

# State modulation in spatial networks with three interneuron subtypes

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## Abstract

Several inhibitory interneuron subtypes have been identified as critical in regulating sensory responses. However, the specific contribution of each interneuron subtype remains uncertain. In this work, we explore the contributions of cell-type specific activity and synaptic connections to dynamics of a spatially organized spiking neuron network. We find that the firing rates of the somatostatin (SOM) interneurons align closely with the level of network synchrony irrespective of the target of modulatory input. Further analysis reveals that inhibition from SOM to parvalbumin (PV) interneurons must be limited to allow gradual transitions from asynchrony to synchrony and that the strength of recurrent excitation onto SOM neurons determines the level of synchrony achievable in the network. Our results are consistent with recent experimental findings on cell-type specific manipulations. Overall, our results highlight common dynamic regimes achieved across modulations of different cell populations and identify SOM cells as the main driver of network synchrony.

## Introduction

As animals navigate the environment, their nervous systems process and react to an ongoing bombardment of sensory information. Internal factors such as motivation, attention, expectations, and arousal strongly impact animals' perception, behavior and decision-making [1, 2, 3, 4]. Inhibitory neurons play an essential role in modulating the information processing and communication in cerebral cortex by tuning cortical oscillations, regulating the time window in which external inputs elicit cortical responses, and modifying the response gain of their excitatory counterparts [5, 6, 7]. Inhibitory neurons, however, cannot be considered as a homogeneous population, but instead exhibit differences in morphology, connectivity, and biophysical properties [8, 9]. Differences in molecular markers distinguish three non-overlapping inhibitory interneuron subtypes: parvalbumin (PV), somatostatin (SOM), and vasoactive intestinal peptide (VIP) expressing neurons. These interneuron subtypes are differentially targeted by neuromodulators and cortical feedback projections [9, 10, 11], and are thought to be involved in the modulation of neural population responses by brain state. Arousal and locomotion state of an animal have been shown to exert diverse influences on the firing rates of interneuron subtypes [12, 13, 14] and to strongly impact the synchrony level of neural population responses [15, 16]. However, the functional role of each interneuron subtype remains unclear.

Advancements in optogenetic techniques enable the use of cell-type-specific stimulation and suppression to study the causal contributions to circuit dynamics by each cell type. Prior work demonstrated diverse effects on cortical firing rates and oscillations elicited by manipulating different target cell classes within cortical microcircuits [6, 10, 17, 18, 19, 20]. Stimulating PV neurons periodically enhances the oscillatory

39 power of the local field potential (LFP) over the gamma frequency range [21, 22]. This is consistent with  
40 previous theories where PV neurons are instrumental in generating gamma oscillations, partly due to their  
41 strong reciprocal connections with the excitatory neurons [23]. However, recent work suggests that SOM  
42 neurons are involved in oscillations in low gamma/beta frequency range (20-40 Hz), while suppressing  
43 PV neurons increases the spectral power of the LFP overall [18, 24]. Suppressing SOM neurons also  
44 reduces the coherence between distant neural ensembles [24], consistent with their broad integration of  
45 lateral excitatory inputs [25]. Stimulating VIP neurons increases the response gain of excitatory neurons,  
46 presumably through the disinhibitory pathway via SOM neurons [26]. Silencing VIP neurons reduces the  
47 sensitivity of excitatory neurons to stimulus context [27] and increases the detection threshold for small  
48 visual stimuli [28]. Despite the proliferative experimental findings, the network mechanisms underlying the  
49 observed changes in neural activity remain elusive, due to the intrinsic nonlinearity of the highly recurrently  
50 connected networks to which all of these cell types belong. Manipulation of one cell type leads to changes  
51 in the activity of the other cell types; however, experimenters typically observe the activity of all neurons  
52 indiscriminately or label one cell type at a time (but see [29, 30]). Therefore, computational models are  
53 needed to parse out the interactions between excitatory neurons and the three interneuron subtypes.

54 Previous models that incorporate multiple interneuron subtypes mostly focus on modulations of firing  
55 rates and do not consider impacts on network synchrony or correlations in neural activity [31, 32, 33, 34, 35].  
56 Some models have suggested that PV and SOM neurons contribute to oscillations of different frequencies  
57 [36, 37]. However, these models are small networks or rate models and do not consider the spatial depen-  
58 dence of synaptic connections. In this work, we studied state modulation in spatially structured spiking  
59 neuron networks including multiple interneuron subtypes. Our past work has shown that such models can  
60 reproduce the irregular and weakly correlated neural population activity commonly observed in cortical  
61 recordings [38]. We applied modulatory input to neurons of each cell type and analyzed the resulting  
62 changes in firing rates and network synchrony. We found that the pattern of activity changes resulting  
63 from activation of excitatory (E) or PV neurons is distinct from that due to activation of SOM or VIP  
64 neurons. Strikingly, SOM firing rates closely aligned with levels of network synchrony across all modulation  
65 cases. We further identified that stronger SOM→E than SOM→PV inhibition is important for maintaining  
66 a weakly synchronized dynamical regime, and that the interaction between E and SOM neurons is essen-  
67 tial for enhancing network synchrony. Our work emphasizes the uniquely critical role of SOM neurons in  
68 regulating the dynamical state of cortical networks.

## 69 Results

70 We developed a spatially-extended network model that includes one E population and three distinct in-  
71 hibitory interneuron populations: PV, SOM, and VIP. Each neuron is modeled as a spiking exponential  
72 integrate-and-fire (EIF) unit [39]. The synaptic connection patterns among the four neuron populations  
73 are constrained by anatomical and physiological data from mouse visual cortex (Figure 1A) [40, 41]. In  
74 particular, we assume there are no reciprocal connections among SOM neurons or among VIP neurons;  
75 VIP neurons mainly inhibit SOM neurons, in what is believed to be an important disinhibitory pathway  
76 [26]; and feedforward inputs only target E and PV neurons [25]. Neurons are randomly distributed on a  
77 two-dimensional plane and synaptic connection probability between neurons decays with distance (Figure  
78 1B; Equation 5). The spatial structure of the network allows for rich spatiotemporal activity patterns,  
79 such as propagating waves and spatiotemporal chaos, with population statistics consistent with cortical  
80 recordings (Figure 1C, S2; Ref [38, 42]). Connections to and from the SOM cells have a larger spatial

81 footprint compared to other connections, which is thought to be involved in surround suppression in visual  
82 cortex [25, 43]. The synaptic timescales of inhibitory connections from SOM and VIP neurons are slower  
83 than that of connections from PV neurons, which is in turn slower than that of excitatory connections,  
84 constrained by physiological data from mouse visual cortex [44]. The network has a total of 50,000 neu-  
85 rons comprising 40,000 E, 4,000 PV, 4,000 SOM, and 2,000 VIP neurons, with the population size ratios  
86 following anatomical data from mouse cortex [45].

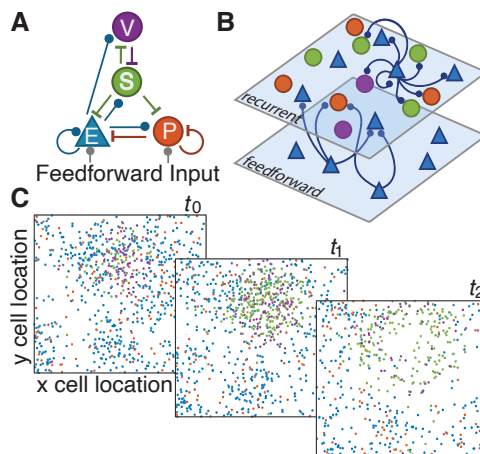


Figure 1: General model scheme and example dynamics. (A) Default network circuit diagram shows excitatory connections in blue (lines with circles) and inhibitory connections (T-lines) in other population-specific colors. (B) The model comprises one recurrent layer with one excitatory population and three inhibitory populations connected as in (A) and a feedforward layer, modeled as independent Poisson units, that provides excitatory input to E and PV neurons. Connection probability decreases with pairwise distance, as is illustrated schematically for E cells here. (C) Three consecutive spike raster snapshots, where a dot with a cell-type-specific color indicates that the neuron at spatial position  $(x, y)$  fired within 1 ms of the time stamp. In this example, local activity of E neurons ( $t_0$ ) recruits more activity of SOM neurons at a later time point ( $t_1$ ), which in turn suppresses the activity of all other neuron populations ( $t_2$ ).

## 87 Network transitions through three dynamical states under variation of cell-type specific 88 inputs

89 To begin our investigation, we apply a static input to each population in the model, one at a time, and  
90 examine network dynamics across a range of input levels. We find that across all input targets, the behavior  
91 of the network transitions through the same three distinct activity patterns, which we call the subcircuit  
92 asynchronous (SA), weakly synchronous (WS) and strongly synchronous (SS) states (Figure 2). We first  
93 define each state and then examine the effects of input modulation separately for each population. In the SA  
94 state (Figure 2Ai-iv; Supplemental video 1), the network behaves essentially as a two population subcircuit  
95 composed of interacting E and PV neurons, while SOM and VIP activity is nearly, if not completely, silent.  
96 The E population is the only excitatory source of input to SOM and VIP. In the SA state, E neurons are  
97 unable to consistently drive SOM and VIP neurons over their respective spiking thresholds (Figure 2Aii-  
98 iii). E neurons exhibit little synchronization or organized activity, as indicated by the near-zero levels of  
99 average spike count correlations between E neuron pairs (Figure 2Aiv). The average spike train coherence  
100 among E or among PV neurons is also low with a peak above 25 Hz (Figure 2Av).

101 Within the WS state, all four populations actively fire (Figure 2Bi-iv). PV neurons exhibit the highest  
102 firing rates, with the other three populations moderately active (Figure 2Bi-iii). The spiking activity of

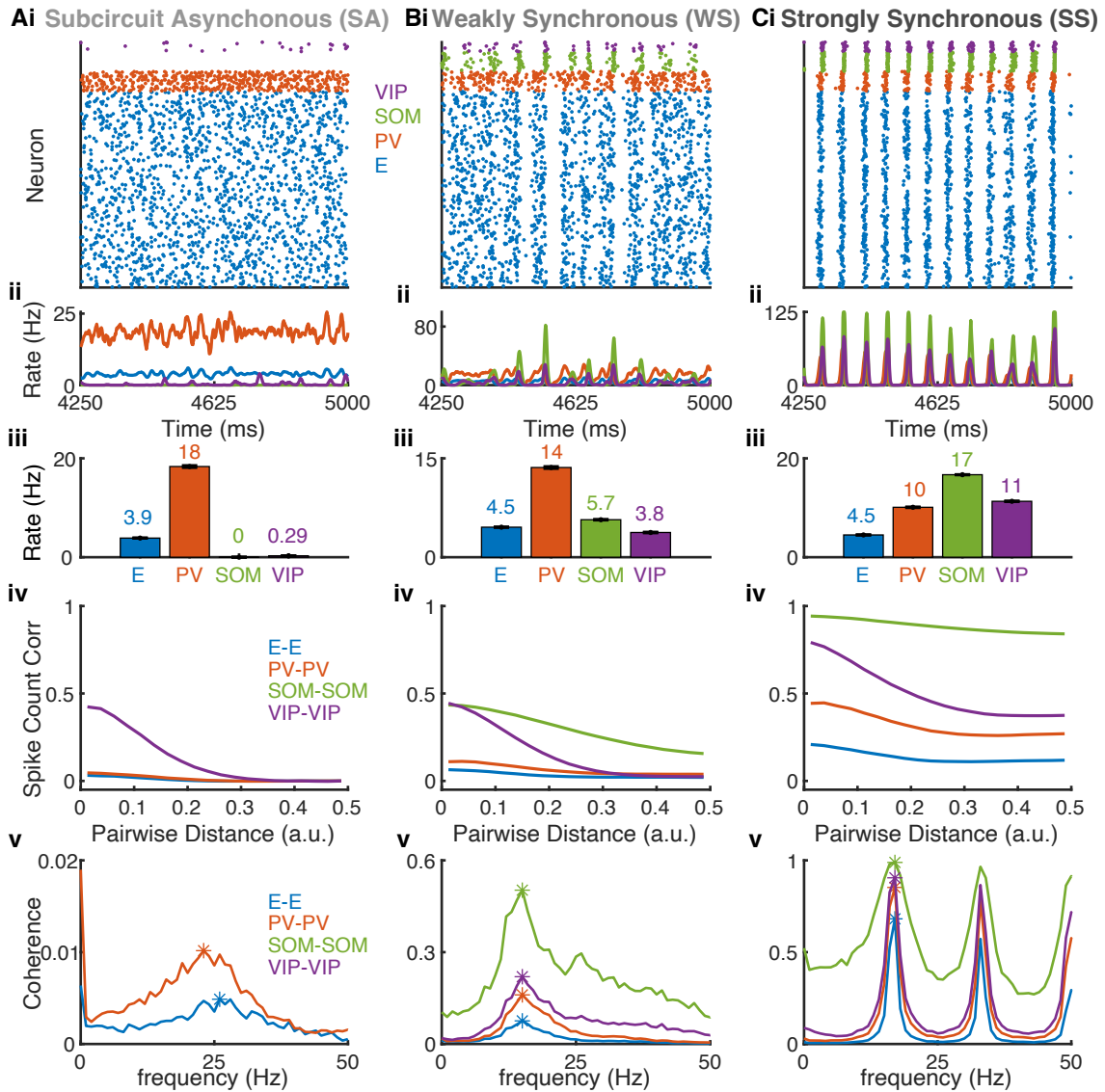


Figure 2: Three representative network states, SA state (Ai-v), WS state (Bi-v) and SS state (Ci-v). Row (i): Spike raster of a subsample of each of the four populations: 400 E (blue), 40 PV (red), 40 SOM (green), and 20 VIP (purple) neurons. The number of neurons of each neuron population shown in the rasters is proportional to the population size. Row (ii): Population-averaged firing rates over the same time course as the spike rasters in row (i). Row (iii): Mean firing rates averaged over neurons and over time for each population. The number on top of each bar is the value of the mean firing rate. Error bars are standard error of mean (SEM). Row (iv): Average spike count correlations (see Methods) as a function of distance for neuron pairs within each population. Row (v): Average pairwise coherence of spike trains (see Methods) versus frequency for neuron pairs within each population. The asterisks mark the maximum coherence over non-zero frequencies. Note the different y-axis scales across panels.

