# PROKARYOTES



## 

# Whole-Genome Sequence of *Chlamydia abortus* Strain GN6 Isolated from Aborted Yak Fetus

# Zhaocai Li,<sup>a</sup> Jinshan Cai,<sup>b</sup> Xiaoan Cao,<sup>a</sup> Zhongzi Lou,<sup>a</sup> Yilin Chao,<sup>b</sup> Wei Kan,<sup>b</sup> Jizhang Zhou<sup>a</sup>

State Key Laboratory of Veterinary Etiological Biology, Lanzhou Veterinary Research Institute, Chinese Academy of Agricultural Sciences, Lanzhou, Gansu, People's Republic of China<sup>a</sup>; Center for Animal Disease Control and Prevention in Qinghai Province, Xi'ning, Qinghai, People's Republic of China<sup>b</sup>

**ABSTRACT** The obligate intracellular Gram-negative bacterium *Chlamydia abortus* is one of the causative agents of abortion and fetal loss in sheep, goats, and cattle in many countries. It also affects the reproductivity of yaks (*Bos grunniens*). This study reports the whole-genome sequence of *Chlamydia abortus* strain GN6, which was isolated from aborted yak fetus in Qinghai-Tibetan Plateau, China.

**C**hlamydia abortus is one of the etiological agents of abortion in pregnant ruminants and some other mammals (1) and represents a zoonotic risk to humans (2, 3). The pathogen is distributed worldwide and has been shown to have relatively low genetic heterogeneity with six multilocus variable-number tandemrepeat analysis (MLVA) genotypes which, to a large extent, are related to their geographical origin (4, 5). Recently, yak flocks have demonstrated the severe problem of abortion associated with *Chlamydia* infection in several regions of the Qinghai-Tibetan Plateau, China (6). We have isolated several *Chlamydia* strains from the aborted yak fetuses and identified them as *C. abortus* belonging to the genotype 2 group (7). For controlling the disease, an inactivated vaccine was developed using one of the isolated strains, GN6, which exhibited high virulence. Administration of the vaccination significantly decreased the abortion rate in the yak flocks with a protective rate of over 95% (data not shown). Here we report the genome sequence of GN6, which will contribute to the understanding of the genetic diversity and antigen properties of *C. abortus*.

The *C. abortus* GN6 genome was sequenced using PacBio sequencing technology. A total of 174,311 reads were obtained, with an  $N_{50}$  read length of 10,815 bp and a mean length of 7,863 bp. Approximately 280× depth was achieved for the bacterial genome. *De novo* assembly was performed with the software program Canu (version 1.0) (8), and the genome sequences were assembled in one scaffold without any gaps. *C. abortus* GN6 is composed of a 1,144,357-bp circular chromosome with a G+C content of 39.86%. A total of 1,031 putative genes in the chromosome were predicted by use of the NCBI Prokaryotic Genome Annotation Pipeline, of which 987 were coding sequences and 44 were RNA genes. The genome contains 38 tRNAs, 3 noncoding RNAs (ncRNAs), and 3 types of rRNAs (55, 165, and 235) in one rRNA operon. There are 62 predicted pseudogenes in the strain, which is more pseudogene content than in the reference strains S26/3 and LLG and indicates the genetic heterogeneity of this isolate (9, 10).

Accession number(s). This complete genome sequence shotgun project of *C. abortus* strain GN6 has been deposited at DDBJ/EMBL/GenBank under the accession no. CP021996, and the version cited in this paper is CP021996.1.

Received 18 July 2017 Accepted 19 July 2017 Published 31 August 2017

Citation Li Z, Cai J, Cao X, Lou Z, Chao Y, Kan W, Zhou J. 2017. Whole-genome sequence of *Chlamydia abortus* strain GN6 isolated from aborted yak fetus. Genome Announc 5:e00893-17. https://doi.org/10.1128/genomeA.00893-17.

**Copyright** © 2017 Li et al. This is an openaccess article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Jizhang Zhou, zhoujizhang@caas.cn.

