

COMMENTARY

 OPEN ACCESS

Sonic hedgehog signaling: A conserved mechanism for the expansion of outer radial glia and intermediate progenitor cells and for the growth and folding of the neocortex

Young-Goo Han 

Department of Developmental Neurobiology, Neurobiology and Brain Tumor Program, St. Jude Children's Research Hospital, Memphis, TN, USA

ABSTRACT

The expansion of outer radial glia (oRGs, also called basal RGs) and intermediate progenitor cells (IPCs) has played a key role in the evolutionary expansion and folding of the neocortex, resulting in superior sensorimotor and cognitive abilities. In particular, oRGs, which are critical for both the increased production and lateral dispersion of neurons, are rare in lissencephalic species but vastly expanded in gyrencephalic species. However, the mechanisms that expand oRGs and IPCs are not well understood. We recently identified Sonic hedgehog (Shh) signaling as the first known signaling pathway necessary and sufficient to expand both oRGs and IPCs. Elevated Shh signaling in the embryonic neocortex leads to neocortical expansion and folding with normal cytoarchitecture in otherwise smooth mouse neocortex, whereas the loss of Shh signaling decreases oRGs, IPCs, and neocortical size. We also showed that SHH signaling activity in fetal neocortex is stronger in humans than in mice and that blocking SHH signaling decreases oRGs in human cerebral organoids. Shh signaling may be a conserved mechanism that promotes oRG and IPC expansion, driving neocortical growth and folding in humans and other species. Understanding the mechanisms underlying species-specific differences in Shh signaling activity and how Shh signaling expands oRGs and IPCs will provide insights into the mechanisms of neocortical development and evolution.

ARTICLE HISTORY

Received 9 August 2016
Revised 12 September 2016
Accepted 25 September 2016

KEYWORDS

gyrification; hedgehog; neocortex; neural progenitor; neural stem cell; radial glia

Introduction

The neocortex, a 6-layered structure that computes high-order sensory, motor, and cognitive processes, is both a hallmark and a remarkably divergent part of mammalian brains. Although the layering and thickness of the neocortex remained relatively constant over the course of evolution, its surface expanded dramatically and folded in certain species, resulting in superior sensorimotor and cognitive abilities. Neocortical expansion and folding require 2 coordinated processes that depend on neural progenitors: the increased production of neural cells and their lateral dispersion.¹⁻⁴ The primary neural progenitors are radial glia (RGs), whose cell bodies reside in the ventricular zone (VZ) at the apical side of the developing brain and are thus called ventricular RGs (vRGs) or apical RGs (aRGs). vRGs have a radial process that extends to the pial surface and serves as a scaffold for the migration of newborn neurons toward the cortical plate, where later-born neurons settle above early

born neurons to form distinct neuronal layers in an inside-out fashion. vRGs produce neurons directly or indirectly via intermediate progenitor cells (IPCs) or outer RGs (oRGs, also called basal RGs) that occupy the subventricular zone (SVZ) basal to the VZ.⁵⁻¹¹ Recent studies suggest that the expansion of oRGs, which not only increases neuron production but also spurs the lateral dispersion of neurons via radial processes of oRGs, plays a critical role in neocortical growth and folding.¹²⁻²⁰ Consistently, oRGs are rare in species with small/smooth brains but are greatly expanded in species with large/folded brains.^{10-14,19} Nonetheless, oRGs are present in all the mammalian lineages that have been examined.^{14,19,21,22} Furthermore, neocortical folding is prevalent in many mammalian lineages, including marsupials and even egg-laying monotremes (<http://neurosciencelibrary.org/index.html>). Therefore, mechanisms to induce oRG expansion and neocortical growth and folding appear to have been conserved from a common

CONTACT Young-Goo Han  young-goo.han@stjude.org  St. Jude Children's Research Hospital, 262 Danny Thomas Place, Memphis, TN, 38105, USA.

© 2016 Young-Goo Han. Published with license by Taylor & Francis.