103 E and PV neurons is largely asynchronous, interspersed with brief coordinated periods of silence (Figure  
104 2Bi; Supplemental video 2). The silent periods in E and PV are preceded by synchronous bouts of rapid  
105 firing in SOM and VIP neurons (Figure 2Bi,Bii). The spike count correlations and coherence of SOM-  
106 SOM and VIP-VIP neuron pairs are larger than those of E-E and PV-PV neuron pairs (Figure 2Biv-v),  
107 consistent with experimental observations in mouse cortex [44, 46]. The correlation between SOM neuron  
108 pairs persists over larger distances than those of other populations, due to the larger spatial footprints of  
109 SOM neuron connections, which is also consistent with cortical recordings [46].

110 The SS state exhibits highly synchronized and oscillatory activity in all populations (Figure 2Ci-iv).  
111 Patterns of firing initially begin with a low number of E and PV spikes, which recruit many more E and  
112 PV neurons to fire, thereby activating a large portion of SOM and VIP neurons (Supplemental video 3).  
113 The elevated firing rates of all three inhibitory populations (Figure 2Cii-iii) supply a significant amount of  
114 inhibitory current, ultimately silencing all neurons until enough feedforward input accumulates to excite  
115 E and PV neurons and to cause the cycle to repeat (Figure 2Ci). Pairwise spike count correlations are  
116 relatively large within each population and only slightly decrease with distance (Figure 2Civ). The average  
117 coherence of each neuron population shows a dominant peak close to 1 at around 20 Hz (Figure 2Cv). Since  
118 spike count correlations depend on the choice of time window for calculating spike counts, the correlation  
119 value can be misleadingly low when the time window coincides with the multiples of the oscillation period.  
120 For this reason, we hereafter use the maximum coherence to measure the level of network synchrony.

121 Comparing the effects of varying a static input current applied to different neuron populations reveals  
122 that external inputs to different targets modulate population dynamics across similar states. Specifically,  
123 we see that activating E neurons increases coherence in all populations (Figure 3A). As input to E increases,  
124 network activity transitions from the SA to the WS to the SS state. The transition is marked by non-  
125 monotonic changes in firing rates in E and PV populations (Figure 3Ai). The counterintuitive decrease of  
126 firing rate with increasing external input to E is due to the enhanced inhibition from SOM neurons. On the  
127 other hand, the firing rates of SOM neurons continue to rise despite reduced mean excitation from E due  
128 to the large increase in the temporal variance of the synaptic input currents they receive (Supplemental  
129 Figure S1Ai-ii).

130 In contrast, we see that increasing the external input to PV neurons results in a reverse order of state  
131 transitions compared to the case with input to E (Figure 3B). Activating PV neurons decreases coherence  
132 in population spiking, moving the network from the SS to the WS to the SA state. The firing rates of E  
133 and PV neurons again exhibit non-monotonic changes, as in the case with input to E neurons (Figure 3A).  
134 In the WS state, driving PV leads to increases in E firing rate because of the reduction in the inhibition  
135 from SOM neurons. The firing rates of SOM drop in the presence of increases in mean excitation due to  
136 the large reduction in the variance of input current (Supplemental Figure S1Bi-ii).

137 Driving SOM neurons increases population coherence and moves the network from the SA to the WS  
138 to the SS state, similar to the effects observed when driving E neurons (Figure 3C). However, the firing  
139 rates of E and PV neurons monotonically decrease as SOM neurons become more active (Figure 3Ci), in  
140 contrast to the non-monotonic changes that result when driving E or PV neurons (Figure 3Ai,Bi). VIP  
141 neurons become suppressed when SOM neurons are sufficiently activated due to inhibition from SOM to  
142 VIP (Figure 3Ci). In contrast, VIP and SOM firing rates co-vary in the same direction when input is  
143 applied to E or PV neurons (Figure 3Ai,Bi).

144 Lastly, varying the external input to VIP neurons yields similar changes to those arising with PV input  
145 variations, but is unable to induce all three of the network states that we have identified (Figure 3D). When  
146 input to VIP is strong, inhibition from VIP to SOM shuts down SOM activity and firing rates of E and PV

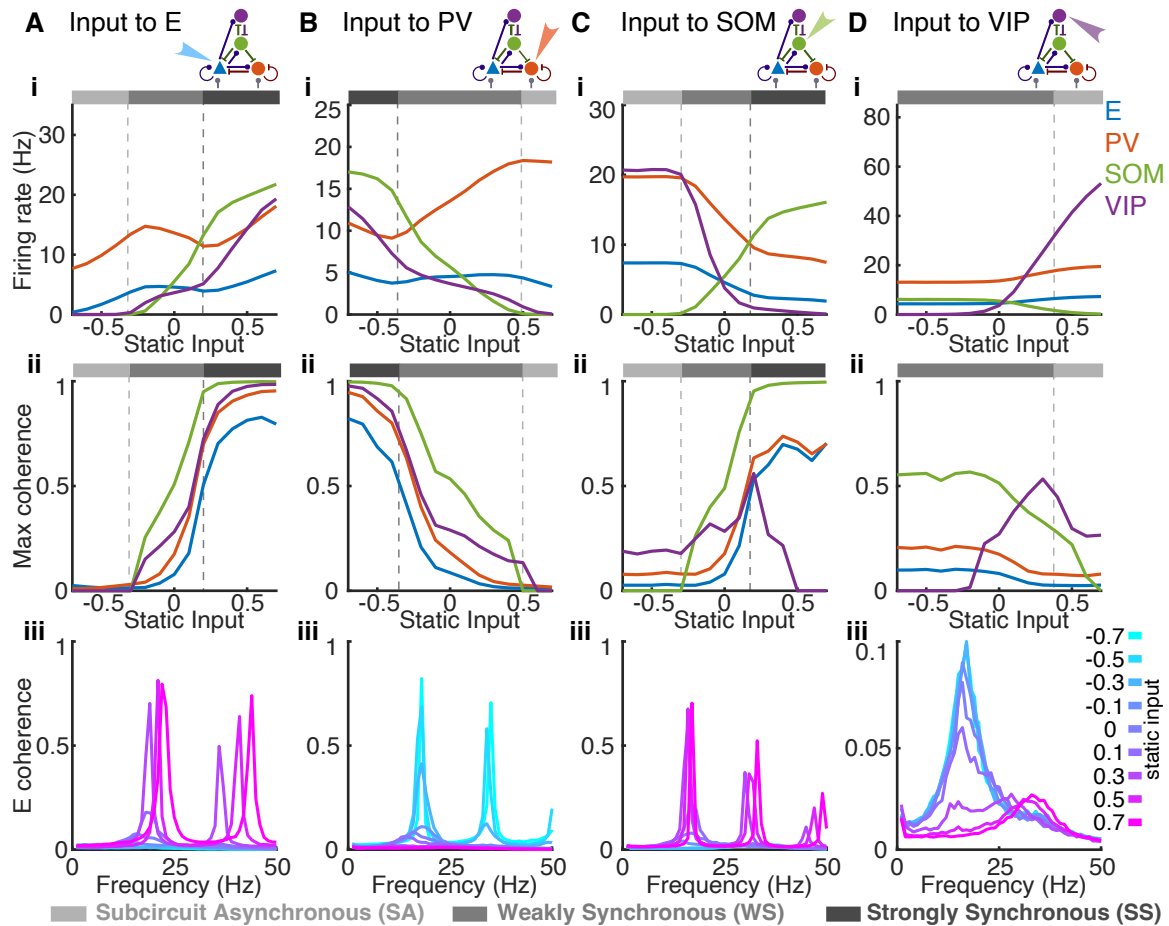


Figure 3: Cell type specific inputs change population firing rates and coherence in distinct ways. Static input is applied to all neurons in one of the four populations: (A) E, (B) PV, (C) SOM or (D) VIP neurons. Row (i): Average population firing rates of each population as a function of static input value. Grey-scale bars above each plot represent network activity state at the corresponding input value (SA: light grey; WS: moderate grey; SS: dark grey). Note the differences in vertical axis scales across panels. Row (ii): Maximum coherence in each population as a function of static input value. Row (iii): E population coherence as a function of frequency for several static input values. Note the distinct vertical axis scale in panel D(iii).

147 neurons increase slightly due to disinhibition. With SOM silenced and VIP having no synaptic connections  
148 to PV and E neurons, the network behaves asynchronously and effectively like a two population E-PV  
149 subcircuit, thus adopting the SA state. When input to VIP neurons is reduced, the drop in inhibition from  
150 VIP to SOM means that SOM starts to fire and VIP firing decreases. The network transitions from the  
151 SA to the WS state, and stays in the WS state once VIP neurons are fully suppressed (Figure 3Di, ii).  
152 Therefore, modulating VIP neurons does not lead the system to the pathological SS state, which makes the  
153 VIP neurons a ideal candidate for moderate state modulations. In addition, we observe that the frequency  
154 of peak coherence within the E population transitions (Figure 3Diii) as a result of changes in static input  
155 to VIP neurons. Activating VIP results in peak frequencies occurring at around 30 Hz, but as VIP reduces  
156 its activity due to reduced input and SOM begins to fire, the peak frequency shifts to approximately 20  
157 Hz, with higher peak levels of E coherence.

158 We find that the spatial structure in the network is important for gradual state transitions, consistent  
159 with the observations in our previous work [38]. In networks with no spatial structure, meaning that the  
160 connection probability between two neurons does not depend on distance, we observe sharp transitions  
161 between SA and SS states as external input varies (Supplemental Figure S2Ai-iv). Therefore, the spatial  
162 structure of the network contributes to maintaining a WS state over a range of input values.

163 In all input cases, we observe similar network state for a given input in multiple simulation runs with  
164 random initial conditions. We also did not observe bistability when comparing network activity with  
165 gradually changing (increasing or decreasing) input (Supplemental Figures S7A,B). Based on the absence  
166 of hysteresis effects, we infer that the transition from the SA to the WS state likely occurs through a  
167 supercritical Hopf bifurcation.

## 168 **Firing rates of SOM neurons co-vary with network synchrony**

169 To directly compare how firing rates and network synchrony change together as input to each neuron  
170 population varies, we summarize the results of four input cases from the previous section with phase plots  
171 of the maximum coherence of the E population versus the firing rate of each neuron population (Figure  
172 4). On these phase plots, each trajectory corresponds to a path of network state transitions as input to  
173 a specific neuron population varies. The arrows represent the directions of transition as input increases  
174 value. We use the maximum coherence of the E population to represent the overall network synchrony  
175 level for two reasons: first, E neurons make up the majority of the total neuron population (80%) and are  
176 recorded most commonly in experimental research and secondly, the coherence of all four populations tend  
177 to vary together, other than some exceptional results in VIP neurons (Figure 3A-Dii).

178 When plotting E population coherence versus E population firing rates, we find that for the cases of  
179 input to E or PV neurons, the network evolves along a common path, with opposite directions of traversal  
180 resulting from similar changes in static input levels (Figure 4A). Similarly, we obtain a common path for  
181 the cases of input to SOM or VIP neurons, but this common path differs from that observed with input  
182 to E or PV neurons. Previously we found that input to VIP never resulted in SS activity (Figure 4Dii),  
183 which explains why the VIP curve (purple) ends at a relatively low coherence value. Within the common  
184 path shared by E and PV stimulation, there exist three regimes: a lower branch where coherence is low ( $\sim$   
185 0) and input changes only affect firing rate (the changing markers on the  $x$ -axis), an upper branch where  
186 coherence remains high ( $> 0.5$ ) over a range of high firing rates, and a transition between the low and  
187 high coherence plateaus, across which coherence changes significantly while firing rates remain relatively  
188 unchanged. These regimes align with the network activity: the lower branch is the SA state, the upper  
189 branch is the SS state, and the transition is the WS state. What is especially remarkable is the precise

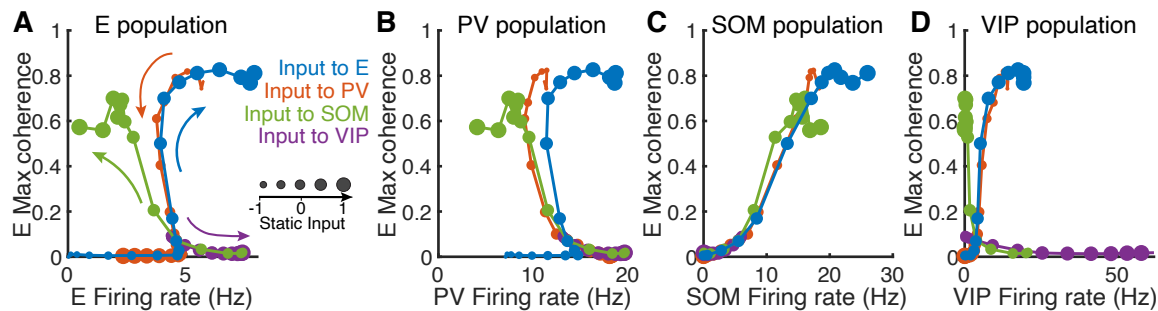


Figure 4: Modulation patterns of population firing rates and E population coherence. Levels of external input are indicated by individual circular markers, where decreasing marker size signifies decreased static input (i.e., progressing from activating to suppressing the target). Colors in each panel indicate which population receives the varying input and colored arrows (A) indicate the direction of increasing input (following the direction of increasing marker size along a single colored curve). Each panel depicts E coherence versus the population-averaged firing rate of one neuron population: (A) E, (B) PV, (C) SOM, and (D) VIP.

