A Journal of the Society for Integrative and Comparative Biology

RESEARCH ARTICLE

A Global Synthesis of the Correspondence Between Epizoic Barnacles and Their Sea Turtle Hosts

John D. Zardus (D*

Department of Biology, The Citadel, 171 Moultrie Street, Charleston, SC 29409, USA

*E-mail: john.zardus@citadel.edu

Synopsis Barnacles that are obligate epizoites of sea turtles are not parasites in the traditional sense. However, they can impair their hosts in some instances, disqualifying the association as strictly commensal. Characterizing these interactions requires knowing which epibionts pair with which hosts, but records of barnacles from sea turtles are scattered and symbiont/host match-ups remain equivocal. The objective of this study was to collate global records on the occurrence of barnacles with sea turtles and describe each species pair quantitatively. Records reporting barnacles with sea turtles were searched spanning the last 167 years, including grey literature, and findings were enumerated for 30,580 individual turtles to evaluate prevalence. The data were summarized globally as well as subdivided across six geographic regions to assess constancy of the affiliations. Patterns of partnering were visualized by hierarchical clustering analysis of percent occurrence values for each barnacle/turtle pair and the relative selectivity of each symbiont and susceptibility of each host were evaluated. After adjusting for synonymies and taxonomic inaccuracies, the occurrence of 16 nominal species of barnacles was recorded from all 7 extant sea turtle species. Mostly, barnacles were not specific to single turtle species, partnering on average with three hosts each. Neither were barnacles entirely host-consistent among regions. Three barnacles were common to all sea turtles except leatherbacks. The most common, widespread, and least selective barnacle was Chelonibia testudinaria, the only symbiont of all turtles. Excluding single-record occurrences, the barnacle Stomatolepas transversa was the only single-host associate of any hard-shell sea turtle (the green sea turtle) and Platylepas coriacea and Stomatolepas dermochelys were exclusive associates of leatherback sea turtles. Green sea turtles were the most vulnerable to epibiosis, hosting 13 barnacle species and Kemp's ridley sea turtles were the least, hosting three. Geographically, there was an average of nine barnacle species per world region, with diversity highest in the Pacific Ocean (12 species) and lowest in the Mediterranean Sea (6 species). It is paradoxical that the flexibility of barnacles for multiple host species contrasts with their overall strict specificity for sea turtles, with each symbiont occupying a virtually unique suite of turtle hosts.

Barnacles, to the undiscerning eye, are as boring as rivets. This is largely attributable to the erroneous impression that they don't go anywhere and don't do anything, ever. The truth of the matter is that they don't go anywhere and don't do anything merely sometimes—and that, other times, barnacle life is punctuated with adventurous travel, phantasmagorical transformations, valiant struggles, fateful decisions, and eating.

David Quammen, Point of Attachment, 1998

Introduction

Barnacles are the epitome of a sessile animal, so it seems incongruous that some are among the widestroving invertebrates on the planet, maybe beyond even what Mr. Quammen has conceived. All

members of the barnacle superfamily Coronuloidea have acquired the borrowed ability to travel many hundreds, or thousands of kilometers over a lifetime as epibionts of their mobile hosts or basibionts. These epizoites of sea turtles, sea snakes, crabs, whales, and other mobile fauna (Darwin 1854; Zann 1975; Hayashi 2013a), utilize their live substratum as a platform for feeding and dispersal, but not as a source of nutrition. The greatest number specializes on sea turtles but their host repertoires and specificity for particular turtle species have not previously been rigorously evaluated. I endeavor to fill this gap to gain greater understanding of these



Fig. 1 Barnacle exemplars that attach to sea turtles by: (A) cementing, Chelonibia testudinaria; (B) clinging, Platyleaps hexastylos; (C); embedding, Stomatolepas elegans [the shell is right side up!]; (D) penetrating, Stephanolepas muricata; and (E) boring, Chelolepas cheloniae (scale bars = 5 mm).

associations and insight on how barnacles select a mobile home.

The nature of barnacle-turtle relationships falls somewhere between parasitism and mutualism. Not a phoretic symbiosis, whereby one animal attaches to another temporarily for conveyance (White et al. 2017), the association is most frequently considered a commensalism. The term refers to feeding at a common table (van Beneden 1876) and results in the commensal deriving benefit from the arrangement while the host remains unaffected. However, the view that turtle hosts are not negatively affected by barnacles is, depending on the situation and the symbiont involved, often plausible, in some cases debatable, occasionally and untenable. Commensalisms are rarely demonstrated as obligate and specific (Wahl and Mark 1999) and many are not stable, tipping toward the benefit or detriment of the host depending on circumstances; indeed, the argument has been made that the concept of commensalism, in the narrow sense, is a theoretical state that cannot be empirically demonstrated (Zapalski 2011). Thus, multiple states apply for epizoic barnacles.

Whatever the designation, advantages certainly accrue to barnacles by living on mobile hosts, including access to reliable currents for passive feeding and protection from benthic predators (Foster 1987). The obligate nature of coronuloid barnacles on their hosts, especially as associates of whales and turtles (Scarff 1986; Hayashi 2013a), is well-demonstrated, with several taxa known from only a single species of cetacean or chelonian (Marloth 1900; Monroe and Limpus 1979). Of the more than 200 species of epibionts documented from sea turtles (Frick and Pfaller 2013), most are facultative associates, but barnacles are the most common obligate taxa. Yet, despite their affinity for sea turtles, a one barnacle—one turtle paradigm is likely the exception rather than

the rule. For example, the common "turtle" barnacle, *Chelonibia testudinaria* (Fig. 1A), has the most elastic host use, regularly utilizing manatees, various crabs, and all species of sea turtles as basibionts (Zardus et al. 2014) and exceptionally, other kinds of reptiles and crustaceans (Ross and Jackson 1972; Badrudeen 2000; Ortiz et al. 2004; Nifong and Frick 2011), even various synthetic substrata (Edmondson and Ingram 1939; Frazier and Margaritoulis 1990; Sloan et al. 2014). Lacking an all-encompassing label to describe the relationship of barnacles with sea turtles, it is perhaps most easily summarized as an obligate association of generally intermediate specificity with neutral to negative consequences for the host.

No reciprocal benefit to sea turtles has been demonstrated by the attachment of barnacles, though the idea that they may provide disruptive camouflage has been suggested (Wahl and Mark 1999; Kobayashi 2000). With most barnacle species, the partnership is benign for turtles under typical conditions, depending primarily on attachment mode. Some species of barnacle cement to or bore into turtle hard parts such as shell and scales, others latch on to leathery epidermis or embed in the supple skin of the neck and limbs. For example, the skin-clinging barnacle Platylepas hexastylos (Fig. 1B) loosely clamps to the outer layers of turtle epidermis but causes no apparent damage or disadvantage (Flint 2009). The superficially-embedding Stomatolepas elegans (syn. S. praegustator) (Fig. 1C) presents a similar but curious case, causing no obvious wounding or irritation as it presses itself, without penetration, into the epidermis, even when densely inhabiting the tongue and gullet (Pilsbry 1910). However, when turtles are injured or debilitated in some way, they may acquire an unusually heavy load of barnacles (Herbert and Jacobson 1995) which reduces their swimming efficiency, especially for sub-adult turtles. Barnacles may also arbitrarily

overgrow the eyes or settle in wounds and exacerbate shell fissuring. A few species of barnacles are unequivocally and intrinsically harmful to sea turtles to varying degrees, but not in a parasitic sense. The barnacle *Stephanolepas muricata* (Fig. 1D) penetrates the epidermis of several species of sea turtles (Frick et al. 2011), mostly along the leading edges of flippers, causing pitting, lacerations, and bleeding. The most pernicious is *Chelolepas cheloniae* (*Tubicinella cheloniae* of some sources) (Fig. 1E) which bores into both epidermis and carapace (Flint et al. 2009), occasionally penetrating to the bone or body cavity, leading to infection and sometimes death of the host (Herbert and Jacobson 1995).

The evolutionary history of the association of barnacles with sea turtles is ancient but not fully resolved. Though both host and symbiont leave fossil remains, barnacle shell plates typically disarticulate at death and their remnants are scarce (Collareta et al. 2019). Fossil evidence indicates the lineage of extant sea turtles, Superfamily Chelonioidea, has its genesis around 130 mya in the Early Cretaceous (Evers and Benson 2019). The leatherback Dermochelys coriacea, is the oldest living taxon, likely originating in the Eocene (Cadena and Parham 2015). Of the hardshell species, green and hawksbill sea turtles (Chelonia mydas and Eretmochelys imbricata, respectively) are next oldest, appearing in the Miocene (perhaps along with the soft-carapaced flatback sea turtle, Natator depressus, as well). They are followed by the loggerhead sea turtle, Caretta caretta, in the Pliocene and lastly the two species of ridley sea turtles, the olive ridley, Lepidochelys olivacea and Kemp's ridley, Lepidochelys kempii (Bowen and Karl 2007; Cadena and Parham 2015). The oldest fossil coronuloid barnacle is an Eocene chelonibiid dating to 34-38 mya and sharing structural features with recent Chelonibia species (Ross and Newman 1967). Although, turtle-fouling per se is argued not to have originated until 20-23 mya with the extinct genus Protochelonibia (Harzhauser et al. 2011). Epizoism in coronuloid barnacles may have commenced with molluscan or crab hosts (Ross and Newman 1967) before transferring to turtles and later whales, though no fossil evidence is known. A counter view is offered by a fossil-calibrated phylogeny (Hayashi et al., 2013) which puts coronuloid origins in the Early Eocene, inferring a mid-Eocene appearance of the clade leading to the skin-attaching barnacles, Stomatolepas, and Platylepas, pre-dating by ~10 million years the shell-cementing Chelonibia clade.

While adult barnacles are largely sessile, they have microscopic free-swimming larvae which, in the case of epizoic species, enable them to reach mobile hosts (Moyse 1961; Molenock and Gomez 1972; Zardus and Hadfield 2004; Nogata and Matsumura 2006; Liu et al. 2016). Barnacles typically reproduce through direct copulation and disperse via a swimming larval phase that disseminates widely in the plankton (Anderson 1994). Following a week or more of planktonic development, passing through seven larval stages, these crustaceans attach to a suitable host only at the final cyprid stage. More than a dozen species of barnacles select their hosts from a smorgasbord of seven extant sea turtle species that vary in habitat use and geographic distribution. Sea turtles mostly inhabit tropical and subtropical regions seeking food variously as herbivores, omnivores, and specialists of sponges and jellyfish, roaming as restricted coastal foragers to open ocean nomads (Spotila 2004). In addition to finding an acceptable basibiont, the barnacles must also target a specific attachment location on the host (Robinson et al. 2019).

For barnacles of rocky shores, it is possible to explain patterns of population connectivity with oceanographic processes (Chan et al. 2012; Wares 2020). But linkage dynamics for species whose dispersal is also influenced by host movements presents a challenge to characterizing biogeographic boundaries. Intuitively, the phylogeography of any hitchhiking associate of a mobile host should follow its hosts' distribution. But contrary to this expectation, some epibionts exhibit genotypic distributions that differ substantially from their hosts'. For instance, a cosmopolitan leech, parasitic with multiple sea turtle species shows, surprisingly, no genetic variation between Atlantic and Pacific oceans like its hosts (Tseng et al. 2018). And whale lice, amphipod crustaceans which dine on sloughing skin, complete their entire life cycle on a single cetacean (Balbuena and Raga 1991) but exchange genes extensively beyond individual host platforms and boundaries of whale subpopulations (Kaliszewska et al. 2005).

