RESEARCH Open Access

Association between oral microbiome diversity and chronic obstructive pulmonary disease in the US population



Xian-xin Xia¹, Chuan-xiang Li¹ and Hong-rong Guo^{1*}

Abstract

Background There is a dearth of population-based studies on the association between the diversity of the oral microbiome and the risk of chronic obstructive pulmonary disease (COPD). The study aims to investigate the association between oral microbiome diversity and COPD.

Methods In this cross-sectional study, data from the National Health and Nutrition Examination Survey (NHANES 2009–2012) were analyzed. The association between the oral microbiome α -diversity and COPD risk was examined via multivariable logistic regression, with Restricted cubic splines revealing potential non-linear trends. The β -diversity disparities between COPD and non-COPD groups were delineated using Principal Coordinate Analysis (PCoA) and Permutational Multivariate Analysis of Variance (PERMANOVA).

Results A total of 6061 participants were included in this study. For α-diversity, the observed ASVs were significantly associated with COPD risk (OR=0.964, 95%Cl: 0.936–0.993, P=0.016). Similarly, Faith's phylogenetic Diversity showed a significant association with COPD risk (OR=0.955, 95%Cl: 0.919–0.993, P=0.020). The Shannon-Weiner index was also associated with COPD risk (OR=0.829, 95%Cl: 0.702–0.981, P=0.029). For β-diversity, PCoA and PERMANOVA analysis showed statistically significant differences in Bray–Curtis, unweighted, and weighted UniFrac distances (all P<0.01) between the COPD and non-COPD groups.

Conclusions Significant differences in oral microbiome α -diversity and β -diversity were found between COPD and non-COPD populations, with α -diversity (observed ASVs, Faith's Phylogenetic Diversity, Shannon-Weiner index) being negatively associated with the risk of COPD.

Keywords Oral microbiome, COPD, Diversity, NHANES

Introduction

Chronic Obstructive Pulmonary Disease (COPD) is a heterogeneous and progressive lung disorder characterized by persistent respiratory symptoms and airflow obstruction due to chronic inflammation of the airways and/or alveoli[1]. It is primarily caused by the complex

interplay of genetic predispositions, such as SERPINA1 mutations, and environmental exposures (like smoking and air pollution from biomass fuels) [1, 2]. Additional risk factors, such as occupational hazards, gender disparities, socioeconomic status, early-life respiratory infections, and history of asthma, all contribute significantly to the global burden of the disease[3–7]. According to the Global Initiative for Chronic Obstructive Lung Disease (GOLD) guidelines, an estimated 10.3% (equating to 391.9 million people) of individuals aged 30–79 years suffer from COPD, with a significant 80.5% residing in low and middle-income countries[8]. By 2050, a 23% rise

*Correspondence: Hong-rong Guo whhxguo@163.com

¹ Department of Respiratory and Critical Care Medicine, The Third Hospital of Wuhan, Wuhan 430030, People's Republic of China



© The Author(s) 2025. **Open Access** This article is licensed under a Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International License, which permits any non-commercial use, sharing, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if you modified the licensed material. You do not have permission under this licence to share adapted material derived from this article or parts of it. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by-nc-nd/4.0/.

in COPD patients is estimated among adults, reaching nearly 600 million patients.

The oral microbiome refers to the diverse community of microorganisms residing in the oral cavity, including bacteria, viruses, fungi, and protozoa. Emerging research has highlighted the significance of the oral microbiome in various diseases beyond the oral cavity, including respiratory diseases[9]. Oral dysbiosis is a significant contributor to pulmonary complications, primarily via the aspiration of oral microflora into the respiratory system, which incites host immune reactions and inflammation [10-12]. Some studies suggested that oral microbiome may play a role in chronic inflammation and the worsening of respiratory symptoms in COPD patients[13, 14], however, there is a dearth of population-based studies on the association between oral microbiome diversity and COPD. Our study aimed to examine the association between oral microbiome diversity (including α -diversity and β -diversity) and the risk of COPD by performing a cross-sectional analysis on a substantial, nationally representative dataset, to enhance the understanding of the potential role of oral microbiome in COPD prevention.

Materials and methods

Study population

National Health and Nutrition Examination Survey (NHANES), with a complex, multistage probability sampling design, is a program conducted by the National Center for Health Statistics (NCHS), aiming to assess the health and nutritional status of adults and children in the United States. There are some methods to provide comprehensive information on various health conditions, including interviews, physical examinations, and laboratory tests. NHANES was approved by the NCHS Institutional Review Board, and informed consent was obtained from all participants.

Data were utilized from NHANES 2009–2010 and 2011–2012 cycles. Participants aged \geq 20 years with detailed information on oral microbiome data and lung function were included. Demographic data, lifestyle factors, and comorbidities were also collected. The flow-chart for participant selection was shown in Fig. 1.

