







Article

A Non-Invasive Neonatal Signature Predicts Later Development of Atopic Diseases

Youssef Sereme ^{1,2}, Moïse Michel ^{1,2,3}, Soraya Mezouar ^{1,2}, Cheick Oumar Guindo ^{1,2}, Lanceï Kaba ^{1,4}, Ghiles Grine ^{1,2,5}, Thibault Mura ^{6,7}, Jean-Louis Mège ^{1,2}, Tu Anh Tran ^{8,9,10}, Pierre Corbeau ^{3,10,11,*}, Anne Filleron ^{8,9,10,*} and Joana Vitte ^{1,2,10,12,*}

- ¹ IHU Méditerranée Infection, 13005 Marseille, France; seremeyoussouf@yahoo.fr (Y.S.); moise0michel@gmail.com (M.M.); soraya.mezouar@univ-amu.fr (S.M.); cheicko86@gmail.com (C.O.G.); lancekaba@yahoo.fr (L.K.); grineghiles@gmail.com (G.G.); jean-louis.mege@univ-amu.fr (J.-L.M.)
 - ² IRD, APHM, MEPHI, Aix-Marseille Université, 13284 Marseille, France
 - ³ Immunology Department, University Hospital Nîmes, 30900 Nîmes, France
 - ⁴ IRD, AP-HM, SSA, VITROME, Aix-Marseille Université, 13284 Marseille, France
 - ⁵ UFR Odontologie, Aix-Marseille Université, 13284 Marseille, France
 - ⁶ INSERM, University of Montpellier, U1061, Neuropsychiatry: Epidemiological and Clinical Research, 34093 Montpellier, France; thibault.mura@chu-nimes.fr
 - ⁷ Laboratoire de Biostatistique, Epidémiologie Clinique, Santé Publique Innovation et Méthodologie (BESPIM), Groupe Hospitalier Caremeau, CHU de Nîmes, Nîmes University Hospital, 30900 Nîmes, France
 - ⁸ Paediatrics Department, University Hospital Nîmes, 30900 Nîmes, France; tu.anh.tran@chu-nimes.fr
 - ⁹ INSERM U1183, Institute for Regenerative Medicine & Biotherapy, 34295 Montpellier, France
 - ¹⁰ Faculty de Medicine, Montpellier University, 34000 Montpellier, France
 - ¹¹ CNRS UMR 9002, Institute of Human Genetics, 34090 Montpellier, France
 - ¹² IDESP, INSERM UMR UA11, Institut Desbrest d'Epidémiologie et de Santé Publique (IDESP) Campus Sante, 34093 Montpellier, France
- * Correspondence: pierre.corbeau@igh.cnrs.fr (P.C.); anne.filleron@chu-nimes.fr (A.F.); joana.vitte@inserm.fr (J.V.); Tel.: +33-4-13-73-20-51 (J.V.); Fax: +33-4-13-73-20-52 (J.V.)
- † These authors contributed equally to this work.



Citation: Sereme, Y.; Michel, M.; Mezouar, S.; Guindo, C.O.; Kaba, L.; Grine, G.; Mura, T.; Mège, J.-L.; Tran, T.A.; Corbeau, P.; et al. A Non-Invasive Neonatal Signature Predicts Later Development of Atopic Diseases. *J. Clin. Med.* **2022**, *11*, 2749. <https://doi.org/10.3390/jcm11102749>

Academic Editor: José Joaquín Cerón

Received: 19 March 2022

Accepted: 10 May 2022

Published: 12 May 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

Abstract: Background: Preterm birth is a major cause of morbidity and mortality in infants and children. Non-invasive methods for screening the neonatal immune status are lacking. Archaea, a prokaryotic life domain, comprise methanogenic species that are part of the neonatal human microbiota and contribute to early immune imprinting. However, they have not yet been characterized in preterm neonates. **Objective:** To characterize the gut immunological and methanogenic Archaeal (MA) signature in preterm neonates, using the presence or absence of atopic conditions at the age of one year as a clinical endpoint. **Methods:** Meconium and stool were collected from preterm neonates and used to develop a standardized stool preparation method for the assessment of mediators and cytokines and characterize the qPCR kinetics of gut MA. Analysis addressed the relationship between immunological biomarkers, Archaea abundance, and atopic disease at age one. **Results:** Immunoglobulin E, tryptase, calprotectin, EDN, cytokines, and MA were detectable in the meconium and later samples. Atopic conditions at age of one year were positively associated with neonatal EDN, IL-1 β , IL-10, IL-6, and MA abundance. The latter was negatively associated with neonatal EDN, IL-1 β , and IL-6. **Conclusions:** We report a non-invasive method for establishing a gut immunological and Archaeal signature in preterm neonates, predictive of atopic diseases at the age of one year.

Keywords: preterm birth; fecal mediator and cytokine; methanogenic Archaea; allergy; atopy

1. Introduction

Preterm birth, defined as delivery at fewer than 37 completed weeks of gestation, is the leading cause of neonatal mortality and morbidity and has long-term adverse health consequences [1]. The global incidence of preterm births was estimated at 10.6 in 2014, and 9.8 in 2000 [2]. The etiology of preterm birth is multifactorial and not yet fully understood [3]. However, factors related to preterm birth include maternal or fetal medical conditions, genetic and epigenetic [4] influences, environmental exposures, infertility treatments, behavioral and socioeconomic elements [3,5]. Preterm infants experience abnormal immune and metabolic programming, which might exert a lasting influence on the risk of future disease [6,7]. Preterm-born children have been shown to have immune mediator dysregulation [8], impaired innate immunity and adaptive responses characterized by reduced levels of immunoglobulin (Ig) G, opsonization and phagocytosis, and increased activation of Th1 cells compared to that of Th2 cells [9]. Cohort studies show that preterm-born children are at increased risk for preschool wheezing and school-age asthma [10], but not for food allergy [11,12] or atopic dermatitis (AD) [11,13].

During fetal life, maternal microbiota produces compounds that are transferred to the fetus and enhance the generation of innate immune cells [14]. This process is halted prematurely in preterm infants, leaving them vulnerable to disease [9]. Preterm infants have an inflammatory and hypoxic state, which has a negative impact on lung maturation, the risk of respiratory infections, and susceptibility to subsequent exposures [9,14,15]. As early as the neonatal period, the gut microbiota imprints a persistent effect on the immune system through multiple mechanisms, including the modulation of epithelial functions, the production of cytokines, and the recruitment and training of immune cells [16–18].

Archaea, considered a separate domain of life from Eukarya, giant bacteria and viruses, are part of the human microbiota [19–21]. Gut methanogenic Archaea consume hydrogen produced by bacterial fermentation, releasing methane and short chain fatty acids (SCFA) and thus taking part in the energy supply to the host [19]. They interact with the host immune system, triggering innate and adaptive immune responses, generation of specific T and B cells, and hypersensitivity responses in animals and humans [22–24]. We have shown that the neonatal gut is colonized by methanogenic Archaea from the first postnatal hours, possibly starting in utero [21,22]. Gut microbiome establishment is altered in preterm and low- birth-weight infants [25,26].

Clinical investigations and research studies in neonates, including those born before term, are usually performed with peripheral blood. However, the search for non-invasive alternatives has gained momentum in recent years [27,28].

We hypothesize the existence of an association between intestinal methanogenic Archaea and the intestinal immunological signature, understood as a pattern of immune biomarkers including cell-specific products, e.g., mast cell tryptase and eosinophil-derived neurotoxin (EDN) and major pro- and anti-inflammatory cytokines. The validation of this hypothesis would open the prospect of a predictive score for the later occurrence of immune disorders, including atopic diseases.

We addressed this question through the development of a non-invasive standardized method for the assessment of the neonatal gut immune and microbial status, implemented in a cohort of preterm infants. The aims of the present study were: (1) establish a non-invasive method adequate for the investigation of preterm neonates, (2) characterize the gut immune and Archaeal components longitudinally from birth to six weeks in the study cohort, and (3) correlate the results of gut immune and Archaeal investigations at birth and up to six weeks to the later occurrence of allergic or atopic conditions.

2. Methods

2.1. Patients and Sampling

Stool samples from 43 preterm neonates were collected without the use of preservatives at the Nimes and Montpellier University Hospitals and stored at -80°C . Samples were collected as meconium ($n = 33$) and later stool samples at 2 ($n = 33$), 4 ($n = 29$), and 6 ($n = 24$) weeks.

2.2. Ethics Statement

2.2.1. A-Immunological Analysis

Preparation of Fecal Samples

One gram of feces was solubilized in two milliliters of an in-house extraction buffer consisting of phosphate buffered saline supplemented with 4 mM 4-(2-aminoethyl)-benzenesulphonyl fluoride, 0.26 mM bestatin, 28 μM E-64, 2 μM leupeptin and 0.6 μM aprotinin, pH 7.4, and a protease inhibitor (Sigma-Aldrich, St. Louis, MN, USA [29]). The stool–buffer mixture was incubated for 20 min at room temperature, prior to centrifugation at 2000 rpm for 15 min at 4°C . The supernatant liquids were freeze-dried for 24 h, resolubilized in one milliliter of extraction buffer, and used for mediator and cytokine determination (Figure 1).

