



Immunohistochemical Study of Expression of Sohlh1 and Sohlh2 in Normal Adult Human Tissues

Xiaoli Zhang¹, Ruihua Liu², Zhongxue Su³, Yuecun Zhang⁴, Wenfang Zhang¹, Xinyu Liu¹, Fuwu Wang¹, Yuji Guo¹, Chuangang Li⁵, Jing Hao¹*

1 Key Laboratory of the Ministry of Education for Experimental Teratology, Department of Histology and Embryology, School of Medicine, Shandong University, Jinan, China, 2 Department of Ultrasound, Yantai Yuhuangding Hospital, Yantai, China, 3 Department of Hepatobiliary Surgery, Shandong Provincial Hospital Affiliated to Shandong University, Jinan, China, 4 Department of Gynecology and Obstetrics, Nanjing Tongren Hospital Affiliated to School of Medicine of Dongnan University, Nanjing, China, 5 Department of Anesthesiology, The Second Affiliated Hospital to Shandong University, Jinan, China

* haojing@sdu.edu.cn



OPEN ACCESS

Citation: Zhang X, Liu R, Su Z, Zhang Y, Zhang W, Liu X, et al. (2015) Immunohistochemical Study of Expression of *Sohlh1* and *Sohlh2* in Normal Adult *Human* Tissues. PLoS ONE 10(9): e0137431. doi:10.1371/journal.pone.0137431

Editor: Wei Shen, Qingdao Agricultural University, CHINA

Received: June 7, 2015

Accepted: August 17, 2015

Published: September 16, 2015

Copyright: © 2015 Zhang et al. This is an open access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: All relevant data are available in the paper.

Funding: This study was supported by the Natural Science Foundation 1 of Shandong Province (NO. ZR2014HM082), the Shandong Department of Science and Technology Plan Project (NO. 2014GSF118085), and the Shandong Province Outstanding Young Scientist Research Award Fund Project (2011BSE27084). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Abstract

The expression pattern of Sohlh1 (spermatogenesis and oogenesis specific basic helixloop-helix 1) and Sohlh2 in mice has been reported in previous studies. Sohlh1 and Sohlh2 are specifically expressed in spermatogonia, prespermatogonia in male mice and oocytes of primordial and primary follicles in female mice. In this report, we studied the expression pattern of Sohlh1 and Sohlh2 in human adult tissues. Immunohistochemical staining of Sohlh1 and Sohlh2 was performed in 5 samples of normal ovaries and testes, respectively. The results revealed that Sohlh genes are not only expressed in oocytes and spermatogonia, but also in granular cells, theca cells, Sertoli cells and Leydig cells, and in smooth muscles of blood vessel walls. To further investigate the expression of Sohlh genes in other adult human tissues, we collected representative normal adult tissues developed from three embryonic germ layers. Compared with the expression in mice, Sohlhs exhibited a much more extensive expression pattern in human tissues. Sohlhs were detected in testis, ovary and epithelia developed from embryonic endoderm, ectoderm and tissues developed from embryonic mesoderm. Sohlh signals were found in spermatogonia, Sertoli cells and also Leydig cells in testis, while in ovary, the expression was mainly in oocytes of primordial and primary follicles, granular cells and theca cells of secondary follicles. Compared with Sohlh2, the expression of Sohlh1 was stronger and more extensive. Our study explored the expression of Sohlh genes in human tissues and might provide insights for functional studies of Sohlh genes.



Competing Interests: The authors have declared that no competing interests exist.

Introduction

Sohlh1 (spermatogenesis and oogenesis helix-loop-helix 1) and Sohlh2 are transcription factors and play a pivotal role in the transition of germ cells? from primordial to primary follicles and in the differentiation of spermatogonia in mice [1-3]. Sohlh1 was detected preferentially in oocytes but not in other mouse cDNA libraries [1, 2, 4]. Sohlh2 was discovered based on the homology with Sohlh2 in the bHLH domains [3-5]. Later it was found that both genes are specifically expressed in germ cell clusters, primordial and early primary oocytes in females and in prespermatogonia and spermatogonia in males. The expression signals disappeared rapidly as oocytes reached the secondary follicle stage and as type A differentiate to type B spermatogonia. Sohlh1 or Sohlh2 null mice were sterile due to the defect in the differentiation of spermatogonia and oocytes. These findings indicate that Sohlhs play crucial roles in spermatogenesis and oogenesis [2, 5, 6].