Although living on mobile hosts certainly benefits dispersal of epizoites, it creates the countermanding challenge of needing to attach to itinerant substrata that vary in material properties and that are highly limited due to rarity. Sea-turtle geospatial distributions, though incompletely characterized on a fine scale, are coarsely known and five of the seven species are circumglobal in tropical and subtropical waters. The exceptions are Kemp's ridley and flatback sea turtles which are both regionally restricted, the former nesting only in the Gulf of Mexico and wandering into the western Atlantic and the latter inhabiting the islands of the Torres Strait and the northern

coast of Australia from the Indian to the Pacific Ocean (Wallace et al. 2010). Where turtles and barnacle larvae meet remains inconclusive but the feeding or nesting localities of adult turtles offer a clue that it happens coastally for most species (Sloan et al. 2014). The particular abilities of barnacles to find and colonize sea turtles may lie in life history strategies that synchronize larval production with host movements and a chemosensitive and highly versatile cyprid attachment organ (Al-Yahya et al. 2016; Dreyer et al. 2020).

That barnacles associate with sea turtles and other mobile megafauna has undoubtedly been recognized since antiquity (Blick et al. 2010), but historical documentation is known only from several hundred years ago (Hayashi 2014). The first edition of Systema Naturae, (Linnaeus 1758) enters green and loggerhead sea turtles into the scientific record along with one of their most common barnacles, C. testudinaria (as Lepas testudinaria). However, taxonomic identities among all extant sea turtles were only stabilized in the latter half of the 20th century through detailed survey work (Carr and Caldwell 1956; Hughes et al. 1967) and phylogenetic study (Bowen et al. 1993). Foundational literature on cirripedes over the last ≥ 150 years (Darwin 1854; Gruvel 1905; Pilsbry 1916; Newman and Ross 1976) set the stage for contemporary systematics of barnacles (Pérez-Losada et al. 2004; 2014), but the modern era of understanding the diversity of barnacles associated with sea turtles dawned with Monroe and Limpus (1979). To date, correspondence between the known barnacle species and their sea turtle hosts remains illdefined, scattered among taxonomic lists of barnacles and survey records of sea turtles, hampered by unrecorded or imprecisely identified hosts in the former and misidentified or unidentified symbionts in the latter. Without a detailed compilation of global records, it cannot be stated with certainty which barnacle species affiliate with which turtle species, how specific their associations are, and whether patterns differ geographically. This hinders both our understanding of host selectivity by barnacles and knowledge of epibiont susceptibility among sea turtle.

The primary objectives of this study were to catalog the association of obligate, epizoic barnacles with sea turtles and assess their degree of specificity. Global records enumerating the numbers of turtles by species hosting each kind of barnacle were searched to collate and quantitatively evaluate the incidence of pairing between each host and symbiont and gauge the relative prevalence of association. The same comparisons were also made within geographic subdivisions to test the fidelity of barnacle/turtle

match-ups globally. Breadth in host utilization was predicted to vary by symbiont, so a barnacle selectivity index was generated, evaluating the number of host species exploited by each taxon. Similarly, a measure of susceptibility to epibiosis for each turtle species was also formulated from barnacle occurrence rates.

Methods

A heterogeneous assemblage of records documenting associations between epizoic coronuloid barnacles and sea turtles was searched from Darwin (1854), the beginning of stable barnacle taxonomies, to the present. Sources fell into three broad categories: (1) "peer-reviewed publications" comprising published journal articles, edited monographs, and books; (2) "technical reports," consisting of government and private reports, theses, and conference proceedings; and (3) "collections," encompassing specimens in museum and private collections. No attempt was made to compile information on species of epizoic stalked barnacles (Lepadomorpha) or opportunistic acorn barnacles. Particular attention was paid to locating records enumerating the number of turtles without accompanying barnacles. Accounts were disregarded that did not mention barnacles or identify them to a useful degree. A catalog was generated from the records, enumerating the sea turtles examined by species and the proportion hosting each species of barnacle (Appendix I). The data were summed and tabulated, first for global metrics then again for geographic comparisons, subdividing the world into six regions: (1) Atlantic Ocean, (2) and Gulf Caribbean Sea of Mexico, Mediterranean Sea, (4) Indian Ocean, (5) Pacific Ocean, and (6) Central Indo-Pacific (defined herein as those seas and straits from the northern coast of Australia in the south to the shores of Southeast Asia bordering the South China Sea in the north, and from the east side of Sumatra across the Indonesian archipelago to the Philippine Islands in the west).

For purposes of calculating percent occurrence, a value of zero was applied to turtles for each barnacle species specifically stated as absent or not mentioned but known to occur with that host in the region. In several instances where barnacle species were not reported they could be inferred from descriptions or photographs. In surveys, where "most" or a "majority" of the turtles were listed as having barnacles, a proportion of 75% was used as an estimate. Misidentifications of host turtles and more commonly the barnacles, especially in early records,

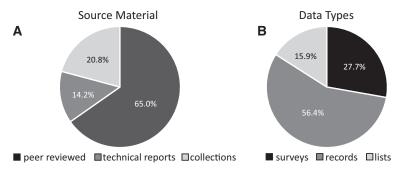


Fig. 2 Proportion of 289 searched records by category (A) and by type of data resulting (B).

were corrected where known with the latest information on synonymies and annotated accordingly in the catalog. For example, recent work confirms that leatherback sea turtles associate with only a few specific barnacles (Robinson et al. 2017b) and for any accounts mentioning P. hexastylos or S. elegans with leatherbacks, the species P. coriacea and S. dermochelys were substituted, respectively. The single report of the sea snake barnacle P. indicus from a leatherback (Fernando 2006) is probably a misidentification of P. coriacea and was treated as such. In some records, locations on the turtle body where barnacles attached were noted, providing useful qualitative information, but such reporting was too infrequent for quantitative comparisons. Likewise, barnacle counts or densities per turtle were not analyzed due to a paucity of information.

Patterns of association between barnacles and sea turtles were visualized by charting hierarchical clustering for each barnacle/turtle pair using the unweighted pair group method with arithmetic mean (UPGMA) of Euclidean distances of the global occurrence rates. From the tabulated data, two indices of association were calculated: first a measure of host selectivity for each barnacle species (S_b), ranging from 10 (most selective) to 0 (least selective), and second a value estimating the susceptibility of each turtle species to barnacle epibiosis (S_t) ranging from 10 (most susceptible) to 0 (least susceptible) using quotients of barnacle and turtle species as follows:

(1) barnacle selectivity index
$$S_b = \left(1 - \frac{T_x}{N_t}\right) \cdot 10$$

(2) turtle susceptibility index $S_t = \frac{B_x}{N_b} \cdot 10$,

where "T" represents the number of sea turtle species found hosting barnacle species "x," " N_t " represents the total number of sea turtle species globally (i.e., 7), "B" represents the number of barnacle species hosted by turtle species "x," and " N_b " represents the total number of named barnacle species globally (i.e., 16).

Results

Searching peer-reviewed publications, technical reports, and collections from the past 167 years, I found 289 records reporting sea turtles hosting named coronuloid barnacles (Fig. 2A). A complete list of sources annotated with counts and explanations for debatable species determinations is provided in Supplementary Material (Appendix I). The data were spread among three categories: "surveys," tallies of targeted sea turtle populations mentioning the presence or absence of barnacles by species and sometimes their abundance; "records," studies of sea turtles mentioning chance reports of barnacles; and "lists," museum or regional taxonomic inventories identifying barnacles and host species (Fig. 2B). In total, percent occurrences of turtles with barnacles were tallied from 30,580 individually observed sea turtles, covering all seven extant species, with 16 nominal species of barnacles identified as obligate associates (Table 1).

Barnacle perspective

The global percent occurrence of sea turtles hosting each barnacle taxon varied widely by species, revealing a continuum from highly selective to indiscriminate host preference by the barnacles. Hierarchical clustering by percent occurrence revealed differentiation of the barnacles into four broad groups based on the type and quantity of hosts they occupied (Fig. 3). The barnacles C. testudinaria, P. hexastylos (and perhaps other unidentified Platylepas species), and S. elegans, comprised a core set of barnacles common to all sea turtles except leatherbacks. Overall, most barnacles partnered with three turtle species each (avg. 2.9). The barnacle calculated to occur numerically more often than any other, was the geographically and host-restricted species Cylindrolepas darwiniana. This is a mathematical bias explained by two unusually large surveys of \geq 12,000 green sea turtles in the Galapagos Islands

Table 1 Global correspondence of epibiont/basibont associations between the seven sea-turtle species of the world and the 19 coronuloid barnacle taxa (16 named species) obligate with them. Values presented are percentages of turtles hosting each epibiont apportioned by turtle species: green sea turtle, *Chelonia mydas* (C.m.); hawksbill sea turtle, *Eretmochelys imbricata*, (E.i.); loggerhead sea turtle, *Caretta caretta*, (C.c.); leatherback sea turtle, *Dermochelys coriacea*, (D.c.); olive ridley sea turtle, *Lepidochelys olivacea*, (L.o.); flatback sea turtle, *Natator depressus*, (N.d.); and Kemp's ridley sea turtle, *L. kempii*, (L.k.) with the global average across all turtles sampled in parentheses. The number of turtles surveyed (Nt) were obtained from the published literature and other sources as detailed in Appendix 1. In some instances a barnacle species was reported present (P) with a species of turtle but no host tally was recorded. Those barnacle taxa not reported occurring with a turtle species are indicated by the null set (Ø). Additionally, the final column and row of the table presents, respectively, the index of increasing host selectivity for each barnacle (Sb) and the index of decreasing vulnerability to epibiosis for each species of sea turtle (Vt) (see text for derivation of indices).

Associated barnacle	Host turtle	C.m.	E.i.	C.c.	D.c.	L.o.	N.d.	L.k.	
Taxa	N _t (30,580)	20,911	5,288	2,679	691	569	328	114	S _b (0–10)
Calyptolepas bjorndale	(0.02)	0.02	Ø	Ø	Ø	Ø	Ø	Ø	8.6
Chelonibia sp. ^a	(4.29)	1.76	12.67	9.97	Ø	1.23	Ø	Ø	
C. caretta	(6.86)	0.11	34.42	9.56	Ø	Ø	Ø	Ø	5.7
C. testudinaria	(16.97)	17.90	3.21	32.03	0.14	25.13	75.00	22.81	0.0
C. ramosa ^b	(<0.01)	< 0.01	Ø	Ø	Ø	Ø	Ø	Ø	8.6
Chelolepas cheloniae	(1.68)	0.17	9.00	0.07	Ø	Ø	0.30	Ø	4.3
(syn. Tubicinella cheloniae)									
Cylindrolepas darwiniana	^f (18.74)	27.41	Ø	Р	Ø	0.18	Ø	Ø	5.7
C. sinica	(0.01)	0.01	Р	Р	Ø	Ø	Ø	Ø	5.7
Platylepas sp. ^c	(1.87)	0.51	1.46	2.24	Ø	Ø	100.00	Ø	
P. coriacea	(1.72)	Ø	Ø	Ø	76.12	Ø	Ø	Ø	8.6
P. decorata	^f (14.38)	20.58	1.68	Ø	Ø	1.05	Ø	Ø	5.7
P. hexastylos	(3.61)	3.41	0.32	9.48	Ø	20.91	Р	1.75	1.4
Stephanolepas muricata	(0.55)	0.09	Р	5.41	Ø	Р	Ø	Ø	4.3
Stomatolepas sp. ^d	(0.03)	0.04	Ø	Ø	Ø	Ø	Ø	Ø	
S. dermochelys	(1.45)	Ø	Ø	Ø	63.97	Ø	Ø	Ø	8.6
S. elegans	(1.58)	0.05	0.08	5.49	Ø	55.36	1.52	0.88	1.4
(syn. S. praegustator)									
S. pilsbryi	(0.01)	Ø	Ø	Ø	0.29	Ø	Ø	Ø	8.6
S. pulchra ^e	(<0.01)	< 0.01	Ø	Ø	Ø	Ø	Ø	Ø	8.6
S. transversa	(0.16)	0.24	Ø	Ø	Ø	Ø	Ø	Ø	8.6
	V _t (10-0)		8.1	5.0	5.0	2.5	3.8	2.5	1.9

^aSpecific epithet not reported in sources but assumed herein not to be *C. ramosa* (see footnote 2).