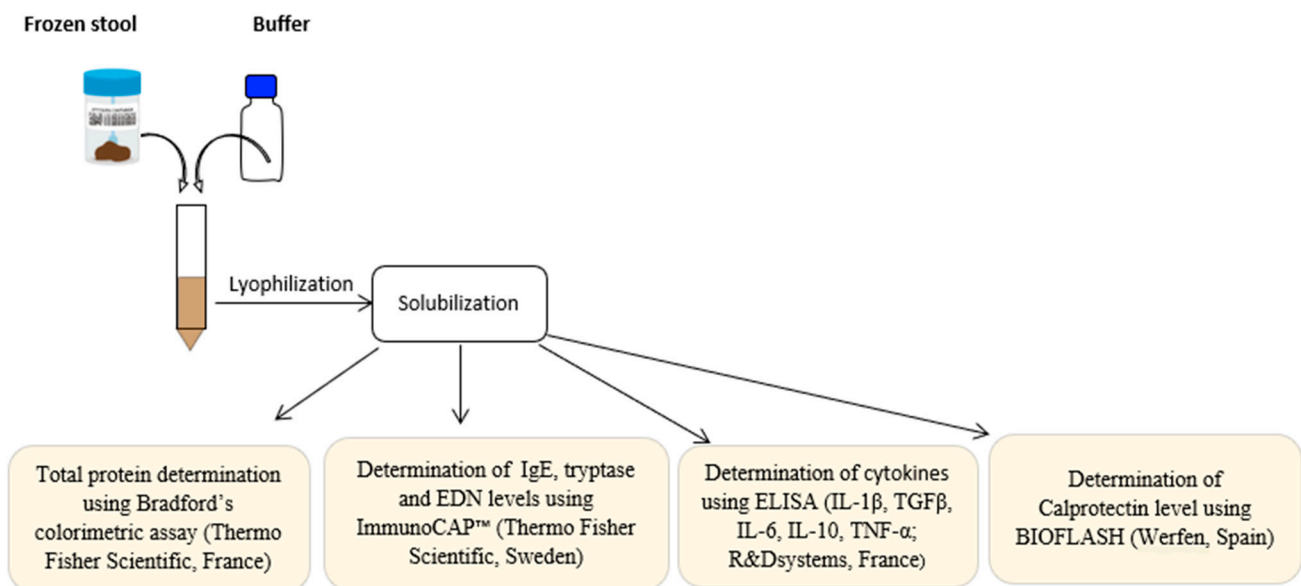


Figure 1. Fecal extraction protocol.

Total IgE, Tryptase, and EDN Determination

The concentration of total IgE, tryptase, and EDN were measured using an automated fluoro-enzymo-immunoassay with the ImmunoCAP™ 250 platform (Thermo Fisher Scientific, Uppsala, Sweden), according to ISO 15,189 standards [30]. The measurement range was 2–5000 kIU/L (4.8–12,000 $\mu\text{g}/\text{L}$) for total IgE, 1–200 $\mu\text{g}/\text{L}$ for tryptase, and 2–200 $\mu\text{g}/\text{L}$ for EDN.

Calprotectin and Total Protein Determination

Fecal calprotectin was measured using the BIOFLASH (Werfen, Barcelona, Spain) chemo-luminescent analyzer platform according to ISO 15,189 standards. Assay sensitivity was greater than 20 $\mu\text{g}/\text{mL}$. The total protein concentration of the samples was measured by the colorimetric method (BCA Protein assay, Thermo Fisher Scientific).

Immunoassays

Cytokines (IL-6, IL-10, IL-1 β , TGF- β , and TNF- α) were measured by ELISA using specific immunoassay kits according to the manufacturer's protocols (R&D systems, Minneapolis, MN, USA). The sensitivity of the assays was 1.0 pg/mL.

2.2.2. A-Microbiological Analysis: Methanogenic Archaea by qPCR DNA Extraction and PCR Assays

For DNA extraction, 0.2 g of each stool sample were mixed in 1.5 mL tubes with 500 µL of G2 lysis buffer from an EZ1[®] DNA Tissue Kit (QIAGEN, Hilden, Germany). Then, 0.3 g of acid-washed beads $\leq 106 \mu\text{m}$ (Sigma-Aldrich, Saint Quentin Fallavier, France) were added in each tube and shaken in a FastPrep BIO 101 device (MP Biomedicals, Illkirch, France) for 45 s for mechanical lysis before 10 min incubation at 100 °C. A 180 µL volume of the mixture was then incubated with 20 µL of proteinase K (QIAGEN, Hilden, Germany) at 56 °C overnight before a second mechanical lysis was performed. Total DNA was finally extracted with an EZ1 Advanced XL extraction kit (QIAGEN) and 50 µL eluted volume. Sterile phosphate buffered saline (PBS) (Fisher Scientific, Illkirch, France) was used as a negative control in each DNA extraction run. Extracted DNA was incorporated into real-time PCR performed using Metha_16S_2_MBF: 5'-CGAACCGGATTAGATACCCG-3' and Metha_16S_2_MBR: 5'-CCCGCCAATTCCTTTAAGTT-3' primers and the FAM_Metha_16S_2_MBP 6FAM-CCTGGGAAGTACGGTCGCAAG probe targeting the 16S DNA gene of methanogens, designed in our laboratory (Eurogentec, Angers, France) as previously described [31]. PCR amplification was done in 20 µL volume including 15 µL of mix and 5 µL of extracted DNA. Five microliters of ultra-pure water (Fisher Scientific, Illkirch, France) were used instead of DNA in the negative controls. The amplification reaction was performed in a CFX96 thermocycler (BioRad, Marnes-la-Coquette, France) incorporating a protocol with a cycle of 50 °C for 2 min, followed by 39 cycles of 95 °C for 45 s, 95 °C for 5 s and finally 60 °C for 30 s. Samples with a CT < 40 were considered positive. Gene amplification and PCR sequencing were performed as previously described [25,26,32–34].

Statistical Analysis

The responses for each quantitative parameter were described using median and 25–75 percentile (interquartile range, IQR) unless otherwise stated. Analyses were performed using the Wilcoxon test when two groups were compared, and the Kruskal–Wallis test when more than two groups were compared. The association between the different biomarkers of interest were analyzed, at each sampling time, using Spearman's correlation coefficient. The association profiles between different biomarkers were also analyzed using a principal component analysis method. Statistical analyses were performed at the conventional two-tailed α level of 0.05, using R 2.13.2 statistical software (R Foundation for Statistical Computing, <https://www.r-project.org> (accessed on 18 March 2022), Vienna, Austria).

3. Results

3.1. Demographic and Clinical Characteristics of Preterm Infants

The 43 preterm neonates included in our study had at birth an average weight of 1160.41 g (range 440–1750 g), an average gestational age of 29 weeks (range 24–32 weeks) and an average height of 37.11 cm (range 32–47 cm). Thirty-five (81%) were born by cesarean section and 8 (19%) by vaginal delivery. Only five mothers (11.6%) had received antibiotic therapy during the peripartum period. As part of the cohort follow-up, clinical evaluation (AF) was conducted at 1 year and assessed the presence or absence of health conditions, including atopic diseases. When necessary, allergy diagnosis was carried out according to current recommendations [35,36]. A total of nine children developed an atopic condition during the first year, manifested as asthma or cow's milk allergy (CMA) in eight and AD in three, with two patients presenting an association of AD, asthma, and CMA (Table 1).

3.2. Immune Profiling

3.2.1. Total Protein Determination

First, we measured the total protein content in all samples. The median concentration of fecal proteins was stable from birth to six weeks, ranging from 4.53 to 9.18 g/L ($p = 0.10$; Kruskal–Wallis) (Table 2).

Table 1. Clinical data for preterm infants investigated for the presence of fecal biomarkers. VD: vaginal delivery; CMA, cow’s milk allergy; CS: cesarean section.

Code	Meconium (M)	Two-Weeks (W2)	Four-Week (W4)	Six-Weeks (W6)	Peripartum Maternal Antibiotic Therapy	Mode of Delivery	Gestational Age	Weight	Size	Asthma or CMA	Atopic Dermatitis
1	0	W2	W4	W6	No	VD	30	1275	37	Yes	Yes
2	M	W2	W4	W6	Yes	CS	27	925	34	Yes	No
3	M	W2	W4	W6	No	CS	26	565	31	Yes	No
4	0	0	W4	0	No	VD	25	820	34	Yes	No
5	M	W2	W4	0	No	CS	32	1260	39	Yes	No
6	M	W2	W4	W6	Yes	CS	27	680	31	Yes	No
7	M	0	W4	W6	No	CS	29	1565	42	No	No
8	M	W2	W4	W6	No	CS	28	890	33	No	No
9	0	0	0	W6	Yes	CS	30	1150	38	No	No
10	M	W2	0	0	No	CS	31	1570	43	No	No
11	M	W2	W4	0	No	CS	32	1575	44	No	No
12	M	W2	W4	0	No	CS	30	1360	39	No	Yes
13	M	W2	0	W6	No	CS	25	870	34	Yes	Yes
14	M	W2	W4	0	No	CS	32	1155	39	Yes	No
15	0	W2	W4	W6	No	CS	25	440	28	No	No
16	0	0	0	W6	No	VD	30	1590	41	No	No
17	M	W2	W4	W6	No	CS	24	530	31	No	No
18	M	W2	0	0	No	CS	26	925	35	No	No
19	M	W2	W4	0	No	VD	30	1480	41	No	No
20	M	W2	W4	0	No	VD	30	1460	38	No	No
21	M	W2	W4	W6	No	CS	29	880	35	No	No
22	0	W2	W4	W6	No	CS	28	840	35	No	No
23	M	W2	W4	0	Yes	VD	30	1670	43	No	No
24	M	W2	W4	W6	No	CS	31	1120	38	No	No
25	0	W2	0	0	No	CS	28	915	36	No	No
26	M	0	W4	W6	No	CS	26	925	35	No	No
27	M	W2	W4	W6	Yes	CS	30	1335	39	No	No
28	M	W2	W4	W6	Yes	CS	30	1355	47	No	No
29	M	W2	0	W6	No	CS	30	1480	39	No	No
30	0	W2	W4	W6	Yes	CS	28	1010	35	No	No
31	M	W2	W4	W6	No	CS	29	1050	39	No	No
32	M	W2	W4	0	No	CS	29	1190	38	No	No
33	0	W2	W4	W6	No	CS	30	1175	39	No	No
34	M	0	0	0	No	CS	32	1930	44	No	No
35	0	W2	W4	0	Yes	CS	29	1430	30	No	No

Table 1. Cont.

