

## Research Article

# Effect of Environmental Disturbance on the Population of Sandflies and *Leishmania* Transmission in an Endemic Area of Venezuela

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The exploitation of new wilderness areas with crops is increasing and traditional crop substitution has been modified by new more productive crops. The results show the anthropogenic disturbance effect on the sandflies population and *Leishmania* transmission in endemic areas of Venezuela. Three agroecosystems with variable degrees of ecological disturbance, forest (conserved), cacao (fragmented), and orangery (disturbed), were selected. Four methods to sandfly capture were used; the specimens were identified and infected with *Leishmania*. Diversity, population structure, ANOVA, Tukey test, and simple correlation analysis were carried out. Shannon traps were able to capture 94.7% of the total sandflies, while CDC light traps, Sticky traps, and direct suction just captured 2.2%, 1.2%, and 0.9%, respectively. The results showed the effect of ecological disturbance degree on the composition of sandflies and population structure, revealing a dominance level increased but decreased on the diversity and richness of sandflies species in the greatest ecological disturbance area in relation to areas with less organic disturbance. Environments more disturbed cause adaptability of certain species such as *Lutzomyia gomezi* and *Lutzomyia walkeri*. These changes on the composition of sandflies population and structure emerging species could cause increasing of leishmaniasis transmission.

## 1. Introduction

The distribution of sandflies correlated with the appearance of cases of leishmaniasis in endemic regions, especially in forested areas. However, with human intervention and the disappearance of their natural habitat, some species appear to have adapted to degraded habitats, contributing to expansion of their spatial distribution and the spread of leishmaniasis [1–3].

The main factors involved in the transmission of tegumentary leishmaniasis are related to deforestation, urbanization, the presence of domestic animals, and the development of agriculture, particularly the cultivation of cocoa, banana, and coffee [4]. Anthropogenic factors tend to alter the com-

position and behavior of populations of sandflies. While some species of sandflies have disappeared, others have become more abundant and have adapted to synanthropic environments by changing their behavior [5–10].

The exploitation of wilderness areas for cultivation is increasing. In particular, this expansion has replaced traditional crops with crops that are more productive, which has led to changes in sandflies populations related to altered patterns of dispersal and spatial distribution of these species in new areas [10–14], because these changes may involve a greater risk of transmission [3, 15]. Thus, an understanding between habitat variation and sandflies populations is essential, and to examine whether these changes can increase the risk of transmission of *Leishmania*, we studied populations

of sandflies in a conserved area and two distinct agroecosystems.

## 2. Materials and Methods

**2.1. Study Area.** The agroecosystems located in the Parroquia Caño El Tigre, Zea Municipality, Merida, Venezuela, were studied. These regions have an average elevation of 300–400 meters above sea level, covering an area of 135 km<sup>2</sup>, which includes 9,595 inhabitants, a tropical rainforest climate, and temperatures that range between 25 and 30°C. The main economic activities of the region are agriculture and cattle.

**2.2. Determination of Environmental and Anthropogenic Variables.** According to methods previously described in the literature, indicators associated with ecoepidemiological levels were recorded using a data sheet that identified the environmental and anthropogenic variables related to the presence of sandflies. These variables included the climatic conditions (elevation, temperature, and relative humidity), the presence of natural or anthropogenic water bodies, dominant vegetation stratum, crops and animals present, and the level of human interference (e.g., logging, burning, and use of fertilizer).

**2.3. Degrees of Disturbance.** The agroecosystems were characterised according to the degree of human modification [3]. The aspects observed concern the vegetation and the presence of both dwellings and animal shelters. Three agroecosystems were selected with varying degrees of ecological disturbance: (1) a conserved area, predominantly forest, characterized by abundant primary vegetation; (2) a fragmented area in which primary vegetation was partially replaced by cocoa crops without management; and (3) a disturbed area with complete replacement of primary vegetation, resulting from the degradation caused by human activity related to citrus cultivation, specifically oranges (Figure 1). The distance between the agroecosystems is approximately 10 km.