(Green 1998; Beaumont et al. 2008) where this barnacle was present with 47% of individuals. Ignoring this result, the globally most common, widespread, and least selective barnacle was *C. testudinaria* (Table 1). The largest and most conspicuous barnacle of sea turtles, it was the only species found associated with all host turtles (albeit rarely with leatherbacks) and other non-turtle hosts. *Platylepas decorata* also occurred at a relatively high frequency but with only three host species. Results for this species were also similarly skewed by the large

number of Galapagos samples mentioned above, along with the fact that it has probably been frequently confused with its congener *P. hexastylos*, the latter occurring more broadly across host species, appearing with all sea turtles except leatherbacks. Though not occurring as frequently, *S. elegans* was the only other barnacle to occur with six of the seven host species.

At the other extreme, seven barnacle species were recorded as highly selective, associating with only a single host species. Of these, four were reported from

^bValidity uncertain, described from one specimen that was destroyed (see Korschelt 1933).

^cSpecific epithet not reported in sources but assumed herein not to be *P. coriacea* (see the text for justification).

^dProbable unidentified species (see Pinou et al., 2013).

^ePossible synonym of S. transversa (see Hayashi, 2013a).

Value skewed upward by an indefinite amount (see the text for explanation).

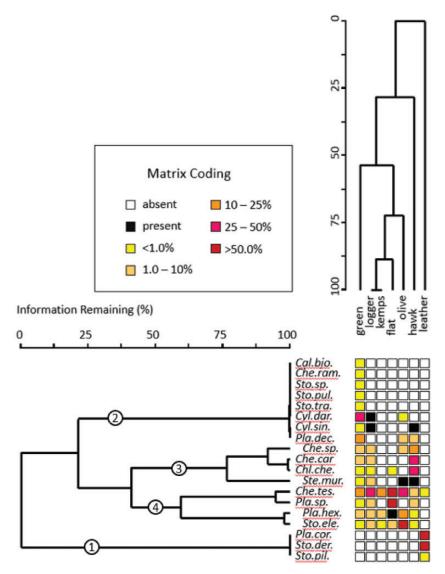


Fig. 3 Hierarchical clustering of barnacle and sea turtle species by global percent occurrence for each barnacle/turtle pair using UPGMA Euclidean distances. Clustering of the seven sea turtle partners is displayed vertically and clustering of the 16 named barnacle partners is presented horizontally. Circled numbers identify four barnacle clusters: 1) barnacles specific to leatherbacks, 2) barnacles with few hosts, 3) barnacles with a medium number of hosts, and 4) barnacles with many hosts. The global percent occurrence for each pairwise association is color-coded according to the matrix binning scheme indicated.

green and three from leatherback sea turtles. However, Calyptolepas bjorndalae, Stomatolepas pulchra, and Chelonibia ramosa are known only from single collection events (and only a single unverified specimen for the latter) and thus are of debatable taxonomic certainty. Stomatolepas pilsbryi, another solo-host affiliate, was described from only two leatherback sea turtles simultaneously, both from the same locality in the western Atlantic (Nova Scotia). Stomatolepas transversa, found in both the Atlantic and Pacific but in every case only with green sea turtles, most commonly occurred in the seams of the plastron and was the only indisputable single-host associate of a hard-shell turtle species. The

two other single-host barnacles were *Platylepas coriacea* and *S. dermochelys*, both of which occurred only on the skin of leatherback turtles; existing at higher global frequencies with their hosts than any other barnacles. The barnacle *Chelonibia caretta* also deserves mention for its specificity; though found with hawksbill, green, and loggerhead sea turtles wherever these turtles occurred, it was by a large margin most often found with hawksbill sea turtles. Overall, *C. caretta* occurred more frequently with hawksbill sea turtles while its congener, *C. testudinaria*, was more common with loggerheads and greens. Occurrences of *C. caretta* on turtles other than hawksbills merits further scrutiny.

A few barnacles occurred at high rates only with certain turtle hosts at select locations. The barnacles C. testudinaria and S. elegans occurred with \geq 50% individuals of particular host species in some locations. However, other than these two species, across a global scale, each barnacle species occurred with 35% or fewer individuals of any sea turtle species except in a large survey of the regionally restricted flatback sea turtle of which 100% of individuals an unspecified platylepadid (Limpus et al. 1983). This was very likely P. hexastylos which has been reported occurring with flatback sea turtles previously (Monroe and Limpus 1979) and is the most common and abundant nonleatherback associate of the genus, though it could also have been P. decorata.

Sea turtle perspective

Hierarchical clustering showed a stepwise differentiation of barnacle incidence among sea turtle species (Fig. 3). The leatherback sea turtle stood out as most distinctive in its barnacle associations due to being the sole host for P. coriacea and S. dermochelys. It was followed by the hawksbill sea turtle which was distinguished by a high incidence of intermediatelyoccurring barnacles, mostly C. caretta and C. cheloniae, two species that were infrequent or absent from other hosts. Green sea turtles, though hosting all common barnacles, differed from other hosts in their associations by the presence of several rare species not occurring with other turtles. Loggerhead, Kemp's ridley, olive ridley, and flatback sea turtles clustered closest together, predominantly due to sharing barnacle species that are common across most sea turtles. Olive ridleys were separated somewhat from this group, possibly due to hosting the barnacles C. darwiniana and P. decorata, sharing the former only with greens and loggerheads and the latter only with greens and hawksbills.

Among all hosts, green sea turtles exhibited the highest diversity of epizoic barnacles, hosting 13 species, lacking only three nominal barnacles associating exclusively with leatherbacks. Thus, green sea turtles ranked highest on the epibiosis susceptibility index (Table 1). Most turtle species were found associated with 6–7 barnacle species (avg. 6.4). Hawksbill, loggerhead, and olive ridley sea turtles hosted an intermediate diversity of barnacles (eight, six, and five species, respectively), followed by leatherback and flatback turtles with four each. With only three associated barnacles, Kemp's ridley turtles hosted the fewest barnacle species, registering the least susceptible to epibiosis. Leatherback sea turtles were

noteworthy in being the only turtle species with nearly reciprocally exclusive barnacle associations. In other words, the barnacles found on leatherbacks were not present on other sea turtle species, except for the rare occurrence of *C. testudinaria*.

Green sea turtles occur in all tropical to subtropical regions of the world ocean and were sampled for barnacles more than any sea turtle species (68.4%), mainly in the Pacific (62.0%) and to a lesser extent in the Indo-Pacific (27.8%). Few were sampled in the Caribbean/Gulf of Mexico (6.8%) and very few from the Indian (2.1%) or Atlantic (1.0%) oceans and Mediterranean Sea (0.2%). *Chelonibia testudinaria* associated with green sea turtles wherever they occurred and overall was the most prevalent barnacle of greens. *Stomatolepas transversa* was the only barnacle specific to green sea turtles.

Hawksbill sea turtles, the second-most frequently sampled turtles worldwide (17.3%), were examined for barnacles in every ocean and sea except the Mediterranean where the species does not occur. Hawksbills in the Atlantic had the greatest diversity of barnacles where they associated with four species. Most hawksbills were assessed from the Caribbean/ Gulf of Mexico (55.9%) and Indo-Pacific (33.5%) with smaller numbers from the Indian (5.5%) and Atlantic (4.8%) oceans and very few from the Pacific (0.4%). Chelonibia barnacles were by far the most common barnacle of hawksbills across all regions with C. caretta being the most prevalent species Caribbean/Gulf of Mexico, overall. In the Chelonibia barnacles were also most frequently encountered with hawksbill sea turtles, but most records did not distinguish between the barnacle species C. caretta and C. testudinaria. This obscures knowing whether C. caretta is the predominant barnacle of hawksbills in the Caribbean as it has been found to be in other regions. In surveys from the Persian Gulf of Iran, a remarkable contrasting distribution was recorded between two sites, both with robust samples sizes. At the island of Hormuz, C. caretta was the exclusive chelonibiid barnacle present with each of 41 hawksbill turtles surveyed (Devin and Sadeghi 2010); whereas, at Nakhiloo Island, \sim 500 km away, *C. testudinaria* was the only barnacle encountered and present with all but one of 122 hawksbill turtles (Razaghian al. et Confirmation is needed whether one or the other sites represents a case of mistaken barnacle identity. Platylepas hexastylos is another barnacle that was widely reported epizoic in the skin of hawksbill sea turtles as well as the harmful boring barnacle C. cheloniae which was frequently found with hawksbills in the Indo-Pacific.

Loggerhead sea turtles were sampled relatively infrequently for barnacles (8.8%) among all sea turtles surveyed. They were assessed most often in the Mediterranean (61.7%) followed by the Atlantic (21.3%), Pacific (11%), and Indian (5.9%) oceans. Very few were sampled in the Caribbean/Gulf of Mexico (0.4%) and none in the Indo-Pacific. As with green sea turtles, *C. testudinaria* was the most common barnacle of loggerheads. It occurred in every region inhabited by the turtle and was the most prevalent species in most instances. The greatest diversity of barnacles on loggerheads was in the Atlantic and Mediterranean. It is notable that *S. elegans* was also found with loggerheads everywhere they were sampled.

Leatherbacks accounted for a small percentage of the sea turtles investigated for barnacles (2.3%). They were sampled for barnacles by an order of magnitude more often in the Pacific (87.0%) than in the Atlantic Ocean (7.4%). But, sampled, at least to a small degree, in every ocean region: Caribbean/ Gulf of Mexico (3.6%), Mediterranean (1.3%), Indian Ocean (0.4%), and Indo-Pacific (0.3%). The barnacles P. coriacea and S. dermochelys were both exclusive to and common on leatherbacks worldwide. Though S. dermochelys was more common on leatherbacks overall, it was less common than P. coriacea in the Pacific. The reverse was true in the Atlantic. In the Atlantic, C. testudinaria was the only non-selective barnacle that was also recorded with leatherback sea turtles, albeit rarely.