190 overlap of pairs of paths, along with the alignment of all paths during the transition region (i.e. the WS  
 191 state), which suggests that the network structure strongly constrains network dynamics. The modulation  
 192 patterns in the full network are distinct from those in the isolated E-PV subcircuit, where firing rate and  
 193 coherence levels tend to vary in the same direction and monotonically as input level varies (Supplemental  
 194 Figure S3).

195 Similarly, curves of E coherence versus PV firing rate overlap significantly for E and PV input cases,  
 196 as do the curves for SOM and VIP input cases (Figure 4B). When comparing the E population coherence  
 197 and SOM population rates (Figure 4C), however, the paths corresponding to application of static input to  
 198 all four target populations largely overlap, no longer showing a distinction across input targets (aside from  
 199 the direction of modulation across states as indicated by changes in marker sizes). Lastly, VIP firing rates  
 200 compared to E coherence for all input cases (Figure 4D) features the dichotomy of trajectories generated  
 201 by inputs to E and PV versus paths from inputs to SOM and VIP (as also observed in Figure 4A, B).

202 Overall, we see that applying excitatory input to E or SOM neurons or inhibitory input to PV or VIP  
 203 neurons tends to increase coherence, although this change is accompanied by distinct changes in firing rates  
 204 for most cell populations. Comparing SOM population rates with the coherence within the E population,  
 205 however, reveals that the two quantities increase together, in a stereotyped way, in all input cases (Figure  
 206 4C). This consistency between E coherence and SOM activity across all input targets leads us to postulate  
 207 that SOM activity plays a central role in dictating the level of network synchrony.

## 208 Strong SOM inhibition to PV drives synchrony

209 We next investigate how synaptic connection strengths in the network shape the modulation patterns of  
 210 network states induced by cell-type specific inputs. Building on our prior observation of the alignment of  
 211 SOM firing rate with network synchrony, we focus on the strengths of connections projecting onto or from  
 212 SOM neurons, specifically SOM→E, SOM→PV, and E→SOM (next section) synapses. Since the influence  
 213 of VIP's inhibitory outputs is restricted to SOM neurons, varying the connection strengths between VIP  
 214 and SOM neurons has little effect on the input-induced transition patterns (Supplemental Figures S4 and  
 215 S5).

216 We find that SOM→E connections are important for generating the non-monotonic changes in E and  
 217 PV firing rates along the transition paths induced by varying input to E or PV neurons (Figures 3Ai,



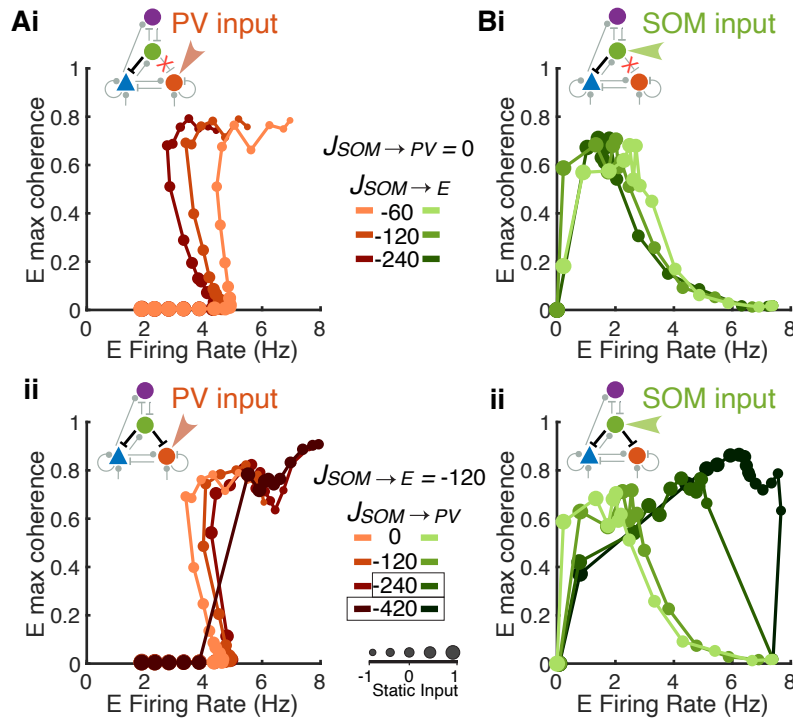


Figure 5: Relative strengths of SOM→E and SOM→PV connections shape modulation patterns of network state. Static input is applied to either PV neurons (Ai, ii) or SOM neurons (Bi, ii). Network state at each input level is represented by E firing rate and E maximum coherence (with the same convention as in Figure 4A). Increasing marker sizes correspond to increasing static input to the target population. (i) SOM inhibition to PV is removed ( $J_{SOM \rightarrow PV} = 0$ ) and increases of  $J_{SOM \rightarrow E}$  correspond to darker curves. (ii) SOM inhibition to E is fixed ( $J_{SOM \rightarrow E} = -120$ ) and increases of  $J_{SOM \rightarrow PV}$  correspond to darker curves. Default values of connections strengths are  $J_{SOM \rightarrow E} = -120$  and  $J_{SOM \rightarrow PV} = -60$ .

218 Bi, and 4A). When we eliminate SOM→PV connections (i.e.,  $J_{SOM→PV} = 0$ ), modulation patterns in  
219 network activity states (Figures 5Ai, Bi) remain qualitatively the same as in the network's default setting  
220 (Figures 3Bi, and 4Ai, Bi; more combinations of SOM→E and SOM→PV connection strengths are in  
221 Supplementary figure S6). Stronger SOM→E connections lead to a larger range of firing rates over the  
222 transition from the SA to the SS state through the middle branch, corresponding to the WS state, where  
223 rate and coherence vary in opposite directions (Figure 5Ai). The changes in state in response to external  
224 input variations are gradual in networks with different SOM→E connection strengths (Figure 5Ai, Bi).  
225 This points to a degree of resilience in the network's responsiveness to external input in the absence of  
226 SOM inhibition to PV.

227 This resilience is disrupted when the synaptic strength of SOM→PV dominates the synaptic strength of  
228 SOM→E, which results in an increased sensitivity of the network to changes in external input. Specifically,  
229 we consistently observe that when SOM inhibition to PV is sufficiently large compared to SOM inhibition  
230 to E, more pronounced and abrupt transitions from the SA to the SS state occur (e.g., the cases in Figure  
231 5Aii with  $J_{SOM→PV} = -420$  and in Figure 5Bii with  $J_{SOM→PV} = -240, -420$ ; Supplemental Figure S6).  
232 That is, dominance of SOM→PV inhibition over SOM→E inhibition increases network sensitivity to input  
233 and reduces or eliminates the range of input levels that result in the transitional activity dynamics, the WS  
234 state. Indeed, in the transition through the WS state, as SOM firing intensifies (Figure 3i), the inhibition  
235 from SOM to E and PV neurons will tend to reduce their firing rates. Yet, the drop in PV firing can  
236 disinhibit E. If this disinhibitory effect is dominant due to sufficiently strong  $J_{SOM→PV}$ , then E firing can  
237 increase rather than decreasing, resulting in a rapid transition through or elimination of the WS state. In  
238 this case, the firing rate and maximum coherence of E neurons tend to vary in the same direction (Figure  
239 5ii, Supplemental Figure S6). Comparing results from increasing and decreasing incremental changes in  
240 input levels, we observe that the abrupt transition between SA and SS states happens at different input  
241 values depending on the direction of change (Supplemental Figure S7). This hysteresis effect suggests that  
242 stronger SOM inhibition to PV neurons changes the criticality of the Hopf bifurcation at which SA stability  
243 is lost, from supercritical to subcritical.

244 These results imply that stronger inhibition from SOM→E neurons than that from SOM→PV neurons  
245 is necessary to observe activity consistent with the WS state and underscores the pivotal influence of SOM  
246 inhibition on the network's dynamical transitions.

## 247 **Dynamic interactions between E and SOM neurons are necessary for SOM-induced** 248 **network synchrony**

249 In this section, we investigate the impacts of E→SOM connections on SOM-induced network synchrony.  
250 What might drive the high coherence among SOM neurons and the rest of the network? Since SOM  
251 neurons do not connect to other SOM neurons and do not receive feedforward input, the high correlation  
252 among SOM neurons is driven by the recurrent input they receive from within the network. There are only  
253 two sources of recurrent inputs to SOM neurons, the excitation from E neurons and the inhibition from VIP  
254 neurons. To investigate the importance of E→SOM connections, we removed the E→SOM connections,  
255 and replaced this recurrent excitation with an external input that mimicked the statistics of the recurrent  
256 excitation.

257 First, we replaced recurrent excitation with colored noise that was independent for each SOM neuron.  
258 The colored noise was constructed as an Ornstein–Uhlenbeck (OU-) process that had equal mean and  
259 variance to the excitatory currents SOM neurons received on average in an intact default network with  
260 no static input (referred to as *baseline*; mean = 0.65 and variance = 0.12). In this decoupled network,

261 the coherence of the network remains low and decreases as we apply static input to SOM neurons, in  
262 addition to the noisy input, to increase their firing rates (Figure 6A). This result is opposite to the large  
263 increase in coherence with SOM rate that we observed in the default network (Figure 4A,C). The firing  
264 rate of E neurons is also suppressed much more abruptly compared to that in the default network as  
265 we increase input to SOM neurons (Supplemental Figure S8 compared to Figure 3C). This suggests that  
266 without E→SOM connections, SOM activity tends to reduce network synchrony mainly by reducing the  
267 E firing rate. The inhibition from VIP alone is not able to correlate SOM neurons. Consistently, varying  
268 VIP→SOM connections has little effect on network coherence (Supplemental Figure S5).

269 Next, we consider the possibility that correlated excitatory inputs are able to synchronize SOM neu-  
270 rons, which in turn synchronize the network as a whole. Because each SOM neuron receives input from a  
271 large number of E neurons (~1200 connections), very weak correlation in E spike trains can result in large  
272 correlation in the pooled excitatory current, as has been demonstrated theoretically [47]. The correlated  
273 excitatory current to SOM neurons cannot be dynamically canceled by inhibition due to the lack of in-  
274 hibitory connections among SOM neurons, which is distinct from the E-PV subcircuit where a balance of  
275 excitation and inhibition can be dynamically achieved [48, 49]. Therefore, excitatory input alone is able to  
276 drive correlated activity in SOM neurons. To demonstrate this, we record SOM spike trains from networks  
277 where SOM neurons receive excitation from E neurons but do not provide feedback inhibition to E and  
278 PV (Figure 6Bi, right column). Static input is applied to SOM neurons to modulate their firing rate. We  
279 then replay the recorded SOM spikes in networks where we remove E→SOM connections but allow SOM  
280 neurons to impact the rest of the network (Figure 6Bi, left column). In this way, E and SOM neurons  
281 are dynamically uncoupled, but SOM neurons receive realistic correlated excitation instead of simplified  
282 independent noise as in Figure 6A. We find that as input to SOM neurons increases, firing rate of SOM  
283 rises rapidly and their coherence level reaches to about 0.3 (Figure 6Bi, right column). This is consistent  
284 with the previous theoretical result that correlation between uncoupled neurons increases with firing rates  
285 [50]. The increased coherence in SOM spiking activity in turn induces synchrony among E neurons until  
286 E neurons are fully suppressed by the inhibition from SOM (Figure 6Bi, left column). Therefore, the  
287 correlated excitatory current to SOM neurons is able to drive the network into a weak synchrony regime  
288 (coherence around 0.15), but the peak coherence is much lower than that in the default network with  
289 E→SOM connections (Figure 6Bii).

290 Lastly, as we gradually restore E→SOM connections ( $J_{E \rightarrow SOM} > 0$ ) to allow for dynamic interaction  
291 between E and SOM neurons, we observe a positive relationship between the increases in coherence and  
292 increases in connection strength (Figure 6C). These results demonstrate that mimicking E→SOM input,  
293 using either colored noise with matched mean and variance (Figure 6A) or recorded SOM spikes from  
294 a decoupled network (Figure 6Bi-ii), is not sufficient to modulate activity through the three identified  
295 network states; rather, it is the dynamic interaction between E and SOM neurons that amplifies the weak  
296 correlation in the E-PV subcircuit and drives the network to strong synchrony.