Lung function

We collected spirometry data including forced expiratory volume in 1 s (FEV1), forced vital capacity (FVC), and FEV1/FVC ratio. According to official documents, participants met the criteria were instructed to perform the spirometry test in a standing position. Testing continued until a reproducible spirogram was achieved, until a maximum of eight spirometry curves were obtained, or until the participant could no longer continue. The goal was for participants to achieve three acceptable

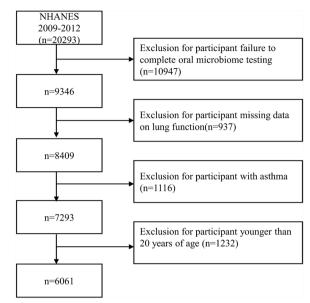


Fig. 1 Flowchart for participant selection

exhalation maneuvers according to the American Thoracic Society (ATS)[15] criteria, where the two highest values for FVC and FEV1 (each from an acceptable forced expiratory maneuver) showed minimal variability: specifically, the two largest FVC values from two acceptable curves should agree within 150 ml, and similarly for the two largest values of FEV1. COPD was defined using the Global Initiative for the GOLD guidelines, with an FEV1/FVC ratio of less than 0.7[16]. In addition, if baseline spirometry indicated the presence of airflow obstruction, bronchodilator testing was performed, and positive responders (an increase of \geq 12% and \geq 200 mL as an absolute value compared with a baseline in either FEV1 or FVC) were excluded from the analysis.

Oral microbiome diversity

Oral microbiome testing was conducted using oral rinse samples from NHANES participants in the 2009–2010 and 2011–2012 cycles. Data processing involved DNA extraction, PCR amplification, sequencing, and bioinformatics analysis to produce relative abundance/read count tables for taxonomic levels from phylum to genus. Microbial sequences were classified into amplicon sequence variants (ASVs) to generate α -diversity metrics (within-sample; observed ASVs, Faith's Phylogenetic Diversity, Shannon-Weiner index, and Simpson Index) and β -diversity metrics (between-sample; unweighted UniFrac, weighted UniFrac, and Bray–Curtis dissimilarity). The α -diversity metrics were calculated using rarefaction values, with random subsampling from 2000 to 10,000 reads[17]. For the final analysis, we used datasets

subsampled to 10,000 reads and averaged the 10 subsamples [18, 19]. The β -diversity was measured between all pairs of samples and represented in a distance matrix, which quantifies the dissimilarity between microbial communities.

Covariates

The following covariates were included, such as age, gender, race, education level, marital status, and PIR (poverty income ratio, ≤ 1.3 ; 1.3-3.5; >3.5), and BMI (body mass index, $<25 \text{ kg/m}^2$, $25-30 \text{ kg/m}^2$, $\geq 30 \text{ kg/m}^2$). Household interviews were conducted to collect data on smoking and drinking behaviors. Diagnoses of hypertension and diabetes were based on self-reported information obtained through questionnaires. The diagnosis of periodontitis was based on the presence of probing depth (PD) $\geq 5 \text{ mm}$ or attachment loss (AL) $\geq 3 \text{ mm}$ at two or more non-adjacent sites[20].

Statistical analysis

The analysis utilized NHANES sampling weights to account for intentional oversampling and ensure the estimates are representative of the US population. For continuous variables, survey-weighted medians (Q1, Q3) and P-values (from survey-weighted linear regression) were reported. For categorical variables, surveyweighted percentages (95% CI) and P-values (from the survey-weighted Chi-square test) were reported. Differences in α-diversity metrics between the COPD and non-COPD groups were assessed, and Pearson correlation analyses were performed to explore associations between α-diversity and lung function (FVC, FEV1, FEV1/FVC). Weighted logistic regression models were used to examine the relationship between α -diversity indicators and COPD risk. Restricted cubic spline (RCS) curves were plotted to visualize the association of the α -diversity with COPD risk. Subgroup analyses were conducted to assess whether the relationship between α -diversity and COPD risk varied by group and interaction analyses were also performed. The β -diversity disparities between COPD and non-COPD groups were delineated using Principal Coordinate Analysis (PCoA) and Permutational Multivariate Analysis of Variance (PERMANOVA). All analyses were performed using R (version 4.2.3), and the P-value < 0.05 was considered statistically significant.

Results

This study included a total of 6061 participants, comprising 5418 non-COPD individuals and 643 COPD individuals. Compared to the non-COPD group, participants in the COPD group were older (55 vs. 41 years), with a higher proportion of males (58.5% vs. 49.7%), a lower proportion of individuals with higher education (58.03%).

vs. 64.5%), and higher proportions of current drinkers (87.01% vs. 81.26%) and smokers (33.97% vs. 14.76%). Regarding lung function, there was no statistically significant difference in FVC between the two groups (P =0.994), but the COPD group had significantly lower FEV1 (2833 ml vs. 3316 ml, P < 0.001) and FEV1/FVC ratios (0.68 vs. 0.81, P< 0.001) compared to the non-COPD group. Regarding α-diversity metrics, the COPD group exhibited significantly lower values in observed ASVs (117.30 vs. 126.10, P = 0.001), Faith's Phylogenetic Diversity (13.53 vs. 14.22, P = 0.003), and the Shannon-Weiner index (4.51 vs. 4.67, P = 0.002), excluding the Simpson Index (0.91 vs. 0.92, P = 0.932), compared to the non-COPD group, as detailed in Table 1. Additionally, the prevalence of comorbidities such as hypertension (P < 0.001), diabetes (P = 0.008), and periodontitis (P < 0.001) were higher in the COPD group than in the non-COPD group.