Interestingly, Sohlh1 is down-regulated in $Sohlh2^{-/-}$ mice, suggesting that the expression of Sohlh1 and Sohlh2 are correlated and the two genes potentially cross-regulate each other's transcription [2, 5]. Newborn ovaries and testes from $Sohlh2^{-/-}$ mice showed very similar molecular changes as those from $Sohlh1^{-/-}$ mice, and it was suggested that Sohlh1 and Sohlh2 could form heterodimers to regulate spermatogonial and oocyte genes to promote the differentiation of germ cells in vivo [2, 5–7].

However, very little is known about the expression of possible cross-regulating *Sohlh1* and *Sohlh2* in normal *human* tissues. Here we provide evidence that *Sohlh1* and *Sohlh2* are widely expressed in normal adult *human* tissues. Using immunohistichemical staining, we revealed a expression pattern that was different from that in *mice*; *Sohlhs* were expressed more extensively in *human* tissues. As expected, the expression pattern of *Sohlh1* and *Sohlh2* is very similar in normal adult *human* tissues probably due to their functional interrelationship. Our exploration of immunoexpression of *Sohlh1* and *Sohlh2* provides a basis for further study of the roles of *human Sohlh1* and *Sohlh2*.

Materials and Methods

Human tissue samples

Normal paraffin-embedded adult *human* tissues (each type of selected tissue is from 5 people) were obtained from the Department of Pathology in Shandong University Affiliated Qilu Hospital and Shandong Provincial Hospital. All the samples are examined by licensed pathologists and histologists and confirmed to be normal. Prior written and informed consent was obtained from every patient and the study was approved by the ethics review board of Shandong University (Permition NO. 201301031).

Reagents

The rabbit anti-*human* polyclonal *Sohlh1* and *Sohlh2* primary antibodies were purchased from Abcam Inc. (Cambridge, MA, USA). Phosphate buffer solution (PBS) was a product of Gibco (CA, USA). Rabbit SABC immunohistochemical kit and DAB color development kit were purchased from Boster Bio-engineering Limited Company (Wuhan, China)

Immunohistochemical staining

To prepare the samples for immunostaining, 5µm sections were deparaffinized in two changes of fresh xylene in 60°C incubator, each for 30 min, followed by treatment in a series of gradient ethanol (100%X2, 95%X2, 90%, 80%, 70% and then PBS; each for 5 min;) Antigens retrieval were performed through incubation in sodium citrate (pH 6.0) for 30min at 96°C. The slides



were naturally coolled down to the room temperature. The immunohistochemical staining was carried out following the procedures described below: Endogenous peroxidases were blocked with 0.3% hydrogen peroxide for 30 min at room temperature and washed three times in PBS, each for 5min; Normal goat serum was then added and incubated with the sections for 15 min to block the nonspecific binding site; Next, the sections were incubated with primary anti-Sohlh1 and Sohlh2 antibodies overnight at 4°C. For negative control, PBS was used instead of the primary antibody. After (insert times of washes) washes with PBS. The sections were incubated with anti-rabbit secondary antibody at 37°C for 1 hour followed by (insert wash times) washes in PBS. To further enhance the staining, SABC was added to the sections and incubated at 37°C for 1 hour. The chromogen diaminobenzidine (DAB) was prepared freshly by mixing one drop of chromogen to 1 ml of buffer in a mixing vial and added to the sections and incubated for 5 min, the sections were then washed in PBS and counterstained with Harris hematoxylin. At the end of staining, the slides were air dried, cleared in xylene and mounted with Neutral balsam. The staining was viewed and photographed under the Olympus U-LH100HG microscope.

Results

1. Sohlh1 and Sohlh2 expression in adult testis and ovary

To study if the *Sohlh1* and *Sohlh2* expression pattern is the same as that in *mice*, we first stained *Sohlh1* and *Sohlh2* in adult *human* testis and ovary using immonohistochemistry. We found that the expression pattern of *Sohlh1* and *Sohlh2* in ovary and testis is very similar, but the staining of *Sohlh1* is stronger and more extensive.

The *Sohlh1* protein was primarily observed in the nuclei of oocytes in primordial and primary follicles. Among the cells of secondary follicles, *Sohlh1* was found highly expressed in granular layer, theca cells and most stromal cells. Similarly, *Sohlh1* signals were found in almost all seminiferous epithelium except spermatids in testis. Intensive signals were found in Leydig cells and myoid cells around seminiferous tubules as well.