Code	Meconium (M)	Two-Weeks (W2)	Four-Week (W4)	Six-Weeks (W6)	Peripartum Maternal Antibiotic Therapy	Mode of Delivery	Gestational Age	Weight	Size	Asthma or CMA	Atopic Dermatitis
36	M	W2	W4	0	No	CS	30	1750	43	No	No
37	M	0	0	0	No	CS	27	600	29	No	No
38	M	W2	W4	W6	No	VD	25	750	32	No	No
39	M	W2	0	W6	No	CS	31	980	36	No	No
40	M	W2	0	W6	No	CS	31	1410	39	No	No
41	M	0	0	0	No	CS	30	770	33	No	No
42	M	0	0	0	Yes	VD	32	1568	41	No	No
43	M	0	0	0	No	CS	30	1680	39	No	No

Table 2. Determination of fecal immune biomarkers.

	Meconium		Two Weeks		Four Weeks		Six Weeks			
	n = 33		n = 33		n = 29		n = 24		p-Value (Frequency)	p-Value (Levels)
	n (%) Detectable	Median IQR	n (%) Detectable	Median IQR	n (%) Detectable	Median IQR	n (%) Detectable	Median IQR		
Total Proteins (g/L)	33 (100)	9.18 (4.51–13.54)	33 (10,055)	5. 4.23–6.05)	29 (100)	4.53 (3.00–5.52)	24 (100)	6.46 (5.39–7.76)	NS	0.10
Total IgE (µg/L)	30 (90.90)	7.3 (6.4–9.9)	32 (97)	8.47 (6.8–9.8)	27 (93.10)	9.74 (3.39–0.26)	24 (100)	115.08 (41.00–193.70)	0.41	<0.0001
Tryptase (µg/L)	3 (9.1)	<1	3 (9.1)	<1	4 (13.79)	<1	14 (58.33)	1.8 (0.0–3.4)	<0.0001	0.61
Calprotectin (µg/L)	33 (100)	310.4 (151.1–771.3)	33 (100)	291.23 (189.41–487.87)	29 (100)	402.44 (300.06–607.3)	24 (100)	422.37 (335.53–823.30)	NC	0.13
EDN (µg/L)	33 (100)	83.2 (19.3–165.0)	33 (100)	70.1 (17.8–152.5)	29 (100)	109.0 (44.2–200.0)	24 (100)	98.1 (57.5–200.0)	NC	0.21
TGF-β (pg/L)	24 (72.7)	121.3 (4.6–258.9)	30 (91)	267.43 (61.71–1000)	26 (89.65)	384.57 (129.60–936)	22 (91.66)	466 (104.36–1430.29)	0.09	0.014
IL-1β (pg/L)	13 (39.4)	0.12 (0.1–2.7)	28 (84.8)	1.53 (0.37–6.53)	25 (86.20)	3.27 (0.31–10.76)	22 (91.66)	6.23 (1.66–20.84)	<0.0001	0.001
IL-10 (pg/L)	4 (12.12)	3.9 (3.9–3.9)	6 (18.18)	3.9 (3.9–3.9)	5 (17.24)	3.9 (3.9–3.9)	5 (20.83)	3.9 (3.9–3.9)	0.85	0.53
IL-6 (pg/L)	25 (75.75)	11.6 (0.5–43.7)	7 (21.21)	0.2 (0.2–0.2)	20 (68.96)	0.2 (0.2–0.2)	20 (83.33)	3.77 (1.66–19.25)	<0.0001	<0.001

Concentrations are expressed as median and interquartile ranges (IQR). n (%): number of samples in which the biomarker was detected (relative frequency of detection). The median and IQR were calculated by restricting the results above the lower LOQ (limit of quantitation) for each analyte. Statistical test: chi-square (frequency), Kruskal–Wallis (concentration). NC, not calculable (calprotectin and EDN were detectable in all samples and at all sampling times).

3.2.2. Immune Cell Markers and Cytokines

Total IgE was detectable in over 90% of the samples at all ages, in increasing amounts between birth (meconium) and six weeks ($p < 0.0001$; Kruskal–Wallis).

Conversely, tryptase detection increased with sampling age, reaching 58% in samples at six weeks, up from less than 15% at earlier times ($p < 0.0001$; Chi-square). As most values were lower than the quantification limit, quantitative comparison was not significant ($p = 0.61$, Kruskal–Wallis) (Figure 2 and Table 2).

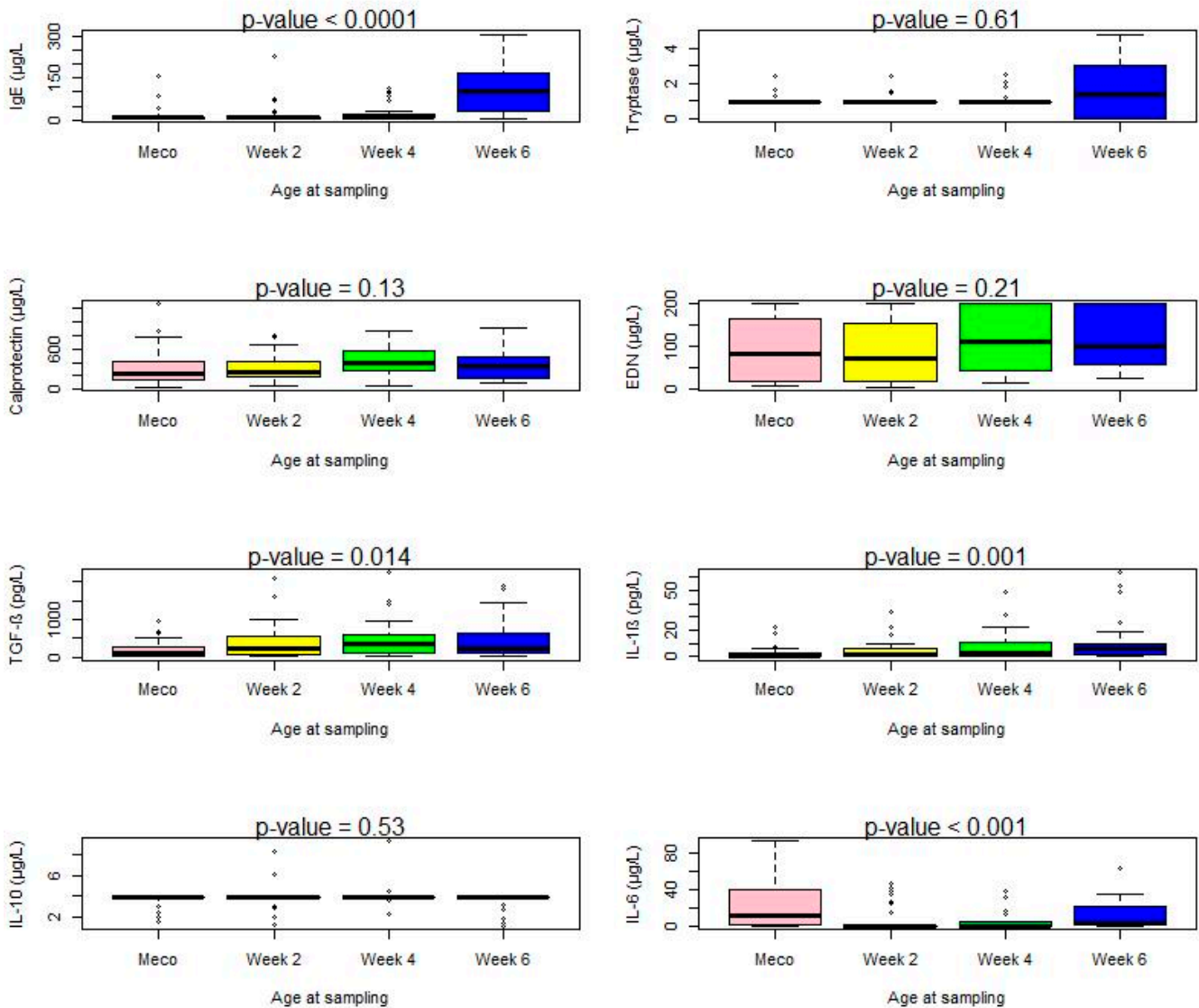


Figure 2. Statistical test: chi-square (frequency), Kruskal–Wallis (concentration). NC, not calculable (calprotectin and EDN were detectable in all samples and at all sampling times).

Calprotectin and EDN were detected in all samples at comparable levels irrespective of age ($p = 0.13$ and 0.21 , Kruskal–Wallis) (Figure 2 and Table 2).

All cytokines except TNF- α were detectable in meconium and fecal samples. TGF- β and IL-6 were the most prevalent, detected in up to 90% of samples, while IL-10 was the less prevalent, found in 20% or less of the fecal samples. The frequency of detection of IL-6 and IL-1 β increased with age ($p < 0.0001$; chi-square), although there was a sharp drop in IL-6 frequency of detection and measured levels between meconium (75%, median 11.6 pg/L) and samples at two weeks (21%, median 0.2 pg/L).

