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International Journal of Medical Informatics

journal homepage: www.elsevier.com/locate/ijmedinf



Predicting prognosis in COVID-19 patients using machine learning and readily available clinical data

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ARTICLE INFO	A B S T R A C T
Keywords: Machine learning COVID-19 Prognostic models Risk assessment Clinical decision support systems	Rationale: Prognostic tools for aiding in the treatment of hospitalized COVID-19 patients could help improve outcome by identifying patients at higher or lower risk of severe disease. The study objective was to develop models to stratify patients by risk of severe outcomes during COVID-19 hospitalization using readily available information at hospital admission. <i>Methods</i> : Hierarchical ensemble classification models were trained on a set of 229 patients hospitalized with COVID-19 to predict severe outcomes, including ICU admission, development of acute respiratory distress syn- drome, or intubation, using easily attainable attributes including basic patient characteristics, vital signs at admission, and basic lab results collected at time of presentation. Each test stratifies patients into groups of increasing risk. An additional cohort of 330 patients was used for blinded, independent validation. Shapley value analysis evaluated which attributes contributed most to the models' predictions of risk. <i>Main results</i> : Test performance was assessed using precision (positive predictive value) and recall (sensitivity) of the final risk groups. All test cut-offs were fixed prior to blinded validation. In development and validation, the tests achieved precision in the lowest risk groups near or above 0.9. The proportion of patients with severe outcomes significantly increased across increasing risk groups. While the importance of attributes varied by test and patient, C-reactive protein, lactate dehydrogenase, and D-dimer were often found to be important in the assignment of risk.
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Conclusions: Risk of severe outcomes for patients hospitalized with COVID-19 infection can be assessed using machine learning-based models based on attributes routinely collected at hospital admission.

1. Introduction

The coronavirus disease (COVID-19) pandemic continues to place world-wide health systems under pressure. The ability to identify patients at greatest risk of developing severe disease would help inform discussions of the risks and benefits of treatments [1–3], target higher risk populations for clinical trials of therapeutic agents, compare outcomes between patients with similar risk, and prioritize limited resources.

Multiple prognostic factors have been identified, including age, certain comorbidities [4–6], neutrophil-to-lymphocyte ratio (NLR), D-

dimer and C-reactive protein (CRP) [4,7–8]. However, the relative influence of these factors on prognosis has been difficult to elucidate.

While traditional statistical modelling has been used to combine multiple attributes to predict outcomes for patients with COVID-19 [9–10], modern machine learning (ML) has added advantage in its capability to discover more complex interactions between correlated attributes. Such methods have shown promise for predicting acute respiratory distress syndrome (ARDS) in diseases other than COVID-19 [11–12] and demonstrated potential utility for predicting poor outcomes in COVID-19 [13–29].

Many patients hospitalized with COVID-19 provide limited medical

Received 11 March 2021; Received in revised form 30 August 2021; Accepted 21 September 2021 Available online 23 September 2021

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https://doi.org/10.1016/j.ijmedinf.2021.104594

history due to not engaging in routine care, not having easily accessible comorbidities data, or being intubated shortly after arrival. Hence, the goal of this study was to create and validate tests to risk stratify COVID-19 patients for progression to severe disease using only readily available clinical, demographic, and laboratory attributes collected at the point of hospital admission. Given the nature of these attributes, the tests stratifying risk of admission to the intensive care unit (ICU), intubation, or progression to ARDS could be validated, blinded to outcome, on an independent patient cohort using data drawn from electronic health records (EHRs).

2. Methods

2.1. Data extraction and patient cohorts

For test development, data was collected for all patients 18 years or older admitted to the University of Colorado Health (UCH) hospital in Aurora, CO with a positive COVID-19 nasopharyngeal polymerase chain reaction test between mid-March and mid-May 2020. Attributes deemed potentially useful at the time were extracted from the EHR by medical students and stored in a REDCap database [30]. Patient data was restricted to first recorded admission and observations for laboratory values and vital measures. ARDS occurrence was defined by any mention of ARDS in the critical care progress notes.

The deidentified dataset was transferred to Biodesix for test development. A set of 26 attributes (indicated with * in Table 1) to be used in classifier training was selected, limited to attributes routinely collected at hospital admission and available in the EHR, where inclusion of an attribute did not reduce the size of the cohort with complete data by more than 10%. The development cohort included patients with complete information for the selected attributes.

An independent validation cohort was derived from data provided by UCH's EHR data warehouse service for blinded validation of the tests. The validation cohort included all patients aged between 18 and 90 years, hospitalized in any UCH system facility with a positive COVID-19 test or diagnosis, with first admission between mid-May 2020 and mid-September 2020 with complete data on the 26 required attributes (Fig. 1). Deidentified data for the validation cohort were transferred to Biodesix for blinded test classification generation. Outcome data for test performance evaluation for the validation cohort was only shared after test classifications were returned to UCH investigators. ARDS occurrence was identified by SNOMED code. More details on data curation and validation are provided in the supplement. The study was reviewed and approved by the Colorado Multiple Institutional Review Board.