Olive ridley sea turtles, which forage in the open sea, do not occur in the Caribbean/Gulf of Mexico nor in the Mediterranean Sea. However, from nesting data they are the most abundant sea turtle in the world's oceans by an order of magnitude (SWOT online database). Relative to all sea turtles sampled for barnacles, few olive ridleys were examined (1.9%), probably due to their open ocean foraging habitus, and their diversity of barnacles was not high. They were assessed for barnacles almost exclusively in the Pacific Ocean (91.2%) where the barnacle S. elegans was its most common associate (60.7%) followed by C. testudinaria (27.6%) and P. hexastylos (22.9%). Olive ridley turtles were not examined for barnacles in the Atlantic Ocean nor in the Indo-Pacific but a few were surveyed in the Indian Ocean (8.8%) where they were reported lacking in barnacles altogether.

Flatback sea turtles are restricted to a single geographic region spanning the islands of the Torres Strait and northern Australia from the Indian to the Pacific Ocean. They were only examined for barnacles at a single location, Crab Island in the Arafura Sea off the York Peninsula, where they hosted four species (Limpus et al. 1983). The most common was an unspecified species of *Platylepas* followed closely by *C. testudinaria*. Uncommonly they hosted *S. elegans* and a single dead turtle was found with *C. cheloniae*.

Kemp's ridley sea turtles, the least abundant sea turtle in the world and least inspected for barnacles, inhabit the most limited territory. Restricted to the Caribbean/Gulf of Mexico and northwestern Atlantic, they were mostly examined in the Gulf of Mexico (93.9%) where they nest and to a smaller degree in the western Atlantic (6.1%) where they occasionally roam. Kemp's ridley turtles hosted three species of barnacles. *Chelonibia testudinaria*, their most frequent associate, but not in great numbers, was common in the Caribbean and Gulf of Mexico but mostly lacking from individuals in the Atlantic where it was replaced by *P. hexastylos* and *S. elegans*.

Geographic perspective

In addition to host- and symbiont-associated patterns, there were macro-geographic components to barnacle occurrence as well, where some barnacles were found at different rates in different regions (Table 2). There was also at least one instance where a barnacle's attachment mode differed between host species. Specifically, *S. elegans* was observed attached to external skin of the neck and body in olive ridleys from the Pacific but fastened to the tongue in loggerheads from the western Atlantic (Lazo-Wasem et al. 2011; Pinou et al. 2013).

Of barnacles associated with sea turtles, there were on average 9.2 species per geographic region, with diversity highest in the Pacific Ocean (12 species) and lowest in the Mediterranean Sea (6 species). The rate of incidence of turtles with barnacles, averaged across all turtle and barnacle species, was highest in the Mediterranean and the Atlantic (9.2% and 9.1%, respectively). It was lowest for the Caribbean/ Gulf of Mexico (2.9%) and intermediate for the Pacific, Indian, and Indo-Pacific (7.2%, 5.4%, and 4.2%, respectively). However, the accuracy of this measure is questionable and must be interpreted cautiously since individual turtles hosting multiple barnacle species were tabulated as separate host occurrences for each barnacle species. Only four barnacle species were found in all geographic regions (C. caretta, C. testudinaria, P. hexastylos, and S. dermochelys) but, except for the last, each was not always primarily associated with the same hosts in each region. For instance, the barnacle P. hexastylos in the Pacific was hosted more frequently by

Table 2 Correspondence of epibiont/basibont associations between the 7 sea-turtle species of the world and the 19 coronuloid barnacle taxa (16 named species) obligate with them, grouped by host turtle species (A-G) and apportioned among 6 ocean regions: Caribbean Sea and Gulf of Mexico (C-GoM), Atlantic Ocean (Atl), Mediterranean Sea (Med), Indian Ocean (Ind), Indo-Pacific (Indo-Pac), and Pacific Ocean (Pac). Values presented are percentages of turtles hosting each symbiont by region with the global average for that host species in parentheses. Numbers of turtles surveyed (NT) were obtained from the published literature and other sources as detailed in Appendix 1. In some instances a barnacle species is reported merely as present (P) because no host turtle tally was taken. Additionally, the last column and row of the table presents, respectively, the number of regions in which each species of barnacle occurs (NR) and the number of barnacle species occurring in each region (NB). Dashes (–) indicate regions in which a particular turtle species does not nest or regularly occur while null symbols (ø) designate regions where a turtle resides but for which no barnacle surveys have been conducted.

A. Green sea turtle, C. mydas

	Ocean region	C-G ₀ M	Atl	Med	Ind	Indo-Pac	Pac	
Associated barnacle taxa	N _T 20,911	1,432	213	35	448	5,811	12,972	N_R
Calyptolepas bjorndale	(0.39)	0.00	2.35	0.00	0.00	0.00	0.00	1
Chelonibia sp.	(4.43)	23.81	0.00	0.00	2.68	0.02	0.10	
C. caretta	(5.35)	0.63	0.00	31.43	0.00	0.03	0.00	3
C. testudinaria	(25.93)	1.14	41.31	60.00	13.39	23.90	16.84	6
C. ramosa	(80.0)	0.00	0.47	0.00	0.00	0.00	0.00	1
Chelolepas cheloniae	(0.50)	0.00	0.00	0.00	2.68	0.22	0.09	3
(syn. Tubicinella cheloniae)								
Cylindrolepas darwiniana	(7.36)	0.00	0.00	0.00	0.00	0.00	44.18	1
C. sinica	(0.01)	0.00	0.00	0.00	0.00	0.03	0.00	1
Platylepas sp.	(4.99)	0.00	29.58	0.00	0.00	0.02	0.32	
P. decorata	(5.53)	0.00	0.00	0.00	0.00	0.00	33.17	1
P. hexastylos	(5.36)	0.14	14.09	0.00	13.17	0.00	4.79	4
Stephanolepas muricata	(0.02)	0.00	0.00	0.00	0.00	0.00	0.14	1
Stomatolepas sp.	(0.01)	0.00	0.00	0.00	0.00	0.00	0.07	
S. elegans	(0.03)	0.07	0.00	0.00	0.00	Р	0.08	3
(syn. S. praegustator)								
S. pulchra	(<0.01)	0.00	0.00	0.00	0.00	0.02	0.00	1
S. transversa	(0.17)	0.00	Р	0.00	0.45	0.02	0.36	4
	N_B	4	6	2	4	7	8	

B. Hawksbill sea turtle, E. imbricata

	Ocean region	C-GoM	Atl	Med	Ind	Indo-Pac	Pac	
Associated barnacle taxa	N _T 5,288	2,954	253	_	293	1,769	19	N_R
Chelonibia sp.	(4.60)	22.65	0.00	_	0.34	0.00	0.00	
C. caretta	(43.40)	2.37	79.05	_	26.28	82.98	26.32	5
C. testudinaria	(12.07)	1.29	0.79	_	42.32	0.17	15.79	5
Chelolepas cheloniae	(5.44)	0.00	0.00	_	0.34	26.85	0.00	2
(syn. Tubicinella cheloniae)								
Cylindrolepas sinica		0.00	0.00	_	0.00	0.00	Р	1
Platylepas sp.	(3.04)	1.22	0.00	_	13.99	0.00	0.00	
P. decorata	(2.69)	2.95	0.00	_	0.00	0.00	10.53	2
P. hexastylos	(7.04)	0.27	1.58	_	0.00	Р	26.32	4
Stephanolepas muricata		0.00	0.00	_	0.00	Р	Р	2
Stomatolepas elegans	(0.10)	0.10	0.40	_	0.00	0.00	0.00	2
(syn. S. praegustator)								
	N_B	5	4		3	5	6	

(continued)

C. Loggerhead sea turtle, C. caretta

	Ocean region	C-GoM	Atl	Med	Ind	Indo-Pac	Pac	
Associated barnacle taxa	N _T 2,679	10	571	1,654	158	0	286	N_R
Chelonibia sp.	(7.74)	0.00	2.98	12.94	22.78	Ø	0.00	
C. caretta	(6.10)	0.00	21.19	7.92	0.00	Ø	1.40	3
C. testudinaria	(26.51)	10.00	68.13	24.49	17.72	Р	12.24	6
Cylindrolepas darwiniana		0.00	Р	0.00	0.00	Ø	Р	2
Cylindrolepas sinica		0.00	0.00	0.00	0.00	Ø	Р	1
Platylepas sp.	(0.75)	0.00	0.18	3.57	0.00	Ø	0.00	
P. hexastylos	(7.53)	Р	13.31	9.61	1.27	Ø	5.94	5
Stephanolepas muricata	(2.53)	0.00	0.18	7.92	0.00	Ø	4.55	3
Stomatolepas elegans	(3.46)	Р	4.90	6.83	0.00	Ø	2.10	4
(syn. S. praegustator)								
	N_B	3	6	5	2	1	7	

D. Leatherback sea turtle, D. coriacea

	Ocean region	C-G ₀ M	Atl	Med	Ind	Indo-Pac	Pac	
Associated barnacle taxa	N _T 691	25	51	9	3	2	601	N_R
Chelonibia testudinaria	(0.33)	0.00	1.96	0.00	0.00	0.00	0.00	1
Platylepas coriacea	(50.01)	36.00	31.37	0.00	100.00	50.00	82.70	5
Stomatolepas dermochelys	(45.18)	4.00	60.78	22.22	66.67	50.00	67.39	6
S. pilsbryi	(1.31)	Ø	3.92	0.000	ø	Ø	0.00	1
	N_B	2	4	1	2	2	2	

(continued)

hawksbill (26.3%) and olive ridley (22.9%) sea turtles but in the Atlantic more often by green, loggerhead, and Kemp's ridley sea turtles in proportions nearly equal to each other (14.1%, 13.3%, and 14.3%, respectively). And *S. muricata* predominantly occurred with green turtles in the major oceans of the world and Caribbean Sea whereas in the Mediterranean it was reported only from loggerheads and in the Indo-Pacific from hawksbill and flatback sea turtles.

Cylindrolepas darwiniana and P. decorata were the most commonly reported barnacles of sea turtles in the Pacific (39.8% and 29.9%, respectively), biased by their high occurrence in the unusually large samples from the Galapagos Islands mentioned above. Otherwise, C. testudinaria was the barnacle most commonly associated with sea turtles in the Pacific (16.4%).

Within the Atlantic Ocean were found 11 barnacle species associated with sea turtles, of which *C. testudinaria* was the most prevalent by a wide margin (43.9%), followed by *C. caretta* (29.3%), then *P. hexastylos* (10.1%). *Chelonibia caretta* was strongly associated with hawksbill (79.1%) and to a lesser degree loggerhead sea turtles (21.2%) while *C. testudinaria* was most often found with loggerhead

(68.1%), followed by green (41.3%), followed by Kemp's ridley (14.3%), sea turtles.

Of eight barnacle species recorded from the Caribbean/Gulf of Mexico, *Chelonibia* barnacles occurred most commonly overall (22.3%) but whether *C. caretta* or *C. testudinaria* predominated was not detectable due to a lack of specificity in identifications. All other barnacle species found in the region occurred with \leq 2.0% of sampled sea turtles.

More sea turtles of the Mediterranean Sea hosted *C. testudinaria* (25.1%) than any other barnacle species. *Platylepas hexastylos, C. caretta*, and *S. muricata* were next most prevalent (9.4%, 8.4%, and 7.7%, respectively). Interestingly, the barnacle *P. coriacea*, commonly associated with leatherbacks throughout the world, was not reported from the Mediterranean. This may be due to limited sampling since the species has an otherwise robust global presence.