Compared to *Sohlh1*, *Sohlh2* was mainly confined to the nuclei of oocytes and very weak in the cytoplasm of theca cells and granular cells in ovary. In testis, *Sohlh2* was found in spermatogonia and Sertoli cells in seminiferous tubule and Leydig cells outside of seminiferous tubules (Fig 1).

2. ohlh1 and Sohlh2 are expressed in adult muscle tissues

The finding that *Sohlh1* and *Sohlh2* were strongly expressed in smooth muscle fibers of blood vessels in ovary promoted us to investigate the expression of *Sohlh1* and *Sohlh2* in all three kinds of muscle tissues-skeletal muscle, cardiac muscle and smooth muscle. The staining revealed that *Sohlh1* and *Sohlh2* were present in all three kinds of muscle tissues. The expression pattern of *Sohlh1* and *Sohlh2* was very similar. The expression intensity of *Sohlh1* and *Sohlh2* was very strong. *Sohlh1* was localized in nucleus, cytoplasm, or both, while the location of *Sohlh2* is mainly confined in the cytoplasm (Fig 2). To detect if the expression is linked to developmental lineages, we also stained a variety of tissues derived from embryonic mesoderm such as kidney and uterine tube. Our results showed that *Sohlh1* and *Sohlh2* were detected in these tissues (data not shown).

3. Sohlh1 and Sohlh2 are expressed in adult cerebral cortex

As we found that *Sohlh* genes can not only be expressed in ovary and testis but also in muscle tissues, we further studied their expressions in the brain.



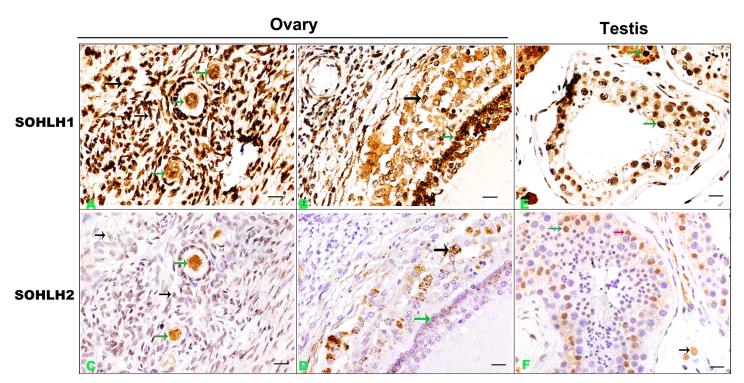


Fig 1. Representative immunohistochemical staining of Sohlh1 and Sohlh2 in ovary and testis (A-F). A and B show expressions of Sohlh1 in ovary. C and D show Sohlh2 expression in ovary. E shows Sohlh1 expression in testis. F shows Sohlh2 expression in testis. Arrows show positive cells and arrows in different color indicate different cell types. Bars indicate 20µm.

doi:10.1371/journal.pone.0137431.g001

The results showed that *Sohlh1* immunostaining was positive in both neurons and neuroglial cells of cerebral cortex. The signals were equally observed in both nucleus and cytoplasm. However, *Sohlh2* signals were mainly confined to the nuclei of the neurons, while they were not detectable in neuroglial cells using immunohistochemical staining method (Fig 3). In addition to the brain, we also detected the expression of *Sohlh* genes in some other tissues derived from embryonic ectoderm including iris, ciliary body, and retina (data not shown).

4. Sohlh1 and Sohlh2 are expressed in epithelia of digestive system and respiratory system

As we investigated *Sohlh* genes expression in mesoderm derived organs and ectoderm derived organs, we then stained *Sohlh* genes in some embryonic endoderm derived tissues. The results showed that *Sohlh1* and *Sohlh2* were present in epithelia of esophagus, lung, liver and pancreas. The expression pattern was similar for *Sohlh1* and *Sohlh2*; but the intensity of *Sohlh1* was much stronger than that of *Sohlh2* and the location of *Sohlh1* was also much diverse than that of *Sohlh2* (Fig 4).