2.2. Test development

A series of classifiers were trained to classify patients as higher or lower risk for each endpoint (ICU admission, intubation, ARDS) using the same set of 26 attributes that were all used in all final tests to predict risk of outcome. These classifiers were arranged in a hierarchy shown in Fig. 2 to give a final classification of up to four possible risk groups. The fundamental model behind each component classifier was the Diagnostic Cortex® platform, a strongly dropout-regularized, feature abstracted, ensemble logistic regression [31]. The classifier at the top of each test's hierarchy was augmented with an additional decision tree model. Hyperparameters for the models were picked on heuristic considerations as in [31] and are given in Table E2, and further optimization was not performed to avoid overfitting. Bootstrap validation results in the development cohort were obtained using out-of-bag estimates [32]. ML methods are described fully in the supplement, and source code demonstrating their application here is provided at [33].

Relative attribute importance for individual patient classification was evaluated using Shapley Values [34–35] for 50 patients in the validation cohort chosen to span different test classification groups and is described in detail in the supplement.

Table 1

Patient characteristics for	the development and	l independent validation cohorts.

		Development Cohort (N = 229)	Validation Cohort (N = 330)
Categorical Attribute	Class	n (%)	n (%)
Race*	White	41 (17.9)	113 (34.2)
	Black/African	52 (22.7)	23 (7.0)
	American		
	Hispanic/Latino	94 (41.0)	164 (49.7)
	Other	31 (13.5)	25 (7.6)
	Unknown	11 (4.8)	5 (1.5)
Sex*	Male	124 (54.1)	172 (52.1)
•CED*	Female	105 (45.9)	158 (47.9)
eGFR*	≥60 mL/min/ 1.73 m ²	180 (78.6)	244 (73.9)
	$30 \ge but < 60$	34 (14.8)	61 (18.5)
	mL/min/1.73 m ²		()
	<30 mL/min/	15 (6.6)	25 (7.6)
	1.73 m^2		
Hypertension	Yes	100 (43.7)	NA
	No	126 (56.3)	NA
Diabetes	Yes	82 (35.8)	NA
	No	147 (64.2)	NA
Continuous Attribut	e	Median (Q1-Q3)	Median (Q1-
			Q3)
BMI ^{**} , kg/m ²		30 (27–36)	
Age*, years		57 (43–68)	57 (44–70)
Temperature*, °C		37 (37–38)	37 (37–38)
Heart Rate*, beats/ minute		98 (84–110)	98 (85–110)
Systolic*, mm Hg		130 (120–150)	130 (120–140)
Diastolic BP*, mm Hg		74 (65–83)	74 (65–84)
Respiratory Rate*, breaths/minute		20 (18–24)	20 (18–24)
Oxygen Saturation			
		92 (87–94)	92 (88–95)
, % Weight, kg		92 (87–94) 82 (72–100)	92 (88–95) 85 (73–100)
*, %			
, % Weight, kg		82 (72–100)	85 (73–100)
, % Weight, kg QTc*,		82 (72–100) 440 (420–460)	85 (73–100) 440 (420–470)
,% Weight, kg QTc*, Sodium*, mmol/L Potassium*, mmol/		82 (72–100) 440 (420–460) 140 (130–140)	85 (73–100) 440 (420–470) 140 (130–140)
, % Weight, kg QTc*, Sodium*, mmol/L Potassium*, mmol/ L Carbon dioxide*		82 (72–100) 440 (420–460) 140 (130–140) 3.8 (3.4–4.0)	85 (73–100) 440 (420–470) 140 (130–140) 3.9 (3.6–4.2)
, % Weight, kg QTc*, Sodium*, mmol/L Potassium*, mmol/L Carbon dioxide* mmol/L BUN* mg/dL Creatinine* mg/dL		82 (72–100) 440 (420–460) 140 (130–140) 3.8 (3.4–4.0) 23 (21–25)	85 (73–100) 440 (420–470) 140 (130–140) 3.9 (3.6–4.2) 23 (21–25) 15 (10–22) 0.87 (0.72–1.2)
, % Weight, kg QTc*, Sodium*, mmol/L Potassium*, mmol/L Carbon dioxide* mmol/L BUN* mg/dL Creatinine* mg/dL Anion Gap*, mmol/ L		82 (72–100) 440 (420–460) 140 (130–140) 3.8 (3.4–4.0) 23 (21–25) 13 (10–20)	85 (73–100) 440 (420–470) 140 (130–140) 3.9 (3.6–4.2) 23 (21–25) 15 (10–22) 0.87 (0.72–1.2) 11 (9–13)