Of the seven barnacles most commonly occurring on sea turtles in the Indian Ocean, *C. testudinaria* (22.3%) was most common followed by *P. hexastylos* (6.4%). Surprisingly, *S. elegans*, though never highly prevalent but nevertheless widespread on multiple turtle species, was only reported from olive ridleys

E. Olive ridley sea turtle, L. olivacea

	Ocean region	C-G ₀ M	Atl	Med	Ind	Indo-Pac	Pac	
Associated barnacle taxa	N _T 569	-	ø	-	50	Ø	519	N_R
Chelonibia sp.	(0.67)	-	Ø	-	0.00	Ø	1.35	
C. testudinaria	(13.78)	-	Ø	-	0.00	Ø	27.55	1
Cylindrolepas darwiniana	(0.06)	-	Ø	-	0.00	Ø	0.19	1
Platylepas decorata	(0.58)	-	Ø	-	0.00	Ø	1.16	1
P. hexastylos	(11.46)	-	Ø	-	0.00	Ø	22.93	1
Stephanolepas muricata		-	Ø	-	0.00	Ø	Р	1
Stomatolepas elegans	(60.69)	-	Ø	-	Р	Ø	60.69	2
(syn. S. praegustator)								
	N_B	-	Ø	-	1	Ø	6	

F. Flatback sea turtle, N. depressus

	Ocean region	C-GoM	Atl	Med	Ind	Indo-Pac	Pac	
Associated barnacle taxa	N _T 328	-	-	-	ø	328	ø	N_R
Chelonibia testudinaria	(75.00)	-	-	-	Ø	75.00	Ø	1
Chelolepas cheloniae	(0.30)	-	-	-	Ø	0.30	Ø	1
(syn. Tubicinella cheloniae)								
Platylepas sp. ^a	(100.0)	-	-	-	Ø	100.00	Ø	
P. hexastylos		-	-	-	Ø	Р	Ø	1
Stomatolepas elegans	(1.52)	-	-	-	Ø	1.524	Ø	1
(syn. S. praegustator)								
	N_B	-	-	-	Ø	4	Ø	

^aPossibly P. hexastylos or P. decorata.

G. Kemp's ridley sea turtle, L. kempii

	Ocean region	C-GoM	Atl	Med	Ind	Indo-Pac	Pac	
Associated barnacle taxa	N _T 114	107	7	-	-	-	-	N_R
Chelonibia testudinaria	(18.83)	23.36	14.29	-	-	-	-	2
Platylepas hexastylos	(7.61)	0.93	14.29	-	-	-	-	2
Stomatolepas elegans	(7.14)	0.00	14.29	-	-	-	-	1
(syn. S. praegustator)								
	N_B	2	3	-	-	-	-	_

in the Indian Ocean. This too may be an artifact of under sampling.

The Central Indo-Pacific was another area of relatively high diversity, with 11obligate sea-turtle barnacles. *Chelonibia testudinaria* was most prevalent (20.7%), followed closely by *C. caretta* (18.6%).

The boring barnacle *C. cheloniae*, only known to range from the Indian to western Pacific Oceans, was modestly present in the center of its range (6.2%). *Stomatolepas pulchra* recorded only once from a single site in the South China Sea (Ren 1980) may be an endemic species though it is likely

a synonym of *S. transversa* (Hayashi 2013a). If endemic, it would have a distribution similar to *Cylindrolepas sinica*, known only from the South China Sea to the western and perhaps central-Pacific (Zardus and Balazs 2007; Hayashi 2009, 2013b).

Discussion

Barnacle substratum selectivity

The present analysis, confirms that a number of coronuloid barnacles are obligate epizoites of sea turtles, but illuminates somewhat of a paradox. Despite affiliating exclusively with particular sea turtles, many barnacle species exhibit relaxed fidelity for several turtle species, associating with overlapping suites of hosts, but with each symbiont occupying a virtually unique host subset. This elasticity might not be surprising were the barnacles all inhabiting turtle hosts in a similar way. But the various barnacle species exploit different microhabitats on turtles, glueing to shell, clamping to skin, or penetrating one or the other of these parts, raising the question why have more of them not become specific to single hosts? Host substratum may or may not vary substantially among turtle species. Both the yielding substratum of skin and the firm surface of shell are composed of keratin (Block and Bolling 1939; Solomon et al. 1986; Alibardi 2005; Kardong 2019), yet these two portions of the turtle body certainly differ physically and perhaps chemically as well. On observation, the surface micro-structure of turtle shell also appears different between species but how biomechanical or biochemical properties vary among hosts and between their parts have not been characterized. Regardless, the key to barnacle flexibility may lie not so much in their ability to detect properties of the substratum but in the versatility of the larval attachment organs, the antennules of the cyprid stage, which across disparate taxa are highly similar in form and structure, targeting substrata that is animate or inanimate, yielding or firm (Al-Yahya et al. 2016; Dreyer et al. 2020). Mechanically assessing the substratum, these larval organs also chemically sense and respond in complex ways to the presence of compounds from conspecific individuals and microbial biofilms (Hadfield 2011), in some cases strengthening adhesion as a response (Zardus et al. 2008). This may in part explain, for instance, how C. testudinaria attaches to keratinous, chitinous, and artificial substrates alike, allowing the maintenance of high host plasticity for millions of generations without specializing (Ewers-Saucedo et al. 2017).

This high versatility in attachment begs the question why are turtle barnacles not found more often on other kinds of hosts? The most common turtle barnacle, C. testudinaria, is the least selective, being the only species reported from all sea turtles. It is also the only "turtle" barnacle occurring on non-turtle hosts excepting the single report of P. hexastylos on a fish (Ryder 1879). Chelonibia testudinaria routinely occurs on various crabs and occasionally on sirenians, crocodilians, and artificial substrata (Zardus et al. 2014). Evolutionary radiation within the Coronuloidea has generated several species of "whale barnacles" that specialize on cetaceans (Scarff 1986; Collareta et al. 2016) and a pair of "sea snake" barnacles (Lanchester 1902; Daniel 1958). But there are no records of Coronuloid barnacles from other prospective living hard substrates such as mollusks or echinoids. Stalked barnacles by comparison are epizoic with a much wider variety of animals (e.g., jellyfish [Pagès 2000], sea urchins [Grygier and Newman 1991], mollusks [Landman et al.1987; Kolbasov and Zevina 1999], sharks [Rees et al. 2019], and opportunistically, pinnipeds, penguins [Reisinger and Bester 2010], and fish [Pilsbry 1907; Sumner et al. 1913; Crozier 1916]). The contrasting question also needs asking: Why are non-obligate, hard-substratum barnacle species not found more frequently on sea turtles? They are occasionally reported, mostly other balanomorph species in the Balanidae (Bugoni et al. 2001; Frick et al. 2004; Hayashi 2017) but also lepadomorph species (Eckert and Eckert 1987; Tachikawa 1995; Frick et al. 1998; Kitsos et al. 2005) and rarely verrucomorphs (Miranda and Moreno 2002). But more often than not, when present, these opportunists are found attached to underlying coronuloid barnacles, not to turtle shell directly.

Single-host barnacles

A few turtle barnacles are very host-selective, associating with only a single species of turtle. For instance, all barnacles occurring with leatherback sea turtles, except for *C. testudinaria*, are found with no other turtles. This degree of selectivity is perhaps not surprising since leatherbacks differ greatly from all other turtles, both phylogenetically and morphologically. The evolutionarily earliest extant sea turtle, the leatherback, pre-dates all others by ~45 mya or more (Cadena and Parham 2015) and instead of being shielded by a shell of keratinous scutes, it is covered by dermis. Somewhat surprising is that among barnacles of hard-shelled sea turtles, excluding single and unverified reports, the barnacle *S. transversa* is

the only single-host species, associating exclusively with the green sea turtle, occupying the very particular niche of the seams of the plastron. This species of turtle is noteworthy in being the most susceptible to barnacle epibiosis, hosting the widest variety of coronuloid cirripeds. At the other extreme, leatherback and olive ridley sea turtles host a low diversity of barnacles, possibly due to their lifestyle of spending most of their time in the open ocean where acorn barnacle larvae are likely less abundant. Stalked barnacles particularly, a component of the open ocean rafting community, are an indication of pelagic existence and are common on leatherbacks (Hughes 1970; Eckert and Eckert 1988; Robinson et al. 2017b), olive ridleys (Robinson et al. 2017a, 2017b), and distantly-straying Kemp's ridley sea turtles (Covelo et al. 2016). Ridley sea turtles are the most recently evolved sea turtles (Dutton et al. 1996; Naro-Maciel et al. 2008) and are known for their waxy surfaces (Weldon et al. 1990), which may be a deterrent to epibiosis, especially Kemp's ridleys which spend much of their lives near coastlines but host the lowest diversity of barnacles of any sea turtle. The barnacle S. muricata has been found associated in the South Pacific with open-ocean loggerhead sea turtles but not with coastal residents (Limpus and Limpus 2003). Sea turtle natural history may also influence rates of epibiosis. Both ridley species nest in arribadas or synchronized, mass assemblages, though olive ridleys can also be solitary nesters (Dornfeld et al. 2015). Synchronized nesting may reduce chances for barnacle attachment by narrowing the window of recruitment opportunity in a region. Activities of mass nesting may also increase barnacle removal by dislodgement (Robinson et al. 2019). But, whether olive ridley turtles carry more barnacles in solitary versus mass nesting populations has not been investigated.

Geographic overlap

The geographic distributions of sea turtles and barnacles overlap but not exactly. Most sea turtles and many of the barnacles occur worldwide but with gaps for various members of each in some regions. In general, barnacles exhibit a greater degree of localization than sea turtles, there being only two species of highly regional turtles (Kemp's ridley and flatback sea turtles) and perhaps six species of regional barnacles. Thus, presenting barnacle incidence rates at large scales for some species is not meaningful. For instance, *S. transversa*, exclusive to green sea turtles, is reported both from the Atlantic and Pacific; but, in the Pacific it has only been reported

from localities in the western Pacific and Indo-Pacific (Nilsson-Cantell 1930; Monroe and Limpus 1979; Hayashi 2012) and in the Atlantic only from Paraíba State in Brazil (Young 1991). A similarly disjunct and enigmatic Atlantic-Pacific distribution has been found among genotypes of S. elegans (Pinou et al. 2013) that cannot be simply explained by linkage through the Panama canal which lies 26 m above sea level and extends 82 km through freshwater Gatun Lake. Barnacle distributions, in addition to being patchy in space, can also be uneven in density, spread perhaps over large areas in generally low abundance but present in high levels at certain locations. For example, C. darwiniana is present in the western Atlantic in low numbers and from both sides of the Pacific associated with several hosts but present on virtually 100% of nesting green sea turtles in the Galapagos Islands (Zullo 1991). Cylindrolepas sinica on the other hand appears localized to a single area, perhaps just a few islands, of the South China Sea (Ren 1980). Yet others, while not abundant, are widely but discontinuously distributed, in particular S. muricata globally (Frick et al. 2011) and C. cheloniae in the Indo-Pacific (Monroe and Limpus 1979 [as T. cheloniae]; Nolte et al. 2020). The dual dispersion experienced by epizoic barnacles, distributed first as larvae in the plankton for several weeks then as adults transported by their hosts, may account for some of these discontinuities.

