestion, and necrosis. The cut surface of the mass showed a variegated appearance with focal greyish white areas. There were also focal areas of mucinous cystic degeneration, haemorrhage, and necrosis. A small bowel segment 8.5 cm in length by 3.2 cm in circumference was found adherent to the mass. On opening the bowel segment, its mucosa was unremarkable. However, the serosal surface adhered to the mass. The second specimen (Figure 5(b)) came with dimensions of $27.9 \times 23 \times 8$ cm. The external surface of the mass was nodular, partially capsulated, and showed foci of congestion, haemorrhage, and necrosis. Along with that, an exudative membrane was found on the surface. The cut surface was glistening, shiny, and having a variegated appearance with focal areas of haemorrhage and necrosis in addition to marked cystic degeneration.

Histopathology examination (Figure 6) of the two huge masses showed features of a malignant neoplasm. It was composed of spindle cells with elongated nuclei arranged in fascicles. Moreover, there was marked pleomorphism and numerous tumour giant cells. The mitotic count was >15/50 HPF. Additionally, there were poorly differentiated areas with 20% necrosis. The tumour was of the high-grade type.

Risk assessment according to Fletcher et al.'s criteria [23] showed that the tumour was of high risk. Margins of the resected small and large bowel loops were free from tumour invasion. However, evidence of lymphovascular invasion was clear. Immunohistochemistry investigations (Figure 7) showed that Vimentin, CD117, and DOG1 were positives, BCL-2 and CD99 were focal positives, Pan-CK, S-100, CD34, Desmin, SMA, and HMB-45 were negative, and Ki-67 was 80%.

Two segments of the small bowel were resected weighing 675 g. The first segment measured 26 cm in length by 4.8 cm in circumference. A polypoid, soft-to-firm, and tan-to-grey in colour mass was found adherent to its serosal surface measuring $8 \times 5.2 \times 1.3$ cm. It was 10.1 cm away from the closest margin, and 17.7 cm away from the distant margin. On opening the small bowel segment, the entire length of mucosa was gangrenous with an area of perforation identified, measuring 7.6 mm, it was 8.5 cm away from the closest margin.

The second segment of the small bowel measuring 27.9 cm in length and a 5.3 cm circumference, with a polypoid grey soft-to-firm mass, was found adherent to the serosal surface, measuring $36.5 \times 6.7 \times 3.4$ cm. The mass was 9.5 cm away from the closest margin and 9.6 cm away from the distant margin. Grossly, it did not appear to involve the mucosa. On opening the lumen, the mucosal surface, at the level of



FIGURE 5: (a) Gross pathology of the first mass showing a huge mass with dimensions of $25.3 \times 20 \times 14$ cm which was greyish white in color with nodular surface and partially capsulated. There were multiple foci of hemorrhage, congestion, and necrosis. (b) Gross pathology of the second mass measuring $27.9 \times 23 \times 8$ cm. Its external surface was nodular, partially capsulated, and showed foci of congestion, haemorrhage, and necrosis.

mass attached to the serosa, was flattened and the remaining mucosa showed an area of stricture measuring 2.5×1 cm, 4.3 cm away from the closest margin with no perforation. Histopathology examination of the bowel loops showed thinning out of the wall with tumour infiltration into the mesenteric fat, reaching up to the muscular layer but the resection margins were negative.

One segment of the sigmoid colon was included and measured 27.9 cm in length by 8 cm in circumference. The specimen was received with an attached mesentery and weighed 299 g. A polypoid grey tan firm mass was found adherent to the mesenteric fat, measuring $10.5 \times 3.6 \times 2.1$ cm, and it was 2.1 cm away from the closest margin. Upon opening the lumen, the mucosal surface along the entire segment was normal with no strictures, perforation, or diverticulosis. Histopathology examination of the sigmoid colonic wall showed tumour infiltration into the mesenteric fat reaching up to the muscular layer with four reactive lymph nodes. The resection margins were clear.

