





Genome Sequences of Two Bacillus Phages Isolated from Indonesia

Yoga A. Handoko, a,b Agustin K. Wardani, a Aji Sutrisno, a Simon B. Widjanarko, a Trever L. Thurgood, C Daniel W. Thompson, C Ruchira Sharma, Dulianne H. Grose

ABSTRACT Here, the genome sequences of two soil bacteriophages isolated from a red chili plantation in Indonesia are presented. The genome of vB_BspS_SplendidRed (42,859 bp) is highly similar to Bacillus phage Ray17 from the United States, while vB_BspM_MarvelLand (156,945 bp) is highly similar to Bacillus phage BC01 from South Korea.

acillus spp. are rod-shaped, Gram-positive, aerobic bacteria capable of forming endospores when exposed to extreme environments (1–3). In the agroindustry, some species of Bacillus, such as B. subtilis, B. thuringiensis, B. licheniformis, B. pumilus, B. anthracis, and B. cereus, can contaminate raw material and food products (4, 5). Bacillus spp. can encode saprophytic enzymes such as α -amylases and proteases, which can damage bread products, pasta, cocoa, milk, meat products, and chili-based products, including chili bo (6-12). Bacteriophages (phages) are viruses that can infect and kill bacteria (4, 13), making them prime candidates for treating bacterial diseases, including Bacillus-related disease (14).

Herein, we announce the genome sequences of two novel Bacillus phages, vB_BspS_SplendidRed (SplendidRed) and vB_BspM_MarvelLand (MarvelLand). These phages were isolated from soil collected from a red chili plantation in Getasan Village, Semarang District, Central Java State, Indonesia (global positioning system coordinates 7°23′09.9"S, 110°27′35.1"E). The Bacillus sp. environmental host was isolated from decayed red chili fruit (Capsicum annuum L.) by plating onto lysis broth (LB) plates, and it was identified as a Bacillus sp. by using 16S rRNA sequencing (with primers described by Liu et al. [15]). The phages were isolated by propagation in an enrichment culture, followed by plaque purification from top agar (adapted from Czajkowski et al. [16] and Sauder et al. [17]).

Phage genomic DNA was extracted using the Norgen Biotek phage DNA isolation kit. The Illumina TruSeq Nano DNA kit was used for genomic library preparation, with each phage having unique barcodes, followed by sequencing on the Illumina HiSeq 2500 platform (250-bp paired-end reads). Geneious version 8.0.5 (18) reported one circular contig for each phage from the de novo assembly algorithm, using 20% of the 1,275,056 reads for MarvelLand (with no preassembly quality checking) and 90% of the 1,220,418 reads for SplendidRed (with the ends trimmed preassembly); the percentage of reads used was determined empirically to achieve a circular contig. The Geneiousbased assembly of MarvelLand resulted in two ambiguities (base pairs 31070 and 31095) that had differential base calling. These ambiguities were resolved by PCR amplification of the region (base pairs 30985 to 31355), followed by Sanger sequencing, which confirmed the most prominent base pairs identified by Illumina sequencing. Phages were annotated using DNA Master (19), giving preference for calls that gave full

Citation Handoko YA, Wardani AK, Sutrisno A, Widjanarko SB, Thurgood TL, Thompson DW, Sharma R, Grose JH. 2019. Genome sequences of two Bacillus phages isolated from Indonesia. Microbiol Resour Announc 8:e01058-19. https://doi.org/10.1128/MRA.01058-19.

Editor J. Cameron Thrash, University of Southern California

Copyright © 2019 Handoko et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Julianne H. Grose, julianne grose@byu.edu.

Received 28 August 2019 Accepted 8 November 2019 Published 12 December 2019

^aDepartment of Agroindustry Technology, Brawijaya University, Malang, East Java, Indonesia

^bDepartment of Agrotechnology, Satya Wacana Christian University, Salatiga, Central Java, Indonesia

^cDepartment of Microbiology and Molecular Biology, Brigham Young University, Provo, Utah, USA



TABLE 1 Basic properties of the two Bacillus phage genomes

	GenBank		Min-max ^a fold coverage	Genome	No. of	No. of	G+C
Phage name	accession no.	SRA accession no.	(avg read depth)	length (bp)	ORFs	tRNAs	content (%)
vB_BspS_SplendidRed	MN013088	SAMN12262426	338-736 (517.2)	42,859	76	NA^b	44.58
vB_BspM_MarvelLand	MN013089	SAMN12262427	1–1,882 (398.9)	156,945	241	18	40.19

^a Min-max, minimum to maximum.

coding potential coverage (20, 21). Default parameters were used for all software unless otherwise stated.