, % Weight, kg QTc*, Sodium*, mmol/L Potassium*, mmol/L Carbon dioxide* mmol/L BUN* mg/dL Creatinine* mg/dL Anion Gap*, mmol/		82 (72–100) 440 (420–460) 140 (130–140) 3.8 (3.4–4.0) 23 (21–25) 13 (10–20) 0.94 (0.69–1.2)	85 (73–100) 440 (420–470) 140 (130–140) 3.9 (3.6–4.2) 23 (21–25) 15 (10–22) 0.87 (0.72–1.2)
, % Weight, kg QTc*, Sodium*, mmol/L Potassium*, mmol/L Carbon dioxide* mmol/L BUN* mg/dL Creatinine* mg/dL Anion Gap*, mmol/ L WBC Count* ×10 ⁹ cells/L Hemoglobin* g/dL		82 (72–100) 440 (420–460) 140 (130–140) 3.8 (3.4–4.0) 23 (21–25) 13 (10–20) 0.94 (0.69–1.2) 12 (10–13)	85 (73–100) 440 (420–470) 140 (130–140) 3.9 (3.6–4.2) 23 (21–25) 15 (10–22) 0.87 (0.72–1.2) 11 (9–13)
, % Weight, kg QTc*, Sodium*, mmol/L Potassium*, mmol/L Carbon dioxide* mmol/L BUN* mg/dL Creatinine* mg/dL Anion Gap*, mmol/ L WBC Count* ×10 ⁹ cells/L Hemoglobin* g/dL Hematocrit*, %		82 (72–100) 440 (420–460) 140 (130–140) 3.8 (3.4–4.0) 23 (21–25) 13 (10–20) 0.94 (0.69–1.2) 12 (10–13) 6.8 (5.3–8.9) 15 (13–16) 44 (40–47)	85 (73–100) 440 (420–470) 140 (130–140) 3.9 (3.6–4.2) 23 (21–25) 15 (10–22) 0.87 (0.72–1.2) 11 (9–13) 7 (5.5–9.1) 14 (13–15) 42 (38–46)
, % Weight, kg QTc*, Sodium*, mmol/L Potassium*, mmol/L Carbon dioxide* mmol/L BUN* mg/dL Creatinine* mg/dL Anion Gap*, mmol/ L WBC Count* ×10 ⁹ cells/L Hemoglobin* g/dL		82 (72–100) 440 (420–460) 140 (130–140) 3.8 (3.4–4.0) 23 (21–25) 13 (10–20) 0.94 (0.69–1.2) 12 (10–13) 6.8 (5.3–8.9) 15 (13–16)	85 (73–100) 440 (420–470) 140 (130–140) 3.9 (3.6–4.2) 23 (21–25) 15 (10–22) 0.87 (0.72–1.2) 11 (9–13) 7 (5.5–9.1) 14 (13–15)
, % Weight, kg QTc*, Sodium*, mmol/L Potassium*, mmol/L Carbon dioxide* mmol/L BUN* mg/dL Creatinine* mg/dL Anion Gap*, mmol/ L WBC Count* ×10 ⁹ cells/L Hemoglobin* g/dL Hematocrit*, % Platelet Count*		82 (72–100) 440 (420–460) 140 (130–140) 3.8 (3.4–4.0) 23 (21–25) 13 (10–20) 0.94 (0.69–1.2) 12 (10–13) 6.8 (5.3–8.9) 15 (13–16) 44 (40–47)	85 (73–100) 440 (420–470) 140 (130–140) 3.9 (3.6–4.2) 23 (21–25) 15 (10–22) 0.87 (0.72–1.2) 11 (9–13) 7 (5.5–9.1) 14 (13–15) 42 (38–46)
, % Weight, kg QTc*, Sodium*, mmol/L Potassium*, mmol/L Carbon dioxide* mmol/L BUN* mg/dL Creatinine* mg/dL Anion Gap*, mmol/ L WBC Count* ×10 ⁹ cells/L Hemoglobin* g/dL Hematocrit*, % Platelet Count* ×10 ⁹ cells/L LDH* U/L D-Dimer* ng/mL		$\begin{array}{c} 82 \ (72-100) \\ 440 \ (420-460) \\ 140 \ (130-140) \\ 3.8 \ (3.4-4.0) \\ 23 \ (21-25) \\ 13 \ (10-20) \\ 0.94 \ (0.69-1.2) \\ 12 \ (10-13) \\ 6.8 \ (5.3-8.9) \\ 15 \ (13-16) \\ 44 \ (40-47) \\ 210 \ (160-260) \\ 320 \ (260-420) \\ 860 \ (530-1500) \end{array}$	85 (73–100) 440 (420–470) 140 (130–140) 3.9 (3.6–4.2) 23 (21–25) 15 (10–22) 0.87 (0.72–1.2) 11 (9–13) 7 (5.5–9.1) 14 (13–15) 42 (38–46) 200 (160–260) 450 (290–750) 740 (410–1500)
, % Weight, kg QTc*, Sodium*, mmol/L Potassium*, mmol/L Carbon dioxide* mmol/L BUN* mg/dL Creatinine* mg/dL Anion Gap*, mmol/ L WBC Count* ×10 ⁹ cells/L Hemoglobin* g/dL Hematocrit*, % Platelet Count* ×10 ⁹ cells/L LDH* U/L		$\begin{array}{c} 82 \ (72-100) \\ 440 \ (420-460) \\ 140 \ (130-140) \\ 3.8 \ (3.4-4.0) \\ 23 \ (21-25) \\ 13 \ (10-20) \\ 0.94 \ (0.69-1.2) \\ 12 \ (10-13) \\ 6.8 \ (5.3-8.9) \\ 15 \ (13-16) \\ 44 \ (40-47) \\ 210 \ (160-260) \\ 320 \ (260-420) \end{array}$	85 (73–100) 440 (420–470) 140 (130–140) 3.9 (3.6–4.2) 23 (21–25) 15 (10–22) 0.87 (0.72–1.2) 11 (9–13) 7 (5.5–9.1) 14 (13–15) 42 (38–46) 200 (160–260) 450 (290–750)