TGF- β and IL-1 β median concentrations increased with age ($p = 0.014$ and 0.001 , respectively; Kruskal–Wallis). The median level of IL-6 was the highest in meconium samples and increased again at six weeks ($p < 0.0001$; Kruskal–Wallis). IL-10 median concentrations did not vary with age (Figure 2 and Table 2).

Maternal antibiotic therapy and route of delivery did not significantly affect the meconium levels of cytokines, total IgE, tryptase, calprotectin, and EDN (Supplementary Table S1). However, analysis according to the development of atopic disease during the first year showed that meconium calprotectin levels were lower in neonates who subsequently developed asthma or CMA compared to those who did not ($p = 0.02$; Wilcoxon test) (Figure 2). Levels of other mediators and cytokines were not associated with the occurrence of an atopic disease (Table 3).

Table 3. Comparison of mediators and cytokines and the occurrence or absence of an atopic condition between years 0 and 1. Statistical test used: Wilcoxon test.

Variables	Allergic Condition (APLV and Asthma)	<i>p</i> -Value			
		Meconium	2 Weeks	4 Weeks	6 Weeks
IgE	Yes	0.27	0.06	0.12	0.03
	No				
Calprotectin	Yes	0.27	0.18	0.91	0.61
	No				
EDN	Yes	0.59	0.19	0.41	0.87
	No				
TGF- β	Yes	0.09	0.18	0.76	0.76
	No				
IL-1 β	Yes	0.37	1.00	0.28	0.91
	No				
IL-10	Yes	0.62	1.00	0.89	0.13
	No				
IL-6	Yes	0.61	0.28	0.37	0.75
	No				

Comparing the different biomarkers according to the presence or absence of an atopic condition, we observed a significant difference at week six for IgE between the presence and absence of cow's milk allergy and asthma. No significant difference was observed for AD (Supplementary Table S2)

3.2.3. Correlation between Biomarkers

As an expected control, significant correlations were found between weight and height ($R = 0.90$; $p < 0.0001$), between gestational age and height ($R = 0.75$; $p < 0.0001$), and between gestational age and weight ($R = 0.75$; $p < 0.0001$).

Total IgE and tryptase levels were strongly correlated in samples taken at any age. IL-10 and IL-6 were correlated at all ages except at two weeks (Table 4).

3.2.4. Meconium Samples

Tryptase levels were correlated to levels of IL-10 ($R = 0.48$, $p = 0.001$) and IL-6 ($R = 0.46$; $p = 0.001$). Tryptase levels were correlated to levels of IgE ($R = 0.91$, $p = 0.0001$), and strong correlations were observed between levels of calprotectin and IL-1 β ($R = 0.90$; $p < 0.0001$). A negative correlation between total protein concentration and TGF- β ($R = -0.36$; $p = 0.01$) was observed (Table 3).

Table 4. Result of the detection of methanogenic Archaea.

	Meconium (n = 33)		Two Weeks (n = 33)		Four Weeks (n = 29)		Six Weeks (n = 26)		p-Value (Frequency)	p-Value (CT)
	n (%)	Median IQR	n (%)	Median IQR	n (%)	Median IQR	n (%)	Median IQR		
CT qPCR	30 (90.9)	36.74 (33.85–38.24)	27 (81.81)	37.20 (36.07–38.33)	23 (79.31)	37.75 (36.13–38.50)	19 (73.03)	38.28 (37.27–39.96)	0.34	0.12

CT methanogenic Archaea are expressed as median and interquartile ranges (IQR). n (%): number of samples in which methanogenic Archaea were detected (relative frequency of detection). The stool concentration factor and median and RDI were not included in our calculations, and the median and RDI were calculated by restricting the results above the lower LOQ (limit of quantitation) for each analyte. Statistical test: Kruskal–Wallis.

3.2.5. Samples at Two Weeks

Strong correlations were observed between levels of calprotectin and IL-1 β ($R = 0.90$; $p < 0.0001$), tryptase and IL-10 ($R = 0.88$; $p < 0.0001$), and total IgE and IL-10 ($R = 0.85$; $p < 0.0001$), while total protein concentration and IL-6 were negatively correlated ($R = -0.61$; $p = 0.0002$) (Table 3).

3.2.6. Samples at Four Weeks

Again, calprotectin and IL-1 β were strongly correlated ($R = 0.74$; $p < 0.0001$). IgE and total protein were also correlated ($R = 0.38$; $p < 0.04$) (Figure 3).

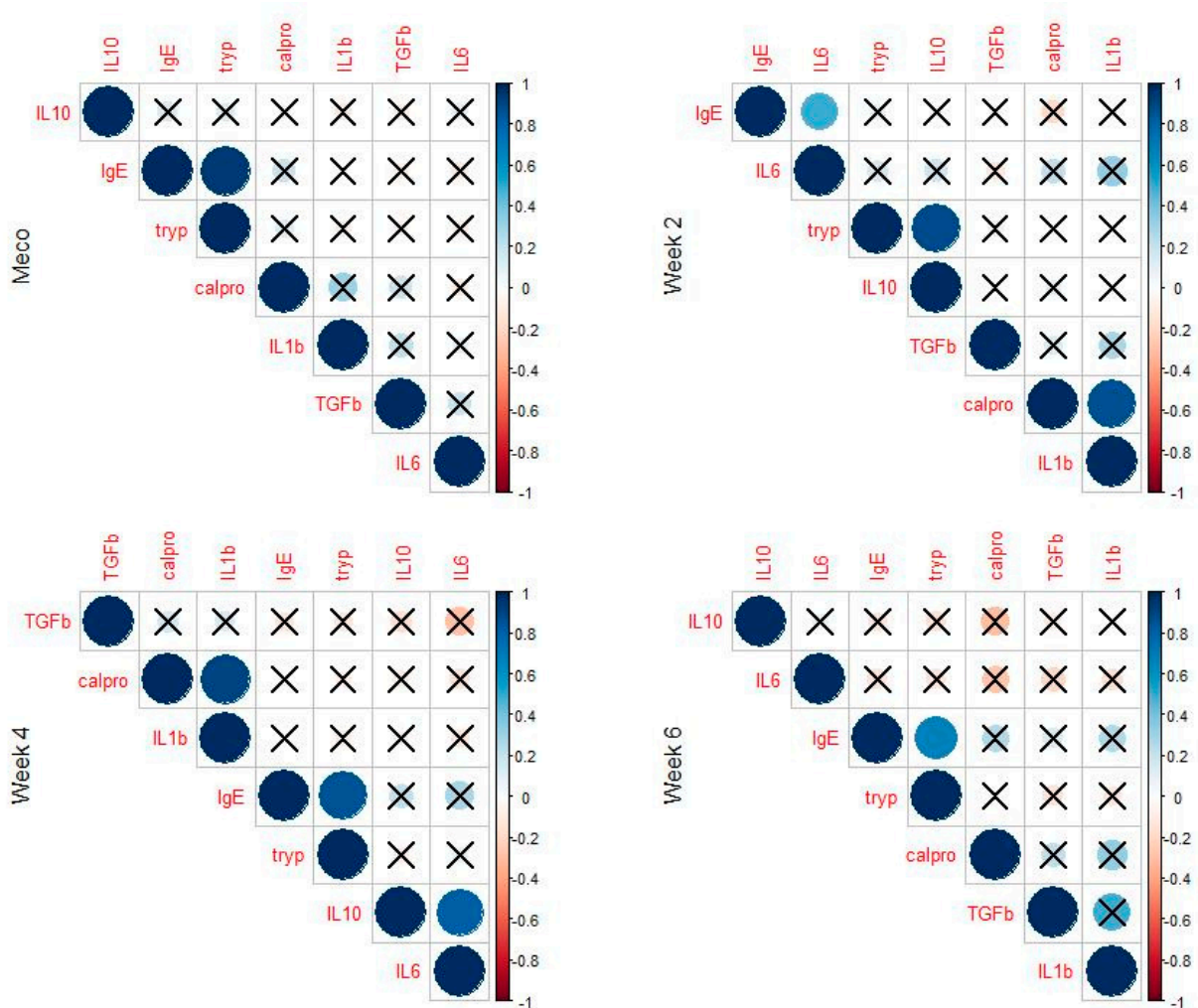


Figure 3. Correlation figure of fecal immune biomarkers.

3.2.7. Samples at Six Weeks

TGF- β was correlated with IL-1 β ($R = 0.49$; $p < 0.01$) and with total proteins ($R = 0.43$; $p < 0.03$). Total proteins were correlated with IL-1 β ($R = 0.47$; $p < 0.02$) and negatively with IL-6 ($R = -0.68$; $p = 0.0003$) (Figure 3).

3.3. Frequency of Detection of Methanogenic Archaea and Relationship with the Subsequent Development of Atopic Diseases

Using real-time PCR with 16S rRNA archaeal gene PCR primers, we detected methanogenic Archaea DNA in 30/33 (90%) meconium samples, 27/33 (81%) two-week-old samples, 23/29 (79%) four-week-old samples, and 19/24 (73%) six-week-old samples, respectively. We found no significant difference in the frequency of detection nor in CTs according to age at sampling (Table 4).

3.4. Unsupervised Analysis of Immunological Markers and Methanogenic Archaea At the Neonatal Period, and the Subsequent Occurrence of AD, Asthma and CMA during the First Year

We performed unsupervised analysis of the immunological data, CT of Archaea, and the clinical information of the occurrence of allergic events during the first year of life. Data were analyzed for each of the four sampling times.