Analysis of the phage genome sequences suggests that SplendidRed (42,859 bp) is a new member of the *Caudovirales Siphoviridae* family, having 93% nucleotide similarity by BLASTN (22) to *Bacillus* phage Ray17, isolated in the United States (23). MarvelLand (156,945 bp) is likely a member of the *Caudovirales Myoviridae* family, having 94% nucleotide similarity by BLASTN to *Bacillus* phage BC01, isolated in South Korea (24). The SplendidRed and MarvelLand genome termini were determined by BLASTN alignment to Ray17 and BC01, respectively. The current annotation suggests that SplendidRed encodes at least 76 open reading frames (ORFs), while MarvelLand encodes 241 ORFs and 18 tRNAs. These phages encode proteins with predicted functions in DNA metabolism, virion structure, packaging, and cell lysis.

Data availability. The GenBank and SRA accession numbers for the *Bacillus* phages SplendidRed and MarvelLand are listed in Table 1.

ACKNOWLEDGMENTS

We thank the Brigham Young University (BYU) Microscopy Lab for phage imaging and the BYU DNA Sequencing Center for phage genome sequencing.

We are grateful to the Department of Microbiology and Molecular Biology (MMBIO), BYU, for research funding. Y.A.H. thanks the Ministry of Research, Technology, and Higher Education, Indonesia, for providing the Sandwich scholarship program. Y.A.H. also thanks Brawijaya University and Satya Wacana Christian University for the strong support enabling this research to be conducted at the Grose Lab, MMBIO Department, BYU.

REFERENCES

- Nicholson WL, Munakata N, Horneck G, Melosh HJ, Setlow P. 2000. Resistance of *Bacillus* endospores to extreme terrestrial and extraterrestrial environments. Microbiol Mol Biol Rev 64:548–572. https://doi.org/10.1128/mmbr.64.3.548-572.2000.
- Setlow P. 2006. Spores of *Bacillus subtilis*: their resistance to and killing by radiation, heat, and chemicals. J Appl Microbiol 101:514–525. https:// doi.org/10.1111/j.1365-2672.2005.02736.x.
- Wood JP, Meyer KM, Kelly TJ, Choi YW, Rogers JV, Riggs KB, Willenberg ZJ. 2015. Environmental persistence of *Bacillus anthracis* and *Bacillus subtilis* spores. PLoS One 10:e0138083. https://doi.org/10.1371/journal.pone.0138083.
- Krasowska A, Biegalska A, Augustyniak D, Łoś M, Richert M, Łukaszewicz M. 2015. Isolation and characterization of phages infecting *Bacillus subtilis*. BioMed Res Int 2015:179597. https://doi.org/10.1155/2015/179597.
- Gillis A, Mahillon J. 2014. Phages preying on *Bacillus anthracis, Bacillus cereus*, and *Bacillus thuringiensis*: past, present and future. Viruses 6:2623–2672. https://doi.org/10.3390/v6072623.
- Ochiai A, Itoh T, Kawamata A, Hashimoto W, Murata K. 2007. Plant cell wall degradation by saprophytic *Bacillus subtilis* strains: gene clusters responsible for rhamnogalacturonan depolymerization. Appl Environ Microbiol 73:3803–3813. https://doi.org/10.1128/AEM.00147-07.
- Collins NE, Kirschner LAM, von Holy A. 1991. Characterization of *Bacillus* isolates from ropey bread, bakery equipment and raw materials. S Afr J Sci 87:62–66.
- Bailey CP, von Holy A. 1993. Bacillus spore contamination associated with commercial bread manufacture. Food Microbiol 10:287–294. https://doi.org/10.1006/fmic.1993.1033.