Definition of abbreviations: eGFR = estimated glomerular filtration rate; BP = blood pressure; QTc = corrected QT interval; BUN = blood urea nitrogen; $CO_2 = carbon$ dioxide; WBC = white blood cell; LDH = lactate dehydrogenase; CRP = C-reactive protein; BMI = body mass index.

* Used for classification.

Only complete for 205 out 229 patients.

2.3. Statistical methods

The validation cohort was analyzed following a prespecified statistical analysis plan, included in the supplement, using SAS Enterprise Guide 8.2 (SAS 9.4) (SAS Institute, Cary, NC). Performance is presented in terms of positive-predictive-value (PPV), specificity, and F1 scores

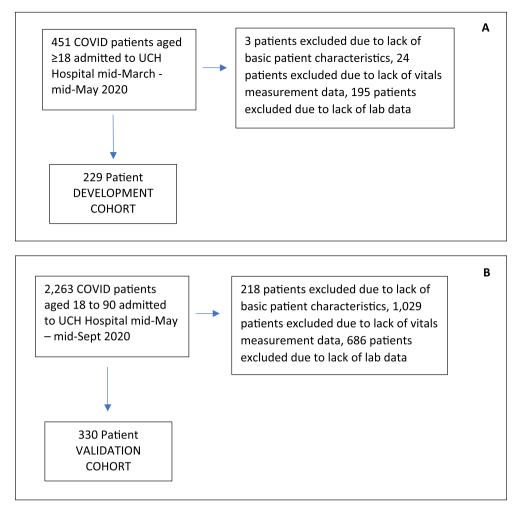


Fig. 1. Consort diagrams of patient selection down to development (A) and validation (B) cohorts.

[36–38]. Cochran-Armitage test and Fisher's exact test were used to assess trends and differences in proportions. Confidence intervals (CIs) for proportions were calculated using the Clopper-Pearson or bootstrap methods.

3. Results

Patients included in the development cohort were generally similar to those excluded due to lack of complete data, although rate of intubation and systolic blood pressure were slightly higher and oxygen saturation and D-dimer level lower. Patients included in the validation cohort exhibited higher rates of severe outcomes than patients excluded due to lack of complete data. The included and excluded cohorts are compared in the supplement.

Two-hundred twenty-nine patients were included in the development cohort: 77 (34%) were admitted to the ICU, 53 (23%) were intubated, and 45 (20%) developed ARDS. The proportions of patients experiencing poor outcomes were smaller in the 330 patients in the validation cohort, of whom 85 (26%) were admitted to the ICU, 42 (13%) were intubated, and 35 (11%) developed ARDS. The patient characteristics of the two cohorts are summarized in Table 1. The racial/ ethnic composure of both cohorts were skewed compared with overall Colorado demographics [39] with White being significantly underrepresented in both sets, and Black being overrepresented only in the development cohort. All attributes used in classification are contained in Table 1 and denoted with an asterisk. The time from data collection to ICU admission was estimated in the validation cohort and is summarized in Fig. 3.

3.1. Predicting ICU admission

Test development for prediction of ICU admission used the schema in Fig. 2B, with the flow of patients through the hierarchical classification scheme shown in Fig. 3. In Fig. 4, we see the test classified 73 (32%), 54 (24%), 51 (22%), and 51 (22%) of the development cohort to the lowest, low, high, and highest risk groups, respectively. Only 12% of patients in the lowest risk group were admitted to the ICU, whereas 65% of the highest risk group were admitted. Performance was similar in the validation cohort, of which 28%, 22%, 27%, and 22% were classified to the lowest, low, high, and highest risk groups, respectively. The proportions of patients that were admitted to the ICU were significantly associated with increasing risk subgroup (Cochran-Armitage p < 0.0001). The proportion of patients admitted to the ICU in the lowest risk group was 11% (95% CI: 5–19%), significantly lower than that in the other groups (Fisher's exact p < 0.0001). The proportion of patients admitted to the ICU in the highest risk group was 51% (95% CI: 39-63%), Fisher's exact p (highest risk vs other) < 0.0001).