This is an Open Access article distributed under the terms of the Creative Commons Attribution-Non-Commercial License (<http://creativecommons.org/licenses/by-nc/3.0/>), which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited. The moral rights of the named author(s) have been asserted.

ancestor of mammals but selectively fortified or inactivated in different lineages, giving rise to large/folded or small/smooth brains.^{14,19,21-23} Recently, we provided evidence indicating that Sonic hedgehog (Shh) signaling has been central to these mechanisms²⁴ (Fig. 1).

Shh signaling expands oRGs and IPCs, leading to neocortical growth and folding in mice

Shh signaling is a conserved mechanism that regulates many aspects of animal development. Notably, mutations that attenuate SHH signaling cause microcephaly in humans,²⁵⁻²⁷ suggesting that SHH signaling regulates brain size in humans. In mice, the loss of Shh signaling in the neocortex decreases its size;²⁸ however, the gain of Shh signaling did not increase the

neocortical size but disrupted the patterning and specification of neural progenitors.²⁹⁻³² To study the role of Shh signaling in neocortical development beyond patterning and specification, we conditionally removed or activated Smoothed (Smo, an activator of Shh signaling) by using *GFAP::Cre*, which induces recombination at embryonic day 13.5, when the patterning and specification of neural progenitors has already been established. The expression of a constitutively active form of Smo (SmoM2) in vRGs and their progenies in *GFAP::Cre; SmoM2^{f/+}* (*SmoM2* mutant) mice significantly increased the size of the neocortex. Remarkably, SmoM2 induced folding in the cingulate cortex without affecting the normal cytoarchitecture. As in the folded brains of larger mammals, in which upper-layer (layer II and III) neurons are much more expanded than are deeper-layer neurons and the white