In Supplementary Figs. 1–4, Pearson correlation analysis was used to evaluate the association between α -diversity indicators and lung function in the total, non-COPD, and COPD populations. These analyses revealed a positive association between α -diversity and lung function (FVC, FEV1, and FEV1/FVC), except for the Simpson Index, which was not associated with FEV1/FVC. Notably, the associations between α -diversity and lung function were stronger in the COPD population compared to the non-COPD population. In the COPD group, four α -diversity indices showed stronger associations with FEV1 (observed ASVs: r = 0.215, P < 0.001; Faith's Phylogenetic Diversity: r = 0.202, P < 0.001; Shannon-Weiner index: r = 0.176, P < 0.001; Simpson Index: r = 0.136, P < 0.001).

Table 2 presented the association between four α-diversity metrics and the risk of COPD. The observed ASVs were significantly associated with COPD risk (unadjusted: OR = 0.938, 95%CI: 0.912-0.964, P < 0.001; model 3: OR = 0.964, 95%CI: 0.936-0.993, P = 0.016). Similarly, Faith's phylogenetic Diversity showed a significant association with COPD risk (unadjusted: OR =0.929, 95%CI: 0.896-0.962, P< 0.001; model 3: OR =0.955, 95%CI: 0.919-0.993, P= 0.020). The Shannon-Weiner index was also associated with COPD risk (unadjusted: OR = 0.775, 95%CI: 0.672-0.894, P < 0.001; model 3: OR = 0.829, 95%CI: 0.702-0.981, P= 0.029). The Simpson Index did not show a statistically significant association with COPD through multiple logistic regression models (all P > 0.05). Additionally, we analyzed the association between α -diversity indicators and the risk of COPD in various subgroups. As shown in Supplementary Figs. 5-8, observed ASVs and Faith's Phylogenetic Diversity were significantly associated with COPD risk, except in the 20-45 age group, the Mexican American

Table 1 Comparison of baseline characteristics between non-COPD and COPD groups, weighted

	Non-COPD (n = 5418)	COPD (n = 643)	P-value
Age (years)	41.00 (30.00,52.00)	55.00 (44.00,62.00)	< 0.001
Gender (%)			0.009
Male	49.70 (47.80,51.61)	58.50 (52.56,64.20)	
Female	50.30 (48.39,52.20)	41.50 (35.80,47.44)	
Race (%)			< 0.001
Mexican American	10.64 (7.48,14.93)	3.57 (2.27,5.58)	
Other Hispanic	6.38 (4.59,8.81)	3.11 (1.93,4.96)	
Non-Hispanic White	62.70 (56.70,68.33)	78.58 (71.56,84.26)	
Non-Hispanic Black	11.82 (9.30,14.91)	8.74 (6.42,11.80)	
Other Races	8.46 (6.90,10.34)	5.99 (3.29,10.67)	
Education level (%)			0.038
High school or less	35.50 (32.10,39.05)	41.97 (34.22,50.14)	
College or more	64.50 (60.95,67.90)	58.03 (49.86,65.78)	
Marital status (%)			0.296
With partner	63.73 (60.74,66.62)	66.46 (61.29,71.26)	
Without partner	36.27 (33.38,39.26)	33.54 (28.74,38.71)	
PIR (%)			0.623
≤ 1.3	22.27 (19.76,24.99)	25.05 (18.97,32.30)	
1.3–3.5	33.54 (30.37,36.86)	33.82 (27.78,40.43)	
> 3.5	44.20 (39.81,48.67)	41.13 (32.81,50.00)	
Drinking status (%)			0.035
Never	9.29 (7.99,10.78)	5.36 (2.80,10.02)	
Ever	9.45 (8.25,10.80)	7.63 (5.60,10.31)	
Now	81.26 (79.09,83.25)	87.01 (82.34,90.58)	
Smoking status (%)			< 0.001
Never	61.49 (58.98,63.93)	31.18 (25.53,37.44)	
Ever	23.76 (21.38,26.31)	34.86 (29.68,40.42)	
Now	14.76 (13.47,16.14)	33.97 (28.71,39.65)	
BMI (kg/m²)	27.39 (24.00,31.90)	27.70 (23.40,31.71)	0.903
Lung function			
FVC (ml)	4097 (3428,4931)	4065 (3351,5106)	0.994
FEV1 (ml)	3316 (2769,3982)	2833 (2231,3430)	< 0.001
FEV1/FVC (%)	0.81 (0.77,0.85)	0.68 (0.64,0.70)	< 0.001
Alpha diversity			
Observed ASVs	126.10 (101.20,155.40)	117.30 (90.60,145.10)	0.001
Faith's Phylogenetic Diversity	14.22 (12.30,16.58)	13.53 (11.31,16.13)	0.003
Shannon-Weiner index	4.67 (4.24,5.04)	4.51 (4.10,4.96)	0.002
Simpson Index	0.92 (0.88,0.94)	0.91 (0.88,0.94)	0.932
Hypertension (%)			< 0.001
No	77.71 (76.05,79.28)	64.08 (59.12,68.76)	
Yes	22.29 (20.72,23.95)	35.92 (31.24,40.88)	
Diabetes (%)			0.008
No	94.05 (93.23,94.77)	89.87 (85.46,93.05)	
Yes	5.95 (5.23,6.77)	10.13 (6.95,14.54)	
Periodontitis (%)			< 0.001
No	62.53 (59.22,65.73)	48.55 (42.56,54.57)	
Yes	37.47 (34.27,40.78)	51.45 (45.43,57.44)	