Test classification was associated with heart rate, respiratory rate, oxygen saturation, sodium, blood urea nitrogen (BUN), anion gap, white blood cell (WBC) count, lactate dehydrogenase (LDH), D-dimer, CRP, and ferritin. Shapley Value analysis showed that the attributes most important for classification generation varied by patient, but that lowest risk classification was most often explained by CRP and D-dimer, with ferritin, LDH, platelet count, heart rate, respiratory rate, oxygen saturation, and WBC also of relatively higher importance. In highest risk patients, classification was related to LDH and D-dimer, although oxygen saturation, respiratory rate, CRP, ferritin, and weight were also

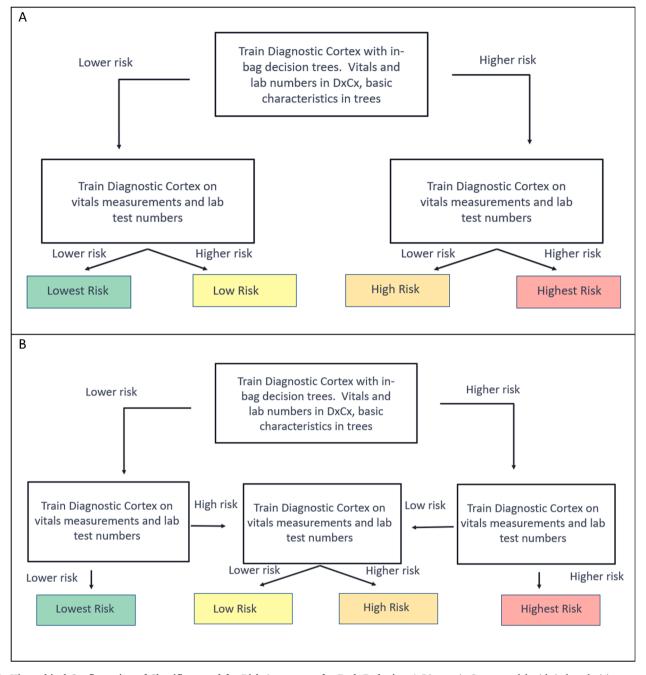


Fig. 2. Hierarchical Configuration of Classifiers used for Risk Assessment for Each Endpoint. A Diagnostic Cortex model with in-bag decision tree model (represented by the top box) was used to stratify the entire development cohort into a higher and lower risk group for each endpoint. Diagnostic Cortex models (middle boxes) without trees were used to split the resulting two groups further according to one of the two schemas. (Schema A was used for the tests predicting risk of any complication and intubation. Schema B was used for the tests predicting risk of ARDS and admission to the ICU.)

important for some patients.

3.2. Predicting ARDS

Test development for prediction of ARDS used the schema of Fig. 2B, with the flow of patients through the hierarchical classification scheme shown in Fig. 5. Note that, for this test, the lowest and lower risk groups were combined into a single lowest risk group, as the proportion of patients with ARDS was found to be similar in both subgroups during test development. In Fig. 4, we see the test predicting risk of developing ARDS classified 142 (62%), 47 (21%), and 40 (17%) patients to the lowest, low, and highest risk groups, respectively, in the development cohort. In the lowest risk group, only 8% of patients developed ARDS,

while in the highest risk group 45% of patients developed ARDS. The proportions of patients assigned to the lowest, high, and highest risk groups were similar in the validation cohort: 190 (58%), 73 (22%), and 67 (20%), respectively. The proportion of patients that developed ARDS increased with increasing risk group (Cochran-Armitage p = 0.02). Although the lowest risk group contained more than half of the patients, only 5% (95% CI: 3–9%) of them developed ARDS (Fisher's exact p (lowest vs other) < 0.0001). In contrast, the percentage of patients developing ARDS in the highest risk group was 27% (95% CI: 17–39%), Fisher's exact p (highest vs other) = 0.0004.

Test classification was associated with age, blood pressure, oxygen saturation, sodium, BUN, WBC, LDH, D-dimer, CRP, and ferritin. Shapley value analysis again showed that the attributes most important

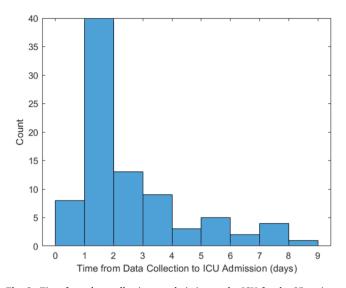


Fig. 3. Time from data collection to admission to the ICU for the 85 patients admitted to the ICU in the validation cohort indicating potential utility for ICU admission risk assessment at hospital admission. A time in the [0, 1] bin indicates the patient was admitted on the same day as the data was collected.

for classification generation varied by patient, but that CRP and D-dimer contributed substantially to a classification of lowest risk, with race and age also contributing for some patients. Highest risk classification was related to LDH, CRP, and BUN in most patients and oxygen saturation in some patients.