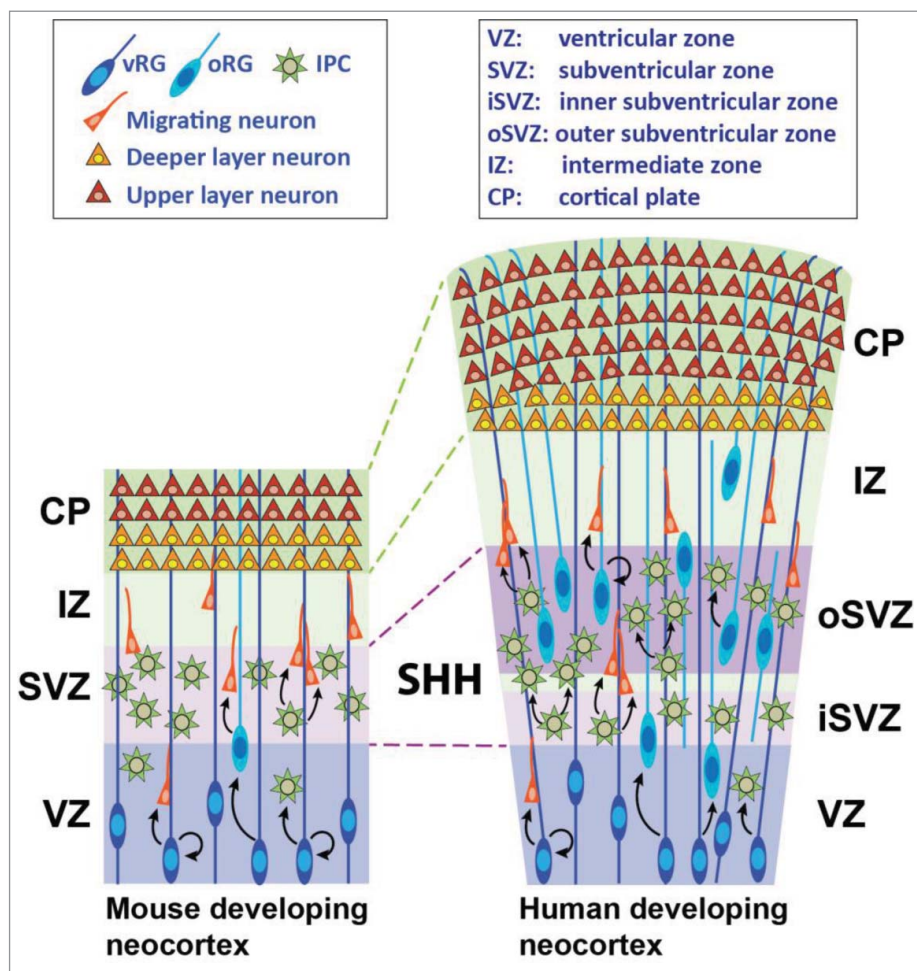


Figure 1. In species with a large and folded neocortex, oRGs and IPCs are expanded in the cortical SVZ, which is divided into the inner and outer SVZs (iSVZ and oSVZ). SHH promotes this expansion, leading to neocortical growth and folding. Mechanistically, SHH expands oRGs by increasing their self-renewal and production from vRGs and expands IPCs by increasing their self-amplifying divisions in the SVZ. *Shh* in mice and *SHH* in humans are highly expressed in the VZ of the ventral forebrain, suggesting trans-ventricular delivery of SHH proteins to the neocortex.

matter extends into the gyri, upper-layer neurons were specifically increased in the folded cingulate cortex of *SmoM2* mutants, and the corpus callosum was extended into the folded area. Upper-layer neurons were not increased in the lateral part of the neocortex that did not show folding, suggesting that increased upper-layer neurons induced neocortical folding. The medial-to-lateral gradient of the upper-layer neuron increase reflected the expression pattern of *SmoM2* in *GFAP::Cre; SmoM2^{fl/+}* mutants. The expression of *SmoM2* in *Nestin::Cre; SmoM2^{fl/+}* or *Nestin::CreER; SmoM2^{fl/+}* mice, which do not show such a *SmoM2* expression gradient, induced folding outside the cingulate cortex too. Therefore, the mechanism that underlies neocortical folding in *SmoM2* mutants must be a general one rather than being specific to the cingulate cortex.

To understand the cellular mechanism by which *SmoM2* expanded upper-layer neurons, we investigated whether and how *SmoM2* changed the number and behavior of neural progenitors. The number of vRGs was not changed, but the numbers of oRGs and IPCs were greatly expanded in *SmoM2* mutants. Notably, *SmoM2* expanded oRGs and IPCs via distinct mechanisms by affecting the behavior of all 3 types of progenitor. *SmoM2* did not affect the proliferation rate of oRGs or vRGs, but it increased self-renewal of oRGs and changed the vRG division modes to produce more oRGs and fewer IPCs and neurons. vRGs dividing on an axis horizontal to the ventricular surface mostly produce neurons or IPCs, whereas those dividing nonhorizontally produce oRGs.^{11,33} Nonhorizontal divisions were markedly increased in *SmoM2* mutants, as compared to controls. Thus, *SmoM2* expanded oRGs by promoting their initial generation from vRGs and their subsequent self-renewal. In contrast, *SmoM2* decreased the generation of IPCs from vRGs but increased their proliferation and self-amplifying divisions, leading to their great expansion in the SVZ. Similarly, IPCs of primates divide to make more IPCs before producing neurons,^{12,34} whereas IPCs of mice and rats mainly divide just once to produce 2 neurons.⁶⁻⁸

Consistent with the results of gain-of-function experiments, we found that endogenous Shh signaling is required to expand oRGs, IPCs, upper-layer neurons, and the neocortex. The loss of Shh signaling in *GFAP::Cre; Smo^{fl/fl}* mutants caused phenotypes opposite to those of *SmoM2* mutants.

Compared to wild-type mice, the *GFAP::Cre; Smo^{fl/fl}* mice had abnormally small brains with fewer upper-layer neurons, significantly fewer oRGs and IPCs (but a similar number of vRGs), and a decreased proportion of vRGs dividing nonhorizontally. Taken together, these findings show that Shh signaling promotes key developmental characteristics of large and folded brains, namely oRG expansion and self-amplifying IPC division, which a comparative study of 102 mammalian brains proposed to be necessary and sufficient for the evolution of an expanded and folded neocortex.³⁵