Discussion

Sohlh1 and Sohlh2 are germ cell-specific spermatogenesis and oogenesis basic helix-loop-helix (bHLH) transcription factors $[\underline{1}-\underline{3}]$. Sohlh1 shares 50% identity with Sohlh2 in bHLH region. Mouse Sohlh2 protein shares 50% identity with its human orthologue, with the highest conservation observed in the bHLH domain. Sohlh1 and Sohlh2 were expressed in mouse spermatogonia and in primordial to primary oocytes in embryonic, neonatal or adult mice $[\underline{1}-\underline{3}]$. Loss of



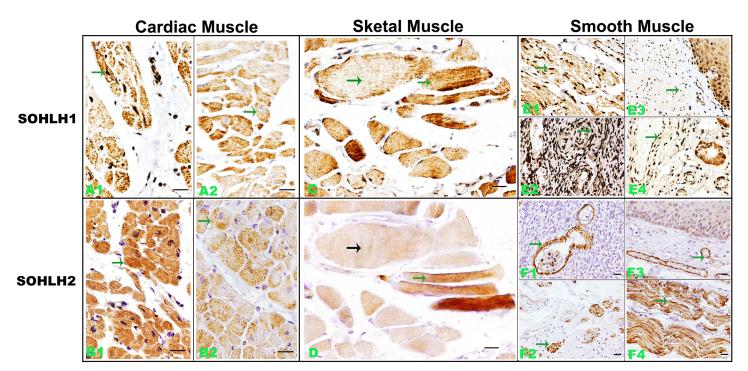


Fig 2. Representative immunohistochemical staining of Sohlh1 and Sohlh2 in muscle tissues (A-F). A and B show Sohlh1 and Sohlh2 expressions in cardiac muscle tissue, respectively, and in different cases (A1-A2 and B1-B2). C and D show Sohlh1 and Sohlh2 expressions in skeletal muscle. E and F show Sohlh1 and Sohlh2 expressions in smooth muscle of different organs (E1-E4 and F1-F4). Arrows show positive cells and arrows in different color indicate different cell types. Bars indicate 20µm.

doi:10.1371/journal.pone.0137431.g002

Sohlh1 or Sohlh2 causes infertility by disrupting spermatogonial differentiation into spermatocytes or ovarian follicle differentiation from primordial to growing follicles [1–2, 5–7]. Sevenday-old testis lacking of Sohlh1 overexpress Sohlh2 [2]. The Sohlh2- null mice downregulated the expression of Sohlh1 indicating an interrelationship between Sohlh1 and Sohlh2 [7]. Sohlh1 and Sohlh2 can form heterodimers or homodimers [7–9]. A Sohlh2/Sohlh1/SP1 ternary complex autonomously and cooperatively regulates Sohlh1 gene transcription during early spermatogenesis and oogenesis [7, 10]. Several other spermatogonial transcriptors could also monitor spermatogenesis by regulating the expression of Sohlh1 or Sohlh2 [11–13].

We studied the expression of *Sohlh1* and *Sohlh2* in normal adult *human* tissues by immuno-histochemical staining and found that they were expressed not only in ovary and testis, but

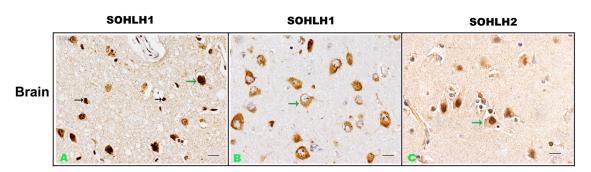


Fig 3. Representative immunohistochemical staining of Sohlh1 and Sohlh2 in brain cortex (A-C). Arrows show positive cells and arrows in different color indicate different cell types. Bars indicate 20µm.

doi:10.1371/journal.pone.0137431.g003



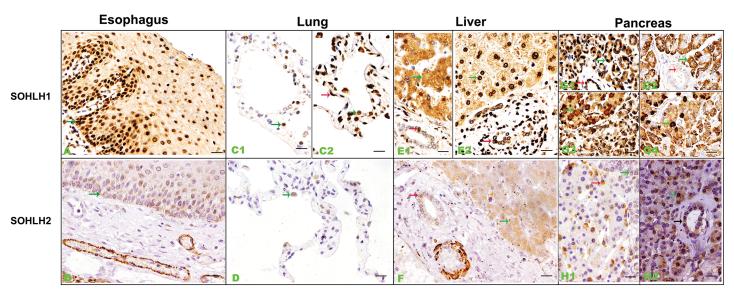


Fig 4. Representative immunohistochemical staining of Sohlh1 and Sohlh2 in epithelia of respiratory and digestive system (A-H). A and B show Sohlh1 and Sohlh2 expressions in esophagus epithelia respectively. C and D show Sohlh1 and Sohlh2 expressions in alveolar cells respectively. E and F show Sohlh1 and Sohlh2 expressions in liver respectively. G and H show Sohlh1 and Sohlh2 expressions in pancreas. Arrows show positive cells and arrows in different color indicate different cell types. Bars indicate 20µm.