Another specimen was retrieved and included two pieces of the peritoneum. The first from the right side measured $13 \times 5.3 \times 2.2$ cm along with a nodule measuring $2.8 \times 2 \times 2.1$

1.5 cm and both weighed 47 g. The second piece came from the left peritoneal layer which measured $18 \times 5.5 \times 0.8$ cm and weighed 25 g. The right peritoneum specimen was positive for tumour infiltration where sections revealed tumour infiltration of the fibrofatty tissue. The left peritoneal layer showed small ectopic adrenal tissue and was positive for tumour infiltration.

Specimens of the ovaries and fallopian tubes revealed a left ovary measuring $3.5 \times 2.4 \times 1.2$ cm with the left fallopian tube measuring 5×1.2 . The overall weight of the left ovary and fallopian tube was 12 g. The right ovary and the attached fallopian tube with surrounding fibrofatty tissue weighed 9 g. The right ovary measured $3.2 \times 1.8 \times 1.4$ cm. The attached right fallopian tube measured 4.2×1.2 cm. The attached fibrofatty tissue showed marked areas of congestion. Histopathology examination of the left ovarian mass revealed a normal ovary and fallopian tube with tumour infiltration into the paratubal tissue. The right ovarian mass was free of invasion.

The conclusion of the pathology assessment came as high-risk GIST (pT4, pN0, pMx) with tumour invasion into the left paratubal tissue, small and large bowel, in addition



FIGURE 6: Histopathology of the tumor. (a, b) The tumor composed of spindle cells with elongation. (c) Areas with necrosis. (d) High mitotic figures.



FIGURE 7: Immunohistochemistry staining. (a) Positive CD117. (b) Positive DOG1. (c) Desmin negative. (d) \$100 negative.

to the right and left peritoneal tissues. The patient had an uneventful postoperative course and was discharged home. Her case was discussed in the national tumour board—MDT, which recommended starting Imatinib therapy and genetic analysis. Unfortunately, genetic analysis was not available in any local centre. Every effort was made to send the tissue blocks abroad for genetic analysis; however, logistical obsta- our kno

cles aborted these endeavours. Although the patient received the first dose of Imatinib, her condition started to deteriorate over the next 2 months after surgery with persistent vomiting and gradual abdominal distention. However, her stoma was functioning and a barium follow-through study confirmed the patency of her bowel. Because of the multiple admissions due to persistent vomiting and abdominal distention, a CT scan was done which confirmed an aggressive recurrence. Because of the severe distention and again abdominal compartment syndrome, the patient was taken to the theatre where an aggressive inoperable tumour was encountered. Laparostomy was performed. The abdomen was left open and covered with a Bogota bag. A few days after the second surgery, the patient passed away.

3. Discussion

The signs and symptoms of GIST depend on the tumour's location and size, with highly malignant GISTs typically being large and symptomatic at the time of diagnosis. Gastrointestinal bleeding is one of the most common symptoms. Other signs and symptoms include abdominal discomfort, pain or distention, intestinal obstruction, and weight loss [24].

In this case, there was a typical presentation with anaemia, melena, and gradual abdominal distention. Although there was no complete obstruction, the patient gave a history of change of stool calibre and constipation, manifestations that go with the external compression exerted by the huge tumour on the bowel.

GISTs arise in the muscularis propria layer of the stomach or intestinal wall. Small GISTs form intramural or serosal nodules and, as they grow and expand, may develop intraluminal, intramural, and extraluminal components to varying degrees. The extraluminal component of a malignant GIST may be so large that it is difficult to determine the site of origin [24, 25].

On the radiological basis, the tumour in our patient was first thought to have originated from the ovaries due to its pelvic-abdominal position and close proximity to the gonads. However, on histopathology assessment, it was proved to invade the left paratubal tissue planes only. Given its huge size and invasion of both the sigmoid colon as well as the small bowels, the origin is not clear; however, the invasion to the small bowel was more aggressive with perforation and stricture formation suggesting a small bowel origin.