## 297 **Heterogeneous external inputs reduce SOM-induced network synchrony**

298 In our previous set of results, adding noise to SOM neurons only slightly reduced the coherence of the  
299 network when E firing rate is small (Figure 6C, compare dark green with grey curves). This observation  
300 suggests that the network can still readily transition into a highly coherent regime even in the presence  
301 of noisy inputs that vary in time. To investigate the impact of noise in the external input, we applied  
302 independent OU input, with equal mean and variance, to each neuron in the SOM population (see Meth-  
303 ods). Increasing the variance of the OU input only weakly impacted the coherence of the network (Figure

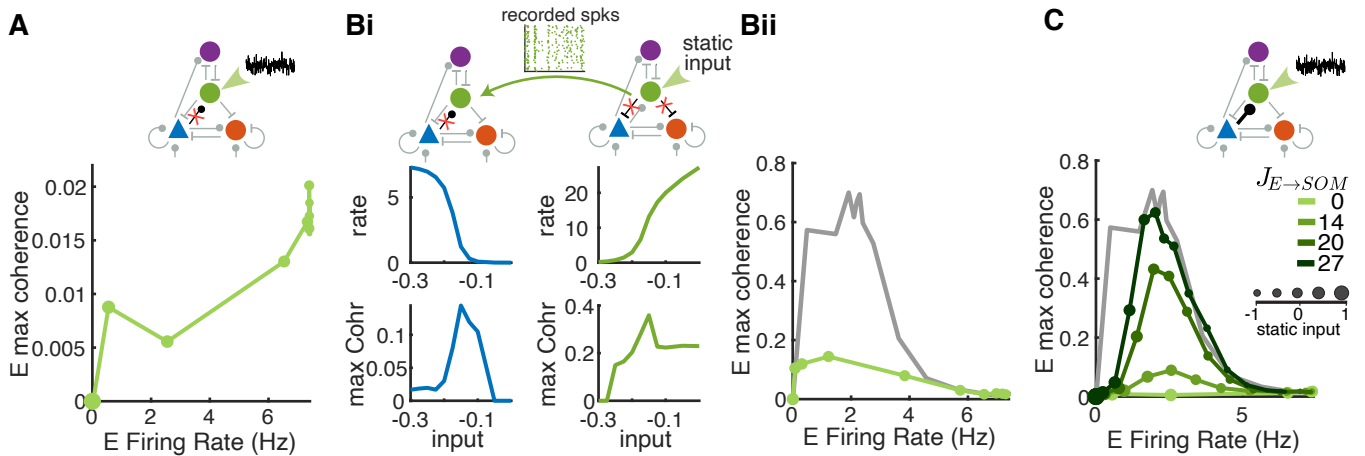


Figure 6: E→SOM connections are critical for SOM-induced network synchrony. (A) Removal of E→SOM connections eliminates coherence, despite the presence of stochastic input (OU process, see Methods) with mean and variance matched those of the excitatory currents to SOM neurons in the intact default network with no static input (mean = 0.65 and variance = 0.12). Larger dots correspond to stronger static inputs to SOM. (Bi-ii) E and SOM firing properties in networks where they are dynamically uncoupled, but SOM neurons receive and provide realistically correlated inputs and outputs, respectively. (Bi) Left column: Firing rate (top) and maximum coherence (bottom) of E neurons in networks with no E→SOM connection and where SOM spikes were replaced with those recorded from the network on the right. Right column: Firing rate (top) and maximum coherence (bottom) of SOM neurons in networks with intact E→SOM but no SOM→E and SOM→PV connections. Static input was applied to SOM neurons in the network on the right. (Bii) Modulation pattern of the firing rate and maximum coherence of E neurons from the network in Bi left (green) and from the default network (grey) with changes in static input to SOM neurons. (C) Increasing E→SOM synaptic strength increases the maximum coherence that can be achieved by varying static input to SOM neurons. SOM neurons receive the same OU noise as in panel A as  $J_{E \rightarrow SOM}$  values are varied. Hence the lightest green curve ( $J_{E \rightarrow SOM}=0$ ) is the same as that in panel A. As in panel Bii, the grey curve shows coherence for the default network with static input for comparison (same data as in Figure 4A green curve).

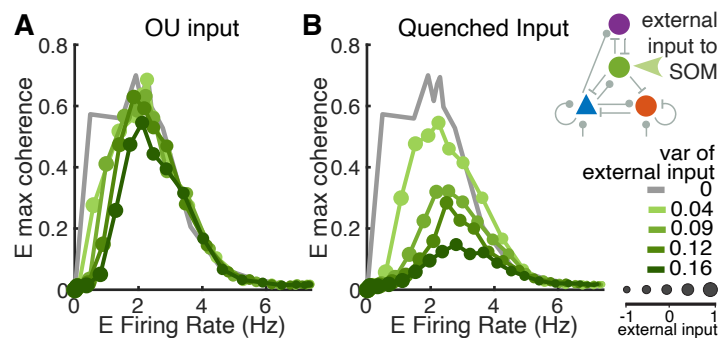


Figure 7: Comparison of two types of external input to SOM neurons. (A) Input to each neuron is modeled as an independent Ornstein–Uhlenbeck (OU-) process with the same mean and variance. (B) Input to each neuron is static in time but the strength is sampled independently from a normal distribution with a specified variance. The case of static input (variance equal to 0) is plotted in grey for comparison in each case (same as the grey curves in Figure 6).

304 7A). We next compared this outcome with the results of applying a different type of applied noise, het-  
 305 erogeneous or quenched noise, which is constant in time but has heterogeneous strengths, sampled from a  
 306 normal distribution of a given variance, across target neurons (see Methods). Implementing the OU input  
 307 and the quenched input allows us to compare the role of time-varying noise versus spatially-varying noise  
 308 in terms of influence on coherence.

309 Interestingly, we find that quenched input to SOM neurons has a much larger impact on network  
 310 synchrony than OU input with the same variance (Figure 7). With quenched input, we observe a substantial  
 311 decrease in E coherence across most firing rates (Figure 7B). Across all cases of external quenched or OU  
 312 inputs, the average population rates evolve similarly to the default network with homogeneous static input  
 313 as input strength is varied (Supplemental Figures S9, S10). Firing rates change more gradually with  
 314 increasing variance in the input, especially in the case of quenched input (Supplemental Figure S10). One  
 315 difference across input types is that SOM is able to suppress E activity at lower values of quenched input  
 316 than it can for static input. Overall, these results show that the transition to a synchronized network state  
 317 resulting from enhancing the activation of SOM neurons is robust against time-varying noisy input that is  
 318 of similar mean strength across the network, whereas a noise signal that has a spatially-varying strength  
 319 is more effective at reducing the network synchrony level.

## 320 Discussion

321 In this study, using a spatially-structured spiking model of a canonical neural circuit comprising E, SOM,  
 322 PV and VIP neurons, we demonstrate that SOM neurons are critical for synchronizing neural population  
 323 activity. As external drive is varied to any target population, the firing rate of SOM neurons is highly  
 324 predictive of the coherence level that emerges in the E population (Figure 4C). Without SOM neurons,  
 325 network synchrony varies much more gradually with the level of input applied to the E-PV subcircuit  
 326 (Figure S3). The spatial structure of the network is necessary for the gradual transition from asynchrony  
 327 to strong synchrony via a weak synchrony state, because it allows for the richer spatiotemporal dynamics  
 328 associated with this transitional regime, consistent with our past work ([38]; Figure S2). In addition, we  
 329 find that when SOM→PV inhibition is strong, the smooth transition through the weak synchrony state is  
 330 disrupted and the network becomes highly sensitive to input changes (Figure 5). We further show that the  
 331 dynamic interaction between E and SOM neurons is a necessary factor in the emergence of SOM-induced

332 network synchrony, as a network in which E and SOM neurons are dynamically uncoupled remains in the  
333 asynchronous state even when SOM firing rate is high (Figure 6).

334 Our model reproduces previous experimental findings where optogenetic inactivation of SOM neurons  
335 led to a reduction in the oscillatory power of the LFP around 30 Hz while inactivation of PV neurons  
336 did the opposite (Figure 3; [18, 24]). Consistent with these experiments [18, 24], SOM neurons contribute  
337 to oscillations of lower frequency ( $\sim 20$  Hz) and PV neurons contribute to oscillations of higher frequency  
338 ( $\sim 35$  Hz) in our model, partly due the differences in their synaptic decay time constants (8 ms for PV  
339 and 20 ms for SOM; [44]). We identified that the firing rate of SOM neurons is tightly correlated with  
340 the overall network synchrony level (Figure 4C), which is also consistent with the previous experimental  
341 finding that the average activity of SOM neurons co-varies linearly with the gamma power of LFP (20-40  
342 Hz) across multiple visual stimulus conditions (Figure S2 in [17]).

343 We find that the firing rate and coherence of E neurons can vary in opposite directions through the  
344 weak synchrony regime in networks with three interneuron subtypes (Figure 4A). In contrast, rates and  
345 coherence are tethered to vary in the same direction in the E-PV subcircuit (Figure S3). SOM neurons  
346 are responsible for the opposite relationship between rate and coherence of E neurons; when SOM neurons  
347 are more active, they suppress E neurons and increase network synchrony, and when SOM neurons are  
348 suppressed, E neurons firing rate increases and network synchrony is reduced. The opposite directionality  
349 of changes in E firing rates versus network synchrony has been observed with changes in spatial attention  
350 [51] and arousal state [15, 52]. The simultaneous increase in firing rate and decrease in synchrony can  
351 presumably enhance the signal-to-noise ratio of neural representations of stimuli [53]. Our results suggest  
352 that incorporating multiple interneuron subtypes supports the robust emergence of this enhanced coding  
353 state.

354 Our model predicts that a stronger or comparable magnitude of inhibition from SOM to E neurons  
355 compared to that from SOM to PV neurons is important for maintaining the weak synchrony regime  
356 (Figure 5, S6). When SOM to PV inhibition is much larger, the network shows abrupt transitions from the  
357 asynchronous to the strongly synchronous regime. This sensitivity arises because the positive feedback in  
358 the SOM $\rightarrow$ PV $\rightarrow$ E $\rightarrow$ SOM disinhibitory loop can lead to instability. Our result is consistent with a previous  
359 model which suggests that SOM inhibition to PV neurons can result in a loss of stability [54]. The presence  
360 of stronger SOM inhibition onto E compared to PV neurons is in agreement with anatomical findings in  
361 cortex [40, 41, 55]. On the other hand, recent experimental work suggests that activating SOM neurons  
362 enhances the reliability of E neuron responses to natural movie stimuli by suppressing PV neurons [29].  
363 The discrepancy between our model and this work could be due to the different temporal patterns of  
364 stimulation across the two. In our model, we only consider sustained application of external input, to  
365 model slow processes like the variation of brain state, while in these experiments [29], pulse stimulation  
366 was used. Further analysis is needed to investigate the dynamic responses of our model to brief, cell-type  
367 specific stimulation.

368 Our results also reveal an advantage of targeting VIP neurons to modulate a network's dynamical  
369 state. That is, targeting VIP neurons flexibly transitions the network between asynchronous and weakly  
370 synchronous regimes without pushing the network to pathologically strong oscillations (Figure 3). Anatom-  
371 ically, VIP neurons reside mostly in superficial layers in cortex and receive mostly long-range projections  
372 from other brain regions [9]. Therefore, they are hypothesized to be the main locus of feedback connec-  
373 tions and neuromodulator release. VIP neurons also have been shown to respond strongly to locomotion  
374 signals [12], novel stimuli and unexpected events [56, 57]. Nevertheless, VIP neurons mainly act through  
375 SOM neurons to regulate the E-PV subcircuit. Therefore, it is the activity of SOM neurons that is mostly

376 reflective of network state in our model.

377 Our model mainly generates fluctuations with spectral power concentrated around 15-40 Hz. However,  
378 past work has shown that activation of SOM neurons reduces low-frequency power (<10 Hz) of LFP in  
379 addition to increasing high-frequency power [10]. Arousal state also tends to have opposing impacts on the  
380 low- versus high-frequency oscillatory power of LFP; high arousal state is associated with reduced power  
381 in the low-frequency band and increased power in the high-frequency band [15, 58, 52]. The lack of slow  
382 time-scale fluctuations in our model means that the model cannot fully account for the impacts of brain  
383 state on population activity. Future work is needed to extend the current model to consider various slow  
384 time variables, such as spike frequency adaption and slow synaptic receptors, that are omitted from the  
385 present work.

386 The brain features a vast diversity of neuronal types, each of which has unique connectivity patterns,  
387 expression of neuromodulator receptors and electrophysiological properties. Different cell types coordinate  
388 their activity to regulate neural population dynamics for flexible computations. Our model provides new  
389 insights and predictions about the different functions that each primary interneuron subtype may serve  
390 in modulating the dynamical state of cortex, highlighting the importance of E-SOM interactions and the  
391 relative strengths of SOM inputs to E versus PV neurons. Altogether, our results emphasize a unique role  
392 of SOM neurons in controlling network synchrony.

## 393 Methods

### 394 Spiking neuron network model

395 The model network consists of a single recurrent layer and a feedforward input layer (Fig. 1B). The  
 396 feedforward layer (population X) is composed of 2,500 ( $N_X$ ) excitatory neurons modeled as independent  
 397 Poisson processes with a uniform rate of 10 Hz. The recurrent layer contains 50,000 neurons ( $N$ ) divided  
 398 into four cell population types,  $N_e = 40,000$  E,  $N_p = 4,000$  PV,  $N_s = 4,000$  SOM, and  $N_v = 2,000$   
 399 VIP neurons. The population size ratios follow anatomical data from mouse cortex [45]. The synaptic  
 400 connection patterns among the four neuron populations are constrained by anatomical and physiological  
 401 data from mouse visual cortex (Figure 1A; [40, 41, 25]). In particular, we assume there are no reciprocal  
 402 connections among SOM neurons or among VIP neurons; VIP neurons only inhibit SOM neurons; and  
 403 only E and PV neurons receive input from the feedforward layer (see Model parameters). Most of model  
 404 parameters are similar to those in our previous work [38] except for some changes to incorporate different  
 405 interneuron subtypes.