Evolutionary patterns

The evolutionary route taken by barnacles in specializing on sea turtles is not entirely clear. Pairwise associations between host and symbiont do not conform to phylogenetic affiliations among the turtles and especially not among the barnacles, as evidenced by scattered patterns in hierarchical clustering of occurrence data (Fig. 3). The leatherback sea turtle hosts the majority of highly selective barnacles which, however, belong to two platylepadid genera that are widely spread across all sea turtles. Though fossil remains suggest chelonibiid barnacles are basal, there is evidence that platylepadids preceded them (Hayashi et al. 2013) and radiated to all turtle hosts. In spite of historical legacies, the underlying template for barnacle/sea turtle associations may be driven by ecology more than phylogeny. Differences in diet and habitat of the hosts may affect barnacle recruitment and survival. Life on a benthic-feeding omnivore that disrupts the benthos while foraging may amplify food quantity and quality for barnacles compared to an herbivorous host foraging in sea grass meadows or a pelagic feeder in the open sea. But which host feeding-mode is most optimal for barnacles is debatable. Herbivorous green sea turtles host the greatest diversity of barnacles whereas omnivorous loggerheads may host the highest barnacle loads and leatherbacks the most widespread species. If narrow host use is a clue, investigating the biology of barnacles tending toward specialization might provide further insight.

Knowledge gaps

Many unknowns remain concerning the association of barnacles with sea turtles. A more complete picture requires further study of both symbionts and hosts, but our knowledge gap of the former is probably larger. To advance understanding, several areas deserve greater attention and exploration. Barnacle shell growth rates, to-date studied only for C. testudinaria (Sloan et al. 2014; Doell et al. 2017), could provide insight on short-term or seasonal movements of sea turtles. The idea of harnessing barnacles to track host movements and trace population boundaries has been explored in California gray whales (Killingley 1980) and sea turtles (Killingley and Lutcavage 1983; Detjen et al. 2015; Pearson et al. 2019, 2020), using isotopic analysis of barnacle shells to infer salinity and temperature signatures of the water inhabited by their hosts over time. Barnacle age structure and infestation rates have also been tested as geospatial indicators for dolphins (Di Beneditto and Ramos 2000). A deeper knowledge of shell growth for other barnacles and the natural history of larval dispersal for any species could further progress. It would be highly valuable to know how the supply of barnacle larvae varies in space a time and could, for instance, help explain the anomaly of different species of Chelonibia on hawksbill turtles found across a relatively short distance in the Persian Gulf of Iran (compare Devin and Sadeghi 2010 to Razaghian et al. 2019). Indeed, where barnacle larvae are distributed and make their rendezvous with sea turtles is perhaps the most crucial and unresolved aspect of the association. Assessing suites of barnacles present on sea turtles might also offer a community level "barnacle fingerprint" mechanism for matching snapshots of epibiont diversity to regional patterns. So might also assaying the distribution of barnacle population genotypes among sea turtles. How barnacle presence varies seasonally or with ontogenetic stages or between genders has also not been thoroughly explored. Potential ecological succession of barnacles has been suggested between home-range and migrating Kemp's ridleys (Lutcavage and Musick 1985),

newly recruiting loggerheads (Limpus and Limpus 2003), and across a nesting interval for female loggerhead sea turtles (Frick et al. 2002). It would also be valuable to examine attachment mode among barnacles and ascertain, as with *S. elegans* (compare Lazo-Wasem et al. 2011 with Pinou et al. 2013), how it may vary with host species.

On the sea turtle side of the association, a lack of comparative information on scute growth, scute shedding, and epidermal turnover, and the impact of these on barnacle attachment, presents a major gap in understanding. As a general model, scutes of turtles grow along their margins and from beneath while shedding layers superficially (Zangerl 1969). But differences in details among host species may influence adaptations in barnacles. Hawksbill sea turtles for instance add new material along the anterior margin of the scutes while material at the posterior edge is eroded but not sloughed and the scutes grow continuously thicker from beneath with age (Tucker et al. 2001). In contrast, loggerhead and green turtles maintain relatively thin scutes by sloughing outer layers at indeterminate rates (López-Castro et al. 2014). As I have observed, the former sheds continuously in flakes or pieces and the latter periodically in large thin sheets. But lacking more detailed characterization of scute growth and shedding, how barnacles respond to different circumstances cannot at present be elucidated.

Results of this study are inevitably constrained by certain limitations and caveats of the data. Information on the occurrence of barnacles on sea turtles comes primarily from nesting females, thus patterns may vary for pelagic individuals, including males and juveniles, which are less easily sampled. Records are also not distributed equally across host species, thus uneven effort may have generated artificial differences between species. That barnacles are not mentioned in many reports of sea turtles does not necessarily indicate their absence, while their presence on the other hand, when recorded, cannot be understood as a complete inventory of the barnacle diversity present, unless stated otherwise in either case. It is likely that greater attention has been paid to the obvious, shell-cementing Chelonibia barnacles. Easy-to-miss species of the skin and those in hardto-reach places such as the plastron, base of limbs, or mouth, are probably under-represented in many records. A large-scale and consistent lack of reports of some barnacles in some locations of the world or with some turtle hosts likely reflects actual distributions but artifacts of underreporting always remain a possible concern, especially for uncommon species. Another significant assumption is that the turtles,

and more especially the barnacles, are correctly reported. In this study, corrections were applied for the analysis where apparent. In particular, one common assumption was that *Platylepas* barnacles on leatherbacks, often reported as *P. hexastylos*, were scored as *P. coriacea*. If incorrect, this assumption overinflates the specificity of these species of barnacles.

In summary, the association of epizoic coronuloid barnacles with sea turtles is both stringent and flexible; narrowly obligate to sea turtles but not tightly linked with single species of hosts except for a few cases. The barnacles are also more taxonomically diverse and geographically subdivided than their chelonian partners, a fact which could be utilized to give insight on where sea turtles travel and how they operate in the environment by tracing various distributions of their barnacle epifauna. The sea turtle/epibiont relationships described herein also offer a valuable baseline for monitoring redistributions of hosts and symbionts in a time of changing climate.

Acknowledgments

A key element in executing this study was gathering a large number of obscure and unconventional documents for which I was ably assisted by the indefatigable efforts of Pamela King of The Citadel's interlibrary loan department. I also gratefully acknowledge valuable discussions with Nathan Robinson about the habits of sea turtles and his thoughts on the occurrence of their epibionts. I am indebted to my colleague Danny Gustafson for proposing the idea of the hierarchical analysis of barnacle occurrence and providing assistance in presenting it graphically. I also wish to extend my deep appreciation to George Balazs and the late Bill Newman, both world experts on sea turtles and barnacles, respectively, who greatly encouraged and aided my early interest in epizoic barnacles.

Supplementary data

Supplementary data are available at *IOB* online.

Declaration of competing interest

The author declares no competing interests.

References

- Al-Yahya HAH, Chen HN, Chan BKK, Kado R, Høeg JT. 2016. Morphology of cyprid attachment organs compared across disparate barnacle taxa: does it relate to habitat? Biol Bull 231:120–9.
- Alibardi L. 2005. Proliferation in the epidermis of chelonians and growth of the horny scutes. J Morphol 265:52–69.

Anderson DT. 1994. Barnacles: structure, function, development and evolution. London: Chapman and Hall.

- Badrudeen M. (2000). On the occurrence of the cirriped barnacle, *Chelonibia patula* (Ranzani) on the sea snake, *Hydrophis cyanocintus* (Daudin). Marine Fisheries Information Service, Technical and Extension Series 164:25.
- Balbuena J, Raga JA. 1991. Ecology and host relationships of the whale-louse *Isocyamus delphini* (Amphipoda: Cyamidae) parasitizing long-finned pilot whales (*Globicephala melas*) off the Faroe Islands (Northeast Atlantic). Can J Zool 69:141–5.
- Beaumont ES, Zárate P, Zardus JD, Dutton PH, Seminoff JA. (2008). Epibiont occurrence in Galapagos green turtles (*Chelonia mydas*) at nesting and feeding grounds. NOAA Technical Memorandum, NMFS-SEFSC-569.
- Blick JP, Zardus JD, Dvoracek D. 2010. The sea turtle barnacle, *Chelonibia testudinaria* (Cirripedia: Balanomorpha: Coronuloidea), from pre-Columbian deposits on San Salvador, Bahamas. Caribbean J Sci 46:228–39.
- Block RJ, Bolling D. 1939. The amino acid composition of keratins: the composition of gorgonin, spongin, turtle scutes, and other keratins. J Biol Chem 127:685–93.
- Bowen BW, Karl SA. 2007. Population genetics and phylogeography of sea turtles. Mol Ecol 16:4886–907.
- Bowen BW, Nelson WS, Avise JC. 1993. A molecular phylogeny for marine turtles: trait mapping, rate assessment, and conservation relevance. Proc Natl Acad Sci USA 90:5574–7.
- Bugoni L, Krause L, Oliveira De Almeida A, Angélica De Pádua Bueno A. 2001. Commensal barnacles of sea turtles in Brazil. Marine Turtle Newslett 94:7–9.
- Cadena EA, Parham JF. 2015. Oldest known marine turtle? A new protostegid from the Lower Cretaceous of Colombia. Paleo Bios 32:1–42.
- Carr AF, Caldwell DE. 1956. The ecology and migrations of sea turtles, 1 results of field work in Florida, 1955. Am Museum Novitates 1793:1–23.
- Chan BKK, Tsang LM, Ng WC, Williams GA, Chu KH. 2012. Biogeography of the widespread intertidal barnacle *Chthamalus malayensis* in Indo-Pacific waters: the interplay of geological history, contemporary ocean circulation patterns and habitat specificity. Integr Compar Biol 52:E27.
- Collareta A, Bosselaers M, Bianucci G. 2016. Jumping from turtles to whales: a Pliocene fossil record depicts an ancient dispersal of *Chelonibia* on mysticetes. Riv Italian Paleontol Stratigrafia 122:35–44.
- Collareta A, Reitano A, Rosso A, Sanfilippo R, Bosselaers M, Bianucci G, Insacco G. 2019. The oldest platylepadid turtle barnacle (Cirripedia, Coronuloidea): a new species of *Platylepas* from the Lower Pleistocene of Italy. Eur J Taxonomy 516:1–17.
- Covelo P, Nicolau L, López A. 2016. Four new records of stranded Kemp's ridley turtle *Lepidochelys kempii* in the NW Iberian Peninsula. Mar Biodiver Rec 9:80.
- Crozier WJ. 1916. On a barnacle, Conchoderma virgatum, attached to a fish, Diodon hystrix. Am Nat 50:636–9.
- Daniel A. 1958. On *Platylepas indicus* n. sp. A new barnacle from the Madras coast of India. Ann Mag Nat Hist 1:755–7.
- Darwin CR. 1854. A monograph on the sub-class Cirripedia, with figures of all the species. The Balanidae, (or sessile cirripedes); the Verrucidae, etc. London: The Ray Society.