Bold represents P-value < 0.05

COPD: Chronic Obstructive Pulmonary Disease; PIR, poverty income ratio; BMI, body mass index; FVC, forced vital capacity; FEV1, forced expiratory volume in 1 s; ASVs, Amplicon sequence variants

Table 2 Association between alpha diversity and risk of COPD in logistic regression models, weighted

	OR (95%CI) P-value				
	unadjusted	Model 1	Model 2	Model 3	
Observed ASVs	0.938 (0.912-0.964) < 0.001	0.976 (0.948–1.004) 0.087	0.974 (0.947–1.001) 0.063	0.964 (0.936-0.993) 0.016	
Faith's phylogenetic Diversity	0.929 (0.896–0.962) < 0.001	0.974 (0.939–1.011) 0.164	0.967 (0.932–1.003) 0.069	0.955 (0.919-0.993) 0.020	
Shannon-Weiner index	0.775 (0.672-0.894) < 0.001	0.851 (0.728-0.994) 0.042	0.857 (0.729-1.007) 0.061	0.829 (0.702-0.981) 0.029	
Simpson Index	1.006 (0.872–1.161) 0.932	0.983 (0.848-1.141) 0.825	0.968 (0.824–1.138) 0.694	0.955 (0.810–1.126) 0.586	

Bold represents P-value < 0.05

ASVs, Amplicon sequence variants; PIR, family poverty income ratio; BMI, body mass index; OR, odds ratio; 95% CI, 95% confidence intervals

Model 1: Adjusted for age, gender, race, education level, marital status, and PIR

Model 2: Adjusted for Model 1 plus BMI, drinking status, and smoking status

Model 3: Adjusted for Model 2 plus hypertension, diabetes, and periodontitis

Note: α diversity changes corresponding to the effect values in the regression models are as follows: for observed ASVs, a 10-unit increase; for Faith's phylogenetic Diversity, a 1-unit increase; for the Shannon-Weiner index, a 1-unit increase; and for the Simpson Index, a 0.1-unit increase

population, and other ethnic subgroups. The Shannon-Weiner index did not show an association with COPD risk in these subgroups, nor other Hispanic populations or never-drinkers. For the Simpson Index, an association with COPD risk was observed in some subgroups, including those aged 46–69 years, with an education level lower than high school, with partner, former alcohol drinkers and smokers, and without a history of periodontitis.

We explored the relationship between different α -diversity indices and the risk of COPD by plotting the RCS curves. As shown in Fig. 2, a non-linear association between α -diversity indicators and the risk of COPD was found (observed ASVs: Pnonlinear = 0.021; Simpson Index: Pnonlinear = 0.003). In contrast, Faith's Phylogenetic Diversity and the Shannon-Weiner index exhibited a linear relationship with COPD risk (P overall < 0.001).

As shown in Fig. 3, we compared the differences in β -diversity metrics between the COPD and non-COPD populations using PCoA analysis. To quantify the extent of these differences in oral microbial β -diversity between groups, PERMANOVA analysis was conducted. The differences in β -diversity between the two groups were statistically significant (Bray–Curtis dissimilarity: R^2 = 0.01; unweighted UniFrac distance: R^2 = 0.01; weighted UniFrac distance: R^2 = 0; all P < 0.01).

Discussion

In this study, there were significant differences in oral microbiome α -diversity and β -diversity between COPD and non-COPD populations. The α -diversity indicators (observed ASVs, Faith's Phylogenetic Diversity, Shannon-Weiner index) were significantly lower in COPD than in non-COPD populations. The β -diversity metrics (Bray–Curtis dissimilarity, unweighted UniFrac distance, weighted UniFrac distance) also indicated significant

differences in oral microbiome diversity between the two groups. Further analyses showed that α -diversity was negatively associated with COPD risk, with higher α -diversity associated with lower risk of COPD.