406 Each neuron in the recurrent layer is modeled as an exponential integrate-and-fire (EIF) neuron with  
 407 membrane potential defined as:

$$C_m \frac{dV_j^\alpha}{dt} = -g_L(V_j^\alpha - E_L) + g_L \Delta_T e^{(V_j^\alpha - V_T)/\Delta_T} + I_j^\alpha(t), \quad (1)$$

408 where neuron  $j$  is a member of the  $\alpha$  population,  $\alpha \in \{e, p, s, v\}$ . When  $V_j^\alpha(t)$  exceeds a threshold  $V_{th}$ ,  
 409 the neuron spikes and the membrane potential is held at  $V_{th}$  for a refractory period  $\tau_{ref}$  and then reset  
 410 to a lower potential value,  $V_{re}$  (see Model Parameters). All membrane potentials are bounded below by  
 411  $V_{lb} = -100$  mV. The total current to neuron  $j$  in population  $\alpha$  is

$$\frac{I_j^\alpha(t)}{C_m} = \sum_{k=1}^{N_X} \frac{J_{jk}^{\alpha X}}{\sqrt{N}} \sum_n \eta_X(t - t_n^{X,k}) + \sum_{\beta=\{e,p,s,v\}} \sum_{k=1}^{N_\beta} \frac{J_{jk}^{\alpha\beta}}{\sqrt{N}} \sum_n \eta_\beta(t - t_n^{\beta k}) + \mu_\alpha + x_j(t), \quad (2)$$

412 where  $n$  indexes the spikes fired by the presynaptic neurons,  $J^{\alpha\beta}$  is the recurrent synaptic strength from  
 413 population  $\beta$  to population  $\alpha$  (which may be 0 in some cases),  $J^{\alpha X}$  is the synaptic strength from the  
 414 feedforward layer to population  $\alpha$  (see Model Parameters),  $\mu_\alpha$  is a constant external input current and  
 415  $x_j(t)$  is input noise (Eq. 6). Note that the strength of each synaptic connection is scaled by  $1/\sqrt{N}$ . In  
 416 equation (2), the postsynaptic current terms are defined as:

$$\eta_\beta(t) = \frac{1}{\tau_{\beta_d} - \tau_{\beta_r}} \begin{cases} e^{-t/\tau_{\beta_d}} - e^{-t/\tau_{\beta_r}}, & t \geq 0 \\ 0, & t < 0 \end{cases}, \quad (3)$$

417 where  $\tau_{\beta_d}$  and  $\tau_{\beta_r}$  (see Model Parameters) are the synaptic decay and rise time constants for population  
 418  $\beta$ . The synaptic timescales of inhibitory connections from SOM and VIP neurons are slower than that of  
 419 connections from PV neurons, which is in turn slower than that of excitatory connections, constrained by  
 420 physiological data from mouse visual cortex [44].

421 Neurons are uniformly distributed on a unit square,  $\Gamma = [0, 1] \times [0, 1]$ . The connection probability  
 422 between a pair of neurons with coordinates  $\mathbf{x} = (x_1, x_2)$  and  $\mathbf{y} = (y_1, y_2)$ , respectively, depends on the  
 423 populations to which the neurons belong and the distance between the two neurons as

$$p_{\alpha\beta}(\mathbf{x}, \mathbf{y}) = \bar{p}_{\alpha\beta} g(x_1 - y_1; \alpha_\beta) g(x_2 - y_2; \alpha_\beta), \quad (4)$$



424 where  $\bar{p}_{\alpha\beta}$  is the mean probability of connections from population  $\beta$  to population  $\alpha$  and  $g(x; \sigma)$  is a  
 425 wrapped Gaussian distribution:

$$g(x; \sigma) = \frac{1}{\sqrt{2\pi\sigma}} \sum_{k=-\infty}^{\infty} e^{-(x+k)^2/(2\sigma^2)} \quad (5)$$

426 with projection width  $\sigma$  (see Model Parameters). Connections to and from the SOM cells have a larger  
 427 spatial footprint compared to other connections, based on findings from mouse visual and auditory cortex  
 428 [25, 43]. A presynaptic neuron is allowed to make more than one synaptic connection to a single postsynap-  
 429 tic neuron. The number of synaptic projections, or out-degree,  $K_{\alpha\beta}$ , from population  $\alpha$  to population  $\beta$  is  
 430 fixed for all neurons in population  $\alpha$ , and indices of postsynaptic neurons are selected randomly according  
 431 to the connection probability in Eq. 4.

For many of our simulations, the external input,  $\mu_\alpha$ , was varied between -1.0 and 1.0 with step size 0.1. Input noise,  $x_j(t)$ , was modeled as an independent Ornstein-Uhlenbeck (OU) process (Figures 6, 7):

$$\tau_{E_d} dx_j = (\mu_n - x_j) dt + \sigma_n dW, \quad (6)$$

432 where  $W$  is a Wiener process, and the time constant of the OU process was chosen to be the same as the  
 433 decay time constant of the excitatory synaptic current,  $\tau_{E_d}$ . The mean of  $x_j(t)$  is  $\mu_n$  and the variance is  
 434  $\sigma_n^2/(2\tau_{E_d})$ . In simulations where we replaced E→SOM connections with an OU process (Figure 6), we set  
 435  $\mu_n = 0.65$  and  $\sigma_n = 1.1$  to match the mean (0.65) and variance (0.12) of the excitatory current from E  
 436 to SOM neurons in default networks without external input. In simulations with quenched input (Figure  
 437 7B), the constant external input,  $\mu_j^\alpha$ , to neuron  $j$  from population  $\alpha$  is sampled from a normal distribution  
 438 with mean  $\mu_\alpha$  and standard deviation  $\Delta_\mu$ .

439 The cellular parameters of the EIF model for each cell type and all network parameters are summarized  
 440 in the Model Parameters section. The differential equations (1) and (2) were solved using a forward Euler  
 441 method with a timestep of 0.05 ms. All simulations were performed on the CNBC Cluster at the Carnegie  
 442 Mellon University. All simulations were written in a combination of C and MATLAB R2021b (9.11),  
 443 MathWorks.

## 444 Model Parameters

445 The following tables specify the parameter values used in our simulations. As above, the symbol X denotes  
 446 the feedforward connections.

### Synaptic time constants

	E	PV	SOM	VIP	X
$\tau_d$ (ms)	5	8	20	40	5
$\tau_r$ (ms)	1	1	1	1	1

### Synaptic connection strengths, $J_{\alpha\beta}$

		from ( $\beta$ )				
		E	PV	SOM	VIP	X
to ( $\alpha$ )	E	30	-90	-120	0	120
	PV	40	-150	-60	0	250
	SOM	27	0	0	-10	0
	VIP	72	0	-10	0	0

### Mean synaptic connection probability, $\bar{p}_{\alpha\beta}$

		from ( $\beta$ )				
		E	PV	SOM	VIP	X
to ( $\alpha$ )	E	0.01	0.04	0.03	0	0.1
	PV	0.03	0.04	0.03	0	0.05
	SOM	0.03	0	0	0.1	0
	VIP	0.01	0	0.1	0	0

### Number of postsynaptic connections, $K_{\alpha\beta}$

		from ( $\beta$ )				
		E	PV	SOM	VIP	X
to ( $\alpha$ )	E	400	1600	1200	0	4000
	PV	120	160	120	0	200
	SOM	120	0	0	400	0
	VIP	20	0	200	0	0

### Connection widths, $\sigma_{\alpha\beta}$

		from ( $\beta$ )				
		E	PV	SOM	VIP	X
to ( $\alpha$ )	E	0.1	0.1	0.2	0	0.1
	PV	0.1	0.1	0.2	0	0.1
	SOM	0.2	0	0	0.2	0
	VIP	0.1	0	0.2	0	0

### EIF Parameters

		E	PV	SOM	VIP
$\tau_m = \frac{C_m}{g_L}$	(ms)	15	10	10	10
$\tau_{ref}$	(ms)	1.5	0.5	1.5	1.5
$V_{lb}$	(mV)	-100	-100	-100	-100
$V_{th}$	(mV)	-10	-10	-10	-10
$\Delta_T$	(mV)	2	0.5	2	2
$V_T$	(mV)	-50	-50	-50	-50
$V_{re}$	(mV)	-65	-65	-65	-65
$E_L$	(mV)	-60	-60	-60	-60

## 447 Quantification and Statistical Analysis

448 **Spike Count Correlations** Spike counts were computed using a sliding window of 100 ms with a step  
449 size of 1 ms. Pearson correlation coefficients were computed for all neuron pairs as a function of distance  
450 (Figure 2iv), except that neurons with rates less than 1 Hz were excluded from correlation calculations.  
451 The membrane potential of each neuron was randomly initialized for each simulation, and connectivity  
452 matrices were regenerated for each input condition. A total of 5 simulations of 15 seconds each were  
453 performed for each input condition. The first 500 ms of each simulation was excluded from the analysis.

454 **Coherence** We measured the average pairwise coherence within each cell type population as an indication  
455 of network synchrony across simulation conditions. Spike trains were first partitioned into 1 ms time bins

456 and these were collected into 1 second time windows with 0.5 second overlap. Mean firing rate of each  
457 sampled neuron was subtracted. Power spectral density,  $S_i$ , of neuron  $i$ , and cross spectral density,  $S_{ij}$ ,  
458 between neuron  $i$  and neuron  $j$ , were calculated using the fast Fourier transform and averaged over time  
459 windows. The coherence between neuron  $i$  and neuron  $j$  at frequency  $f$  was calculated as

$$C_{ij}(f) = \frac{S_{ij}(f)}{\sqrt{S_i(f)S_j(f)}}. \quad (7)$$

460 Pairwise coherence was averaged across all sampled neuron pairs within a population. Note that the  
461 coherence definition used here is not magnitude-squared, because the magnitude-squared coherence is  
462 always positive even when the network is asynchronous. We excluded neurons with rates less than 1 Hz  
463 and ensured that 500 neurons were sampled from each population. The first second of each simulation was  
464 removed.

465 **Activity State Definitions** We identified three network states that were observed for the range of  
466 input levels considered, based on mean firing rates and maximum coherence. Specifically, the subcircuit  
467 asynchronous (SA) state occurs when the average firing rate of SOM neurons is less than 1 Hz and the  
468 maximum coherence of E neurons is less than 0.1. The weakly synchronous (WS) state arises when the  
469 maximum coherence of E neurons is between 0.1 and 0.5 and the average firing rate of SOM neurons is  
470 larger than 1 Hz. The strongly synchronous (SS) state is when the maximum coherence of E neuron is  
471 larger than 0.5 and the average firing rate of SOM neurons is larger than 1 Hz.

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## 476 Author Contributions

477 M.E., J.R. and C.H. conceived the project; M.E. performed the simulations and analysis, in consultation  
478 with J.R. and C.H.; C.H. supervised the project; all authors contributed to writing the manuscript.

## 479 Data and Software Availability

480 Computer code for all simulations and data analysis will be available online upon publication.

## 481 Declaration of Interests

482 The authors declare no competing interests.

## 483 References

- 484 [1] Albright TD, Stoner GR. Contextual influences on visual processing. *Annual review of neuroscience*.  
485 2002;25(1):339–379.
- 486 [2] Harris KD, Thiele A. Cortical state and attention. *Nature reviews neuroscience*. 2011;12(9):509–523.
- 487 [3] McCormick DA, Nestvogel DB, He BJ. Neuromodulation of Brain State and Behavior. *Annual Review*  
488 *of Neuroscience*. 2020;43(1):391–415.
- 489 [4] Flavell SW, Gogolla N, Lovett-Barron M, Zelikowsky M. The emergence and influence of internal  
490 states. *Neuron*. 2022;110(16):2545–2570.
- 491 [5] Isaacson JS, Scanziani M. How Inhibition Shapes Cortical Activity. *Neuron*. 2011;72(2):231–243.
- 492 [6] Cardin JA. Inhibitory Interneurons Regulate Temporal Precision and Correlations in Cortical Circuits;  
493 2018.
- 494 [7] Ferguson KA, Cardin JA. Mechanisms underlying gain modulation in the cortex. *Nature Reviews*  
495 *Neuroscience*. 2020;21(2):80–92.
- 496 [8] Ascoli GA, Alonso-Nanclares L, Anderson SA, Barrionuevo G, Benavides-Piccione R, Burkhalter A,  
497 et al. Petilla terminology: nomenclature of features of GABAergic interneurons of the cerebral cortex.  
498 *Nature Reviews Neuroscience*. 2008;9(7):557–568.
- 499 [9] Tremblay R, Lee S, Rudy B. GABAergic interneurons in the neocortex: from cellular properties to  
500 circuits. *Neuron*. 2016;91(2):260–292.
- 501 [10] Chen G, Rasch MJ, Wang R, Zhang Xh. Experience-dependent emergence of beta and gamma band  
502 oscillations in the primary visual cortex during the critical period. *Scientific Reports*. 2015;5(1):17847.
- 503 [11] Zhang S, Xu M, Kamigaki T, Hoang Do JP, Chang WC, Jenvay S, et al. Long-range and local circuits  
504 for top-down modulation of visual cortex processing. *science*. 2014;345(6197):660–665.
- 505 [12] Fu Y, Tucciarone JM, Espinosa JS, Sheng N, Darcy DP, Nicoll RA, et al. A cortical circuit for gain  
506 control by behavioral state. *Cell*. 2014;156(6):1139–1152.
- 507 [13] Pakan JM, Lowe SC, Dylida E, Keemink SW, Currie SP, Coutts CA, et al. Behavioral-state modulation  
508 of inhibition is context-dependent and cell type specific in mouse visual cortex. *Elife*. 2016;5:e14985.
- 509 [14] Dipoppa M, Ranson A, Krumin M, Pachitariu M, Carandini M, Harris KD. Vision and locomotion  
510 shape the interactions between neuron types in mouse visual cortex. *Neuron*. 2018;98(3):602–615.
- 511 [15] Vinck M, Batista-Brito R, Knoblich U, Cardin JA. Arousal and Locomotion Make Distinct Contri-  
512 butions to Cortical Activity Patterns and Visual Encoding. *Neuron*. 2015;86(3):740–754.
- 513 [16] McGinley MJ, David SV, McCormick DA. Cortical Membrane Potential Signature of Optimal States  
514 for Sensory Signal Detection. *Neuron*. 2015;87(1):179–192.
- 515 [17] Veit J, Handy G, Mossing DP, Doiron B, Adesnik H. Cortical VIP neurons locally control the  
516 gain but globally control the coherence of gamma band rhythms. *Neuron*. 2023;111(3):405–417.e5.  
517 doi:10.1016/j.neuron.2022.10.036.