- Detjen M, Sterling E, Gómez A. 2015. Stable isotopes in barnacles as a tool to understand green sea turtle (*Chelonia mydas*) regional movement patterns. Biogeosci Discus 12:4655–69.
- Devin ML, Sadeghi P. 2010. Barnacles on hawksbill sea turtles, *Eretmochelys imbricata*, in Hormoz Island, Iran (Reptilia: Cheloniidae). Zool Middle East 49:45–8.
- Di Beneditto APM, Ramos RMA. 2000. Records of the barnacle *Xenobalanus globicipitis* (Steenstrup, 1851) on small cetaceans of Brazil. Biotemas 13:159–65.
- Doell SA, Connolly RM, Limpus CJ, Pearson RM, Van De Merwe JP. 2017. Using growth rates to estimate age of the sea turtle barnacle *Chelonibia testudinaria*. Mar Biol 164:222.
- Dornfeld TC, Robinson NJ, Santidrián Tomillo P, Paladino FV. 2015. Ecology of solitary nesting olive ridley sea turtles at Playa Grande, Costa Rica. Mar Biol 162:123–39.
- Dreyer N, Zardus JD, Høeg JT, Olesen JM, Yu MC, Chan BKK. 2020. How dolphin barnacles attach to their hosts and the paradox of remarkably versatile attachment structures in cypris larvae. Organ Divers Evol 20:233–49.10.1007/s13127-020-00434-3
- Dutton PH, Davis SK, Guerra T, Owens DW. 1996. Molecular phylogeny for marine turtles based on sequences of the ND4-leucine tRNA and control regions of mitochondrial DNA. Mol Phylogenet Evol 5:511–21.
- Eckert KL, Eckert SA. 1987. Growth rate and reproductive condition of the barnacle *Conchoderma virgatum* on gravid leatherback sea turtles in Caribbean waters. J Crustacean Biol 7:682–90.
- Eckert KL, Eckert SA. 1988. Pre-reproductive movements of leatherback sea turtles (*Dermochelys coriacea*) nesting in the Caribbean. Copeia 1988:400–6.
- Edmondson CH, Ingram WM. 1939. Fouling organisms in Hawaii. Occasion Paper Bishop Museum 14:251–300.
- Evers SW, Benson RBJ. 2019. A new phylogenetic hypothesis of turtles with implications for the timing and number of evolutionary transitions to marine lifestyles in the group. Paleontology 62:93–134.
- Ewers-Saucedo C, Chan BKK, Zardus JD, Wares JP. 2017. Parallel patterns of host-specific morphology and genetic admixture in sister lineages of a commensal barnacle. Biol Bull 232:171–85.
- Fernando SA. 2006. Monograph on Indian barnacles. Kochi, India: Ocean Science & Technology Cell, Cochin University of Science & Technology.
- Flint M, Patterson-Kane JC, Limpus CJ, Work TM, Blair D, Mills PC. 2009. Postmortem diagnostic investigation of disease in free-ranging marine turtle populations: a review of common pathologic findings and protocols. J Vet Diagnost Invest 21:733–59.
- Foster BA. 1987. Barnacle ecology and adaptation. In: Southward AJ, editor. Barnacle biology Rotterdam, Netherlands: A.A. Balkema. p. 113–34.
- Frazier JG, Margaritoulis D. 1990. The occurrence of the barnacle, *Chelonibia patula* (Ranzani, 1818) on an inanimate substratum (Cirripedia, Thoracica). Crustaceana 59:213–8.
- Frick MG, Pfaller JB. 2013. Sea turtle epibiosis. In: Wyneken J, Lohmann KJ, Musick JA, editors. The biology of sea turtles. Boca Raton (FL): CRC Press. p. 399–426.

- Frick MG, Williams KL, Markesteyn EJ, Pfaller JB, Frick RE. 2004. New records and observations of epibionts from loggerhead sea turtles. Southeast Nat 3:613–20.
- Frick MG, Williams KL, Robinson M. 1998. Epibionts associated with nesting loggerhead sea turtles (*Caretta caretta*) in Georgia, USA. Herpetol Rev 29:211–4.
- Frick MG, Williams KL, Veljacic DC, Jackson JA, Knight SE. (2002). Epibiont community succession on nesting loggerhead sea turtles, *Caretta caretta*, from Georgia, USA. NOAA Technical Memorandum, NMFS-SEFSC-477.
- Frick MG, Zardus JD, Ross A, Senko J, Montano-Valdez D, Bucio-Pacheco M, Sosa-Cornejo I. 2011. Novel records and observations of the barnacle *Stephanolepas muricata* (Cirripedia: Balanomorpha: Coronuloidea); including a case for chemical mediation in turtle and whale barnacles. J Nat Hist 45:629–40.
- Green D. (1998). Epizoites of Galapagos green turtles. NOAA Technical Memorandum, NMFS-SEFSC-412.
- Gruvel A. 1905. Monographie des cirrhipèdes ou thécostracés. Paris, France: Masson et Cie.
- Grygier MJ, Newman WA. 1991. A new genus and two new species of Microlepadidae (Cirripedia: Pedunculata) found on western Pacific diadematid echinoids. Galaxea 10:1–22.
- Hadfield MG. 2011. Biofilms and marine invertebrate larvae: what bacteria produce that larvae use to choose settlement sites. Ann Rev Mar Sci 3:453–70.
- Harzhauser M, Newman WA, Grunert P. 2011. A new Early Miocene barnacle lineage and the roots of sea-turtle fouling Chelonibiidae (Cirripedia, Balanomorpha). J Syst Palaeontol 9:473–80.
- Hayashi R. 2009. New host records of the turtle barnacle, *Cylindrolepas sinica*: a case study of sea turtles' behaviour and their epibionts. Mar Biodivers Rec 2:1–4.
- Hayashi R. 2012. Atlas of the barnacles on marine vertebrates in Japanese waters including taxonomic review of superfamily Coronuloidea (Cirripedia: Thoracica). J Mar Biol Assoc UK 92:107–27.
- Hayashi R. 2013a. A checklist of turtle and whale barnacles (Cirripedia: Thoracica: Coronuloidea). J Mar Biol Assoc UK 93:143–82.
- Hayashi R. 2013b. Intraspecific variation in the turtle barnacle, *Cylindrolepas sinica* Ren, 1980 (Cirripedia, Thoracica, Coronuloidea), with brief notes on habitat selectivity. ZooKeys 327:35–42.
- Hayashi R. 2014. Past biodiversity: historical Japanese illustrations document the distribution of whales and their epibiotic barnacles. Ecol Indicat 45:687–91.
- Hayashi R. 2017. First documentation of the barnacle Fistulobalanus albicostatus (Cirripedia: Balanomorpha) as an epibiont of loggerhead sea turtle Caretta caretta. Mar Biodivers 47:157–8.
- Hayashi R, Chan BKK, Simon-Blecher N, Watanabe H, Guy-Haim T, Yonezawa T, Levy Y, Shuto T, Achituv Y. 2013. Phylogenetic position and evolutionary history of the turtle and whale barnacles. Mol Phylogenet Evol 67:9–14.
- Herbert LH, Jacobson ER. 1995. Diseases of marine turtles. In: Bjorndal KA, editor. Biology and conservation of sea turtles. Washington (DC): Smithsonian Institution Press. p. 593-6
- Hughes GR. 1970. Further studies on marine turtles in Tongaland. III. Lammergeyer 12:7–25.

Hughes GR, Bass AJ, Mentis MT. 1967. Further studies on marine turtles in Tongaland. I. Lammergeyer 3:5–54.

- Kaliszewska ZA, Seger JON, Rowntree VJ, Barco SG, Benegas R, Best PB, Brown MW, Brownell RL, Carribero A, Harcourt R, et al. 2005. Population histories of right whales (Cetacea: *Eubalaena*) inferred from mitochondrial sequence diversities and divergences of their whale lice (Amphipoda: *Cyamus*). Mol Ecol 14:3439–56.
- Kardong KV. 2019. Vertebrates: comparative anatomy, function, evolution. 8th ed. New York (NY): McGraw-Hill.
- Killingley JS. 1980. Migrations of California gray whales tracked by oxygen-18 variations in their epizoic barnacles. Science 207:759–60.
- Killingley JS, Lutcavage M. 1983. Loggerhead turtle movements reconstructed from ¹⁸O and ¹³C profiles from commensal barnacle shells. Estuarin Coast Shelf Sci 16:345–9.
- Kitsos MS, Christodoulou M, Arvanitidis C, Mavidis M, Kirmitzoglou I, Koukouras A. 2005. Composition of the organismic assemblage associated with *Caretta caretta*. J Mar Biol Assoc UK 85:257–61.
- Kobayashi M. 2000. Relationship between brightness or size and the presence of barnacles on the carapace of the hawksbill turtles (*Eretmochelys imbricata*). Curr Herpetol 19:91–6.
- Kolbasov GA, Zevina GB. 1999. A new species of *Paralepas* (Cirripedia: Heteralepadidae) symbiotic with *Xenophora* (Mollusca: Gastropoda); with the first complemental male known for the family. Bull Mar Sci 64:391–8.
- Korschelt E. 1933. Über zwei parasitäre Cirripedien, *Chelonibia* und *Dendrogaster*, nebst angaben über die beziehungen der Balanomorphen zu ihrer unterlage. Zool Jahrbüch Abteil Syst Geogr Biol Tiere 64:1–40.
- Lanchester WF. 1902. On the crustacea collected during the "Skeat expedition" to the Malay Peninsula. Part II Anomura, Cirripedia and Isopoda. Proc Zool Soc Lond 2:371–2.
- Landman NH, Saunders WB, Winston JE, Harries PJ. 1987. Incidence and kinds of epizoans on the shells of live *Nautilus*. In: Saunders WB, Landman NH, editors. Nautilus: the biology and paleobiology of a living fossil. New York (NY): Plenum Press. p. 163–79.
- Lazo-Wasem EA, Pinou T, Peña De Niz A, Feuerstein A. 2011. Epibionts associated with the nesting marine turtles *Lepidochelys olivacea* and *Chelonia mydas* in Jalisco, Mexico: a review and field guide. Bull Peabody Museum Nat Hist 52:221–40.
- Limpus CJ, Limpus DJ. 2003. Biology of the loggerhead turtle in western South Pacific Ocean foraging areas. In: Bolten AB, Witherington B, editors. Biology and conservation of the loggerhead sea turtle. Washington (DC): Smithsonian Institution Press. p. 93–113.
- Limpus CJ, Parmenter CJ, Baker V, Fleay A. 1983. The Crab Island sea turtle rookery in the north-eastern gulf of Carpentaria. Austral Wildlife Res 10:173–84.
- Linnaeus C. 1758. Systema naturae per regna tria naturae: secundum classes, ordines, genera, species, cum characteribus, differentiis, synonymis, locis. 10th ed. Stockholm, Sweden: Laurentii Salvii.
- Liu JCW, Høeg JT, Chan BKK. 2016. How do coral barnacles start their life in their hosts? Biol Lett 12:20160124.