For meconium, calprotectin, EDN, and IL-1 β levels were negatively and significantly ($p < 0.001$) correlated ($r = -0.64$) with subsequent development of AD. Calprotectin, EDN and IL-1 β had the largest and most significantly ($p < 0.01$) correlated positive correlation coefficients, which were 0.79, 0.53, and 0.51, respectively. No correlation was observed for the Archaea CT with the other parameters (Figure 4b(A)).

At two weeks, IL-1 β ($r = 0.88$) and calprotectin ($r = 0.82$) had a strong positive correlation with each other, followed by IL-6 ($r = 0.62$) and EDN ($r = 0.59$). These biomarkers were significantly ($p < 0.001$) associated. Archaea CTs had a weak ($r < 0.5$) but positive and significant ($p < 0.01$) association with the biomarkers IL-1 β , calprotectin, IL-6, and EDN (Figure 4b(B)).

At four weeks, later occurrence of AD, CMA, and asthma was positively correlated with calprotectin ($r = 0.61$), IL-1 β ($r = 0.58$), EDN ($r = 0.57$), and TGF- β ($r = 0.57$), and negatively correlated with IL-6 ($r = -0.71$) and IL-10 ($r = -0.61$). Calprotectin, IL-1 β , and EDN were significantly associated with each other ($p < 0.001$). Archaea CTs were positively associated with calprotectin, IL-1 β , IL-6, and EDN with a significant correlation ($p < 0.001$), however, they were inversely correlated with allergic events, although the correlation coefficient was low (Figure 4b(C)).

At six weeks, only IL-6 correlated negatively ($r = -0.51$) with the other biomarkers and allergic events. IL-1 β ($r = 0.68$), IL-10 ($r = 0.67$), and AD ($r = 0.58$) showed the strongest positive correlations, with AD significantly ($p < 0.001$) associated with IL-1 β and IL-10. Archaea CTs were weakly correlated ($r < 0.5$) with IL-1 β , calprotectin, IL-6, and EDN. However, allergic events were negatively associated with CT, and AD had almost no correlation (Figure 4b(D)).

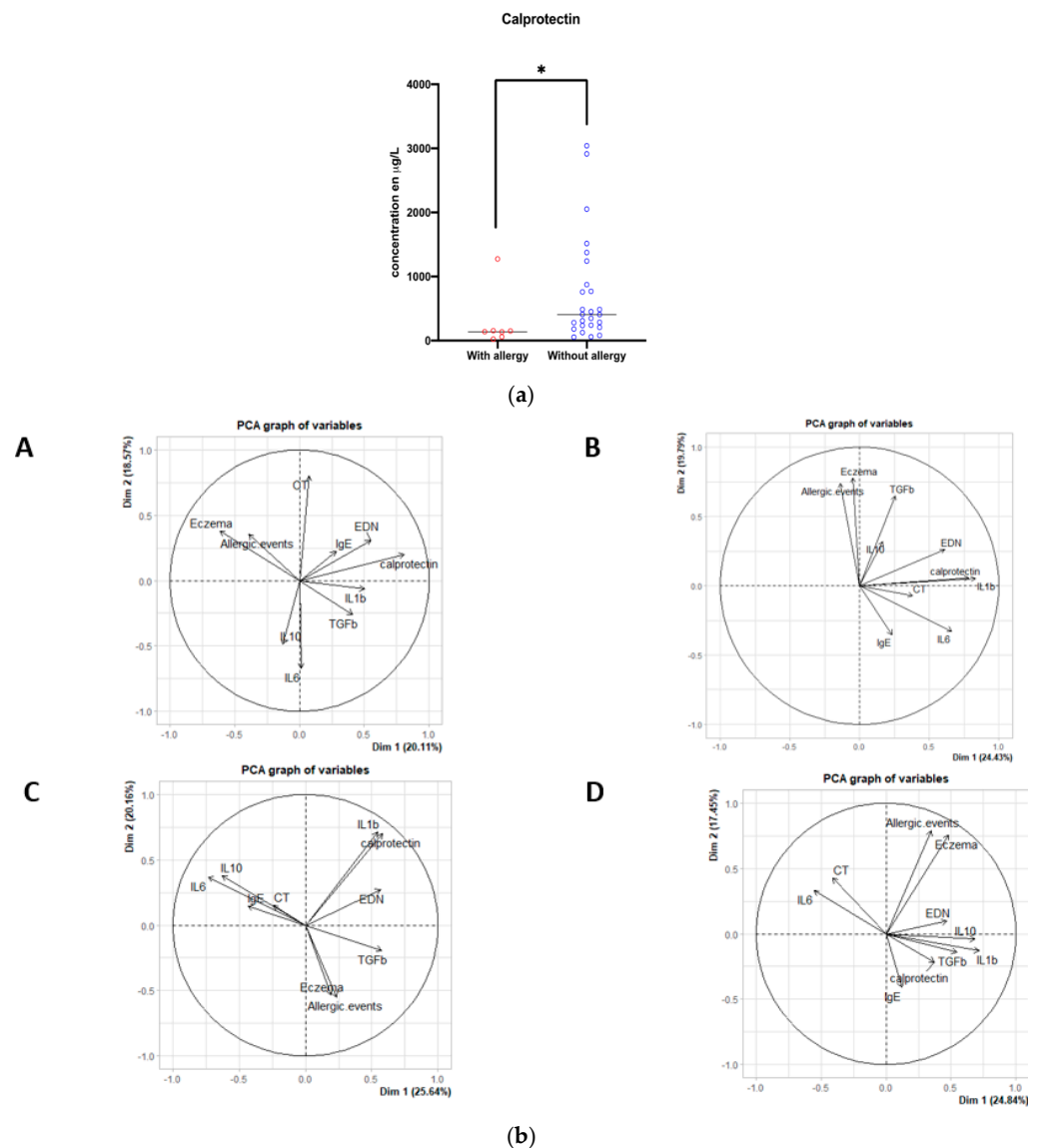


Figure 4. (a). Comparison of meconial calprotectin concentration according to the later occurrence of asthma or cow’s milk allergy. (b). Principal component analysis of neonatal immune and archaeal biomarkers as a function of later occurrence of atopic conditions. (A) Negative and significant ($p < 0.001$) correlation ($r = -0.64$) between the occurrence of atopic dermatitis with calprotectin, EDN, and IL-1 β . Strong positive and significant correlation ($p < 0.01$) between calprotectin ($r = 0.79$), EDN (0.53), and IL-1 β (0.51). No correlation between Archaea TCs and other parameters. (B) Correlation between allergic events (asthma or cow’s milk allergy) and atopic dermatitis. Strong positive and significant ($p < 0.001$) correlation between IL-1 β ($r = 0.88$) and calprotectin ($r = 0.82$), positive correlation between IL-6 ($r = 0.62$) and EDN ($r = 0.59$), significant ($p < 0.001$). Low positive ($r < 0.5$) and significant correlation between Archaea TCs and biomarkers IL-1 β , calprotectin, IL-6, and EDN. (C) Positive and significant correlation ($p < 0.001$) between atopic dermatitis, allergic events with the markers calprotectin ($r = 0.61$), IL-1 β ($r = 0.58$), EDN ($r = 0.57$), and TGF- β ($r = 0.57$). Negative correlation between IL-6 ($r = -0.71$) and IL-10 ($r = -0.61$) with atopic dermatitis and allergic events. Positive and significant correlation between Archaea TCs with calprotectin, IL-1 β , IL-6, and EDN. Negative correlation between Archaea Ct and allergic events. (D) Negative correlation between IL-6 ($r = -0.51$) with other biomarkers and allergic events. Strong positive and significant correlation ($p < 0.001$) between IL-1 β ($r = 0.68$), IL-10 ($r = 0.67$), and atopic dermatitis ($r = 0.58$). Low correlation ($r < 0.5$) between Archaea TCs with IL-1 β , calprotectin, IL-6, and EDN. Negative correlation between allergic events and TC of Archaea.

4. Discussion

In this study, we describe a non-invasive screening method for profiling neonatal immunity and its validation in a preterm neonate cohort as a predictive tool for subsequent development of atopic diseases. The method was also applied to meconium samples, which reflect intrauterine processes and contain almost 1000 identified proteins with important functions [37]. The clinical endpoints of this study were evaluated at the age of one year, while the total duration of cohort follow-up will be three years.

Total IgE was detected in over 90% of the samples, at increasing concentrations with age. Transplacental delivery of allergens and preterm sensitization have long been recognized, possibly inducing sensitization and detectable meconial IgE [38,39]. Transplacental transport of maternal IgE able to sensitize fetal mast cells has been recently demonstrated [40], but its role in neonatal immune defenses or subsequent immune disorders is only speculative.

Addressing mast cell tryptase in meconium and later samples, we found that it was detectable only in a minority of meconium samples and during the first month of life, however, it became a common finding at the end of the neonatal period, represented by samples collected at six weeks. Fecal tryptase and IgE levels were strongly associated at each of the studied time points. These results suggest that gastrointestinal mast cells, as opposed to skin mast cells [40], are mostly recruited postnatally, and mature after birth with IgE levels exerting a positive effect. Conversely, maturity of gastrointestinal mast cell populations might be attained during late pregnancy. Tryptase is a serine protease able of autocrine activation of mast cells and induction of proinflammatory effects such as proteolytic cleavage and activation of PAR2 receptors and inactivation of VIP (Vasoactive Intestinal Peptide), associated with smooth muscle relaxation [41]. Through PAR-2 activation, luminal tryptase can contribute to the dysfunction of the gut epithelial barrier [42]. The presence of tryptase in stool samples has been associated with food allergic diseases, dietary exposure and/or mast cell stimulation or increased intestinal mast cell count [43,44]. In addition, fecal tryptase has also been shown to be associated with inflammatory bowel disease and irritable bowel syndrome [45,46].