The observed ASVs reflect species richness, Faith's Phylogenetic Diversity captures evolutionary diversity, and the Shannon-Weiner index accounts for both richness and evenness, providing a comprehensive measure of microbiome complexity. Our results indicated that higher α -diversity was associated with a reduced risk of COPD. Specifically, for every 10-unit increase in observed ASVs, the risk of COPD decreased by 3.6%, while each 1-unit increase in Faith's Phylogenetic Diversity and the Shannon-Weiner index was associated with a 4.5% and 17.1% reduction in COPD risk, respectively. These findings suggested that a more diverse oral microbiome may better resist pathogenic invasion, potentially reducing COPD risk. Previous studies have shown that COPD patients with frequent exacerbation exhibit lower microbiome diversity, both in the oral cavity and sputum, compared to those with infrequent exacerbations[21]. Moreover, another study has indicated that reduced microbiome diversity has been associated with frequent acute exacerbations of chronic obstructive pulmonary disease (AECOPD)[22]. At the same time, a multicenter study found that the dysbiosis in microbiome during exacerbations was linked to increased exacerbation severity[23]. A meta-analysis also found that patients with AECOPD had a lower Simpson Index for airway microbiome compared to healthy controls, further suggesting reduced microbiome diversity during acute episodes[24]. Additionally, the sputum microbiome α -diversity was found to be an important predictor of one-year mortality in COPD patients, with non-survivors exhibiting significantly lower diversity. For each 1-unit increase in observed

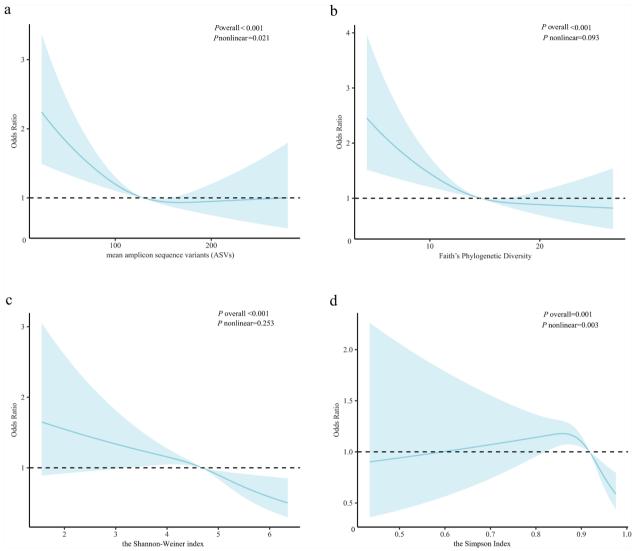


Fig. 2 Restricted cubic spline curves between alpha diversity metrics and COPD risk, **a** observed ASVs; **b** Faith's Phylogenetic Diversity; **c** Shannon-Weiner index; **d** Simpson Index

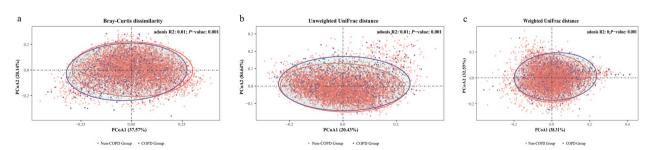


Fig. 3 Comparison of oral microbial beta diversity in COPD and non-COPD populations, **a** Bray–Curtis dissimilarity; **b** unweighted UniFrac distance; **c** weighted UniFrac distance

ASVs, the risk of COPD was reduced by 4%, while a 47% reduction in risk was associated with the Shannon Index and a 38% reduction with Faith's Phylogenetic Diversity[25]. These results underscore a significant association between α -diversity and COPD risk, and emphasize the importance of maintaining a diverse and balanced oral microbiome to improve patient outcomes.

The significant difference in β -diversity was identified between COPD patients and non-COPD patients, which is consistent with previous findings[14, 26, 27]. The magnitude of this variation can be ascertained through the utilisation of diverse analytical approaches, including Bray-Curtis dissimilarity, unweighted and weighted UniFrac distance. This finding signifies that there is a substantial variation in the composition of the oral microbiome community between these two groups. For example, Haldar et al. identified a significant difference in the microbiomes of healthy individuals and COPD patients through the weighted UniFrac distances (P =0.01)[26]. Similarly, several studies based on the Bray-Curtis distance have shown significant β-diversity differences between healthy controls, stable COPD patients, those with AECOPD, and those in recovery[14, 27]. We used PCoA and PERMANOVA analysis to assess the association between β-diversity and COPD, and assessed the models' fit by comparing model R² values. P-values were from the Adonis to test the significance of difference between the COPD and non-COPD groups [28]. We acknowledge that the R² values from the PERMANOVA analyses are indeed low, despite being statistically significant. A plausible rationale for this discovery would be the significant between-sample differences in oral microbiome diversity between individuals with COPD and those without COPD[29]. Notably, the large sample size in our study guaranteed the statistical power to discern subtle variations in oral microbiome, especially those impacted by the fraction of a small number of taxa[18]. The between-sample variation observed in our study may reflect the complex nature of the oral microbiome, where overall microbiome diversity could be more indicative of COPD risk than the presence or absence of specific bacteria. For example, some studies have demonstrated differences in microbiome diversity between healthy individuals and COPD patients, with variations in microbiome diversity linked to disease status[14, 26, 30]. Therefore, these studies have demonstrated distinct oral microbiome diversity between COPD and non-COPD groups, supporting the significant differences in oral microbiome β -diversity observed in the present study.