#### ACKNOWLEDGMENTS

Zhaocai Li, Jinshan Cai, Xiaoan Cao, and Jizhang Zhou designed this project. Zhaocai Li and Wei Kan grew the isolate, purified the EBs, and conducted DNA genomics. Zhaocai Li and Xiaoan Cao analyzed the sequence, and Wei Kan, Jinshan Cai, and Yilin Chao performed experiments.

We thank Shenzhen Zhongnong Jing Yue Biotechnology Co., Ltd., for technological assistance.

This work was supported by NSFC grant 31502081 and the Central Public-interest Scientific Institution Basal Research Fund (1610312016024).

### REFERENCES

- Longbottom D, Coulter LJ. 2003. Animal chlamydioses and zoonotic implications. J Comp Pathol 128:217–244. https://doi.org/10.1053/jcpa .2002.0629.
- Hagemann JB, Simnacher U, Longbottom D, Livingstone M, Maile J, Soutschek E, Walder G, Boden K, Sachse K, Essig A. 2016. Analysis of humoral immune responses to surface and virulence-associated *Chlamydia abortus* proteins in ovine and human abortions by use of a newly developed line immunoassay. J Clin Microbiol I54:1883–1890. https://doi .org/10.1128/JCM.00351-16.
- Walder G, Hotzel H, Brezinka C, Gritsch W, Tauber R, Würzner R, Ploner F. 2005. An unusual cause of sepsis during pregnancy: recognizing infection with *Chlamydophila abortus*. Obstet Gynecol 106:1215–1217. https://doi.org/10.1097/01.AOG.0000161060.69470.9c.
- Sait M, Clark EM, Wheelhouse N, Spalding L, Livingstone M, Sachse K, Markey BK, Magnino S, Siarkou VI, Vretou E, Caro MR, Yaga R, Lainson FA, Smith DG, Wright F, Longbottom D. 2011. Genetic variability of *Chlamydophila abortus* strains assessed by PCR-RFLP analysis of polymorphic membrane protein-encoding genes. Vet Microbiol 151:284–290. https:// doi.org/10.1016/j.vetmic.2011.03.005.
- Laroucau K, Vorimore F, Bertin C, Mohamad KY, Thierry S, Hermann W, Maingourd C, Pourcel C, Longbottom D, Magnino S, Sachse K, Vretou E, Rodolakis A. 2009. Genotyping of *Chlamydophila abortus* strains by

multilocus VNTR analysis. Vet Microbiol 137:335–344. https://doi.org/10 .1016/j.vetmic.2009.01.029.

- Chen Q, Gong X, Zheng F, Cao X, Li Z, Zhou J. 2014. Seroprevalence of *Chlamydophila abortus* infection in yaks (*Bos grunniens*) in Qinghai, China. Trop Anim Health Prod 46:503–507. https://doi.org/10.1007/ s11250-013-0519-8.
- Li Z, Cao X, Fu B, Chao Y, Cai J, Zhou J. 2015. Identification and characterization of *Chlamydia abortus* isolates from yaks in Qinghai, China. BioMed Res Int 2015;658519. https://doi.org/10.1155/2015/658519.
- Berlin K, Koren S, Chin CS, Drake JP, Landolin JM, Phillippy AM. 2015. Assembling large genomes with single-molecule sequencing and locality-sensitive hashing. Nat Biotechnol 33:623–630. https://doi.org/10 .1038/nbt.3238.
- Thomson NR, Yeats C, Bell K, Holden MT, Bentley SD, Livingstone M, Cerdeño-Tárraga AM, Harris B, Doggett J, Ormond D, Mungall K, Clarke K, Feltwell T, Hance Z, Sanders M, Quail MA, Price C, Barrell BG, Parkhill J, Longbottom D. 2005. The *Chlamydophila abortus* genome sequence reveals an array of variable proteins that contribute to interspecies variation. Genome Res 15:629–640. https://doi.org/10.1101/gr.3684805.
- Sait M, Clark EM, Wheelhouse N, Livingstone M, Spalding L, Siarkou VI, Vretou E, Smith DG, Lainson FA, Longbottom D. 2011. Genome sequence of the *Chlamydophila abortus* variant strain LLG. J Bacteriol 193: 4276–4277. https://doi.org/10.1128/JB.05290-11.