3.3. Predicting intubation

Test development to predict intubation used the schema of Fig. 2A, with the flow of patients through the hierarchical classification scheme shown in Fig. 5. In Fig. 6, we see the test predicting risk of intubation classified 74 (32%), 62 (27%), 55 (24%), and 38 (17%) patients of the development cohort to the lowest, low, intermediate, and highest risk groups, respectively and 86 (26%), 120 (36%), 67 (20%), and 57 (17%), respectively, in the validation cohort. The patients of the validation cohort were assigned 86 (26%):120 (36%):67 (20%): 57 (17%) to the lowest, low, high and highest risk groups. Only 1% (95% CI: 0–6%) of patients in the lowest risk group were intubated (Fisher's exact p (vs other <0.0001), while 33% (95 %CI: 21–47%) of patients in the highest risk group were intubated (Fisher's exact p (vs other) < 0.0001). There was a significant association of increase in intubation with increasing risk group (Cochran-Armitage p < 0.0001).

Test classification was associated with respiratory rate, oxygen saturation, BUN, creatinine, anion gap, WBC, LDH, D-dimer CRP, and ferritin. Shapley value analysis revealed that multiple attributes, varying by patient, were of increased relevance for classification. However, CRP was found to be generally most relevant for generation of a lowest risk classification, with D-dimer, LDH, BUN, race, and age also important factors. A highest risk classification was related to LDH, oxygen saturation, CRP, and BUN.

3.4. Comparison with other tests

An attempt was made to compare the performance of the tests to similar published classifiers [8,10,15]. The classifier described in Xiao et al. [12] applied on the development cohort yielded very similar areas under the receiver-operating characteristic (ROC) curves to those for the first classifiers in our hierarchies for each endpoint. Using the specified cutoff [10], relatively small, but pure lowest risk groups were observed for all four endpoints. While the purity was similar to that of our tests, the lowest risk group sizes were roughly half the size. The classifier

described by Liu et al. [8] used age and NLR to stratify patients into four risk groups. Across all four endpoints, purities in the lowest risk groups were similar to those presented here but were roughly half the size. The highest risk groups were nearly twice the size of those for our tests but had purities a factor of 2–3 smaller. Performance for predicting ICU admission was poor, with the low risk group containing a similar proportion of patients admitted to the ICU as the highest risk group. Many studies have looked at binary classification to similar prognostic endpoints [20–27]. While our tests stratify patients into more than 2 groups, the top classifier in each hierarchy achieved comparable performance in terms of AUROC.

4. Discussion

Using EHR-derived data recorded at the time of hospital admission, we developed and blindly validated a clinical decision support system composed of three different ML-based risk tests predicting three measures of progression to severe disease in patients hospitalized with COVID-19; namely risk of progression to ICU admission, intubation, and ARDS diagnosis. Given that many patients admitted with COVID-19 may not be in regular care and aware of comorbidities or may be critically ill upon presentation and unable to provide a detailed medical history, we utilized only attributes readily accessible in an emergency care setting.

Multiple previous studies have used ML-based algorithms to predict COVID-19 prognosis by combining basic patient demographics, vital signs, laboratory measurements, and comorbidities [14–16], sociodemographic information, comorbidities and current medications [19], or chest computed tomography (CT) scan alone [18,19] or with patient demographics [15]. The performance of our tests compared favorably with that of two existing risk assessment models for which we were able to generate classifications for our development set, with our test achieving similar negative predictive values in the lowest risk groups but superior specificity. However, for other simple nomogram-based models, we were not able to compare performance due to lack of information for required attributes, as these were not obtainable from collected EHR data.

We validated three tests by generating test classifications blinded to all outcome data and analyzing the results according to a prespecified statistical analysis plan. This validation study was possible using data directly extracted from the EHR, without any further curation, demonstrating that the data required for risk assessment was easily and readily accessible. Test performance was similar between the development and validation cohorts, illustrating the ability of the ML platform to generalize to unseen datasets. The successful validation of the performance of our models observed in test development shows that our selection of minimal attributes was rich in predictive information. Figs. 4–6 demonstrate significant enhancement in the outcome of interest from the baseline in each cohort to the final risk groups.

As with all modern ML-based modeling, it was not a priori obvious which attributes would most inform the results for an individual patient. Recent work using Shapley values has directly addressed model explainability at an individual patient level [40]. Our ML architecture made the calculation of exact Shapley values for all classifiers for 50 patients from the validation cohort computationally feasible without the need for commonly made approximations [41-42], supplement). The Shapley value analysis found that, when used in combination by the ML, simple clinical measures of risk and disease severity (age, weight, oxygen saturation) and general biomarkers of inflammation and acute phase reactants (CRP, D-dimer, LDH) were important for risk assessment. CRP, D-dimer, and LDH have been linked to COVID-19 severity in other studies, likely reflective of a pronounced inflammatory, systemic immune response with heightened thromboembolic risk [43-44]. Increasing age and body mass index are well-established risk factors for influenza severity [45], and may, in part, exacerbate the inflammatory response [46–47]. While race/ethnicity was not found to play a key role in explaining risk classification in the Shapley value analysis, a white