The reduction in the diversity of the oral microbiome may be a contributing factor in the development of COPD, through mechanisms such as dysbiosis of the oral-lung axis and inflammation[12, 31–33]. The oral and

lung microbiomes exhibit a high degree of compositional similarity, a phenomenon attributable to the anatomical connection between the oral cavity and the respiratory tract[34]. It is hypothesised that oral microorganisms may enter the respiratory system through clinical microaspiration and enter the respiratory tract directly, exacerbating lung flora imbalance and inducing lung injury[12]. The association between the oral microbiome and COPD extends beyond direct microbial transfer, encompassing potential host-pathogen interactions that may influence lung health by instigating systemic inflammation or modulating host immune defense mechanisms[12]. Dysregulation of the oral microbiome may be associated with the progression of COPD, especially during acute exacerbations, as changes in the oral microbiome may exacerbate lung inflammation[31, 35, 36]. Furthermore, oral microbiome have been demonstrated to contribute to the development of COPD by promoting the attachment of respiratory pathogens, influencing the production of inflammatory cytokines, and participating in biofilm formation[32, 37, 38]. Overall, the potential mechanistic pathways between oral microbiome and COPD remain unclear and require further investigation.

In addition, evidence suggests that probiotics and appropriate dental care can have a positive impact on oral and respiratory health; maintaining good oral hygiene and increasing intake of beneficial bacteria is an intervention strategy to reduce the risk of developing COPD[39, 40]. Therefore, healthcare professionals should consider monitoring patients' oral health as an integral part of comprehensive COPD management. Meanwhile, larger prospective studies can be conducted in the future to further validate the association between oral microbiome diversity and the risk of COPD and its associated mechanisms, and to explore the role of specific microbial species in the pathogenesis of COPD.

Strengths and limitations

This study has several strengths. The NHANES database provides a nationally representative dataset with extensive health information, ensuring broad representation and credibility, which enhances the external validity and generalizability of our findings. This study is among the largest population-based analyses examining the link between oral microbiome diversity and COPD risk, with a large sample size that enhances the statistical power and robustness. Our study employed multiple measures of α -diversity and β -diversity to thoroughly evaluate the oral microbiome, enabling us to explore its association with COPD risk from various perspectives and providing a comprehensive understanding of its potential role in COPD development. This study also has several limitations. Firstly, the

cross-sectional design of the study, in conjunction with the collection of data at the same time points, resulted in difficulties in determining the direction of causality between variables. The recommendation for future studies is the implementation of prospective studies or longitudinal analysis, to enhance the comprehension of potential causal mechanisms. Secondly, the NHANES database lacks composition of oral microbiome, preventing us from analyzing the association between it and COPD risk. Thirdly, the cross-sectional nature of the study and the reliance on publicly available datasets, such as NHANES, make it challenging to account for all potential confounders. Although we tried to adjust covariates as much as possible, some variables could be further studied, including diet, exercise, air pollution, and use of inhaled glucocorticoids[1, 41-44].

Conclusions

In this study, we found significant differences in the α -diversity and β -diversity of oral microbiome between COPD and non-COPD populations, with α -diversity negatively associated with the risk of COPD. The findings provide a basis for further exploration of the role of the oral microbiome in COPD.

Abbreviations

AECOPD Acute exacerbations of chronic obstructive pulmonary disease AL Attachment loss ASVs Amplicon sequence variants ATS American Thoracic Society BMI Body mass index

COPD Chronic obstructive pulmonary disease FEV1 Forced expiratory volume in 1 s

FVC Forced vital capacity

GOLD Global Initiative for Chronic Obstructive Lung Disease National Center for Health Statistics

NHANES National Health and Nutrition Examination Survey

PCoA Principal coordinate analysis

PD Probing depth

PERMANOVA Permutational multivariate analysis of variance

Poverty income ratio RCS Restricted cubic spline

Supplementary Information

The online version contains supplementary material available at https://doi. org/10.1186/s12967-025-06553-9.

Additional file 1

Acknowledgements

We sincerely thank NHANES for their invaluable data contribution to our research.

Author contributions

X. Xia: Conceptualization, Methodology, Software, Formal analysis, Data Curation, Writing—Original Draft. C. Li: Investigation, Writing—Review & Editing. H. Guo: Writing—Reviewing and Editing, Project administration, and had primary responsibility for the final content. All authors read and approved the final manuscript.

Funding

This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

Availability of data and materials

The dataset (s) supporting the conclusions of this article are available in the NHANES repository, with data accessible via the CDC's website at http://www. cdc.gov/nchs/nhanes/index.htm.

Declarations

Ethics approval and consent to participate

Approval was granted by the National Center for Health Statistics (NCHS) Research Ethics Review Board.

Consent for publication

Not applicable.

Competing interests

The authors declare no competing interests.