**2.4. Capture of Sandflies.** Captures of adult sandflies specimens were performed for 12-month period, from January 2012 to January 2013 at three agrosystems. The captures were conducted at the peridomicile areas, using one Shannon traps, three CDC traps, six Sticky traps, and direct suction with an oral grabber. Sampling was conducted after sunset, when sandflies are most active, between 18:30 h and 20:00 h; with minimum of one capture by months each collection agrosystem. Shannon traps were conducted in peridomicile areas with three collectors, the CDC light traps were placed in proximity of houses (poultry houses, breeding pigs, tree, etc.), and Sticky strips (white paper sheets 21.6 × 27.9 cm coated with castor oil) were placed indoors or outdoors in proximity of houses. The traps were distributed over 1 ha of the agrosystems and arranged in transect with at least 20 m of distance between each trap.

Conserved



Fragmented



Disturbed



FIGURE 1: Different degrees of ecological disturbance of the agroecosystems.

**2.5. Determination of Natural Infection and Sandflies Identification.** To determine the presence of *Leishmania* promastigotes [16], the digestive system was extracted via the dissection of live females and examined using phase contrast microscopy at 400x magnification. We then performed rapid identification of fresh sandflies individuals, and body or representative segments were subsequently cleared in Nesbitt solution for 24 hours and were prepared and mounted on slides using Berlese's medium to identify females for corroboration of the species by comparative external and internal morphology [17].

**2.6. Analysis.** The methods used were based on community structure, proportional abundance, dominance index, and Margalef's index which was used to calculate biodiversity [18]. An analysis of the different captures among and agrosystems was conducted using a cluster analysis which was performed using PCORD.5 software (License belonging to ICAE). The comparison for the different agrosystems was conducted using analysis of variance (ANOVA) which was

TABLE 1: Ecoepidemiological characteristics and the degree of disturbance of the agroecosystems.

	Forest	Cocoa	Orangery
Environmental influence			
Temperature	24.8–27.9°C	26.7–32°C	24.8–26.9°C
Relative wetness	65.5–82%	52–75.7%	75.2–83%
Water bodies	Yes	No	No
Type of vegetation	Arboreal	Shrubby and herbaceous	Shrubby
Anthropogenic influence			
Animal presence	No	Yes (domestic)	Yes (domestic and breeding)
Crop presence	No	Yes (cocoa, banana)	Yes (citrus fruit)
Human influence	Felling of trees	Garbage	Chemical contamination
Stored and irrigation water	No	Cistern	Irrigation
Ecological disturbance	Conserved	Fragmented	Disturbed
Level of disturbance	Low	Medium	High

TABLE 2: The species abundance in the agroecosystems studied.

Species	Agroecosystems											
	Forest				Cocoa				Orangery			
	<i>N</i>	%	pi	( $\lambda$ )	<i>N</i>	%	pi	( $\lambda$ )	<i>N</i>	%	pi	( $\lambda$ )
<i>L. gomezi</i> *	128	41.16	0.41	0.17	25	29.76	0.30	0.09	73	79.35	0.79	0.63
<i>L. ovallesi</i> *	126	40.51	0.41	0.16	36	42.86	0.43	0.18	7	7.61	0.08	0.01
<i>L. walkeri</i> *	24	7.72	0.08	0.01	3	3.57	0.04	0.00	3	3.26	0.03	0.00
<i>L. trinidadensis</i> **	2	0.64	0.01	0.00	12	14.29	0.14	0.02	5	5.43	0.05	0.00
<i>L. panamensis</i> *	11	3.54	0.04	0.00	—	—	—	—	—	—	—	—
<i>L. atroclavata</i> **	2	0.64	0.01	0.00	1	1.19	0.01	0.00	1	1.09	0.01	0.00
<i>L. cayennensis</i>	1	0.32	0.00	0.00	—	—	—	—	0	—	—	—
<i>L. hernandezii</i>	6	1.93	0.02	0.00	—	—	—	—	2	2.17	0.02	0.00
<i>L. migonei</i> *	2	0.64	0.01	0.00	—	—	—	—	—	—	—	—
<i>L. olmeca nociva</i>	—	—	—	—	1	1.19	0.01	0.00	—	—	—	—
<i>L. pilosa</i>	1	0.32	0.00	0.00	—	—	0.00	—	—	—	—	—
<i>L. puntigeniculata</i>	2	0.64	0.01	0.00	—	—	0.00	—	—	—	—	—
<i>L. shannoni</i>	3	0.96	0.01	0.00	1	1.19	0.01	0.00	—	—	—	—
<i>L. spinicrassa</i> *	—	—	—	—	—	—	—	—	1	1.09	0.01	0.00
<i>L. venezuelensis</i> **	1	0.32	0.00	0.00	4	4.76	0.05	0.00	—	—	—	—
<i>L. youngi</i> *	2	0.64	0.01	0.00	1	1.19	0.01	0.00	—	—	—	—
Total	311	100	1	0.34	84	100	1	0.30	92	100	1	0.64