Shh signaling is required for human oRG expansion

Based on our mouse study, we predicted that Shh signaling activity would correlate with the number of oRGs and IPCs and be stronger in gyrencephalic species than in lissencephalic species. Indeed, by comparing RNAseq data and the results of *in situ* hybridization experiments, we found that SHH signaling activity is stronger in human fetal neocortex than in mouse embryonic neocortex. Furthermore, the developmental change in SHH signaling activity correlated with oRG expansion in human fetal cortex. In mice, the regional difference in Shh signaling activity in the neocortex correlated with the number of oRGs. A previous study in ferrets showed that Shh signaling activity is significantly higher in the VZ area that gives rise to the thick SVZ containing many oRGs than in the VZ area that gives rise to the thin SVZ containing fewer oRGs.³⁶

To functionally test whether SHH signaling expanded human oRGs and IPCs, we employed human cerebral organoids that recapitulate key features of the developing human cortex, including abundant oRGs.³⁷⁻⁴¹ In contrast to mouse vRGs, but similar to human vRGs in slice culture,³³ more than half of the vRGs in the organoids divided nonhorizontally. SANT1 (a Smo inhibitor) strongly decreased the incidence of nonhorizontal division, similar to the low incidence of nonhorizontal division in mouse vRGs, and subsequently decreased the number of oRG-like cells outside the VZ, whereas neither effect was seen with SAG (a Smo agonist). Accordingly, we showed that SHH signaling was intrinsically active in the organoids and could be blocked by SANT1 but could not be further increased by SAG. The number of IPCs was

very low and was not significantly affected by SANTI or SAG. These results suggest that Shh signaling promotes oRG expansion in gyrencephalic species.

Conclusion and future directions

Our study showed that Shh signaling promotes oRG and IPC expansion, leading to neocortical growth and folding. Shh signaling is the first signaling pathway with these properties to be identified. This role of SHH signaling appears to be conserved, at least in mice and humans. SHH signaling activity is stronger in human fetal cortex than in mouse embryonic cortex and correlates with the number of oRGs in both species, suggesting that Shh signaling may have played important roles in the evolutionary growth and folding of the neocortex. These findings linking Shh signaling with oRG and IPC expansion, neocortical growth, and evolution raise important questions: what are the mechanisms underlying the difference in Shh signaling activity in the developing neocortex of humans and mice; what are the molecular mechanisms by which Shh signaling differentially affects 3 different neural progenitor types; and are these mechanisms conserved? The answers to these questions will provide fundamental insights into the development and evolution of mammalian brains.

Disclosure of potential conflicts of interest

No potential conflicts of interest were disclosed.

Funding

The work in my laboratory is supported by NIH/NCI Cancer Center Core Support grant CA021765 (SJCRC), the Sontag Foundation Distinguished Scientist Award, a Whitehall Foundation research grant, and ALSAC.