Given the weight and volume of our tumour (overall weight of 7.3 kg— $25.3 \times 20 \times 14 \text{ cm}$ for the first mass and $27.9 \times 23 \times 8 \text{ cm}$ for the second mass), it is one of the largest GISTs reported in the literature. Koyuncuer et al. [26] reported a huge GIST originating from the stomach in a 43-year-old male, which was 6.109 kg in weight and measured $39 \times 27 \times 14 \text{ cm}$. Also, Cappellani et al. [27] reported a gastric GIST in a 67-year-old male, weighing 8.5 kg and measuring $37 \times 24 \times 13 \text{ cm}$. Moreover, Cruz et al. described a gastric GIST in a 37-year-old male, which was $32 \times 25 \times 21$ in size and 3.75 kg in weight. Accordingly, to the best of

our knowledge, our patient had the largest reported GIST in the literature with a volume of $25.3 \times 20 \times 14 + 27.9 \times 23 \times 8$ cm and the second heaviest tumour weighing 7.3 kg.

GISTs are potentially aggressive forms of cancer that may develop anywhere in the GI tract [28]. They most frequently metastasize within the abdominal cavity, especially the liver and peritoneum. Bone and lung metastases are far less common [28, 29]. Very rare metastases to the skeletal muscles, adrenal gland, brain, testicles, and heart have also been reported [30–33]. Sizes larger than 10 cm cystic changes, high cellularity, mitotic figures >10/50 HPF, and coagulative necrosis are all more likely to be associated with metastasis [34].

In our patient, the histopathology assessment classified the tumour as a high-risk malignant tumour. Although distant spread to the liver or bones is excluded based on radiological staging, the tumour is proved to be locally aggressive spreading to the peritoneum, bowels, and paratubal tissues. This is expected given the huge nature of the tumour, a mitotic count of >15/50 HPF, and Ki-67of 80%.

Association between GIST and NF1 is established since an old autopsy study had documented a GIST in one-third of the NF1 patients [35]. NF1-associated GISTs are typically reported as multiple, generally low-grade tumours that affect the jejunum, ileum, duodenum, and stomach [15, 36]. Salvi PF et al. [19] in a systematic review of 252 patients with NF1-associated GISTs reported that patients affected by NF1-associated GISTs were younger and tumours were significantly smaller. Moreover, tumours were located mainly in the jejunum and ileum in the NF1 subgroup, whereas the main localization in the sporadic group was the stomach. They also reported a prevalence of low-risk criteria in the NF1 subgroup compared with the sporadic GISTs.

In our case, the patient was young (43 years old), with the tumour most probably originating from the small bowel—-criteria that go with the previous literature. However, the tumour was huge and of the high-risk category, which were both striking features.

The genetic basis for the association between NF1 and GISTs is being elucidated. The genetic defect of NF1 disease is a mutation in the NF1 gene which encodes neurofibromin, a tumour suppressor protein that regulates the cellular proliferation via inactivation of RAS through GTPase activity. RAS is known to stimulate signal transduction through the MAP kinase pathway when activated. The resultant loss of the function of neurofibromin predisposes to the development of benign and malignant tumours [37-39]. NF1-related GI disease has been described to occur in three principal forms: neurogenic tumours, stromal tumours, and neuroendocrine tumours [40-42]. The most common of these GI manifestations in NF1 is GIST with one study indicating an incidence of 7% in the NF1 population and another reporting a 150-fold increased risk as compared to the general population [35, 38].

GISTs in adults are typically sporadically occurring and are associated with somatic mutations in the KIT or PDGFR α gene. However, 10%–15% of GISTs lack KIT or PDGFR α mutations. These GISTs can be associated with NF1 or can represent succinate dehydrogenase (SDH)-deficient GISTs that include GIST in the Carney triad and the Carney-Stratakis syndrome [43, 44].

GISTs associated with Carney Triad, Carney Stratakis syndrome, along with young and paediatric GISTs have been documented to have a loss of succinate dehydrogenase subunit B (SDHB) expression [20, 45, 46].

Based on the SDHB expression, it has been recently proposed that GISTs could be differentiated into two characteristic subgroups: type 1 SDHB-positive and type 2 SDHB-negative [45]. SDHB-positive GISTs usually occur in adults with no predilection of the tumour's locations, show homogeneous male-to-female ratio, present KIT or PDGFR α mutations, and, generally, may benefit from the Imatinib treatment. SDHB-negative GISTs occur usually in paediatric and young female patients, locate almost exclusively in the stomach and present an epithelioid morphology. These tumours are usually c-kit and PDGFR α wild-type and do not respond to the molecular treatment with Imatinib [47, 48].