Received: 19 September 2024 Accepted: 4 May 2025 Published online: 17 May 2025

References

- Agustí A, Celli BR, Criner GJ, Halpin D, Anzueto A, Barnes P, et al. Global initiative for chronic obstructive lung disease 2023 report: GOLD executive summary. Am J Respir Crit Care Med. 2023;
- Greene CM, Marciniak SJ, Teckman J, Ferrarotti I, Brantly ML, Lomas DA, et al. α1-Antitrypsin deficiency. Nat Rev Dis Primer. 2016;2:16051.
- DeMeo DL, Ramagopalan S, Kavati A, Vegesna A, Han MK, Yadao A, et al. Women manifest more severe COPD symptoms across the life course. Int J Chron Obstruct Pulmon Dis. 2018;13:3021-9.
- Townend J, Minelli C, Mortimer K, Obaseki DO, Al Ghobain M, Cherkaski H, et al. The association between chronic airflow obstruction and poverty in 12 sites of the multinational BOLD study. Eur Respir J. 2017;49:1601880.
- McGeachie MJ, Yates KP, Zhou X, Guo F, Sternberg AL, Van Natta ML, et al. Patterns of growth and decline in lung function in persistent childhood asthma, N Fngl J Med, 2016;374;1842-52
- Martínez-García MÁ, Faner R, Oscullo G, la Rosa-Carrillo D, Soler-Cataluña JJ, Ballester M, et al. Chronic bronchial infection is associated with more rapid lung function decline in chronic obstructive pulmonary disease. Ann Am Thorac Soc. 2022;19:1842-7
- Paulin LM, Diette GB, Blanc PD, Putcha N, Eisner MD, Kanner RE, et al. Occupational exposures are associated with worse morbidity in patients with chronic obstructive pulmonary disease. Am J Respir Crit Care Med. 2015;191:557-65
- Adeloye D, Song P, Zhu Y, Campbell H, Sheikh A, Rudan I, et al. Global, regional, and national prevalence of, and risk factors for, chronic obstructive pulmonary disease (COPD) in 2019: a systematic review and modelling analysis. Lancet Respir Med. 2022;10:447-58.
- Pathak JL, Yan Y, Zhang Q, Wang L, Ge L. The role of oral microbiome in respiratory health and diseases. Respir Med. 2021;185: 106475.
- Dong J, Li W, Wang Q, Chen J, Zu Y, Zhou X, et al. Relationships between oral microecosystem and respiratory diseases. Front Mol Biosci. 2022;8.
- 11. Garmendia J, Cebollero-Rivas P. Environmental exposures, the oral-lung axis and respiratory health: the airway microbiome goes on stage for the personalized management of human lung function. Microb Biotechnol. 2024:17: e14506.
- 12. Mammen MJ, Scannapieco FA, Sethi S. Oral-lung microbiome interactions in lung diseases. Periodontol. 2000;2020(83):234-41.