- Sorokulova IB, Reva ON, Smirnov VV, Pinchuk IV, Lapa SV, Urdaci MC. 2003. Genetic diversity and involvement in bread spoilage of *Bacillus* strains isolated from flour and ropy bread. Lett Appl Microbiol 37: 169–173. https://doi.org/10.1046/j.1472-765x.2003.01372.x.
- Te Giffel MC, Beumer RR, Leijendekkers S, Rombouts FM. 1996. Incidence of *Bacillus cereus* and *Bacillus subtilis* in foods in the Netherlands. Food Microbiol 13:53–58. https://doi.org/10.1006/fmic.1996.0007.
- Leisner JJ, Rusul G, Wee BW, Boo HC, Muhammad K. 1997. Microbiology of chili bo, a popular Malaysian food ingredient. J Food Prot 60:1235–1240. https://doi.org/10.4315/0362-028X-60.10.1235.
- Zaini NAM, Harith HH, Olusesan AT, Zulkifli AH, Bakar FA, Osman A, Hamid AA, Saari N. 2010. Level of chemical and microbiological contaminations in chili bo (paste). J Food Prot 73:541–546. https://doi.org/10 .4315/0362-028x-73.3.541.
- 13. Brovko LY, Anany H, Griffiths MW. 2012. Bacteriophages for detection and control of bacterial pathogens in food and food-processing environment. Adv Food Nutr Res 67:241–288. https://doi.org/10.1016/B978 -0-12-394598-3.00006-X.
- Grose JH, Jensen GL, Burnett SH, Breakwell DP. 2014. Erratum to: genomic comparison of 93 *Bacillus* phages reveals 12 clusters, 14 singletons and remarkable diversity. BMC Genomics 15:1184. https://doi.org/10.1186/1471 -2164-15-1184.
- Liu Y, Han JX, Huang HY, Zhu B. 2005. Development and evaluation of 16S rDNA microarray for detecting bacterial pathogens in cerebrospinal fluid. Exp Biol Med (Maywood) 230:587–591. https://doi.org/10.1177/ 153537020523000810.
- 16. Czajkowski R, Ozymko Z, Lojkowska E. 2014. Isolation and characterization

Volume 8 Issue 50 e01058-19 mra.asm.org **2**

^b NA, no tRNAs were identified.



- of novel soilborne lytic bacteriophages infecting *Dickeya* spp. biovar 3 ("D. solani"). Plant Pathol 63:758–772. https://doi.org/10.1111/ppa.12157.
- Sauder AB, Quinn MR, Brouillette A, Caruso S, Cresawn S, Erill I, Lewis L, Loesser-Casey K, Pate M, Scott C, Stockwell S, Temple L. 2016. Genomic characterization and comparison of seven *Myoviridae* bacteriophage infecting *Bacillus thuringiensis*. Virology 489:243–251. https://doi.org/10 .1016/j.virol.2015.12.012.
- Kearse M, Moir R, Wilson A, Stones-Havas S, Cheung M, Sturrock S, Buxton S, Cooper A, Markowitz S, Duran C, Thierer T, Ashton B, Meintjes P, Drummond A. 2012. Geneious Basic: an integrated and extendable desktop software platform for the organization and analysis of sequence data. Bioinformatics 28:1647–1649. https://doi.org/10.1093/bioinformatics/bts199.
- Lawrence J. 2012. DNA Master, v5.0.2. http://cobamide2.bio.pitt.edu/ computer.htm.
- Grose JH, Jensen JD, Merrill BD, Fisher JNB, Burnett SH, Breakwell DP. 2014.
 Genome sequences of three novel *Bacillus cereus* bacteriophages. Genome Announc 2:e01118-13. https://doi.org/10.1128/genomeA.01118-13.
- Sharma R, Berg JA, Beatty NJ, Choi MC, Cowger AE, Cozzens BJR, Duncan SG, Fajardo CP, Ferguson HP, Galbraith T, Herring JA, Hoj TR, Durrant JL, Hyde JR, Jensen GL, Ke SY, Killpack S, Kruger JL, Lawrence EEK, Nwosu IO, Tam TC, Thompson DW, Tueller JA, Ward MEH, Webb CJ, Wood ME, Yeates EL, Baltrus DA, Breakwell DP, Hope S, Grose JH. 2018. Genome sequences of nine *Erwinia amylovora* bacteriophages. Microbiol Resour Announc 7:e00944-18. https://doi.org/10.1128/MRA.00944-18.
- Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. 1990. Basic local alignment search tool. J Mol Biol 215:403–410. https://doi.org/10.1016/ S0022-2836(05)80360-2.
- 23. Showalter R, Adat I, Raab R, Temple L. 2019. Complete genome sequence of *Bacillus amyloliquefaciens* bacteriophage Ray17. Microbiol Resour Announc 8:e00134-19. https://doi.org/10.1128/MRA.00134-19.
- Kim J, Kim G-H, Lee N-G, Lee J-S, Yoon S-S. 2018. Whole-genome sequencing and genomic analysis of a virulent bacteriophage infecting *Bacillus cereus*. Intervirology 61:272–280. https://doi.org/10.1159/000499068.

Volume 8 Issue 50 e01058-19 mra.asm.org **3**