Even though NF1-associated GISTs have been documented to be type 1 SDHB-positive tumours [20], they could be differentiated by several features including the predilection of localization to the jejunum and small intestines, common tumour multiplicity, the lack of GIST-specific mutations (kit and PDGFR α wild type); and they are KIT-positive with hyperplasia of ICCs. Moreover, and alike the SDHB type 2 tumours, they do not respond well to Imatinib treatment [15, 20, 49–51].

Given the above, the therapeutic challenges for advanced NF1-associated GISTs are evident with expected resistance to Imatinib therapy. However, Sunitinib is the second-line tyrosine kinase inhibitor and has shown activity with clinical benefit in 56% of wild-type patients and can be used for the patients who develop resistance to Imatinib as well [52].

4. Conclusion

NF1-related GISTs represent a unique entity of GISTs relevant to their different molecular basis and presentation. When advanced and unresectable, they pose a great challenge due to resistance to Imatinib therapy. Here, we present one of the largest GISTs reported in the currently available literature with an overall weight of 7.3 kg and volume of $25.3 \times 20 \times 14 + 27.9 \times 23 \times 8$ cm, which was resected on an emergency basis due to the rapid deterioration and development of abdominal compartment syndrome.

Ethical Approval

This study is approved by the ethical committee.

Consent

Written informed consent was obtained from the patient for publication of this case report and accompanying images.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

References

- S. Phongkitkarun, C. Phaisanphrukkun, J. Jatchavala, and E. Sirachainan, "Assessment of gastrointestinal stromal tumors with computed tomography following treatment with imatinib mesylate," *World Journal of Gastroenterology*, vol. 14, no. 6, pp. 892–898, 2008.
- [2] I. K. Skandalos, N. F. Hotzoglou, K. C. Matsi, X. A. Pitta, and A. I. Kamas, "Giant extra gastrointestinal stromal tumor of lesser omentum obscuring the diagnosis of a choloperitoneum," *International Journal of Surgery Case Reports*, vol. 4, no. 10, pp. 818–821, 2013.
- [3] K. Søreide, O. M. Sandvik, J. A. Søreide, V. Giljacac, A. Jureckovad, and V. R. Bulusue, "Global epidemiology of gastrointestinal stromal tumours (GIST): a systematic review of population-based cohort studies," *Cancer Epidemiology*, vol. 40, pp. 39–46, 2016.
- [4] A. Mekni, I. Chelly, H. Azzouz et al., "Extragastrointestinal stromal tumor of the urinary wall bladder: case report and review of the literature," *Pathologica*, vol. 100, no. 3, pp. 173– 175, 2008.
- [5] J. R. Goldblum, "Gastrointestinal stromal tumorsChapter 26 - Mesenchymal Tumors of the GI Tract," in Surgical Pathology of the GI Tract, Liver Biliary Tract, and Pancreas, R. D. Odze and J. R. Goldblum, Eds., pp. 681–694, Saunders, Philadelphia, PA, USA, 2nd edition, 2009.
- [6] M. Vij, V. Agrawal, A. Kumar, and R. Pandey, "Cytomorphology of gastrointestinal stromal tumors and extragastrointestinal stromal tumors: a comprehensive morphologic study," *Journal of Cytology*, vol. 30, no. 1, pp. 8–12, 2013.
- [7] M. Benesch, "Gastrointestinal stromal tumors," in *Rare Tumors in Children and Adolescents*, Pediatric Oncology, D. Schneider, I. B. Brecht, and T. A. Olson, Eds., pp. 279-280, Springer, Berlin, Berlin, 2012.
- [8] I. Judson, R. Bulusu, B. Seddon, A. Dangoor, N. Wong, and S. Mudan, "UK clinical practice guidelines for the management of gastrointestinal stromal tumours (GIST)," *Clinical Sarcoma Research*, vol. 7, no. 1, article 6, 2017.
- [9] J. Lasota and M. Miettinen, "KIT and PDGFRA mutations in gastrointestinal stromal tumors (GISTs)," *Seminars in Diagnostic Pathology*, vol. 23, no. 2, pp. 91–102, 2006.
- [10] B. P. Rubin, GIST, EGIST, Enzinger and Weiss's Soft Tissue Tumors, Saunders, Philadelphia, PA, USA, 6th edition, 2013.
- [11] A. Ahmad, F. Mahmood, C. Shen, S. Cabezon, and V. Rao, "Malignant gastrointestinal stromal tumour – a case report," *Journal of Case Reports: Clinical & Medical*, vol. 1, no. 3, p. 125, 2018.
- [12] Y. Maeda, T. Shinohara, T. Katayama, A. Nagatsu, N. Futakawa, and T. Hamada, "Gastrointestinal stromal tumor of the stomach with an abscess excised by laparoscopic surgery," *Case Reports in Gastroenterology*, vol. 10, no. 2, pp. 399–405, 2016.
- [13] S. Yardimci, T. K. Uprak, F. E. Kombak, H. Kaya, and S. C. Yegen, "Ruptured gastric stromal tumour into gastric lumen with an abscess," *ANZ Journal of Surgery*, vol. 84, no. 9, pp. 687–689, 2014.
- [14] N. De la Fuente, M. Rodríguez Blanco, G. Cerdán, and V. Artigas, "Hemorragia digestiva aguda en paciente con neurofibromatosis tipo 1 afecto de múltiples GIST y ganglioneuromatosis intestinal," *Cirugía Española (English Edition)*, vol. 97, no. 4, pp. 237–239, 2019.