- López-Castro M, Bjorndal KA, Bolten AB. 2014. Evaluation of scute thickness to infer life history records in the carapace of green and loggerhead turtles. Endanger Species Res 24:191–6.
- Lutcavage M, Musick JA. 1985. Aspects of the biology of sea turtles in Virginia. Copeia 1985:449–56.
- Marloth R. 1900. Notes on the mode of growth of *Tubcinella trachaelis*, the barnacle of the southern right whale. Trans South African Philos Soc 11:1–6.
- Miranda L, Moreno RA. 2002. Epibiontes de *Lepidochelys olivacea* (Eschscholtz, 1829) (Reptilia: Testudinata: Cheloniidae) en la región centro sur de Chile. Rev Biol Mar Oceanografia 37:145–6.
- Molenock J, Gomez ED. 1972. Larval stages and settlement of the barnacle *Balanus* (Conopea) *galeatus* (L.) (Cirripedia Thoracica). Crustaceana 23:100–8.
- Monroe R, Limpus CJ. 1979. Barnacles on turtles in Queensland waters with descriptions of three new species. Memoirs Queensland Museum19:197–223.
- Moyse J. 1961. The larval stages of *Acasta spongites* and *Pyrgoma anglicum* (Cirripedia). Proc Zool Soc Lond 137:371–92.
- Naro-Maciel E, Le M, Fitzsimmons NN, Amato G. 2008. Evolutionary relationships of marine turtles: a molecular phylogeny based on nuclear and mitochondrial genes. Mol Phylogenet Evol 49:659–62.
- Newman WA, Ross A. 1976. Revision of the balanomorph barnacles; including a catalog of the species. *Memoirs of the San Diego Society of*. Nat Hist 9:1–108.
- Nifong JC, Frick MG. 2011. First record of the American alligator (*Alligator mississippiensis*) as a host to the sea turtle barnacle (*Chelonibia testudinaria*). Southeast Nat 10:557–60.
- Nilsson-Cantell CA. 1930. Diagnoses of some new cirripedes from the Netherlands indies collected by the expedition of his royal highness the prince leopold of Belgium in 1929. Bull Musé Royal Hist Nat Belgique 6:1–2.
- Nogata Y, Matsumura K. 2006. Larval development and settlement of a whale barnacle. Biol Lett 2:92–3.
- Nolte CR, Nel R, Pfaff MC. 2020. Determining body condition of nesting loggerhead sea turtles (*Caretta caretta*) in the south-west Indian Ocean. J Mar Biol Assoc UK 100:291–9.
- Ortiz M, Lalana R, Varela C. 2004. Caso extremo de epibiosis de escaramujos (Cirripedia: Balanomorpha), sobre una esquila (Hoplocarida: Stomatopoda), en Cuba. Rev Investigaciones Mar 25:75–6.
- Pagès F. 2000. Biological associations between barnacles and jellyfish with emphasis on the exoparasitism of *Alepas pacifica* (Lepadomorpha) on *Diplulmaris malayensis* (Scyphozoa). J Nat Hist 34:2045–56.
- Pearson RM, Van De Merwe JP, Connolly RM. 2020. Global oxygen isoscapes for barnacle shells: application for tracing movement in oceans. Sci Total Environ 705:135782.
- Pearson RM, Van De Merwe JP, Gagan MK, Limpus CJ, Connolly RM. 2019. Distinguishing between sea turtle foraging areas using stable isotopes from commensal barnacle shells. Sci Rep 9:6565.
- Pérez-Losada M, Høeg JT, Crandall KA. 2004. Unraveling the evolutionary radiation of the thoracican barnacles using molecular and morphological evidence: a comparison of

- several divergence time estimation approaches. Syst Biol 53:244–64.
- Pérez-Losada M, Høeg JT, Simon-Blecher N, Achituv Y, Jones DS, Crandall KA. 2014. Molecular phylogeny, systematics and morphological evolution of the acorn barnacles (Thoracica: Sessilia: Balanomorpha). Mol Phylogenet Evol 81:147–58.
- Pilsbry HA. 1907. The barnacles (Cirripedia) contained in the collections of the U.S. National Museum. Bull US Natl Museum 60:1–122.
- Pilsbry HA. 1910. *Stomatolepas*, a barnacle commensal in the throat of the loggerhead turtle. Amn Nat 44:304–6.
- Pilsbry HA. 1916. The sessile barnacles (Cirripedia) contained in the collections of the U.S. National Museum; including a monograph of the American species. Bull US Natl Museum 93:1–366.
- Pinou T, Lazo-Wasem EA, Dion K, Zardus JD. 2013. Six degrees of separation in barnacles? Assessing genetic variability in the sea-turtle epibiont *Stomatolepas elegans* (Costa) among turtles, beaches, and oceans. J Nat Hist 47:2193–212.
- Razaghian H, Esfandabad BS, Hesni MA, Shoushtari RV, Toranjzar H, Miller J. 2019. Distribution patterns of epibiotic barnacles on the hawksbill turtle, *Eretmochelys imbricata*, nesting in Iran. Region Stud Mar Sci 27:100527–6.
- Rees DJ, Noever C, Finucci B, Schnabel K, Leslie RE, Drewery J, Theil Bergum HO, Dutilloy A, Glenner H. 2019. *De novo* innovation allows shark parasitism and global expansion of the barnacle *Anelasma squalicola*. Curr Biol 29:R562–3.
- Reisinger RR, Bester MN. 2010. Goose barnacles on seals and a penguin at Gough Island. Afr Zool 45:129–32.
- Ren X. 1980. Turtle barnacles of the Xisha islands, Guangdong province, China. Stud Mar Sin 17:187–97.
- Robinson NJ, Lazo-Wasem E, Butler BO, Lazo-Wasem EA, Zardus JD, Pinou T. 2019. Spatial distribution of epibionts on olive ridley sea turtles at Playa Ostional, Costa Rica. PLoS One 14:e0218838.
- Robinson NJ, Gatto C, Figgener C, Lazo-Wasem EA, Paladino FV, Tomillo PS, Zardus JD, Pinou T. 2017a. Assessing potential limitations when characterising the epibiota of marine megafauna: effect of gender, sampling location, and inter-annual variation on the epibiont communities of olive ridley sea turtles. J Exp Mar Biol Ecol 497:71–7.
- Robinson NJ, Lazo-Wasem EA, Paladino FV, Zardus JD, Pinou T. 2017b. Assortative epibiosis on leatherback, olive ridley, and east Pacific green turtles in the Eastern Tropical Pacific. J Mar Biol Assoc UK 97:1233–40.
- Ross A, Jackson CG. Jr. 1972. Barnacle fouling of the ornate diamondback terrapin, *malaclemys terrapin macrospilota*. Crustaceana 22:203–5.
- Ross A, Newman WA. 1967. Eocene Balanidae of Florida, including a new genus and species with a unique plan of "turtle-barnacle" organization. Am Museum Novitates 2288:1–21.
- Ryder JA. 1879. Strange habitat of a barnacle on a gar pike. Am Nat 13:453.
- Scarff JE. 1986. Occurrence of the barnacles *Coronula diadema*, *C. Reginae* and *Cetopirus complanatus* (Cirripedia) on right whales. Sci Rep Whales Res Inst 37:129–53.

- Sloan K, Zardus JD, Jones ML. 2014. Substratum fidelity and early growth in *Chelonibia testudinaria*, a turtle barnacle especially common on debilitated loggerhead (*Caretta care*tta) sea turtles. Bull Mar Sci 90:581–97.
- Solomon SE, Hendrickson JR, Hendrickson LP. 1986. The structure of the carapace and plastron of juvenile turtles, *Chelonia mydas* (the green turtle) and *Caretta caretta* (the loggerhead turtle). J Anat 145:123–31.
- Spotila JR. 2004. Sea turtles: a complete guide to their biology, behavior, and conservation. Baltimore (MD): Johns Hopkins University Press.
- Sumner FB, Osburn RC, Cole LJ. 1913. A biological survey of the waters of Woods Hole and vicinity, section III. A catalogue of the marine fauna. Bull US Bureau Fish 31:545–794.
- Tachikawa H. 1995. Notes on three species of stalked barnacles found from a turtle barnacle on the carapace of a green turtle, *Chelonia mydas*. Nanki Seibutu 37:67–8.
- Tseng CT, Leu JH, Cheng IJ. 2018. On the genetic diversity of two species of the genus *Ozobranchus* (Hirudinida; Ozobranchidae) from the Atlantic and Pacific oceans. J Mar Biol Assoc UK 98:955–60.
- Tucker AD, Broderick D, Kampe L. 2001. Age estimation of *Eretmochelys imbricata* by schlerochronology of carapacial scutes. Chelon Conservat Biol 4:219–22.
- van Beneden PJ. 1876. Animal parasites and messmates. New York (NY): Appleton and Co.
- Wahl M, Mark O. 1999. The predominately facultative nature of epibiosis: experimental and observational evidence. Mar Ecol Prog Ser 187:59–66.
- Wallace BP, DiMatteo AD, Hurley BJ, Finkbeiner EM, Bolten AB, Chaloupka MY, Hutchinson BJ, Abreu-Grobois FA, Amorocho D, Bjorndal KA, et al. 2010. Regional management units for marine turtles: a novel framework for prioritizing conservation and research across multiple scales. PLoS One 5:e15465.
- Wares JP. 2020. Small, flat, and cryptic: diversity in Chthamalidae as context for global study of marine biogeography. J Crustac Biol 40:1–16.
- Weldon PJ, Mason RT, Tanner MJ, Eisner T. 1990. Lipids in the Rathke's gland secretions of Kemp's ridley sea turtle (*Lepidochelys kempii*). Compar Biochem Physiol 96:705–8.
- White PS, Morran LT, De Roode JC. 2017. Phoresy. Curr Biol 27:R578–91.
- Young PS. 1991. The superfamily Coronuloidea Leach (Cirripedia, Balanomorpha) from the Brazilian coast, with redescription of *Stomatolepas* species. Crustaceana 61:190–212.
- Zangerl R. 1969. The turtle shell. In: Gans C, editor. Biology of the reptilia. New York (NY): Academic Press. p. 311–39.
- Zann LP. 1975. Biology of a barnacle (*Platylepas ophiophilus* Lanchester) symbiotic with sea snakes. In: Dunson WA, editor. The biology of sea snakes. Baltimore (MD): University Park Press. p. 267–86.
- Zapalski MK. 2011. Is absence of proof a proof of absence? Comments on commensalism. Palaeogeogr Palaeoclimatol Palaeoecol 302:484–8.
- Zardus JD, Balazs GH. 2007. Two previously unreported barnacles commensal with the green sea turtle, *Chelonia mydas* (Linnaeus, 1758), in Hawaii and a comparison of their attachment modes. Crustaceana 80:1303–15.

- Zardus JD, Hadfield MG. 2004. Larval development and complemental males in *Chelonibia testudinaria*, a barnacle commensal with sea turtles. J Crustac Biol 24:409–21.
- Zardus JD, Nedved BT, Huang Y, Tran C, Hadfield MG. 2008. Microbial biofilms facilitate adhesion in biofouling invertebrates. Biol Bull 214:91–8.
- Zardus JD, Lake DT, Frick MG, Rawson PD. 2014. Deconstructing an assemblage of 'turtle' barnacles: species
- assignments and fickle fidelity in *Chelonibia*. Mar Biol 161:45–59.
- Zullo VA. 1991. Zoogeography of the shallow-water cirriped fauna of the Galápagos Islands and adjacent regions in the Tropical Eastern Pacific. In: James MJ, editor. Galápagos marine invertebrates: taxonomy, biogeography, and evolution in Darwin's islands. New York (NY): Plenum Press. p. 173–92.