Focusing on two secreted biomarkers of innate immune cells, neutrophil-derived calprotectin, and eosinophil-derived EDN, we found that fecal samples at all studied ages contained detectable and stable levels of both biomarkers. The levels measured in our preterm cohort were much lower than the reference values [47], but similar to those reported during the first postnatal month in another preterm cohort [48]. These results suggest that the preterm gut contains small numbers of granulocytes, or that such granulocytes are not activated. Indeed, lower neutrophils were reported in preterm infant cord blood [49]. An association between low levels of fecal calprotectin and adverse health conditions, including obesity and sepsis, by age two has been suggested [48].

Proinflammatory and anti-inflammatory cytokines were detected in the meconium and later samples of preterm infants, with the notable exception of TNF- α which was not demonstrated in any sample. Different temporal patterns were demonstrated: IL-6 levels were higher in the meconium than in later samples, while IL-10 was seldom detected and TGF- β and IL-1 β displayed a progressive increase between birth and six weeks. Mostly undetectable IL-10 levels were also reported in a pilot study of fecal biomarkers in preterm infants [8]. Although we did not determine the cellular source of fecal cytokines, a shift in immune cells lining the intestine has been demonstrated for macrophages, with resident fetal macrophages being replaced after birth by bone-marrow-derived macrophages [50]. Macrophage cytokine production, most notably of proinflammatory IL-6 and IL-1 β , can be persistently altered by metabolic conditions [51]. The increase in TGF- β levels from birth to six weeks might provide a counter-acting mechanism in a proinflammatory environment. Allergic events (asthma or cow's milk allergy) and atopic dermatitis were also positively correlated with EDN, IL-1 β , IL-10, and IL-6 at four and six weeks. The high production of fecal EDN, IL-1 β , and IL-10 during the first weeks of life may therefore be an indicator for later risk of allergic diseases.

The neonatal period is paramount for the establishment of the intestinal microbiota. Intrauterine life is associated with low levels of maternal microbial translocation [52]. However, we have recently demonstrated the presence of the viable methanogenic Archaea *Methanobrevibacter smithii* in the meconium, suggesting intrauterine colonization of the fetus by this microorganism [26]. Here, we provided evidence for postnatal persistence of methanogenic Archaea in fecal samples and suggest a possible role in the orientation of intestinal immunity, supported by the negative association between Archaea abundance (inversely proportional to CT values) and the concentrations of EDN, IL-1 β , and IL-6. We also found that Archaea abundance at four and six weeks was positively associated with later occurrence of allergic events. Archaea have been shown to produce SCFA which induce regulatory T cell differentiation, downregulate proinflammatory cytokines, and may protect against the occurrence of atopic conditions [19,53–57]. However, in a cohort study, the protective effect of methanogenic Archaea was restricted to the species *Methanobrevibacter stadtmanae* [58]. A decrease in the load of beneficial methanogenic Archaea during the first years of life could therefore favor the occurrence of allergic events during the first years of life.

A third line of contribution to protection or increased risk of developing atopic conditions is the genetic background. As an example, a del/del genotype (–2549 –2567 del18) of Vascular Endothelial Growth Factor (VEGF) has been associated with asthma occurrence and irreversible bronchoconstriction [59].

The strengths of our study are methodological and medical:

- (1) miniaturization and standardization, using small quantities of stool (1 g) and small volumes of extraction buffer (2 mL). The dilution of the samples was corrected by the freeze-drying process, as the lyophilizates were contained in 1 mL of buffer.
- (2) prevention, thanks to the use of protease inhibitors, of the risk of potential contamination of the handler.
- (3) suitability for a microarray platform yielding patterns of immune responses rather than individual measurements.
- (4) suitability for combined immune and microbiological assessment.
- (5) proof of concept of the immune profiling of fecal mediators in meconium and neonatal samples as predictors of later development of atopic disorders.
- (6) proof of concept for non-invasive investigation of the immune status of preterm neonates.

The main weakness of this study is the lack of microbiological data outside Archaea. Further studies are warranted for longitudinal immuno-microbiological profiling of meconium and neonatal samples, in preterm and at-term infants. Its validation as a non-invasive diagnostic method will be in line with the currently unmet needs in terms of non-invasive diagnosis of allergy.

5. Conclusions

This study allowed us to highlight the presence of mediators in the meconium and feces of preterm infants. We provide proof of concept of the feasibility and value of a standardized fecal mediator assay for non-invasive profiling of neonatal immunity. Such assays can be used for early characterization of the immune status of a newborn. Technical optimization for a multiplex assay could facilitate the implementation of fecal immune profiling in clinical and research laboratories. We report evidence of a correlation between the meconial and neonatal load of methanogenic Archaea and selected fecal biomarkers, and later occurrence of atopic conditions in preterm children.

Supplementary Materials: The following supporting information can be downloaded at: <https://www.mdpi.com/article/10.3390/jcm11102749/s1>, Table S1: Comparison of biomarkers and CT of Archaea according to the presence or absence of maternal antibiotic therapy and according to the mode of delivery, Table S2: Comparison of median CT of Archaea in preterm infants according to the presence or absence of allergic events during the first year of life, Table S3: Comparison of

median CTs of Archaea in preterm infants according to the presence or absence of atopic dermatitis during the first year of life.

Author Contributions: Y.S. developed the stool preparation method, performed the immunology experiments, analyzed the data, and wrote the first draft of the manuscript. M.M. and S.M. performed the experiments, analyzed the data, and proofread the manuscript. Y.S., C.O.G., L.K. and G.G. performed the microbiology experiments and analyzed the data. T.M. analyzed the data. G.G. and T.M. proofread the manuscript. J.-L.M., T.A.T., P.C., A.F. and J.V. designed the study and supervised the experiments. T.A.T. and A.F. collected and analyzed the clinical data. A.F. included patients and performed clinical examination. T.A.T., P.C., A.F. and J.V. wrote the manuscript. All authors have read and agreed to the published version of the manuscript.

Funding: Y.S. is supported by a “Fondation Méditerranée Infection” doctoral position. S.M. was first supported by a “Fondation pour la Recherche Médicale” postdoctoral fellowship (SPF20151234951) and then by a “Fondation Méditerranée infection” postdoctoral fellowship. This work was supported by the French Government under the “Investissements d’avenir” (Investments for the Future) program managed by the “Agence Nationale de la Recherche” (reference: Méditerranée Infection 10-IAHU-03), by the Région Provence-Alpes-Côte d’Azur and the European funding FEDER IHU PRIMMI, and the PHRC program for clinical research Primibiota.

Institutional Review Board Statement: This study was embedded in the Primibiota “Influence of Intestinal Microbiota Implantation in Preterm Infants on Microbiota and Immune Orientation at 3 Years” (NCT02738411, principal investigator AF), a population-based prospective cohort study from birth to the age of 3 years, enrolling children born preterm in the University Hospitals of Nîmes and Montpellier, France. The ancillary study presented here was approved by a joint committee of the Clinical Research Departments of the University Hospitals of Nîmes, France and the University Hospitals of Marseille, France (Research collaboration agreement 2018.1238).

Informed Consent Statement: Informed consent was obtained from both parents at infants’ birth.

Data Availability Statement: Not applicable.

Acknowledgments: We thank Michel Drancourt and Bernard La Scola for conceptual and methodological assistance; Patricia Blanchard, Céline Chartier, Robert Perrier, and Simon Pinchemel for technical assistance with the ImmunoCAP platform.

Conflicts of Interest: J.V. reports speaker and consultancy fees in the past five years from Astra Zeneca, Meda Pharma (Mylan), Novartis, Sanofi, Thermo Fisher Scientific, outside the submitted work. The other authors declare no competing interests in relation to this study.

Abbreviations

AD: atopic dermatitis; CMA, cow’s milk allergy; CT, cycle threshold; IQR, interquartile range; SCFA, short chain fatty acid; EDN, eosinophil-derived neurotoxin.

References

1. Crump, C.; Sundquist, J.; Winkleby, M.A.; Sundquist, K. Gestational age at birth and mortality from infancy into mid-adulthood: A national cohort study. *Lancet Child Adolesc. Health* **2019**, *3*, 408–417. [[CrossRef](#)]
2. Chawanpaiboon, S.; Vogel, J.P.; Moller, A.B.; Lumbiganon, P.; Petzold, M.; Hogan, D.; Landoulsi, S.; Jampathong, N.; Kongwatanakul, K.; Laopaiboon, M.; et al. Global, regional, and national estimates of levels of preterm birth in 2014: A systematic review and modelling analysis. *Lancet Glob. Health* **2019**, *7*, e37–e46. [[CrossRef](#)]
3. Goldenberg, R.L.; Culhane, J.F.; Iams, J.D.; Romero, R. Epidemiology and causes of preterm birth. *Lancet* **2008**, *371*, 75–84. [[CrossRef](#)]
4. Schuster, J.; Uzun, A.; Stablia, J.; Schorl, C.; Mori, M.; Padbury, J.F. Effect of prematurity on genome wide methylation in the placenta. *BMC Med. Genet.* **2019**, *20*, 116. [[CrossRef](#)]
5. Beck, S.; Wojdyla, D.; Say, L.; Betran, A.P.; Merialdi, M.; Requejo, J.H.; Rubens, C.; Menon, M.; Look, V.F.P. The worldwide incidence of preterm birth: A systematic review of maternal mortality and morbidity. *Bull. World Health Organ.* **2010**, *88*, 31–38. [[CrossRef](#)]
6. Gagneur, A.; Piquier, D.; Quach, C. Immunization of preterm infants. *Hum. Vaccines Immunother.* **2015**, *11*, 2556–2563. [[CrossRef](#)]
7. Marshall, H.; Clarke, M.; Rasiah, K.; Richmond, P.; Buttery, J.; Reynolds, G.; Andrews, R.; Nissen, M.; Wood, N.; McIntyre, P. Predictors of Disease Severity in Children Hospitalized for Pertussis During an Epidemic. *Pediatr. Infect. Dis. J.* **2015**, *34*, 339–345. [[CrossRef](#)]