- [15] T. Nishida, M. Tsujimoto, T. Takahashi, S. Hirota, J. Y. Blay, and M. Wataya-Kaneda, "Gastrointestinal stromal tumors in Japanese patients with neurofibromatosis type I," *Journal of Gastroenterology*, vol. 51, no. 6, pp. 571–578, 2016.
- [16] Y. Hakozaki, S. Sameshima, T. Tatsuoka et al., "Rectal carcinoma and multiple gastrointestinal stromal tumors (GIST) of the small intestine in a patient with neurofibromatosis type 1: a case report," *World Journal of Surgical Oncology*, vol. 15, no. 1, article 160, 2017.
- [17] L. Al Momani, L. Crosnoe-Shipley, J. Phemister et al., "Recurrent GIST tumor in a patient with neurofibromatosis," *American Journal of Gastroenterology*, vol. 112, pp. \$1380-\$1381, 2017.
- [18] M. Miettinen and J. Lasota, "Histopathology of gastrointestinal stromal tumor," *Journal of Surgical Oncology*, vol. 104, no. 8, pp. 865–873, 2011.
- [19] P. F. Salvi, L. Lorenzon, S. Caterino, L. Antolino, M. S. Antonelli, and G. Balducci, "Gastrointestinal stromal tumors associated with neurofibromatosis 1: a single centre experience and systematic review of the literature including 252 cases," *International Journal of Surgical Oncology*, vol. 2013, Article ID 398570, 8 pages, 2013.
- [20] J. H. Wang, J. Lasota, and M. Miettinen, "Succinate dehydrogenase subunit B (SDHB) is expressed in neurofibromatosis 1-Associated gastrointestinal stromal tumors (GISTs): implications for the SDHB Expression Based classification of GISTs," *Journal of Cancer*, vol. 2, pp. 90–93, 2011.
- [21] A. W. Kirkpatrick, D. J. Roberts, J. De Waele et al., "Intraabdominal hypertension and the abdominal compartment syndrome: updated consensus definitions and clinical practice guidelines from the World Society of the Abdominal Compartment Syndrome," *Intensive Care Medicine*, vol. 39, no. 7, pp. 1190–1206, 2013.
- [22] M. L. Malbrain, "Aspects respiratoires du syndrome du compartiment abdominal," *Réanimation*, vol. 16, no. 1, pp. 49–60, 2007.
- [23] C. D. Fletcher, J. J. Berman, C. Corless et al., "Diagnosis of gastrointestinal stromal tumors: a consensus approach," *International Journal of Surgical Pathology*, vol. 10, no. 2, pp. 81–89, 2002.
- [24] A. D. Levy, H. E. Remotti, W. M. Thompson, L. H. Sobin, and M. Miettinen, "From the Archives of the AFIP," *Radio-graphics*, vol. 23, no. 2, pp. 283–304, 2003.
- [25] A. D. Levy, M. A. Manning, W. B. Al-Refaie, and M. M. Miettinen, "Soft-tissue sarcomas of the abdomen and pelvis: radiologic-pathologic features, part 1-common sarcomas: from the radiologic pathology archives," *Radiographics*, vol. 37, no. 2, pp. 462–483, 2017 Mar-Apr.
- [26] A. Koyuncuera, L. Gönlüs, and A. V. Kutsalb, "A rare case of giant gastrointestinal stromal tumor of the stomach involving the serosal surface," *International Journal of Surgery Case Reports*, vol. 12, pp. 90–94, 2015.
- [27] A. Cappellani, G. Piccolo, F. Cardì et al., "Giant gastrointestinal stromal tumor (GIST) of the stomach cause of high bowel obstruction: surgical management," *World Journal of Surgical Oncology*, vol. 11, no. 1, article 172, 2013.
- [28] A. El-Menyar, A. Mekkodathil, and H. Al-Thani, "Diagnosis and management of gastrointestinal stromal tumors: an upto-date literature review," *Journal of Cancer Research and Therapeutics*, vol. 13, no. 6, pp. 889–900, 2017.