Number of sandflies (*N*); abundance (pi); Simpson index ( $\lambda$ ); anthropophilic species (\*); zoophilic species (\*\*).

performed with a level of significance of 0.005, Tukey's test. To investigate the possible association between species distribution and ecosystems a simple correspondence analysis was carried out using the IBM SPSS statistical software package, which is publicly available for download at <http://ibm-spss-statistics.softonic.com>.

### 3. Results

The ecoepidemiological characteristics and the degree of disturbance of the 3 agroecosystems are summarized in Table 1. The environmental characterisation demonstrated that

the studied areas presented different degrees of anthropogenic modification. The forest was more preserved and the orange-ry was more modified.

The Shannon traps, CDC light traps, Sticky traps, and direct suction captured 94.7%, 2.2%, 1.2%, and 0.9% of the sandflies, respectively. *L. gomezi* was the most abundant species in the area, present in all environments studied. According to the abundance values of sandfly specimens collected, *L. gomezi*, *L. ovallesi*, *L. walkeri*, *L. trinidadensis*, and *L. panamensis* were the main species identified in the 3 agroecosystems. These species were found at different abundance levels, although *L. panamensis* was only detected in the conserved forest (Table 2).

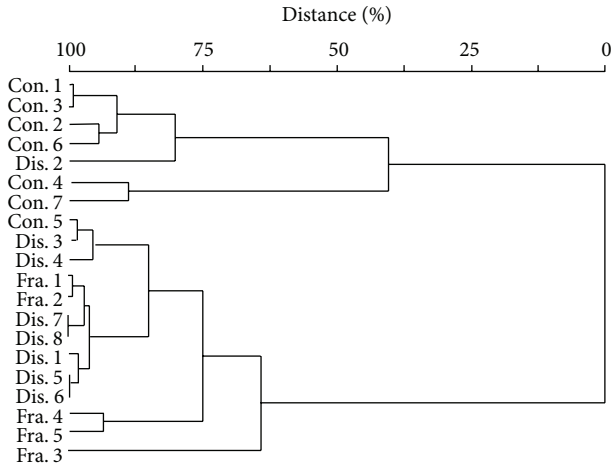


FIGURE 2: Cluster analysis of the capture in the agroecosystems.

TABLE 3: Analysis of variance (ANOVA) between the agroecosystems.

Agroecosystems	$\alpha$
Forest-cocoa	0.041*
Forest-orangery	0.073
Cocoa-orangery	0.001*

\*Significant differences in multiple comparison (alpha: 0.05).

Cluster analysis was performed to assess the segmentation of each capture, and we identified 2 groups of homogeneous captures with 46% similarity. These groups corresponded to captures in conserved and fragmented environments (Figure 2).

The ANOVA results showed significant differences between the populations of sandflies identified in each agroecosystem (one-way ANOVA,  $F = 551$ ,  $df = 16$ ,  $P = 0.000$ ). To further evaluate these differences, a *post hoc* Tukey's test was performed for paired agroecosystems, specifically between forest and cocoa agroecosystems and cocoa and orange agroecosystems (Table 3).

The highest values of diversity and species richness occurred in the most conserved agroecosystem, the forest (2.26 and 14, resp.). Moreover, the values for diversity and species richness decreased with an increasing degree of ecological disturbance, as observed with the cocoa (1.80 and 9) and orange agroecosystems (1.32 and 7, resp.). The dominance level was 0.34 in the forest and increased with an increasing degree of ecological disturbance, with the highest value corresponding to the orange grove agroecosystem (0.64) (Figure 3).