ORCID

Young-Goo Han  <http://orcid.org/0000-0002-4008-294X>

References

- [1] Lui JH, Hansen DV, Kriegstein AR. Development and evolution of the human neocortex. *Cell* 2011; 146:18-36; PMID:21729779; <http://dx.doi.org/10.1016/j.cell.2011.06.030>
- [2] Florio M, Huttner WB. Neural progenitors, neurogenesis and the evolution of the neocortex. *Development* 2014; 141:2182-94; PMID:24866113; <http://dx.doi.org/10.1242/dev.090571>
- [3] Sun T, Hevner RF. Growth and folding of the mammalian cerebral cortex: from molecules to malformations. *Nat Rev Neurosci* 2014; 15:217-32; PMID:24646670; <http://dx.doi.org/10.1038/nrn3707>
- [4] Dehay C, Kennedy H, Kosik KS. The outer subventricular zone and primate-specific cortical complexification. *Neuron* 2015; 85:683-94; PMID:25695268; <http://dx.doi.org/10.1016/j.neuron.2014.12.060>
- [5] Noctor SC, Flint AC, Weissman TA, Dammerman RS, Kriegstein AR. Neurons derived from radial glial cells establish radial units in neocortex. *Nature* 2001; 409:714-20; PMID:11217860; <http://dx.doi.org/10.1038/35055553>
- [6] Noctor SC, Martínez-Cerdeño V, Ivic L, Kriegstein AR. Cortical neurons arise in symmetric and asymmetric division zones and migrate through specific phases. *Nat Neurosci* 2004; 7:136-44; PMID:14703572; <http://dx.doi.org/10.1038/nn1172>
- [7] Haubensak W, Attardo A, Denk W, Huttner WB. Neurons arise in the basal neuroepithelium of the early mammalian telencephalon: a major site of neurogenesis. *Proc Natl Acad Sci U S A* 2004; 101:3196-201; PMID:14963232; <http://dx.doi.org/10.1073/pnas.0308600100>
- [8] Miyata T, Kawaguchi A, Saito K, Kawano M, Muto T, Ogawa M. Asymmetric production of surface-dividing and non-surface-dividing cortical progenitor cells. *Development* 2004; 131:3133-45; PMID:15175243; <http://dx.doi.org/10.1242/dev.01173>
- [9] Gal JS, Morozov YM, Ayoub AE, Chatterjee M, Rakic P, Haydar TF. Molecular and morphological heterogeneity of neural precursors in the mouse neocortical proliferative zones. *J Neurosci* 2006; 26:1045-56; PMID:16421324; <http://dx.doi.org/10.1523/JNEUROSCI.4499-05.2006>
- [10] Wang X, Tsai JW, LaMonica B, Kriegstein AR. A new subtype of progenitor cell in the mouse embryonic neocortex. *Nat Neurosci* 2011; 14:555-61; PMID:21478886; <http://dx.doi.org/10.1038/nn.2807>
- [11] Shitamukai A, Konno D, Matsuzaki F. Oblique radial glial divisions in the developing mouse neocortex induce self-renewing progenitors outside the germinal zone that resemble primate outer subventricular zone progenitors. *J Neurosci* 2011; 31:3683-95; PMID:21389223; <http://dx.doi.org/10.1523/JNEUROSCI.4773-10.2011>
- [12] Hansen DV, Lui JH, Parker PRL, Kriegstein AR. Neurogenic radial glia in the outer subventricular zone of human neocortex. *Nature* 2010; 464:554-61; PMID:20154730; <http://dx.doi.org/10.1038/nature08845>
- [13] Fietz SA, Kelava I, Vogt J, Wilsch-Bräuninger M, Stenzel D, Fish JL, Corbeil D, Riehn A, Distler W, Nitsch R, Huttner WB. OSVZ progenitors of human and ferret neocortex are epithelial-like and expand by integrin signaling. *Nat Neurosci* 2010; 13:690-9; PMID:20436478; <http://dx.doi.org/10.1038/nn.2553>
- [14] Reillo I, de Juan Romero C, García-Cabezas MÁ, Borrell V. A role for intermediate radial glia in the tangential expansion of the mammalian cerebral cortex. *Cereb Cortex* 2011; 21:1674-94; PMID:21127018; <http://dx.doi.org/10.1093/cercor/bhq238>
- [15] Smart IHM, Dehay C, Giroud P, Berland M, Kennedy H. Unique morphological features of the proliferative zones