- 518 [18] Chen G, Zhang Y, Li X, Zhao X, Ye Q, Lin Y, et al. Distinct inhibitory circuits orchestrate cortical  
519 beta and gamma band oscillations. *Neuron*. 2017;96(6):1403–1418.
- 520 [19] Jang HJ, Chung H, Rowland JM, Richards BA, Kohl MM, Kwag J. Distinct roles of parvalbumin  
521 and somatostatin interneurons in gating the synchronization of spike times in the neocortex. *Sci Adv*.  
522 2020;6(17).
- 523 [20] Wood KC, Blackwell JM, Geffen MN. Cortical inhibitory interneurons control sensory processing.  
524 *Curr Opin Neurobiol*. 2017;46:200–207.
- 525 [21] Cardin JA, Carlén M, Meletis K, Knoblich U, Zhang F, Deisseroth K, et al. Driving fast-spiking cells  
526 induces gamma rhythm and controls sensory responses. *Nature*. 2009;459(7247):663–667.
- 527 [22] Sohal VS, Zhang F, Yizhar O, Deisseroth K. Parvalbumin neurons and gamma rhythms enhance  
528 cortical circuit performance. *Nature*. 2009;459(7247):698–702.
- 529 [23] Buzsáki G, Wang XJ. Mechanisms of gamma oscillations. *Annual review of neuroscience*.  
530 2012;35(1):203–225.
- 531 [24] Veit J, Hakim R, Jadi MP, Sejnowski TJ, Adesnik H. Cortical gamma band synchronization through  
532 somatostatin interneurons. *Nature Neuroscience*. 2017;20(7):951–959.
- 533 [25] Adesnik H, Bruns W, Taniguchi H, Huang ZJ, Scanziani M. A neural circuit for spatial summation  
534 in visual cortex. *Nature*. 2012;490(7419):226.
- 535 [26] Pi HJ, Hangya B, Kvitsiani D, Sanders JI, Huang ZJ, Kepecs A. Cortical interneurons that specialize  
536 in disinhibitory control. *Nature*. 2013;503(7477):521–524.
- 537 [27] Keller AJ, Dipoppa M, Roth MM, Caudill MS, Ingrosso A, Miller KD, et al. A Disin-  
538 hibitory Circuit for Contextual Modulation in Primary Visual Cortex. *Neuron*. 2020;108(6).  
539 doi:<https://doi.org/10.1016/j.neuron.2020.11.013>.
- 540 [28] Ferguson KA, Salameh J, Alba C, Selwyn H, Barnes C, Lohani S, et al. VIP interneurons regulate  
541 cortical size tuning and visual perception. *Cell reports*. 2023;42(9).
- 542 [29] Rikhye RV, Yildirim M, Hu M, Breton-Provencher V, Sur M. Reliable sensory processing in mouse  
543 visual cortex through cooperative interactions between somatostatin and parvalbumin interneurons.  
544 *Journal of Neuroscience*. 2021;41(42):8761–8778.
- 545 [30] Potter C, Bassi C, Runyan CA. Simultaneous interneuron labeling reveals population-level interactions  
546 among parvalbumin, somatostatin, and pyramidal neurons in cortex. *bioRxiv*. 2024;.
- 547 [31] Litwin-Kumar A, Rosenbaum R, Doiron B. Inhibitory stabilization and visual coding in cortical  
548 circuits with multiple interneuron subtypes. *Journal of Neurophysiology*. 2016;115(3):1399–1409.
- 549 [32] Garcia del Molino LC, Yang GR, Mejias JF, Wang XJ. Paradoxical response reversal of top-down  
550 modulation in cortical circuits with three interneuron types. *eLife*. 2017;6:e29742.
- 551 [33] Palmigiano A, Fumarola F, Mossing DP, Kraynyukova N, Adesnik H, Miller KD. Common rules  
552 underlying optogenetic and behavioral modulation of responses in multi-cell-type V1 circuits. *bioRxiv*.  
553 2020; p. 2020–11.

- 554 [34] Waitzmann F, Wu YK, Gjorgjieva J. Top-down modulation in canonical cortical circuits with short-  
555 term plasticity. *Proceedings of the National Academy of Sciences*. 2024;121(16):e2311040121.
- 556 [35] Beerendonk L, Mejías JF, Nuiten SA, de Gee JW, Fahrenfort JJ, van Gaal S. A disinhibitory circuit  
557 mechanism explains a general principle of peak performance during mid-level arousal. *Proceedings of*  
558 *the National Academy of Sciences*. 2024;121(5):e2312898121.
- 559 [36] Vierling-Claassen D, Cardin JA, Moore CI, Jones SR. Computational modeling of distinct neocortical  
560 oscillations driven by cell-type selective optogenetic drive: separable resonant circuits controlled by  
561 low-threshold spiking and fast-spiking interneurons. *Frontiers in human neuroscience*. 2010;4:198.
- 562 [37] Hahn G, Kumar A, Schmidt H, Knösche TR, Deco G. Rate and oscillatory switching dynamics of a  
563 multilayer visual microcircuit model. *Elife*. 2022;11:e77594.
- 564 [38] Huang C, Ruff DA, Pyle R, Rosenbaum R, Cohen MR, Doiron B. Circuit Models  
565 of Low-Dimensional Shared Variability in Cortical Networks. *Neuron*. 2019;101:337–348.e4.  
566 doi:10.1016/j.neuron.2018.11.034.
- 567 [39] Fourcaud-Trocmé N, Hansel D, Van Vreeswijk C, Brunel N. How spike generation mechanisms deter-  
568 mine the neuronal response to fluctuating inputs. *Journal of neuroscience*. 2003;23(37):11628–11640.
- 569 [40] Campagnola L, Seeman SC, Chartrand T, Kim L, Hoggarth A, Gamlin C, et al. Local connectivity  
570 and synaptic dynamics in mouse and human neocortex. *Science*. 2022;375(6585).
- 571 [41] Pfeffer CK, Xue M, He M, Huang ZJ, Scanziani M. Inhibition of inhibition in visual cortex: the logic  
572 of connections between molecularly distinct interneurons. *Nature neuroscience*. 2013;16(8):1068.
- 573 [42] Mosheiff N, Ermentrout B, Huang C. Chaotic dynamics in spatially distributed neuronal networks  
574 generate population-wide shared variability. *PLOS Computational Biology*. 2023;19(1):e1010843.
- 575 [43] Kato HK, Asinof SK, Isaacson JS. Network-level control of frequency tuning in auditory cortex.  
576 *Neuron*. 2017;95(2):412–423.
- 577 [44] Karnani MM, Jackson J, Ayzenshtat I, Tucciarone J, Manoocheri K, Snider WG, et al. Cooperative  
578 subnetworks of molecularly similar interneurons in mouse neocortex. *Neuron*. 2016;90(1):86–100.
- 579 [45] Kim Y, Yang GR, Pradhan K, Venkataraju KU, Bota M, García del Molino LC, et al. Brain-wide  
580 Maps Reveal Stereotyped Cell-Type-Based Cortical Architecture and Subcortical Sexual Dimorphism.  
581 *Cell*. 2017;171(2):456–469.e22.
- 582 [46] Khoury CF, Fala NG, Runyan CA. The spatial scale of somatostatin subnetworks increases from  
583 sensory to association cortex. *Cell Rep*. 2022;40(10):111319.
- 584 [47] Rosenbaum R, Trousdale J, Josic K. Pooling and correlated neural activity. *Frontiers in computational*  
585 *neuroscience*. 2010;4:1209.
- 586 [48] Renart A, de la Rocha J, Bartho P, Hollender L, Parga N, Reyes A, et al. The Asynchronous State  
587 in Cortical Circuits. *Science*. 2010;327:587–590. doi:10.1126/science.1179850.
- 588 [49] Van Vreeswijk C, Sompolinsky H. Chaos in neuronal networks with balanced excitatory and inhibitory  
589 activity. *Science*. 1996;274(5293):1724–1726.

- 590 [50] De La Rocha J, Doiron B, Shea-Brown E, Josić K, Reyes A. Correlation between neural spike trains  
591 increases with firing rate. *Nature*. 2007;448(7155):802–806.
- 592 [51] Cohen MR, Maunsell JHR. Attention improves performance primarily by reducing interneuronal  
593 correlations. *Nature Neuroscience*. 2009;12(12):1594–1600.
- 594 [52] Reimer J, Froudarakis E, Cadwell CR, Yatsenko D, Denfield GH, Tolias AS. Pupil fluctuations track  
595 fast switching of cortical states during quiet wakefulness. *neuron*. 2014;84(2):355–362.
- 596 [53] Kohn A, Coen-Cagli R, Kanitscheider I, Pouget A. Correlations and Neuronal Population Information.  
597 *Annual Review of Neuroscience*. 2016;39(1):237–256.
- 598 [54] Bos H, Oswald AM, Doiron B. Untangling stability and gain modulation in cortical circuits with  
599 multiple interneuron classes. *bioRxiv*. 2008;doi:10.1101/2020.06.15.148114.
- 600 [55] Xu H, Jeong HY, Tremblay R, Rudy B. Neocortical somatostatin-expressing GABAergic interneurons  
601 disinhibit the thalamorecipient layer 4. *Neuron*. 2013;77(1):155–167.
- 602 [56] Garrett M, Manavi S, Roll K, Ollerenshaw DR, Groblewski PA, Ponvert ND, et al. Experience shapes  
603 activity dynamics and stimulus coding of VIP inhibitory cells. *elife*. 2020;9:e50340.
- 604 [57] Najafi F, Russo S, Lecoq J. Unexpected events modulate context signaling in VIP and excitatory cells  
605 of the visual cortex. *bioRxiv*. 2024; p. 2024–05.
- 606 [58] Akella S, Ledochowitsch P, Siegle JH, Belski H, Denman D, Buice MA, et al. Deciphering neuronal  
607 variability across states reveals dynamic sensory encoding. *bioRxiv*. 2024; p. 2024–04.
- 608 [59] Tsodyks MV, Skaggs WE, Sejnowski TJ, McNaughton BL. Paradoxical effects of external modulation  
609 of inhibitory interneurons. *Journal of neuroscience*. 1997;17(11):4382–4388.
- 610 [60] Ozeki H, Finn IM, Schaffer ES, Miller KD, Ferster D. Inhibitory stabilization of the cortical network  
611 underlies visual surround suppression. *Neuron*. 2009;62(4):578–592.
- 612 [61] Sanzeni A, Akitake B, Goldbach HC, Leedy CE, Brunel N, Histed MH. Inhibition stabilization is a  
613 widespread property of cortical networks. *Elife*. 2020;9:e54875.

614 **Supplemental information**

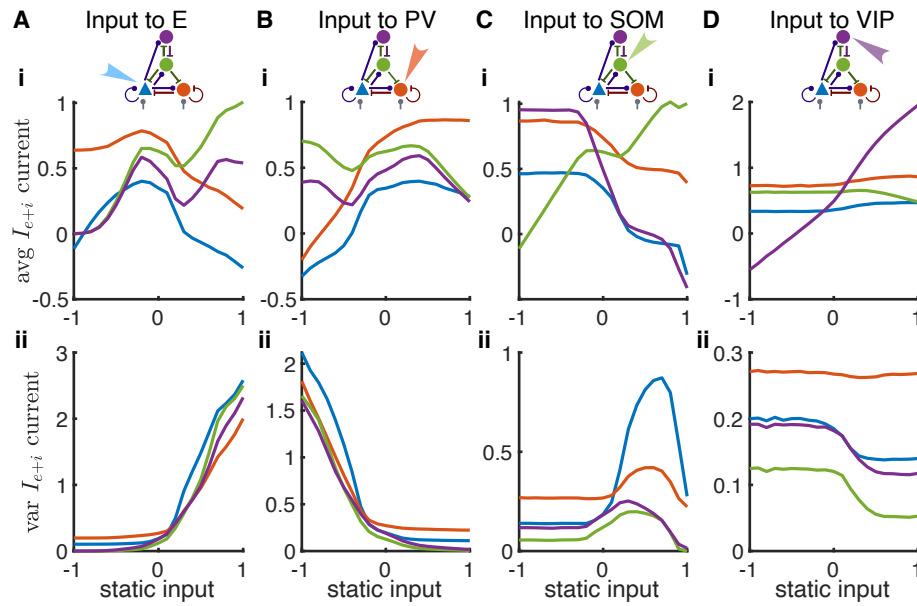


Figure S1: **Changes in synaptic currents as input is applied to each population. Related to Figure 3.** Static input is applied to the E (A), PV (B), SOM (C) or VIP (D) population. Row (i): Average total synaptic current to each population. Row (ii): Population-averaged variance of the total synaptic current to each population.