- [29] C. Cauchi, J. C. Trent, K. Edwards et al., "An unusual site of metastasis from gastrointestinal stromal tumor," *Rare Tumors*, vol. 2, no. 4, article e58, p. 58, 2010.
- [30] U. Bashir, A. Qureshi, H. A. Khan, and N. Uddin, "Gastrointestinal stromal tumor with skeletal muscle, adrenal and cardiac metastases: an unusual occurrence," *Indian Journal of Pathology & Microbiology*, vol. 54, no. 2, pp. 362–364, 2011.
- [31] B. Hughes, D. Yip, D. Goldstein, P. Waring, V. Beshay, and G. Chong, "Cerebral relapse of metastatic gastrointestinal stromal tumor during treatment with imatinib mesylate: case report," *BMC Cancer*, vol. 4, no. 1, article 74, 2004.
- [32] M. Dorić, S. Radović, M. Babić et al., "Testicular metastasis of gastrointestinal stromal tumor of the jejunum," *Bosnian Journal of Basic Medical Sciences*, vol. 7, no. 2, pp. 176–179, 2007.
- [33] D. Siamkouris, M. Schloesser, A. Yousef, and E. Offers, "A rare case of gastrointestinal stromal tumor with a liver metastasis infiltrating the inferior vena cava and extending to the right atrium with an early recurrence after surgical extraction," *Case Reports in Cardiology*, vol. 2019, Article ID 2623403, 6 pages, 2019.
- [34] M. Vasundhara, G. Parvathi, and A. B. Lakshmi, "Our experience with gastrointestinal stromal tumors over a period of three years from a tertiary care centre," *International Journal* of Research in Medical Sciences, vol. 4, pp. 3709–3713, 2016.
- [35] M. E. Zoller, B. Rembeck, A. Oden, M. Samuelsson, and L. Angervall, "Malignant and benign tumors in patients with neurofibromatosis type 1 in a defined Swedish population," *Cancer*, vol. 79, no. 11, pp. 2125–2131, 1997.
- [36] J. Andersson, H. Sihto, J. M. Meis-Kindblom, H. Joensuu, N. Nupponen, and L. G. Kindblom, "NF1-associated gastrointestinal stromal tumors have unique clinical, phenotypic, and genotypic characteristics," *The American Journal of Surgical Pathology*, vol. 29, no. 9, pp. 1170–1176, 2005.
- [37] M. Kiuru and K. J. Busam, "The NF1 gene in tumor syndromes and melanoma," *Laboratory Investigation*, vol. 97, no. 2, pp. 146–157, 2017.
- [38] R. E. Ferner, S. M. Huson, N. Thomas et al., "Guidelines for the diagnosis and management of individuals with neurofibromatosis 1," *Journal of Medical Genetics*, vol. 44, no. 2, pp. 81–88, 2007.
- [39] Y. Hirata, H. Brems, M. Suzuki et al., "Interaction between a domain of the negative regulator of the Ras-ERK pathway, SPRED1 protein, and the GTPase-activating protein-related domain of neurofibromin is implicated in Legius syndrome and neurofibromatosis type 1," *The Journal of Biological Chemistry*, vol. 291, no. 7, pp. 3124–3134, 2016.
- [40] S. Cheng, M. J. Huang, T. L. Yang et al., "Neurofibromatosis with gastrointestinal stromal tumors: insights into the association," *Digestive Diseases and Sciences*, vol. 49, no. 7-8, pp. 1165–1169, 2004.
- [41] C. E. Fuller and G. T. Williams, "Gastrointestinal manifestations of type 1 neurofibromatosis (von Recklinghausen's disease)," *Histopathology*, vol. 19, no. 1, pp. 1–12, 1991.
- [42] A. Agaimy, N. Vassos, and R. S. Croner, "Gastrointestinal manifestations of neurofibromatosis type 1 (Recklinghausen's disease): clinicopathological spectrum with pathogenetic considerations," *International Journal of Clinical and Experimental Pathology*, vol. 5, no. 9, pp. 852–862, 2012.
- [43] R. Ricci, "Syndromic gastrointestinal stromal tumors," *Hered-itary Cancer in Clinical Practice*, vol. 14, no. 1, article 15, 2016.