- and postmitotic compartments of the neural epithelium giving rise to striate and extrastriate cortex in the monkey. *Cereb Cortex* 2002; 12:37-53; PMID:11734531; <http://dx.doi.org/10.1093/cercor/12.1.37>
- [16] Lukaszewicz A, Savatier P, Cortay V, Giroud P, Huisoud C, Berland M, Kennedy H, Dehay C. G1 phase regulation, area-specific cell cycle control, and cytoarchitectonics in the primate cortex. *Neuron* 2005; 47:353-64; PMID:16055060; <http://dx.doi.org/10.1016/j.neuron.2005.06.032>
- [17] Stahl R, Walcher T, De Juan Romero C, Pilz GA, Cappello S, Irmeler M, Sanz-Aquela JM, Beckers J, Blum R, Borrell V, et al. *Trnp1* regulates expansion and folding of the mammalian cerebral cortex by control of radial glial fate. *Cell* 2013; 153:535-49; PMID:23622239; <http://dx.doi.org/10.1016/j.cell.2013.03.027>
- [18] Florio M, Albert M, Taverna E, Namba T, Brandl H, Lewitus E, Haffner C, Sykes A, Wong FK, Peters J, et al. Human-specific gene *ARHGAP11B* promotes basal progenitor amplification and neocortex expansion. *Science* 2015; 347:1465-70; PMID:25721503; <http://dx.doi.org/10.1126/science.aaa1975>
- [19] Martínez-Cerdeño V, Cunningham CL, Camacho J, Antczak JL, Prakash AN, Cziep ME, Walker AI, Noctor SC. Comparative analysis of the subventricular zone in rat, ferret and macaque: evidence for an outer subventricular zone in rodents. *PLoS One* 2012; 7:e30178; <http://dx.doi.org/10.1371/journal.pone.0030178>
- [20] Nonaka-Kinoshita M, Reillo I, Artegiani B, Martínez-Martínez MÁ, Nelson M, Borrell V, Calegari F. Regulation of cerebral cortex size and folding by expansion of basal progenitors. *EMBO J* 2013; 32:1817-28; PMID:23624932; <http://dx.doi.org/10.1038/emboj.2013.96>
- [21] García-Moreno F, Vasistha NA, Trevia N, Bourne JA, Molnár Z. Compartmentalization of cerebral cortical germinal zones in a lissencephalic primate and gyrencephalic rodent. *Cereb Cortex* 2012; 22:482-92; <http://dx.doi.org/10.1093/cercor/bhr312>
- [22] Kelava I, Reillo I, Murayama AY, Kalinka AT, Stenzel D, Tomancak P, Matsuzaki F, Lebrand C, Sasaki E, Schwamborn JC, et al. Abundant occurrence of basal radial glia in the subventricular zone of embryonic neocortex of a lissencephalic primate, the common marmoset *Callithrix jacchus*. *Cereb Cortex* 2012; 22:469-81; PMID:22114084; <http://dx.doi.org/10.1093/cercor/bhr301>
- [23] Hevner RF, Haydar TF. The (not necessarily) convoluted role of basal radial glia in cortical neurogenesis. *Cereb Cortex* 2012; 22:465-8; PMID:22116731; <http://dx.doi.org/10.1093/cercor/bhr336>
- [24] Wang L, Hou S, Han YG. Hedgehog signaling promotes basal progenitor expansion and the growth and folding of the neocortex. *Nat Neurosci* 2016; 19:888-96; PMID:27214567; <http://dx.doi.org/10.1038/nn.4307>
- [25] Heussler HS, Suri M, Young ID, Muenke M. Extreme variability of expression of a Sonic Hedgehog mutation: attention difficulties and holoprosencephaly. *Arch Dis Child* 2002; 86:293-6; PMID:11919111; <http://dx.doi.org/10.1136/adc.86.4.293>
- [26] Derwińska K, Smyk M, Cooper ML, Bader P, Cheung SW, Stankiewicz P. *PTCH1* duplication in a family with microcephaly and mild developmental delay. *Eur J Hum Genet* 2009; 17:267-71; PMID:18830227; <http://dx.doi.org/10.1038/ejhg.2008.176>
- [27] Izumi K, Hahn A, Christ L, Curtis C, Neilson DE. Familial 9q22.3 microduplication spanning *PTCH1* causes short stature syndrome with mild intellectual disability and dysmorphic features. *Am J Med Genet* 2011; 155A:1384-9; PMID:21567912; <http://dx.doi.org/10.1002/ajmg.a.33959>
- [28] Komada M, Saitsu H, Kinboshi M, Miura T, Shiota K, Ishibashi M. Hedgehog signaling is involved in development of the neocortex. *Development* 2008; 135:2717-27; PMID:18614579; <http://dx.doi.org/10.1242/dev.015891>
- [29] Wang H, Ge G, Uchida Y, Luu B, Ahn S. *Gli3* is required for maintenance and fate specification of cortical progenitors. *J Neurosci* 2011; 31:6440-8; PMID:21525285; <http://dx.doi.org/10.1523/JNEUROSCI.4892-10.2011>
- [30] Dave RK, Ellis T, Toumpas MC, Robson JP, Julian E, Adolphe C, Bartlett PF, Cooper HM, Reynolds BA, Wainwright BJ. Sonic hedgehog and Notch signaling can cooperate to regulate neurogenic divisions of neocortical progenitors. *PLoS One* 2011; 6:e14680; PMID:21379383; <http://dx.doi.org/10.1371/journal.pone.0014680>
- [31] Shikata Y, Okada T, Hashimoto M, Ellis T, Matsumaru D, Shiroishi T, Ogawa M, Wainwright B, Motoyama J. *Ptch1*-mediated dosage-dependent action of Shh signaling regulates neural progenitor development at late gestational stages. *Dev Biol* 2011; 349:147-59; PMID:20969845; <http://dx.doi.org/10.1016/j.ydbio.2010.10.014>
- [32] Yabut OR, Fernández G, Huynh T, Yoon K, Pleasure SJ. Suppressor of fused is critical for maintenance of neuronal progenitor identity during corticogenesis. *Cell Rep* 2015; 12:2021-34; PMID:26387942; <http://dx.doi.org/10.1016/j.celrep.2015.08.031>
- [33] LaMonica BE, Lui JH, Hansen DV, Kriegstein AR. Mitotic spindle orientation predicts outer radial glial cell generation in human neocortex. *Nat Commun* 2013; 4:1665; PMID:23575669; <http://dx.doi.org/10.1038/ncomms2647>
- [34] Betizeau M, Cortay V, Patti D, Pfister S, Gautier E, Bellemin-Ménard A, Afanassieff M, Huissoud C, Douglas RJ, Kennedy H, Dehay C. Precursor diversity and complexity of lineage relationships in the outer subventricular zone of the primate. *Neuron* 2013; 80:442-57; PMID:24139044; <http://dx.doi.org/10.1016/j.neuron.2013.09.032>
- [35] Lewitus E, Kelava I, Kalinka AT, Tomancak P, Huttner WB. An adaptive threshold in mammalian neocortical evolution. *PloS Biol* 2014; 12:e1002000; PMID:25405475; <http://dx.doi.org/10.1371/journal.pbio.1002000>
- [36] de Juan Romero C, Bruder C, Tomasello U, Sanz-Anquela JM, Borrell V. Discrete domains of gene expression in germinal layers distinguish the development of gyrencephaly.