- Mao X, Li Y, Shi P, Zhu Z, Sun J, Xue Y, et al. Analysis of sputum microbial flora in chronic obstructive pulmonary disease patients with different phenotypes during acute exacerbations. Microb Pathog. 2023;184: 106335.
- Su L, Qiao Y, Luo J, Huang R, Li Z, Zhang H, et al. Characteristics of the sputum microbiome in COPD exacerbations and correlations between clinical indices. J Transl Med. 2022;20:76.
- Miller MR, Hankinson J, Brusasco V, Burgos F, Casaburi R, Coates A, et al. Standardisation of spirometry. Eur Respir J. 2005;26:319–38.
- Cadham CJ, Oh H, Han MK, Mannino D, Cook S, Meza R, et al. The prevalence and mortality risks of PRISm and COPD in the United States from NHANES 2007–2012. Respir Res. 2024;25:208.
- 17. Oral Microbiome Data Documentation.
- Feng X, Patel EU, White JL, Li S, Zhu X, Zhao N, et al. Association of oral microbiome with oral human papillomavirus infection: a population study of the National Health and Nutrition Examination Survey, 2009–2012. J Infect Dis. 2024;230:726–35.
- Hu Y, Luo J, Xie J, Wang Y, Bao D, Hua L. Exploring the relationship between the oral microbiota and fasting blood glucose level based on the analysis of NHANES data. Panminerva Med. 2023;
- Eke PI, Page RC, Wei L, Thornton-Evans G, Genco RJ. Update of the case definitions for population-based surveillance of periodontitis. J Periodontol. 2012;83:1449–54.
- Pragman AA, Knutson KA, Gould TJ, Isaacson RE, Reilly CS, Wendt CH.
 Chronic obstructive pulmonary disease upper airway microbiota alpha diversity is associated with exacerbation phenotype: a case-control observational study. Respir Res. 2019;20:114.
- Pragman AA, Hodgson SW, Wu T, Zank A, Reilly CS, Wendt CH. Sputum microbiome α-diversity is a key feature of the COPD frequent exacerbator phenotype. ERJ Open Res. 2024;10:00595–2023.
- 23. Wang Z, Singh R, Miller BE, Tal-Singer R, Van Horn S, Tomsho L, et al. Sputum microbiome temporal variability and dysbiosis in chronic obstructive pulmonary disease exacerbations: an analysis of the COPDMAP study. Thorax. 2018;73:331–8.
- Kou Z, Liu K, Qiao Z, Wang Y, Li Y, Li Y, et al. The alterations of oral, airway and intestine microbiota in chronic obstructive pulmonary disease: a systematic review and meta-analysis. Front Immunol. 2024;15:1407439.
- Leitao Filho FS, Alotaibi NM, Ngan D, Tam S, Yang J, Hollander Z, et al. Sputum microbiome is associated with 1-year mortality after chronic obstructive pulmonary disease hospitalizations. Am J Respir Crit Care Med. 2019;199:1205–13.
- Haldar K, George L, Wang Z, Mistry V, Ramsheh MY, Free RC, et al. The sputum microbiome is distinct between COPD and health, independent of smoking history. Respir Res. 2020;21:183.
- Wang J, Chai J, Sun L, Zhao J, Chang C. The sputum microbiome associated with different sub-types of AECOPD in a Chinese cohort. BMC Infect Dis. 2020;20:610.
- Li S, Vogtmann E, Graubard BI, Gail MH, Abnet CC, Shi J. fast.adonis: a computationally efficient non-parametric multivariate analysis of microbiome data for large-scale studies. Bioinform Adv. 2022;2:vbac044.
- Putra P, Hasan H. The influence of digital marketing and content on vacation decisions at Lowita Beach Tourism Malabo Village Kae'E Regency Pinrang. Int J Multicult Multireligious Underst. 2023;10:9.
- Park H, Shin JW, Park S-G, Kim W. Microbial communities in the upper respiratory tract of patients with asthma and chronic obstructive pulmonary disease. PLoS ONE. 2014;9: e109710.
- Wen S, Zhang Z, Ouyang Y, Liu J, Liang Z, Pi Y, et al. The role of oral microbiota in chronic obstructive pulmonary disease. Respiration. 2022;101:859–68.
- Slocum C, Kramer C, Genco CA. Immune dysregulation mediated by the oral microbiome: potential link to chronic inflammation and atherosclerosis. J Intern Med. 2016;280:114–28.
- Gupta A, Saleena LM, Kannan P, Shivachandran A. The impact of oral diseases on respiratory health and the influence of respiratory infections on the oral microbiome. J Dent. 2024;148: 105213.
- Kitsios GD, Sayed K, Fitch A, Yang H, Britton N, Shah F, et al. Longitudinal multicompartment characterization of host-microbiota interactions in patients with acute respiratory failure. Nat Commun. 2024;15:4708.
- Liu J, Ran Z, Wang F, Xin C, Xiong B, Song Z. Role of pulmonary microorganisms in the development of chronic obstructive pulmonary disease. Crit Rev Microbiol. 2021;47:1–12.

- Zhang J, Wu Y, Liu J, Yang Y, Li H, Wu X, et al. Differential oral microbial input determines two microbiota pneumo-types associated with health status. Adv Sci. 2022;9:2203115.
- 37. Shah SA, Ganesan SM, Varadharaj S, Dabdoub SM, Walters JD, Kumar PS. The making of a miscreant: tobacco smoke and the creation of pathogen-rich biofilms. Npj Biofilms Microbiomes. 2017;3:26.
- 38. Li Y, Xing Z, Wang S, Wang Y, Wang Z, Dong L. Disruption of biofilms in periodontal disease through the induction of phase transition by cationic dextrans. Acta Biomater. 2023;158:759–68.
- 39. Wang D, Dai L, Cui Z, Xing W, Huang X, Yang H, et al. Association between periodontal diseases and chronic obstructive pulmonary disease: evidence from sequential cross-sectional and prospective cohort studies based on UK Biobank. J Clin Periodontol. 2024;51:97–107.
- Shen H-T, Fang Y-T, Tsai W-H, Chou C-H, Huang M-S, Yeh Y-T, et al. A lactobacillus combination ameliorates lung inflammation in an elastase/ LPS-induced mouse model of chronic obstructive pulmonary disease. Probiotics Antimicrob Proteins. 2024;
- Urban S, Chmura O, Wątor J, Panek P, Zapała B. The intensive physical activity causes changes in the composition of gut and oral microbiota. Sci Rep. 2024;14:20858.
- Shoer S, Shilo S, Godneva A, Ben-Yacov O, Rein M, Wolf BC, et al. Impact of dietary interventions on pre-diabetic oral and gut microbiome, metabolites and cytokines. Nat Commun. 2023;14:5384.
- 43. Wade WG. Resilience of the oral microbiome. Periodontol. 2000;2021(86):113–22.
- Yang IA, Ferry OR, Clarke MS, Sim EH, Fong KM. Inhaled corticosteroids versus placebo for stable chronic obstructive pulmonary disease. Cochrane Database Syst Rev. 2023;2023:CD002991.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.