The simple correspondence analysis between sandflies species and agroecosystems identified a strong association between *L. gomezi* and *L. atroclavata* with disturbed agroecosystems and a strong association between *L. ovallesi*, *L. walkeri*, *L. shannoni*, *L. hernandezi*, *L. panamensis*, *L. migonei*, *L. cayenensis*, and *L. pilosa* with conserved agroecosystems; species such as *L. trinidadensis*, *L. olmeca nociva*, and *L. spinicrassa* showed no association with any agroecosystem

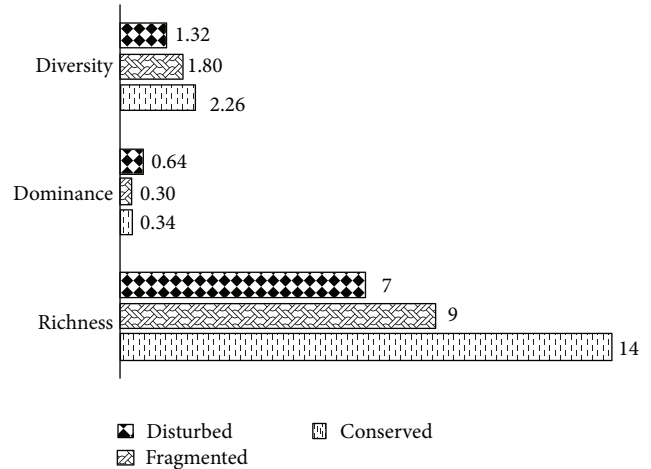


FIGURE 3: Dominance, diversity, and species richness in the three agroecosystems.

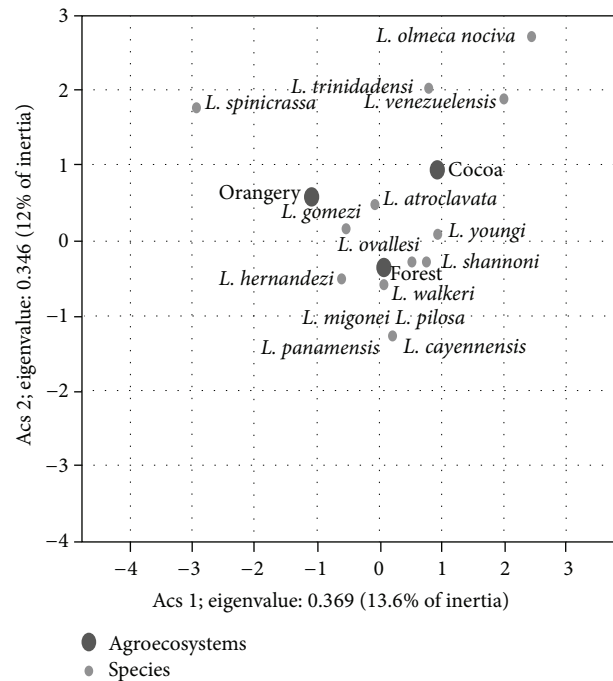


FIGURE 4: Association between sandflies species and agroecosystems by the simple correspondence analysis.

( $\chi^2$ : 124.7;  $df = 30$ ;  $P = 0.005$ ) (Figure 4). In the conserved agroecosystem, *L. gomezi*, *L. ovallesi*, and *L. walkeri* demonstrated natural infection with *Leishmania* species, which were identified as the subgenera *Leishmania* and *Viannia*.

#### 4. Discussion

Human encroachment on forest ecosystems is driven by logging and agricultural conversion, resulting in sharp and rapidly moving gradients between the relatively cool and

humid primary forest and the cultured land, which show strong insolation, higher temperature, and lower humidity. Tropical areas are characterized by a great diversity and wide distribution of sandflies fauna [19, 20]. In Brazil, it has been reported that the devastation of natural areas, which includes natural habitats for sandflies, increases the adaptability of these species to environments with human intervention, as observed by the increasing number of cases of leishmaniasis in urban environments [21].