doi:10.1371/journal.pone.0137431.g004

also in many other tissues. The expression patterns of *Sohlh1* and *Sohlh2* were very similar, which was not surprising given the previous observations of the relationship between *Sohlh1* and *Sohlh2* in *mice*. The proteins were found in both nucleus and cytoplasm. We were able to detect *Sohlh1* and *Sohlh2* in testis tissues such as spermatogonia, Sertoli cells and Leydig cells and in ovary cells including oocytes, early primary follicles, granular cells, and theca cells in secondary follicles. Because we did not find any oocytes in all of the secondary follicles, it was difficult to tell if *Sohlh1* and *Sohlh2* were expressed in oocytes of secondary follicles.

In the mammalian ovary and testis, progressive activation of primordial follicles or spermatogonia serves as the source of fertilizable ova and sperms, and disorders in the development of primordial follicles or spermatogonia lead to various diseases [14–20]. The polymorphisms of the *Sohlh* 2 gene could be the genetic risk factors for nonobstructive azoospermia (NOA) in the Chinese population [16]. A splice-acceptor site mutation of the *Sohlh1* gene also leads to nonobstructive azoospermia [17]. Novel variants in the *Sohlh2* gene were also found in women with premature ovarian failure (POF) of both Chinese and Serbian [18]. *Sohlh2* was expressed at very low levels in epithelial ovarian cancer (EOC) samples probably by the epigenetic mechanisms [21–25]. These findings strongly suggest the important roles of *Sohlh2* in various diseases and promote us to study the expression patterns of these genes in normal *human* tissues.

The most notable finding in the current study is that *Sohlh1* and *Sohlh2* seem to be expressed ubiquitously and not to be associated with developmental lineages. *Sohlh1* and *Sohlh2* were found in various tissue types like cerebral cortex, muscle tissues and epithelial tissues of esophagus, lung, liver and pancreas. The above studies indicate that *Sohlh1* and *Sohlh2* may play very important roles in normal *human* tissues, and our exploration for the expression of *Sohlh1* and *Sohlh2* provides the basis for further study of functions of *Sohlh1* and *Sohlh2* in relevant academic fields.

The notable difference of expression pattern with that in *mice* is not uncommon. Bonnet A observed the constant expression of *Sohlh2* in *sheep* granular cells and in oocytes during early follicular development until the small antral (SA) stage which is also quite different from that



in *mice* [26]. They speculated that such different *Sohlh2* expression pattern suggests the existence of different mechanisms that need further investigation. The difference also underlines the importance of acquiring expression data from different species and highlights certain species specificities.

As to our knowledge, this study is the first to investigate the expression of *Sohlh1* and *Sohlh2* in normal adult *human* tissues. Like the study in the rhesus monkey, we also found the difference of *Sohlh1* and *Sohlh2* expression between *human* beings and *mice*. For cells in the same section, some signals are confined in the nucleus, *and* some signals are found in the cytoplasm and some signals are found in both nucleus and cytoplasm. In regard to the location of the proteins, Suresh et al. [27] discovered that the spermatogonial *Sohlh1* nucleocytoplasmic shuttling was associated with the initiation of spermatogenesis in the rhesus monkey and suggested that in the monkey, nuclear location of *Sohlh1* is closely associated with spermatogonial differentiation. We surmise that it could also be the nucleocytoplasmic shuttling mechanism of *Sohlh1* and *Sohlh2* that determine the different state (proliferation or differentiation) of the cells in *human* tissues. Consistent with this, our current study confirmed that *Sohlh1* and *Sohlh2* in *human* were localized in both nucleus and cytoplasm.

The expression pattern of *Sohlh1* and *Sohlh2* in ovary is important to the *human* reproductive expert to decipher the critical molecular processes and the complexity of the communication between oocytes, granular cells and theca cells. Similarly, the different expression pattern of *Sohlh1* and *Sohlh2* in testis could be illuminating of scientific researchers in male reproductive field to explore the relationships among spermatogonial cells, Sertoli cells, Leydig cells, or even the myoid cells around seminiferous tubule during spermatogenesis.