8. Gómez, M.; Moles, L.; Espinosa-Martos, I.; Bustos, G.; de Vos, W.; Fernández, L.; Rodríguez, M.J.; Fuentes, S.; Jiménez, E. Bacteriological and Immunological Profiling of Meconium and Fecal Samples from Preterm Infants: A Two-Year Follow-Up Study. *Nutrients* **2017**, *9*, 1293. [\[CrossRef\]](#)
9. Helmo, F.R.; Alves, E.A.R.; Moreira, R.A.D.A.; Severino, V.O.; Rocha, L.P.; Monteiro, M.L.G.D.R.; Marlene Antônia dos Reis, D.A.M.; Etchebehere, M.R.; Machado, R.J.; Corrêa, M.R.R. Intrauterine infection, immune system and premature birth. *J. Matern.-Fetal Neonatal Med.* **2018**, *31*, 1227–1233. [\[CrossRef\]](#)
10. Sonnenschein-van der Voort, A.M.; Arends, L.R.; de Jongste, J.C.; Annesi-Maesano, I.; Arshad, S.H.; Barros, H.; Basterrechea, M.; Bisgard, H.; Chatzi, L.; Corpeleijn, E.; et al. Preterm birth, infant weight gain, and childhood asthma risk: A meta-analysis of 147,000 European children. *J. Allergy Clin. Immunol.* **2014**, *133*, 1317–1329. [\[CrossRef\]](#)
11. Wooldridge, A.L.; McMillan, M.; Kaur, M.; Giles, L.C.; Marshall, H.S.; Gatford, K.L. Relationship between birth weight or fetal growth rate and postnatal allergy: A systematic review. *J. Allergy Clin. Immunol.* **2019**, *144*, 1703–1713. [\[CrossRef\]](#)
12. Mitselou, N.; Hallberg, J.; Stephansson, O.; Almqvist, C.; Melén, E.; Ludvigsson, J.F. Cesarean delivery, preterm birth, and risk of food allergy: Nationwide Swedish cohort study of more than 1 million children. *J. Allergy Clin. Immunol.* **2018**, *142*, 1510–1514. [\[CrossRef\]](#)
13. Barbarot, S.; Gras-Leguen, C.; Colas, H.; Garrot, E.; Darmaun, D.; Larroque, B.; Roze, J.C.; Ancel, P.; Ancel. Lower risk of atopic dermatitis among infants born extremely preterm compared with higher gestational age. *Br. J. Dermatol.* **2013**, *169*, 1257–1264. [\[CrossRef\]](#)
14. Al Nabhani, Z.; Eberl, G. Imprinting of the immune system by the microbiota early in life. *Mucosal Immunol.* **2020**, *13*, 183–189. [\[CrossRef\]](#)
15. Stokholm, J.; Blaser, M.J.; Thorsen, J.; Rasmussen, M.A.; Waage, J.; Vinding, R.K.; Schoos, M.A.; Kunøe, A.; Fink, R.N.; Chawes, C.B.; et al. Maturation of the gut microbiome and risk of asthma in childhood. *Nat. Commun.* **2018**, *9*, 141. [\[CrossRef\]](#)
16. Gomez Perdiguero, E.; Klapproth, K.; Schulz, C.; Busch, K.; Azzoni, E.; Crozet, L.; Garner, H.; Trouillet, C.; Bruijn, D.F.M.; Geissmann, F.; et al. Tissue-resident macrophages originate from yolk-sac-derived erythro-myeloid progenitors. *Nature* **2015**, *518*, 547–551. [\[CrossRef\]](#)
17. Schirmer, M.; Smeekens, S.P.; Vlamakis, H.; Jaeger, M.; Oosting, M.; Franzosa, E.A.; Horst, T.R.; Jansen, A.; Jacobs, L.; Bonder, J.M.; et al. Linking the Human Gut Microbiome to Inflammatory Cytokine Production Capacity. *Cell* **2016**, *167*, 1897. [\[CrossRef\]](#)
18. Martens, E.C.; Neumann, M.; Desai, M.S. Interactions of commensal and pathogenic microorganisms with the intestinal mucosal barrier. *Nat. Rev. Microbiol.* **2018**, *16*, 457–470. [\[CrossRef\]](#)
19. Sereme, Y.; Mezouar, S.; Grine, G.; Mege, J.L.; Drancourt, M.; Corbeau, P.; Vitte, J.L. Methanogenic Archaea: Emerging Partners in the Field of Allergic Diseases. *Clin. Rev. Allergy Immunol.* **2019**, *57*, 456–466. [\[CrossRef\]](#)
20. Dridi, B.; Henry, M.; El Khéchine, A.; Raoult, D.; Drancourt, M. High prevalence of *Methanobrevibacter smithii* and *Methanosphaera stadtmanae* detected in the human gut using an improved DNA detection protocol. *PLoS ONE* **2009**, *4*, e7063. [\[CrossRef\]](#)
21. Gaci, N.; Borrel, G.; Tottey, W.; O'Toole, P.W.; Brugère, J.F. Archaea and the human gut: New beginning of an old story. *World J. Gastroenterol.* **2014**, *20*, 16062–16078. [\[CrossRef\]](#)
22. Bang, C.; Weidenbach, K.; Gutschmann, T.; Heine, H.; Schmitz, R.A. The Intestinal Archaea *Methanosphaera stadtmanae* and *Methanobrevibacter smithii* Activate Human Dendritic Cells. *PLoS ONE* **2014**, *9*, e99411. [\[CrossRef\]](#)
23. Vierbuchen, T.; Stein, K.; Heine, H. RNA is taking its Toll: Impact of RNA-specific Toll-like receptors on health and disease. *Allergy* **2019**, *74*, 223–235. [\[CrossRef\]](#)
24. Bernatchez, E.; Gold, M.J.; Langlois, A.; Blais-Lecours, P.; Boucher, M.; Duchaine, C.; Marsolais, D.; McNagny, M.K.; Blanchet, R.M. *Methanosphaera stadtmanae* induces a type IV hypersensitivity response in a mouse model of airway inflammation. *Physiol. Rep.* **2017**, *5*, e13163. [\[CrossRef\]](#)
25. Grine, G.; Boualam, M.A.; Drancourt, M. *Methanobrevibacter smithii*, a methanogen consistently colonising the newborn stomach. *Eur. J. Clin. Microbiol. Infect. Dis.* **2017**, *36*, 2449–2455. [\[CrossRef\]](#)
26. Sereme, Y.; Guindo, C.O.; Filleron, A.; Corbeau, P.; Tran, T.A.; Drancourt, M.; Vitte, J.; Grine, G. Meconial *Methanobrevibacter smithii* suggests intrauterine methanogen colonization in preterm neonates. *Curr. Res. Microb. Sci.* **2021**, *2*, 100034. [\[CrossRef\]](#)
27. Baumann, R.; Untersmayr, E.; Zissler, U.M.; Eyerich, S.; Adcock, I.M.; Brockow, K.; Biedermann, T.; Ollert, M.; Chaker, M.A.; Pfaar, O.; et al. Noninvasive and minimally invasive techniques for the diagnosis and management of allergic diseases. *Allergy* **2021**, *76*, 1010–1023. [\[CrossRef\]](#)
28. Bao, R.; Hesser, L.A.; He, Z.; Zhou, X.; Nadeau, K.C.; Nagler, C.R. Fecal microbiome and metabolome differ in healthy and food-allergic twins. *J. Clin. Investig.* **2021**, *131*, e141935. [\[CrossRef\]](#)
29. Sereme, Y.; Zarza, S.M.; Medkour, H.; Amona, I.; Fenollar, F.; Akiana, J.; Mezouar, S.; Orain, N.; Vitte, J.; Davout, D.; et al. Stool Serology: Development of a Non-Invasive Immunological Method for the Detection of Enterovirus-Specific Antibodies in Congo Gorilla Faeces. *Microorganisms* **2021**, *9*, 810. [\[CrossRef\]](#)
30. Lambert, C.; Sarrat, A.; Bienvenu, F.; Brabant, S.; Nicaise-Roland, P.; Alyanakian, M.-A.; Apoil, P.A.; Capron, C.; Couderc, R.; Evrard, B.; et al. The importance of EN ISO 15189 accreditation of allergen-specific IgE determination for reliable in vitro allergy diagnosis. *Allergy* **2015**, *70*, 180–186. [\[CrossRef\]](#)