- EMBO J 2015; 34:1859-74; PMID:25916825; <http://dx.doi.org/10.15252/embj.201591176>
- [37] Lancaster MA, Renner M, Martin C-A, Wenzel D, Bicknell LS, Hurles ME, Homfray T, Penninger JM, Jackson AP, Knoblich JA. Cerebral organoids model human brain development and microcephaly. *Nature* 2013; 501:373-9; PMID:23995685; <http://dx.doi.org/10.1038/nature12517>
- [38] Mariani J, Coppola G, Zhang P, Abyzov A, Provini L, Tomasini L, Amenduni M, Szekely A, Palejev D, Wilson M, et al. FOXP1-dependent dysregulation of GABA/glutamate neuron differentiation in autism spectrum disorders. *Cell* 2015; 162:375-90; PMID:26186191; <http://dx.doi.org/10.1016/j.cell.2015.06.034>
- [39] Camp JG, Badsha F, Florio M, Kanton S, Gerber T, Wilsch-Bräuninger M, Lewitus E, Sykes A, Hevers W, Lancaster M, et al. Human cerebral organoids recapitulate gene expression programs of fetal neocortex development. *Proc Natl Acad Sci U S A* 2015; 112:15672-7; PMID:26644564; <http://dx.doi.org/10.1073/pnas.1508055112>
- [40] Otani T, Marchetto MC, Gage FH, Simons BD, Livesey FJ. 2D and 3D stem cell models of primate cortical development identify species-specific differences in progenitor behavior contributing to brain size. *Cell Stem Cell* 2016; 18:467-80; PMID:27049876; <http://dx.doi.org/10.1016/j.stem.2016.03.003>
- [41] Qian X, Nguyen HN, Song MM, Hadiono C, Ogden SC, Hammack C, Yao B, Hamersky GR, Jacob F, Zhong C, et al. Brain-region-specific organoids using mini-bioreactors for modeling ZIKV exposure. *Cell* 2016; 165:1238-54; PMID:27118425; <http://dx.doi.org/10.1016/j.cell.2016.04.032>