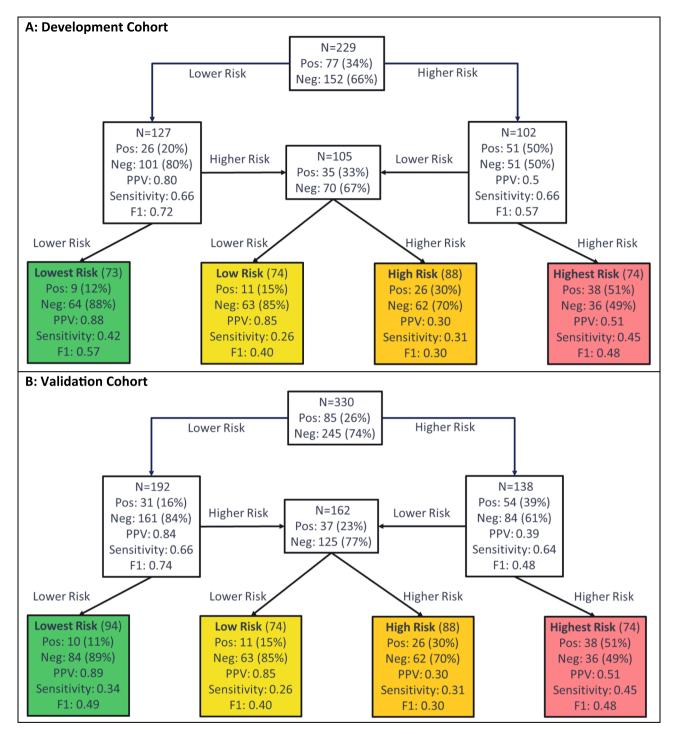


Fig. 4. Performance Flow Chart for the Risk Assessment Test for ICU Admission for (A) the Development Cohort and (B) the Validation Cohort. Each uncolored box represents a classifier with the contents reflecting the set of patients to be classified by the classifier. The colored boxes represent the final risk groups with the contents reflecting composition of the groups and test performance. Bootstrap 95% confidence intervals for performance metrics are given in the supplement. Pos = Positive (Admitted to ICU); Neg = Negative (Not Admitted to ICU), PPV = Positive Predictive Value.

race was observed to contribute towards a lower risk classification, while all other choices did not contribute substantially to risk category (see Fig. E22 in supplement). These observations were consistent with the associations we observed when we compared patient characteristics between our risk subgroups (see supplement). However, the Shapley values also revealed that our tests combined information across the 26 attributes in non-trivial ways and the attributes that explained patient classification differed from patient to patient. For some patients, all attributes had similar importance, while for others, some attributes

pointed towards higher risk, while others pointed towards lower risk.

In contrast to the single center development cohort, the validation cohort included patients from the statewide UC health system. However, the cohort size and restriction to Colorado are limitations of this study. While patients excluded from the development cohort due to missing data were generally similar to those included, in validation, patients with complete data (only 15% of the total available) exhibited higher rates of severe disease and generally worse prognostic factors (laboratory and vital signs) than those without. Further validation of the tests in

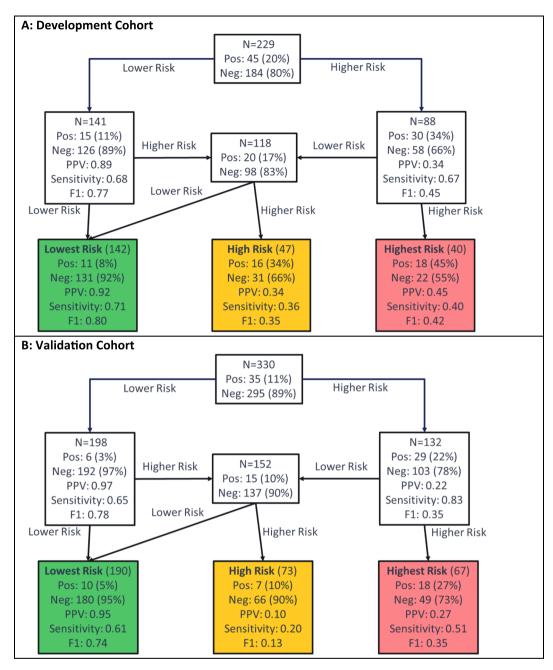


Fig. 5. Performance Flow Chart for the Test Predicting Risk of Developing ARDS in (A) the Development Cohort, (B) the Validation Cohort. Each uncolored box represents a classifier with the contents reflecting the set of patients to be classified by the classifier. The colored boxes represent the final risk groups with the contents reflecting composition of the groups and test performance. Bootstrap 95% confidence intervals for performance metrics are given in the supplement. Pos = Positive (Developed ARDS); Neg = Negative (Did not develop ARDS), PPV = Positive Predictive Value.

larger cohorts derived from other health systems and geographic areas is necessary.