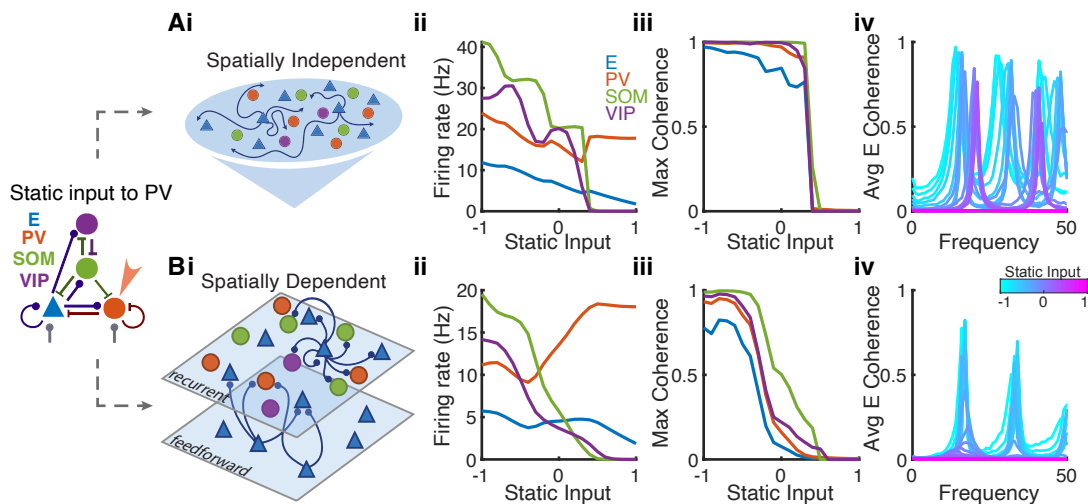


Figure S2: **Sharp transitions in networks with no spatial structure. Related to Figure 3.** Static input was applied to PV neurons. (Ai-iv) Firing rate and coherence in the four neuron populations in networks with no spatial structure, meaning that the connection probability between two neurons does not depend on distance. A sharp transition from the asynchronous to the strongly synchronous state occurs as input is increased. (Bi-iv) The same quantities for networks with spatial structure, as also shown in Figure 3B. The spatially dependent network exhibits a more gradual transition and the existence of weakly synchronous state over a range of input values. The parameters of the networks in A and B are the same except for the connection widths,  $\sigma_{\alpha\beta}$  (see Model Parameters in Methods).



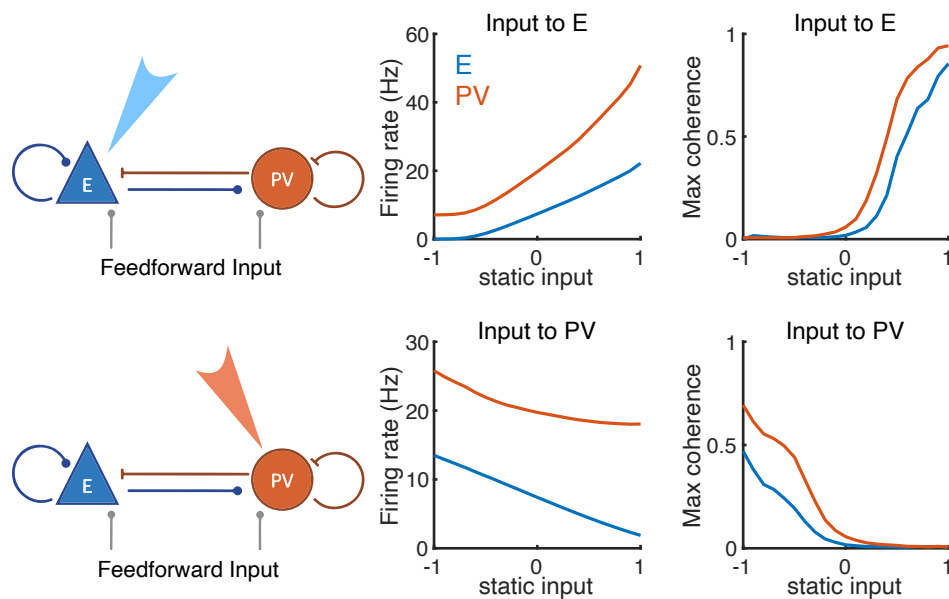


Figure S3: **Modulations of firing rates and maximum coherence in the E-PV subcircuit. Related to Figure 4.** Static external input was targeted to E (top row) or PV (bottom row) neurons. Firing rates and network synchrony are tethered to change in the same direction in the E-PV subcircuit. That is, stimulating E neurons increases the firing rates and coherence of both E and PV neurons, while stimulating PV neurons decreases firing rates and coherence in both populations. The paradoxical effect where stimulating PV leads to a reduction in PV firing rate suggests that the E-PV subcircuit is in the inhibition-stabilized regime [59, 60, 61]. Network parameters were the same as those Figures 1-4 except that we removed SOM and VIP populations.

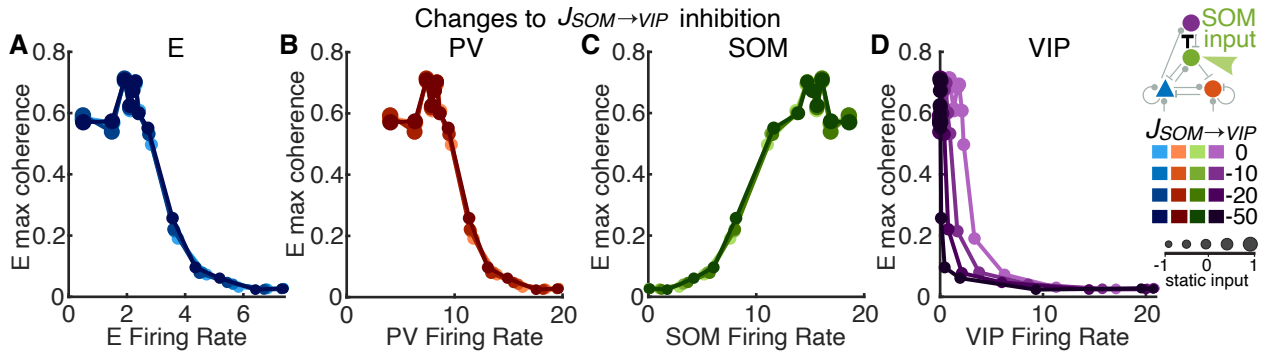


Figure S4: **SOM→VIP connection strength has little effect on modulation patterns. Related to Figure 5.** Static input was applied to SOM neurons. (A) E, (B) PV, (C) SOM, and (D) VIP population rates compared to E maximum coherence in networks with different SOM→VIP connection strengths,  $J_{SOM \rightarrow VIP}$ . Only the relation to VIP firing rate (panel D) is different in networks with different  $J_{SOM \rightarrow VIP}$ . In networks with larger  $J_{SOM \rightarrow VIP}$  inhibition, SOM is able to suppress VIP at a lower rate, resulting in the darkened curves shifting leftward (D). Therefore, altering the connection strength of  $J_{SOM \rightarrow VIP}$  exclusively affects the VIP population and does not influence how the rest of the network responds to external input. Note that  $J_{SOM \rightarrow VIP} = -10$  is the default circuit parameter used in the main text.

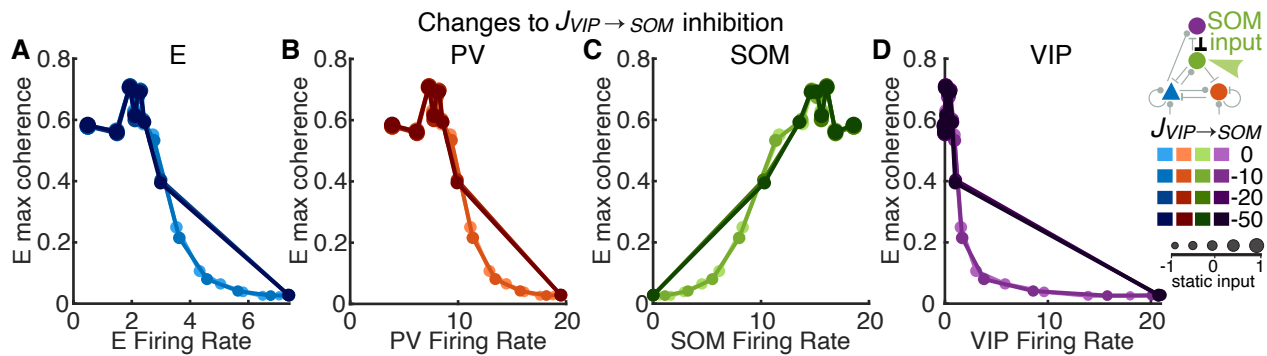
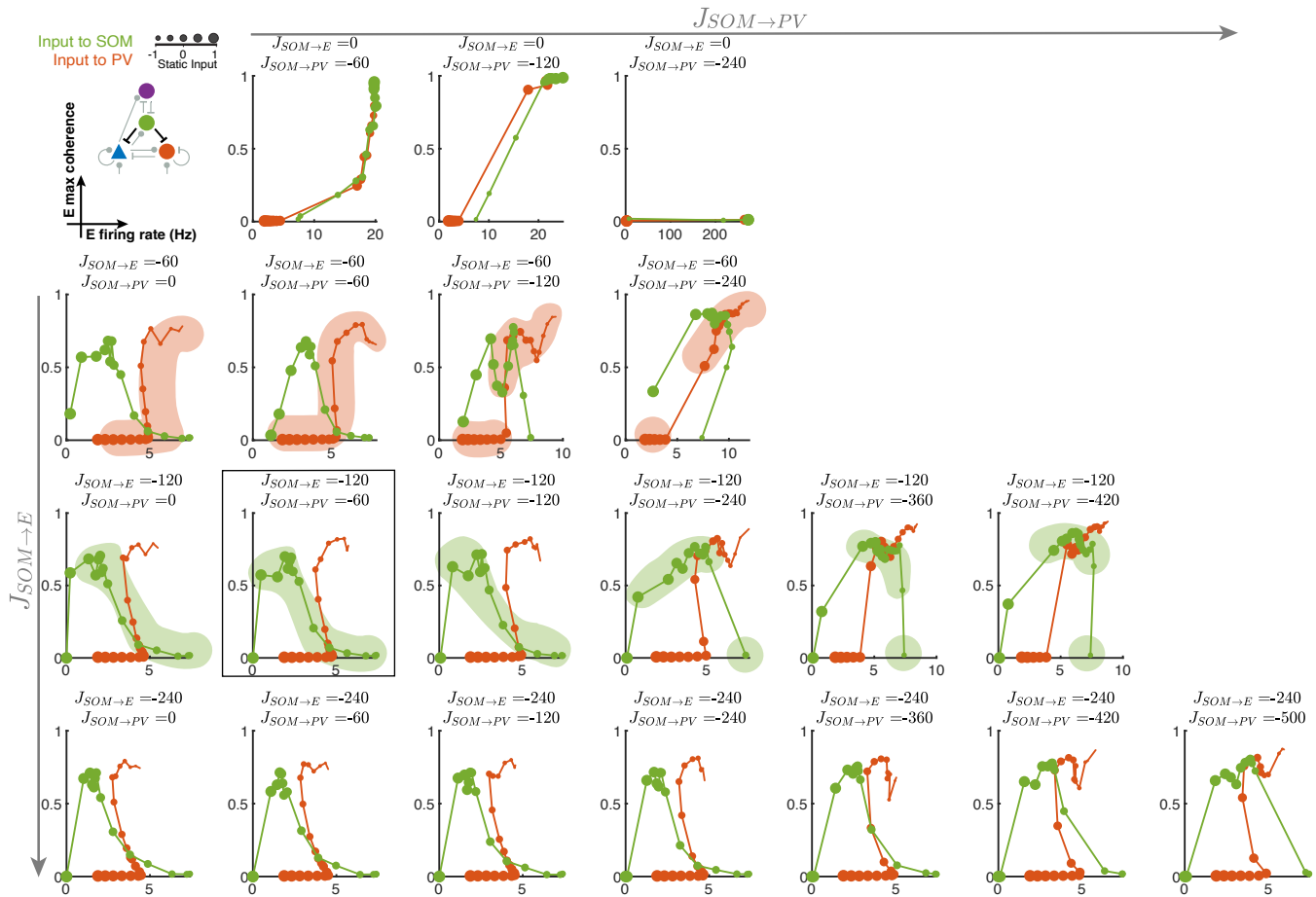
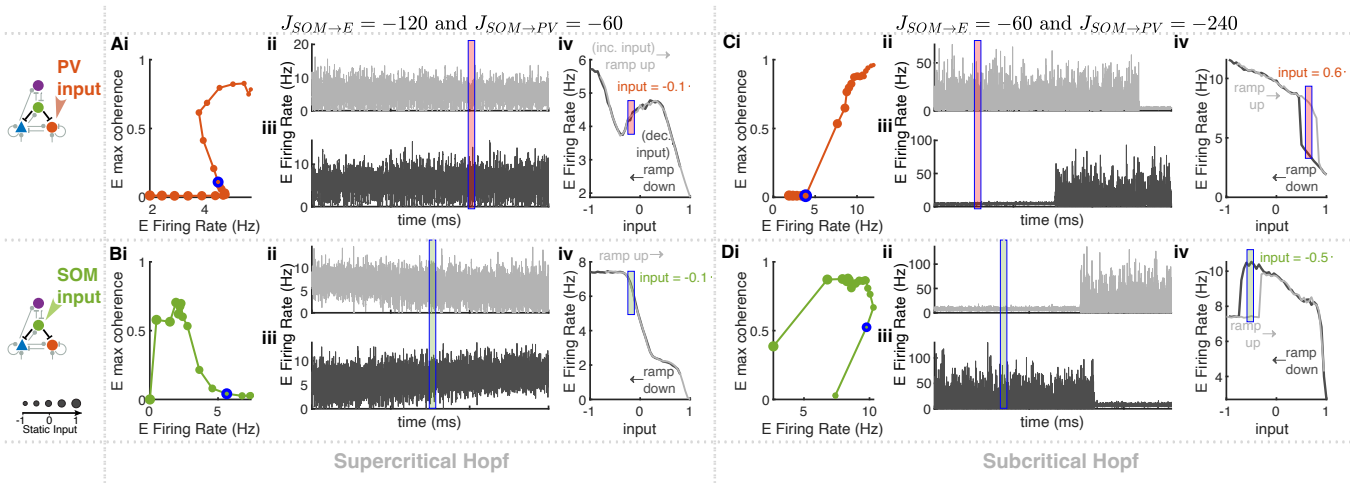


Figure S5: **VIP→SOM connection strength has little effect on modulation patterns. Related to Figures 5, 6.** Same format as Supplemental Figure S4. Static input was applied to SOM neurons. Modulation patterns are the same across VIP→SOM connection strengths  $J_{VIP \rightarrow SOM}$  except for in the network with the largest strength (darkest color). Networks with large  $J_{VIP \rightarrow SOM}$  become sensitive to small changes in external input to SOM. The firing rate of SOM neurons switches from zero to above 10 Hz and the rate of VIP neurons switches from about 20 Hz to near zero as input to SOM increases slightly (one step in panels C,D). Therefore, networks with large inhibition from VIP to SOM exhibit the WS state over only a limited parameter range and switch relatively abruptly between the SA and the SS states as input varies. Note that  $J_{VIP \rightarrow SOM} = -10$  is the default circuit parameter used in the main text.