We hope our study can be a starting point for further investigation of the function of *Sohlh1* and *Sohlh2* in *human* tissues, not only in the reproductive system but also in various academic fields.

Acknowledgments

We thank Dr. Baojie Li and Tengguo Li very much for the language editing.

Author Contributions

Conceived and designed the experiments: X. Zhang R. Liu JH ZXS Y. Zhang. Performed the experiments: X Zhang W. Zhang X. Liu YG FW. Analyzed the data: X. Zhang R. Liu JH. Contributed reagents/materials/analysis tools: ZXS Y. Zhang R. Liu YG FWW CGL. Wrote the paper: X. Zhang R. Liu JH.

References

- Pangas SA, Choi Y, Ballow DJ, Zhao Y, Westphal H, Matzuk MM, et al. Oogenesis requires germ cellspecific transcriptional regulators Sohlh1 and Lhx8, Proc Natl Acad Sci U S A. 103 (2006) 8090–8095. PMID: 16690745
- Ballow D, Meistrich ML, Matzuk M, Rajkovic A. Sohlh1 is essential for spermatogonial differentiation, Dev Biol. 294 (2006) 161–167. PMID: 16564520
- Ballow DJ, Xin Y, Choi Y, Pangas SA, Rajkovic A. Sohlh2 is a germ cell-specific bHLH transcription factor, Gene Expr Patterns. 6 (2006) 1014–1018. PMID: 16765102
- Rajkovic A, Yan M S C, Klysik M, Matzuk M. Discovery of germ cell-specific transcripts by expressed sequence tag database analysis, Fertil Steril. 76 (2001) 550–554 PMID: <u>11532480</u>
- Choi Y, Yuan D, Rajkovic A. Germ cell-specific transcriptional regulator Sohlh2 is essential for early mouse folliculogenesis and oocyte-specific gene expression, Biol Reprod. 79 (2008) 1176–1182. doi: 10.1095/biolreprod.108.071217 PMID: 18753606