It is likely that habitat degradation and climate change greatly impact the abundance and richness of sandflies. The results of this study highlight differences in the sandflies population composition and structure across 3 agroecosystems, characterized by the different degrees of ecological disturbance that were surveyed. Few studies on sandflies have focused on this aspect, as most reports have been limited to epidemiological studies and the documentation of naturally infected species [21]. In addition, other studies have focused on how the population composition changes in different areas, such as the home and peridomestic or wild environments [22–24], or according to the type of capture method used [25]. The results are in concert with others who have proposed that changes in habitat may have a marked impact on the sandflies populations [5–10].

The relationship between leishmaniasis and agricultural activity has been recorded and the relationship between coffee cultivation and the transmission of *Leishmania* by sandflies has been recorded in Venezuela, Colombia, Brazil, and Mexico [11, 26–29]. This could be explained by the suitability of shade-grown coffee plantations for the resting and breeding of sandflies. Moreover, this type of agroecosystem presents high biodiversity and promotes the presence of many vertebrates, which in turn act as reservoirs of *Leishmania* and potential feeding sources for sandflies [30, 31].

In this study, the effect of human intervention was reflected in the disturbed agroecosystems as an increase in dominance and a decline in diversity and species richness, relative to less ecologically disturbed areas such as the conserved agroecosystem, where dominance is lower and diversity and species richness are greater. These results are supported by those of previous studies [3, 21, 32]. Most diversity and species richness in the forest, conserved area, could be caused by higher accumulation organic material to accumulate as a result of the decomposition of leaves and vegetation waste lying on the soil favoring larval development.

Environments with significantly disturbed wilderness areas cause certain species to adapt to these new spaces, as observed in our study. Moreover, our results show that anthropogenic modification can favor certain species to colonize these disturbed environments, such as was reported for *L. longipalpis* [33] and *L. flaviscutellata* in urban areas of Brazil [34]. Few species are able to adapt to high levels of anthropogenic disturbance, consequently, demographic parameters such as mortality and birth rates for each species are affected differently, and ecosystem structure and dynamics are in turn affected; yet based on the abundance values, our results suggest that *L. gomezi* was the species with the greatest ability to exploit disturbed environments [3, 35].

Both *L. gomezi* and *L. ovallesi* have been considered as important vectors of *Leishmania* [36]. The type of agroecosystem affected the abundance of *L. gomezi* and *L. ovallesi* which have an important effect on the probability of humans being bitten by one of these two vectors. *L. gomezi* has been reported to have a marked preference for biting humans around homes where vegetation is scarce [37], and this species has also been known to invade the inside of the home [36, 37]. These findings suggest a greater risk of transmission of the disease in these areas.

The abundance of *L. ovallesi*, a species that transmits *Leishmania braziliensis*, has also been confirmed as a vector of *Leishmania mexicana* in Venezuela, and in conserved areas such as forests, a potential natural habitat and fragmented areas with cocoa plantations confirm the association of this species with woody vegetation [38–40]. The sympatric relationship between *L. ovallesi* and *L. gomezi* is comparable to what was reported in Brazil between *L. intermedia* and *L. neivai* [21], where *L. ovallesi* is the species with a greater dependence on conserved areas than *L. gomezi*, predominated near the peridomestic, indicating a process of adaptation, mainly to this environment of less dense vegetation. *L. gomezi* and *L. ovallesi* as predominant species of primary forest, as the deforestation extended, there was a tendency for *L. ovallesi* to disappear, suggesting that this species is more dependent on the primary forest than *L. gomezi*.

In the conserved forest agroecosystem, *L. gomezi*, *L. ovallesi*, and *L. walkeri* demonstrated natural infection with *Leishmania* of the subgenera *Leishmania* and *Viannia*, and this seems to indicate that these species may be transmitting the leishmaniasis agent in the forest agroecosystem area. If these areas have a greater diversity of sandflies species, it would be expected that there would be a greater coexistence of various species of *Leishmania*, given the specificity between the sandflies vector and *Leishmania*. Moreover, the increased abundance of *L. gomezi* in disturbed agroecosystems indicates that this species has adapted to new environments modified by humans. The altered environments favor adaptation of *L. gomezi*; these results suggest that the transmission pattern may be changing.