**Figure S6: Modulation patterns in networks with different SOM→E and SOM→PV connection strengths. Related to Figure 5.** Static input was either applied to PV neurons (red) or SOM neurons (green). Each plot represents the maximum coherence of E neurons versus the average firing rate of E neurons. Marker sizes correspond to increasing static input to the target population. Rows represent (negative) increases in synaptic connection strength of SOM→E. Columns represent (negative) increases in synaptic connection strength of SOM→PV. The boxed plot features the same parameters as the default network. We find that when  $|J_{SOM \rightarrow E}| > |J_{SOM \rightarrow PV}|$  (lower triangle of the plots), the shapes of the modulation patterns for both input cases remain qualitatively consistent. When  $|J_{SOM \rightarrow PV}|$  is much larger than  $|J_{SOM \rightarrow E}|$ , the network exhibits abrupt changes from the SA to the SS state, and the firing rate and coherence of E neurons tend to vary in the same direction over all levels of input to PV. The orange shading in row two and green shading in row three highlight examples of changes in modulation patterns as  $|J_{SOM \rightarrow PV}|$  increases (across columns in the same row). Discontinuities in the shading are abrupt changes between adjacent dots (i.e., small changes in external input leading to large changes in coherence) along the modulation path.



**Figure S7: Hysteresis effects in networks with strong SOM→PV inhibition in response to ramping input. Related to Figures 3,4,5.** (A,B) The default network ( $J_{SOM \rightarrow E} = -120$  and  $J_{SOM \rightarrow PV} = -60$ ; Figures 3,4) with input applied to PV (A) or SOM (B) neurons. (C,D) Same as A,B for a network with stronger SOM to PV inhibition ( $J_{SOM \rightarrow E} = -60$ ,  $J_{SOM \rightarrow PV} = -240$ ) with input applied to PV (C) or SOM (D) neurons. Panel (i): Modulation path of E population firing rates versus E maximum coherence with varying static input (same format as Figures 4A, S6). The blue outlined marker in (i) represents the input value that is indicated in panels (ii)-(iv) by colored rectangles. Note that each panel (i) was generated with a sequence of fixed values of static input to the indicated population, and not with ramping input, which changes in time. Panel (ii): The population-averaged firing rate of E neurons as a function of time with slowly increasing input (*ramp up* case). Panel (iii): same as panel (ii) for slowly decreasing input (*ramp down* case). In the ramping input cases, external inputs were increased or decreased by a small incremental change,  $\pm 0.05$ , every 5 seconds. The 5-second interval allows sufficient time for the network to converge to a stationary state at the given input value. The colored rectangle in panels (ii) and (iii) indicates time intervals of the same input value in both ramping cases, which are aligned in time for comparison. Panel (iv): E population-average firing rates calculated within each 5-second interval of ramping input. The initial 250 ms of each interval was excluded to avoid transient activity. In the default network with  $|J_{SOM \rightarrow E}| > |J_{SOM \rightarrow PV}|$  (A,B), the E firing rate is the same for each fixed input value in both *ramp up* and *ramp down* cases. This suggests that there is no co-existence of multiple network states for any input value and that the transition from the SA to the SS state is likely through a supercritical Hopf bifurcation where the amplitude of oscillation increases gradually after bifurcation. In contrast, in a network with  $|J_{SOM \rightarrow E}| < |J_{SOM \rightarrow PV}|$  (C,D), the same input value results in different dynamic states in the *ramp up* and *ramp down* cases (Cii-Civ, Dii-Div, regions indicated by colored rectangles). This hysteresis effect demonstrates the co-existence of two network solutions, one asynchronous and one synchronous oscillation, over a range of input values. This suggests that oscillations arise via a subcritical Hopf bifurcation, where there is a sudden jump in the amplitude of oscillations after the bifurcation point, in networks with strong SOM→PV inhibition.

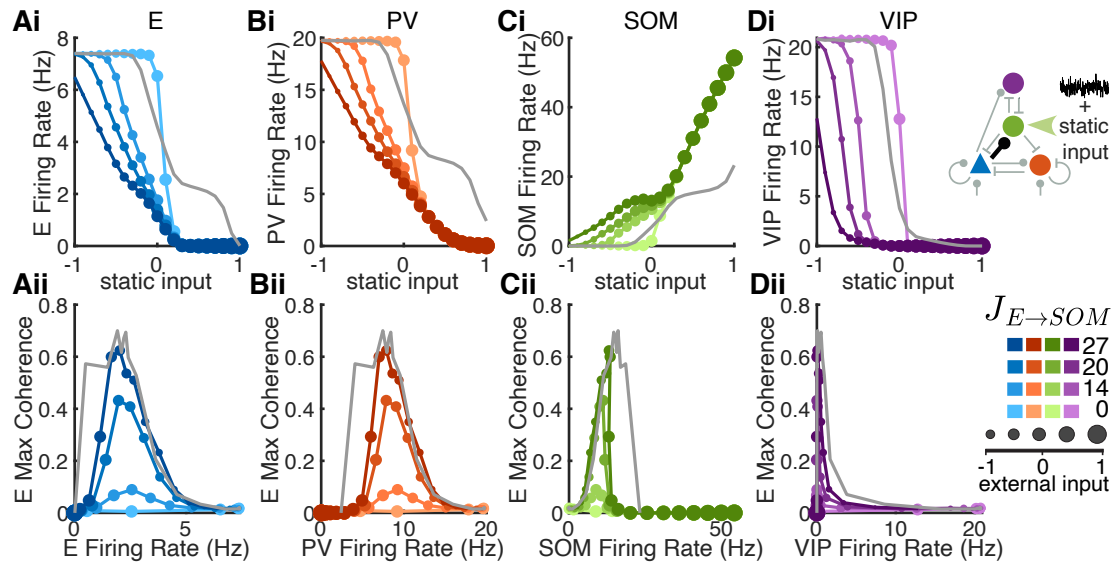


Figure S8: **Population rates and coherence in networks with different  $E \rightarrow SOM$  connection strengths. Related to Figure 6.** Both static input and colored noise were applied to SOM neurons. The colored noise was constructed as an OU process to match the mean and variance of the recurrent excitatory input that SOM neurons receive in the default network without external input (same noise input as in Figure 6A,C). Static input varied from -1 to 1. Column: (A) E, (B) PV, (C) SOM, (D) VIP population. Row (i): Average firing rates of each cell population with respect to static input value. Row (ii): The maximum coherence of E neurons compared to the population firing rates of each cell type. The grey curves are from the default network ( $J_{E \rightarrow SOM} = 27$ ), with SOM neurons receiving static input without the OU noise.

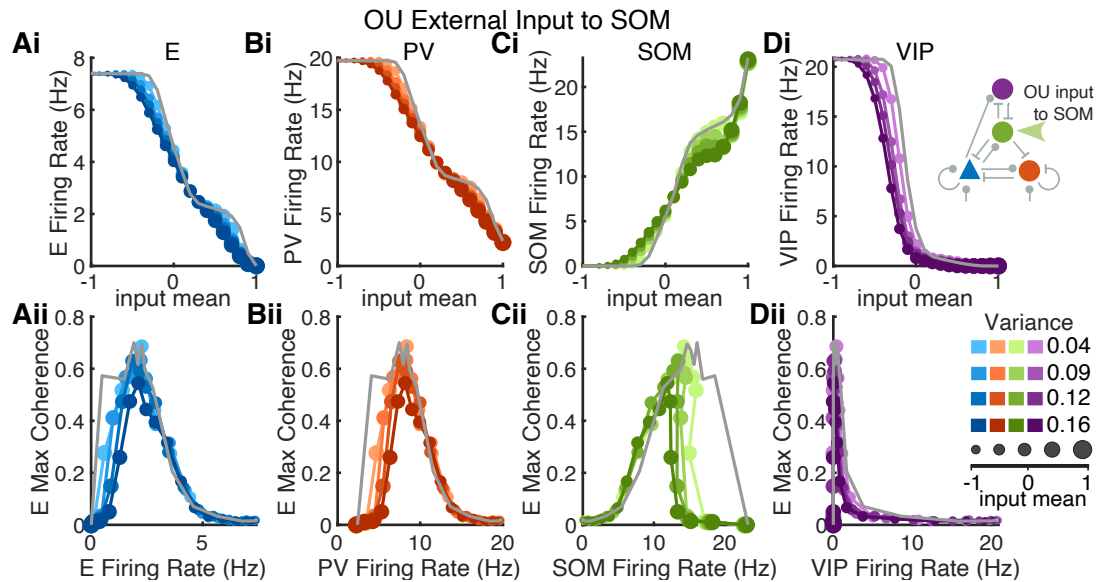


Figure S9: **Weak impacts of dynamic noise input parameters on population rates and coherence.** Related to Figure 7. Independent temporally-varying noise, modeled as an OU process of given mean (dot size) and variance (color shade), is applied to each SOM neuron. Columns show firing rate and coherence of (A) E, (B) PV, (C) SOM, (D) VIP populations. Row (i): Average firing rate of each cell population with respect to the mean value of OU input. Row (ii): The maximum coherence of E neurons compared to the population firing rates of each cell type. The grey curves are from the default network with SOM receiving static input.

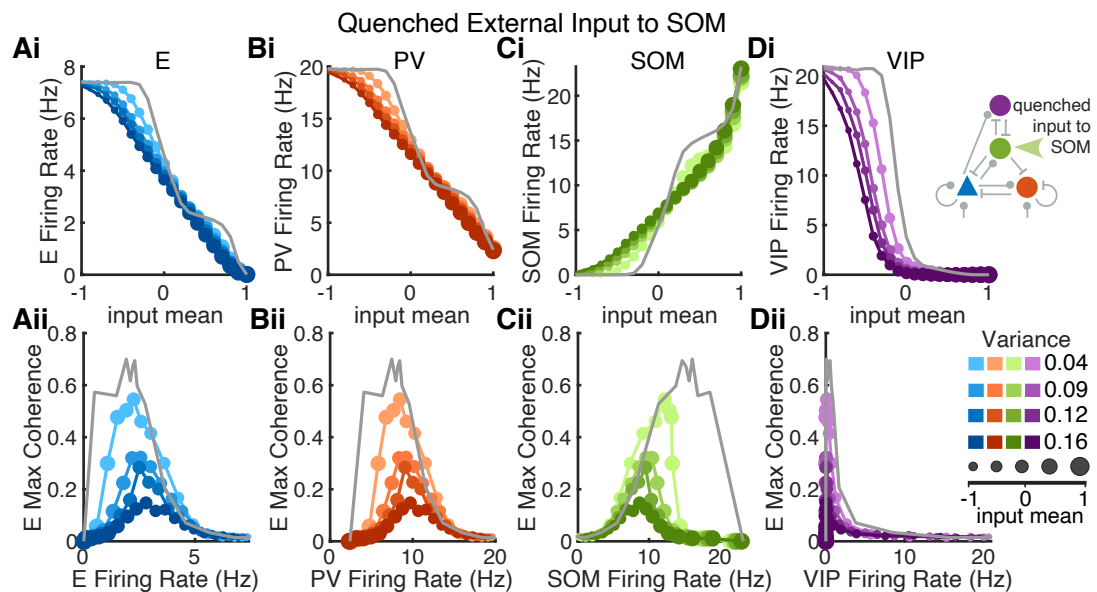


Figure S10: **Impacts of quenched inputs on population rates and coherence.** Related to Figure 7. Quenched input is spatially variable, but temporally invariant. Each SOM neuron receives an input value that is sampled from a Gaussian distribution with given mean (dot size) and variance (color shade). Columns show firing rate and coherence of (A) E, (B) PV, (C) SOM, (D) VIP populations. Row (i): Average firing rate of each cell population with respect to the mean value of the quenched input. Row (ii): The maximum coherence of E neurons compared to the population rates of each cell type. The grey curves are from the default network with SOM receiving static input.

615        Supplementary Video 1: Spiking activities of the spatially dependent spiking neuron network in the  
616 subcircuit asynchronous (SA) state (same parameters as in Figure 2A). Each dot indicates that the neuron  
617 at spatial position  $(x, y)$  fired within one millisecond of the time stamp shown on top. Color of each dot  
618 indicates the cell type of the neuron that fired (blue: E; red: PV; green: SOM; purple: VIP).

619

620        Supplementary Video 2: Same as Video 1 for the network in the weakly synchronous (WS) state (same  
621 parameters as in Figure 2B).

622

623        Supplementary Video 3: Same as Video 1 for the network in the strongly synchronous (SS) state (same  
624 parameters as in Figure 2C).

625