31. Drancourt, M.; Djemai, K.; Gouriet, F.; Grine, G.; Loukil, A.; Bedotto, M.; Lévasseur, A.; Lépidi, H.; Khalil, B.J.; Khelaifia, S.; et al. Methanobrevibacter smithii archaeemia in febrile patients with bacteremia, including those with endocarditis. *Clin. Infect. Dis.* **2020**, *73*, e2571–e2579. [[CrossRef](#)] [[PubMed](#)]
32. Grine, G.; Terrer, E.; Boualam, M.A.; Aboudharam, G.; Chaudet, H.; Ruimy, R.; Drancourt, D. Tobacco-smoking-related prevalence of methanogens in the oral fluid microbiota. *Sci. Rep.* **2018**, *8*, 9197. [[CrossRef](#)] [[PubMed](#)]
33. Nkamga, V.D.; Huynh, H.T.T.; Aboudharam, G.; Ruimy, R.; Drancourt, M. Diversity of human-associated Methanobrevibacter smithii isolates revealed by multispacer sequence typing. *Curr. Microbiol.* **2015**, *70*, 810–815. [[CrossRef](#)] [[PubMed](#)]
34. Guindo, C.O.; Davoust, B.; Drancourt, M.; Grine, G. Diversity of Methanogens in Animals' Gut. *Microorganisms* **2020**, *9*, 13. [[CrossRef](#)]
35. Muraro, A.; Werfel, T.; Hoffmann-Sommergruber, K.; Roberts, G.; Beyer, K.; Bindslev-Jensen, C.; Bindslev-Jensen, C.; Cardona, V.; Dubois, A.; duToit, G.; et al. EAACI food allergy and anaphylaxis guidelines: Diagnosis and management of food allergy. *Allergy* **2014**, *69*, 1008–1025. [[CrossRef](#)]
36. Papadopoulos, N.G.; Arakawa, H.; Carlsen, K.H.; Custovic, A.; Gern, J.; Lemanske, R.; Le Souef, P.; Mäkelä, M.; Roberts, G.; Wong, G.; et al. International consensus on (ICON) pediatric asthma. *Allergy* **2012**, *67*, 976–997. [[CrossRef](#)]
37. Lisowska-Myjak, B.; Skarżyńska, E.; Wojdan, K.; Nasierowska-Guttmejer, A. Protein and peptide profiles in neonatal meconium: Classification of meconium proteins. *J. Obstet. Gynaecol. Res.* **2019**, *45*, 556–564. [[CrossRef](#)]
38. Kolmannskog, S.; Marhaug, G.; Haneberg, B. Fragments of IgE Antibodies in Human Feces. *Int. Arch. Allergy Immunol.* **1985**, *78*, 358–363. [[CrossRef](#)]
39. Szépfalusi, Z.; Pichler, J.; Elsässer, S.; van Duren, K.; Ebner, C.; Bernaschek, G.; Urbanek, R. Transplacental priming of the human immune system with environmental allergens can occur early in gestation. *J. Allergy Clin. Immunol.* **2000**, *106*, 530–536. [[CrossRef](#)]
40. Msallam, R.; Balla, J.; Rathore, A.P.S.; Kared, H.; Malleret, B.; Saron, W.A.A.; Liu, Z.; Hang, W.J.; Dutertre, A.C.; Larbi, A.; et al. Fetal mast cells mediate postnatal allergic responses dependent on maternal IgE. *Science* **2020**, *370*, 941–950. [[CrossRef](#)]
41. Lyons, J.J.; Yi, T. Mast cell tryptases in allergic inflammation and immediate hypersensitivity. *Curr. Opin. Immunol.* **2021**, *72*, 94–106. [[CrossRef](#)]
42. Edogawa, S.; Edwinson, A.L.; Peters, S.A.; Chikkamenahalli, L.L.; Sundt, W.; Graves, S.; Gurunathan, V.S.; Breen-Lyles, K.M.; Johnson, S.; Dyer, B.R.; et al. Serine proteases as luminal mediators of intestinal barrier dysfunction symptom severity in IBS. *Gut* **2020**, *69*, 62–73. [[CrossRef](#)]
43. Raithel, M.; Winterkamp, S.; Pacurar, A.; Ulrich, P.; Hochberger, J.; Hahn, E.G. Release of mast cell tryptase from human colorectal mucosa in inflammatory bowel disease. *Scand. J. Gastroenterol.* **2001**, *36*, 174–179. [[CrossRef](#)]
44. Peterson, C.G.B.; Hansson, T.; Skott, A.; Bengtsson, U.; Ahlstedt, S.; Magnussons, J. Detection of local mast-cell activity in patients with food hypersensitivity. *J. Invest. Allergol. Clin. Immunol.* **2007**, *17*, 314–320.
45. Carroccio, A.; Brusca, I.; Mansueto, P.; Soresi, M.; D'Alcamo, A.; Ambrosiano, G.; Pepe, I.; Iacono, G.; Lospalluti, L.M.; Chiusa, L.M.S.; et al. Fecal assays detect hypersensitivity to cow's milk protein and gluten in adults with irritable bowel syndrome. *Clin. Gastroenterol. Hepatol.* **2011**, *9*, 965–971. [[CrossRef](#)]
46. Lettesjö, H.; Hansson, T.; Peterson, C.; Ung, K.-A.; Ringström, G.; Abrahamsson, H.; Simrén, M. Detection of inflammatory markers in stools from patients with irritable bowel syndrome and collagenous colitis. *Scand. J. Gastroenterol.* **2006**, *41*, 54–59. [[CrossRef](#)]
47. Roca, M.; Rodriguez Varela, A.; Donat, E.; Cano, F.; Hervas, D.; Armisen, A.; Vaya, J.M.; Sjölander, A.; Ribes-Koninckx, C. Fecal Calprotectin and Eosinophil-derived Neurotoxin in Healthy Children Between 0 and 12 Years. *J. Pediatr. Gastroenterol. Nutr.* **2017**, *65*, 394–398. [[CrossRef](#)]
48. Willers, M.; Ulas, T.; Völlger, L.; Vogl, T.; Heinemann, A.S.; Pirr, S.; Pagel, J.; Fehlhaber, B.; Halle, O.; Schöning, J.; et al. S100A8 and S100A9 Are Important for Postnatal Development of Gut Microbiota and Immune System in Mice and Infants. *Gastroenterology* **2020**, *159*, 2130–2145. [[CrossRef](#)]
49. Olin, A.; Henckel, E.; Chen, Y.; Lakshmikanth, T.; Pou, C.; Mikes, J.; Gustafsson, A.; Bernhardsson, K.A.; Zhang, C.; Bohlin, K.; et al. Stereotypic Immune System Development in Newborn Children. *Cell* **2018**, *174*, 1277–1292. [[CrossRef](#)]
50. Torow, N.; Marsland, B.J.; Hornef, M.W.; Gollwitzer, E.S. Neonatal mucosal immunology. *Mucosal Immunol.* **2017**, *10*, 5–17. [[CrossRef](#)]
51. Goretzki, A.; Lin, Y.J.; Schülke, S. Immune metabolism in allergies, does it matter?—A review of immune metabolic basics and adaptations associated with the activation of innate immune cells in allergy. *Allergy* **2021**, *76*, 3314–3331. [[CrossRef](#)]
52. Macpherson, A.J.; de Agüero, M.G.; Ganai-Vonarburg, S.C. How nutrition and the maternal microbiota shape the neonatal immune system. *Nat. Rev. Immunol.* **2017**, *17*, 508–517. [[CrossRef](#)]
53. Singh, N.; Gurav, A.; Sivaprakasam, S.; Brady, E.; Padia, R.; Shi, H.; Thangaraju, M.; Prasad, D.P.; Manicassamy, S.; Munn, H.D.; et al. Activation of Gpr109a, receptor for niacin and the commensal metabolite butyrate, suppresses colonic inflammation and carcinogenesis. *Immunity* **2014**, *40*, 128–139. [[CrossRef](#)]
54. Furusawa, Y.; Obata, Y.; Fukuda, S.; Endo, T.A.; Nakato, G.; Takahashi, D.; Nakanishi, Y.; Uetake, C.; Kato, K.; Kato, T.; et al. Commensal microbe-derived butyrate induces the differentiation of colonic regulatory T cells. *Nature* **2013**, *504*, 446–450. [[CrossRef](#)]
55. Atarashi, K.; Tanoue, T.; Ando, M.; Kamada, N.; Nagano, Y.; Narushima, S.; Suda, W.; Imaoka, A.; Setoyama, H.; Nagamori, T.; et al. Th17 Cell Induction by Adhesion of Microbes to Intestinal Epithelial Cells. *Cell* **2015**, *163*, 367–380. [[CrossRef](#)]

56. Vonk, M.M.; Blokhuis, B.R.J.; Diks, M.A.P.; Wagenaar, L.; Smit, J.J.; Pieters, R.H.H.; Garssen, J.; Knippels, J.M.L.; Esch, V.M.A.C.B. Butyrate Enhances Desensitization Induced by Oral Immunotherapy in Cow's Milk Allergic Mice. *Mediat. Inflamm.* **2019**, *2019*, 9062537. [[CrossRef](#)]
57. Roduit, C.; Frei, R.; Ferstl, R.; Loeliger, S.; Westermann, P.; Rhyner, C.; Schiavi, E.; Barcik, W.; Rodriguez-Perez, N.; Wawrzyniak, N.; et al. High levels of butyrate and propionate in early life are associated with protection against atopy. *Allergy* **2019**, *74*, 799–809. [[CrossRef](#)]
58. Barnett, D.J.M.; Mommers, M.; Penders, J.; Arts, I.C.W.; Thijs, C. Intestinal archaea inversely associated with childhood asthma. *J. Allergy Clin. Immunol.* **2019**, *143*, 2305–2307. [[CrossRef](#)]
59. Gomulka, K.; Liebhart, J.; Jaskula, E.; Lange, A.; Medrala, W. The –2549 –2567 del18 Polymorphism in VEGF and Irreversible Bronchoconstriction in Asthmatics. *J. Investig. Allergol. Clin. Immunol.* **2019**, *29*, 431–435. [[CrossRef](#)]