This study provides a basis for further in-depth studies to assess how anthropogenic changes can modulate vector composition and distribution and could also help to explain how this might affect the transmission of tegumentary leishmaniasis in Merida and potentially disease risks.

## 5. Conclusion

These results clearly show that sandflies fauna exhibited changes in species number as well as population structure in degraded environments. As a result, changes in the determinants of transmission can lead to the development of new outbreaks.

## Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

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## References

- [1] A. E. Jimenez, J. C. Rojas, F. Vargas, and M. V. Herrero, "Temporal and spatial variation of phlebotomine (Diptera: Psychodidae) community diversity in a cutaneous leishmaniasis endemic area of Costa Rica," *Journal of Medical Entomology*, vol. 37, no. 2, pp. 216–221, 2000.
- [2] B. L. Travi, G. H. Adler, M. Lozano, H. Cadena, and J. Montoya-Lerma, "Impact of habitat degradation on phlebotominae (Diptera: Psychodidae) of tropical dry forests in Northern Colombia," *Journal of Medical Entomology*, vol. 3, no. 22, pp. 451–456, 2002.
- [3] A. Valderrama, M. G. Tavares, and J. D. A. Filho, "Anthropogenic influence on the distribution, abundance and diversity of sandfly species (Diptera: Phlebotominae: Psychodidae), vectors of cutaneous leishmaniasis in Panama," *Memórias do Instituto Oswaldo Cruz*, vol. 106, no. 8, pp. 1024–1031, 2011.
- [4] P. Desjeux, "The increase in risk factors for leishmaniasis worldwide," *Transactions of the Royal Society of Tropical Medicine and Hygiene*, vol. 95, no. 3, pp. 239–243, 2001.
- [5] R. Bonfante, S. Barroeta, M. Mejía, E. Meléndez, C. Arredondo, and R. Urdaneta, "Leishmaniasis tegumentaria urbana en Barquisimeto, Venezuela," *Boletín de la Oficina Sanitaria Panamericana*, vol. 97, pp. 105–109, 1984.
- [6] J. F. Walsh, D. H. Molyneux, and M. H. Birley, "Deforestation: effects on vector-borne disease," *Parasitology*, vol. 106, pp. S55–S75, 1993.
- [7] C. M. Aguilar, E. Fernández, R. Fernández et al., "Urban visceral leishmaniasis in Venezuela," *Memórias do Instituto Oswaldo Cruz*, vol. 93, no. 1, pp. 15–16, 1998.
- [8] L. A. Agudelo, J. Uribe, D. Sierra, F. Ruíz, and I. D. Vélez, "Presence of American cutaneous leishmaniasis vectors surrounding the City of Medellín, Colombia," *Memórias do Instituto Oswaldo Cruz*, vol. 67, no. 5, pp. 641–642, 2002.
- [9] E. E. Bejarano, S. Uribe, W. Rojas, and I. Darío Vélez, "Presence of *Lutzomyia evansi*, a vector of American visceral leishmaniasis, in an urban area of the Colombian Caribbean coast," *Transactions of the Royal Society of Tropical Medicine and Hygiene*, vol. 95, no. 1, pp. 27–28, 2001.
- [10] F. D. O. P. Dias, E. S. Lorosa, and J. M. M. Rebêlo, "Blood feeding sources and peridomiciliation of *Lutzomyia longipalpis* (Lutz & Neiva, 1912) (Psychodidae, Phlebotominae)," *Cadernos de Saúde Pública*, vol. 19, no. 5, pp. 1373–1380, 2003.
- [11] J. Scorza, "Cambios epidemiológicos de la leishmaniasis tegumentaria en Venezuela," *Boletín de la Dirección de Malaria y Saneamiento Ambiental*, vol. 25, pp. 7–14, 1985.
- [12] J. Scorza, "La epidemiología de la leishmaniasis tegumentaria en Venezuela: situación actual," *Boletín de la Dirección de Malaria y Saneamiento Ambiental*, vol. 28, pp. 3–4, 1988.