- [44] C. A. Stratakis and J. A. Carney, "The triad of paragangliomas, gastric stromal tumours and pulmonary chondromas (Carney triad), and the dyad of paragangliomas and gastric stromal sarcomas (Carney Stratakis syndrome): molecular genetics and clinical implications," *Journal of Internal Medicine*, vol. 266, no. 1, pp. 43–52, 2009.
- [45] A. J. Gill, A. Chou, R. Vilain et al., "Immunohistochemistry for SDHB divides gastrointestinal stromal tumors (GISTs) into 2 distinct types," *American Journal of Surgical Pathology*, vol. 34, no. 5, pp. 636–644, 2010.
- [46] J. Gaal, C. A. Stratakis, J. A. Carney et al., "SDHB immunohistochemistry: a useful tool in the diagnosis of Carney-Stratakis and Carney triad gastrointestinal stromal tumors," *Modern Pathology*, vol. 24, no. 1, pp. 147–151, 2011.
- [47] B. Pasini, S. R. McWhinney, T. Bei et al., "Clinical and molecular genetics of patients with the Carney-Stratakis syndrome and germline mutations of the genes coding for the succinate dehydrogenase subunits SDHB, SDHC, and SDHD," European Journal of Human Genetics, vol. 16, no. 1, pp. 79–88, 2008.
- [48] P. J. Oppelt, A. C. Hirbe, and B. A. Van Tine, "Gastrointestinal stromal tumors (GISTs): point mutations matter in management, a review," *Journal of Gastrointestinal Oncology*, vol. 8, no. 3, pp. 466–473, 2017.
- [49] C. Mussi, H. U. Schildhaus, A. Gronchi, E. Wardelmann, and P. Hohenberger, "Therapeutic consequences from molecular biology for gastrointestinal stromal tumor patients affected by neurofibromatosis type 1," *Clinical Cancer Research*, vol. 14, no. 14, pp. 4550–4555, 2008.
- [50] J.-L. Lee, J. Y. Kim, M.-H. Ryu et al., "Response to imatinib in KIT- and PDGFRA-wild type gastrointestinal stromal associated with neurofibromatosis type 1," *Digestive Diseases and Sciences*, vol. 51, no. 6, pp. 1043–1046, 2006.
- [51] W. J. Jessen, S. J. Miller, E. Jousma et al., "MEK inhibition exhibits efficacy in human and mouse neurofibromatosis tumors," *The Journal of Clinical Investigation*, vol. 123, no. 1, pp. 340–347, 2013.
- [52] M. C. Heinrich, R. G. Maki, C. L. Corless et al., "Primary and secondary kinase genotypes correlate with the biological and clinical activity of sunitinib in imatinib resistant gastrointestinal stromal tumor," *Journal of Clinical Oncology*, vol. 26, no. 33, pp. 5352–5359, 2008.