The clinical utility of these tests will depend on their application. All tests achieved good performance in identifying patients who did not experience adverse outcome in the lowest risk groups in development and validation. Performance in identifying patients who experienced adverse outcomes in the highest risk groups was promising, but the risks of false positives and negatives would need to be carefully considered in an application where identifying patients likely to experience adverse outcome is desired.

In summary, we have developed and validated a suite of tests able to assess the risk of a poor outcome for patients hospitalized with COVID-19 based on information easily and routinely collected at time of hospital admission. Additional validation, preferably in a prospective setting, is required to further demonstrate the clinical utility of this risk assessment tool beyond clinical assessment alone. However, with readily-derived and quickly-available EHR data, a risk assessment at or near the time of admission can inform prognosis, guide discussions on the risks and benefits of treatments (including intubation), or identify low or high-risk patients for limited resources or enrollment in clinical trials. Furthermore, the methods here may be implemented in the care of future patients with novel viral infections.

5. Summary Table

What is already known?

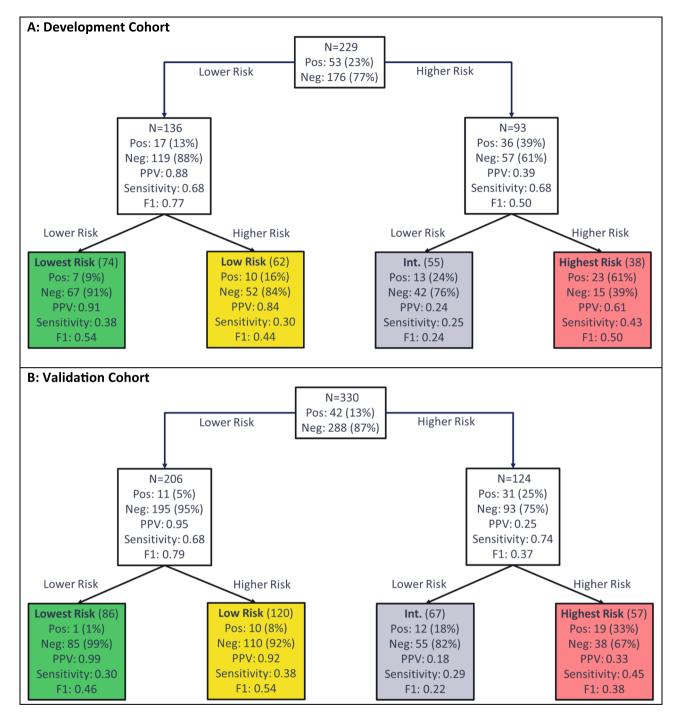


Fig. 6. Performance Flow Chart for the Test Assessing Risk of Intubation for (A) the Development Cohort and (B) the Validation Cohort. Each uncolored box represents a classifier with the contents reflecting the set of patients to be classified by the classifier. The colored boxes represent the final risk groups with the contents reflecting composition of the groups and test performance. Bootstrap 95% confidence intervals for performance metrics are given in the supplement. Pos = Positive (Intubated); Neg = Negative (Not intubated), PPV = Positive Predictive Value.

(continued)

- Basic lab results and other clinical and demographic attributes have predictive power in predicting risk of adverse events during COVID-19 hospitalization
- These include things like age, BMI, general inflammatory markers, and others
 Machine learning can be used to combine these attributes to make reliable
- Machine learning can be used to combine these attributes to predictions of COVID-19 prognosis

What does this study add?

• Robust, validated, multi class machine learning risk prediction for 3 endpoints during COVID-19 hospitalization using only easily collectable attributes that are common in EHRs

(continued on next column)

(continued)

 Rigorous explainability analysis to describe which attributes contributed more to the machine learning algorithms' assignment of risk not found in any similar work

CRediT authorship contribution statement

Thomas W. Campbell: Conceptualization, Formal analysis, Methodology, Software, Writing – original draft, Writing – review and editing, **Melissa P. Wilson:** Data Curation. **Heinrich Roder:** Supervision, Methodology, Software, Writing – original draft, Writing – review and editing. Samantha MaWhinney: Supervision, Writing – review & editing. Robert W. Georgantas: Supervision, Writing – review and editing. Laura K. Maguire: Formal analysis, Writing – review and editing. Joanna Roder: Supervision, Methodology, Formal analysis, Writing – original draft, Writing – review and editing. Kristine M. Erlandson: Conceptualization, Resources, Writing - review & editing.

Declaration of Competing Interest

The authors declare the following financial interests/personal relationships which may be considered as potential competing interests: 'The authors have no major conflicts of interest to disclose. TC, HR, RG, LM, and JR are named as inventors on a provisional patent assigned to Biodesix relevant to the work and hold stock and/or stock options in Biodesix. KE reports grant funding from the NIH during the conduct of the study and grant funding from Gilead and personal fees from Theratechnologies and ViiV outside of the conduct of the study'.

Appendix A. Supplementary material

Supplementary data to this article can be found online at https://doi.org/10.1016/j.ijmedinf.2021.104594.

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