- [13] L. Muñiz, R. Rossi, H. Nietzsche, W. Monteiro, and U. Teodoro, "Estudo dos hábitos alimentares de flebotomíneos em área rural no sul do Brasil," *Memórias do Instituto Oswaldo Cruz*, vol. 19, pp. 1087–1093, 2006.
- [14] W. Monteiro, H. Nietzsche, M. Lonardon, T. Silveira, M. Ferreira, and U. Teodoro, "Distribuição geográfica e características epidemiológicas da leishmaniose tegumentar americana em áreas de colonização antiga do estado do Paraná, Sul do Brasil," *Memórias do Instituto Oswaldo Cruz*, vol. 24, pp. 1291–1303, 2008.
- [15] C. M. D. O. Legriffon, K. R. Reinhold-Castro, V. C. Fenelon, H. C. Nietzsche-Abreu, and U. Teodoro, "Sandfly frequency in a clean and well-organized rural environment in the State of Paraná, Brazil," *Revista da Sociedade Brasileira de Medicina Tropical*, vol. 45, no. 1, pp. 77–82, 2012.
- [16] R. Lainson and J. Shaw, "The Role of animals in the epidemiology of South American leishmaniasis," in *Biology of the Kinetoplastida*, pp. 1–116, Academic Press, London, UK, 1979.
- [17] D. Young and M. Duncan, "Guide to the identification and geographic distribution of *Lutzomyia* sandflies in México, the West Indies, Central and South America (Diptera: Psychodidae)," *Memoirs of the American Entomological Institute*, vol. 54, pp. 779–881, 1994.
- [18] A. Magurran, *Ecological Diversity and Its Measurement*, vol. 101, Croom Helm Ltd Londres, Gran Bretaña, UK, 1988.
- [19] K. Kuhn, "Global warming and leishmaniasis in Italy," *Tropical Medicine of Institute Health*, vol. 7, pp. 1–2, 1999.
- [20] R. Killick-Kendrick, "The life: cycles of *Leishmania* in the sand fly and transmission of leishmaniasis by bite," *International Canine Leishmaniasis Forum*, vol. 57–59, 2002.
- [21] L. Saraiva, G. De Lima, C. Castilho, D. Alves, and J. Dilermando, "Biogeographical aspects of the occurrence of *Nyssomyia neivai* and *Nyssomyia intermedia* (Diptera: Psychodidae) in a sympatric area of the Brazilian savannah," *Memórias do Instituto Oswaldo Cruz*, vol. 107, no. 7, pp. 867–872, 2012.
- [22] R. Borges, B. Blanco, H. De Lima, M. Ortega, J. Morales, and W. Galindo, "Epidemia de leishmaniasis tegumentaria americana en el municipio El Hatillo del estado Miranda," *Gaceta Médica Caracas*, vol. 112, p. 249, 2004.
- [23] D. Cazorla and P. Morales, "Fauna flebotomina (Diptera: Psychodidae) del estado Falcón, Venezuela," *Revista Peruana Biología*, vol. 19, no. 1, pp. 75–80, 2012.
- [24] T. Machado, M. Lima, C. Lima, and J. Andrade, "Species diversity of sandflies (Diptera: Psychodidae) during different seasons and in different environments in the district of Taquarucú, state of Tocantins," *Memórias do Instituto Oswaldo Cruz*, vol. 107, no. 7, pp. 955–959, 2012.
- [25] A. M. Da Silva, N. J. De Camargo, D. R. Dos Santos et al., "Diversity, distribution and abundance of sandflies (Diptera: Psychodidae) in Parana State, Southern Brazil," *Neotropical Entomology*, vol. 37, no. 2, pp. 209–225, 2008.
- [26] A. Warburg, J. Montoya-Lerma, C. Jaramillo, A. L. Cruz-Ruiz, and K. Ostrovska, "Leishmaniasis vector potential of *Lutzomyia* spp. in Colombian coffee plantations," *Medical & Veterinary Entomology*, vol. 5, no. 1, pp. 9–16, 1991.
- [27] B. Alexander, E. Barbosa De Oliveria, E. Haigh, and L. Leal De Almeida, "Transmission of *Leishmania* in coffee plantations of Minas Gerais, Brazil," *Memórias do Instituto Oswaldo Cruz*, vol. 97, no. 5, pp. 627–630, 2002.
- [28] B. Alexander, L. A. Agudelo, J. F. Navarro et al., "Relationship between coffee cultivation practices in Colombia and exposure