- Hao J, Yamamoto M, Richardson TE, Chapman KM, Denard BS, Hammer RE, et al. Sohlh2 knockout mice are male-sterile because of degeneration of differentiating type A spermatogonia, Stem Cells. 26 (2008) 1587–1597. doi: 10.1634/stemcells.2007-0502 PMID: 18339773
- Toyoda S, Miyazaki T, Miyazaki S, Yoshimura T, Yamamoto M, Tashiro F, et al. Sohlh2 affects differentiation of KIT positive oocytes and spermatogonia, Dev Biol. 325 (2009) 238–248. doi: 10.1016/j.ydbio.2008.10.019 PMID: 19014927
- Barrios F, Filipponi D, Campolo F, Gori M, Bramucci F, Pellegrini M, et al. Sohlh1 and Sohlh2 control Kit expression during postnatal male germ cell development, J Cell Sci. 125 (2012) 1455–1464. doi: 10.1242/jcs.092593 PMID: 22328502
- Suzuki H, Ahn HW, Chu T, Bowden W, Gassei K, Orwig K, et al. Sohlh1 and Sohlh2 coordinate spermatogonial differentiation, Dev Biol. 361 (2012) 301–312. doi: 10.1016/j.ydbio.2011.10.027 PMID: 22056784
- Toyoda S, Yoshimura T, Mizuta J, Miyazaki J. Auto-regulation of the Sohlh1 gene by the Sohlh2/ Sohlh1/SP1 complex: implications for early spermatogenesis and oogenesis, PLoS One. 9 (2014) e101681. doi: 10.1371/journal.pone.0101681 PMID: 25003626
- Zhang T, Murphy MW, Gearhart MD, Bardwell VJ, Zarkower D. The mammalian Doublesex homolog DMRT6 coordinates the transition between mitotic and meiotic developmental programs during spermatogenesis, Development. 141 (2014) 3662–3671. doi: 10.1242/dev.113936 PMID: 25249458
- Matson CK, Murphy MW, Griswold MD, Yoshida S, Bardwell VJ, Zarkower D. The mammalian doublesex homolog DMRT1 is a transcriptional gatekeeper that controls the mitosis versus meiosis decision in male germ cells, Dev Cell. 19 (2010) 612–624. doi: 10.1016/j.devcel.2010.09.010 PMID: 20951351
- Song HW, Wilkinson MF. Transcriptional control of spermatogonial maintenance and differentiation, Semin Cell Dev Biol. 30 (2014) 14–26. doi: 10.1016/j.semcdb.2014.02.005 PMID: 24560784
- Zheng W, Zhang H, Gorre N, Risal S, Shen Y, Liu K. Two classes of ovarian primordial follicles exhibit distinct developmental dynamics and physiological functions, Hum Mol Genet. 23 (2014) 920–928. doi: 10.1093/hmg/ddt486 PMID: 24087793
- Suzumori N, Pangas SA, Rajkovic A. Candidate genes for premature ovarian failure, Curr Med Chem. 14 (2007) 353–357. PMID: 17305537
- Song B, Zhang Y, He XJ, Du WD, Ruan J, Zhou FS, et al. Association of genetic variants in Sohlh1 and Sohlh2 with non-obstructive azoospermia risk in the Chinese population, Eur J Obstet Gynecol Reprod Biol. 184 (2015) 48–52. doi: 10.1016/j.ejogrb.2014.11.003 PMID: 25463635
- Choi Y, Jeon S, Choi M, Lee MH, Park M, Lee DR, et al. Mutations in Sohlh1 gene associate with nonobstructive azoospermia. Hum Mutat. 31 (2010) 788–793. doi: 10.1002/humu.21264 PMID: 20506135
- Qin Y, Jiao X, Dalgleish R, Vujovic S, Li J, Simpson JL, et al. Novel variants in the Sohlh2 gene are implicated in human premature ovarian failure, Fertil Steril. 101 (2014) 1104–1109 doi: 10.1016/j. fertnstert.2014.01.001 PMID: 24524832
- Jagarlamudi K, Rajkovic A. Oogenesis: transcriptional regulators and mouse models, Mol Cell Endocrinol. 356 (2012) 31–39 doi: 10.1016/j.mce.2011.07.049 PMID: 21856374
- Zheng P, Dean J. Oocyte-specific genes affect folliculogenesis, fertilization, and early development, Semin Reprod Med. 25 (2007) 243–251 PMID: <u>17594605</u>
- Woloszynska-Read A, Zhang W, Yu J, Link PA, Mhawech-Fauceglia P, Collamat G, et al. Coordinated cancer germline antigen promoter and global DNA hypomethylation in ovarian cancer: association with the BORIS/CTCF expression ratio and advanced stage. Clin Cancer Res. 17 (2011) 2170–2180. doi: 10.1158/1078-0432.CCR-10-2315 PMID: 21296871
- Zhang H, Zhang X, Ji S, Hao C, Mu Y, Sun J, et al. Sohlh2 inhibits ovarian cancer cell proliferation by upregulation of p21 and downregulation of cyclin D1. Carcinogenesis. 35 (2014) 1863–1871. doi: 10.93/carcin/bgu113 PMID: 24858206
- 23. Pan B, Chao H, Chen B, Zhang L, Li L, Sun X, et al. DNA methylation of germ-cell-specific basic helix-loop-helix (HLH) transcription factors, Sohlh2 and Figlα during gametogenesis, Mol Hum Reprod. 17 (2011) 550–561. doi: 10.1093/molehr/gar017 PMID: 21427160
- Shen L, Kondo Y, Guo Y, Zhang J, Zhang L, Ahmed S, et al. Genome-wide profiling of DNA methylation reveals a class of normally methylated CpG island promoters, PLoS Genet. 3 (2007) 2023–2036. PMID: 17967063
- 25. Lim EJ, Choi Y. Transcription factors in the maintenance and survival of primordial follicles, Clin Exp Reprod Med. 39 (2012) 127–131. doi: 10.5653/cerm.2012.39.4.127 PMID: 23346521
- 26. Bonnet A, Bevilacqua C, Benne F, Bodin L, Cotinot C, Liaubet L, et al. Transcriptome profiling of sheep granulosa cells and oocytes during early follicular development obtained by laser capture microdissection, BMC Genomics. 12 (2011) 417 doi: 10.1186/1471-2164-12-417 PMID: 21851638



 Ramaswamy S, Razack BS, Roslund RM, Suzuki H, Marshall GR, Rajkovic A, et al. Spermatogonial Sohlh1 nucleocytoplasmic shuttling associates with initiation of spermatogenesis in the rhesus monkey (Macaca mulatta), Mol Hum Reprod. 20 (2014) 350–357. doi: 10.1093/molehr/gat093 PMID: 24324034