- to infection with *Leishmania*,” *Transactions of the Royal Society of Tropical Medicine and Hygiene*, vol. 103, no. 12, pp. 1263–1268, 2009.
- [29] J. Pérez, A. Virgen, J. C. . Rojas et al., “Species composition and seasonal abundance of sandflies (Diptera: Psychodidae: Phlebotominae) in coffee agroecosystems,” *Memórias do Instituto Oswaldo Cruz*, vol. 109, pp. 259–266, 2014.
- [30] G. Ibarra-Núñez, “Los artrópodos asociados a cafetos en un cafetal mixto del Soconusco, Chiapas, México. I. Variedad y abundancia,” *Folia Entomologica Mexicana*, vol. 79, pp. 207–231, 1990.
- [31] I. Perfecto, R. A. Rice, R. Greenberg, and M. E. Van Der Voort, “Shade coffee: a disappearing refuge for biodiversity: shade coffee plantations can contain as much biodiversity as forest habitats,” *BioScience*, vol. 46, no. 8, pp. 598–608, 1996.
- [32] M. Souza, M. Marzochi, R. Carvalho et al., “Ausência da *Lutzomyia longipalpis* em algumas áreas de ocorrência de leishmaniose visceral no Município do Rio de Janeiro,” *Cadernos de Saúde Pública*, vol. 19, pp. 1881–1888, 2004.
- [33] R. Lainson, “Demographic changes and their influence on the epidemiology of the American leishmaniasis,” *Demography and Vector-Borne Diseases*, vol. 85, p. 106, 1989.
- [34] R. Souza, J. Lima, F. Souza, A. Gadelha, and V. Braga, “Establecimiento de un criadouro natural de *Lutzomyia longipalpis*,” *Revista da Sociedade Brasileira de Medicina Tropical*, vol. 32, pp. 214–215, 1999.
- [35] A. T. Peterson and J. Shaw, “*Lutzomyia* vectors for cutaneous leishmaniasis in Southern Brazil: ecological niche models, predicted geographic distributions, and climate change effects,” *International Journal for Parasitology*, vol. 33, no. 9, pp. 919–931, 2003.
- [36] E. Nieves, N. Villarreal, M. Rondón, M. Sánchez, and J. Carrero, “Evaluación de conocimientos y prácticas sobre la leishmaniasis tegumentaria en un área endémica de Venezuela,” *Biomédica*, vol. 28, pp. 347–356, 2008.
- [37] D. Feliciangeli, “La fauna flebotómica (Diptera, Psychodidae) en Venezuela: I Taxonomía y distribución geográfica,” *Boletín de la Dirección de Malaria y Saneamiento Ambiental*, vol. 28, no. 34, p. 25, 1988.
- [38] N. Añez, E. Nieves, D. Cazorla, M. Oviedo, A. Lugo de Yarub, and M. Valera, “Epidemiology of cutaneous leishmaniasis in Merida, Venezuela. III. Altitudinal distribution, age structure, natural infection and feeding behaviour of sandflies and their relation to the risk of transmission,” *Annals of Tropical Medicine and Parasitology*, vol. 88, no. 3, pp. 279–287, 1994.
- [39] D. Feliciangeli, “Sobre los flebotomos (Diptera: Psychodidae: Phlebotominae), con especial referencia a las especies conocidas en Venezuela,” *Acta Biologica Venezuelana*, vol. 26, no. 2, pp. 61–80, 2006.
- [40] A. Jorquera, R. González, E. Marchán-Marcano, M. Oviedo, and M. Matos, “Multiplex-PCR for detection of natural *Leishmania* infection in *Lutzomyia* spp. captured in an endemic region for cutaneous leishmaniasis in state of Sucre, Venezuela,” *Memorias do Instituto Oswaldo Cruz*, vol. 100, no. 1, pp. 45–48, 2005.